# **VOLUME 1**

# COASTAL FISHES OF THE WESTERN UNDER THE WESTERN UNDER THE SECOND STATE OF THE SECOND S

**EDITED BY** 

Phillip C Heemstra • Elaine Heemstra • David A Ebert • Wouter Holleman • John E Randall

# COASTAL FISHES OF THE WESTERN INDIAN OCEAN

# **VOLUME 1**

EDITED BY Phillip C Heemstra Elaine Heemstra David A Ebert Wouter Holleman John E Randall



Copyright © South African Institute for Aquatic Biodiversity, a National Research Facility of the National Research Foundation (NRF-SAIAB)

Published by the South African Institute for Aquatic Biodiversity, Private Bag 1015, Makhanda, 6140, South Africa

First edition 2022

Print ISBNs:

#### Hard covers

Soft covers	
978-1-990951-27-5	(Volume 5)
978-1-990951-26-8	(Volume 4)
978-1-990951-25-1	(Volume 3)
978-1-990951-24-4	(Volume 2)
978-1-990951-23-7	(Volume 1)
978-1-998950-40-9	(Set)

978-1-998950-41-6	(Set)
978-1-998950-35-5	(Volume 1)
978-1-998950-36-2	(Volume 2)
978-1-998950-37-9	(Volume 3)
978-1-998950-38-6	(Volume 4)
978-1-998950-39-3	(Volume 5)

#### Electronic (PDF) ISBNs:

978-1-990951-28-2	(Volume 1)
978-1-990951-29-9	(Volume 2)
978-1-990951-30-5	(Volume 3)
978-1-990951-31-2	(Volume 4)
978-1-990951-32-9	(Volume 5)

Copyright in the text is vested in the authors.

All rights reserved. No part of this publication may be reproduced or transmitted in any form or by any means, electronic or mechanical, including photocopying, recording, or any such information storage or retrieval system, without prior permission in writing from the publisher.

The authors and publisher have made every effort to obtain permission for and to acknowledge the use of any copyrighted material. Should any infringement of copyright have occurred, please contact the publisher, and every effort will be made to rectify omissions or errors in the event of a reprint or future edition.

The publisher gratefully acknowledges the funding from the South African Department of Science and Innovation (DSI) toward the publication of this work.

The views expressed in this publication do not reflect those of the National Research Foundation or the South African Institute for Aquatic Biodiversity, unless specifically stated.

Copy editor: Cindy Kulongowski

Proofreaders: Jenny Gon, Liz Gowans and Robert McKenzie Design and layout: Ink Design Publishing Solutions, Cape Town, www.inkdesign.co.za Front cover photographs: Allen Walker Photography (www.awphotosa.com) Inside cover maps: Willem Coetzer and Susan Abraham Printed by CADAR Printers, Gqeberha, South Africa

# CONTENTS

#### INTRODUCTION AND SUPPLEMENTARY MATERIAL

Foreword: Dr Peter Nick Psomadakis 2
Foreword: Dr Philemon Mjwara 4
Origins
Acknowledgements 7
Introduction: about this book 11
How to use this book
Naming organisms and determining their relationships 16
Aspects of the biology of fishes
Mimicry and protective resemblance in WIO fishes 33
The oceanography of the Western Indian Ocean
The origins and geology of reefs of the
Western Indian Ocean 46
Socotra Archipelago: fishes and fisheries in the eastern
Gulf of Aden 64
Fisheries of the Western Indian Ocean78
The oceanography and fisheries along the west coast of
India and at Lakshadweep 89
The collectors: ichthyological exploration of the
Western Indian Ocean
Contributors
Abbreviations and acronyms
Glossary
Bibliography 153

#### SYSTEMATIC ACCOUNTS

#### **MYXINIFORMES AND PETROMYZONTIFORMES:**

LIVING REPRESENTATIVES OF JAWLESS FISHES, AND THEIR
RELATIONSHIPS WITH OTHER VERTEBRATES
FAMILY MYXINIDAE – Hagfishes
CLASS CHONDRICHTHYES – CARTILAGINOUS FISHES
Origin and evolution of the Chondrichthyes 386
Coastal chondrichthyans of the Western Indian Ocean 394
Anatomy of sharks, rays and chimaeras
Key to families of chondrichthyans 404
ORDER HEXANCHIFORMES410
FAMILY CHLAMYDOSELACHIDAE – Frilled sharks 410
FAMILY HEXANCHIDAE – Cow sharks

ORDER ECHINORHINIFORMES416
FAMILY ECHINORHINIDAE – Bramble sharks
ORDER SQUALIFORMES418
FAMILY SQUALIDAE – Dogfishes
FAMILY CENTROPHORIDAE – Gulper sharks 424
FAMILY ETMOPTERIDAE – Lanternsharks
FAMILY SOMNIOSIDAE – Sleeper sharks
FAMILY OXYNOTIDAE – Roughsharks or
prickly dogfishes 441
FAMILY DALATIIDAE – Kitefin sharks
ORDER PRISTIOPHORIFORMES
FAMILY PRISTIOPHORIDAE – Sawsharks
ORDER SQUATINIFORMES450
FAMILY SQUATINIDAE – Angel sharks
ORDER HETERODONTIFORMES452
FAMILY HETERODONTIDAE – Bullhead sharks 452
ORDER ORECTOLOBIFORMES454
FAMILY HEMISCYLLIIDAE – Bamboo sharks
FAMILY STEGOSTOMATIDAE – Zebra shark
FAMILY GINGLYMOSTOMATIDAE – Nurse sharks 459
FAMILY RHINCODONTIDAE – Whale shark
ORDER LAMNIFORMES463
FAMILY CARCHARIIDAE – Raggedtooth shark 463
FAMILY ODONTASPIDIDAE – Sandtiger sharks 464
FAMILY MITSUKURINIDAE – Goblin shark
FAMILY PSEUDOCARCHARIIDAE – Crocodile shark 467
FAMILY MEGACHASMIDAE – Megamouth shark 468
FAMILY ALOPIIDAE – Thresher sharks
FAMILY CETORHINIDAE – Basking shark
FAMILY LAMNIDAE – Mackerel sharks
ORDER CARCHARHINIFORMES476
FAMILY PENTANCHIDAE – Deepwater catsharks 476
FAMILY SCYLIORHINIDAE – Catsharks
FAMILY PROSCYLLIIDAE – Finback catsharks 493
FAMILY PSEUDOTRIAKIDAE – False catsharks 496
FAMILY TRIAKIDAE – Houndsharks
FAMILY HEMIGALEIDAE – Weasel sharks 505
FAMILY GALEOCERDONIDAE – Tiger shark
FAMILY CARCHARHINIDAE – Requiem sharks 510
FAMILY SPHYRNIDAE – Hammerhead sharks538

ORDER TORPEDINIFORMES543
FAMILY TORPEDINIDAE – Torpedo rays 543
FAMILY NARCINIDAE – Numbfishes
FAMILY NARKIDAE – Sleeper rays552
ORDER RHINOPRISTIFORMES558
FAMILY PRISTIDAE – Sawfishes558
FAMILY RHINIDAE – Shark ray and wedgefishes 562
FAMILY RHINOBATIDAE – Guitarfishes and
shovelnose rays 565
FAMILY GLAUCOSTEGIDAE – Giant guitarfishes 572
ORDER RAJIFORMES576
FAMILY RAJIDAE – Hardnose skates
FAMILY ARHYNCHOBATIDAE – Softnose skates 591
FAMILY ANACANTHOBATIDAE – Legskates
FAMILY GURGESIELLIDAE – Pygmy skates
ORDER MYLIOBATIFORMES

FAMILY PLESIOBATIDAE – Deepwater stingray 598
FAMILY DASYATIDAE – Stingrays 598
FAMILY GYMNURIDAE – Butterfly rays
FAMILY MYLIOBATIDAE – Eagle rays619
FAMILY AETOBATIDAE – Pelagic eagle rays
FAMILY RHINOPTERIDAE – Cownose rays
FAMILY MOBULIDAE – Devilrays628
ORDER CHIMAERIFORMES634
FAMILY CALLORHINCHIDAE – Elephantfishes 634
FAMILY RHINOCHIMAERIDAE – Longnose chimaeras 635
Colour plates see separate PDF

Scientific and common name indexes (Volume 1)

	• • •	see separate PDF
--	-------	------------------

COASTAL FISHES OF THE WESTERN INDIAN OCEAN

INTRODUCTION AND SUPPLEMENTARY MATERIAL

# FOREWORD

## Dr Peter Nick Psomadakis

*Coastal Fishes of the Western Indian Ocean* is the third of a series of extremely popular ichthyological books on southern African and Seychellois marine fish fauna produced by NRF-SAIAB. It fills a longstanding gap as existing reference publications, such as the *FAO Species Identification Sheets for Fishery Purposes, Western Indian Ocean (Fishing Area 51)* and NRF-SAIAB's *Smiths' Sea Fishes,* both released in the mid-1980s, are now largely out of date.

This authoritative publication follows the fine tradition of producing multi-authored, well-illustrated volumes on fisheries resources by FAO and on ichthyofauna by NRF-SAIAB. The format is similar to the one presented in Smiths' Sea Fishes but is improved in two fundamental aspects. Well-illustrated keys to all taxa, including orders for bony fishes are now included. An ordinal-level key to bony fishes was not available in Smiths' Sea Fishes and its inclusion is an important innovation that greatly assists the non-expert user in the complex task of keying out an unidentified specimen starting from the ordinal level, passing through the familial level and down to its specific identity. The second important improvement is the coverage of nearly all species included in this publication with colour photographs of species in fresh/live condition or with beautifully coloured illustrations. This will be of great use to the non-expert user who is unfamiliar with discoloured specimens held in museum collections and finds the black & white illustrations/line drawings more difficult to apply for the purpose of species identification. Also of note is the inclusion of photographs/colour illustrations of colour morphs of some species (e.g. Rhinobatos punctifer), as well as many instances of juvenile colour patterns which are especially helpful to correctly identify those species where juveniles strikingly differ from adult colouration.

While the Western Indian Ocean is home to about 20% of the world's marine fish fauna, current marine biodiversity research efforts are inadequate and/or unequal across the region. As a result, identities and distributions of several species are inaccurately understood and reported in the literature. Taxonomic uncertainty and unavailability of formal species names implies that species are frequently misidentified or not recognised in fishery statistics which could have implications for their sustainable management and conservation.

In a scenario of climate change and global impacts on fish populations, it is important that fishery resources are managed according to an ecosystem approach to fisheries (EAF) which incorporates the marine ecosystem and biodiversity concerns into fisheries management. The principles of EAF are all embedded within the Code of Conduct of Responsible Fisheries and its importance is highlighted in the Blue Transformation initiative of FAO.

Fisheries research surveys at sea offer a unique opportunity to provide further information on marine species and document marine biodiversity. Since 1975, surveys at sea have been an integral part of the Norway & FAO Nansen Programme, and marine fish taxonomists employed at NRF-SAIAB have participated in selected surveys in the region on board the R/V Dr Fridtjof Nansen to support the work with identification of species in the catches and develop capacities of local scientists and for sample collection. NRF-SAIAB also hosts some of the reference collections for the Programme. Throughout time, this collaboration has yielded important scientific outputs, including the discovery and formal description of numerous new fish species, especially from surveys conducted in Oman, Yemen, Mozambique, Tanzania, Kenya, Madagascar, Seychelles, Maldives and the Mascarene Plateau. These discoveries show the value of surveys at sea for increasing our understanding of marine biodiversity while providing coastal states with important knowledge for sustainable management of their resources.

According to a recent IUCN assessment, the Western Indian Ocean has the highest level of uncertainty in species conservation status (16.9% of species assessed as Data Deficient) compared to other tropical regions. This means

that fundamental biological and ecological information to quantify trends in the population status of species occurring in the region is lacking—a condition that may hamper the progress towards international targets for biodiversity conservation, such as the United Nations Sustainable Development Goals (SDGs) and the Strategic Plan for the Convention on Biological Diversity (CBC).

*Coastal Fishes of the Western Indian Ocean* is the end product of over twenty years of collaboration between the editors Phillip C Heemstra (deceased), Elaine Heemstra, David A Ebert, Wouter Holleman, John E Randall (deceased) and more than 100 globally recognised experts (including scientific illustrators) on the various families of marine fishes occurring in the region. It will become the reference work for many years to come for both professional fish taxonomists and non-experts and is expected to improve the quality of identification and stimulate research on the biology and ecology of fish species in the region.

In congratulating the editors, authors and illustrators for this excellent production, I hope that the Institute will continue to lead research in marine fish biodiversity in the region, building on the legacy of the Smiths and all those who have devoted time and energy to the conception, production and realisation of this benchmark work.

#### Dr Peter Nick Psomadakis

EAF-Nansen Programme, Fisheries and Aquaculture Division, Food and Agriculture Organization of the United Nations (FAO) and Research Associate, Ichthyology – South African Institute for Aquatic Biodiversity (NRF-SAIAB)



# FOREWORD

## Dr Philemon Mjwara

Within South Africa's National System of Innovation, 'Aquatic Biodiversity' is a fragmented and unevenly attended discipline. As a National Research Facility of the National Research Foundation, which is an agency of the Department of Science and Innovation (DSI), the South African Institute for Aquatic Biodiversity (NRF-SAIAB) serves as a unifying force, covering the discipline from pure to applied science and infrastructure provision.

Twenty years since the Institute became a National Facility, the need for a national unifying force in the aquatic sciences is becoming increasingly relevant. South Africa's aquatic environment has key economic, social and environmental opportunities that are codified in the global Sustainable Development Goals, National Development Plan, Medium Term Strategic Framework, DSI White Paper and the DSI Decadal Plan.

Following the success of its previous international collaborative publication project, *Smiths' Sea Fishes* (first published in 1986), NRF-SAIAB – at that time known as the JLB Smith Institute of Ichthyology – embarked on a similar, but more ambitious, long-term project that provided the means to identify known shallow-water fishes from the tropical and warm-temperate waters of the entire Western Indian Ocean (WIO).

This project generated renewed interest worldwide in the diversity of fishes in the WIO, to the extent that new species of marine fishes from the WIO are still being described almost every month by scientists from Australia, Brazil, Britain, Japan and elsewhere. Over the years, several taxonomists from other countries have travelled to South Africa to come and work on the fish collections at NRF-SAIAB with taxonomists based at the Institute. This multi-volume work covers some 3 600 species of fishes in the largest area of ocean a book like this has ever covered – from Cape Point [18° E] in South Africa, to the Red Sea, Persian/Arabian Gulf and west coast of India, as far as Kanyakumari (formerly Cape Comorin), the southernmost point of India [~80° E] – as well as all the WIO islands north of 40° S and west of 80° E.

The resulting publication, far more substantial than JLB Smith could have imagined, is arranged in five volumes, and covers species that occur in waters generally shallower than 200 m. In addition, the publication includes the oceanography of the WIO, the origins of coral reefs, an account of the people who laid the foundations of our knowledge of the fishes of the WIO, the evolution of bony fishes, maps of the WIO, and more.

Globally South Africa is considered to be the 3rd most diverse country in the world. This current publication reflects the important contributions that the country is making to the world classification of species. The DSI congratulates the authors and the editors who have contributed to this seminal work that is globally highlighting South African expertise.

mit unisara

#### Dr Philemon Mjwara

Director-General: Department of Science and Innovation, South Africa



Science & innovation Department: Science and Innovation REPUBLIC OF SOUTH AFRICA

# ORIGINS

## Michael N Bruton

After JLB Smith had described the first living coelacanth, caught off East London in December 1938, he predicted that it was a stray from tropical reefs up the east coast of Africa. As the internal organs of the first coelacanth had been discarded, he was determined to find and examine another specimen. After World War II, he mounted a series of fishcollecting expeditions into East Africa with his wife, Margaret, searching for coelacanths while also making extensive collections of other sea fishes. He had realised that, in order to fully understand the ichthyofauna of South Africa, he needed to know more about East African fishes, as many local species occurred or had close relatives further north.

Smith's research on the collections that he and Margaret Smith made in South and East Africa made it possible for him to write the authoritative book, *The Sea Fishes of Southern Africa*, illustrated by Margaret and a team of artists, and published in 1949. But Smith was not satisfied. In the early 1950s he began planning a more comprehensive book on the fishes of the Western Indian Ocean as a companion volume to the *Sea Fishes* book. In 1949 he told a Trustee of the Natal Museum that he was planning a major work on the fishes of the Western Indian Ocean that would be "... at least 4 times the size of my current volume and would probably occupy 6–10 years". Smith had briefly contemplated accepting the post of Director of the Natal Museum so that he could be closer to the Western Indian Ocean region in order to complete this book, but this appointment never materialised.

Smith had, however, not as yet abandoned his lofty ambition. In 1950 he received a grant of £1 800 for the WIO book project from the CSIR. In 1955 he applied for (and received) a further grant of £5 000 for the WIO project but used these funds, quite legitimately, for publication of the *Ichthyological Bulletin*, a periodical produced by Rhodes University's Department of Ichthyology.

In the introductory pages to the 1953 edition of the *Sea Fishes* book Smith wrote, "A vast collection of East African fishes and many hundreds of valuable photographs and colour sketches have been assembled. There is at present in preparation a Companion East African Volume, in the same scale as this, and as fully illustrated, so as to cover the fishes of the whole Western Indian Ocean". But this project also never came to fruition.

As it turned out, this was wishful thinking. By the end of 1956 Smith realised that he had underestimated the enormity

of the project and expressed doubts that he would ever be able to complete it. On 1 December 1959 Stephan Meiring Naude, President of the CSIR, wrote to Smith, suggesting that he should reduce his work load and stating that his "... work is of such inestimable value to the country that you owe it to South Africa to take good care of yourself". Yet, on 12th June 1967, just six months before he died, Smith still wrote to Naude that he was hoping to "... live long enough to complete [the project] as it is likely to be the biggest work of all", and referred to it as the 'Sea Fishes of Mozambique'. This suggested that he had scaled down his vision for a WIO fishes book.

Instead, JLB and Margaret produced a series of carefully researched and well-illustrated scientific papers on the taxonomy and distribution of Western Indian Ocean fishes, family by family, which were published in the Ichthyological Bulletin, starting in 1956. By the time of JLB's death in January 1968, they had produced 32 reviews in the series, with a total of 682 pages, 100 pages more than the original Sea Fishes book. They had, therefore, achieved their goal from the scientific point of view, but the Ichthyological Bulletin publications were not designed for use by lay people. Fishes of the Seychelles, published by the Smiths in 1963, by contrast, was "designed to be of optimal value to anglers, skin divers, and other naturalists as well as scientists". This book covered 880 species, of which 95% are found in areas of the tropical Western Indian Ocean other than the Seychelles. The book was in great demand internationally and a second edition and reprints with addition of further species, especially of sharks, rays and clupeoids, were subsequently published.

Between the publication of the 1986 edition of *Smiths' Sea Fishes* (SSF) and the death of Margaret Smith in September 1987, informal discussions were held between then Director of the JLB Smith Institute of Ichthyology, Mike Bruton, past Director, Margaret Smith, and Phillip (Phil) Heemstra, Senior Curator of Marine Fishes at the Institute, on the feasibility of producing a WIO book, and the idea for the project was revived and its development encouraged, although it took some time to get off the ground. Paul Skelton, who became Director of the Institute in 1995, prodded it along, and by 2008 a small number of manuscripts had been submitted to Phil Heemstra. The project then gathered momentum, particularly when David Ebert agreed to coordinate the entire chondrichthyan fauna, and by 2014 most manuscripts had been submitted. Paul Skelton handed both the Institute, now the South African Institute for Aquatic Biodiversity (NRF-SAIAB), and the book project on to Angus Paterson, who became Managing Director in 2015 and took the project to fruition.

The dream of producing a book on the fishes of the Western Indian Ocean has now finally been realised by the staff of NRF-SAIAB and many international collaborators. The first steps were taken in 1993 by Phil Heemstra, with the backing of John (Jack) Randall of the Bishop Museum in Hawaii. A major contribution to this huge task was Phil's revision of 16 fish families for *FAO Species Identification Sheets for the Western Indian Ocean*. In 2004, after a great deal of further field and laboratory work, Phil and his wife, Elaine, Senior Illustrator at the Institute, also published *Coastal Fishes of Southern Africa* (CFSA).

JLB Smith's original dream has now been fulfilled with the publication by NRF-SAIAB of this multi-volume treatise, *Coastal Fishes of the Western Indian Ocean*, edited by Phillip C Heemstra, with Elaine Heemstra, David A Ebert, Wouter Holleman and John E Randall. It is the culmination of the work of more than 100 authors, photographers and illustrators from 16 countries over a period of more than 20 years, with major contributions from Phillip Heemstra, David Ebert, David Smith, Bruce Collette, Stuart Poss, John Randall, Gerald Allen, Helen Larson, Dannie Hensley, Kunio Amaoka, Eric Anderson and Keiichi Matsuura.

The fine tradition of producing multi-authored, wellillustrated volumes on South and East African sea fishes, pioneered by JLB and Margaret Smith, has been successfully continued by Phillip Heemstra. This world-class, multivolume book, *Coastal Fishes of the Western Indian Ocean*, is dedicated to the memory of JLB and Margaret Smith, and Phillip Heemstra.

# ACKNOWLEDGEMENTS

## The Editors

This project could not have been conceived – and realised – without the expertise, many hours of labour and endless patience from more than 100 contributors. In recognition, this first volume includes biographical sketches of these contributors. These contributors also provided many of the photographs used. Colleagues, friends and strangers (all fellow fish watchers) around the world were also approached for permission to use their photographs and, without exception, several hundred photographs were generously donated. Without them many species would not have been illustrated. Most prominent was the late John E Randall who contributed hundreds of images from his extensive library of species photographs.

It also takes the unflagging effort and encouragement of many people over many years to bring a project of this magnitude to fruition. It is almost inevitable that in recording the editors' collective thanks, names of people and institutions who have contributed in one way or another will somehow 'slip through the net', and we apologise at the outset for such omissions.

Our first thanks must go to the three Directors of the South African Institute for Aquatic Biodiversity ([NRF-SAIAB] formerly the JLB Smith Institute of Ichthyology): Michael Bruton, under whose watch *Coastal Fishes of the Western Indian Ocean* (CFWIO) was conceived in 1993; Paul Skelton, who was responsible for bringing the project into focus; and Angus Paterson, the current Managing Director, who supported and encouraged it through to completion.

The funding for such a venture is paramount and this book would not have been completed without the generous support of the South African Department of Science and Innovation and the National Research Foundation.

Dennis and Sally Polack provided the initial sponsorship for the project – without such seed funding the task would have been much more difficult. In addition, their friendship and enthusiastic support over many years has been invaluable. They also hosted several scientists during the course of CFWIO field work.

This work would not have been possible without the backup and support of all the staff at the NRF-SAIAB. In particular, we record our thanks to: Roger Bills, Linda Coetzee, Willem Coetzer, Kholiwe Dubula, Andrew Grant, Vuyani Hanisi, Penny Haworth, Hanoria Kalamashe, Bafo Konqobe, Mark Lisher, Sherwyn Mack, Bernard Mackenzie, Nkosinathi Mazungula, Maditaba Meltaf, Phumeza Mpambani, Vusi Mthombeni, Naniswa Nyoka, Jean Pote, Elvis Rungqu, Sally Schramm, Margie Shaw, Leslie Ter Morshuizen, Garth van Heerden and Joan Wright.

And then there are the people in the background without whom no work ever becomes published. Cindy Kulongowski did a fine job in copy editing the manuscripts which were then proofread by Robert McKenzie and Liz Gowans prior to layout. Stephen Walker of WalkerDigital was responsible for scanning all the plates of painted fishes. Roseanne Palmer contributed excellent drawings of photographs where needed. Susan Abraham helpfully recreated some of the illustrations for the introductory chapters, as well as finalising the inside-cover maps, in collaboration with Willem Coetzer. The bibliography was first compiled by Sheila Hicks from disparate sources. Aidan Wood contributed to completing the bibliography and also assisted with checking the page proofs. Particular thanks are due to Jenny Gon for her meticulous final proofreading of the page proofs.

Mike Schramm of NISC (Pty) Ltd was the independent eye who asked searching questions and whose publishing expertise guided the project through the later stages of the publication process. Liz Gowans tactfully guided the editorial and proofreading team; her efficient organisation, and coordination with the Ink Design design and layout team, greatly facilitated the process. Ink Design's creativity, layout proficiency, amenability to making changes, and endless patience during the lengthy production process is greatly appreciated. The end product is an accolade that cannot be bettered.

Sections of these volumes were reviewed by Gerald R Allen, M Eric Anderson, Geremy Cliff, Matthew T Craig, Brit Finucci, Phillip C Heemstra, Hisashi Imamura, Horst Kaiser, Rob W Leslie, John E McCosker, Hiroyuki Motomura, Richard L Pyle, Helen A Randall, Barry C Russell, David G Smith, Wayne C Starnes, Nico Straube, James van Hasselt, Simon Weigmann and Sabine P Wintner.

The invaluable reference resource provided by *Eschmeyer's Catalog of Fishes* underpinned this book. Where we were unsure, William Eschmeyer, Ronald Fricke and Richard van der Laan patiently and promptly answered the numerous questions our work generated.

Fish survey work which provided new species and fish distributions were undertaken with the help of many people who provided logistic support, hospitality, friendship and local expertise over two decades: Mark Addison and staff of Blue Wilderness, KwaZulu-Natal, South Africa; Guy and Caroline Fothergill and staff of Island Ventures, Grande Comore; Tara Lynch, Tom Hooper, Eric Blais and Sabrina Meunier of Shoals of Capricorn, Rodrigues; Riaz Aumeeruddy, Jude Bijoux, and staff of Seychelles Fishing Authority; staff of Albion Fisheries Research Centre and the Mauritius Oceanography Institute, both Mauritius; Neville and Wendy Ayliffe, Brian and Corlia Ring and staff of Reefteach, and Jenny Smart of Odysea, all from Sodwana Bay, South Africa; and Mike and Valda Fraser, and Ruth and Ginger Seipp, all from KwaZulu-Natal, South Africa. Many Rhodes students enthusiastically volunteered their help with diving, collection, sorting and preservation of fishes. Andrew Bentley, Brian Godfrey, Steven Warren and Malcolm Smale helped as dive supervisors.

NRF-SAIAB has over many decades benefitted from the input and help of fish enthusiasts, photographers, divers, aquarists, and anglers. There is a long history of citizen scientists photographing, collecting and sending interesting specimens that they had not seen before, often to find that the NRF-SAIAB ichthyologists did not recognise the species either. Or if they did, did not know of them occurring in that particular area. Their photographs (several of which appear in this book), specimens and detailed observations have led to the description of many new species and also documented species distributions. Their names and the many others who provided photographs are listed below.

A. EM Abdussamad, Robert Abela, Moteah Shaikh Aideed, KV Akhilesh, Gerald R Allen, Tim Allen, Vincent Altamirano, Oddgeir Alvheim, Kunio Amaoka, R Charles Anderson, M Eric Anderson, Susan G Anderson, T Aoki, RS Atherton, Neville Ayliffe, Abdul P Azeez. B. Avi Baranes, Alain Barrère, Georg Barsch, Hans Bath, Andrew C Bentley, Adam Ben-Tuvia, Ricardo Betancur-R, Gabriella Bianchi, Roger Bills, KK Bineesh, J Black, Sergey V Bogorodsky, A Bollen, Emma Booysen, Philippe Bouchet, Bruce Bowden, Brian Bowen, M Boyer, Dianne J Bray, Eran Brokovich, Marie Broodryk, Geer Brovad, JJ Brown, Henrich Bruggemann, Richard T Bryant, Suzanne Bullock, Colin D Buxton. C. Christopher Caine, Prosanta Chakrabarty, Russell Chalmers, Rong-Hua Chang, Simon Chater, CY Chen, Mikhail V Chesalin, Amber Childs, Paul J Clerkin, Rui Coelho, Allan D Connell, Paul D Cowley, P Crabb, Matthew T Craig, CSIRO Fish Collection, Digby P Cyrus. D. Gavin Dally, Ryan Daly, David Darom, Brian W Darvell, Armand Daydé, Helmut Debelius, Charles de la Harpe, John Dench, Greg de Valle, Padmavathi Devarapalli, Carlie Devine, Dominique A Didier, Alain Diringer, Jackie Docherty, Andrey Dolgov, Ryu Douichi, Christopher E Dowling, Clinton Duffy, Bernard Dupont, Alexei Dyer, Mike Dykhouse. E. David A Ebert, Graham J Edgar, Mark V Erdmann, Kerstin Erler, Jessica EscobarPorras, Bernadine Everett, Janet Eyre. F. Alessandro Falleni, Mike Farquhar, Sean T Fennessy, Claude Ferrara, Carl Ferraris, Richard Field, Alfredo Carvalho Filho, Darko First, Camilla Floros, Chris Fourie, Thomas Fraser, Valda & Mike Fraser. G. Qasem Gharibi, Anthony C Gill, Thomas Gloerfelt-Tarp, Dawn Goebbels, Daniel Golani, A Golubev, Martin F Gomon, Ofer Gon, Albrecht Götz, Gavin Gouws, Andrew Grant, Brian Gratwicke, David W Greenfield, Jim Greenfield, J Griffiths, T Britt Griswold, Stefano Guerrieri, Meneeka Gurroby. H. M Hackenberg, J Hall, Mark Harris, T Harrison, Eusuf Hasan, Kiyotaka Hatooka, Bert Hazes, Marika Hedin, Elaine Heemstra, Phillip C Heemstra, Rose Henderson, Dai Herbert, Juergen Herler, Gary Hermann, Yusuke Hibino, Hsuan-Ching Ho, Douglas Hoese, Wouter Holleman, Karen Honevcutt, John Hoover, Brett A Human. I. Hisashi Imamura, Yuoko Ishikawa, Yukio Iwatsuki. J. Zeehan Jaafar, Glenn Jacoby, Laith A Jawad, PT Jinesh, G David Johnson, Roel Jonkman, Gareth Jordaan, Jean-Lou Justine. K. Tatsuya Kaga, Sven Kahlbrock, Japie Kamminga, George Karamanos, Emma Karmovskaya, Eri Katayama, Shoishi Kato, Toshio Kawai, Philippe Keith, Maroof A Khalaf, M Moazzam Khan, Khin May Chit Maung, Hiroyuki Kimura, Seishi Kimura, Dennis R King, H Kishimoto, Robert Koch, Marc Kochzius, Keita Koeda, Kevin Kohen, Albert Kok, Marcelo Kovačić, Tassapon Krajangdara, Frank Krasovec, Yannick Kratochvil, Uwe Krumme, Friedhelm Krupp, Rudie H Kuiter, A Biju Kumar. L. Helen K Larson, Peter R Last, Pieter Laubscher, Martyn Lee, Po-Feng Lee, Jeffrey M Leis, Rob W Leslie, Ewald Lieske, Kelvin Lim, Hsin-Lua Lin, P-L Lin, Mark Lisher, Paul Loiselle, F Lombard, Pieter Loubser, Patrick Louisy, Roger Lubbock, Angela Lund. M. Gavin Macauley, Jade Maggs, Miles Mander, Mabel Manjaji-Matsumoto, Bruce Mann, Patrice Marker, Andrea D Marshall, Eve Marshall, A Arockia Mary, Keiichi Matsuura, C McCarthy, Linda J McCarthy, Jonathan KL Mee, R & U Menon, Nico K Michiels, Michael M Mincarone, Alec BM Moore, Kate A Moots, E Morcel, Michelle Morris, Hiroyuki Motomura, Andrew Murch, Ed Murdy, Monica Mwale, Robert F Myers. N. Hiroshi Nagano, Hiroyuki Nagano, Rekha Nair, Jeremy Neech, Amanda Northrop. O. Anker Odum, Makoto Okamoto, orangkucing, Pamela Osborn, Wayne Osborn, Hamid B Osmany. P. Stephen Pain, Ryan Palmer, Maurice Parmentier, Thomas Paulus, Adrian S Pearton, Daniel Pelicier, Juan Pereira, Rohan Pethiyagoda, Roberto Pillon, Dennis A & Sally Polack, Gianluca Polgar, Duncan Pritchard, Kim Prochazka, Artem Prokofiev, Peter N Psomadakis, Richard L Pyle. R. Helen A Randall, John E Randall, Sandra J Raredon, Sahat Ratmuangkhwang, Sohrab G Rezvani, Sonia Ribes, Laura Rich, Brian Ring, Ross Robertson, James Robey, Jean Louis Rose, Jean Michel Rose, Mark Rosenstein, Robert Rostol, Laila Rouhani, Ourban Rouhani, Barry C Russell, Andrey Ryanskiy. S. Raju Sarovan, D Sasaki, Kunio Sasaki, Ukkrit Satapoomin, Otto Schlumberger, Werner

Schwarzhans, Lucy Scott, Sue Scott, Hiroshi Senou, Seotara, Bernard Serét, Ilia Shakhovskoy, Kwang-Tsao Shao, David Shen, Charles RC Sheppard, Koichi Shibukawa, Masato Shiina, Maria I Shtaut, Ulrike E Siebeck, Kerry Sink, Hans Sjöholm, Malcolm J Smale, David G Smith, Tim Smith, Peter Southwood, John S Sparks, Geoffrey Spiby, A Spreinat, Victor G Springer, James Stapley, Matthias FW Stehmann, Darren Stevens, Richard Steventon, Robin E Stobbs, Jenny Strömvoll, Rick Stuart-Smith, Montri Sumontha, Andrey Suntsov, Alan Sutton, Déwald Swanepoel, Johan J Swanepoel, L Swart. T. T Takita, Heok Hui Tan, William Tan, Fumiya Tanaka, Hiroyuki Tanaka, Yi-Kai Tea, N Teichert, D Terver, Ralf Thiel, Sujitha Thomas, Craig Thomassen, Anthony R Thorpe, Kenneth Tighe, Peter Timm, Sven Tränkner, Alan Townsend, Michael N Trevor, Cheng-Yi Tsai. U. Franz Uiblein. V. Mark van Coller, Herman van der Bank, Carel van der Colff, Rudy van der Elst, Carel van der Westhuizen, Christo van Jaarsveld, Richard Vari, Benjamin C Victor, Hugues Vitry, Elyaperumal Vivekanandan, Archie Viviers. W. Allen Walker, Thorsten Walter, Dagny Warmerdam, Simon Weigmann, Will White, Alan K Whitfield, Jeffrey T Williams, Rex Williams, Keith DP Wilson, Rick Winterbottom, Peter Wirtz, A Wright, Ian Wyness. Y. Sukran Yalçin-Ozdilek, Z-L Yang. Z. Diana Zaera-Perez, Uwe Zajonz, Rickard Zerpe, Guido Zsilavecz.

Special thanks are due to illustrators who contributed paintings, diagrams and scientific illustrations. Their names (where available) are Brittany Atkinson, John Bass, M Brown, Mildred H Carrington, Pamela C Caruso, Peter HJ Castle, Manuela D'Antoni (Manu), Denys M Davis (DMD), Ryu Douichi, Barbara Duckworth, Ronald Fricke, Mary H Fuges (MHF), N Gasco, Anthony C Gill, Celia M Godkin, William Gosline, VM Gregory, Patti Hansen (now Hadley), Kiyotaka Hatooka, Elaine Heemstra (EG & EH), Phillip C Heemstra, Rose Henderson, Hsuan-Ching Ho, Mary Ann Holloway, Zeehan Jaafar (ZJ), Zbigniew T Jastrzebski, Toshio Kawai, Cindy Kulongowski, Helen K Larson, Paulo Lastrico (LP), O Lidonnici (Olid), Moira Lambert (ML), Anne Lefebvre (AL), Hester W Locke (HWL), Yasue Matthews, Virginia McCrostie (VMcR), Randolph McKay, Peter Miller, Michael Mincarone, SG Monden, K Hiratsuka Moore, Atsunobu Murase, Pierre Opic, Patricia Parkin (PP), Tracy D Pedersen (TDP), Mathieu Pinault, GN Pokhilskava, G Rankin, Kunio Sasaki, Jack R Schroeder (JRS), Margaret Smith (MMS), Liz Tarr (EMT), Betty Thorn, Dave Voorvelt (DPV), Jean Michel Vincent (JMV), Rick Winterbottom (RW), Thosaporn Wongratana (TW), Frances W Zweifel.

Many individuals need to be thanked for their help (in some cases, additional to that already noted above): EM Abdussamad, KV Akhilesh, Abdullah Al-Nahdi, Oddgeir Alvheim (IMR), Lee-Ann Anderson, Brenda Appelbaum-Golani (HUJ), Riaz Aumeeruddy, Satoshi Awato, Rhet Bennett, KK Bineesh, Heiko Bleher, Sergey V Bogorodsky, Philippe Bouchet (MNHN), Michael Bougaardt (Iziko-SAM), V Brown, John Butler, P Crabb (NHM), Matthew T Craig (NOAA), Oliver Crimmen (NHM), David Darom, Hiromitsu Endo (Kochi University), Sean T Fennessy (ORI/SAAMBR), Richard Field, Ralph Foster (Iziko-SAM), Mike & Valda Fraser, Daniel Golani (HUJ), Menachem Goren, Alistair Graham, Richard L Haedrich, Julia Heemstra, Lydia Heemstra, Yusuke Hibino, Hitoshi Ida, Hisashi Imamura, Tatsuva Kaga, Japie Kamminga, Emma Karmovskava, John Keulder, Maroof A Khalaf, Seishi Kimura, Dennis R King, Peter Koenigshof, Alexander Kotlyar, Fareed Krupp (SMF), Rudie H Kuiter, Yefim Kukuev, Hsin-Hua Lin, Paul Loiselle, Eben Lourens, James Maclaine (NHM), Bruce Mann (ORI/SAAMBR), Mizuki Matsunuma (KAUM), Peter Miller, Ed Murdy, Gota Ogihara, Loreen R O'Hara (BPBM), Sean O'Hara, Alexei Orlov, Lisa Palmer, Nik Parin, Mathieu Pinault, Patrice Pruvost (MNHN), Peter N Psomadakis (FAO), Sandra J Raredon (USNM), Gareth Roocroft, Mark H Sabaj Pérez (ANSP), Etsuro Sawai (HU), Kwang-Tsao Shao (ASIZ), Gento Shinohara, Kerry Sink (SANBI), Yonela Sithole (NRF-SAIAB), Matthias FW Stehmann, Pkee Stopforth, Tore Strømme, Arnold Suzumoto (BPBM), Sarah Sworder, Fumiya Tanaka, Harry Taylor (NHM), Ralf Thiel (UH), Alexis Tindall (NRF-SAIAB), Hans-Peter Tschorsnig, James Tyler, David van Beuningen, Diego Vaz, Sarah Viana, E Vignieux, Elena Voronina, Florian Wicker (SMF).

We would also like to acknowledge the help and contributions of the following institutions and their staff: ACEP — African Coelacanth Ecosystem Programme; ACRSD - Andaman Coastal Research Station for Development, Kasetsart University, Bangkok, Thailand; AFRC — Albion Fisheries Research Centre, Petite Riviere, Mauritius; AMNH — American Museum of Natural History, New York, USA; ANSP - Academy of Natural Sciences of Philadelphia, (now Academy of Natural Sciences of Drexel University), Philadelphia, USA; ASCLME - Agulhas and Somali Current Large Marine Ecosystems programme; ASIZ -Academia Sinica, Institute of Zoology, Taipei, Taiwan; AMS -Australian Museum, Sydney, Australia; AV2010 — Atimo Vatae expedition to Madagascar of the Muséum National d'Histoire Naturelle de Paris / Pro-Natura International / Institut Halieutique et des Sciences Marines / Wildlife Conservation Society Madagascar Program Expedition "Our Planet Reviewed" Initiative; BMRI-UMS - Borneo Marine Research Institute, Universiti Malaysia Sabah, Malaysia; BPBM — Bishop Museum, Honolulu, Oahu, Hawaii, USA; CAS - California Academy of Sciences, San Francisco, California, USA; CCMAR - Centre for Marine Sciences, Faro, Portugal; Chinese Academy of Fishery Sciences; CMFRI — Central Marine Fisheries Research Institute (Indian Council of Agricultural Research), India; CMI -College of the Marshall Islands, Majuro, Marshall Islands;

COMU - Canakkale Onsekiz Mart University, Turkey; CSIRO and CSIRO Australian National Fish Collection, Hobart, Tasmania, Australia; DPIRD — Department of Primary Industries and Regional Development, Western Australia; FAO — Food and Agriculture Organization of the United Nations, Original Scientific Illustrations Archive, Rome, Italy; FLMNH — Florida Museum of Natural History, Gainesville, USA; FRC-S — Fisheries Research Center-Salalah (Ministry of Agriculture and Fisheries Wealth), Oman; FRLM — Fisheries Research Laboratory, Mie University, Shima, Mie Prefecture, Japan; GCRL - Gulf Coast Research Laboratory, University of Southern Mississippi, Ocean Springs, Mississippi, USA; HUJ — Hebrew University of Jerusalem, Israel; HUMZ — Fisheries Science Center, Hokkaido University Museum, Hokkaido, Japan; ICAR —Indian Council of Agricultural Research, New Delhi, India; IIP - National Institute of Fisheries Research, Maputo, Mozambique; IMR - Institute of Marine Research, Bergen, Norway; IORAS - PP Shirshov Institute of Oceanology, Russian Academy of Sciences, Moscow, Russia; KAUM — Kagoshima University Museum, Kagoshima, Japan; KAUST — King Abdullah University of Science and Technology, Thuwal, Saudi Arabia; KPM -Kanagawa Prefectural Museum of Natural History, Kanagawa, Japan; KU — Kasetsart University, Bangkok, Thailand; also, University of Kansas Biodiversity Institute, Lawrence, Kansas, USA; LSU - Louisiana State University, Baton Rouge, Louisiana, USA; MAGNT - Museum and Art Gallery of the Northern Territory, Australia; MAMU — Macleay Museum, Sydney, Australia; MFDK — Marine Fisheries Department, Karachi, Pakistan; MMF — Marine Megafauna Foundation, Tofo, Mozambique; MNHN — Muséum national d'Histoire naturelle, Paris, France; MOI - Mauritius Oceanography Institute, Albion, Mauritius; MSNDC - MS Naicker Degree College, Kakinada, India; MUFS — Department of Plant and Animal Sciences, University of Miyazaki, Miyazaki, Japan; MZUSP — University of Sao Paulo, Museu

de Zoologia, Sao Paulo, Brazil; NHM — Natural History Museum, London, UK; NMMB-P — National Museum of Marine Biology and Aquarium, Pintung, Taiwan; NMSA -KwaZulu-Natal Museum, Pietermaritzburg, South Africa; NMV - Museum of Victoria, Melbourne, Australia; NOAA -National Oceanic and Atmospheric Administration, USA; NRF-SAEON - South African Environmental Observation Node, South Africa; NRF-SAIAB - South African Institute for Aquatic Biodiversity, Grahamstown/Makhanda, South Africa; OMMSFC — Oman Marine Science and Fisheries Centre, Oman; ORI/SAAMBR — Oceanographic Research Institute, Durban, South Africa; PEM - Port Elizabeth Museum, Gqerberha, South Africa; PINRO — Polar Research Institute of Marine Fisheries and Oceanography, Murmansk, Russia; PMBC — Phuket Marine Biological Centre, Thailand; RMBR — Raffles Museum of Biodiversity Research, Singapore; RU-DIFS — Department of Ichthyology and Fisheries Science, Rhodes University, Grahamstown/Makhanda, South Africa; SAM — South Australian Museum, Adelaide, Australia; SANBI — South African National Biodiversity Institute, Pretoria and Cape Town, South Africa; SFA - Seychelles Fisheries Authority, Mahé, Seychelles; SMF - Senckenberg Research Institute and Natural History Museum, Frankfurt, Germany; SURG — Southern Underwater Research Group, Cape Town, South Africa; Two Oceans Aquarium, Cape Town, South Africa; UFRJ — Universidade Federal do Rio de Janeiro, Brazil; UH - Biocenter Grindel und Zoological Museum, University of Hamburg, Germany; UJ – University of Johannesburg, South Africa; UoK — University of Kerala, India; Uni Wien - University of Vienna, Austria; USNM -National Museum of Natural History, Smithsonian Institution, Washington, DC, USA; UWAR — University of Warwick, UK; ZIHU — Hiroshima University, Department of Biological Sciences, Hiroshima, Japan; ZMT - Leibniz Center for Tropical Marine Ecology, Bremen, Germany; ZUMT -Department of Zoology Museum, University of Tokyo, Japan.

# **INTRODUCTION: ABOUT THIS BOOK**

The primary purpose of this book is to provide a means of identifying the more than 3 200 species of coastal fishes known to occur in the Western Indian Ocean (WIO). Coastal fishes are those that inhabit waters generally less than ~200 m deep, the waters over continental and insular shelves, and upper continental slopes. The book also includes some oceanic species and species that live in deeper water, but are sometimes caught in trawls in less than 200 m, or that migrate into shallower waters at night to feed.

The Western Indian Ocean (WIO), as treated in these volumes, is the area between Cape Point, South Africa, and 77°34' E, at Kanyakumari (formerly Cape Cormorin), the southernmost point of India, and to 40° S, just south of St Paul Island. Although considered as separate water bodies, the Red Sea and Persian/Arabian Gulf have been included. Some contributors have also chosen to include species from Sri Lanka. The region thus encompasses the entire east and southern coasts of Africa, Madagascar and the various island clusters of the Comoros, the Seychelles, the Maldive and Lakshadweep islands, the Chagos Archipelago and the islands and sea mounts of the Mascarene Plateau, to as far as 40° S. and thus some fishes from St Paul and Amsterdam Islands have been included. This large expanse, stretching from tropical waters of the northwestern Indian Ocean to the warm temperate waters of False Bay, South Africa, includes a number of poorly known biogeographic areas.

A map of the entire Indian Ocean is placed on the inside front cover of each printed volume, with some areas in greater detail on the inside back cover. The book does not include distribution maps for species, but gives localities from which species are known, with emphasis on WIO localities; our understanding of distributions of many species is often incomplete.

Fishes are the most abundant and diverse group of vertebrates and have colonised every aquatic habitat on Earth: the oceans, lakes, rivers and caves, from polar seas at -2 °C to hot, freshwater springs at 44 °C, and from tropical reefs and mangrove forests to the deepest ocean depths. Fishes are also the most poorly known group of vertebrates. In the 2006 edition of Joseph Nelson's *Fishes of the World* the estimate of the number of species of extant fishes worldwide stood at about 23 000. This number is growing annually, and was thought to be about 33 460 species at the end of 2016

(www.fishwisepro.com). Between the years 2000 and 2015 an average of 150 new species of marine fishes were described each year – of which 10% of the total (156 species) were from the WIO. The WIO is home to about 15% of all the marine fish species in the world's oceans.

Another measure of the diversity of fishes of this area is its relatively high level of endemicity, particularly around southern Africa and in the Red Sea. About 13% of southern African marine fishes are endemic, most of these in only five families: Clinidae with about 44 endemic species, Gobiidae with 28, Sparidae with 28, Pentanchidae with 6, and Batrachoididae with 7 endemic species. In the Red Sea at least 170 of the more than 1100 species are endemic.

The WIO region is also home to a large human population, representing a wide range of ethnic and cultural backgrounds. The area includes the countries of South Africa, Mozambique, Tanzania, Kenya, Somalia, Eritrea, Sudan, Egypt, Israel, Jordan, Saudi Arabia, Yemen, Oman, United Arab Emirates, Qatar, Bahrain, Kuwait, Iraq, Iran, Pakistan, India and Sri Lanka, as well as the many island nations and territories. Many of the people living in coastal areas are dependent on fish catches and other marine resources for both sustenance and often a livelihood, as highly diversified artisanal fisheries make up the bulk of the fishing effort in the region.

And, as elsewhere in the world, many of the fish resources have been compromised by commercial interests (including those of other countries), often leaving fish stocks in a poor state. This book has a number of purposes, all of which coalesce around providing users with a better understanding of the area's fishes and their environment. Accordingly, it includes a number of background chapters covering subjects as diverse as the oceanography of the region, and the history and evolution of the bony fishes.

In recent years genetic analysis has proved to be a powerful tool for taxonomists. In many instances molecular results have caused taxonomists to rethink both the definitions of certain taxa and the interrelationships of taxa. In some instances, what were long considered cohesive (monophyletic) taxa were found to include groups of fishes that are in fact not closely related (paraphyletic), while in other instances taxa thought to be distinct were found not to be, meriting their merging with other exiting taxa. At times, long-accepted family groups have been divided into two or more distinct families, or separate families have been combined into a single one. Where possible such changes in our understanding of the relationships of fishes are reflected in these volumes. Where some contributors have taken a more conservative approach by awaiting more research and not adopting these changes, alternative taxonomies are noted (see also the introductory chapter on **Naming organisms and determining their relationships**).

For each species in the book, the literature pertinent to that species in the WIO is given: the original species description reference, synonyms for the region and other important taxonomic and biological references. For many commercially important species or fishes of interest to anglers there is additional information on life history, size and capture, and for some but not all species, their IUCN conservation status if Near Threatened, Vulnerable, Endangered or Critically Endangered (in the first instance, valid at the time of writing. See www.iucnredlist.org for current information. *Note:* we have not included the IUCN conservation status where species are of Least Concern or Data Deficient).

Most species are illustrated with photographs, drawings or paintings. Colour photographs and paintings are provided on plates for each volume. The coverage of each of the volumes of this book is outlined in the next chapter.

# **HOW TO USE THIS BOOK**

The *Coastal Fishes of the Western Indian Ocean* has family and genus accounts, keys, species descriptions, illustrations, paintings and photographs of most of the known coastal fish species from the Western Indian Ocean (WIO).

The book has been divided into five volumes. The first part of Volume 1 contains all introductory material, including chapters of general interest, plus a list of abbreviations, a comprehensive glossary and a bibliography. The second part contains the cartilaginous fishes (hagfishes, sharks, rays and chimaeras). Volume 2 contains the bony fishes (teleosts) – eels, herrings, flyingfishes, seahorses, scorpionfishes, and others. Volumes 3 to 5 contain the many families of perciform (perchlike) fishes, as well as the flatfishes, spikefishes, triplespines, triggerfishes, filefishes, boxfishes, pufferfishes, porcupinefishes, molas and the coelacanth.

The book has been designed to help identify the shallowwater fishes (to ~200 m) found in the area: along continental and insular localities, in tidepools, estuaries and mangroves, along rocky coastlines and sandy beaches, on inshore and offshore reefs, banks and seamounts, as well as pelagic species of interest to commercial and sport fishermen. The various keys to orders, families, genera and species are for species that occur in the WIO and are not necessarily applicable in other areas.\*

The maps printed on the inside covers (or separate PDF for digital version) show the areas covered in these volumes. Coordinates of place names used in the text can be accessed from a number of websites, such as www.geonames.org. The place names used in synonymies of species have been rendered in their currently accepted form, and not as they were originally used.

Each volume has an index to scientific names for the species described in the volume, as well as an index to English common names used for their families. Names that appear in synonymies are *not* included in the indexes, and can be found in *Eschmeyer's Catalog of Fishes* (see link at the end of this chapter). In addition, two full alphabetical lists of all family names (scientific and common) are provided with their corresponding volume number. These are included to assist the reader in identifying the correct volume for a particular family.

# **CLASSIFICATION**

All living organisms are grouped in a hierarchical system – their classification – which is determined by how we understand the way in which the various groups of organisms are related to each other. The chapter on taxonomy explains how organisms are classified and given their names.

# **IDENTIFYING FISHES**

To find more information about a particular fish species, you will need to identify the family to which the fish belongs. Scientific names of families end in -idae (e.g., Serranidae). If you know the family of the fish you want to identify, e.g., Serranidae (perchlets and seabasses), or Carcharhinidae (requiem sharks), use the index to go to the family account in the book.

If you do not know family, genus or species, decide if your fish is a cartilaginous fish - a shark, ray, hagfish or chimaera or a bony fish. If it is a cartilaginous fish (a chondrichthyan), you will find the Key to Families of Chondrichthyans in Volume 1. If it is a bony fish, you will find the Key to Orders of Bony Fishes in Volume 2. If this key takes you to the Order Perciformes (the perch-like fishes), you will find the Key to Perciform Families in Volumes 3 to 5. The keys to families will help find the family account and the related keys to genera and species of that family. Some families have several different 'typical' shapes, and the outline drawings of the shapes also help place a fish in a family. Once you have identified a likely family, turn to the family account. Compare your fish with the drawing, painting or photograph. A plate reference will direct you to the relevant plate(s) if there are colour photographs or paintings for the species. Photographs and paintings have been grouped together in families to make identification easier. If the colour or shapes of the species illustration do not match, read the species description to check for variation within the species; check the account for the genus for details not

\*Note: the drawings contained in the keys were mostly drawn for this book or sourced from Smiths' Sea Fishes, unless otherwise indicated.

duplicated in the species account. Distributions can confirm whether your fish is known from the area. However, bear in mind that the WIO distributions, as known at present, are often a result of sparse and uneven sampling. Many species not previously known from an area, or depths <200 m, have been brought to the attention of scientists by enthusiastic amateur ichthyologists.

The photographs of some species have been included without accompanying species diagnoses in the text. This is indicated by # after the caption and indicates that the species is likely to occur in the WIO but has not yet been confirmed, or is known from the area but a species account has not been included because of time constraints, or the species has not been described.

The families are placed in evolutionary (phylogenetic) order in the book. In other words, the oldest, supposedly most 'primitive', cartilaginous fishes (Class Chondrichthyes) are in Volume 1, with an account of their origins and evolution, followed by the more recently evolved or 'derived' bony fish families (Class Osteichthyes) in the other volumes. There is also an account of the origins and evolution of the bony fishes at the beginning of Volume 2.

The sequence in which the families are placed generally groups similar fishes together (e.g., all catsharks [Pentanchidae and Scyliorhinidae], gobies [Gobiidae], or jobfish and snappers [Lutjanidae]), which makes comparisons and identification easier. Within families, genera and species are arranged alphabetically by scientific name. A common English name has been given for most species; however, some species are known only from one or two specimens and have no common names; some well-known species may have several. The common names of many fishes can be accessed at www.fishbase.org and www.fishwisepro.com.

## **FAMILY ACCOUNTS**

Each family account starts with a brief description of the family characteristics and biology. The worldwide geographic distribution of the family is given, as is the estimated number of genera and species in the family.

# **GENUS ACCOUNTS**

Genus accounts give a short list of the characters of the species in that genus. *When certain characters, such as fin spine and ray counts are common to all species in the genus, they are usually not repeated in the species accounts.* The number of known species in the genus worldwide (at time of publication), as well as in the WIO, are given.

# **SPECIES ACCOUNTS**

Each species account starts with a scientific and (usually) a common name. The scientific name comprises the genus and the species epithet, followed by the author(s) of that species and the year in which the name was published and the locality where the holotype was found. The oldest name is usually the valid name (or senior synonym) and is the one that is used. More recent names are considered junior synonyms (see synonymy below). The author is the person (or persons) who identified that fish as a new species and gave it its scientific name and description of the species. An author's name in parentheses indicates that the genus to which the species is currently allocated is different from the one in which it was originally placed. Some species have yet to be given a scientific name and have sp. (the abbreviation of species) after the genus name. Some families and genera are noted as 'monotypic', meaning there is only a single genus with a single species in the family, or only a single species in the genus. A few species names in the text and captions to illustrations are also given as 'Genus cf. species', meaning the species is considered very similar to the name given, but the author is not absolutely sure.

Below the name (the heading) is the original name of the species, the author(s) and date of publication, and the type locality, i.e., where the fish was collected or caught. The type locality is usually given in parentheses; if the locality was not stated in the original description, but is known, it is written in square brackets. Next is a *synonymy* which lists the names used by various authors over time, including junior synonyms and misidentifications. A misidentification has the incorrectly used species name, and (in parentheses) *non* with the author of

the misidentified species to show that the name given is valid but incorrectly applied to that species. A ? (question mark) means that the identification is not certain, and 'in part' means that only part of the reference applies to the species. *In most instances, only the names used in the Western Indian Ocean have been listed.* This has been done in the interests of brevity, as some species have very lengthy synonymies.

The first paragraph of the species account is the diagnosis of the species and includes fin ray and body scale counts, as well as other measurements and features that aid identification. Diagnostic data given for the family or genus are not usually repeated in species accounts. Measurements of body features are given as a ratio of a larger measurement, such as body length or depth, head length, fork length, standard length or total length, or as a percentage of a different length.

Colour descriptions are usually given where known, particularly when there is sexual dichromatism (different colour patterns for females and males), which is common for many species. Descriptions are accompanied by an illustration or photograph of the fish (or both, where available) and these may also document changes in colouration and body shape from juveniles to adults. Some species accounts are therefore accompanied by more than one image. If known, the length for each fish illustrated is given in the caption. For a number of species, however, there are few photographs, and the best image available has been used. The lengths are mostly given to the nearest centimetre or millimetre. Localities, where known, are in parentheses. To make comparisons easier, some drawings and photographs have been reversed so that the fish faces left, a convention used around the world when illustrating fishes. The colour notes include the known maximum size of the species.

The distribution of a species is given as the localities in WIO where a species is known to occur, while for *outside WIO*, *usually only approximate distribution ranges are noted*.

Finally, some additional remarks are often included that provide other relevant information about the species that may be of interest or importance, e.g., information about fisheries, feeding, habitat, reproduction, questions about identity, and IUCN conservation status where appropriate.

Many worthy sources of information on fishes, including taxonomic information, are available on the internet and we've listed a number of useful URLs below.

#### **Eschmeyer's Catalog of Fishes:**

https://researcharchive.calacademy.org/research/ ichthyology/catalog/fishcatmain.asp

**FishBase**: www.fishbase.org – *global information system on fishes* 

**FishWise Professional**: www.fishwisepro.com – *relational database that can be interrogated* 

International Union for Conservation of Nature: www.iucn.org

John E. Randall's Fish Photos: http://pbs.bishopmuseum. org/images/JER/images.asp

South African Institute for Aquatic Biodiversity: www.saiab.ac.za

# NAMING ORGANISMS AND DETERMINING THEIR RELATIONSHIPS

## Wouter Holleman and Gavin Gouws

The system of binomial nomenclature by which all plants and animals are given a genus name and a species name, was established by Carl Linnaeus in the first half of the 18th century. The names are based on Latin and Greek and, for animals, are governed by a set of rules laid down in the *International Code of Zoological Nomenclature*. The 10th edition of Linnaeus's *Systema Naturae*, published in 1758, is considered the starting point for all zoological nomenclature.

Organisms are given scientific names for several reasons. The first is for consistency so that everyone knows which species, genus or family is being referred to. The second is that the names are a reflection of the assumed relationship of organisms. The species in a particular genus are assumed to be more closely related to each other than to other less similar species, which are then placed in a different genus. In Linnaeus's words: *"Characterem non constituero Genus, sed Genus Characterem*", which Darwin commented on as follows:

Such expressions as that famous one of Linnaeus ... that *the characters do not make the genus, but that the genus gives the characters*, seem to imply that something more is included in our classifications, than mere resemblance. I believe that something more is included; and that propinquity of descent – the only known cause of the similarity of organic beings – is the bond, hidden as it is by various degrees of modification, which is partially revealed to us by our classifications. (Darwin 1859)

These relationships – the "propinquity of descent" – are implicit in Darwin's theory of evolution, as expounded in *On the Origin of Species by Means of Natural Selection*, first published in 1859. In the book he established the notion that all animals (and plants) vary and that species evolve by the process of natural selection so that, in effect, all species are descended from a single, common ancestor. In fact, the only illustration to appear in his book is a small hypothetical 'tree' that shows the relationships between three taxa.

Over the years, taxonomy and systematics have incorporated increasingly more information and techniques, thereby contributing to a better understanding of what species are.

For many years animals and plants have been grouped together based on their morphological similarity. Species that look most similar were assumed to be most closely related: note the similarity between the various species of butterfly fishes, the Chaetodontidae (Volume 3), with their deep, compressed bodies, bristle-like teeth and eye-catching colours, often in combinations of yellow, black, white and silver. This way of classifying is called *gradistic* taxonomy. Very often, when the similarities were derived from a common ancestor, the taxa that were created in this way were reflections of the actual relationship between organisms.

In 1963, Robert R Sokal and Peter HA Sneath developed a classification system which grouped organisms into taxonomic units by statistical methods, in which certain characters or character states were given numerical values. These 'numerical taxonomists' aimed to create a taxonomy using numeric algorithms like cluster analysis, rather than using subjective evaluation of similarity. They divided the field into *phenetics* in which classifications are formed based on the patterns of overall similarities, and *cladistics* in which classifications are based on the branching patterns of the estimated evolutionary history of the taxa.

Although this was intended to be objective, in practice, the choice and implicit or explicit weighting of characteristics was too often influenced by the data available and by the research interests of the investigator. What was objective were the explicit steps that were used to create dendrograms and cladograms using numerical methods, rather than any subjective synthesis of the data.

At about the same time, a German entomologist, Willi Hennig, developed a different approach, one based on the notion that species evolved suites of characters that enabled them to adapt to a particular environment. Such adaptations – or 'specialisations' – were named 'apomorphies' by Hennig, and if they were shared by a group of taxa they were termed 'synapomorphies'. A Hennigian system of classifying organisms thus was based not merely on the similarities between taxa, but on whether those similarities were both derived and shared in order to reflect the actual evolutionary relationships between taxa. In other words, the relationships were reflections of genealogies.

To discover the genealogical relationships between taxa involved comparing selected characters (bones, muscles and ligaments, colour patterns, behaviour patterns and so on) in one group of organisms or taxa, and then comparing the characters with the same suite of characters in a similar taxon called the 'outgroup'. By this method one could then determine which characters were 'specialisations' shared by the organisms or taxa of the 'in-group', and which thus were 'synapomorphies'. These were distinguished from characters spread more widely, and beyond the group under consideration, which are 'plesiomorphic' characters, or characters that were the result of parallel evolution, where two unrelated taxa evolved similar traits. Such characters are called 'anologues', and 'homoplasies' when occurring in two or more otherwise closely related groups. So, for example, all butterfly fishes (Chaetodontidae) possess many peculiar, bristle-like teeth on the jaws, not found in any other fishes. They do, however, have compressed bodies and yellow and black colour patterns similar to many rabbitfishes (Siganidae), but rabbitfishes have two spines in their pelvic fins, one on either side of five central rays, with the innermost spine connected to the body by a membrane, a character not found in any other group of fishes.

Another example, not from the world of fishes, will further illustrate this. All birds share a particular specialisation (synapomorphy), in that they have feathers and wings for flight. However, not all birds fly - think of rheas, emus and ostriches. These birds are called ratites, and have secondarily lost the ability to fly, which is in itself a derived or apomorphic characteristic. Birds also all lay shelled eggs, an adaptation for living on dry land (fishes and amphibians lay their eggs in water to prevent the embryos drying out). But, lizards, snakes, crocodiles and tortoises also lay eggs, as did many dinosaurs we know of. Since the shelled egg is an apomorphic character that it is shared by such a large groups of seemingly unrelated animals, it suggests that birds, lizards, tortoises, crocodiles and dinosaurs are all related. And so, because various anatomical and molecular data show clearly that they are in fact related, they are grouped together in a large taxon called the Archosauria.

This way of classifying organisms is now referred to as 'cladistics' or 'phylogenetic systematics', a clade being a number of genealogically related taxa. A clade with a single, common ancestor is considered the correct reflection of the evolutionary relationships between its constituent taxa, and is called 'monophyletic'. Most taxonomists today apply cladistic methodology to their work. Taxonomists also began to look beyond mere morphological characters and include behaviour, physiology and various aspects of ecology in their suites of characters.

The 1960s saw the rise of the genetic sciences and the use of DNA analysis for a multitude of different purposes. Genetic information has always been an intrinsic part of marine fish systematics and taxonomy insofar as the characters used to define taxonomic units (whether species, genera, families or higher taxa) and the systematic arrangements of these are heritable and largely genetically determined. However, the development of techniques that allowed insights into the genetics of the organisms at the molecular level, either directly or by inference, has had a profound influence on ichthyology. The variety of techniques and markers for generating genetic data, and the range of applications of such information are myriad. This overview thus focuses only on the taxonomic and systematic applications, and those techniques that have been widely used in ichthyology.

Genetic data began to take hold in the 1960s, with the development of protein or allozyme electrophoresis. This involved placing a crude protein solution, extracted from particular tissues of the specimens of interest, in a gel matrix under an electric current. Protein molecules would migrate through the matrix, their mobility determined by the size, shape and charge of the molecules. Differences in the extent of movement of a specific protein, reflecting differences in the phenotype of that protein and amino acid variation among individuals, provided a rough indication of differences at the DNA level for the genes encoding these proteins. Individuals could then be genotyped and compared using the profiles of several proteins, and this data subjected to statistical analysis. However, the technique was crude; genetic diversity and divergence were often underestimated, and aspects of the genotyping were somewhat subjective. It also became increasingly difficult to apply to increasing taxonomic rank, as the homology of the specific proteins and alleles examined became difficulty to establish. Nonetheless, the technique was widely applied - molecular systematists commonly refer to the 1970s and 1980s as the "find 'em and grind 'em" era (in reference to the production of the protein extract). In ichthyology, this technique and the data generated were used extensively to examine diversity within and among species. It was a particularly powerful tool for testing species boundaries and for species delineation; any fixed differences (e.g., different sets of alleles) in the genotypes of these Mendelian-inherited markers among populations could be used to infer reproductive isolation among biological species. Hybridisation was also frequently studied, and the technique was invaluable for investigating population and fishery genetics.

While allozymes provided an indirect assessment of variation at the DNA-sequence level, restriction analyses allowed a closer look at the DNA molecule itself. In Restriction Fragment Length Polymorphisms (RFLPs), the DNA molecule is 'cut up' using endonuclease restriction enzymes. The resulting 'digestion profiles', sometimes referred to as 'fingerprints', are compared among samples. The presence of a particular fragment in one and its absence in another is as a result of differences in the DNA sequence at the restriction site, such that the enzyme was unable to cleave the DNA at that specific point in the DNA strand. This approach, used mostly to examine patterns of genetic variation and diversity within species, was short-lived and was superseded by DNA sequencing. However, the general principle is still applied in more modern and routinely used techniques, such as Amplified Fragment Length Polymorphisms (AFLPs).

The full power of genetic data was revealed following the invention of Sanger sequencing in 1977 and, particularly, Polymerase Chain Reactions (PCR) in 1985. Through these techniques it became possible to isolate, amplify, sequence, and directly visualise and analyse specific DNA sequences. This spawned a wealth of DNA-sequence-based studies, and the approach was firmly entrenched and widely used by the early to mid-1990s. Initially, these studies used mitochondrial DNA gene regions almost exclusively.

Mitochondrial DNA (mtDNA) is maternally inherited and is haploid; it gives only one perspective with respect to genetic structure, relationships and evolutionary history. However, the benefits of mtDNA are that the molecule is present in high numbers in the cells, and is technically simple to work with. Very quickly a range of primers and laboratory protocols were in place to amplify mtDNA fragments.

Technology developed, protocols were refined and competencies honed, and more studies began to incorporate more nuclear DNA markers, which are diploid markers and reflect the contributions of both parents in sexually reproducing species. They thus provide a more holistic picture of relationships. Moreover, computational advances and the incorporation of novel statistical approaches meant that larger datasets (with more markers and data, and/or wider taxonomic representation) could be analysed in less time and with more rigour and complexity.

Incorporating data from more and more gene regions meant that the final relationships revealed by the analyses of the various genes, each providing their own patterns of relationship of the gene itself (the gene tree), better approximated the true relationships and evolutionary history (the species tree) of the taxa under study. With this complexity, studies moved beyond just examining species boundaries and identifying new species, detecting hybridisation, and proposing and testing phylogenetic relationships and hypotheses (with obvious implications for taxonomy, systematic arrangements and character evolution), to investigating aspects of biogeography and dating evolutionary events and the divergence of taxa.

The taxonomic arrangements suggested by molecular or DNA data have not been without controversy. Some have been widely accepted, but others are still robustly debated among the morphological and molecular camps. Among the more controversial arrangements include the placement of the Scaridae (parrotfishes) within the Labridae (wrasses) rather than as an independent family; the inclusion of the fusiliers (Caesionidae) as a subfamily within the snappers (Lutjanidae); and the raising of the subfamily Epinephelinae (groupers, podges and soapfishes) from the larger group of serranoids (groupers, podges, soapfishes, sea basses, basslets and perchlets) as an independent family. The continuous state of flux of generic arrangements one sees in large groups, such as the Apogonidae, demonstrates the interaction between the molecular perspectives and the reappraisal of morphological characters. There is no doubt that many other taxa will experience similar rearrangements as DNA sequencing progresses.

A significant advancement, which has led to an exponential increase in the amount of DNA-sequence data available for marine fishes over the last decade, is DNA barcoding. Molecular systematists have long known that one can identify known species and detect new species using DNA-sequence data. However, an initiative was established for this particular application of DNA analysis, and was popularised by Paul Hebert and colleagues at the University of Guelph, Canada, under the banner of DNA barcoding. Their objective was to create a DNA-sequence (termed 'barcodes') reference library for all organisms on Earth. DNA barcodes included in the database (www.barcodinglife.org) would be derived from and underpinned by expertly identified, taxonomically validated specimens, lodged in reputable scientific/natural history collections, and accompanied by taxonomic information, repository and accession details, collection and geographic information, and an image. Standardised genetic markers were to be used. These would have sufficient information content to accurately and reliably discriminate species and be applicable across many different organismal groups. A fragment of the mitochondrial cytochrome c oxidase subunit I (COI) gene was chosen for most animals. For most marine fishes, this has proved to be effective, but additional markers have been considered for certain groups, such as the elasmobranchs, that show enigmatic mtDNA evolution. The reference database would enable reliable and unambiguous species identification for users without (or without access to) taxonomic expertise. Users would include ecologists, biologists, resource managers, biodiversity practitioners, conservationists, compliance and enforcement agencies, customs and excise officials, and the general public. Among the benefits, other than having rapid and accurate species identifications for unknown specimens, included assigning a species identity to different life-history stages or sexes, where one form might be unknown, to the forensic applications in being able to identify a species from a fragment or a piece of tissue. Barcoding was also suggested as a boon to solving the 'taxonomic impediment', the dwindling number of taxonomists and capacity in taxonomy in the face of immense undescribed and imperilled biodiversity. Here, barcoding could provide assistance to taxonomists, indicating new species requiring formal description or highlighting problematic groups.

DNA barcoding has not been without its detractors. Its validity as a scientific discipline (rather than a technical exercise), its assumptions, the depth of the analyses routinely

used, its accuracy and performance, and its contribution towards solving the 'taxonomic impediment' have all been questioned. Nonetheless, for marine fishes, it has been a valuable tool in terms of compiling inventories, in forensic applications, and in linking eggs and larvae to adult forms. The last decade has also seen a plethora of new species described, combining DNA barcoding, morphological analyses and traditional taxonomy.

Recently, new technologies have brought us Next-Generation Sequencing (NGS). NGS 'parallelises' DNA sequencing, such that several million different sequences (upwards of 50 million on new platforms) can be generated in single reactions, rather than the single sequence produced by Sanger sequencing. Continuous improvements are increasing the numbers and lengths of the fragments that can be produced. The technology and the applications are still relatively expensive, but the high-throughput nature and the amount of data that is obtained result in a low 'cost per sequence'. As such, it is now feasible to generate full genomes, by piecing together these fragments, for the samples and species of interest and to analyse relationships using these. Alternatively, a 'shotgun' approach will randomly generate sequences from the genomes of many taxa, and researchers can select markers or fragments of DNA that are common to the species or samples studied, or that show appropriate levels of variation, to address the question at hand. Examples of the two approaches are the recent publication (in the journal *Nature*) of the coelacanth genome, and the large phylogenies involving several thousand species, being constructed using Ultra-Conserved Elements, to examine the relationships, evolutionary history and taxonomic arrangements within the percomorphs. The systematic discipline of phylogenetics has been expanded to accommodate the use and analysis of such genomic data, as phylogenomics. The full genomic information also gives one access to markers that are under selection, and a variety of markers such as Single Nucleotide Polymorphisms (SNPs) and microsatellites for population genetic studies. Similarly, these techniques can also be applied to produce data relevant to studies of gene expression and epigenetics. Analysis of such large datasets is always a constraint but computing (especially cluster or 'super' computing), bioinformatics and data analysis pipelines are continually being developed to serve this advancing field. The growth in NGS capability has been astounding, and one can only speculate what will be available to ichthyological geneticists in the future.

Carl Linnaeus was born on 23 May 1707, in Stenbrohult in southern Sweden. His father was a Lutheran pastor and an avid gardener, and from early on, Carl developed a deep love of plants and a fascination for their names. He studied medicine at the University of Uppsala, but actually spent most of his time studying plants. In 1731, while still a student with little money, he made an expedition to Lapland to collect plants, and in 1734 he made another expedition to central Sweden. The following year he went to Holland to study further at the universities of Hardewijk and Leiden and in the same year he published the first edition of a slim little book on the classification of living organisms, his Systema Naturae.

Linnaeus returned to Sweden in 1738 where he practiced medicine and lectured at the University of Uppsala, becoming a professor at the university in 1741. While in Stockholm, he restored the university's botanical garden, arranging the plants according to his system of classification.

Linnaeus arranged for some of his students to go on trade and exploration voyages all over the world. Daniel Solander, perhaps his most famous student, was naturalist on Captain James Cook's first voyage around the world. Another student, Anders Sparrman, was botanist on Cook's second voyage. These students brought back to Europe the first plant collections from Australia and the South Pacific. Others brought back specimens from Japan, North America, Southeast Asia, Africa and the Middle East. As more and more plant and animal specimens were brought to Linnaeus, he continued to add to and revise his Systema Naturae, until it comprised many volumes.

In 1758 he bought an estate called Hammarby, near Uppsala, and there built a museum for all his collections. Three years later he was made a nobleman and became Carl von Linné. When Linnaeus died in 1778, his son Carl, became professor in his place. But Carl died five years later and Linnaeus's mother and sisters sold Linnaeus's library, manuscripts and collections to Sir James Edward Smith, who founded the Linnean Society in London to look after them.

#### GLOSSARY

**allele** – a specific and unique form or variant of a particular gene or genetic marker; alleles typically refer to pairs of genes found on each of the two homologous chromosomes of a diploid organism.

**allozyme** – particular form of a given protein, coded by a particular allele of the gene responsible for encoding for that protein.

**charge (of a molecule)** – molecules possess a net electrical charge, determined by the structure of the molecule (amino acids in the case of proteins), the atomic make-up of the molecule itself and the arrangement and nature of subatomic particles, under certain physical conditions (e.g., pH, chemical composition of the medium).

**cleave (DNA)** – the breaking of the DNA molecule/strand into two (or more) fragments by breaking the phosphate bonds that link the individual nucleotides in the chain together. This can be done with the use of enzymes, chemically, physically or by radiation.

**diploid** – having both chromosomes of the paired homologous chromosome set and, thus, both copies of each gene present in the cell nucleus; resulting from sexual reproduction.

**encoding (genes and proteins)** – the function and process whereby a particular gene produces a protein or enzyme; a gene responsible for a certain character, product, process, behaviour or phenotypic trait is said to *encode* for that feature.

**endonuclease restriction enzymes** – bacterial enzymes that cleave (break) DNA molecules wherever a particular sequence of nucleotides (the restriction site) is recognised.

**genotype** – the genetic make-up (profile) of the individual organism; the specific combination of genetic variants (e.g., a DNA sequence or a combination of alleles) for a genetic marker being studied or underpinning a character (phenotype) being considered.

**haploid** – having only one chromosome (of the normally paired homologous chromosome set) and, thus, one copy of each gene present in the cell nucleus or cell organelle.

homology - sharing the same evolutionary origins.

**Mendelian inheritance** – the rules of genetic inheritance in diploid organisms, as deduced by Gregor Mendel's seminal 'genetic' hypotheses based on breeding, cross-breeding and observing the phenotypes of garden peas. This work suggested that phenotypes are controlled by two factors (alleles in a genetic context), one obtained from each parent; that these factors assort or act independently of factors for other phenotypes during cell division (meiosis) and fertilisation; that these factors can be dominant or recessive with respect to influencing the phenotype, and that the phenotypic outcomes can be predicted numerically/statistically, based on the phenotypes present in the population and by tracing a genealogy.

**mitochondrial DNA** – an extranuclear (i.e., occurring outside the cell nucleus), double-stranded, circular DNA molecule found in the mitochondria of animal cells; the molecule is maternally inherited and haploid.

**percomorph** – fishes belonging to the Percomorpha, a large clade of bony fishes, mostly perciform, characterised by having bony fin spines, a ductless swimbladder and ctenoid scales.

**phenotype** – physical features of the organism or aspects of the organism's biochemistry, physiology or behaviour that can be observed and are thought to reflect variation in the genotype or the interaction of the genotype and the environment.

**population** – a group of organisms belonging to the same species that co-occur in a distinct area.

**protocols** – a set of procedures or a methodology to be followed in the laboratory.

taxon (pl. taxa) – a group of organisms such as a species, genus or a family that share a common ancestor.

# **ASPECTS OF THE BIOLOGY OF FISHES**

This brief introduction to some aspects of the biology of fishes serves as a background to the many species covered in these volumes, and has been included for two reasons. The first is that many of the topics covered, such as colour and aspects of reproduction and physiology, are important taxonomic tools used to identify different species. The second is to give some insight into the way fishes live in an environment very different to our own.

## SHAPE, SIZE, SWIMMING AND SPEED

Fishes exhibit a tremendous variety of shapes: compressed (flattened from side to side, e.g., butterflyfish and moonies); elongate (e.g., snoek and cutlassfish); robust and streamlined (e.g. tunas); depressed (flattened from back-to-belly e.g., monkfishes and stingrays); spherical (e.g., porcupinefishes); or box-shaped (e.g., many boxfishes). The boxfish, *Tetrosomus reipublicae*, is triangular in cross-section. The ocean sunfish, *Mola mola*, resembles a truncate disc or the front end of a fish that has been cut in two, hence the name 'head-fish' by which it is sometimes known (Figure 1).

The largest fish in the world is the whale shark, which attains a length of at least 12 m, and perhaps as much as 19 m. Like the blue whale, which grows considerably larger, the whale shark also feeds on plankton. The smallest fishes in the world are *Paedocypris progenetica* (7.9 mm SL) from a peat swamp in Indonesia, *Schindleria brevipinguis* (8.4 mm SL) from the Great Barrier Reef and Coral Sea, the pygmy goby *Pandaka pygmaea* (9.0 mm SL) from the Philippines and *Trimmatom nanus* (8.5 mm SL), a tiny goby known from the Maldives and Chagos Archipelago in the Western Indian Ocean (WIO) (Figure 2).

Most fishes swim by moving the caudal peduncle and caudal fin. In the fastest fishes (tunas and other scombrids), the body flexes hardly at all, and the caudal fin provides all the thrust. In eels and some other elongate fishes, the whole body flexes in an undulating wave that passes from head to tail, and this backward-moving wave drives the eel forward. The majority of fishes fall in between these two extremes, with peduncles more flexible than tunas, but less flexible than eels.

Scombrids have several anatomical adaptations that contribute to their remarkable swimming abilities. The stiff, deeply lunate caudal fin is well adapted for rapid movements and minimal drag. A speeding tuna can drive its caudal fin at the incredible rate of 10 beats/second. The massive body muscles are connected to tendons that run along the narrow peduncle (which acts as a pulley) to attach at the base of the caudal-fin rays. Horizontal keels on the peduncle strengthen it and are thought to reduce turbulence from the rapid lateral movements as the tail beats back and forth. The body is very streamlined, with grooves and recesses for the first dorsal and pectoral fins when they are folded back, to reduce drag. This sustained high-speed swimming of scombrid fishes requires a high metabolic rate, which means a high oxygen consumption. The gills of scombrids have a much greater surface area than the gills of non-scombrid fishes, and are therefore much more efficient at extracting oxygen from the water.



Figure 1 Mola mola, the ocean sunfish (Indonesia). © M Fraser



Figure 2 One of the smallest fishes, the goby *Trimmatom nanus*, 8.5 mm SL (Palau). © R Winterbottom, ROM

Unlike the typical cold-blooded fishes, the tunas and lamnid sharks (mako, porbeagle and white sharks) are able to maintain their inner muscular temperature several degrees above ambient water temperature. In tunas, the blood leaving the body muscles passes through a special network of capillaries that function as a countercurrent heat exchanger that transfers the metabolic heat generated by the muscles to the cold blood running into the muscles and digestive system in order to assimilate food faster, and thus become ready for the next meal. The swimming muscles are thus able to operate at the higher temperatures necessary for the higher metabolic rate required for sustained high-speed swimming. Of all fishes, the bluefin tunas have the most efficient heat exchangers, and they are the scombrids that penetrate farthest into cold waters, often occurring along the coasts of Norway and Alaska, from 40° to 50° S. The Atlantic bluefin tuna (Thunnus thynnus) can maintain a core body temperature of about 25 °C in water temperatures as low as 2.8 °C. Another discovery is that the opah or moonfish (Lampris guttatus), an inhabitant of cold waters, is entirely endothermic, and circulates warmed blood throughout its entire body (Wegner et al. 2015).

There are other fast-swimming fishes besides the scombrids. Kingfishes (Carangidae), dolphinfishes (Coryphaenidae), marlins and sailfishes (Istiophoridae) and the lamnid sharks (Lamnidae) all have a stiff, lunate caudal fin and narrow peduncle. This shape of caudal fin is the most efficient for fast cruising. A broad truncate or rounded caudal fin, as in groupers (Epinephelidae) or kob (Sciaenidae), is better for the sudden fast acceleration required by a lurking predator.

Many fishes use their pectoral fins for propulsion. Wrasses, parrotfishes, damselfishes, and the oceanic opahs (*Lampris*) cruise by flapping their pectoral fins like wings. The pectoral fins of opahs are long and stiff, and the muscles that move them are enormous, so that they 'fly' through the water, much like penguins flapping their wings. Although wrasses and parrotfishes depend on their pectoral fins for cruising, they also use their caudal fin when they are in a hurry. Stingrays, manta rays and most other batoids use undulatory waves of their broad pectoral fins for propulsion and appear to fly effortlessly through the water. Electric rays, guitarfishes and sawfishes use their tail and caudal fin for swimming.

Ocean sunfishes (Molidae) have stiff paddle-like dorsal and anal fins which are their primary means of propulsion. In pufferfishes (Tetraodontidae), porcupinefishes (Diodontidae) and boxfishes (Ostraciidae), the short-based dorsal and anal fins as well as the pectoral fins are used for slow swimming. But, unlike the sunfishes which have no caudal fin, these fishes use the caudal fin when they want to swim faster. Fishes with long-based dorsal and anal fins, like triggerfishes and filefishes, use these fins for slow swimming by means of undulatory waves passed from the front to the rear of the fin.

Accurate measurements of the speed of the fastest fishes are difficult to make. The cruising speed of scombrids is in

the order of 2–10 km/h, but their burst swimming speed, which they use when chasing prey or fleeing from predators, is much faster. A wahoo (*Acanthocybium*), reputedly the fastest swimming fish, has been timed at 77 km/h for short bursts of speed, and a yellowfin tuna achieved 74 km/h for a 5-second period (Walters & Fierstine 1964). The billfishes (marlin, sailfish, swordfish, etc.) are also very fast swimmers, and could rival the tunas at their top speeds.

# **FISH MIGRATION**

Migrations are regular mass movements of animals from one place to another, usually for purposes of breeding or feeding. Many fishes undertake vertical migrations of several hundred metres every evening in order to feed on the abundant plankton in shallower, epipelagic waters. By feeding in surface waters at night, they are safe from birds and other predators that hunt visually. At dawn they return to depths of 100–1 000 m.

The elf or bluefish (*Pomatomus saltatrix*) migrates from KwaZulu-Natal to the southern Cape coast of South Africa during the late summer months to feed, and returns to warmer northern waters to breed from September to December. This species also occurs along the Atlantic coast of the USA and on the south coast of Australia where it is well known as a migratory fish.

Freshwater eels (*Anguilla* spp.) that live in the rivers of the south and east coasts of Africa and Madagascar must return to the open ocean to spawn. The transparent larvae (leptocephali) drift south in the Mozambique current from somewhere near Madagascar. When they enter freshwater, they turn into the wormlike little elvers that ascend the rivers, even wriggling up dam walls. Females go further up rivers than males, and the eels stay and grow in freshwater for about 14 years. When they are ready to return to the sea to breed, they stop eating, their eyes enlarge, and their dark skin becomes silvery. They migrate to off the east coast of Madagascar to spawn, but once they leave the rivers they are never seen again.

Marine fishes that live in freshwater and return to the sea to spawn are called catadromous. Freshwater eels are *obligate* catadromous species, in that they *have* to live their juvenile and subadult life in freshwater. On the other hand, the freshwater mullet (*Myxus capensis*) is a *facultative* catadromous species that spends its juvenile and subadult life either in estuaries or in their adjacent rivers, but returns to the sea to spawn. If an estuary does not provide access to freshwaters, it does not affect the life cycle of the mullet.

The annual migration of the pilchard (*Sardinops ocellatus*) to KwaZulu-Natal waters, known as the 'Sardine Run', is probably a feeding migration. These schools of pilchard appear to be following the abundant plankton which occur in the countercurrent flowing northward along the South African east coast in winter.



Figure 3 Larvae of various fish species. A Mola mola, B Dactylopterus sp., C a species of Holocentridae, D Naso sp., E. Forcipiger sp., at 3.3 and 5.2 mm, F Poecilopsetta sp., at 9.2 and 26.5 mm (two different spp.; note the migration of the eye). © A–D: GD Johnson; E–F: JM Leis & S Bullock

# LARVAL FISH DISPERSAL

Most of the coastal fish species found in the WIO occur on or in association with reefs of some kind. The vast majority of these species have complex life histories that include pelagic larvae that spend a few days to a few weeks in the open ocean before settling into a 'reef existence'. When adult, these species are closely associated to a particular part of the reef – they are site-attached.

It was long assumed that larval fishes were passively dispersed by ocean currents, often over very large distances, which accounted for the way various populations were connected. Larvae were referred to as planktonic and were assumed to form part of the passively drifting biomass of the sea, and they could settle anywhere the ocean current took them. Reef fish populations were thus seen as 'open' as there was very little self-recruitment into local populations.

This traditional view of larval dispersal was supported by the fact that the larvae are small compared to the distances current could disperse them, and would generally not survive the distances. It was also thought that larvae lacked any kind of ability that could influence their dispersal. These assumptions were, however, based on the larval dispersal patterns for groups of fishes found in temperate waters, none of which are associated with tropical reefs.

In the 1990s these ideas were challenged and subsequent research showed that larvae of tropical reef fishes are not passively dispersed. This was not to say that the larvae of many reef fishes were not dispersed over very long distances, which is why species are found at oceanic islands separated by considerable stretches of deep ocean. There is clear evidence that long-distance dispersal does take place over 100s to 1000s of kilometres, but most fish larvae find a reef home much closer to their natal reef. This was first shown by studies on fish larvae collected by ship-towed nets that found the larvae of different reef fishes, originating on the same reef, had different distributions (see Leis 2015 and references therein). Over time, laboratory and field research has shown that the tiny larvae are strong swimmers and are able to orient themselves in a featureless ocean. Further, as they develop morphologically, their swimming and orientation abilities develop and improve. These abilities also differ among species. Orientation abilities include a sense of sight which allow them to pick up celestial clues, the ability to hear reefs from several kilometres away, and a sense of smell that helps them find an appropriate place to settle (see Leis 2015 and references therein).

# FOOD AND FEEDING

Fishes exploit a wide range of foods, from detritus and microand macro-plankton, to algae, a variety of invertebrates and vertebrates, including fishes, marine mammals, birds, turtles and sea snakes. Some fishes concentrate on invertebrate prey – tunicates, jellyfish, live coral, echinoderms, crustaceans, molluscs, worms and sponges – while others feed mainly on fishes or cephalopods. Most fishes, however, are essentially opportunists and will eat whatever is plentiful and easy to catch.

The teeth of a fish usually reveal much about its diet. Large canines are typical of predators that feed on fishes, mammals, birds or cephalopods; herbivores use incisors; and planktivores have small, villiform teeth or none at all. Molars indicate a diet of hard-shelled prey – crabs, gastropods or bivalves.

Herbivores such as surgeonfishes, angelfishes, some parrotfishes and some damselfishes feed on algae, seagrass or detritus. They are active by day and are usually seen near the bottom where they spend most of their time grazing. They often have small mouths with small incisors and long intestines to cope with the long digestive times demanded by such a diet. Planktivores are grouped into filter-feeders and plankton pickers. Bony fish filter-feeders such as piIchards, sardines and anchovies, have many, long gill rakers projecting between the gill arches. The mouth is moderate in size, and teeth minute or absent. The whale shark and manta rays are filter-feeders with a grid-like structure between the gill arches and, like bony fish filter-feeders, swim through concentrations of plankton with their mouths open. As the water current passes through the gills, minute planktonic animals (copepods, amphipods, mysids, arrow worms, larval fish, crustaceans, molluscs and echinoderms) are filtered out of the water and passed into the gullet. Pelagic bony fish filter-feeders are active in the open water during the day, making them vulnerable to predators. This accounts for their schooling behaviour, which makes it more difficult for predators to target individual fish.

Plankton pickers are either obligate pickers or facultative planktivores. Obligate pickers, such as fusiliers (Caesionidae), goldies (*Pseudanthias* spp.) and aggregating damselfish (*Chromis* spp.), occur in groups in midwater above the reef. They are active by day and have short snouts and large eyes with good binocular vision for targeting larger zooplankton, such as amphipods, mysids, fish larvae and salps that they pick out of the plankton. Like filter-feeding planktivores, they have many long gill rakers, a more or less protrusile upper jaw, and minute or no teeth. They also tend to live close to reefs which provide refuge from predators. Facultative planktivores include various omnivores, such as galjoen (*Dichistius* spp.), some damselfishes, chubs and batfishes (*Platax* spp.), which feed on a variety of plant and animal food, but will concentrate on plankton when it is abundant.

Many damselfishes, such as *Plectroglyphidodon* and *Stegastes* spp., are territorial herbivores that defend and feed on specific patches of benthic algae. These algae are covered with a layer of detritus, and a variety of small invertebrates as well as bacteria, fungi, diatoms and dinoflagellates that live on the algae. The damselfishes carefully weed out undesirable algae, which may be less nutritious or less conducive to the growth of the epilithic algal matrix.

Most fishes change their diet with growth. Juvenile groupers eat mainly crustaceans, but many adult groupers include a large portion of fish and cephalopods in their diet.

## AGE AND GROWTH

The growth of fishes, like that of other poikilothermic animals, is dependent on the temperature of their environment. The

availability of food is the other main factor that influences fish growth. Determining the age of fishes that live in temperate waters is relatively easy, because they grow faster in summer than in winter. This annual variation in growth is recorded in the form of growth marks (called annuli) in the otoliths (the ear bones), bones of the skeleton and scales of a fish, much like the growth rings of a tree. Fishes that live in tropical waters or in the deep sea, where water temperature is pretty constant all year round, do not have such marked growth rings and are often difficult to age (Figure 4).

The otoliths of fishes are often used to determine how old they are, because their annuli are more distinct than those of other bones or scales. In long-lived species, the accurate determination of age for a fish older than 10 or 20 years becomes complicated, because mature fishes grow more slowly than juveniles. After maturity the annuli become more crowded together and difficult to count. For older fishes it may be necessary to weigh or section the otolith to determine the growth increments. The ratio of radioactive isotopes of lead and radium in the sectioned otoliths of rockfish (a North Pacific scorpaenid fish) has shown that this cold-water species may attain an age of 80 years, despite growing to only 46 cm.

Larger fish species generally live longer than smaller ones, provided that the large species also grow slowly. The dolphinfish (Coryphaena hippurus), which attains a length of 2 m, is an exceptionally fast-growing species; it is mature at an age of one year (at 55 cm FL), and probably does not live more than four years. The enormous tunas and billfishes are also very fast growing and apparently do not live much past the age of 15. Tagging studies of sharks, together with otolith data and bomb radiocarbon dating, have given us ages for several species: white sharks more than 40 years; sandbar sharks 30 years and more; and spiny dogfish can live longer than 45 years. Some deep-sea bony fishes live very long - the orange roughy lives to ~150 years, maturing at 30+ years of age. It is likely that some deep-sea sharks are also long-lived. At the other end of the scale, several small gobies of the genus Trimma pack an entire life-cycle into only a few months: Trimma nasa lives for only about 87 days, spending some 35 days in the plankton (Winterbottom & Southcott 2008). Coelacanths are known to live to 100 years.



Figure 4 A Electron micrograph of otolith of Argyrosomus japonicus, B horizontal section of otolith, C section showing growth rings of 18-year-old fish [see also Figure 9]. 
(a) A: MJ Smale; B, C: A Childs, RU-DIFS

# SENSES AND PHYSIOLOGY

## Vision

For most fishes, as for almost all other animals, vision is important for finding food, avoiding predators, and identifying potential mates.

Fishes show a tremendous diversity in the development of their eyes. The major difference between a fish's eye and that of non-aquatic vertebrates is that the lens is spherical, whereas most vertebrates have a 'lens-shaped' lens, i.e., oval in cross-section. The human eye and the eyes of most other vertebrates are focused by the initial refraction of light rays by the cornea and by muscles that flatten an elastic lens. The cornea of the eye of most fishes is optically non-functional, as it has the same refractive index as water, except for fishes like mudskippers (Periophthalmus spp.) that use their eyes out of water. Focusing is done by moving the lens backwards and forwards in the eye. Fishes are generally long-sighted, especially in their lateral field of view, but their sharpest vision is in the forward field of view, because the back part of the retina has the greatest density of visual cells. As in many other vertebrates, the eyes of fishes have two types of sensory cells, 'rods' and 'cones'. Rods are most sensitive to low light, while cones give very sharp vision, and have the ability to distinguish colours.

In well-lit epipelagic and nearshore waters, fishes have a retina that is rich in cone cells. Fishes living at depths of 500–1 000 m, where light levels are low, usually have large eyes with mostly rods in the retina. In the deepest parts of the sea (below ~2 000 m), the absence of sunlight and rarity of bioluminescent fishes renders vision largely obsolete, and many fishes have vestigial eyes. Deep reef fishes such as the coelacanth and sharks have eyes with a layer at the back (the *tapetum lucidum*) that shines like a cat's eye at night and reflects light back through the retina.

The amazing array of colours and patterns of the many different fishes on a coral reef have evolved for many ecological reasons. Conspicuous colour patterns have two functions. The first is interspecific communication such as the possession and defence of territory and in recognising individuals of the opposite sex. This is particularly true for those species where males and females have strikingly different colour patterns, as in the sexually dimorphic parrotfishes and wrasses. The second is aposematic colouration, used to advertise that the fish is inedible, as with the soapfishes. Fishes are otherwise cryptically coloured or camouflaged to avoid predation (see section **Mimicry and protective resemblance in WIO fishes** in this chapter).

Seeing colours under water is, however, different to seeing colours above the water: water absorbs long-wavelength light on the red end of the spectrum (i.e., wavelengths of >600 nm), so that almost all red-orange light has been absorbed below 10 m. The red pigments on fishes at these depths appear dark grey. In recent years it has been discovered that more than 180, mostly small, species of fishes can generate red fluorescent light, often in species-specific patterns (Michiels *et al.* 2008; Gerlach *et al.* 2014; Meadows *et al.* 2014) (Figure 5).



Figure 5 Red fluorescence in six species of reef fishes from the Red Sea: Corythoichthys flavofasciatus, C. schultzi, Enneapterygius pusillus, E. destai, E. abeli and Helcogramma steinitzi. © NK Michiels, University of Tübingen



**Figure 6** Colours and ultra-violet facial patterns of four individuals of two species of damselfish: *Pomacentrus amboinensis* (above), *P. moluccensis* (below). © UE Siebeck, UQ



Figure 7 Periophthalmus takita (Australia). © T Takita

Fluorescent patterns usually include the eye ring and parts of the head and chest. Emission of red fluorescence by fish, a colour absent from the background, may function as a means of communication or to attract conspecifics. Fluorescent light emission from the eyes in particular might assist in detection of cryptic prey (Meadows *et al.* 2014). Red fluorescence has been found to be associated with guanine crystals made by iridophores, well-known for producing iridescence in many bony fishes.

Reflective red pigmentation is widespread amongst reef fishes, and appears grey or black below around 10 m, allowing a fish to blend in with its background.

Another area where our knowledge of colour vision in fishes is poor, is their ability to detect ultraviolet light (UV). It is thought that UV signals must be bold, as they are scattered in air and water, and because, if present, there are few UVsensitive cones in the eye. Recently researchers have discovered that two similar species of damselfishes use ultraviolet light *patterns* for face recognition (Siebeck *et al.* 2010) (Figure 6). This has led to the hypothesis that some fishes use UV as a secret channel for communication that is hidden from potential predators (see section **Colouration and bioluminescence**).

Fishes with eyes on the sides of their head typically have a wide field of view. Many species can also rotate their eyes independently, like a chameleon. Mudskippers (*Periophthalmus* spp.) (Figure 7) spend most of their time out of the water climbing about the roots of mangrove trees, and can see quite well in air. They can retract their protruding, periscope-like eyes and roll them around in the eye socket to keep them moist, erecting them again to scan the area for predators or possible prey. Soles are flattened and lie on their sides on the bottom, so well-camouflaged that they are almost invisible. They do this by being able to match the colour of the upper (or eyedside) of the body to the colour of the substrate. If a sole is blinded it loses the ability to blend with its surroundings (see section **Mimicry and protective resemblance in WIO fishes** in this chapter).

## Hearing and lateral-line sense

The density of water is about 1 000 times greater than that of air, and water is thus much less compressible than air. This greater density and resistance to compression greatly affect the production and transmission of sound. In air, sound travels relatively slowly, as a periodic form of compression waves (Figure 8). In water, sound travels about five times faster, by particle displacement as well as by compression waves. The particle displacement form of sound energy is strongest close to the sound source, and is termed the near-field effect; compression waves extend farther from the sound source and are thus known as the far-field effect.



Figure 8 Sound waves travel in air as compression waves (above) and in water by particle displacement (below).

Fishes detect sounds with their lateral-line receptors and their ears. The sensory organs (called 'hair cells' or 'neuromasts') of the lateral line and the ears are very similar in structure. Lateral-line neuromasts are mechanoreceptors which are more-or-less directly exposed to the water surrounding the fish, and are especially sensitive to the near-field form of sound energy. Experiments have shown that lateral-line receptors are sensitive only to low-frequency sounds (10–200 Hz); the movements of fishes or other animals through the water primarily produce such low-frequency sound waves. Lateralline neuromasts are usually found in small, perforated tubes running just under the skin of the head and/or along the side of the body (hence the 'lateral line').

The ability to detect the motion of other animals is useful to both predators and prey, especially at night or in deep sea where sunlight is non-existent. The greatly elongated lateral line of some midwater fishes, such as the frostfishes (Trichiuridae) and ribbonfishes (Trachipteridae), is very efficient at detecting any movements in their vicinity. Although the lateral line is absent from the body of most clupeid fishes (herrings, sardines and pilchards), the well-developed lateralis canals of the head are important for schooling behaviour by providing information on fish movements in their immediate vicinity.

The ears of fishes are convoluted, fluid-filled organs, called labyrinths, enclosed in bony chambers (otic capsules) on either side of the skull. The upper part of each labyrinth consists of three curved membranous tubes (semicircular canals) that join a central chamber called the utriculus. Two of the semicircular canals lie in vertical planes approximately at right angles to each other; the third canal is located in the horizontal plane. Movements of the head result in displacements of the fluid (endolymph) in the semicircular canals and utriculus, and the fluid displacements deform the sensory hair cells on the inner wall of the labyrinth. Stimulation of these hair cells then sends nervous impulses to the brain, to be read as information on the movement and orientation of the fish. A fish's perception of gravity appears to be primarily a function of the utriculus. The lower part of the labyrinth has two adjoining chambers, the sacculus and the lagena, each with a dense calcareous otolith, or 'ear bone'. The utriculus (which also contains an otolith) is usually joined to the sacculus (Figure 9).

Fishes are sensitive to sounds of a wide range of frequencies. The hair cells, located inside the otic capsules, are primarily sensitive to far-field sound energy. This pressure wave is either transmitted directly through the tissues of the skull to the ear, or, in the case of fishes with a connection between the swimbladder and the ears, the sound wave induces vibrations of the gas-filled swimbladder, and these vibrations are conveyed directly to the ears. Fishes with a connection between the swimbladder and the ears are especially sensitive to highfrequency sounds (up to 180 kHz). At the labyrinth, the sound wave vibrations are transmitted to the endolymph and otoliths,



Figure 9 Labyrinth of a bony fish. Source: CFSA

and the vibrations of the otoliths stimulate the hair cells with which they are in contact.

## Touch

Although fishes apparently lack the specialised touch receptors of birds and mammals, they are able to register touch sensations by means of a network of free nerve endings in the skin. The barbels of catfishes and goatfishes are equipped with taste buds as well as many free nerve endings, and they probably function as taste and touch receptors when the fish is foraging for prey. The enlarged free pectoral-fin rays of gurnards, fingerfins and threadfins probably serve the same functions.

# Smell

A sense of smell is well developed in most fishes. The olfactory organs are round to oval plates of folded sensory epithelium located in nasal capsules on either side of the snout. Sharks (and most other elasmobranchs) have a keen sense of smell, as indicated by their well-developed olfactory organs and the large area of their brain that is concerned with olfaction. It is well known that sharks will approach a bait from downstream, following a scent trail to its source.

# Taste

The taste receptors of fishes are not restricted to the mouth; they are also found on the lips, barbels and certain modified fin rays, and even scattered over the head and body (see above paragraph on the sense of touch). Sharks are greatly repelled by the taste of the Moses sole (*Pardachirus marmoratus*) which secretes a milky toxin from pores at the base of its dorsal- and anal-fin rays. The soapfishes, the Grammistinae, also secrete a toxin from glands in their skin. The name 'soapfishes' alludes to the thick slimy mucus that covers the skin in these fishes. If a predator tries to eat a soapfish, the bitter-tasting mucus will cause the predator to spit out the fish, which is usually not seriously injured. Predators soon learn to recognise and avoid soapfishes.

# Electrical detection

Sharks, rays and some bony fishes, such as the marine catfishes of the genus *Plotosus*, have electroreceptors sensitive to the minute electric fields generated by all living animals. These receptors are called ampullary organs, or, in the case of elasmobranchs, ampullae of Lorenzini, after the man who first described them, and are located mainly on the front of the head in sharks. Each ampulla is a slender, flask-shaped organ with a long canal leading to a pore at the surface of the skin. Ampullary organs can detect minute electric fields caused by breathing prey buried in the substrate, and experiments have confirmed that benthic sharks and rays use this electrosensory ability to find their food.

We also know that fishes use their electroreceptors to detect the electric fields produced as the fish swims through the magnetic field of the Earth, and this sensory input might be used in navigation. A white shark, tagged at Gansbaai near Cape Town on 7 November 2003, was recorded 11 000 km away, off the northwest coast of Australia on 28 February the next year from a pop-up tag, and was then photographed off Cape Town, six months later, on 21 August 2004 (Bonfil *et al.* 2005).

## Pain

Do fishes feel pain? This a much-debated question and the subject of considerable research. There are many tales of fishermen who have hooked a fish, let it go, only to hook it a short while later. This has led to the notion that a fish does not feel the pain of the hook in its mouth. Sharks seem to be indifferent to wounds, and thus presumably feel little or no pain.

This apparent inability to feel pain may have to do with the number and kind of receptors in the skin. One group of researchers (Sneddon *et al.* 2003) found nociceptors on the face and snout of a salmon. These are sensory receptors that respond to potentially damaging stimuli, such as heat or chemicals, by sending nerve signals to the spinal cord, which then triggers a reflex reaction. In a review article, Rose *et al.* (2014) record that the typical human cutaneous nerve cell contains 83% C-type trauma receptors (those that are sensitive to extreme pain), whereas sharks have none. The debate is not settled and it is likely that fishes sense 'pain' differently from other vertebrates. Another aspect of this debate is whether fishes are *sentient* or not: do they 'feel' in any way? In an extensive review paper, Key (2015) presents evidence that while fishes react to noxious stimuli, that drugs do modify their behaviour, and they do seem to react to physiological stress, fishes do not have the neural receptors or parts of the brain which birds and mammals have evolved that enable them to feel pain in the way that we understand it. Fishes may thus sense pain differently from other vertebrates, but the current state of knowledge does not explain how fishes sense pain.

# PHYSIOLOGY

# Respiration

The gills of fishes are equivalent to the lungs of other vertebrates. The bright red colour of the gills of a live fish indicates the abundant blood supply. Gas exchange take place at the thin walls of the gill filaments. Each gill filament has numerous lamellae, where the blood is separated from the water flowing through the gills by a very thin membrane. Oxygen diffuses from the water into the blood, and carbon dioxide from the blood into the water across this membrane. A further adaptation that facilitates oxygen uptake by the gills is a *countercurrent* system, where the flow of blood in the lamellae is opposite to the flow of water across the lamellae (Figure 10).

Fast-swimming fishes like scombrids have a high metabolic rate and much higher oxygen consumption, compared with more sluggish fishes like catsharks or anglerfishes. The gills of the more active fishes have become adapted for more efficient uptake of oxygen: the lamellar walls are thinner, with many more lamellae, providing a much greater surface area for diffusion of oxygen and carbon dioxide.

The water current for gill ventilation is provided either by a 'branchial pump' or by simple 'ram ventilation', where the fish simply swims with its mouth open, and the water flow through the mouth and gills is accomplished by the movement of the fish through the water. Ram ventilation is used by fastswimming fishes (such as tunas), and by most plankton feeders (such as whale sharks), which swim about with the mouth







Figure 11 A whale shark, Rhincodon typus. © M Fraser

open to capture plankton. A branchial pump is used by less active fishes to force water through the gills. By alternately expanding and contracting the mouth and gill chambers, water is sucked into the mouth and pushed through the gills. There is also a flap of skin on the roof of the mouth that acts as a valve so that when the mouth is closed the water does not flow back out of it. This 'push-pull' way of breathing can be seen by watching fishes in an aquarium as they expand and contract their gill covers.

Some fishes have developed aerial respiration as a means for breathing out of the water, for a variety of reasons. Air breathing is best developed in freshwater lungfishes, which have lungs as well as gills. Freshwater eels (Anguilla spp., which spawn at sea; see Volume 2) normally take up about 10% of their oxygen through the skin, but if the eel is out of the water when it migrates from one water body to another, about 66% of their oxygen is taken in through the skin and the rest is extracted through the gills. A few marine fishes also use aerial respiration to supplement oxygen obtained from water. Mudskippers (Periophthalmus spp.), as mentioned previously, live out of the water for long periods during low tides. Here they have territories, with a burrow and often with low 'walls', which they will defend. When out of the water they acquire oxygen in two ways, in the same way as Anguilla spp. eels, through the skin, and by holding water in the gill chamber and quickly rocking the head from side to side to moisten the gills to absorb oxygen from the gill chamber (Jaafar & Murphy 2017).

Some species of mullet (Mugilidae) frequently jump out of the water. In warm stagnant water, which contains little dissolved oxygen, mullet jump more frequently than they do in well-oxygenated habitats. They apparently jump out of the water to take in air, which is then kept in bubbles in a chamber above the gills. These air bubbles provide an additional source of oxygen where oxygen is scarce.

# Osmoregulation

This is the process by which the salt concentration of blood and other tissues is regulated within the limits necessary to sustain life. When one has two liquids on either side of a very thin membrane, water (or any other liquid) will flow from the side with lower concentration of dissolved substances to the side with higher concentrations, so as to equalise the concentration of these substance on either side of the membrane.

In marine bony fishes, the salt concentration of the blood and tissues is lower than that of sea water, and they thus lose water by diffusion through the gills and skin. To make up for this loss of water, bony fishes take in sea water and absorb the water (along with salts which they do not need) through the gut. The unwanted salt ions are then excreted via the gills, urine and faeces.

The salt content of cartilaginous fishes (Chondrichthyes) is only slightly higher than that of teleosts. However, because of the high concentration of urea in the blood, there is no tendency to lose water through the gills or skin. Cartilaginous fishes (and coelacanths, which also use urea to osmoregulate) are somehow able to tolerate concentrations of urea in their blood that would be fatal to most other animals. The influx of salt ions is countered by excretion of the ions via the gills, urine and a rectal gland. Rectal glands appear to be unique to coelacanths and chondrichthyans, and excrete most of the Na<sup>+</sup>Cl<sup>-</sup> (sodium and chlorine ions).

Euryhaline fishes, such as the bull shark (*Carcharhinus leucus*), are able to tolerate wide fluctuations in salinity. This shark is often found in rivers; juveniles are known from Malawi and the Kruger National Park in South Africa. Estuarine fish fauna comprises euryhaline species like moonies (*Monodactylus*), mullet (Mugilidae) and kob (*Argyrosomus*). In hagfishes, the salt concentration of the blood is similar to that of sea water, and hagfishes cannot tolerate even half-strength sea water.

## Temperature

With a few notable exceptions, fishes are cold-blooded (poikilothermic or ectothermic) animals. This means that their body temperature is dependent on the temperature of the surrounding water. Except for certain oceanic fishes that often move through the thermocline, fishes cannot tolerate rapid changes of more than a few degrees. When an east wind blows along South Africa's southern coast, or when the northeast monsoon blows onto the coast of Somalia, it causes an upwelling of cold bottom water; this influx of cold water in the nearshore environment often kills fishes or makes them very lethargic.

Some large, fast-swimming fishes (tunas, marlins and lamnid sharks) are able to maintain their body temperatures above ambient water temperature and at a fairly constant level. See section **Shape**, **size**, **swimming and speed** for more about these warm-bodied — or endothermic (homeothermic) — fishes.

# Swimbladder, buoyancy and sound production

Most fishes have a gas-filled swimbladder in the upper part of the abdominal cavity. In some primitive fishes, such as lungfishes and the freshwater bowfins of North America, the swimbladder is connected to the oesophagus. A swimbladder's main function is to provide bouyancy.

In the spiny-rayed fishes the swimbladder has no connection with the gut, and any change in the amount of gas in the swimbladder is a slow process of secretion or resorption. If the fish is pulled rapidly to the surface from a depth of 20 m or more, the amount of gas in the swimbladder cannot be adjusted to the sudden decrease in external pressure (the pressure in the water changes by one atmosphere for every 10 m of depth). The result is a huge expansion of the gas and thus of the swimbladder, so that it may rupture or become pushed into the mouth. Anglers know that if they hook a large grouper in deep water and can bring it halfway up to the surface, the battle is won, as the expanding gas in the swimbladder makes it very difficult for the fish to swim back down to the bottom.

To get around the problem of rapid gas expansion with sudden changes in depth, some fishes have either replaced the gas-filled swimbladder with a partly or wholly fat-filled swimbladder, or they have lost their swimbladder completely. The swimbladder of the coelacanth (*Latimeria*) is completely filled with lipid (mainly low-density wax esters), and the body muscles are also rich in wax esters. The oilfish (*Ruvettus*) is notorious for the purgative properties of its flesh, the result of an abundance of wax esters in the body muscles. *Latimeria* and *Ruvettus* have thus evolved a similar solution to the problem of buoyancy: these two quite unrelated species occupy the same habitat at the Comoro Islands, and make extensive vertical migrations — and hence depth changes — in their search for the same prey.

Sharks (and other chondrichthyans) have no swimbladder, but also produce large amounts of lipid to provide buoyancy. This lipid is stored in the liver as oil (mainly squalene, a very low-density oil that incidentally also makes an excellent lubricant for fine machinery). The livers of some sharks are enormous, up to 25 per cent of their total weight.

The swimbladder is also used by some fishes to produce or amplify sounds. By contracting muscles attached to a gasfilled swimbladder, vibrations are produced that can be heard as growls, grunts, hoots and drumming noises. The muscles are either wholly attached to the swimbladder (intrinsic muscles) or attached by one end to the swimbladder and by the other to the skull, vertebral column or ribs (extrinsic muscles). These noises may be used in courtship and the defence of territory. Toadfishes (Batrachoididae) have a welldeveloped swimbladder with large intrinsic muscles. The cave bass (*Dinoperca petersi*) has a large, thick-walled swimbladder that is almost completely enveloped by three pairs of intrinsic muscles. Divers familiar with this species report that it is often seen at the entrance to its cave, making a loud drumming noise when approached.

## **REPRODUCTION AND DEVELOPMENT**

Fishes reproduce in two main ways, by spawning eggs (oviparity), or by producing live young (viviparity). Most fishes are oviparous, and produce eggs that are fertilised, develop and hatch outside the body of the female. Unfertilised eggs are released into the water by a female at the same time as a male releases sperm to fertilise the eggs.

Oviparous species are the most fecund of all animals, some species releasing hundreds of thousands of eggs at a time. Tropical species generally spawn more than once a year and usually in small groups or in pairs. The small pelagic eggs float near the surface and take only a day or two to hatch. Soon after hatching, the rapidly growing larvae exhaust their small yolk supply and must begin feeding. In tropical waters larvae will spend about eight weeks drifting in the surface waters and feeding on plankton. In temperate seas, the larval stage may last four or five months (12 months or more for *Anguilla* spp. eels). Such long-lived pelagic larvae are often carried far from their place of origin and are thus a means of dispersal for shallow-water fishes (see section **Larval fish dispersal**).

Some oviparous species produce non-buoyant (demersal) eggs, which are larger than pelagic eggs and are generally attached to the substrate. Many species with demersal eggs, such as damselfishes (Pomacentridae) and triggerfishes (Balistidae) also guard their eggs until they hatch. A further adaptation for protecting the eggs is some form of incubation. A male seahorse has a pouch on his belly in which he keeps the eggs until they hatch; he then releases the baby seahorses from his pouch by bending his body and apparently pushing the juveniles out with contractions of his abdominal muscles. Once released, the little seahorses must fend for themselves. Sea catfishes (Ariidae) and many cardinalfishes (Apogonidae) practice oral incubation, in which the male keeps the eggs in his mouth until they hatch.

Larvae that hatch from demersal eggs are bigger than most larvae that hatch from pelagic eggs, and their larval stage is correspondingly briefer (sometimes even non-existent). In chondrichthyans (chimaeras, sharks and rays) the hatchlings
are miniature copies of the adult, and the eggs that produce these well-developed juveniles are quite large — a whale shark egg-case measures  $30 \times 15$  cm.

Although most oviparous fishes fertilise their eggs after they have been released from the body of the female, a few bony fishes and all chondrichthyans fertilise their eggs inside the female. Internal fertilisation is usually accomplished with special copulatory structures called intromittent organs, but some species, such as the coelacanth (*Latimeria*), somehow manage internal fertilisation without an intromittent organ. In bony fishes, the intromittent organ can either be a fleshy 'penis' (as in Indo-Pacific Clinidae), or a modified anal fin, the andropodium of Hemirhampidae. A male chondrichthyan has paired intromittent organs (misnamed claspers) that are modified parts of the pelvic-fin skeleton.

Teleosts, such as Indo-Pacific Clinidae, and most elasmobranchs are viviparous and give birth to active, freeswimming young. Species that provide their embryos with no extra nourishment (other than their original yolk supply) are called ovoviviparous, amongst them the spiny dogfishes (Squalus spp.), the whitespotted smoothhound (Mustelus *palumbes*), and the coelacanth. The developing embryo of most viviparous elasmobranchs is provided with some nourishment from the mother in addition to its original yolk supply. Carcharhinoid sharks (e.g., Carcharhinus spp., Mustelus mustelus, and the hammerheads [Sphyraenidae]) have a yolksac-placental connection between the foetus and the oviduct of the mother. This placental source of nutrients develops during the later months of the gestation period, after the initial yolk supply has been exhausted. In some shark species the foetus is nourished by feeding on eggs and smaller siblings that may be found in the oviduct. These oophagous or adelphophagous (intrauterine cannibalistic) sharks (e.g., Carcharias taurus) give birth to only one or two large pups (one per uterus) at a time. In the oophagous foetal nourishment scheme, the ovary

enlarges and functions as a kind of milk gland, sending little packets of yolk into each uterus to feed the developing foetus.

The occurrence of both sexes in the same individual, hermaphroditism, is normal and common in many bony fishes. Functional hermaphroditism may be either synchronous or sequential. The gonads of synchronous hermaphrodites produce ripe ova and sperm at the same time. Although synchronous hermaphrodites are capable of fertilising their own eggs, this does usually not occur, at least not in those species where spawning has been observed. The synchronous serranids that have been studied (Serranus and Hypoplectrus spp.) spawn in pairs and take turns fertilising each other's eggs. Successive hermaphrodites change sex, beginning the reproductive period of their lives either as females (protogynous) and after a few years or spawning periods they change to males, or they begin as males (protandrous) and then change to females. Although hermaphroditism is rare in freshwater fishes, it is common in marine fishes, especially in mesopelagic and coral-reef species. Many coral-reef fishes such as Labridae, Scaridae, Serranidae, Anthiadidae, Sparidae, Lethrinidae, Polynemidae, and others - are successive hermaphrodites.

An interesting reproductive pattern has evolved in the sexually dichromatic fishes of the genus *Pseudanthias*. Species form small schools and occupy an area of a reef — a territory. Each school comprises a few (sometimes only one or two) large, distinctively coloured males, several smaller adult females and numerous juveniles coloured like the females. Each male spawns with several females in a harem-like fashion. If a male is removed from the group, the dominant (largest) female changes sex and colour pattern to take the place of the missing male. This sex and colour pattern change occurs within a few days, but if another male is introduced into the colony before the dominant female has completed her sex change, she returns to her female role and livery (Figure 12).



Figure 12 Pseudanthias cooperi (males with white streak under eye, females mauve, paler ventrally) and P. squamipinnis (males with yellowish sides and red pectoral fins, female orange) (South Africa). © M Fraser

#### COLOURATION AND BIOLUMINESCENCE

Fishes living in the well-lit epipelagic region of the sea are countershaded (dark above and silvery below). Viewed from above, their dark greenish blue backs match the colour of the deep ocean. Seen from below, these fishes look like silvery streaks against a silvery mirror. Below 300–400 m, fishes are mostly uniform black; a few are bright red, but appear black in their natural environment because the red component of sunlight is filtered out in the first 10–20 m of water. In the neritic zone (the shallow area over a continental shelf) pelagic fishes, such as kingfishes, sardines and mackerels, are generally silvery and countershaded, but the more sedentary sublittoral fishes are often coloured in a variety of colours and patterns. The many beautiful coral-reef fishes are among the most colourful of all animals.

The silvery iridescent colours of fishes are due to special cells in the skin or scales. These cells, called iridophores, contain crystals of guanine arranged in layers which reflect several colours depending on the angle of the incoming light and position of the observer. The other (non-iridescent) colours of fishes are produced by chromatophores. These cells contain various pigments, and each chromatophore has only one kind of pigment. A chromatophore is typically irregular in shape, with numerous branching processes extending outwards from its centre. The pigment granules can be moved within the chromatophore: when the pigment is dispersed throughout the cell, the colour is intensified; when the granules are concentrated in the centre of the cell, the colour is diminished. The skin of fishes may contain several different chromatophores (red, yellow, black, etc.) and it is the alternating contraction and expansion of the pigment in the various chromatophores that produces the changes in colour pattern.

Rapid colour changes are under nervous control. For example, a dark galjoen (*Dichistius capensis*) swimming over dark rocks can change immediately to a pale colour when it swims over a light sandy area. The immediate contraction of the melanophores is responsible for this quick change. Fishes often have a night colour pattern that is quite different from their daytime colouration, and a fish under stress may display its night pattern during the day.

Long-term colour changes, such as the transition from juvenile to adult patterns, are affected by increases or decreases in the number of various chromatophores. This type of colour change is under hormonal control, as is that of sexually dimorphic fishes that change their colours when they change sex or become mature. Some fishes maintain their colour to match their habitat.

Disruptive colouration is exhibited by many coral-reef fishes. Bold black bars or stripes break up the general outline of the fish, and irregular patches of contrasting colour or tones distract the eye from the overall shape of the fish. When a boldly patterned fish is swimming over the complicated, multicoloured background of a coral reef, the fish is difficult to recognise and hard to follow.

Another type of disruptive colouration is the eye stripe, a black band or bar on the head that passes across the eye and makes it less obvious. Some butterflyfishes not only have a black eye stripe, but also have a false eye spot or conspicuous ocellus at the rear end of the dorsal fin. This false eye spot is supposed to deflect the attack of a predator to a less vulnerable part of the fish (see also section **Mimicry and protective resemblance in WIO fishes** in the following pages).

Different colour patterns probably also serve in species for sex recognition. Sexual dichromatism seems to be most highly developed in hermaphroditic species, such as parrotfishes, wrasses and some angelfishes. This conspicuous difference in males and females is obviously important in the reproductive behaviour of these species. Some fishes exhibit sexual dichromatism only when they are about to spawn. Species recognition is important not only for mating, but also for fishes that establish and defend a territory, and for those that want to 'advertise' their presence to others.

Bioluminescence, the ability to produce light, is better developed in fishes than in any other group of animals. At least 45 families of marine fishes have species with light-emitting organs (no freshwater fishes are light-producing). Most of the light-producing fishes are found at depths of 300–1 000 m, but there are some light-producing shallow-water species, such as the pineapple fishes (*Monocentris*), flashlight fishes (*Photoblepharon*) and cardinalfishes of the genus *Siphamia*.

Most luminous fishes produce substances that react chemically to produce light without heat. This reaction takes place in light organs (photophores) and is under nervous control. Some species rely on symbiotic luminous bacteria for their light. These bacterial light organs glow continuously, and the light can be turned off only by covering or occluding the light organ in some way. In the flashlight fish (*Anomalops*), the large bacterial light organ under the eye is 'turned off' by rotating it down into a black-pigmented pocket under the eye.

### MIMICRY AND PROTECTIVE RESEMBLANCE IN WIO FISHES

Helen A Randall and John E Randall



Figure 13 A The cleaner wrasse, Labroides dimidiatus (Oman); B the mimic blenny, Aspidontus taeniatus (Maldives). © JE Randall, Bishop Museum



Figure 14 An aquarium photograph of the sargassumfish, *Histrio histrio* (Hawaii). © JE Randall, Bishop Museum



Figure 15 A Antennarius commerson, anglerfish, and Porites lobata, coral (Hawaii); B same fish as above, lifted off bottom (Hawaii). © JE Randall, Bishop Museum

The word mimicry is derived from the Greek term *mimetikos*, meaning imitative, and was first used in reference to people. Among animals other than *Homo sapiens*, the word has most often been used for insects. However, a surprising number of mimics have been discovered among marine fishes (Randall 2005). Classic examples include the resemblance of the black-and-white-barred snake eel *Myrichthys colubrinus* to the very venomous sea snake *Platurus colubrinus* (McCosker & Rosenblatt 1993), and the close resemblance of the false scorpionfish *Centrogenys vaigiensis* (from Indonesia) to the golden scorpionfish *Parascorpaena aurita*, with its full complement of venomous dorsal-fin spines (Whitley 1935).

There is a distinction between protective resemblance and mimicry. Poulton (1898) defined them as follows: protective resemblance is when an animal resembles some object which is of no interest to a predator, and is thus concealed; mimicry is when an animal resembles an object which is well known and avoided by predators, and in so doing becomes conspicuous.

Observations of the cleaner wrasse Labroides dimidiatus (Figure 13A) and the mimic blenny Aspidontus taeniatus (Figure 13B) led Randall & Randall (1960: 445) to revise Poulton's definition of mimicry to "... an animal resembles an object which is well known and is avoided, or not preyed upon by its enemy, and in so doing becomes conspicuous." Far from being avoided, the cleaner wrasse is sought by other fishes of the reef community to remove ectoparasites. It has a conspicuous pattern of bright blue with a broad horizontal black band from the snout to the caudal fin, and occupies a particular area (territory) of the reef, above which it swims in a distinctive, jerky up-and-down fashion. Fishes recognise the cleaner and come to its 'cleaning station' to be picked over for their parasites, and the cleaner is thus protected from becoming someone else's meal. But, the cleaner wrasse also serves as the model for the mimic blenny (also called the false cleaner) enabling the latter to get close enough to nip pieces from the fins of other fishes that mistake it for a cleaner wrasse. As long as the mimic is relatively rare (compared to the model), it is afforded some safety from predation and can successfully practise its game of deception.

Examples of protective resemblance in fishes are legion. Many fishes that rest on the bottom have the ability to blend with the background in colour, and some, such as scorpaenids (scorpionfishes), lophiids (monkfishes) and antennariids (frogfishes) have cirri or cutaneous flaps that augment their concealment. The venomous stonefish looks just like a lump of coral rock covered with algae, and is a real hazard to waders who might tread on it. The sargassumfish (*Histrio histrio*) (Figure 14) is among the first to come to mind. It is very difficult to detect when it is hiding in drifting masses of *Sargassum* weed. Another, *Antennarius commerson* (Figure 15), is well concealed by looking like a small colony of the coral *Porites lobata*. Flatfishes often match the pattern and colour of the substrate so well that they are almost impossible to see unless they move.

There are many fishes that resemble plants, reviewed by Breder (1946). Atz (1951) published a popular account and Randall & Randall (1961) added further examples, one being the deep-bodied yellow juveniles of *Platax orbicularis* (Figure 16) that is often seen flat on its side, on the surface of the water, drifting among yellow leaves of *Hibiscus tiliaceus*.



Figure 16 Juvenile *Platax orbicularis* floating on side at surface where it resembles similar drifting yellow hibiscus leaves. © JE Randall, Bishop Museum



Figure 17 Juvenile Novaculichthys taeniourus drifting like a small mass of detached algae (Hawaii). © JE Randall, Bishop Museum



**Figure 18** The pipehorse *Syngnathoides biaculeatus* in a seagrass bed (Philippines). © JE Randall, Bishop Museum



Figure 19 The tiny seahorse *Hippocampus* bargibanti on the seafan *Muricella* (Indonesia). © JE Randall, Bishop Museum

The fish was noticed only when it moved in a different direction from the leaves. A similar example was described for the juvenile of the tripletail (*Lobotes surinamensis*) by Schmid & Randall (1997) in the Red Sea. The juvenile of the peacock razorfish (*Iniistius pavo*) was mistaken as a blackened leaf moving to and fro with other plant debris from surge in shallow water over white sand. Its long filamentous dorsal-fin spine was held forward and looked like the stem of the leaf. Again, the fish was noticed only when it moved differently from the plant debris and, as it was approached, it quickly dived into the sand. Another remarkable juvenile wrasse, the rockmover (*Novaculichthys taeniourus*), looks like a small drifting mass of algae (Figure 17), and can be green, olive, maroon or brown.

Several fishes that inhabit seagrass beds or substrata with dense beds of algae are coloured like these plants. Prominent green examples include the alligator pipehorse *Syngnathoides biaculeatus* (Figure 18), the ghost pipefish *Solenostomus halimeda*, the wrasses *Novaculoides macrolepidotus*, *Pseudojuloides argyreogaster* and *Pteragogus pelycus*, the raggedtooth parrotfish *Calotomus spinidens*, the filefish *Acreichthys tomentosus*, and individuals of the klipfish, *Clinus superciliosus*, that live in kelp. Other individuals of this species will take on the colour of the substrate the juveniles settle on when they leave the plankton.

Seahorses are famous for being protectively coloured; our favourite is the tiny *Hippocampus bargibanti* that is difficult to spot until it moves on the sea fan *Muricella* sp. (Figure 19). It is not known from the Indian Ocean, but it is so easily overlooked when it might be present.

The most intriguing examples of mimicry in fishes are those where a harmless species mimics one with some repelling property such as a toxic species, or one with a venomous bite or fin spines. An example of the latter from the Red Sea is the fangblenny *Meiacanthus nigrolineatus*, which has a vicious venomous bite. It is strikingly mimicked by the blenny *Ecsenius gravieri* (Figure 20). A second mimicking blenny, *Plagiotremus nigrolineatus* enjoys the same benefit, despite its slender body (Springer & Smith-Vaniz 1972). Some polyclad flatworms are known to secrete toxins that makes them repellant to predators. A few species of soles have effectively mimicked these flatworms, as has the juvenile of *Platax pinnatus* (Randall & Emery 1971) (Figure 21). The young of the African pompano (*Alectis ciliaris*) (Figure 22) have very long, filamentous dorsal- and anal-fin rays that resemble the long thread-like tentacles of venomous jellyfishes, such as those of the virulent box jellyfishes of Indonesia and the Coral Sea.



**Figure 20** The blenny *Ecsenius gravieri* (left), a mimic of the sabretooth blenny *Meiacanthus nigrolineatus* to the right (Red Sea). © VG Springer, USNM



*pinnatus*, a polyclad flatworm mimic (Palau). © JE Randall, Bishop Museum



**Figure 22** A juvenile *Alectis ciliaris* with long dorsal and anal rays that resemble jellyfish, with long, thread-like, stinging tentacles (Hawaii). © JE Randall, Bishop Museum

Tobies (*Canthigaster* spp.) are well known to harbour crinitoxins that cause predaceous fishes to eject them when they attempt to feed on them. Species of the filefish genus *Paraluteres* have successfully mimicked these small puffers, thereby enjoying relative freedom from predation (Clark & Gohar 1953). The example here (Figure 23) is unusual in having both model *Canthigaster valentini* (lower) and its mimic *Paraluteres prionurus* (top) together, as if fooled by their own trickery. The mimicking filefish keeps its dorsal spine folded flat to augment this deception. The juvenile of the grouper *Plectropomus laevis* may also be a mimic of this puffer, though not as perfect.

The juveniles of some predatory fishes, such as the groupers *Anyperodon leucogrammicus* and *Epinephelus jayakari* and the snapper *Lutjanus bohar*, have evolved to mimic harmless fishes such as damselfishes or small species of wrasses, thereby being able to approach their prey more closely. Figure 24 shows the juvenile of *E. jayakari* from the Persian/Arabian Gulf that mimics the damselfish *Neopomacentrus sindensis*.

The biological basis for mimicry is usually clear. However, it took considerable time to determine why a surgeonfish of the genus Acanthurus should gain any benefit from mimicking a small angelfish of the genus Centropyge. Juveniles of Acanthurus pyroferus in the Pacific and eastern Indian Ocean and A. tristis in the Indian Ocean precisely mimic adults of the small angelfishes *Centropyge flavissimus* and *C*. eibli (Figure 25), respectively. When the surgeonfishes grow larger, they revert to the relatively drab colour of adults. It was first thought that the Centropyge models would be cleaners and enjoy freedom from predation. Hours were spent observing individuals of the little angelfishes, hoping to see them pick at the bodies of other fishes, but this was not ever seen. Then it was thought that they must have some toxic property to make them unpalatable. However, when served as bait, they were taken by predators as readily as control fish of the same size and shape. Finally, it was found that these little angelfishes never move far from shelter. They feed on algae and detritus in the immediate vicinity of their shelter. Surgeonfishes, by contrast, range widely in search of their preferred filamentous algae, and are therefore much more vulnerable to predation. If they resemble a small angelfish of the genus Centropyge, a potential predator moves on to easier prey.

Dafni & Diamant (1984) proposed the term 'school-oriented mimicry' when a normally solitary fish species mingles with a similar, aggregating species for the advantage of schooling. Their example is the juvenile of the fangblenny *Meiacanthus nigrolineatus* that in the Red Sea and Gulf of Aden joins a school of apogonid fishes of similar colour pattern. Randall & McCosker (1993) preferred the term 'social mimicry' for this. Their example from the Pacific, is a mixed planktonfeeding school of three anthiine fishes, a damselfish, and a blenny, all with a yellow and pink colour pattern. They also cited *Mulloidichthys mimicus* Randall & Guézé of the Marquesas and Line Islands that schools with the snapper *Lutjanus kasmira*. Uiblein (2011) described a new goatfish *Mulloidichthys ayliffe* from the Western Indian Ocean, and noted that it is similar to *M. mimicus*, and that it also schools with *Lutjanus kasmira*, as well as *M. vanicolensis* (Figure 26).



Figure 23 The model toby Canthigaster valentini, with its mimic, the filefish Paraluteres prionurus, above. © J Hall



**Figure 24** A The Arabian demoiselle *Neopomacentrus* sindensis, and **B** its mimic, the juvenile Muscat grouper *Epinephelus jayakari* (both Persian/Arabian Gulf). © JE Randall, Bishop Museum



**Figure 25 A** The blacktail angelfish *Centropyge eibli* and **B** its mimic, surgeonfish *Acanthurus tristis* (both Indonesia). © JE Randall, Bishop Museum



**Figure 26** *Mulloidichthys ayliffe* (centre) schooling with *M. vanicolensis* (lower right) and *Lutjanus kasmira* (in background) (South Africa). © DA Polack

### THE OCEANOGRAPHY OF THE WESTERN INDIAN OCEAN

#### Johann RE Lutjeharms and Wayne S Goschen

For an appropriate understanding of the life histories and hence the geographic distributions of marine fishes, we need to know something about the basic oceanography (or oceanology) of a region. This includes the hydrographic and chemical characteristics of the main water masses, such as temperature, salinity and nutrient content, the major ocean currents, regions of upwelling, and prevailing primary productivity.

Observation, theory and numerical modelling are all used to describe ocean currents. These can provide a reasonable approximation of the largest water masses and their movements, though global or regional theories and models generally do not account for nonlinear and turbulent processes. In situ observations of ocean currents are relatively meagre in time and space, yet different water masses can be tracked according to their potential temperature and salinity (TS scatter plots) because these properties are retained from the place of formation. Remote sensing has become an important tool in determining surface currents, while subsurface floats, moorings and profiles can provide an account of the vertical water structure. Such knowledge is especially important for understanding the physical aspects of the Indian Ocean and its adjacent seas as this expanse covers a wide spectrum of ocean circulation patterns and conditions.

As described in the Introduction, the WIO region (Figure 1) is here defined as bounded by the southern coast of Africa, northward to the Arabian Sea, and eastwards to the longitude of Kanyakumari (formerly Cape Comorin) at the southern tip of India; southwards the region extends to the Subtropical Convergence Zone, the generic border between waters of tropical origin and the Southern Ocean.

By this definition, WIO encompasses a number of oceanic systems. The greater Agulhas Current system flows between the northern end of the Mozambique Channel and southern Cape coast of South Africa.

This system includes the various sources of the Agulhas Current proper as well as part of its outflow (Figure 2). To the north of this system, the powerful influence of the monsoonal winds drives the Somali Current and circulation in the Arabian Sea, which alters direction dramatically with transitions between seasons. The regional seas, the Red Sea and Persian/ Arabian Gulf, comprise a separate grouping of ocean-like systems. In many aspects those systems are 'artificial' because water is continually exchanged between the peripheral water body and WIO, hence influencing each other. For instance, Red Sea Intermediate Water, with very high salinities, is formed in the Red Sea and Persian/Arabian Gulf from where it moves southward into WIO at depths to about 1 000 m, and still retains its identity upon reaching the southern end of the African continent and beyond.



Figure 1 The geography of the Western Indian Ocean and its bottom configuration. Depths are given in metres at three levels. The names of some prominent ridges and plateaux are: A Agulhas Bank, B Southwest Indian Ridge, C Madagascar Ridge and Walters Shoals, D Mozambique Ridge, E Mascarene Plateau, F Southeast Indian Ridge, G Carlsberg Ridge, H Somali Basin, I Madagascar Basin, J Mozambique Basin, K Agulhas Basin, L Southwest Indian Basin, M Chagos-Laccadive Ridge.

### THE GREATER AGULHAS CURRENT SYSTEM

#### South Equatorial Current

In the WIO, the South Equatorial Current occurs as a broad stream of mainly surface water, generally south of 10° S, which forms the westward component of the wind-driven Southwest Indian Ocean subgyre (Figure 2); this is the main source of the greater Agulhas Current system. The current is most strongly developed in the upper 300 m, and its surface waters exceed 28 °C at the end of the Southern Hemisphere summer (March) but are <26 °C in June to November. The current has a mixed-



Figure 2 A stylised portrayal of the main surface currents of the Western Indian Ocean. Currents in the northern Indian Ocean are given as solid for the Southwest Monsoon period and as broken for the Northeast Monsoon. Blue regions indicate upwelling. The most prominent circulation features are: A South Equatorial Current, B northern branch of the East Madagascar Current, C southern branch of the East Madagascar Current, **D** South Indian Ocean Countercurrent, **E** Mozambigue Channel eddies, F intermittent retroflection of the southern branch of the East Madagascar Current, G Agulhas Current, H Agulhas Current retroflection, I Agulhas Return Current, J South Indian Ocean Current, K Subtropical Convergence (blue broken line), L Port Alfred upwelling cell, M Angoche upwelling cell, N South Madagascar upwelling cell, O South Equatorial Current, P Somali Current during Southwest Monsoon, Q Southwest Monsoon Current, Equatorial Countercurrent during Northeast Monsoon, R East Arabian Current during Southwest Monsoon, S West Indian Current, **T** North Equatorial Current during Northeast Monsoon, **U** Somali upwelling during onset of Southwest Monsoon.

layer depth of ~40 m in March, and increases to 100 m by August. It has a shallow salinity minimum (i.e., an intrusion of low-salinity water) of Indian Tropical Surface Water, overlying higher-salinity South Indian Subtropical Surface Water (Figure 3). The speed of this current is relatively low, usually less than 0.3 m/s, and it extends to 8° S, but expands northward to 6° S in September.



**Figure 3** A Potential temperature–salinity and **B** potential temperature– dissolved oxygen relationships for the Western Indian Ocean. South Indian Central Water is identified by SICW, Sub-Antarctic Mode Water by SAMW, North Atlantic Deep Water by NADW, Antarctic Bottom Water by AABW, Antarctic Surface Water by AASW, Antarctic Intermediate Water by AAIW, Sub-Antarctic Surface Water by SAASW, Lower Circumpolar Deep Water by LCDW and Upper Circumpolar Deep Water by UCDW. Source: Gordon *et al.* 1987

The South Equatorial Current flows from east to west; it receives a contribution of Pacific Ocean water at the Indonesian Throughflow and strengthens as it moves westward. South of the Seychelles, it is largely disrupted by the shallow Mascarene Plateau (Figure 1), which affects the flow even at 50 m depth. Water moves through two gaps in this plateau, so that the South Equatorial Current west of the plateau consists of two relatively intense jets: one at 13° S and the other at 18° S. The more intense northern jet passes the northern coast of Madagascar and the Comoros (Figure 2) and then divides at the coast of East Africa, with some water flowing southward into the Mozambique Channel and the remainder northward along the coast as the East African Coastal Current. These currents seem to be strongest during the Southwest Monsoon in May-September, with peak speeds in July.

#### South Indian Ocean Countercurrent

It is generally accepted that the surface flow of the southern Indian Ocean circulates in an anticlockwise direction, as typical of a subtropical gyre. The northern branch consists of the South Equatorial Current (described above). The main southern branch is the intense Agulhas Return Current, and east of about 70° E is the South Indian Ocean Current (Figure 2). Consequently, the border between the average eastward flow and westward flow lies at about 30° S, the latitude of the port city of Durban, South Africa. There is one notable and curious exception, however. From the southern end of Madagascar a narrow South Indian Ocean Countercurrent carries water at about 25° S across the greater width of the subtropical Indian Ocean, opposite the main general flow. Notwithstanding this current's limited width, it carries a substantial volume of water, about  $10 \times 10^3$  m<sup>3</sup>/s, and is evident to a depth of at least 800 m. The origin of the water in the South Indian Ocean Countercurrent is not yet known with certainty, but it is presumed to originate at the southern end of the southern branch of the East Madagascar Current (described below). Significantly, this countercurrent also presents a means for passive, non-swimming organisms from the tropics to be carried eastward across the southern Indian Ocean.

#### East Madagascar Current

Where the southern jet of the South Equatorial Current impinges on the east coast of Madagascar, at about 17° S, the current splits to form the northern and southern branches of the East Madagascar Current. Both branches closely follow the continental shelf edge and increase in speed and volume flux downstream. In their well-developed state, the waters reach a speed of 1.5 m/s and a volume flux of about  $25 \times 10^6$  m<sup>3</sup>/s. At this point they reach a width of about 80 km and depth of 600+ m and have no known distinctive seasonality. The northern branch of the East Madagascar Current joins the northern jet of the South Equatorial Current as it flows past the northern end of Madagascar. The southern branch terminates south of Madagascar in either a retroflection that feeds the above-mentioned South Indian Ocean Countercurrent, or else as a jet moving westward towards the coast of Africa.

The upper 50 m of the East Madagascar Current consist of Indian Tropical Surface Water, with temperatures of >23 °C and salinities of <35 (Figure 3). Offshore and below this water is South Indian Subtropical Surface Water, with salinities of >35.6 and a subsurface oxygen minimum of 4 ml/l at a depth of 200 m. The influence of this narrow and intense current on the adjacent coastal waters is unknown – but it can be assumed to be strong along the narrow continental shelf. A distinct upwelling cell forms on the wider continental shelf southeast of Madagascar, with a decrease of about 2 °C in temperature and an increase in chlorophyll *a* concentrations; maximum chlorophyll fluorescence is attained at depths of 40–100 m. Some of this chlorophyll-rich continental shelf water may be taken up by clockwise eddies and shed in a westward direction.

East of the East Madagascar Current, in the open ocean, a strong seasonal phytoplankton bloom develops. It usually begins in January, is most intense during February–March, and may last until April. During its peak it extends over most of the Madagascar Basin, moving eastward (up to 70° E) as a wave of enhanced phytoplankton concentration. However, the bloom has failed in some years, which is assumed to be a result of seasonally increased wind strength that deepens the layer that is mixed, thus entraining nutrients into the photic zone. (The wind mixes the surface-layer waters to the same temperature; below this surficial mixed layer is the thermocline, defined by a sudden decrease in temperature with depth. Nutrients are trapped in the upper mixed layer, the photic zone, by the thermocline.)

#### Mozambique Channel circulation

Circulation in the Mozambique Channel was poorly understood until recently when it was discovered that the Mozambique Current consists largely of a series of clockwise and anticlockwise eddies, formed at (or just north of) the channel's narrows and which then drift southward. Water from the South Equatorial Current (Figure 2) leaks into the Mozambique Channel from the north, along Mozambique's coast. This course of water may be only ~150 m deep and 250 km wide, and likely flows at <0.5 m/s. It moves through the Comoros Basin south of the Comoro Islands (Figure 1), generally circulating anticlockwise. The eddies formed in the basin and at the Channel's narrows are about 300 km in diameter, extend all the way to the ocean floor, and move southward at ~5 km/day. Generally, one or more eddies are formed each year and collectively carry about  $15 \times 10^6$  m<sup>3</sup>/s of water southward through the Channel.

These eddies seem to have two noticeable effects on the local ecosystem. First, they may carry chlorophyllrich water from the continental shelf to the centre of the Mozambique Channel; second, primary productivity seems to be concentrated at their margins, because frigate birds and other predators have been seen to feed there. It is likely that concentrations of tuna are found at these eddy edges, too. Furthermore, passing eddies may stimulate upwellings where the coastline forms bights, large bays, or capes (collectively termed offsets). For instance, intense upwelling has been observed off the town of Angoche (16°14' S, 39°55' E) where the highest concentrations of chlorophyll *a* in the Mozambique Channel have been recorded.

A deep countercurrent in the channel has been observed at intermediate depths, at  $\sim$ 1 000 m, and carries water towards the equator at a speed of  $\sim$ 0.5 m/s.

The continental shelf of Mozambique is wide, and in places this may be supported by runoff from major rivers. At the major shelf area at Sofala Bank, where the Zambezi River flows into the sea, the surface outflow signature can cover an area of 50-170 km<sup>2</sup>. Salinities as low as 20 have been measured under flood conditions and as far as 50 km from the river mouth. Such relatively freshwater is usually restricted to the upper 10 m. Interestingly, in the case of Sofala Bank, this freshening of the marine waters of the continental shelf by river outflow may be compensated by an outflow of water with salinities as high as 37, originating from the salt marshes at the southern end of the bank. Low-lying coastal areas and estuaries, in Mozambique especially, contain extensive areas of salt marshes and mangrove swamps formed as a result of a tidal range of between 2 and 5 m. Over the wide, shallow areas of the Sofala Bank, strong tidal currents also cause the continuous movement of sand banks and create other sedimentary seabed features, such as sand ripples.

#### **Delagoa Bight circulation**

The Delagoa Bight, off the city of Maputo, Mozambique, forms the largest offset on the East African coastline. The circulation in the Bight is different to the circulation along the rest of Mozambique's continental shelf: often there is a clockwise eddy of surface waters flowing northward past Maputo and eastward along the northern border of the Bight (Figure 2). The sediment-laden waters of the rivers that drain into the Bight, in particular the Limpopo River on its northern side, then usually move along the coast with these currents. The cold water at the centre of the Delagoa Bight eddy is upwelled from a depth of ~900 m, and though this water may be rich in plant nutrients, such as nitrates, phosphates and silicates, currently there is no evidence that this addition of nutrients into the upper water layer leads to enhanced primary productivity.

What drives this eddy? It was long assumed that the passing and overshooting Mozambique Current induces the Delagoa Bight eddy as a trapped lee eddy. However, the discovery that the Mozambique Current is not a continuous boundary current discounts this hypothesis. The irregular passage of Mozambique Channel eddies, with their anticlockwise rotation, may possibly be sufficient to drive the Delagoa Bight eddy intermittently, but it is not known how prevalent this eddy is. It is possible that the eddy may sometimes escape from the confines of the Bight's area, dramatically modifying the circulation in the Bight until it gets spun around again. If and how often this happens, remains uncertain. At other places in the WIO where a boundary current, for example the Agulhas Current, passes from a narrow to a wide area of continental shelf, there is evidence of upwelling. If a constant Mozambique Current existed, one would similarly expect an upwelling cell at the northeastern bend of the Delagoa Bight. Such an upwelling arises only intermittently though; this suggests that water circulation in the Bight is driven by the occasional passage of Mozambique Channel eddies.

The water masses in the Delagoa Bight consist of South Indian Subtropical Surface Water, with salinities of 35.5–35.6 in the surface layers, with possible insertion of Indian Tropical Surface Water of lower salinities (<35.6) at times. Below these surface waters the South Indian Central Water extends to a depth of at least 1 000 m.

#### The northern Agulhas Current

It is not known exactly where the southwestward-flowing Agulhas Current becomes fully formed and starts to carry water past the African continental shelf. Sediment distributions suggest that the current takes form somewhere between Richards Bay in South Africa and Maputo in Mozambique (Figure 2). South of Richards Bay, the Agulhas Current increases in depth and volume, to a full volume flux of  $\sim 65 \times 10^6$  m<sup>3</sup>/s, with surface speeds of more than 2 m/s, and a surface width of roughly 100 km. Here, the current follows the shelf edge closely (Figure 4) and is underpinned by a countercurrent occurring at depths of 800+ m and which hugs the continental slope and carries mostly cold Antarctic Intermediate Water northward towards the Equator. Similar undercurrents are found in the Mozambique Channel and also east of Madagascar, but models using water-volume data suggest that those undercurrents are not connected to the Agulhas Undercurrent.



Figure 4 Satellite image in the thermal infrared of the surface layers of the Agulhas Current. The temperature scale (°C) is given on top; white areas represent clouds. The continental shelf edge at 200 m depth is indicated by a black line. Note how the northern Agulhas Current, up to Port Elizabeth, is nearly rectilinear whereas the southern Agulhas Current exhibits considerable sideways meandering and the formation of shear edge eddies and plumes of warm water. Regions shaded blue on the west coast indicate wind-driven, shelf-edge upwelling. Water of similar temperatures to the south represents cold Subtropical Surface Water poleward of the Subtropical Convergence. Source: Lutjeharms 2011



**Figure 5** Satellite image in the thermal infrared of the surface temperatures of the Agulhas Current showing a Natal Pulse south of Port Elizabeth. The cyclonic eddy which nestled in the offshore portion of the meander is evident as are other shear cyclonic eddies, plumes and filaments on the inshore side of the Agulhas Current. Possible eddies in the KZN Bight and off Durban are evident. Water with higher temperature is represented by shades of red and cooler shelf water by shades of green. Coastal upwelled water around Port Elizabeth is shown as shades of purple and blue. Source: Lutjeharms 2006

The water masses of the northern Agulhas Current are largely Indian Tropical Surface Waters in the current's surface layers, with South Indian Subtropical Surface Water offshore. Subtropical Surface Water is also found within the current at a depth of 150–200 m. Plugs of Red Sea Water are found at intermediate depths on the inshore side of the current. The surface temperatures show a decline of ~2 °C moving southward along the length of the northern Agulhas Current and are ~28 °C in summer and ~23 °C in winter.

The northern Agulhas Current flows along a generally narrow continental shelf (Figure 4). The current is intermittently absent (generally 1.6 instances per year, but >3 times per year is not uncommon) from the shelf edge. This is the result of the passage of large, solitary meanders of the current, the so-called Natal Pulses, that are thought to be triggered at the KwaZulu-Natal Bight (an unusually wide offset on an otherwise narrow shelf) by an offshore eddy encroaching on the coast (Figure 5). Natal Pulses move downstream at ~20 km/day, grow laterally as they move, and carry an embedded clockwise eddy. This eddy may reverse a portion of the currents on the shelf for a short period as it passes, and such reversals, as well as accompanying decreases in temperature, may have important effects on the distribution and movement of biota, but currently no data are available to confirm these suppositions.

#### KwaZulu-Natal Bight circulation

The KwaZulu-Natal (KZN) Bight between St Lucia and Durban forms an offset in the eastern coastline of South Africa (Figure 6), which in many aspects is similar to the Delagoa Bight to the north. The KZN Bight is much narrower and shallower, however, resulting in a very different circulation pattern, generally found to be clockwise, but which may contain a number of eddies at any given time, and it may be rapidly disrupted by strong winds. Nonetheless, the water masses and circulation of this bight are characterised by some interesting and important features.

The eastern side of the KZN Bight is bounded by the Agulhas Current proper. At Durban the shelf narrows sharply and the Agulhas Current overshoots it, only to join the shelf edge again further downstream. Thus, off Durban, a lee eddy with clockwise motion is generated. This eddy may be carried away with the formation of a Natal Pulse, but it otherwise seems a fairly persistent feature, bringing deeper waters closer to the surface at its centre. By contrast, at the northern edge of the bight a peculiar upwelling cell is located between Richards Bay and St Lucia.



**Figure 6** A conceptual illustration of the general cyclonic circulation patterns in the KZN Bight. The circulation of the bight consists of cyclonic eddies and is constrained in the coastline and the Agulhas Current, which is generally located near the 200 m isobath. Isobaths are in metres. Source: Schumann 1987

This upwelling is both wind-driven and formed by the Agulhas Current as it moves from a narrow to a wider area of the continental shelf. The upwelled water originates from the South Indian Central Waters and is rich in nutrients.

This colder, nutrient-rich water moves along the seafloor at the KZN Bight, but can easily become mixed throughout the relatively shallow water column during a strong wind. The resulting increase in primary productivity at the St Lucia upwelling cell is well documented.

The phytoplankton generated in this cell is sometimes transported southwards in a strand of water along the shoreward edge of the Agulhas Current. Whether this upwelling cell has a noticeable influence on the benthic or pelagic ecosystems of the Natal Bight as a whole is still unknown.

The water masses of the KZN Natal Bight are mostly South Indian Subtropical Surface Water and Indian Tropical Surface Water, with an insertion of South Indian Central Water at the St Lucia upwelling cell. The slightly fresher (with lower salinity) Indian Tropical Surface Water is probably inserted onto the KZN Natal Bight sporadically as it may be found in varying amounts and degrees of purity in the Agulhas Current. The Thukela River is the only major river draining into this bight, but even during floods the dilution of water on the continental shelf by river water is only noticeable very close to the river mouth.

#### The southern Agulhas Current

Downstream of Port Elizabeth, in the Eastern Cape, the southern Agulhas Current can be distinguished from the northern Agulhas Current because it follows a quite different path. Whereas the northern portion is characterised by its nearly constant location at the shelf break, with the exception of the intermittent passage of a Natal Pulse, the southern portion meanders to either side of its 'mean' path along the shelf edge, with accompanying shear-edge eddies and plumes of warm surface water (Figure 7). These meanders grow as the current moves downstream. The warm-water threads are usually only ~50 m deep; their contribution to the water mass and heat budget on the adjacent shelf remains unknown, although the warmer water occasionally extends entirely to the coastline. The induced shear-edge eddies on the shoreward border of the southern Agulhas Current draw up deeper water in their centres, but this water has not been recorded on the Agulhas Bank, the wide shelf region south of Africa.



**Figure 7** A conceptual portrayal of the Agulhas Current system. Areas shallower than 3 000 m are shaded. The edge of the continental shelf is marked by a dotted line. Intense currents are black and the general background circulation is indicated by open arrows. Cyclonic eddies are open; anti-cyclonic rings and eddies are black. Source: Lutjeharms 2006

The southern Agulhas Current sometimes moves southward at Port Elizabeth under the influence of a large Natal Pulse, and joins the Agulhas Return Current and thus does not continue along the shelf edge of the Agulhas Bank. Such a short circuit will usually last only a few weeks, but an unusual occurrence where the Agulhas Current has turned back on itself and flows eastward instead of westward for six months has been recorded.

The southern Agulhas Current extends to the seafloor and carries an estimated  $70 \times 10^6$  m<sup>3</sup>/s. In the upper 200 m, Indian Tropical Surface Water has a temperature of 16 °C or more; South Indian Subtropical Surface Water is usually found below this, with higher salinity; and deeper yet, South Indian Central Water has temperatures of 6–14 °C. At greater depths the southern Agulhas Current carries Antarctic Intermediate Water, a small amount of Red Sea Intermediate Water, and North Atlantic Deep Water. The warm surface waters of the current cool significantly as they move downstream. The temperature also changes with the seasons: during February (the Southern Hemisphere summer) it is ~23 °C, whereas in August (winter) it is only ~17 °C. As can be expected, the southern Agulhas Current has a strong influence on the circulation and water masses of the Agulhas Bank.

#### Agulhas Bank circulation

The Agulhas Bank (Figure 7) is the widest part of the continental shelf of South Africa, and it represents the centre of South African commercial fishing grounds. The Bank is unique as it lies between a westward-flowing boundary current on its east side, namely the Agulhas Current, and an eastward-flowing boundary current to the west, the Benguela Current of the South Atlantic Ocean gyre. These two currents largely influence the circulation, water masses and vertical stratification of the Agulhas Bank. These influences are reinforced by the bathymetry, with the shallow Alphard Banks shoal in the centre separating the circulation on the eastern and western sides of the Agulhas Bank.

On the eastern side of the Bank most of the bottom waters of the continental shelf follow the southwestwardmoving Agulhas Current. There is occasional input onto the Bank of warm surface water via plumes from shearedge eddies, as mentioned above. On the western side of the Bank the main water movement is northwestward. However, occasionally intense Agulhas Rings, which are large eddies rotating anticlockwise, may substantially increase the current speeds along the shelf edge. Just west of the southern tip of the Agulhas Bank, the Agulhas Current may also generate a clockwise lee eddy that forces water near the shelf edge towards the southeast. This eddy may escape westward or dissipate and lose its influence on the shelf circulation. Strands of warm surface water (~50 m deep) from the Agulhas Current also sporadically move along this western shelf edge. Coastal upwelling is common along the coastline of the Agulhas Bank under suitable wind conditions, particularly at coastal headlands.

The Agulhas Bank shows seasonal stratification of its waters. During summer the surface waters are warmed and a thermocline typical for the subtropics is formed. At the same time, however, the waters are cooled from below, considerably increasing the intensity of the thermocline. During winter this vertical stratification is destroyed by vigorous wind mixing to a depth of at least 75 m.

But, what cools the water from below? At the eastern corner of the Agulhas Bank, where the Agulhas Current diverges from the coastline, an upwelling cell forms off Port Alfred, Eastern Cape, drawing the colder South Indian Central Water into this cell. Although disputed, it is believed that the cell then moves onto the Agulhas Bank and forms an unusually cold bottom layer. The Agulhas Current itself may drive cold bottom waters far onto the continental shelf. It is possible that a resulting, exceptionally intense seasonal thermocline provides a more hospitable environment for spawning anchovy. The marine organisms occurring in the vicinity of the Port Alfred upwelling cell are also cold-water species, hence it has been suggested that this transitional area is a significant barrier to shelf biota along the southern and eastern shelf areas of South Africa.

#### Agulhas retroflection

After the Agulhas Current passes the southernmost tip of the Agulhas Bank, it abruptly turns to flow back into the southern Indian Ocean: this is known as the Agulhas retroflection (Figure 2). This retroflection is not geographically stable but one of the most variable worldwide. The retroflection loop, with a diameter of ~400 km, moves into the southeastern Atlantic Ocean, occludes and forms an Agulhas Ring that then moves off into the South Atlantic Ocean. About 6–9 rings are formed each year, and most disintegrate close to where they are formed as a result of instability or interaction with other rings. Those that remain intact may move across the full width of the South Atlantic Ocean while slowly spinning down over a couple of years. The water masses of the rings are identical to those found in the Agulhas Current, but with heat loss, evaporation, and wind action the top layers may become colder, more saline, and substantially mixed by convection.

As part of the process of ring shedding, intrusions of cold, nutrient-rich, Sub-Antarctic Surface Water penetrate between a newly formed ring and the new retroflection. These intrusions have temperatures of <17 °C, salinities of <34.9, and are present in the region about 40% of the time. These water masses and their biological diversity make this a remarkably variable region. Biologically, the most important aspect of the southern termination of the Agulhas Current is that it affords access for Indo-Pacific species to be carried to the Atlantic Ocean by surviving in intact Agulhas Rings until these rings reach warmer latitudes more amenable to the survival of subtropical and even tropical species.

#### Agulhas Return Current

Most waters of the Agulhas Current, having passed through the Agulhas retroflection, eventually enter the Agulhas Return Current which flows eastward at about 40° S (Figures 2 and 7). Gradual cooling occurs as the waters move eastward and become mixed with ambient water masses, so that at ~70° E the last vestige of the Agulhas Current has dissipated.

The Agulhas Return Current flows roughly parallel to the Subtropical Convergence, and in the process large turbulent eddies are formed, with ever-changing meanders and eddy shedding. The first equatorward meander in the eastward flow of the current occurs as it passes the Agulhas Plateau (Figure 7). This meander is considered relatively stable in shape and location (although further research is needed), whereas meanders farther downstream are more variable.

The Agulhas Return Current is approximately 100 km wide, extends to depths of 4 000+ m, and carries about  $44 \times 10^6$  m<sup>3</sup>/s in the upper 1 000 m alone. At its origin (the Agulhas retroflection) the surface speeds are little different to those of the Agulhas Current. The surface speed at 25° E has been measured at 1.3 m/s, whereas at 60° E it is only 0.4 m/s. At intermediate depths of ~1 000 m the current speed is 0.4 m/s, at longitudes to approximately 40° E. One notable water characteristic of the Agulhas Current is a subsurface oxygen minimum, but this is rapidly mixed farther out in the Agulhas Return Current. An unusual and important feature of the Agulhas Return Current in combination with the Subtropical Convergence Zone is the enhanced primary productivity of this frontal system.

#### The Subtropical Convergence

The Subtropical Convergence Zone is the generic southern boundary of the Indian Ocean. This is where colder Sub-Antarctic Surface Water, with a high nutrient content (excepting silica) and an equatorward drift component subducts below the warmer and nutrient-poor Subtropical Surface Water. In the southwestern Indian Ocean, the southern border of the Agulhas Return Current sometimes meets with the front (Figure 7) but remains separate from it, thus a distinct Agulhas Front can be observed, particularly when the meridional temperature and salinity gradients of the Subtropical Convergence become steeper, rendering it one of the most intense fronts in all oceans. A temperature gradient of 5 °C has been observed over a distance of only 35 km.

The Subtropical Convergence lies in the vicinity of latitude ~40° S, in both the southeastern Atlantic and southwestern Indian Ocean; south of Africa, the zone may be pushed as much as 5° southward by the impact of the Agulhas Current. In general, the zone has a width of ~220  $\pm$  140 km. This wide range is a result of a high degree of mesoscale turbulence resulting from north–south shifts of the front, as well as eddy shedding to either side (west and east). Furthermore, as a result of eddy formation and the presence of an Agulhas Front, many different fronts may exist at any one time, each with a different gradient and intensity. A special feature of this convergence, particularly west of 70° E, where the north–south gradients are strongest, is the presence of enhanced primary productivity.

The atmosphere above the Subtropical Convergence Zone is ordinarily a cloudy region. To obtain a relatively cloud-free view from space, satellite observations over a long period combine all the cloud-free expanses. This has led to the belief that the whole Subtropical Convergence Zone has enhanced (higher than usual) primary productivity. Research ships passing the front have occasionally recorded high concentrations of chlorophyll a, but at other times have found no evidence of such high levels. Studies using satellite technology and modelling have shown that primary productivity at the convergence occurs in the form of highly specific phytoplankton blooms of relatively short duration and limited geographic extent. The blooms are largely aided by eddy formation where high nutrient loads, originating south of the front, and a high degree of vertical stratification are combined to create favourable conditions. Furthermore, from a biological perspective, the Subtropical Convergence Zone is not just a border between sub-Antarctic and subtropical ecosystems, but forms a distinct ocean ecosystem of its own,

with many endemic species of fauna. This is substantiated by assuming that most eddies that are shed at the front are eventually forced back and re-absorbed, thus continually reinforcing the front and not allowing much mixing or escape.

#### THE MONSOONAL SYSTEM

#### East African Coastal Current

The monsoonal system influencing the northern Indian Ocean involves of a complex set of ocean currents and water masses, all in some way affected by the seasonal monsoonal winds. Beginning in the south, the first current encountered is the East African Coastal Current, also known as the African Coastal Current or the Zanzibar Current. Fed by the South Equatorial Current (Figure 2), this is one of the few currents in the region that may be considered a permanent feature, even though it is affected by the seasonality of its source. During the Northeast Monsoon (December-March) the northward flow of the East African Coastal Current is opposed by the southward-flowing Somali Current. The boundary between these two currents begins near ~1° N at the beginning of the season, but shifts to ~4° S towards the end of the season. On meeting the opposing Somali Current, the waters of the East African Coastal Current then flow eastward into the Equatorial Countercurrent.

Where the two opposing currents meet, a large coastal upwelling system may form in the vicinity of 3° N, thus separating the currents along the coastline. As a result, two gyres are formed, a northern anticlockwise gyre carrying  $\sim 27 \times 10^6$  m<sup>3</sup>/s, and a southern clockwise gyre carrying  $\sim 22 \times 10^6$  m<sup>3</sup>/s, both in the upper 100 m of the water column. Phytoplankton from this upwelling area may be advected a long distance into the northern Indian Ocean by the Equatorial Countercurrent. The surface speeds of the East African Coastal Current may attain up to 2 m/s by the end of April, at which time the current flow volume reaches  $15 \times 10^6$  m<sup>3</sup>/s.

#### Somali Current

The inflow into the coastal currents of East Africa as well as their outflow into equatorial east–west currents varies with the monsoon season. The most important feature of the Somali Current is that it changes direction with season. During the Northeast Monsoon (December–March) the Somali Current flows southward along the Horn of Africa, yet south of 5° N the East African Coastal Current sets northward, as mentioned above. As the season advances the southward Somali Current strengthens and pushes increasingly farther south (up to ~ 4° S by the end of summer), its water deriving from the North Equatorial Current and flowing away from the coast in the Equatorial Countercurrent. The surface-water speeds of the Somali Current at the start of this monsoon season (in December) are 0.7–1.0 m/s, but by June the speed may already be as high as 3.5 m/s.

During the Southwest Monsoon (July–September) the Somali Current direction changes completely and flows northward, in contrast to all other currents along the African and Arabian coasts, thereby feeding directly into the East Arabian Current. South of 10° N the Somali Current is only ~150 m deep during this monsoon season, but its depth increases as the current moves northward into the Arabian Sea.

#### Arabian Sea circulation

The Arabian Sea fills the ocean basin to the west of the Indian subcontinent. Although called a sea, it has none of the characteristics of a regional sea such as the Red Sea or Persian/ Arabian Gulf, but forms an inherent part of the broader circulation of the northern Indian Ocean. Water movement in the Arabian Sea is likewise dominated by the monsoonal winds and is therefore strongly seasonal.

During November–January the Northeast Monsoon causes a weak flow of less than 0.2 m/s in the Arabian Sea. Along the western continental shelf of India a current sets northwestward, approximately 400 km wide and 200 m deep, but increasing in depth as it progresses. The East Indian Winter Jet injects fresher water from the Bay of Bengal, past Kanyakumari (Cape Comorin) at the southern tip of India, and into the Arabian Sea. In the north, cold winds from over the land cause convective overturning. South of 15° N, at ~65° E, the current flows westward, and by the end of this monsoon season the flow is clockwise in the entire basin.

By April the Southwest Monsoon starts and the northward Somali Current is formed, moving water northward along the Horn of Africa towards the Arabian Sea. By May the continuation of this current, the East Arabian Current, is formed along the Arabian Peninsula, and carries water at ~0.7 m/s northward, while water movement in the greater part of the basin is clockwise (eastward). During the Southwest Monsoon strong coastal upwelling occurs along the continental shelf of the Arabian Peninsula, accompanied by a decline in water temperature of ~5 °C, as well as at the western shelf of India.

The circulation patterns in the adjacent seas are very different, as discussed below.

#### Red Sea circulation

The salinity of the Red Sea is higher than the world average of the oceans, as evaporation vastly exceeds precipitation; a salinity as high as 42 may be attained in the northern limits

of the Gulf of Suez and Gulf of Aqaba. Its average depth is ~560 m, with a maximum depth of possibly 2 900 m, and a sill depth of ~110 m at the Strait of Bab-el-Mandeb, its entrance to the Gulf of Aden and WIO. The Red Sea's shape and its high salinity allow it to act as an inverse estuary. Inflow of ocean water occurs at the surface in winter and moves northward aided by seasonal winds. As this water moves northward, evaporation increases the salinities from averages of 36 to 40. In the northern reaches, the temperature of the surface water is 27-30 °C in summer, but decreases to 20 °C in winter when active convection takes place. In summer, deeper water begins to flow out over the sill and descends to a depth of 500-1 000 m in the northern Indian Ocean; here it has a temperature of 13–14 °C and salinity of >36.5. In summer the water is saturated with oxygen, but at such high temperatures the dissolved oxygen concentration is only about 4 ml/l (water with a salinity of 35 at 15 °C carries 8.5 ml/l oxygen). Waters with these particular characteristics are found in some form of dilution at intermediate depths throughout WIO, and water similar to Red Sea Water may correspondingly originate in the Persian/Arabian Gulf.

#### Persian/Arabian Gulf circulation

The Persian/Arabian Gulf is similar to the Red Sea in terms of area, and both are concentration basins. However, the Red Sea has a deep trough in the centre, whereas the Persian/ Arabian Gulf basin forms an inherent part of the continental shelf and has a mean depth of only 25 m. Water in the interior of the Persian/Arabian Gulf also has very high salinity (~40), and the inflow of freshwater from the Euphrates and Tigris rivers makes little difference to this. The Gulf, however, has no pronounced sill holding back deep, dense waters. Just outside the mouth of the Gulf, at the Strait of Hormuz leading to the Gulf of Oman, the slightly diluted water retains a high salinity (~38) and an oxygen maximum. Although Persian/Arabian Gulf Water has a lower density than Red Sea Water and therefore does not descend as deeply, at large distances from these sources it becomes difficult to distinguish between waters of the Red Sea and the Persian/Arabian Gulf.

#### CONCLUSIONS

The WIO region shows high diversity in physical and chemical conditions and ocean ecosystems. From its border with colder sub-Antarctic waters, and northward into tropical waters near the equator, a great spectrum of temperature, salinity, nutrient content, mixed-layer depths and environments can be expected. A vast amount of this ocean region remains as yet unexplored and unmeasured, particularly the southwestern areas, and the hydrodynamics are imperfectly understood. Moreover, the peripheral seas of WIO, the Red Sea and Persian/Arabian Gulf possess some unique features and extreme environmental conditions. In certain portions of WIO, but specifically in much of the equatorial expanse, the currents may be quite stable, whereas in other portions, such as at the Agulhas retroflection south of Africa, some of the greatest current variability in the world's oceans can occur. While many of this region's ocean systems are currently under intense investigation, linking what is learnt through physical and chemical oceanography to the life histories and distributions of fauna, particularly fishes, will present a challenge to researchers for some time to come.

#### GLOSSARY

advect - to convey a fluid by horizontal mass movement.

**convective overturning** – reversals of water masses with welldefined temperature and salinity regimes/characteristics.

**fluorescence** – the property of absorbing light of short wavelength and emitting light of longer wavelength.

**Indonesian Throughflow** – an ocean current important for global climate, which moves warmer freshwater from the Pacific to the Indian Ocean, where most of the water flows from north of the Equator, past the south of the Philippines, between Borneo and Sulawesi and south between Bali and Lombok, Indonesia, into the eastern Indian Ocean.

meridional – along a meridian or in a north-south direction.

**photic zone** – the surface layer of the ocean that is reached by sunlight, and hence where photosynthesis can take place (roughly to 80+ m depth).

**phytoplankton bloom** – a large and sudden increase in microscopic algae caused by an increase in nutrient content in an area, common in the world's oceans, and often changing the local colour of the water.

**salinity** – the measure of dissolved salt content in water, usually expressed as a ratio in parts per thousand (ppt); average ocean salinity is ~35 ppt (or 3.5%, or 3.5 g of salt per litre of water).

**silica** – silicon dioxide, a mineral used by marine organisms such as diatoms to build the cell wall.

spinning down - losing energy, slowing down and dissipating.

**thermocline** – an abrupt transition between two layers of water with typically large temperature differences, with the colder water beneath the warmer.

**volume flux** – a measure of water flow, where a particular volume (m<sup>3</sup>) flows across a unit area (m<sup>2</sup>) in a unit of time.

# THE ORIGINS AND GEOLOGY OF REEFS OF THE WESTERN INDIAN OCEAN

Charles RC Sheppard, Mebrahtu Ateweberhan, Nicholas AJ Graham and Bernhard Riegl

#### INTRODUCTION

The Western Indian Ocean (WIO) is a biologically coherent subdivision of the world's largest biogeographic province, the tropical Indo-Pacific. Its partial separation from the eastern portion of the Indian Ocean is largely a result of currents, great depths and substrate types, and not of land masses or to marked water temperature differences.

Reef types present in the WIO include all of Darwin's (1842) classic reef types: fringing reefs, barrier reefs and atolls. Added to these are numerous variations resulting from substantial vertical and horizontal movements of land related to plate tectonics, and to sea levels which have changed markedly over recent geological time. The area's two main embayments, the Persian/Arabian Gulf and the Red Sea, have very different reef conditions, which support fundamentally similar Indian Ocean marine biota, but with several characteristic and endemic species.

The distribution of substrate suitable for reef development is uneven (Figure 1), and is partly determined by depth and by a range of other factors which favour, or preclude, reef development (Sheppard 2000). One of the largest reef systems is the chain of reefs in the central Indian Ocean, most of which are classic atolls. Further to the west lie the granitic Seychelles and the volcanic Mascarene islands of Réunion, Mauritius and Rodrigues. Between and around these are numerous, extensive, submerged limestone platforms, sometimes called the Shoals of Capricorn, because they are located on the Tropic of Capricorn. These areas have thousands of square kilometres of reefal substrate lying within the photic zone, where water temperature is suitable for reef development. Further west of these shoals lie numerous groups of atolls, most of which are politically part of the Seychelles. Madagascar has very extensive fringing reefs, a very large barrier reef in the southwest, and numerous patch reefs and atollshaped structures dotted around it, mainly on its northwestern side. The shores of East Africa and its many islands have many variations of fringing and barrier reefs which extend southwards to South Africa.

Near the Horn of Africa, heavily sedimented shores preclude the development of extensive reefs, though reefs do occur in patches, and the Socotra archipelago is fringed by extensive coral reefs. In the Gulfs of Aden and Oman, cold water upwelling results in patchy reef development, though coral growth may be extensive in some areas.

Along the coasts of Pakistan and western India the marine substrate is mainly soft or heavily sedimented so that corals are

few and reef development is poor or absent. These conditions persist as far as southern India and Sri Lanka where there are some extensive fringing reefs (Sheppard 2000). Elsewhere there are extensive submerged limestone platforms that are separated by large expanses of deep ocean or, around the rim of the ocean, by extended stretches of muddy shallow water not conducive to reef life. These areas are still largely unexplored.



**Figure 1** Map of the western and central Indian Ocean. Blackshaded areas represent ocean areas that are shallow enough to support coral reefs; in such areas reefs may be prolific and abundant as in mid-ocean sites, or they may be largely soft sediment with scattered reefs in some continental sites, such as western India. Redrawn from Sheppard 2000

Burgeoning human populations, poor management and overexploitation of marine resources has raised deep concern over the resilience of reefs and their ability to recover from impact. About 70% of the region's reef systems have been either destroyed, damaged, or impacted to some degree (Figure 2), and the existence and integrity of these huge geological features that are key to supporting much of the marine life are profoundly threatened.



Figure 2 Pie chart of Indian Ocean coral reefs categorised into four main groups, from effectively lost to low threat and largely undamaged. Source: data from Wilkinson 2008



**Figure 3** Sea level during the past 140 000 years, showing that major fluctuations have occurred during this period. Especially important today is the low sea level stand about 14 000–20 000 years before present, when sea level was depressed nearly 150 m relative to the present day. Redrawn from Hopley 1982

#### **POST-HOLOCENE SEA LEVELS**

Present-day reef distribution depends on past distribution; modern reefs grow on old reefs where they can. During the last ice ages, sea level oscillated from a little higher than it is today to ~150 m below today's level, as a result of a series of warming and cooling episodes (Figure 3). Then, at the end of the last ice age, between 18 000 and 15 000 vears ago, there was a marked rise in sea level which continued for ~10 000 years. As sea levels rose, reef growth probably continued in the shallowest 20–50 m. Where there were brief still-stands, horizontal reef accretion increased, but where sea level rise was continuous and rapid, reefs simply grew upwards — some near their maximum possible rate until the sea level stabilised near its present level. This is, however, a very simplified description and clearly did not apply everywhere: much more reef substrate exists in ~5-50 m deep in the WIO than that which supports surfacereaching reefs today. This is illustrated by the well-mapped example from the Chagos Archipelago in the central Indian Ocean, where the area of reef substrate that remains submerged to several metres greatly exceeds the area which has reached the surface of the sea today (Figure 4) (Dumbraveanu & Sheppard 1999).

Area at each depth



Depth (m) below low water

**Figure 4** Area distribution from 0 to 100 m deep of a complex of coral atolls and banks in the Chagos Archipelago. Top line (red) represents the main atolls plus submerged banks and reefs; bottom line (blue) represents the five main atolls only. Source: Dumbraveanu & Sheppard 1999

### THE NORTHWEST INDIAN OCEAN AND PERIPHERAL (ADJACENT) SEAS

## Persian/Arabian Gulf, Gulf of Oman and Arabian Sea

The Persian/Arabian Gulf is a largely carbonate basin, where most substrate is <20 m deep (Figure 5). Only a narrow strip along its northern, Iranian, shore is deeper. The Arabian peninsula shoreline is geologically stable, while the Asian shoreline is unstable (Sheppard et al. 2000; Subba Rao & Al-Yamani 2000). During the Last Glacial Maximum (LGM), ~20 000 years ago, this Gulf was completely dry except for the channel which drained rivers such as the Tigris and Euphrates into the Indian Ocean. With the rise in sea level the Gulf flooded with water from the Indian Ocean, bringing with it an Indian Ocean biota. Today the marine climate is very hostile, being highly saline with enormous annual temperature ranges. These harsh conditions also create a kind of carbonate substrate which is rather uncommon in the Indian Ocean region: biogenic hard grounds of limestone dominated by microfauna and chemical depositions (Purkis 2011). While there is still reef growth, the diversity of limestone depositing species that can tolerate conditions in the Gulf is a quarter or less than that in the Indian Ocean (Figure 6), and these species become less diverse as salinity and temperature become more extreme.

Most of this vast limestone platform in the Gulf is thinly covered by carbonate sands, which form flat, featureless expanses with remarkably little relief, except for some raised mounds that resemble patch reefs. The summits of some of these are near low water level and, while several have rich coral growth, many have few corals because of the severe environmental conditions, and instead support large brown algae. These limestone platforms are formed in part by underlying salt domes forcing surface limestone upwards, a mechanism which also led to the creation of the archipelagos in the southern Red Sea.

Reef terraces are found at Bahrain and other small island groups such as the 10- to 15-m high islands off the Hawar Archipelago. Likewise, the small islands off Saudi Arabia probably also have uplifted limestone at their core, although these are now coral cays ringed by modern coralline algal and coral reefs that provide the most diverse hard substrate habitats known in the Gulf. The southernmost Gulf supports a complex mixture of limestone patches and sedimented areas colonised by seagrasses and algae, with extensive corals in patches. Large amounts of limestone sediments, dominated by foraminifera, form banks and shoals. The sediment becomes rapidly cemented to form extensive limestone crusts in both shallow subtidal and intertidal waters, the crusts sometimes dominated by algae or corals, or by both. The Iranian side is poorly known. It has a steep slope, supports reefs in patches and, given the general anticlockwise circulation in the Gulf, may be the source of most of the propagules of corals and other marine life in the Arabian or southern side.

The entire Arabian Peninsula has rotated through ~7° anticlockwise since the opening of the Red Sea (Sheppard et al. 2000). Northeastern Arabia is being pushed into the Asian plate leading to fjord-like, mountainous structures in the Musandam Peninsula, a remarkable feature jutting into the Straits of Hormuz, the entrance of the Persian/Arabian Gulf. Today the peninsula is a 90-km spur of limestone mountains and cliffs of Cretaceous origin, long recognised as being an unusual geological structure, whose linear coastline is probably nearly 900 km (Figure 7). The substrate is mainly reefal, built by algae, and later by scleractinian corals, whose layers are folded and tilted. This area is biologically important because it provides substantial rocky substrate where, for hundreds of kilometres on either side, the shore is formed by soft substrate largely uncolonisable by reef biota. The area is spectacularly beautiful and, on land at least, inhospitable because the steep-sided hills shut off any breeze. The phrase 'going round the bend' is said to have originated here: officials posted to the telegraph relay station round a bend in a fjord in the 1860s used to be driven to distraction by boredom and heat. Missing the marvellous opportunities to study the reefs, they gave this area the reputation of being the most oppressive place in the world.

The Gulf of Oman is sedimentary and the sea water has near-normal oceanic salinity. The northern shore, Balochistan, remains almost completely unstudied. Its southern shoreline in Oman is a gently sloping sandy bay over 300 km long. The Daymaniyat Islands (Figure 8) provide almost the only significant coral-reef substrate, and the reefs are among the best examples of reef formation in the Sultanate, but they are isolated, relatively tiny expanses of reef in a very large sedimented area. The mainland shore is composed mainly of various sandstone and dolomite outcrops along with limestones and metamorphosed basalts, interspersed with beaches. There are some reefs, including extensive monospecific stands of Pocillopora (Figure 9). At 2-10 m above sea level there are early Pleistocene corals, some being 'stacked bioherms' rather than true fossil reefs, in which layered coral communities alternate with coarse wadi deposits and storm debris, and the coral layers are not fully consolidated reefs. Semi-fossilised drowned reefs have also been discovered along a 20-m depth contour on this section of the northern coast of coast.

The Arabian Sea shoreline of Oman is poorly known. From Ras Al Hadd (the easternmost part of Arabia) the shore has large, sweeping sandy bays backed by low-lying dunes, salt pans and, towards the south, cliffs. Masirah Island has some fairly



Figure 5 Map of the Persian/Arabian Gulf region. This shallow sea has high salinity and seasonally both high and low water temperatures. © CRC Sheppard







**Figure 7** The Musandam Peninsula forming the southern part of the entrance to the Straits of Hormuz. This is a heavily indented coastline with steep, deep slopes that are very suitable for coral growth and which is unlike the coastline that extends for many hundreds of kilometres east or west of it. © CRC Sheppard



**Figure 8** The Daymaniyat Islands of Oman, being made of fossil reef and raised rock, provide hard substrate for coral growth which otherwise is very scarce in the Gulf of Oman. © CRC Sheppard



**Figure 9** Underwater in the Daymaniyat Islands there exist reefs, or more properly bioherms, made of vast expanses of single species of corals. This example is of *Pocillopora*; further east there are similar stands of *Montipora*. © CRC Sheppard



Figure 10 Off Masirah Island in the Arabian Sea are large almost monospecific reefs of the leafy coral genus *Montipora*. © R Baldwin



**Figure 11** Kelp and coral reefs. In upwelling areas such as southern Oman in the Arabian Sea, the benthic community is a mixed one of macroalgae characteristic of cold-water stands growth amongst corals. The kelp is *Ecklonia*, the tall alga is *Sargassopsis* (both genera of brown algae) and the coral is an *Acropora* species. © R Baldwin

well-known reefs with large expanses of *Montipora* (Figure 10). Midway down Oman the extensive dunes of the Wahiba Sands continue into the sea, precluding reef development.

Further south, rocky cliffs alternate with long stretches of littoral sand dunes and generally terminate near low water level. These areas are heavily scoured, so that the sublittoral of this high-energy coastline is predominantly a soft substrate environment below a few metres depth. Coastal rock is a complex mix of Tertiary shales, limestone and gypsum, with chert and marly bands interbedded with some igneous rock. Some remarkable locations occur where both seasonal and permanent stands of the Southern Hemisphere kelp Ecklonia and other related large brown algae form kelp forests amongst the coral communities, in a remarkable overlap between these two usually well-separated ecosystems (Figure 11). This is attributed to a seasonal, nutrient-rich, cold-water upwelling. This pattern continues along the Yemen coast, south to Aden, supporting patches of reefs, but mostly of corals veneering the underlying rock (Pichon et al. 2010).

The Socotra Archipelago lies on the Carlsberg Ridge. The southern rocky shores of the four main islands are steep and exposed to the full force of the summer monsoon, but the sheltered northern shores support significant coral communities between rocky outcrops dominated by macroalgae. The islands themselves lie on a broad and shallow, sedimented platform from which originates much of the sediment that controls the coral-reef communities. In some areas this accumulates into immense beaches (Figure 12).



Figure 12 A Socotra Island in the Gulf of Aden is part of Yemen politically but is closer to the African coastline. This group of islands provides significant hard substrate suitable for corals in this sea. B Socotra shoreline. © CRC Sheppard

#### Red Sea

The Red Sea and the Gulf of Aden, both rich in reefs (Figure 13) (Sheppard 2000), were formed by tectonic rifting of Arabia from Africa that started ~180 million years ago (mya) during the fragmentation of Gondwana. The separation formed the Afar Triple Junction (Figure 14) during late Oligocene-early Miocene, ~30 mya. This Junction forms the centre of a continuous rift between the Arabian, Nubian and Somalian plates. The Red Sea is considered to be a young ocean, its formation initiated by the magmatism that began ~31 mya. The trough started to form in the south-central part 25 mya (Bosworth et al. 2005), and was followed by a wave of volcanism throughout the entire Red Sea, from Afar and Yemen to northern Egypt. At ~20 mya the principal phase of shoulder uplift took place and water depth increased dramatically through the entire length of the sea. Rifting in the Gulf of Aden started earlier than in the Red Sea, as evident from the wider spreading. The rift connects directly to the worldwide system of oceanic plate boundaries via the Sheba Ridge in the Arabian Sea.



**Figure 13** Sketch of the Red Sea and Gulf of Aden areas that are rich in coral reefs (darker-grey shaded areas). The Red Sea is particularly deep, being formed by tectonic rifting of Arabia from Africa. It is more diverse in corals and fishes than many parts of the Indian Ocean.



Figure 14 Map of the Afar Triple Junction. https://commons.wikimedia.org/wiki/File:ATJ\_map\_%28color%29.jpg#filelinks

The Red Sea is connected to the Indian Ocean by the narrow and shallow Hanish Sill (at ~137 m depth) at the Bab-el-Mandab (the 'Gate of Lamentations', perhaps so named because of its stormy character). The Sea is also now connected in a minor way to the Mediterranean in the north by the Suez Canal. As a part of the great rift system which extends from eastern Africa, through the Gulf of Aqaba to the Dead Sea (Braithwaite 1987; Bosworth *et al.* 2005; Tiberi *et al.* 2005), the central depths of the Red Sea are both very hot and highly saline as a result of volcanic activity.

The region's climate is largely controlled by seasonal changes in atmospheric pressures (Edwards 1987), and weather variations are associated with the relatively large North–South gradient in atmospheric pressure. Much of the airflow to the Red Sea is from Northeast Trade Winds, and the mountain ranges along the sides of the sea ensure that the main wind systems blow predominantly along its length. The air near the sea surface is dry as a result of the desert and semidesert surroundings. Associated with the variation in wind direction and water circulation, the Red Sea can be separated into two parts, north and south of 20° N. Palaeoclimatic evidence from fossil and recent coral cores show that the northern part is influenced by non-stationary circulation regimes associated with the ENSO teleconnections. Before 1970 patterns were associated with strong Pacific–North Atlantic teleconnection; those after 1970 are related to ENSO conditions and weaker Pan Pacific–North Atlantic Circulation regimes (Rimbu *et al.* 2003; Felis *et al.* 2004). The southern Red Sea seems to respond entirely to the Indian Ocean monsoon system (Klein *et al.* 1997).

Thermoclines established by the warm conditions prevent vertical mixing and cycling of nutrients from deeper water, so that the main source of nutrients is incoming Indian Ocean water. This exchange is stronger in winter when planktonrich Gulf of Aden water flows into the Red Sea at the surface (Ponomareva 1968). Recent satellite observations, however, indicate higher chlorophyll *a* concentration in the south during summer months. The northern Red Sea, north of 18° N, is less affected by seasonal changes, and water movement is consistently southward throughout the year. It has lower nutrient levels, and surface chlorophyll *a* concentration is less than half that of the southern Red Sea.

The minimum water level in the early Holocene was close to the depth of the sill at the entrance. It is unlikely that the sill was exposed completely, so the Red Sea was probably not completely cut off (Siddal *et al.* 2003; Fennadez *et al.* 2006). However, the depth of the sill would have been very shallow so that water exchange with the Indian Ocean would have been extremely restricted. There is considerable evidence that the Red Sea became hypersaline during this period, and its principal biota may have been temperature- and salinityresistant micro-organisms (Siddal *et al.* 2003; Fennadez *et al.* 2006). Under such conditions reef growth would have ceased, only to be resumed when sea levels rose and water flooded in over the sill, thus lowering salinities.

As a result, the biota forms distinct northern and southern biogeographic entities. While popular accounts continue to assume that this sea is fringed with rich coral reefs throughout, the southern third is, in fact, relatively poor in reefs, and its sedimentary character favours macroalgae, seagrasses and mangroves more than reefs. Reef structure in this part of the Red Sea resembles that in the Persian/ Arabian Gulf more than it does that of the northern Red Sea (Sheppard et al. 1992). Reefs in the north commonly extend 60 m deep while many reefs in the south are limited to ~10 m, terminating on sandy bottoms (Loya 1972; Roberts et al. 1992; Sheppard et al. 1992). However, new investigations indicate equal or higher species richness of several groups, such as corals (JEN Veron et al. unpublished data), butterflyfishes, seaweeds and seagrass (Lipkin & Silva 2002; Ateweberhan & Van Reine 2005).

During the Pleistocene ice ages, reef growth in shallow water was spread over a vertical range of at least 100 m,



**Figure 15 A** Ages (bars) and elevations (line) of Red Sea reefs that align along both the African and Arabian shores. Present fringing reefs have surfaces at low sea level, while older, fossil reefs are found further inland on both shores at generally increasing elevations. **B** Photograph of the present reef flat which is submerged (left), while to the right is the youngest fossil reef elevated by 2–3 m. **A** Redrawn from Medio *et al.* 2000 **B** © CRC Sheppard

resulting in the development of several substantial limestone ridges. Today these form modest-sized barrier reef systems offshore, and present-day coral reefs are therefore built on and veneer older reef systems that were formed during earlier interglacial periods. Many reefs also veneer alluvial fans formed from outwashed terrestrial material, and in some areas, cores have revealed alternating layers of alluvial fan and reef superimposed on one another. Because of the tectonic uplift of the shores, and the widening of the Red Sea by several millimetres per year, raised fossil reefs are found on the shores in a series of different steps in which height is related to age (Figure 15).

While classic, steeply sloping, fringing reefs are the best known, several other forms occur too, many given different names of greater or lesser popularity. Reef construction will, however, occur wherever substrate and environmental conditions allow, irrespective of how we classify them, and in the tectonically active Red Sea where variation in substrate is considerable, it leads to a corresponding variety of reef types (Figure 16).

- 'Contour reefs' develop on sites where mountains submerge into the sea. These are usually thin, typically only 3–4 m wide, but may extend outwards as much as 1 km in embayments and old wadi systems.
- Barrier reefs have developed on older offshore limestone platforms, now lying some distance from land along depth contours of 50–200 m along old Pleistocene shorelines.
- Patch reefs typically occur where temperature and salinity and, most importantly, high sedimentation constrain greater coral-reef development in the lee of barrier or fringing reefs. These rise from sandy substrate at 1–5 m depth.

- 'Ridge reefs' were first described as longitudinal ridges lying along the axis of the Red Sea, their base being the result of normal faulting and underlying salt deposits moving upwards along these faults (Guilcher 1988).
- Sanganeb, off Sudan, has been described as the only atoll in the Red Sea.

The most northerly reefs of the entire Indian Ocean region are in the sedimented Gulf of Suez. The patch reefs on its eastern shore reach 1–3 m in height above the soft substrate. The Gulf of Aqaba is like a small-scale Red Sea. It has a 200-m deep sill at its entrance and supports continuous fringing reefs that conform to their rock basement, and include some of the most recreationally used reefs anywhere in the world. These are mostly narrow reefs, sometimes only 10 m from the beach, but in one or two areas they broaden to almost 1 km.

The southern part of the Red Sea is generally poor in reefs along the mainland shores, as a result of increasing sedimentation, shallowing bathymetry and higher nutrient levels which favour algae. The continental shelf becomes broad and fringing reefs become intermittent, being replaced in many places by sand, mud and mangroves (Figure 17). Reefs occur where a gradual subsurface slope has allowed them to extend considerable distances seaward. Here, a different type of reef, constructed by calcareous red algae, and dominated by algae can be found. These reefs seem to be analogous to oceanic coral-reef crests. They rise from sandy substrates at 2-4 m deep, support few corals and are typically covered in Sargassum, whose fronds form thick mats on the water surface. These coralline reefs provide extensive hard substrate in otherwise sandy areas. Offshore, the Farasan and Dhalak archipelagos support rich reefs which extend into deeper water.



**Figure 16** Variation in reef type is considerable within the Red Sea, from classic fringing reef (1) with reef flat and steep slope, through a range of patch reefs (2–4) to a few atoll-like structures (5). Sketch 6 shows two transects from shore to ~50 km to seaward that show numerous irregular pinnacles of coral growth and, in the right one-third of each trace, barrier reef structures. © CRC Sheppard



**Figure 17** In the centre and south of the Red Sea, mangroves become increasingly common. These form extensive habitats of soft substrate unsuitable for coral growth although seaward of them where water is clear, coral reefs again flourish. © CRC Sheppard

#### Pakistan, India and Sri Lanka

The shores of western India and Pakistan are largely sedimented, with little coral-reef growth, although corals may be found in a few areas with hard substrate (Bakus *et al.* 2000). One exception is in the Gulf of Kutch in northwest India, where corals are abundant and where rudimentary reefs occur.

Coral diversity shows a marked gradient along this coast, with ~40 species of reef-forming corals in the north, to 200 or more in the south. Curiously, there are no modern branching corals in the north, though semi-fossilised *Acropora* has been found (Bakus *et al.* 2000). Fringing and patch reefs occur especially between India and Sri Lanka. In general, the reefs of this coastline have been heavily degraded by human exploitation, including extensive coral and reef mining for construction.

Around Sri Lanka, fringing and barrier reefs occur close to shore within the 30-m depth contour. However, little research has been done, and for many years civil conflict has made the northern area inaccessible. Some areas are declared protected (e.g., Hikkaduwa Reef), but heavy human exploitation has severely damaged most reefs.

# Lakshadweep, Maldives and Chagos chain of atolls

The largest single chain of atolls in the Indian Ocean extends from the Lakshadweep (part of India and formerly the Laccadives), southwards to the Chagos Archipelago. This chain contains several hundred islands, and tens of thousands of square kilometres of coral reefs, formed by a deep hot spot under the Earth's crust. As India moved north with the break-up of Gondwanaland, this hot spot produced a chain of volcanoes over millions of years, and these eventually became seamounts and islands. Crustal movements later placed the hot spot under Mauritius, and today it lies under Réunion (Figure 18). Most reefs in this chain are classic atolls. The chain itself lies on the Ninety East Ridge. The island states on this ridge are commonly included in 'South Asia' in political terms, though ecologically they are part of the WIO.

Land area consists of mostly small islands on rims of coral atolls, invariably low-lying: most are less than 2 m above high tide. A typical atoll has a reef flat near present low water level and seaward reef slopes that are generally steep, plunging in steps or continuously to depths of hundreds of metres. Most atolls in the WIO have lagoons that are shallower than the depths to which sea level dropped just before the Holocene; most are between 10 and 20 m deep, and some as little as 2 m, while others reach as much as 60 m. Thus, even modest falls in sea level would expose the reef flats and much of a lagoon floor, greatly increasing the area of dry land. The sea level rise in the early Holocene thus greatly increased the area available for active reef growth.

Lakshadweep comprises 12 atolls with a total of 36 islands, with a surface area of just 32 km<sup>2</sup>, and about eight submerged banks and reefs. The islands are heavily populated and the people are substantially subsidised by India (Bakus *et al.* 2000). The extraction of coral rock and rubble has destroyed many of the shallow reefs, leading to severe disturbances (Bakus *et al.* 2000).

The Maldives are a double chain of 26 atolls with more than 1 200 islands, extending another 1 000 km southwards (Risk & Sluka 2000). Some atolls are several tens of kilometres in diameter, and many have islands and circular reefs called 'faroes' within the lagoon, perhaps formed from earlier episodes of sub-aerial erosion of existing limestone reef that was periodically exposed above water level (Figure 19). There is a gradient of atolls from north to south: "in the north they are broad banks, discontinuously fringed by reefs, with small peripheral islands and many faroes. To the south, atoll lagoons are deeper, faroes are rare or absent, and the atoll rim becomes stronger and more cemented" (Risk & Sluka 2000). As in the Lakshwadeep Islands, exploitation of marine resources, accompanied with heavy extraction of limestone rock, has caused severe ecological disturbances in several areas of the archipelago.



**Figure 18** Sketch of movement of the Earth's crust over the deep Indian Ocean hotspot, from the Deccan Traps in India to the present and youngest islands of the Mascarene archipelago, that result from the hot spot.



Figure 19 Maldives faroes. Vaavu atoll in the Maldives, Indian Ocean. The Maldives contain several atolls whose lagoons contain numerous annular reef structures called 'faroes'.

Google Earth, 2015. Data: SIO, NOAA, US Navy, NGA, GEBCO, Landsat Image, Image © 2015 Digital Globe

The Chagos Archipelago lies at the southern end of the Laccadive-Chagos ridge, in the geographical centre of the tropical Indian Ocean (Sheppard 2000). The archipelago consists of a limestone cap of a few hundred metres to a few kilometres thick which rests on volcanic rock (Francis & Shor 1966). The archipelago has five atolls with islands, and a greater number which are awash or completely submerged. Why some have islands, some are awash and others are drowned to 5 m or more is not known. The central feature is the Great Chagos Bank which is the world's largest atoll at 200 km by 100 km in area. The Bank is mostly submerged, but there are eight islands on its western and northern rims. Chagos contains the largest expanse of unimpacted reef in the WIO, as well as some of the most abundant with life. With the exception of part of Diego Garcia, the reefs have been undisturbed for at least 40 years.

The five atolls with islands and the great number of submerged atolls and reefs, all have profusely growing corals (Figure 20). Those reaching the surface, including Blenheim Atoll which is entirely awash at high tide, have the largest wave-resistant algal ridges and spurs in the Indian Ocean (Figure 21). Exposure in some areas is such that even lagoonal reefs support small spur and groove systems. Oceanic swell is substantial in Chagos, even in periods of relative calm. On some large deeper reefs, such as Speakers Bank and the Great Chagos Bank, corals and seagrasses are profuse, indicating high benthic productivity. The Great Chagos Bank, the largest system of all, has a complex pattern of reefs that have several ring-shaped structures within the main atoll rim, which attest to a complex past history of growth and erosion. Lagoonal faroes, such as those found at the Maldives, do not occur here.

The 1998 El Niño event warmed Chagos and other coral islands in the WIO so severely that coral-reef mortality was substantial. An investigation into the speed of recovery from this event, and of its chemical contamination, reef condition, abundance of corals and fish, made Chagos the least contaminated reef site in the Indian Ocean. In 2010, the United Kingdom declared the archipelago a no-take marine protected area out to the 200-nautical-mile limit, but recently that declaration was found to violate international law, and Mauritius claims legal rights over the archipelago.





Figure 20 Rich and diverse coral cover on reefs of the Chagos Archipelago, central Indian Ocean. A Ocean-facing slope, B lagoon slope. © CRC Sheppard





**Figure 21** Spur and groove structures that are found on the edge of many high-energy reefs in the Indian Ocean. These structures are essential to the survival of such reefs. They are made by a calcareous red algae which is particularly hard and resistant to erosion and which can survive high temperatures when exposed at low tides. Image on the left is an underwater scene showing both the calcareous red algae and a general absence of corals in this high-energy location. © CRC Sheppard

#### Seychelles

The Seychelles consists of 41 tall granitic islands and 74 coralline islands with a total land area of 455 km<sup>2</sup>. Despite this small area, the country has an Exclusive Economic Zone of almost 1.4 million km<sup>2</sup>, and extensive coral reefs (Jennings et al. 2000). The archipelago can be broadly split into the 'inner' granitic islands to the north and the 'outer' coralline islands and atolls to the south and west. The inner islands contain most land, the largest islands being Mahé, Praslin and La Digue, where 97% of the people live. These are high granitic islands on the Seychelles Bank which broke away from the ancient supercontinent of Gondwanaland ~135 million years ago. This Bank, ~43 000 km<sup>2</sup> in area, is of Precambrian origin, dating to ~650 million years. Mean water depths over the Bank are between 44 and 65 m, and the islands rise to 914 m above sea level (Figure 22). The Amirantes Bank of ~6 300 km<sup>2</sup>, lies to the south and is built on a base of basalt with 13 km of granite above, and is capped by a carbonate sediment layer of ~500 m thick. Low-lying coral islands of sand, such as Bird Island, have formed several metres above sea level on both these banks. Further to the southwest lie higher limestone islands forming the atolls of Aldabra, Assumption, Astove, Cosmoledo and St Pierre. Like other atolls, these are formed on the basalts of old volcanoes that emerge from depths of over 2 000 m, and now rise to 8 m above sea level.

The reefs of Seychelles fall into three categories: fringing and platform reefs and atolls. The fringing reefs are most welldeveloped around the granitic islands of Mahé and Praslin, covering 20 and 27 km<sup>2</sup> respectively, whereas such reefs



**Figure 22** The central high islands of the Seychelles are unusual in being granitic, and are fringed with coral reefs. These are near Baie Ternay, Mahe. © CRC Sheppard

occupy only 10 km<sup>2</sup> in the outer islands. These reefs typically only extend down to 10 m, and sometimes to 20 m. The most extensive and continuous reefs off Mahé are found on the more exposed eastern side of the island, whereas those to the north and western sides are more patchy and broken up, with most coverage in sheltered bays. The fringing reefs off Praslin are large and can extend up to 3 km offshore. Granitic outcrops in the sea also commonly have corals growing on them. The 1998 coral-bleaching event heavily impacted the reefs of the inner Seychelles, killing off nearly 90% of the corals (Goreau *et al.* 2000). Recovery is very slow and, with many of the dead reefs eroding, the diversity of reef fishes has declined (Graham *et al.* 2006).

Platform reefs cover ~200 km<sup>2</sup> and can be found in the Amirantes and Farquhar groups. These reefs either form shallow lagoons and reef areas with very little emergent land, such as the African Banks, Île Plate and Providence, or raised platform reefs, reaching 8 m above sea level, such as Assumption and St Pierre islands.

The atolls in the Seychelles may be low-lying or raised. Low-lying atolls include Farquhar, St Joseph, St Francois and Alphonse with a total reef area of some 20 km<sup>2</sup>. Raised atolls include Aldabra, Cosmoledo and Astove, with ~200 km<sup>2</sup> reef area.

#### The Mascarenes

The Mascarene basin is over 4 000 m deep, and is bordered on the east by the Mascarene Plateau, which contains the banks of Saya de Malha (28 350 km<sup>2</sup>), Nazareth (7 625 km<sup>2</sup>), St Brandon Shoals (1 208 km<sup>2</sup>), Soudon and Rodrigues (1 688 km<sup>2</sup>). The largest islands are Mauritius (8 my old), Réunion (2.1 my old) and Rodrigues (1.3–1.6 my old) (Naim *et al.* 2000). They are all volcanic in origin and were formed by the same hot spot that produced the Lakshadweep, Maldives and Chagos archipelagos.

Mauritius has a 200-km long coastline surrounded by 243 km<sup>2</sup> of lagoons, fringing reefs and barrier coral reefs. Réunion only has 10–12 km of fringing reef, located on the southwest. Reef initiation at Réunion is likely to have kept pace with rising sea levels from 8 000 yrs BP. Rodrigues is situated on a shelf of 950 km<sup>2</sup>, and is enclosed by a large fringing reef and lagoon complex totalling ~200 km<sup>2</sup>. This complex is up to 13 km wide to the south. St Brandon Shoals consists of 55 low-lying sand cays and islands, with a total reef area of 190 km<sup>2</sup>.

The extensive Shoals of Capricorn banks situated between the Mascarenes and Seychelles consist of carbonate sediment overlaying a thick band of granite on a basalt base. Very little is known about the ecology of the banks, but reports indicate large sandy areas interspersed with coral outcrops and areas dominated by algae (Turner *et al.* 2000).

#### The Comoros

These are located at the northern end of the Mozambique Channel, between Madagascar and mainland Africa and consist of four main islands: Grande Comore, Moheli, Anjouan and Mayotte. They are all isolated volcanic seamounts increasing in age toward the east. Grande Comore is an active volcano with limited fringing reef growth. Moheli and Anjouan both have extensive fringing reef growth, and Mayotte, the oldest island, has patch reefs, fringing reefs, a barrier reef that extends for 197 km, and a rare double-barrier reef system (Quod *et al.* 2000).

#### Africa and Madagascar

Africa and Madagascar both have huge continental shelves where the slopes are much more gradual compared to atoll slopes, especially along the coast of Africa. Rising and falling sea levels, together with the resulting extensive movement of the coastline have influenced both the nature of the reefs and their extent. In several areas tectonic rise and fall of the shore has added to these effects, modifying the patterns greatly. Many patch reefs located midway on shelves probably lie on substrate that lay in water much shallower than today, their growth matching rising sea levels. Chains of present-day patch reefs are situated on top of old reefs which used to line palaeoshores, with some excellent examples in Somalia.

#### PRESENT-DAY GEOMORPHIC FEATURES

The coast from South Africa to Central Mozambique is mostly composed of unconsolidated Quaternary to Recent sandy sediments, interspersed with heavier-textured alluvium at river mouths. The considerable deposition and transport of sediments has caused significant seaward growth of the shoreline in central Mozambique. North of small areas of Tertiary basalt at ~16° S, heavily-faulted Cretaceous to Tertiary sediments line the coast of northern Mozambique (Tinley 1971; Rodriguez *et al.* 2000).

The area from northern Mozambique to ~8° N in northern Somalia consists of the top of a Pliocene–Pleistocene sequence of sedimentary deposits, the result of aeolian and biogenic processes. A combination of these processes, sealevel fluctuations, Holocene climate changes and tectonic movements produced the modern coastlines characterised by intertidal reef platforms, beach cliffs and rocky platforms. There are deeper platforms at depths below 8 m and 35 m.

These coastal areas also have extensive limestone coral cliffs and numerous coralline islets formed during the

Pleistocene that became exposed by a subsequent drop in sea level. Some exceptions are Mafia Island in central Tanzania, which originated during tectonic uplift of the continental shelf, and Pemba Island located in northern Tanzania, which broke away from the mainland ~10 mya along the Pemba Rift (Kent *et al.* 1971; Gallup *et al.* 1994; Ngusaru 1997). Pemba is separated from the continental shelf by a 400-m deep channel. Palaeoecological investigation of the rocky cliffs and platforms suggests similar ecological conditions as the present, with transition from early colonising forms to *Acropora* dominance at intermediate stages, and to the dominance by massive forms, e.g. *Porites* and faviids, at the final stage. A comparison of gastropods between Pleistocene and modern reefs shows 1–21% loss in the number of species (Crame 1980, 1981, 1986).

# Coastal habitats of East Africa and Madagascar

The continental shelf along most of East Africa is narrow, averaging 8–10 km wide, except around some of the archipelagos. Coral reefs, seagrass meadows, algal beds and mangrove forests are found along the entire coastline. Deepwater channels, river outlets and bays break the dominantly fringing coral reefs in places. High sedimentation and low salinities restrict coral-reef development near river mouths to a few offshore islands. Luxuriant stands of estuarine mangroves occur at the mouths of the major rivers: the Limpopo, Zambezi, Tana, Juba and Shebeli rivers, and the Rufiji Delta. The Rufiji Delta supports the largest mangrove forest area in East Africa at ~100 km<sup>2</sup> (Darwall & Guard 2000). Many of the shallow waters and bays along archipelagos and islands support luxuriant growth of coral reefs, seagrass beds and mangroves.

The reefs of South Africa represent the southernmost distribution of coral reefs in the WIO (Reigl *et al.* 1995). The southward-flowing Agulhas Current creates warm subtropical climatic conditions that are suitable for reef development. The reefs are, however, not well-developed, lack well-defined zonation and have a modest coral diversity of 143 species in 43 genera. Environmental conditions do not appear to limit coral growth and low abundance seems to be the main reason for the low accretion. Overall, soft corals dominate the reefs, though stony coral cover can reach as high as 63% in some places (Schleyer 2000).

Mozambican reefs are among the least studied, but include some of the highest coral diversity in the region – Pemba Island reefs, for example, have 208 species in 51 genera. Here the coast is divided into three regions. North of the Rovuma River (17°20' N), extending for ~770 km, are the most extensive, welldeveloped reefs in the country. The barrier reef surrounding Primeiras and Segundas Islands that form the Quirimbas Archipelago is well known for its diverse coral-reef life. The central coast is an area of many rivers and reef development and is limited by low salinity, high turbidity and sedimentation. The area, however, supports well-established mangrove stands. The area south of Bazaruto Island (26°50' S), the third region, is characterised by a patchy distribution of reefs.

The reefs of Tanzania are amongst the best developed and, except near river mouths, are widely distributed. Coral diversity is high, with 200-280 species in 50-60 genera (Johnstone et al. 1998; Obura 2003; Obura & Machano 2004; Obura et al. 2000, 2006). The Tanzanian coastal area is broadly divided into northern and southern regions. The area south of Dar es Salaam consists of complex coastal ecosystems of estuarine, mangrove, coral-reef and marine-channel habitats (Darwell & Guard 2000). The waters of Mafia Island, the Songo Songo Archipelago and Mtwara-Msimbati are well known for their well-developed reefs with coral cover (in the early 2000s) of 50-60%. The northern section includes the island of Pemba and Zanzibar and the mainland coasts of Tanga, Dar es Salaam and Bagamoyo. A fringing reef along the mainland coast is broken by bays and estuaries, with many patch reefs adjacent to the continental shelf. The reefs of Tanga are among the best along the Tanzanian mainland coastline (Horrill et al. 2000). There is also a well-developed fringing reef along the seaward sides of islands near Dar es Salaam that make up the Dar es Salaam Marine Reserve System. These are separated from the mainland by a 3-km lagoon, where coral cover reaches ~70% on back reef edges at 2-4 m depth, and where the dominant species are adapted to high turbidity. Reef development around Zanzibar is variable. There is some patchy fringing reef on the northern and eastern coasts, but Mnemba Island, north of Unguja, has a well-developed reef (Horrill et al. 2000)

Kenya has well-developed reefs along its southern coast, whereas reef development is less extensive in the north mainly because of freshwater influx. Most older studies on Kenyan reefs have concentrated on the Malindi-Watamu National Marine Park, but recent investigation has included sites further south, near Mombasa and Kisite Island, near the border with Tanzania (McClanahan & Shafir 1990; McClanahan 1994, 1997). Reefs in the north still remain the least studied (Obura & Church 2004). Coral species diversity in the south is estimated at 200 species in 55 genera (Obura *et al.* 2000). Generally, the reefs are characterised by dominance of poritiid, acroporid and faviid species arranged broadly into four depth zones (Obura *et al.* 2000). There seems a clear separation of the coral community of Kisite Island which resembles more northern Tanzanian reefs compared to other Kenyan reefs (McClanahan *et al.* 2007).

Although Somalia has the longest national coastline in Africa with an estimated shelf area of 32 500 km<sup>2</sup>, cold water upwelling and freshwater influx restricts reef development along most of the coast (Carbone & Accordi 2000; Carbone et al. 2003). Well-developed fringing reefs are found in the Bajuni Archipelago, north of the border with Kenva. Fleshy macroalgae dominates most hard substrate, while Thalassodendron seagrass beds dominate the shallow soft substrate. Benthic communities of typical mobile sandy substrates are limited to beach ridges and shoals. Coral carpets of dense assemblages of massive and encrusting corals are found along with crustose corallines, soft corals, sponges and Halimeda in some areas where mobile sandy cover and Thalassodendron meadows are missing. Most mangrove forests of Somalia are in the south, although some isolated stands are found in the northern coast. In a few places wide salt flats occur along some channels.

Madagascar has probably the most variable reefs in the region because of its diverse oceanographic provinces. The main coral-reef areas are located in the northeast (417 km), north and northwest (578 km) and southwest (458 km in extent) (Gabrié et al. 2000). The northern and northwestern part of the country have the highest coral species diversity in the WIO, with 318 species in 62 genera (Pichon 1978; McClanahan & Obura 1998; Veron & Turak 2005). However, information on environmental conditions, reef development and ecology is still very limited (Gabrié et al. 2000; McClanahan et al. 2009). Reefs in the southwest are probably the most developed, with fringing reefs, barrier reefs and reefs with sand cays. The Grand Recif of Toliara is the best known; the mid-west is poorly known, although with high sedimentation from river discharge, reef development is probably very limited (Gabrié et al. 2000). There are isolated reefs at the Barren Islands in the south and Pracel Shoal in the north, where there is also a submerged barrier reef further offshore. Reef development is better further north where the submerged ancient reef extends 10-60 km towards the open sea and supports 100% coral cover dominated by Pachyseris speciosa on the outer slopes (Gabrié et al. 2000). Except near estuaries and bays, fringing reefs surround the islands and line the mainland coasts of the northwest. Fore-reef areas are dominated by Acropora, back-reef areas by massive Porites, and lagoons by seagrass beds. The northeastern reefs of Masoala are less well-known, but modern coral growth dominated by Acropora borders the exposed ancient reefs.

### PRESENT-DAY ENVIRONMENTAL CONSTRAINTS TO REEF GROWTH

Reef growth and size depends to only a very minor degree on the number of species of the main reef-building organisms, corals and coralline algae. Low-diversity areas can produce substantial reefs, just as high-diversity coral areas may produce small, weakly developed reefs because of various factors involved in limestone deposition. Thus, except at extreme boundaries of the WIO reef province, such as South Africa, there is probably very little correlation between coral diversity and reef development.

Environmental constraints include temperature (both high summer and cold winter temperatures), salinity, sedimentation and nutrients. Preferred temperature ranges of most corals have been understood at least since Darwin, and for most – but not all – species the lowest survivable temperatures are from 15–18 °C, to a high of ~30 °C. Several species tolerate greater extremes. Table 1 shows the species in one or two locations in extreme areas around Arabia, and the temperature ranges they survive. Increasing salinity likewise results in declining numbers of coral species (see Figure 6). Similarly, freshwater from rivers impedes coral growth so that reefs do not occur near estuaries, a major constraint along parts of the African shoreline. This is also true of the northern Arabian Sea.

The Arabian Sea has exceptionally high oceanic productivity, especially adjacent to the coast of Oman, where seasonal upwelling between May and September brings nutrient-rich, cool water to the surface, impeding reef development. Here, although the diversity of corals exceeds 100 species, sometimes occurring in thick stands which have the superficial appearance of a reef, most are not truly accreting corals. Instead, the corals are attached directly to sublittoral and non-limestone rock. In such marginal reef areas, macroalgae develop which, visually at least, are as dominant as the corals.

Reef development is generally associated with waters of poor productivity, and in the Indian Ocean productivity ranges widely. Expressions like 'stressful environment' or 'increasing stress' are meaningless, since stress for any one group of organisms may be inconsequential or even desirable for another. For reef-building organisms increasing stress refers to an environmental gradient along which, at some point, corals can no longer live. Severe stress kills off reef-building organisms and hinders reef development. Table 2 shows physical characteristics of reef and other hard substrate areas where corals live but where reefs do not develop and where fleshy macroalgae dominate much of the shallow substrate along with corals.

Along an environmental gradient, the demise of reef growth may take place at quite different points according to prevailing combinations of temperature, salinity, nutrients and other factors such as exposure to severe waves and sedimentation. While we now understand that reefs do not simply develop because of coral growth, we still do not really understand the main mechanisms of reef growth which require explanations that incorporate microbial and micro-chemical processes.

 Table 1
 Coral species that survive temperature extremes of the range indicated (data from Sheppard 2000; Sheppard & Loughland 2002).

Temperature fluctuation (°C) 8–15	15–20	20–28	30
Acropora horrida>			
Stylophora pistillata>			
Porites nodifera>		Sargassun	i only
Cyphastrea microphthalma		·····>	
Siderastrea savignyana		>	
Porites compressa	>		
Platygyra daedalea	·;	>	
Porites lutea	>		
Psammocora contigua		>	
Pavona varians	>		
Coscinaraea monile		>	
Leptastrea purpurea		->	
Favia pallida	>		
Favia speciosa	>		
Favia favus	>		
Favites pentagona	>	>	
Turbinaria crater	·····>		

**Table 2** Physical characteristics of reef and other hard substrate areas where corals live in conditions where reefs do not develop and where fleshy macroalgae dominate some or all of the shallow substrate. This excludes coralline algal ridges and reef crests made mainly from encrusting red algae (from Sheppard 2000).

REEF/ALGAL CHARACTERISTICS	RAISED SALINITY	HIGH SUMMER TEMPERATURE	LOW SUMMER TEMPERATURE	LOW WINTER TEMPERATURE	RAISED SEDIMENTATION	RAISED NUTRIENTS	AREAS
Red algal patch reefs covered with <i>Sargassum</i> , very low coral cover	No	Yes	No	No	Yes	Yes	Southern Red Sea
High to moderate coral cover, <i>Sargassum</i> dominated reef crest but coral domination on reef slope	Slight, 36 to 40 ppt	Yes	No	No	No	Yes	Central, southern Red Sea
Limestone domes, mainly devoid of corals, in 1–5 m deep	Yes, > 40 ppt	Yes	No	Yes	Yes	No	Central and eastern Arabian Gulf
Reef flats at 0–1 m below low-water springs, high- energy areas, abundant algae on reef flats, but high coral diversity deeper	No	No	No	No	Yes, some areas	Yes, some areas	e.g., Western Sri Lanka
Upwelling areas, mainly dominated by <i>Fucales</i> but including <i>Ecklonia</i>	No	No	Yes	No	No	Yes	Oman to Somalia, <i>Ecklonia</i> in southern Oman, possibly also Yemen
Simplest 'corals-on- corals' mode of reef development	No	No	No	No	Yes	Yes, some areas	Embayments, these are notably formed by <i>Porites</i>

#### INDIAN OCEAN REEFS IN THE FUTURE – THE ANTHROPOCENE

Given that the limestone foundations of reefs are hundreds of metres thick, and have persisted for millions of years, it is tempting to imagine that coral reefs are timeless and immutable. But their history has been punctuated by long episodes when they did not grow at all, and this may be happening again today. Reefs are constructed for the most part by living organisms which have undergone enormous stress in the past. Noticeable declines in coral reefs were first noticed more than a century ago (Gardiner 1936). By the 1960s it was commonly asserted that traditional (mostly subsistence) fishing was centuries old, implying they made little or no impact on an ecosystem, but subsistence fishing was soon realised to be no less destructive of marine habitats than subsistence farming was of land (Ray 1969). In many countries many reefs have been reduced to rubble as a result of destructive fishing, amongst other factors (Salm 1983).

Reefs are extremely productive environments (Table 3) and essential to human well-being. The poor condition of at least two-thirds of the reefs of the Indian Ocean (see Figure 2) is probably surprising to those who encounter them only as holiday destinations which, naturally, are located at the best places. It is difficult to see how an environment conducive to reef growth can continue when runoff of pesticides and soils from poor agricultural practices continues to increase, and where overfishing persists. More than 30 years ago it was already known that many tons of topsoil per hectare per year was being washed into the WIO as a result of poor agricultural practices, and that the total exceeded 500 million m<sup>3</sup> per year. This has obliterated many coastal reefs, effectively converting them into soft substrate habitats. Finn (1983) summarised this: "this is not a pretty picture. It will be extremely difficult for the region's governments to confront such large scale and diffuse problems."

Since then the picture has become even uglier. It is estimated that human population will increase by another

three billion between now and the middle of this century. About half of these people will be in countries bordering or in the Indian Ocean. Pressures on reef resources will increase regardless of any improvements in reef management. Climate change, which will cause further coral bleaching and ocean acidification, is expected to interact with other human-induced disturbances. The WIO, particularly the central–northern half, suffered one of the highest global coral-reef mortalities during the strong 1998 ENSO event.

Although regional average sea surface temperature (SST) is increasing by a little more than 0.1 °C per decade, the frequency and intensity of extreme temperature events are expected to increase further, resulting in recurring bleaching events and extinction of coral-reef organisms. Such fundamental shifts may cause negative feedback loops, weaken the critical role of scleractinian corals as reef ecosystem engineers and their major ecosystem functions and services, including biodiversity, fisheries and tourism.

The consequences to coral reefs are dire, to the extent that living coral reefs will quite possibly become the first major earth habitat to become functionally extinct. Coral reefs create a large proportion of the hard, sublittoral substrate in tropical seas and, with increase in ocean acidification, coral bleaching and diseases, this substrate itself is turning rapidly from being biogenetic structures to erosional ones, further threatening biodiversity, and ecosystem goods and services that are critical to the many, largely poor, communities. Indeed, since the first draft of this chapter, further ocean heatwaves have severely reduced the coral cover on reefs in this region.

**Table 3** Values for productivity of Indian Ocean systems, from numerous sources (Sheppard 2000). Some of the generalised data are from outside the Indian Ocean, for which much less data exist than for the Pacific and Caribbean. Values are  $g C/m^2/d$ .

	PRODUCTIVITY (			
SYSTEM	COMMON OR AVERAGE VALUES (g C/m²/d)	EXTREME RECORDS (g C/m²/d)	P/R RATIO*	
Reefs				
Whole reef systems	3.2-4.0	2.3–6.0	Usually ~1	
Outer slopes	2.0–7.1		0.7-1.1	
'High activity areas'	9.0–14.0	8.0-23.0	0.6–1.7	
Reef flats, lagoons	3.0–7.0	2.9–19	0.7–2.5	
Corals with zooxanthellae	2.63			
Sandy reef areas	0.9–1.5	0.6–2.7	0.6-1.1	
Mangroves				
Global averages	6.0–15.0		1.2	
'Dwarf' in Red Sea	<6.0			
Seagrasses				
Global averages	6.0–15.0		1.2	
Thalassia, Syringodium, Lakshadweep	5.8			
Northwest Indian Ocean	3.0			
Algae				
Turfs and pavement	2.0–7.0	1.0-14.0	0.5-13.7	
Coralline algae	0.8–1.0	7.0	1.3–1.4	
Sargassum belts	60	(dry organic wt)		
Tropical kelp (Oman)	4.0	(dry organic wt)		
Halimeda beds	0.2–6.0			
'Mixed algae' Lakshadweep	1.9			
Cyanophytes	0.62-1.39			
Endolithic algae (Ostrobium)	0.4–0.6			
Benthic diatoms (sand, 5 m)	0.41			

\* P/R Ratio is ratio of production to respiration

	PRODUCTIVITY (			
SYSTEM	COMMON OR AVERAGE VALUES (g C/m²/d)	EXTREME RECORDS (g C/m²/d)	P/R RATIO*	
Bacterial				
Pelagic	0.007–0.11			
In sediments	1.2			
On coral rubble	0.01–0.1			
Planktonic				
Open ocean	0.04-0.06	0.003-0.4	0.3-1.4	
Arabian upwelling, Southwest Monsoon	1.16			
Arabian upwelling, Northeast Monsoon	0.23			
Arabian Sea, central	0.12-0.76			
East Africa, Southwest Monsoon	0.83			
East Africa, Northeast Monsoon	0.42			
Bay of Bengal	0.21			
Atoll lagoons	0.10-0.42			
Near shore or reefs	0.2–0.9	0.1-1.0		
North Red Sea	0.21-0.50			
Central Red Sea	0.39			
Southern Red Sea	1.60			

\* P/R Ratio is ratio of production to respiration

#### ACKNOWLEDGEMENT

GLOSSARY

We are grateful to Oxford University Press for permission to use material from the volume: McClanahan TR, Sheppard CRC, Obura DO (eds). **2000**. *Coral Reefs of the Western Indian Ocean: Ecology and Conservation*. Oxford University Press, Oxford, 522 pp.

aeolian - biogenic	- relating to the action of wind; wind-blown. – produced by living organisms	<b>palaeoclimatic evidence</b> – what geological information can tel about climates thousands to millions of years ago.		
<b>ENSO</b> – El Niño–Southern Oscillation; a recurring climate pattern involving changes in the temperature of waters in the central and	<b>photic zone</b> – the depth of water in which photosynthesis takes place.			
eastern tr	eastern tropical Pacific Ocean.	<b>propagule</b> – a vegetative/living structure that becomes detached from a parent to give rise to a new organism; an agent of reproduction		
<b>ENSO teleconnections</b> – the link between what happens in the Pacific Ocean during an El Niño event and what happens in the Indian Ocean.	e.g., a sucker on a plant or the eggs fishes or corals shed into the wat			
	scleractinian corals – stony or hard corals that form coral reefs today			
<b>hypersal</b> than 35 p	<b>ine</b> – with salinities higher than that of sea water, i.e., more arts per thousand (ppt).	<b>thermocline</b> – the boundary between two layers of water with large differences in temperature, the colder water beneath the warmer.		
monospe	ecific – composed of a single species.			

#### Links with other chapters in this volume:

The oceanography of the Western Indian Ocean Socotra and islands in the eastern Gulf of Aden: their fishes and fisheries

### SOCOTRA ARCHIPELAGO: FISHES AND FISHERIES IN THE EASTERN GULF OF ADEN

Uwe Zajonz

#### **INTRODUCTION**

The Socotra Archipelago, part of Yemen, lies in the northwestern Indian Ocean, at the Horn of Africa. It is globally recognised for its outstanding biodiversity and endemism, the reason why the entire island group was designated a UNESCO World Heritage Site in 2008 (Scholte *et al.* 2011; Van Damme & Banfield 2011). While Socotra's recognition as the 'Galapagos of the Indian Ocean' often alludes primarily to the diversity of its terrestrial ecosystems, the marine ecosystems are also highly diverse and unique in several ways (Cheung & DeVantier 2006).

#### HISTORY

The time of the first colonisation of Socotra by earlier settlers is not known, but dates at least from the first millennium BCE, according to earliest archaeological evidence. The origins of the first human population also remain unclear, but are likely to have been tribespeople from South Arabia, and possible occasional arrivals from Africa.

Recent discoveries of writings in several languages tell of the diverse origins of the people who used the islands as a trading base between the 3rd century BCE and the 6th century CE. There is a tradition that the islands and their inhabitants were occupied by Greeks sent by Alexander the Great around 330 BCE, and subsequently visited by Thomas the Apostle (in 52 CE), Marco Polo (c. 1293 CE) and the Chinese fleet of Admiral Zheng He (1420 CE). Around 50 CE Socotra likely belonged to the Hadramaut state of Eleazos. The Arab geographer Abu Mohammad al-Hasan al-Hamdan recorded in the 10th century CE that most of the inhabitants were Christians. Between 900 and 1300 CE the islands were notorious as a base for pirates. From about 1200 CE, and for several centuries, Socotra gained strategic importance for maritime trade and transport. After a brief conquest of Socotra Island by the Portuguese in 1507 as a strategic base to control sea traffic from the Red Sea en route to the Indian Ocean, it was abandoned four years later, and the archipelago came again under the control of the Mahra Sultanate of Qishn and Socotra, established early in the 16th century, and included what is now eastern Yemen and the

island of Socotra. It thus became finally Islamised (Islam may have gradually taken roots already from 700 CE onwards). In spite of strong Omani and European interest to take control of Socotra, the Mahri sultans maintained more or less continuous power over Socotra until the mid-18th century. From around 1600, however, the islands were on the maps of especially British and Dutch trading fleets for replenishment of water and stocking of local produce such as aloe. In the first half of the 18th century the building of the Suez Canal prompted European powers, Britain, France and Austria, to compete for the control of the Gulf of Aden, including Socotra. In 1886 the islands finally became part of the British Protectorate that included Aden and the Mahri Sultanate (while locally the Sultanate retained power), and remained so until the British withdrew from Aden in 1967. Soon after, the islands became part of the socialist People's Democratic Republic of Yemen (PDRY, commonly South Yemen). Socialist policies resulted in an initial wave of development in Socotra during this period that included the building of roads, schools and clinics. It also came under the influence of the Soviet Union as a result of the PDRY's political association with the latter which impacted on, e.g., its military and scientific activities. In 1990 Yemen was unified and in 1999 the islands were administratively assigned part of the Governorate of Hadramaut with its capital Al Mukallā. In 2013 Socotra was declared a governorate of its own as part of the political reforms in Yemen. This current status grants Socotra relative independence and protection from the political instability prevailing at the Yemeni mainland (composed from Cheung & DeVantier 2006 and https://en.wikipedia.org/wiki/Socotra - 25.07.2016).

#### RESEARCH

In the late 19th century Socotra had been the subject of a series of scientific expeditions, led by Balfour in 1880, by Riebeck and Schweinfurth in 1881, by Forbes and Ogilbie-Grant in 1897–1898, and by Paulay and Simony in 1898. Very little research took place in the first half of the 20th century, but in the 1970s interest in the former South Yemen was renewed. Scientific efforts gained momentum from 1996 onwards, with the so-called 'Socotra Biodiversity Project' (SBP) of the United Nations Development Programme (UNDP) (Cheung & DeVantier 2006). The 43 reports that were generated by an international team of more than 30 scientists contributed substantially to greater understanding of its marine ecosystems and biota (Krupp et al. 2002). These included key works on coastal and nearshore marine habitats (Klaus & Turner 2004), coral reefs and their communities (e.g., DeVantier et al. 2004), and macroalgae communities (e.g., Schils & Coppejans 2003). Research on the fish communities and fisheries was documented in a large number of reports, yet only a few of the results have been published to date (e.g., Kemp 1998, 2000; Gill & Zajonz 2003; Gill & Zajonz 2011; Lavergne et al. 2016).

After the termination of the SBP in 2002 the author continued to investigate the fish communities of the island group. From 2007 to 2015 funding for a follow-on research programme became available through the Senckenberg Biodiversity and Climate Research Centre (BiK-F). Some of the results of this work have already been published (e.g., Lavergne et al. 2013, 2014, 2016; Zajonz et al. 2016, 2019), and various publications are currently in preparation. Results of a survey in 2014 by a team of King Abdulasiz University, Jeddah, contributed to a number of recent publications (e.g., DiBattista et al. 2015). Since 2011 fieldwork around the Socotra Archipelago has frequently been hampered by the political situation on mainland Yemen.

#### THE ISLANDS AND THEIR **OCEANOGRAPHY**

The Socotra Archipelago lies in the northwestern corner of the Indian Ocean at 53°00'-54°35' E and 12°05'-12°43 'N, at the junction between the Gulf of Aden and the Arabian Sea (Figure 1). It includes the main island of Socotra, together with three smaller islands, Samha, Darsa, Abd al-Kuri and twoislets, Sabuniya and Kal Farun. The westernmost island Abd al-Kuri is separated from mainland Africa by the Socotra Passage, a narrow strip of water only 95 km wide, and the eastern tip of Socotra Island is 330 km from the nearest point of mainland Arabia, at Ras Fartak, Yemen. The main island of Socotra is 134 km long and has a total area of about 3 600 km<sup>2</sup> — after Madagascar it is the second largest island in the Western Indian Ocean.



Figure 1 Map showing position of Socotra off the Horn of Africa.



**Figure 2** Schematic diagram of the seasonal variation of the major ocean currents and circulatory features in the northern Indian Ocean and Gulf of Aden region. Solid black arrows indicate surface currents (0–100 m), dashed grey arrows indicate subsurface currents (100–400 m depth) when they flow opposite to surface currents. Black shaded areas show the areas of seasonal cold-water upwelling. Abbreviations: South Equatorial Countercurrent (SECC), Somali Current (SC), Southern Gyre (SG), Great Whirl (GW), Socotra Eddy (SE), North Socotra Warm Eddy (NSWE), and North Socotra Cold Eddy (NSCE). Modified from Klaus & Turner 2004, compiled and adapted from several sources

The islands are subject to the alternating monsoon seasons in the northern Indian Ocean: the weak and wet winter, or northeast monsoon (November to February), and the forceful and dry summer, or southwest monsoon (May to September). This seasonally reversing monsoon system creates particular oceanographic conditions around these islands. The most important is the cool-water, nutrient-rich upwelling during the summer monsoon season which temporarily establishes more temperate ('pseudo-temperate') sea conditions in an otherwise tropical environment. With the onset of the southwest (summer) monsoon the surface flow of the Somalia Current is dramatically reversed along the African mainland coast, from southward to northward. The reversed Somali Current, in combination with the southwest monsoon winds, results in the formation of two distinct upwelling systems at the coast of Somalia and at the southeastern Arabian coast of Yemen and Oman (Figure 2) (e.g., Wyrtki 1973; Schott et al. 1990; Fischer et al. 1996).

The high primary productivity resulting from these monsoonal dynamics provide the basis for exceptionally high productivity levels of fishes and other organisms (see below). The fish biomass is exploited by a large and productive fishery (IFAD 2010; Zajonz *et al.* 2016): "The north-eastern part of the Gulf of Aden and the area south of Socotra are among the most productive marine areas in the world, with productivity levels comparable to those off the coasts of Peru and West Africa" (Hariri *et al.* 2002).

The monsoon-driven oceanographic dynamics and the coastal upwelling also influence the spatial and (especially) the temporal distribution and composition of the benthic and associated fish communities (e.g., Zajonz & Khalaf 2002; Klaus & Turner 2004; Zajonz *et al.* 2016). Moreover, the situation of the islands and the dynamics of changing monsoons places the island group literally at the crossroads (DeVantier *et al.* 2004) between the regional ocean currents, which has implications for the dispersal and genetic connectivity of marine organisms, and therefore for patterns of marine diversity and biogeography in the wider northern and western Indian Ocean (Kemp 1998; Zajonz & Khalaf 2002; DeVantier *et al.* 2004; Schils 2006; DiBattista *et al.* 2015; Zajonz *et al.* 2019; Zajonz *et al.* in press).

#### HABITATS AND BIODIVERSITY

The shallow, nearshore benthic habitats around the Socotra Archipelago support highly heterogeneous and spatially variable benthic communities. These range from sandy habitats with seagrass beds and other soft sediment habitats, to rocky habitats with turf and macroalgae and sponges, mixed macroalgal and hard and soft coral communities, and highly diverse hard coral dominated communities (Klaus & Turner 2004). Coral-reef growth is constrained as a result of the marginal 'pseudo-temperate' conditions and the generally gently sloping or flat coastal topography. Where reefs are found they are typically situated in the lee of headlands, or within embayments that are sheltered from the monsoon winds. Coral-dominated communities usually grow directly on a rock substrate and coral cover and diversity is high, with some
253 scleractinian species (58 genera, 16 families) recorded to date (DeVantier *et al.* 2004; Klaus & Turner 2004).

Diverse macroalgae-dominated assemblages cover a substantial part of the inshore areas, particularly on the south coast of Socotra Island, which is most exposed to the southwest monsoon and upwelling. Here algal communities have the highest number of species and a relatively homogeneous community structure, which shows a lower affinity with the (sub-)tropical Indian Ocean flora, and is marked by disjunctly distributed species. Along the northern coast more species commonly known from the tropical Indian Ocean occur (Schils 2002; Schils & Coppejans 2003).

The main island of Socotra has about 30 estuaries and one lagoon. Most of the estuaries are separated from the sea by berms of gravel or sand and are only temporarily connected to the sea after flash floods or by storm surges (Klaus & Turner 2004; Lavergne *et al.* 2016). As a result, and in combination with the local topography, they vary greatly in morphology, freshwater and seawater inputs and ecological conditions; the monsoonal changes contributing to the marked seasonal changes in water flow, salinity and temperature regimes and, hence, very variable and heterogeneous estuarine environments. There are no such ecosystems on the other islands.

#### **FISH DIVERSITY**

The Socotra Archipelago lies at the centre of a region with relatively poorly known coastal and marine fish faunas, with limited information published on the taxonomy and ecology of the coastal fishes of the islands. Fishes were sampled for the first time on Socotra Island during the German expedition of Riebeck and Schweinfurth in 1881-1882, resulting in an account of eight species being published by Taschenberg (1883). His records are, however, not entirely reliable. Steindachner published more extensively, on material collected by the Austrian Expedition to Socotra and South Arabia in 1898-1899. He listed 56 species of marine and brackish water fishes and described Gerres socotranus and Hirundichthys socotranus as new to science (Steindachner 1902; Lavergne et al. 2016). A number of expeditions visited the island group after that, but did not contribute substantially to our knowledge of its fishes. Kemp (1998, 2000) was the first to assess the inshore fish communities and reported 215 species based on visual records. He also made the first regional zoogeographical analysis based on distribution data of four families. The SBP inventories and quantitative fish community

surveys made in 1999–2002, found the fish communities to be rich in species, with some unique regional reef fish assemblages (Zajonz *et al.* 2000; Zajonz & Khalaf 2002; Zajonz & Saeed 2002). Investigation of inshore and estuarine fish assemblages continued, with the first description of a new species of fish (*Halidesmus socotraensis*) from the islands for more than a century (Gill & Zajonz 2003). This was followed by a study of pseudochromine and pseudoplesiopine dottyback fishes, which included the descriptions of *Pseudochromis chrysospilus* and *P. socotraensis* (Gill & Zajonz 2011).

# Inshore and coral-associated assemblages

A preliminary account of the coastal fishes reported 682 confirmed and 51 unconfirmed species, totalling 733 species in 108 families. Using incidence-based richness models of Colwell et al. (2012), between 830 and 890 species (Figure 3) are predicted to occur (Zajonz et al. 2016; Zajonz et al. 2019). Of the 86 families encountered in the field, the Labridae are the most speciose, with about 10% of the species, followed by the Gobiidae, Pomacentridae and Serranidae. When compared to the species richness recorded in adjacent regions, the species diversity of Pseudochromidae (13 spp.), Chaetodontidae (29 spp.) and Acanthuridae (29 spp.) were found to be particularly high. In comparing the species richness of eight key families (Acanthuridae, Balistidae, Chaetodontidae, Labridae, Pomacentridae, Pomacanthidae, Pseudochromidae and Epinephelidae, with 411 species in total) used to analyse the relatedness of 10 putative Arabian ecoregions, the richness at Socotra was the highest (Figure 4) (modified from Spalding et al. 2007; see section on biogeography). This diversity is especially striking when comparing species-area relationships with neighbouring ecoregions (Figure 5). Certain 'reef' associated families and functional groups are as diverse, or more so, than these groups in the entire Red Sea, even though the islands' coastline is but one-eighth in length that of the Red Sea, and supports considerably fewer biogenic reefs (Zajonz et al. 2019; Zajonz et al. in press). The islands thus appear to be endowed with the highest diversity of marine fishes of any reasonably comparable biogeographic unit in the wider Arabian region (Zajonz et al. 2019). Certain small, cryptic, and nocturnal groups are still under-assessed and further work is expected to reveal the presence of more species, particularly of Blenniidae, Gobiidae, Tripterygiidae, Labridae, Platycephalidae, Mugilidae, Ophichthidae, Scorpaenidae, Syngnathidae and various families of flatfishes.



**Figure 3** Fish species-richness modelling for Socotra Archipelago using incidence-based richness estimators (Chao 2, ICE, Jackknife 2), based on 68 fish inventory sites (FIS) and 487 species (six of 74 FIS were excluded from the richness modelling in order to avoid spatial bias). Source: Zajonz *et al.* 2016



**Figure 4** Comparison of actual species richness of eight key families in 10 putative ecoregions (ecoregions modified from Spalding *et al.* 2007, see Zajonz *et al.* 2019) comprising the wider Arabian region and Kenya as an external reference (compare Zajonz *et al.* in press for ecoregional delimitations and a regional species distribution list for these families). \*comprises approximately the 'Central Somali Coast' and 'Northern Mons oon Coast' ecoregions of Spalding *et al.* (2007). Source: Zajonz *et al.* 2016 modified, Zajonz *et al.* in press



Figure 5 Comparison of fish species-area relationships (SAR) between Socotra Archipelago and neighbouring geographical units, using **A** the number of species per 100 km of coastal length, and **B** the number of species per 100 km<sup>2</sup> of coral area. Source: comparative data from FishBase (Froese & Pauly 2017), www.fishbase.org, sourced 2012; and ReefBase, www.reefbase.org, sourced 2015, verified and adjusted (Zajonz *et al.* 2016)

# Estuarine and lagoonal assemblages

Lavergne *et al.* (2016) analysed estuarine and lagoon fish diversity and fish assemblage structure at 13 sites at Socotra Island, and compared them to three sites on the coast of Yemen. Their study included collections and surveys made between 1999 (SBP) and 2009. A total of 65 species in 32 families were recorded from Socotra, with but 20 species in 17 families from mainland Yemen. Twenty-one species were new records for Socotra. If Steindachner's historical records are included, the number of fish species of the estuaries and lagoons is 76, and richness models suggest that the actual estuarine fish species richness might be even higher than this.

Mugilidae and Gobiidae are the most speciose estuarine families, followed by Lutjanidae, Gerreidae and Sparidae. Five species dominate occurrence and abundance frequencies: *Terapon jarbua*, *Hyporhamphus sindensis*, *Aphanius dispar*, *Ambassis dussumieri* and *Chelon macrolepis*.

#### BIOGEOGRAPHY

The Socotra Archipelago has long been considered to be located at the intersection of several distinct biogeographic entities, by Rosen (1971), Klausewitz (1972), Briggs (1974), Hayden et al. (1984), and others. Based on the zoogeography of the corals, the archipelago was thought to be at the junction of the Arabian and the Western Indian Ocean subprovinces, within an Indian Ocean marine province, or, in terms of fishes, just north of that boundary (e.g., Klausewitz 1978; Ormond & Edwards 1987; Sheppard & Salm 1988; Klausewitz 1989; Sheppard & Sheppard 1991; Sheppard et al. 1992). Klausewitz (1989) suggested the archipelago was located close to the boundaries between the Eritrean and South Arabian section, and between the South Arabian and "Persian" section of the Arabian subprovince, and close to a centre of speciation and endemism identified on the Arabian Sea coast of Oman (see Randall 1995; Randall & Hoover 1995). These studies were, however, based on very limited data for the archipelago.

The marine biogeographic affinities of the archipelago were summarised by Kemp (1998), DeVantier *et al.* (2004), and others. DeVantier *et al.* (2004) applied the term 'zoogeographic cross-roads' to characterise the particular biogeographic attributes of the archipelago. The location of hypothetical biogeographical boundaries and the evidence for putative barriers that give rise to faunal and floral breaks in the seas around the archipelago, have been and are still a matter of considerable scientific debate (see, for example, Sheppard & Salm 1988; DiBattista *et al.* 2015; Priest *et al.* 2015; Hodge & Bellwood 2016).

Kemp (1998) was the first to study the zoogeography of the coral-reef fishes of the archipelago, and substantially advanced our understanding of the marine biogeography of the northwestern Indian Ocean (Kemp 2000; Kemp & Benzoni 2000). He identified a distinct South Arabian region, combining parts of southern Oman and eastern Yemen, and recognised strong affinities of this region to the Socotra Archipelago, together with an East African influence. These studies indicate that the Gulf of Aden (including the Socotra Archipelago) is probably not a homogeneous biogeographic entity; an important result that - surprisingly - was largely ignored in most subsequent publications that referred to Kemp's work (e.g., Spalding et al. 2007; Briggs & Bowen 2012; Kulbicki et al. 2013; DiBattista et al. 2015). The data Kemp (1998) primarily dealt with covered the regional distribution of five families of reef-associated fishes, with a focus on the Chaetodontidae, and has now been superseded by newer inventories of Zajonz et al. (2016) and Zajonz et al. (2019) (Figure 6).



**Figure 6** Comparison of coastal and marine ('All species') fish species richness between Socotra Archipelago and neighbouring geographical units ('Coastal' species number follows the total archipelagic richness extrapolated from the species richness models; 'All species' includes an estimate of expected deep-dwelling species based on unpublished regional data of Zajonz & Bogorodsky). Source: comparative data from Froese & Pauly 2014, www.fishbase.org, sourced 2012, verified and adjusted

Recently, a string of primarily phylogeographic and phylogenetic studies have contributed information relevant to the ichthyogeography of both the Socotra Archipelago and the Gulf of Aden, and have been reviewed by Zajonz et al. (2019) and Zajonz et al. (in press). Of these studies, the work of DiBattista et al. (2015) is especially noteworthy because it recognises the archipelago as a prominent hotspot for marine fish hybridisation. None of these recent studies, however, aimed at reviewing the distributional or ecological ichthyogeography or marine biogeography of the Socotra Archipelago, the wider Gulf of Aden and the adjacent seas. To date, the results of contemporary global metastudies as well as of taxon-specific regional studies do not yet correspond to one another in a satisfactory way, and are altogether based on outdated fish species lists for the Socotra Archipelago and southern Arabia; since then number of recorded species has more than tripled (Zajonz et al. 2016; Zajonz et al. 2019). Based on updated inventories Zajonz et al. (in press) analysed the distributional ichthyogeography of the archipelago, challenging current ecoregional concepts.

# Species distribution ranges and converging distribution limits

Based on records of 658 species with reliably known distribution ranges the fish assemblages of the archipelago are dominated by species from the Indo-Pacific (almost half) and the northwestern Indian Ocean (Zajonz *et al.* 2019). A relatively high number of species have both pan-Indian Ocean and wider, Indo-Pacific, distribution ranges.

Among 234 species of 8 key families (see Figure 4), the proportion of species with distribution range limits converging on Socotra is >50% (Zajonz et al. in press). Such a high rate of convergence inevitably leads to the sympatric co-existence of sister species or very closely related species, and also offers an explanation for the comparatively high rate of hybridisation, and a relatively low number of only five endemic species (Zajonz et al. 2019). The island group can then also be characterised as a 'faunal melting pot'. Given that the number of species at their northern distribution boundary is more than twice as high as those at their southern boundary, and that species with a northern Indian Ocean distribution pattern are substantially underrepresented (compared to 'Arabia' as a whole, sensu Manilo & Bogorodsky 2003), strongly suggests that the archipelago's role in promoting species dispersal in a northward direction is more pronounced than it is in a southward direction (Zajonz et al. in press).

Kemp (2000) was the first to recognise the eastern Gulf of Aden coastline as an important region for the hybridisation of reef fishes, based *inter alia* on the occurrence of *Pomacanthus maculosus* × *semicirculatus* at the Hadramaut and Shabwa coast of the Yemeni mainland. The occurrence of this hybrid on Socotra Archipelago was first reported by Zajonz & Khalaf (2002). Recently, DiBattista *et al.* (2015) identified seven fish hybrids from Socotra Island – combinations of 14 species – which led them to recognise the archipelago as a global hotspot for fish hybridisation. Based on this and additional observations from both Socotra Archipelago and the eastern Arabian mainland coast, it seems reasonable to assume an 'eastern Gulf of Aden hybrid zone' which encompasses the eastern Arabian hybrid zone proposed by Kemp (2000), and the Socotra hybridisation hotspot proposed by DiBattista *et al.* (2015) (Zajonz *et al.* in press).

# FISH BIOMASS PRODUCTIVITY

The impressive biological diversity of fishes and many other marine taxa, and the unique community and biogeographic features, are matched by the high fish biomass productivity around the island group, which sustains a substantial local fishery sector (see below). Fishing is thus the second-most important source of income and food for the islanders after its pastoralist economy (Cheung & DeVantier 2006).

The generally high fish biomass productivity in the eastern Gulf of Aden and the wider Arabian Sea was determined by a series of fisheries research cruises during the 'Indian Ocean Programme' (IOP) of the FAO and UNDP, conducted by the research vessel Dr Fridtjof Nansen between 1975 and 1984. Saetersdal et al. (1999) reported a standing stock of demersal and pelagic (small pelagics and mesopelagics) resources for the area Somalia-NE Arabian Sea in the order of 110 t/nmi<sup>2</sup>, which corresponds to standing biomass in excess of 3 million tons for the area Somalia-southern Arabia based on 1975–1977 data (matching earlier data of Cushing 1971). Surveys in 1983–1984 confirmed standing biomass of small pelagics and mesopelagics of 0.93 million tons for Somalia and Yemen alone (Saetersdal et al. 1999). While these figures highlight that the Socotra Archipelago is situated in one of the most productive seas and upwelling areas globally, very few data on inshore and reefal fish biomass exist from this region.

Inshore biomass estimates based on visual underwater length-frequency counts were started in 2007 by the author as a proxy for the coastal secondary and tertiary productivity of the Socotra Archipelago, adding to the permanent transect-based ecological monitoring program operated since 2000 (Zajonz & Saeed 2002). Standing crop in 2007 (pre-monsoon) averaged 2.08 t/ha across eight sites (six at Socotra Island, one at Darsa Island and one at Samha Island) and 3.04 t/ha at five deeper sites (> 6 m). These biomass values are impressive, especially considering that fishing activities had already harvested the coastal productivity for the six months following the summer monsoon. Pre-monsoonal maxima of 5.13 t/ha (SD  $\pm$  5.64) were recorded around Socotra and 5.95 t/ha (SD  $\pm$  9.34) around Darsa in 2007. In 2011 the post-monsoon maxima recorded at individual sites were 5.03 t/ha (SD  $\pm$  1.67) and 6.77 t/ha (SD  $\pm$  3.06). These maxima rank among the highest in the Indian Ocean (e.g., Garpe & Öhman 2003; McClanahan et al.

2007; Burt *et al.* 2011; McClanahan 2011; McClanahan *et al.* 2011) and are only surpassed by biomass estimates recorded from the Chagos Archipelago (>7 t/h), which is one of the most strictly enforced fishery closures globally (Sheppard *et al.* 2012).

Comparisons between pre- and post-monsoon biomass of 113 fish species in 2011 indicate a substantial replenishment of biomass following the summer monsoon. Biomass increases of 1012% (SD  $\pm$  722%) were recorded from 12 relatively undisturbed sites on the northeast coast of Socotra, representing an increase from an average of 0.183 t/ha (SD  $\pm$  0.275) in May to 2.035 t/ha (SD  $\pm$  2.052) in November 2011. Increases in biomass were non-linearly related to an increase of more than fourfold in the in number of fishes and of 40% in the number of species. The increase in biomass is also related to higher average weight per individual (+270%) in the post-monsoon period (Zajonz *et al.* 2016 and unpubl. data).

These findings correspond with the marked seasonal effects of the summer monsoon. The replenishment appears to be mainly and closely related to enhanced primary productivity rates during and following the monsoon seasons, notably the more pronounced summer monsoon, as inferred from sea surface temperatures and of chlorophyll *a* concentrations (Figure 7).



0 0.1 0.2 0.3 0.4 0.5 0.6 0.7 0.8 0.9 1.0 1.5 2.0 2.5 3.0 4.0 5.0 10.0 15.0 >20.0

**Figure 7** Mean monthly Chl *a* concentration 2000–2011 around the Socotra Archipelago. Source: OceanColourWeb (sourced 2013)

#### **FISHERIES**

The Socotra Archipelago supports productive fisheries that supply both domestic and commercial markets. Research into these fisheries has however been limited. Most of the information available about these fisheries up to 2002, come from two documents: the *Fisheries Management Plan* prepared during the SBP (Nichols 2001; Esseen & Al-Saqaf 2002; Hariri 2002), the only one of its kind in Yemen, and the *Fisheries Feasibility Study*, prepared as part of the Socotra Archipelago Master Plan study (Reid *et al.* 2002).

Most of the fishing activities around the islands are essentially artisanal, beach-landing fisheries. These operate within 10 km of the coast, over the shelf, to 200 m depth, and cover an area of 14 417 km<sup>2</sup> (Nichols 2001). The islands are also targeted by industrial-scale fishers, notably longliners, midwater and a few demersal trawlers, and gillnetters, especially during the summer monsoon, despite the 12 nmi conservation buffer zone and a 5 nmi non-trawling zone. This poaching is deplored by local fishers.

Artisanal fisheries cease during the summer monsoon season in almost all areas. The average fishing season is then only 8 to 9 months out of the year, during which a fisherman will spend 160 to 180 days – 220 at most – fishing (Hariri & Yusif 1999; Nichols 2001; Reid *et al.* 2002), depending on various factors. Fishers operate from some 86 coastal villages and settlements across the islands, ten of which are on Samha and Abd al-Kuri. For many years there was a single cooperative, the Socotra Fishermen Cooperative, established in 1970. In 1993 it had 1 500 members, but it ceased to operate in 1994. There are now 25 cooperatives after the concept was promoted by the SBP and other fisheries development projects (Yusuf & Kassem 2000; Riet *et al.* 2000; Nichols 2001).

Socotran fishermen predominantly use two types of Yemen-crafted boat. The first is an open, fibre-glass reinforced plastic skiff, known locally as *bushenda*, *qareeb*, *qa'difa*, *plastic* or *houri*. These are 7.7–9.5 m long and equipped with one or two 15–40 hp outboard engines and have a crew of 2–4 fishers. These are the most abundant and are used for dayrange activities. The second is larger and is known as a *sanbuq* or *sambuq*. They are up to 15 m in length, made of wood or fibreglass, have insulated holds with ice to store the catch, and are equipped with 40–75 hp outboard engines. They take 5 to 7, and occasionally as many as 12, crew. *Sambuqs* are used primarily for gillnetting, longlining, and catch-buying journeys lasting several days (Figure 8).



**Figure 8** Main types of fishing crafts in Socotra's artisanal fishery: **A** fibre-glass reinforced skiff with outboard engine (*houri, qa'difa, bushenda*); **B** wooden (as here) or fibre-glass boat with inboard diesel engine (*abriy, sambuq*). © U Zajonz

The fisheries primarily target shallow demersal and 'reef' fishes inshore, and large pelagic species and sharks offshore. There is also a rock lobster and sea-cucumber fishery, and a more limited deep demersal fishery, which have only developed over the last decade. Some of the larger estuaries are also opportunistically or seasonally fished during spawning and foraging migrations of valued species (e.g., Nichols 2001; Mohsen 2002; Zajonz pers. obs).

The large pelagic fishery targets primarily kingfish (*Scomberomorus commerson*), yellowfin tuna (*Thunnus albacares*), rainbow runner (*Elegatis bipinnulata*), dorado (*Coryphaena hippurus*), kawakawa (*Euthynnus affinis*) and longtail tuna (*Thunnus tonggol*), with sailfishes, marlin and other tunas being caught incidentally. These boats primarily use surface-set gillnets, pelagic longlines and trolling lines.

The shark fishery targets 12–15 species of requiem, hammerhead and mako sharks, using longlines and gillnets, but also baited single hook-and-line. The catches are predominantly silvertip shark (*Carcharhinus albimarginatus*), spottail shark (*C. sorrah*) and scalloped hammerhead (*Sphyrna lewini*), as well as the shortfin mako (*Isurus oxyrinchus*). In catches landed along the southern coast of Socotra Island, species such as the blacktip reef shark (*Carcharhinus melanopterus*), Human's whaler shark (*Carcharinus humani*) and tiger shark (*Galeocerdo cuvier*) are also abundant (Saeed 2000; Nichols 2001).

The shallow demersal and reef fishery targets about 120 different species, primarily in shallow coastal habitats, particularly species of high-value families, such as groupers (Epinephelidae), trevallies or jacks (Carangidae), snappers (Lutjanidae), sweetlips (Haemulidae) and emperors (Lethrinidae). Species of many other families are taken opportunistically and, if not sold, are used for domestic consumption. The dominant gear is a baited vertical hook-and line. Traps are used occasionally at discrete sites, with beach seines and cast-nets being resorted to by older and poorer fishers for subsistence fishing.

# Fisheries catch and effort

As with other multi-gear, multi-species artisanal fisheries, there are numerous isolated landing sites, and a large number of stakeholders with different vested interests. Catches are marketed directly on the beach or to buyer vessels, most of which come from the mainland. Unlike the rest of Yemen, fish are sold by the piece and not by weight, and there are no direct records of these transactions. Instead, the value (price) is supposed to be reported to the fisheries office which uses price–weight coefficients by species to recalculate the weight – obviously not very accurate a recording procedure. The dispersed nature of the landing sites, the method by which catches are sold, combined with a local fisheries administration with limited capacity, make monitoring fisheries around Socotra a serious challenge.

Efforts to establish a fishery monitoring programme on Socotra started in 1998, during the SBP. The data collected were partly of limited usefulness (Nichols 2001) on the one hand, and partly put to limited use only on the other hand. Further, the data analysis was restricted to 6 of 17 fishing areas, simply because of the enormity of the task. The data were never used to calculate catch-and-effort trends, or make stock assessments, and then ceased to be used after the project ended in 2005.

The basic data for the estimates presented in this account (Zajonz et al. 2016) were partly compiled from the reports of fisheries consulting teams that visited the islands between 1998 and 2002, on projects funded by the UNDP-GEF and the EU. The data were then verified and updated using various other unpublished sources, reports and personal observations. Data on catch and effort before 1998 and after 2001 are primarily derived from records from the fishing cooperatives and accounts from fisheries authorities, are consequently very unreliable (Nichols 2001; Reid et al. 2002; Zajonz pers. obs.), and required careful verification, as explained below. Data on fishing effort from before 1993 are not available and those data that do exist only refer to the numbers of vessels and fishermen, and estimates of fishing days per unit and per year, and has been reported by Esseen & Khanbash (1999), Hariri & Yusif (1999), Yusuf & Kassem (2000), Zajonz et al. (2010), Scholte et al. (2011), and others. Catch-and-effort data between 2005 and 2014 were collected de novo from the fisheries stakeholders.

The effort data from these sources were reviewed altogether and used to produce two time-series curves (Figure 9) illustrating the lower and upper range of the total numbers of boats and fishers over the past 21 years. These data show that between 1996 and 2014 the number of boats increased by 41% to approximately 1 100, and the number of fishers rose by 32%, to about 3 800. Overall effort seems to have reached a maximum in 2006 before levelling out. One of the reasons for the increase in number of fishers on Socotra is a result of population growth. Another is the increasing number of Bedouins from the island interior that have joined the fishery during the last decade, to supplement family incomes derived from traditional pastoral livelihoods.

There is some uncertainty with these data because recent statistics refer only to fishers registered (organised) with the cooperatives, whereas earlier figures also included 'unorganised' fishers i.e., among the approximately 5 000 fishers given in Hariri & Yusif (1999) and IFAD (2010). There may thus be an additional few hundred fishers, parttime fishers, deck-hands and helpers that are not accounted for as they have not joined one of the cooperatives, and the total numbers are probably closer to 1 300 boats and 4 500 fishers, with unconfirmed reports of 2 000 boats and up to 6 000 fishers.



**Figure 9** Basic consolidated effort trends for Socotra Archipelago from 1993–2014, based on the numbers of fishing craft and active fishers, showing for both the upper and lower range margin of plausible data. Years that are completely data deficient are not listed on the X-axis. Source: Zajonz *et al.* 2016

The substantial increase of overall fishing effort around Socotra during the last 20 to 30 years corresponds with similar trends observed for the Gulf of Aden mainland (Zajonz & Akester 2005) and Yemen as a whole (e.g., Tesfamichael *et al.* 2012). Zajonz *et al.* (2010) reported that the number of boats in Yemen had more than tripled between 1997 and 2006, from 7 150 to 22 600, and was likely to have reached 29 000 by 2010. Likewise, the number of fishermen had doubled between 1997 (37 600) and 2006 (63 200), and further increased to 83 200 by 2010 (Zajonz *et al.* 2010).

# Total catch

The data available for the main fisheries and the total fisheries production for the Socotra island group are, like the effort data, also limited and unreliable, and for several years such data are not available (Zajonz et al. 2016). There are principally two sources for data, each with different biases and levels of accuracy. The first source is actual 'catch records' (and associated revenue data) registered by the fishing cooperatives, and partly compiled and summarised by resource group (e.g., 'sharks', 'demersals', 'large pelagics', etc.), by the local branch of the Ministry of Fish Wealth of Yemen. The second source is 'catch estimates' produced by trained fishery scientists or technicians, and is based on field samples of catch and effort and fleet counts. They are usually higher than actual records, as explained below. No catch records were available for years the estimates were produced for, and thus cross-validation is not possible.

In the 1990s the catch records seem to have been checked for accuracy, and corrected by fishery scientists of the then Marine Science and Resources Research Centre in Aden (MSRRC) of the Ministry of Fish Wealth (MFW), and appear relatively plausible (see Saeed 2000 or Mohsen 2002). From 2000 onward no catch data that have been quality-guided and consolidated by local fisheries experts from the cooperatives or MFW, are available. When the SBP database system was in operation there were no data from the cooperatives (c. 1998–2004). Data for 2005–2007 are in a range that could be considered plausible, whereas data from 2008 onwards appear to be pretty inaccurate, and several years' records are missing.

The available data were then used to reconstruct long-term catch trends over a period of 25–30 years (Figure 10). As with the effort trends, two time-series curves are illustrated, showing the lower and upper margins of the plausible range of fisheries production. The same data were also used to extrapolate mid-term catch trends for the period 1990 to 2014 (Figure 11), based on the relative proportion the main resource groups had on average in 2005–2007: sharks (49%), large pelagics (27%), reefal-demersals (23%), and other (1%). During this 25-year period, data for reefal-demersals are available for 12 years, while data for the other groups are fewer and more scattered.



**Figure 10** Consolidated long-term trend of total finfish catch for Socotra Archipelago from 1981–2014, based on available records, qualified estimates and partial reconstruction, showing the upper (T2) and lower (T1) range margin of plausible data. From 1981–1993 and 2005–2014 T2 is based on reconstructed catch data. Trend curves (T1, T2) are given as continuous lines bridging data-deficient years. The figures of 1999/2000 refer to fishing season and not to calendar year but were used in the absence of alternative data. Years that are completely data deficient are not listed on the X-axis. Source: Zajonz *et al.* 2016



**Figure 11** Mid-term trend extrapolation of total finfish catch for Socotra Archipelago from 1990–2014, based on available records and reconstruction of missing data according to the average proportional shares of the main resource groups in 2005–2007. Data from 2005–2014 represent actual catch data adjusted to account for assumed under-reporting. Years that are completely data deficient are not listed on the X-axis. Source: Zajonz *et al.* 2016

The reefal-demersal data were used to estimate the missing potential catch data of the other groups (retaining extant values) using average group shares for 2005–2007, the only years for which data are concurrently available.

Records available for 1981–1996 from the then Socotra Fishing Cooperative indicate a total annual catch of 391 metric tons (mt) in 1981, 1036 mt in 1990 and 705 mt in 1996 (Nichols 2001). According to unpublished estimates of the SBP (in collaboration with international development agencies), annual production rose dramatically to 7 284 mt in 1998, based on a fleet of 843 craft, a 160-day working season and an average catch of 54 kg per boat per day (also in Reid *et al.* 2002).

For the same fleet and season Hariri & Yusif (1999) provide an estimate of 8 270 mt, based on a breakdown of production by boat type. Based on recorded length-frequencies (which are converted into weights) for the catch of a subsample of the actual total boat trips, the SBP team (unpubl.), using lengthweight coefficients, extrapolated a total annual catch of 1 516 mt for 6 of 17 fishing areas for the 1998/1999 season. According to Nichols (2001), artisanal catch estimates for the main groups for the 1999/2000 season ranged between 6 030 and 6 139 mt in total. Substituting the implausibly low shark-catch figure used by Nichols (2001) with the median of 5 000 mt for the estimated range of the shark catch given by Huntington & Al-Sakaf (2000) [unavailable, see Reid et al. 2002], the catch production of Socotra's fisher fleet was probably some 9 885 mt in 1999/2000. The alternative values from the reefal-demersal-based extrapolations are between 13-25% higher, at 11 560 mt in 1999/2000. These figures mark the peak catch production for the 35 years for which data are available. These statistics represent a sharp rise in production from the early 1990s, followed by a steep decline, with current production figures reported by the cooperatives similar to those reported from the early 1980s.

An assessment of the buyers' records of a single large fishing cooperative on the Hadramaut coast was made to validate these findings, and gives a total of 3 612 mt of fish traded in from Socotra from 2007 to 2011. These data also show a severe decline in catch available to buyers, from 2 384 mt in 2007, to a mere 244 mt in 2009, and to only 68.8 mt in 2011, corroborating the negative trend observed from the Socotra records.

Figure 12 shows the annual total finfish production per boat and fisher as basic proxies of catch-per-unit-effort (CPUE), and the trend from 1993 to 2014. The CPUEs peaked in 1999–2000 ranging between 8.1 and 11.4 mt per boat and 3.1 and 3.9 mt per fisher, depending on the ranges of boats, the number of fishers and total catch, with a potential maximum of 14.6 mt per boat in 1999. It has slumped dramatically since then, even though there is no doubt that the actual catch production is still substantially higher than the available official records would suggest. This negative trend most likely reflects a period of unsustainable yields for the past decade, and has severely impacted the exploited fish populations over the long term.



**Figure 12** Mid-term trend of basic catch-per-unit-effort (CPUE) for Socotra Archipelago from 1993–2014, based on the means of the plausible value-ranges for the number of crafts and fishers. Years that are completely data deficient are not listed on the X-axis. Source: Zajonz *et al.* 2016

There are four main reasons why actual exploitation levels in the waters around Socotra are substantially higher than those reflected in the available catch records. First, the fishermen pay a 3% levy to their cooperative, based on catch value registered by them, which creates a disincentive to provide the correct values. Second, a similar disincentive applies to the buyers, who are (theoretically) obliged to pay a 5-7% levy on the total purchase value, which is shared between the cooperatives, the MFW, and the governorate. Thus, the catch reports from Socotra (which include local consumption and marketing, and exports to the mainland) should actually be higher than the trading records from the islands to the mainland. The fact that one out of 11 trading records (the mainland cooperatives recorded not only their own catch, but also purchases from elsewhere) exceeded the direct records on Socotra itself, illustrates how flawed and severely underreported the Socotra data are (see Zajonz & Akester 2005). The misreporting of catch in order to evade fees and taxes, led IFAD (2010) to re-estimate the total catch production of Yemen to be in the order of 250 000 mt instead of the 109 000 mt officially reported by the MFW to FAO in 2009. This would translate into the actual catch on Socotra ranging from 22 600 to 26 500 mt for 1999/2000. Third, a substantial proportion of the catch has thus far gone completely unrecorded in Socotra's record books. This includes the catches of Yemeni fishers landed at sites on the mainland, predominantly at Hadramaut, Al-Mahara and Shabwa, and

occasionally elsewhere such as at Oman. These catch numbers have thus far never been added to the catch statistics for the island group. Hariri & Yusif (1999) noted that the shark production from Socotra, which is landed at Hadramaut and Al-Mahara, is about as high as the production registered on Socotra itself (although the actual figures provided by them are too low). Fourth, comparing the average annual production per boat in Al-Mahara with that on Socotra, strongly suggests that the Socotra CPUE resulting from present data is much too low to be credible. In 2003, a total of 7 197 fishers operating 2 479 crafts (crew of 3) landed a total catch of 75 818 mt in Al-Mahara (Zajonz & Akester 2005), equivalent to 30.6 mt annual production per boat and 10.5 mt per fisher, compared to 8.1-11.4 mt per boat and 3.1-3.9 mt per fisher at Socotra in 2000 (crew of 3). Most of the Socotran fishermen would evidently migrate to fish at the mainland if the productivity of their 'home turf' were really that much lower. In reality, however, more mainland fishers are attracted to Socotra than the other way around. This illustrates how severely flawed the catch records from Socotra have become, essentially rendering them useless to inform sustainable fisheries management.

Socotran fisher communities have in the past been recognised for managing their stocks wisely, according to traditional management methods (Morris 2002; Reid et al. 2002; Cheung & DeVantier 2006). These included banning fishing for small pelagics and the use of fish traps, imposing temporal or spatial closures and temporal bans on nets, or, more recently, promoting traps in the lobster fishery instead of nets. Strong peer pressure and community-imposed penalties on infringements were reported as resulting in high levels of compliance with the customary management system. It used to be common sense that the local fishery is more or less sustainable with only a few target species such as sharks and rock lobsters being overexploited (Hariri & Schotah 1999; Nichols 2001; Cheung & DeVantier 2006). The customary management norms of the fishermen of Socotra have long been unique in Yemen and probably the entire Arab region. Indeed, many of the existing traditions that have helped the Socotrans manage their fisheries to date are what organisations elsewhere are only now trying to reinstate by establishing Locally Managed Marine Areas (LMMAs) (Rocliffe et al. 2014). At the same time, the relative isolation of the islands, their limited fish processing facilities, long transportation routes and ill-developed marketing paths created a 'natural' control mechanism that prevented an unsustainable growth of the fishing sector. Historical catches (1981-1996) for all main groups (sharks, kingfish, reef fish and rock lobster) were considered as being at "light exploitation levels", and stocks in the 1999/2000 season generally "in a healthy condition" (Nichols 2001; Hariri et al. 2002, on reefal-demersals). These were most probably already unjustified statements at the time.

It is highly questionable whether the small-scale fishery on Socotra is sustainable nowadays. The increase of the local fleet size over 20 years, an increase in local ice-making, cold storage and primary processing facilities, and an ever-increasing share of production exported to buyers on the mainland, makes Socotra's fishery today a more commercial enterprise and undermines its traditional, artisanal roots and best customary practices. Other underlying causes include rapid population growth, increasing coastal development and expansion of tourism, which exert increasing pressure and higher demand on living marine resources, particularly at key tourist sites. More recently, Socotra's fishing industry has partly come under the control of Arabian Gulf states which adds a lot to the existing fishing pressure. Overfishing now presents a major challenge to the Socotran fisheries sector, not least because the strongly negative trend in fish production, as inaccurate as catch statistics might be, is corroborated by fishers and others in the fisheries sector who unanimously report that catches and CPUE are declining.

# CONCLUSIONS

The Socotra Archipelago has a unique marine environment with tropical and 'pseudo-temperate' features that are generated by particular regional oceanographic conditions driven by the Indian monsoon. Although the diversity and ecology of its fish communities have been studied for about 20 years, the information available has only recently been pulled together. Even less was known about the local fisheries, which had long been sustainably managed through traditional, community-based practices. Furthermore, no information on fish productivity (in terms of biomass) on which these fisheries were based was available. At a time when Yemen is torn by a severe political and humanitarian crisis its living marine resources are likely to experience increasing pressures, as are those of the archipelago.

The coastal fish diversity of Socotra Archipelago is the highest of any 'ecoregional unit' (e.g., Spalding *et al.* 2007) in the northwestern Indian Ocean, and ranks high compared to similar areas in the wider WIO, especially when considering species richness in relation to length of coastline, shelf area and reef area. It is especially striking that many typical reef fish species – such as 29 species of chaetodontids – occur here, and that the island group supports unique reefal and coastal fish communities, in spite of the limited extent of coral reefs. The Socotra Archipelago is therefore critically important to the region's marine and fish conservation management.

The marine biogeographic affinities of the archipelago are characterised as being at a crossroads. The location of hypothetical biogeographical boundaries and barriers in the seas around the archipelago are still a matter of scientific debate. The most eminent biogeographic studies do not yet correspond to one another, and used to be based on outdated fish inventories for Socotra. The archipelago is perhaps less a source than a sink for regional fish diversity. It represents a globally important hybridisation hotspot for marine fishes, forming likely part of an eastern Gulf of Aden hybrid zone and it is a faunal melting pot and key stepping stone for regional marine biogeographic patterns.

The coastal fish biomass productivity of the inshore waters of Socotra Archipelago is high, and certain sites rank among the most productive in the Indian Ocean. Biomass productivity and the underlying communities are seasonally highly dynamic, driven by the seasonal upwellings created by the northeast monsoon.

Recent data point to a sharp decline of standing crop and fisheries productivity over at least the past ten to fifteen years, pointing to serious socio-ecological and livelihood problems in the future. Whether the once traditionally managed smallscale fishery on Socotra is sustainable is extremely doubtful. The following statement of Reid et al., in the fisheries section of the Socotra Master Plan study (2002), is sobering: "... the current liberal management regime, lack of regulation and the ready availability of finance [lending for investment in the fisheries sector] is likely to lead to an unsustainable and poorly structured industry that will not properly serve the Socotran people". Likewise, Hariri et al. (2002) predict that "Socotra's fisheries will come under threat if the islands' societal structure changes markedly or outside interests begin to exploit resources on an unsustainable basis". Overfishing is now prevalent around Socotra Archipelago posing a major problem. The markedly increased fishing efforts, combined with the severe, real, decrease of the fisheries production, raises serious concerns about the future sustainability of Socotra Archipelago's fisheries and not least, the food security for the islands' coastal communities. The deterioration of the traditional, community-based fishing regulations needs to be halted, and the poor enforcement of the regulation effecting existing marine protected areas (MPAs) must be overcome. Effective management of MPAs including 'no-take-zones' will allow recovery of populations, increase fish biomass and will, in the long term, benefit adjacent use-areas through the supply of adult fish and larvae for recruitment (spillover). Active fisheries management through gear and catch

restrictions and export limitations will further limit overfishing and habitat destruction, and thus improve the sustainability of the local fisheries. First and foremost, however, it is pivotal to organise and fund fishery research based on biological and fisheries data in tandem, in order to understand the resource base. Effective evidence-based fisheries management has to take priority over perpetual investment into the production, processing and marketing sector, or else these severely negative trends will not be reversed.

# ACKNOWLEDGEMENTS

This account, of necessity, has drawn heavily on the author's scientific reports and peer-reviewed articles. All co-authors of these joint papers are gratefully acknowledged for their contributions. Many individuals and institutions collaborated during field work or facilitated research in the archipelago. While there are too many to be named individually, a few persons deserve special thanks: Friedhelm Krupp and Edoardo Zandri for providing me with the initial opportunity to work on Socotra, Rebecca Klaus for providing benthic and remote sensing data, Edouard Lavergne for the fruitful and

amicable scientific collaboration over many years and his help in bringing most of the illustrations into shape, Sergey Bogorodsky for the productive cooperation in the field of fish faunistics and taxonomy, Jonathan Brown for his contributions to the fishery analyses, and Moteah Sheikh Aideed and Fouad Naseeb Saeed for providing data and good company in the field. I am most grateful to all of them for their long collaboration in my Socotra research.

Much of the data reviewed in this account were initially collected between 1998 and 2002, under the auspices of the project "Conservation and Sustainable Use of Socotra Archipelago", led by the United Nations Development Programme (UNDP) and funded by the Global Environment Facility (GEF). From 2008 to 2014, research at the Biodiversity and Climate Research Center (BiK-F) of the Senckenberg Society for Nature Research, Frankfurt a.M., was financially supported by the research-funding program 'LOEWE – Landes-Offensive zur Entwicklung Wissenschaftlichökonomischer Exzellenz' of Hesse's Ministry of Higher Education, Research, and the Arts.

Finally, I would like to thank the people of Socotra, notably the fishers and coastal communities and the Environment Protection Authority Yemen and Socotra, who kindly provided access to their knowledge and to the areas under their care.



Figure 13 Scenes from Socotra's artisanal fishery: A–B pelagic fishery, C–E shark fishery, F–G reefal-demersal fishery, H lobster fishery, traps. © A: JJ Brown; B, C, F, G: SV Bogorodsky; D: A Bollen; E, H: U Zajonz

# FISHERIES OF THE WESTERN INDIAN OCEAN

#### Rudy van der Elst

# **INTRODUCTION**

The thousands of fishes described in these volumes reflect the marvellous diversity of Western Indian Ocean (WIO) marine life. The species are nestled in an equally diverse array of ecosystems and habitats that make up the region's natural environment. Not surprisingly, this great biodiversity provides the basis for a range of fisheries and fishers who depend on this resource for food, employment and other ecosystem benefits, including medicinal, scientific and cultural. From humble but often innovative subsistence fishers to sophisticated industrial operations, the WIO region sustains the livelihoods of millions of people and their dependants. Notwithstanding the great value placed on the fishery resources, sustainable harvests can only be assured if they operate within the regenerative capacity of species and their ecosystems. In turn, this calls for the wise and restrained use of resources as well as collaboration between stakeholders. Such regional initiatives do exist and many undertakings are in place to manage the use of WIO resources.

About half the world's population lives in countries that border on the Indian Ocean, and, in most cases, seafood represents an important source of food security, employment and a national resource that contributes to the gross domestic product of the countries. It is estimated that some 30 million people in the 10 countries of the WIO coastal zone depend directly on harvesting marine fishes and contribute to elements of socioeconomic stability in these coastal regions. While large industrial fisheries, such as those targeting tuna and shrimp, supply the world's markets, most developing countries in the region have a large contingent of artisanal fishers that contribute to local food security and the economy.

# THE NATURE OF WIO FISHERIES

There is an amazing variety of fisheries taking advantage of almost every ecosystem and habitat. The most elementary are subsistence fisheries, which include large numbers of fishers engaged in a daily chore of providing food security to their families. Many simply harvest food gathered from the intertidal zone, whereas others might use traps, canoes, handlines and small seine nets to secure their catch. In some cases the fishers supply local markets, mostly using modest, low-technology fishing gear. Such artisanal fishers are common throughout the WIO region and fulfil an important role in the local socioeconomics of different areas. Artisanal and subsistence fisheries are mostly located in the shallower coastal zone. Some fisheries are classed as semi-industrial, making use of larger motorised vessels and various forms of technical innovation, such as GPS devices, radios, icing facilities, and even diving. Still larger vessels, capable of fishing in oceanic waters, are termed industrial fisheries and use fishing gear such as trawls, purse seines and longlines. Many industrial vessels operating in the WIO region are foreign owned and operate under bilateral fisheries agreements, whereby the catch may be taken directly to the vessels' flag country. In some cases the catch is trans-shipped at sea or directly supplied to canneries in the region, such as the large fish processing factory in the Seychelles. Fisheries can also be grouped by the type of gear used, as reflected in the WIOFish Database (http://www. wiofish.org) in which 211 different fisheries are described (Table 1).

GEAR	SEYCHELLES	MAURITIUS	KENYA	TANZANIA	MOZAMBIQUE	SOUTH AFRICA	TOTAL
Diving	2	0	6	4	4	5	21
Harpoon	1	0	0	0	0	1	2
Hook & line	15	10	3	5	7	14	54
Industrial nets	1	1	3	3	4	3	15
Large nets	0	1	2	0	1	0	4
Mixed gears	0	2	0	0	0	0	2
Other	0	0	0	2	0	1	3
Shore gathering	6	2	7	11	3	18	47
Small nets	6	3	8	8	6	10	41
Traps	5	1	4	2	5	3	20
Unknown	2	0	0	0	0	0	2
TOTAL	38	20	33	35	30	55	211

Table 1 Different fisheries' gear types used per country. Source: WIOFish

#### **FISHERY TRENDS IN WIO**

Global trends in fisheries landings are documented by the Food and Agriculture Organization of the United Nations (FAO). The FAO divides the world's marine waters into 19 major fishing areas, and each nation or territory that harvests marine resources in any of the areas voluntarily submits statistics to the FAO. This global repository of fisheries statistics can be accessed through FishStat (http://www.fao.org/fishery/statistics/ global-production/en) and includes national and regional information on total capture production from 1950 to nearcurrent. The Western Indian Ocean, designated as Area 51, is the fifth largest of these fishing areas; it is some 29.3 million square kilometres in extent, or about 8.1% of the global ocean area, stretching from latitude 45°00' S roughly to the southern Cape coast of South Africa (30°00' E longitude) to Somalia and the Arabian Sea, Red Sea and Persian/Arabian Gulf, and eastwards to the southern tip of India (Figure 1). The WIO region is complex with respect to geopolitical boundaries and Exclusive Economic Zones (EEZs), which extend 200 nautical miles (370 km) out from the coast, and in some cases territorial claims are disputed. Not insignificantly, ownership of a tiny atoll can infer the rights to exploit a much larger surrounding EEZ. For example, France is granted extensive fishing rights in WIO by virtue of its sovereignty of numerous small islands and atolls in the region. Also, of consequence is that national and EEZ waters account for approximately one-fifth of Area 51, hence leaving a huge expanse of WIO as international waters. This has received attention in the development of certain international agreements, such as the Southern Indian Ocean Fisheries Agreement (SIOFA), which aims to document catches and regulate landings in remote ocean expanses, such as fisheries for tunas, orange roughy, Patagonian toothfish, sharks, and other oceanic and deepwater species.



Figure 1 African section of the WIO study area (FAO-51), showing national Exclusive Economic Zones (EEZs). Source: FAO

The records collated each year by the FAO focus primarily on formal, more industrialised fisheries, and provide useful insight into broad trends and the status of particular species stocks. Although the FAO continues to report a decline in the global fisheries catch, Area 51 is one of the few fisheries regions of the world that contradicts this trend. Landings in Area 51 peaked at 4.5 million metric tons (mt) in 2012, which



Figure 2 Growth in relative tonnage of fish types harvested annually in the WIO, 1950–2019. Source: FAO



Figure 3 Growth in annual tonnage of catch taken in the WIO by African nations which reported at least 30 000 mt. Period 1950–2019. Source: FAO

represented about 5.7% of the global industrialised fish catch, considerably more than the 1.6 million mt harvested in the nutrient-rich southeastern Atlantic (FAO 2014).

Figure 2 shows the broad composition of WIO fish landings over time. The total fish catch in WIO has increased substantially over the decades, largely attributable to greater fishing effort and a growing tuna harvest that exceeds 1 million mt per annum. Indeed, the tuna canning factory in the Seychelles is the second largest in the world and annually processes around 300 000 mt of tuna. The catch of several other species has also increased, notably that of the Indian oil sardine, which accounts for 300 000 mt per annum.

While the increasing total catch may superficially seem reassuring, in reality it masks several areas of concern. One of these relates to the number of species being harvested, which has risen from at least 85 in 1970 to more than 150 today. This increase is partly due to better reporting and enhanced technology, such as the use of fish aggregating devices, but also reflects a wider demand for 'new' species in the context of declining economic viability of stocks of traditional species. Such trends could be an indication of serial overfishing, whereby one species replaces a depleted species, so as to retain fishing capacity in the region. The Southwest Indian Ocean Fisheries Commission reported that <25% of the region's fisheries were exploited at unsustainable levels in 2010, moderately better than the global average of 30%. While this may seem positive, it leaves no room for complacency, especially as it does not take into consideration the effects of ecosystem overfishing.

As catches in several of the world's fishing zones decline, so the fishing fleets shift their focus to new regions. The number of countries that have reported catches from WIO has reached 60, up from approximately 46 in 2005. About half of these are nations that do not border WIO but operate in the region as part of fisheries agreements with local nations. In some cases foreign vessels operate outside the declared EEZs, mostly in pursuit of tuna, sharks and other oceanic species. Clearly, this highlights the global demand for fish. Based on FAO data it appears that about 20% of the total catch in WIO is taken by such foreign operators. Some insight into the respective and growing harvest taken by countries operating in WIO can be gauged from Figure 3.

While the FAO reporting system is an innovative and useful tool to evaluate the overall conservation status of marine resources, it is imperfect. There are clear indications that not all catch records are fully reported by every country, especially the under-reporting from artisanal fisheries. Some estimates have suggested that the total catch may well be double that which is reported to FAO.

Based on country surveys and research projects, some estimates of the artisanal fisheries in several WIO countries have been made; a preliminary overview is given in Table 2. These fisheries are restricted mostly to inshore areas and carried out over a range of coastal habitats, such as sandy beaches, estuaries, coral reefs, lagoons, wetlands, bays, mangrove forests and seagrass beds, but rarely in oceanic waters. The participants are local coastal communities who fish on a small scale using simple equipment (often boats without engines), fishing for their own use or to supply local markets and in rare cases surplus for export. Typically, individual daily catches per fisher are a few kilograms. Significantly, however, this low-yield but widespread fishery accounts for more than 80% of the total marine catch in Tanzania, Kenya, the Comoros and Madagascar. Regardless of the exact figure of total fish landings from WIO, based on the *declared landings* there is growing concern that the region may be approaching its maximum harvest potential.

NO. ARTISANAL ARTISANAL CATCH COUNTRY **PRINCIPAL FISH FAMILIES** FISHERS (mt/yr) Carangidae, Lutjanidae, Scombridae, Lethrinidae, Epinephelidae, Maldives 14 000 ~100 000 Coryphaenidae, Istiophoridae Comoros 5 500-7 507 - 13 500 ('97) Scombridae, Gempylidae, Carangidae 8 000-8 500 Carangidae, Sphyraenidae, Scombridae, Siganidae, Epinephelidae, Seychelles 1 700-1 800 4 000-5 000 Scaridae Mugilidae, Epinephelidae, Carangidae, Gerreidae, Hemiramphidae, 10 651 12 382-70 000 Madagascar Elopidae Mauritius 2 365 950 Lethrinidae, Mugilidae, Siganidae, Acanthuridae, Scaridae Réunion 521 866 Scombridae, Carangidae Somalia 4 200 6 0 0 0 Lobsters, sharks Lethrinidae, Siganidae, Scaridae, Lutjanidae, Epinephelidae, Scombridae, Kenva 10 000 8 000 - 16 000 Carangidae, shrimp Lethrinidae, Epinephelidae, Siganidae, Mullidae, Lutjanidae, Carangidae, Tanzania 58 000 70 000 (incl. Zanzibar) Scombridae, Clupeidae Mozambique 100 000 - 120 000 Siganidae, Lethrinidae, Sparidae, Labridae, shrimp 70 000 South Africa 5 183 2 1 5 3 Mugilidae, Carangidae, Sparidae, Scombridae, Leiognathidae, Gerreidae (KZN only)

Table 2 An overview of selected artisanal fisheries for several WIO countries (modified from UNEP/Nairobi Convention, 2009).

#### SELECTED COUNTRY PROFILES

The salient features of the fishery sector in several countries of the WIO region and trends in annual landings are presented in Figure 4.







Figure 4 Annual catches (mt) declared to FAO by WIO countries.

# Comoros

This archipelago of four small volcanic islands consists of French-controlled Mayotte and the three islands that constitute the sovereign nation of the Union of the Comoros. These islands only have a small continentalshelf area for artisanal fishing, despite the population's considerable dependence on fish for food security and employment. The islands have no industrial fisheries of their own, although a total catch of 12 000 mt is reported annually. Instead the nation has fisheries agreements with the European Union (EU) authorising 60-70 purse seine or longline vessels to catch about 5 000 mt of tuna, marlin and related species annually. Despite its oceanic setting, artisanal and subsistence fishing is prominent among its half-million people, with about 5 000 fishers operating from dugout canoes (pirogues) or engine-powered vedette boats, using handlines, gillnets and traps. Prominent artisanal landings include species of the families Lutjanidae, Lethrinidae, Epinephelidae and Carangidae. At times these fishers also venture offshore to catch oilfish and the occasional coelacanth. Recently, permits have been issued to some countries that authorise the targeting of sharks with gillnets, raising concern in some circles. Nevertheless, most of the fisheries in the Comoros are considered to be fully exploited.

# Seychelles

An archipelago of 115 islands (with only a few inhabited) just south of the Equator, the Seychelles provides many fishing opportunities, which include industrial, artisanal and recreational. Domestic and foreign industrial fleets land up to 400 000 mt of tuna and billfish every year, creating employment and providing the country with critical foreign exchange. Artisanal fisheries are equally important, with about 1 000 fishers on 400 vessels, providing food security, employment and cultural identity. Smaller boats (especially pirogues) operate in inshore areas, with handlines, traps and seine nets, whereas larger fully decked inboard-engine vessels ('schooners') operate on offshore banks and ledges of the Mahé Plateau. The total catch from the artisanal sector has remained relatively constant since 2000, with landings between 4 000 and 5 000 mt per year. Besides lobster, crab, octopus and sea cucumber, the most popular fishes are diverse species of the families Lutjanidae (particularly Lutjanus sebae and species of Pristipomoides and Aprion), Siganidae, Carangidae and Epinephelidae. In most cases the nation's marine resources are considered to be

moderately to fully exploited. However, the fisheries for Siganidae (rabbitfishes) and especially the much-valued *L. sebae* are of concern and known to be fully exploited.



Figure 5 A French purse-seiner in the Seychelles. © B Everett, ORI/SAAMBR



Figure 6 Offloading tuna to the cannery in the Seychelles. © B Everett, ORI/SAAMBR



**Figure 7** A Seychellois cucumber fisher. © B Everett, ORI/SAAMBR



Figure 8 A typical Seychellois catch. © B Everett, ORI/SAAMBR



Figure 9 A Seychellois fisher. © B Everett, ORI/SAAMBR

# Mauritius

This small island nation includes the islands of Mauritius and Rodrigues, several outer islands (including Agaléga, and those of St Brandon Shoals), the Chagos Archipelago and extensive areas of the Mascarene Plateau. The Republic of Mauritius, with a huge EEZ covering some 1.9 million km<sup>2</sup>, is the custodian of considerable fishery resources, specifically for tuna, deepwater shrimps, and so-called 'distant bank fisheries' that comprise captures of Lutjanidae, Lethrinidae and Sparidae, including a new fishery for *Polysteganus baissaci* (Frenchman seabream). However, the total reported landings are modest, at approximately 6 000 mt per annum, about onethird of the peak in the mid-1990s. This is in part attributable to tight management controls and limits to access imposed by the Mauritian authorities, although there remains concern about the status of the reef fisheries.

Fisheries in Mauritius provide direct and indirect employment to approximately 10 000 people, with nearly 3 000 fishers, mostly engaged in the artisanal fishery. Besides octopus (mostly from Rodrigues), a wide range of shallowwater reef fishes are traditionally taken in the lagoons and fringing reefs using pirogues equipped with handlines, basket traps, large nets, gillnets, harpoons and hand spears. As the lagoon fisheries have been badly depleted, authorities are providing incentives for fishers to either exit the fishery or shift to less vulnerable offshore fisheries employing fish aggregating devices. Recreational fisheries play a significant role in the tourist economy, with about 350 mt of marlin and other gamefish caught annually.

# Réunion

Although politically an overseas department of France, Réunion has its own fishing industry that generates valuable fishery products, primarily tuna and billfishes, for export. This small volcanic island has little continental shelf and a coastline of just over 200 km. Hence, industrial fisheries predominate. These comprise mainly tuna fisheries as well as a semiindustrial billfish operation that produces high-quality smoked products. The total declared industrial catch has decreased to less than 3 000 mt after peaking at 4 600 mt in 2005, largely attributable to a lower tuna catch. An artisanal fishery engages some 500 fishers operating from small outboard-powered boats using handlines, driftlines and trolling. Collectively, these fishers take 300-800 mt of linefish around the main island annually, mainly species of Scombridae but also Carangidae (e.g., Selar crumenophtalmus), Clupeidae (e.g., Sardinella), Mullidae, Epinephelidae and Lutjanidae. Catches have declined in recent years, suggesting that the exploitation levels may have exceeded sustainability. On outer islands and reefs within the large EEZ of Réunion, larger mother ships with attendant dories harvest reef fishes using handlines to catch mainly

Lethrinidae, especially *Lethrinus mahsena* and *L. variegatus*. Réunion also serves as a base for French vessels that undertake trawling for Antarctic fishes around the Kerguelen Is. and lobsters from Saint-Paul and Amsterdam Is.

# Madagascar

With a coastline of 5 600 km and a shelf area of 177 000 km<sup>2</sup>, this large island supports a great diversity of fisheries, ranging from traditional to industrial operations. The total declared industrial catch increased enormously, from 20 000 mt in 1980 and peaking at almost 120 000 mt in 2004, but has since slowed to about 80 000 mt per year. Notable is the offshore tuna fishery, which is accessed by roughly 100 vessels, mostly from the EU. Shrimps are an important but declining resource, with domestic trawlers landing about 8 000 mt per year. Artisanal fisheries involve at least 60 000 fishers, mostly using dugout canoes cleverly rigged with sails and traditional gear such as gillnets, traps and beach seines. Although poorly documented, their landings are believed to be in the region of 75 000 mt annually, and this may rival the industrial catch. These traditional fishers contribute significantly to the nation's food security. Besides catches of fish as food, the harvesting of sea cucumbers, octopus, crabs and aquarium fish for export is popular in some locations.



Figure 10 Pirogues drawn up on the beach (Madagascar). © R van der Elst, ORI/SAAMBR

# Maldives

This tropical archipelago of 26 atolls comprising more than 1 100 small coral islands depends heavily on its fish resources as the main source of protein supply, for employment and export revenue, and as a basis for its tourist industry. The declared industrial catch peaked near 186 000 mt in 2005, but has since declined by one-third, possibly a signal of climatechange impact on the tuna catch. Almost 1 000 domestic fishing boats are operational, the majority being mechanised pole-and-line vessels, but an additional large component of the catch is taken by traditional handlines, often making use of live bait. Skipjack tuna (*Katsuwonus pelamis*) predominates in catches, but other tunas and a variety of oceanic (*Coryphaena hippurus, Elagatis bipinnulata* and *Scomberomorus* spp.) and coastal fishes (Carangidae, Lutjanidae, Lethrinidae and Epinephelidae) comprise about one-third of the total catch. Although many of the islands are remote, improved access has led to increased reef-fish exploitation, and negative catch trends of Epinephelidae (groupers) are of concern. Illegal fishing is also believed to have contributed to declining trends.

# Somalia

The Horn of Africa is an area of huge seasonal upwelling, providing a basis for high concentrations of fishes, including valuable small pelagic species and larger tunas. However, the nomadic lifestyle of the Somali people and political turmoil have thwarted fisheries in the country. For a number of years foreign vessels harvested large quantities of tuna (often illegally), and numerous, now defunct, fish factories were established with foreign aid in anticipation of harvesting small pelagic species. Recently, the fisheries have been dominated by 3 000-4 000 locally based artisanal operators that supply limited local markets with a diversity of linefish. Shark fins are also a popular commodity, with extensive gillnets set parallel along many sections of the coast. Also prominent is an inshore lobster fishery along the northeastern Puntland coast, which uses a variety of gear types to harvest up to 300 mt annually for export. Remarkably, a number of the species of shellfish and finfish harvested in Somalia are similar to species taken in KwaZulu-Natal, South Africa, suggesting an antitropical distribution. Included among these are Pomatomus saltatrix, Pomadasys olivaceum, Dinoperca petersi, Argyrosomus japonicus, the lobster Panulirus homarus and, periodically, Cape fur seals.

#### Kenya

While Kenya's 640-km tropical coastline and its 142 000 km<sup>2</sup> EEZ are rich in fish, only 5% of the national catch comes from the sea; the remainder (especially the highly valued Nile perch, *Lates niloticus*) is taken from Kenya's Rift Valley lakes. Declared industrial marine landings have been stable at just below 7 000 mt for several years. This includes shrimp trawling in Ungwana Bay, where trawler damage to seagrass meadows, a large bycatch, high sea turtle mortality, and conflict with artisanal fishers over access has led to closure of this fishery for several years.

The artisanal sector is much larger, with some 10 000 fishers who land up to 16 000 mt per year. Dhows, outrigger canoes and planked boats are used to fish with gillnets, handlines and tangle-nets. Popular fishing zones include Lamu, Malindi, the Tana River Delta, and the south coast around Majoreni, Gazi and Vanga. Here, fishers target Scombridae, Sphyraenidae, Siganidae, Mugilidae, Carangidae, Lutjanidae and Lethrinidae. After years of growth, the artisanal fishery is now considered fully exploited, with intense fishing and even overfishing reported from coral reefs and seagrass beds in several locations. Reef degradation has further adversely affected the productivity and diversity. However, several sites in Kenya are famed for sport fishing and attract tourist anglers from around the world; although the total landings are not known, this fishery represents an important socioeconomic benefit.





Figure 11 Fisher and dhows (Kenya). © B Everett, ORI/SAAMBR

Figure 12 Going fishing (Kenya). © B Everett, ORI/SAAMBR

# Mozambique

Mozambique is a fishing nation, drawing on the rich resources of Lake Malawi (known as Lago Niassa in Mozambique) as well as its extensive coastline and wide continental shelf. The fishing industry has been one of the country's largest generators of foreign revenue, with shrimp contributing up to 50% of the export value in the late 1990s. Annual declared industrial landings increased to more than 70 000 mt in 2006, following decades of reported landings of approximately 30 000 mt (this dramatic increase is possibly attributable to the recent inclusion of data from artisanal landings, however).

While industrial fishing at various levels contributes significantly to the economy, it is the artisanal fisheries that provide a livelihood for more than 70 000 fishers and their families, thereby supplying food to a large part of the population. Some 700 landing sites and 15 000 artisanal vessels are in operation, mostly wooden, non-motorised canoes and similar craft. Handlines, cast-nets, beach seines, gillnets, trolling lines, traps and cages are popular gear. Total landings vary but exceed 100 000 mt annually, encompassing a wide range of species, from coral-reef fishes, estuarine species, and inshore pelagics such as Carangidae and Scombridae. Sharks and rays are an increasingly popular resource for both food security and fins for export. Apart from fishes and shrimps, other important exploited marine resources near urban centres include invertebrates such as crabs, clams and sea urchins.



Figure 13 Mozambique artisanal fisher. © B Everett, ORI/SAAMBR



Figure 14 Fishing boat launch in Bazaruto (Mozambique). © R van der Elst, ORI/SAAMBR

Recreational fishing (mainly by South African tourists) has increased significantly over the years, but this has occurred with low levels of compliance, many reports of excess catches, and conflicts with local operators. Research indicates that the majority of fisheries in Mozambique are moderately to fully exploited, so that conservation measures are being progressively implemented.

# Tanzania

Although access to the Rift Valley lakes provides Tanzania with high catches of freshwater fishes, marine fish landings are persistently high as well, at approximately 70 000 mt per year, inclusive of some 20 000 mt taken by Zanzibar fishers. Virtually the entire marine catch is taken by artisanal fishers, who now number more than 58 000, and many are associated with coral-reef ecosystems. Catches are landed from dhows and outrigger canoes (ngalawas), using gillnets, beach seines, handlines, fixed traps and basket traps. Technically, beach seines are illegal yet popular, while the use of poison, dynamite, spear guns and other destructive gear persists. A wide variety of species are landed, ranging from reef fishes to small and large pelagics. Sharks, rays, lobsters, octopus and squid are also freely taken. Industrial fisheries often take the form of joint ventures between Tanzanian and foreign companies. Recently, deepwater tuna fishing has been established in Zanzibar, based on a projected potential of 8 000-24 000 mt per year from 70 licensed vessels. Compliance is generally poor, with sea cucumbers and several species of fish overexploited, including



Figure 15 Ngalawas drawn up on the beach, Pemba Island (Tanzania). © R van der Elst, ORI/SAAMBR



**Figure 16** Community fishing dhow, Zanzibar (Tanzania). © R van der Elst, ORI/SAAMBR



Figure 17 Trawlers off Dar es Salaam (Tanzania). © B Everett, ORI/SAAMBR

sharks for the fin trade. While clearly important in Tanzania, artisanal fishing has contributed to considerable degradation of the marine environment and reduced overall catches.



Figure 18 Zanzibar artisanal fish market (Tanzania). © R van der Elst, ORI/SAAMBR



Figure 19 Prawn trawler catch, KwaZulu-Natal, South Africa. © J Robey, ORI/SAAMBR

# South Africa

Although South Africa is a significant fishing nation, with annual landings that can attain 1 million mt, only about 1 000 mt is taken in WIO Area 51. These include shrimps, deepwater lobster, swordfish, and a range of bony fishes taken by small-scale operators. Although the purse-seine fisheries for small pelagic species (anchovy, pilchard, round herring and juvenile horse mackerel) are traditionally located on South Africa's west coast (in the southeastern Atlantic), recent landings have moved further east, closer to Area 51. A range of small, engine-powered boats are used to access sea conditions that can be challenging. Mainly hook-and-line gear is used, but beach seines are used at times, especially during the winter sardine run of *Sardinops sagax*. Recreational fishing is also especially important along the coastline.

While the South African fisheries are generally wellmanaged, the status of many endemic species has fallen below acceptable threshold levels. This includes especially the Sparidae and some of the Scianidae. The trend in WIO catches taken in KwaZulu-Natal, for example, has steadily declined since the peaks attained in the early 1980s.

# SPECIFIC CHALLENGES FOR WIO FISHERIES

Many of the fisheries in WIO share similar problems. The growing need for food security and employment in generally low-income locations are primary drivers that escalate fishing pressure to or beyond levels of sustainability. Globalisation is also a factor, with the industrial fisheries of many nations reaping an ever-growing harvest in the region. Ultimately, it is a will to implement and enforce scientifically based management policies that is required for the region to meet its development challenges while protecting its biodiversity. There are several specific issues:

- *Fishing capacity.* There are few limits on fishing volume in the region and many of the fisheries are in essence open access. Clearly, this impacts on the entire fish resource and lowers the productivity of individual fishers. Fishing is often an 'employment of last resort,' but overexploitation is a collective threat to all stakeholders. Consequently, a major challenge is to ensure that fishing effort is maintained within levels of sustainability, even while access to fisheries resources becomes equitably allocated.
- *Fisheries data.* Notwithstanding the value of FAO statistics, most fisheries are poorly documented and data are incomplete. In more than half of the WIO fisheries the existing level of monitoring and reporting is considered to be inadequate for management, while approximately only 4% of the fisheries have been scientifically fully assessed.
- **Destructive fishing practices.** In many cases the fishing gear used is destructive to the environment. For example, trawling gear often degrades the biodiversity of traditional fishing grounds, including the ecosystems on which fishes depend for nurseries and reproduction. Most disconcerting is that fishing with dynamite is still practised, albeit illegally, in several areas. The use of inappropriately small mesh-sizes,

gillnets, beach seines and fish poisons also raises concern. Trampling in shallow, intertidal areas while collecting octopus, shellfish, sea cucumbers and seaweeds also causes physical damage to sensitive habitats and fishing grounds.

- *Habitat destruction.* Of ever-growing concern is the physical alteration of marine habitats. Climate change, coral bleaching, mangrove-forest clearance and seagrass-meadow destruction all pose threats to essential habitats that are being transformed or destroyed with consequent impact on the ichthyofauna.
- *Inadequate enforcement.* While all countries of the WIO region have domestic fishing regulations, ensuring compliance is a huge and costly task, especially as most fishing activities are carried out in remote areas. A lack of funds and technical support frequently hampers the effective management of marine fisheries. Only 21% of the 211 fisheries surveyed in WIO are considered to have an adequate management plan.

#### MANAGEMENT APPROACHES

Fisheries management is always challenging. Dealing with people in a mainly open-access resource, large numbers of dispersed operators, and, in WIO, a very large number of species, with many not adequately studied, continually complicates fisheries management. Each of the WIO nations has its own domestic policy and regulations to manage the harvesting of fishes in its waters. However, these domestic fisheries regulations alone may be inadequate as many species are common to more than one country and straddle national borders. Many other species migrate freely across different areas of WIO, so that a collective approach for management is required.

Such an approach can be achieved, in part, through the programmes of the FAO. As signatories to the United Nations Convention on the Law of the Sea (UNCLOS), the WIO nations participate in regional fisheries management through the FAO's activities. Besides the key annual submission of catch information, the FAO provides support in a number of ways. For example, in order to ensure a fair distribution of shared species among nations, several conventions, agreements and structures have been established. Thus, there are FAOsupported regional fisheries management organisations (RFMOs). For WIO, there are the Southwest Indian Ocean Fisheries Commission (SWIOFC) and the Indian Ocean Tuna Commission (IOTC). Both these bodies assess annual trends and make recommendations to their member countries about levels of exploitation.

Several international agreements and protocols have been adopted by most countries in the region. Specifically, the Southern Indian Ocean Fisheries Agreement (SIOFA) for stocks outside of any EEZ, the 1995 United Nations Agreement for the Conservation and Management of Straddling Fish Stocks and Highly Migratory Fish Stocks, and the 1995 Code of Conduct for Responsible Fisheries are all designed to guide best fisheries management practices. In recognition of the important link between the environment and fisheries, the FAO promotes an ecosystem approach to fisheries management (EAFM) framework, which is designed to consider fisheries impacts on the environment as well as the social implications. For countries in the Southern African Development Community (SADC) region there is the SADC Protocol on Fisheries, an instrument that spells out a common approach of member countries to ensure a sustainable approach to fishing in their collective waters. A further notable instrument is the Nairobi Convention (UNEP/Nairobi Convention Secretariat 2009), which is a UNEP-hosted structure that promulgates protocols relating to land-based sources of pollution, and protection of coastal biodiversity and its resources. While the level of compliance of these important instruments by WIO countries is variable, they do play an increasing role in guiding management approaches that will ensure greater levels of sustainability and better equip authorities to deal with diverse management problems, including climate change.

# CONCLUSIONS

There can be some consolation that WIO fisheries may be generally in a better state than the fisheries of other regions, despite the several negative trends that have emerged. However, there is no room for complacency as the need for implementation of wise approaches to fisheries management is more pressing than ever. Currently, several large regional programmes are contributing to transboundary collaboration, which bodes well for the future.

**Note:** There are many inconsistencies in the modestly available statistics, which also change over time. Much of the information presented here is based on the sources given below. It is suggested that these websites are visited periodically to obtain updated information: http://www.fao.org: for global fisheries statistics. http://www.unep.org/ http://www.wiofish.org: for updated information on different

http://www.wiofish.org: for updated information on different WIO fisheries.

# THE OCEANOGRAPHY AND FISHERIES ALONG THE WEST COAST OF INDIA AND AT LAKSHADWEEP

Elayaperumal Vivekanandan

# **INTRODUCTION**

Fisheries along the west coast of India are unique in the Indian Ocean with respect to the diversity of resources, catch volume, magnitude of fishing activities, economic returns and livelihood opportunities provided. Marine fisheries along the eastern Arabian Sea are among the largest in the Indian Ocean, providing vast food security and large contributions to export earnings and the gross domestic product of India. Long timeseries of fisheries statistics on catches and fishing effort with high spatial and temporal resolution are available and provide insight into the development issues facing the sector.

The west coast of India extends 3 335 km and is characterised by a wide shelf in the north (about 250 km wide off Mumbai), which narrows to some 60 km wide off Kochi in the south (Figure 1). However, while nearly 80% of the continental shelf area lies along the northwestern coast, more than half of the fisheries catch comes from the southwestern coast.



Figure 1 Map of India showing extent of the continental shelf.

Table 1 Fishing areas in the EEZ along the west coast of India.

STATE/UNION TERRITORY	COAST LENGTH (km)	CONTINENTAL SHELF AREA (000 km²)
Kerala	590	40
Karnataka	300	25
Goa	104	10
Maharashtra	720	112
Gujarat	1 600	164
Daman & Diu	21	0.1
TOTAL	3 335	351.1

From an oceanographic and fisheries perspective, the west coast of India can be divided into two geographical areas, north and south of 15° N. The region is generally highly productive; the main features contributing to this productivity are an upwelling during the summer monsoon, and a cooling of the northern Arabian Sea during winter. These two processes bring large amounts of nutrients into the upper water layers, which enhance primary productivity and thus fundamentally contribute to the fisheries. Marine fish landings along this coast have increased in the last six decades and continue to rise (as also along the east coast of India and in the Bay of Bengal). However, the threats to sustainability with the increase in fisheries production masks several issues of concern. Fishing and other anthropogenic impacts on finite resources.

#### METEOROLOGICAL AND OCEANOGRAPHIC FEATURES

The Arabian Sea is influenced by the southwest and northeast monsoons. Nearly 80% of the rainfall in the region occurs during the southwest monsoon, which lasts from June to September. The northeast monsoon lasts from October to January. Seasonality in the Arabian Sea can be clearly divided into four seasons, albeit with some variability from one year to the next. This is also reflected in seasonal variations in productivity that differ between regions. During the spring inter-monsoon period (March–May), a transition from winter to summer, the entire Arabian Sea generally has very low primary productivity (14-21 mmol C m<sup>-2</sup> d<sup>-1</sup>) and low levels of chlorophyll (~45 mmol C m<sup>-2</sup>). A more or less similar scenario prevails during September-October, the transition from summer to the winter monsoon. During these periods the waters have an increased sea surface temperature (SST) of ~28 °C, shallow mixed-layer depths of ~20-30 m, and strong stratification. Many nutrients, especially nitrate, the main limiting factor in primary production, are then undetectable in the surface waters. With the onset of the summer monsoon in June, the situation changes considerably. The current, which flows northward along the coast during the northeast monsoon, reverses and flows southward, whereas the wind blows from the west and west-northwest. During the southwest monsoon, a strong southward drift, especially along the southwestern coast, is prominent in the upper water layers; and there is a northward countercurrent along the lower boundary of the thermocline, but it is comparatively weaker and discontinuous.

## Southwestern coast

The prime oceanographic feature on the southwestern coast is an upwelling during the southwest monsoon. During July–August the surface mixed-layer becomes more or less obliterated, with the temperature decreasing to 26.5 °C and the oxygen-deficient layer reaching upwards, even to the surface. This indicates a coastal upwelling, which extends from Kanyakumari to Karwar (~8–14° N), though the intensity is much less south of Quilon (~9° N). It has also been noticed that this upwelling starts at the southern tip of the west coast in late May/early June and then extends northwards with the progress of the southwest monsoon season. These conditions prevail until late August/early September, when the upwelling subsides. No major upwelling occurs north of Goa, at ~16° N.

While the strong wind along the shore induces upwelling in the south, the cross-shore component of the wind in the north modifies the density structure of the water surface in the northern part to reduce the intensity of the upwelling (Muraleedharan & Kumar 2014). The upwelling increases the nutrient content, thereby increasing the productivity of the region, and at its peak the bloom is on par with some of the most fertile waters in the world's oceans. The productive value of upwelling is reflected in the total zooplankton biomass, which in turn is associated with high-volume fisheries along this coast during the southwest monsoon (Banse 1959, and others).

The southwest monsoon is also the period when mud-banks are formed at places along the southwestern coast of India, especially along the Kerala Coast. The mud-bank extent is inconsistent, varying from 5 to 10 km in length and normally 2 to 5 km outwards from the shore. The source of mud for these banks is the sea bottom and the huge, adjoining Vembanad Lake (the longest in India), which empties into the sea at ~10° N. The mud-banks are not formed every year, but when they do, they remain for 2–3 months during June to August. The mud, which contains a high phosphorus content, is largely composed of very fine particles, with a relatively high organic carbon content. Plant pigments and carbohydrates are present in low concentrations, consequently rendering the mud of low caloric and poor nutrient value. The advantage of mudbank formation from a fisheries perspective is that organisms, especially the shrimp *Parapenaeopsis stylifera*, aggregate and become trapped in the calm, shallow muddy waters, resulting in plentiful catches of this high-value species.

# Northwestern coast

Notwithstanding that convectional upwelling does not occur along this coast, the waters of the northwestern coast also have a fairly high abundance of phytoplankton and zooplankton during the southwest monsoon, probably as a result of land/ river run-off. An additional feature of this segment of the coast is the winter cooling effects of the northern Arabian Sea (north of ~15° N). From November to February, the cold, dry continental air blowing over the northern Arabian Sea cools the sea surface to ~24 °C, and the denser surface waters sink, leading to deep mixed-layers below 100 m. The weak winds do not generate convective mixing in summer yet do inject nutrients into the surface layers, thus leading to the higher productivity.

In summary, the seasonally higher productivity in the eastern Arabian Sea is mainly through upwelling during summer, to  $\sim 15^{\circ}$  N along the southern coast, and cooling north of  $\sim 15^{\circ}$  N during winter. In both cases the photic zone obtains some nutrients from the deeper layers, which results in higher productivity.

# **FISHERIES PROFILE**

The favourable meteorological and oceanographic conditions along the west coast of India have resulted in highly productive, high-volume fisheries, making India by far the largest fish producer in the WIO, contributing over 46% of total production. However, the number of fish-landing centres along the west coast has not been constant over recent decades, as new ones have been opened and others became defunct; even so, they remain at around 640. With an increasing population, the number of active fishermen involved in actual fishing on the west coast has increased threefold over five decades, from 118 000 in 1961/62 to 350 000 in 2011. Meanwhile, the catch per active fisherman has persisted at around 5 metric tons (mt) per year (Table 2).

Table 2 Fisheries profile of the west coast of India during different time periods (CMFRI 2006, 2012).

YEAR	LANDING CENTRES	ACTIVE FISHER POPULATION (000s)	FISH CATCH (mt/fisher)	NUMBER OF CRAFT		
				MECHANISED	MOTORISED	NON-MOTORISED
1961–62	NA	118	5.1	5 456	0	40 147
1973–77	639	171	5.1	6 030	0	41 570
1980	727	218	4.6	14 304	0	67 774
1998	NA	NA	NA	32 926	22 800	24 656
2005	582	326	4.7	37 626	30 200	28 644
2010	594	350	5.5	41 801	30 150	13 961

NA – not available.

Table 3	Number of mechanised boats in the fishery along the west
coast of	India (CMFRI 2006, 2012).

	SOUTHWE	ST COAST	NORTHWEST COAST		
CRAFITYPE	2005	2010	2005	2010	
Trawler	7 327	7 359	12 536	17 732	
Gillnetter	1 729	672	5 083	7 486	
Dolnetter	0	0	6 905	6 607	
Liner	38	29	257	10	
Ring-seiner	480	495	0	0	
Purse-seiner	755	778	160	435	
Others	635	169	1 721	29	
TOTAL	10 964	9 502	26 662	32 299	

Three types of craft are operated in the fishery. The larger boats with inboard engines (mechanised boats) are 13-20 m in length, with engines of 100-400 hp (Figure 2). The number of these boats in operation increased from 5 456 in 1961/62 to 41 801 in 2010; the increase, especially of trawlers, was ~40% along the northwestern coast between 2005 and 2010 (Table 3). Fishing efficiency also increased substantially; with the increase in the size and engine power of boats, extended fishing trips to distant fishing grounds (lasting five days or more) became possible. These boats employ a variety of gear, of which trawls are the most prominent. A second type of craft is smaller, with outboard engines of 10-40 hp (Figure 3). Outboard engines were introduced in the mid-1980s and became popular among small-scale fishermen, particularly as the government provided subsidies for the engines to empower these fishermen. In 2010, more than 30 000 boats operated along the northwestern coast (Table 3). Consequently, the number of a third type and non-motorised craft, which depend on the wind, decreased substantially.





Figure 2 Typical boat with inboard engine. © E Vivekanandan

Figure 3 Boat with outboard engine. © E Vivekanandan



Figure 4 Ring-seine operation, Karwar, Karnataka, India. © E Vivekanandan

A wide variety of target-specific and non-specific fishing gear is used along the entire coast, and these can be broadly classified as trawls, gillnets, lines, seines and bag nets. Improvements are regularly made to the gear to achieve better efficiency. High-opening trawlnets, monofilament gillnets of a wide range of mesh sizes, longlines, purse seines, ring seines and dolnets are intensively operated (Figure 4).

# **FISH LANDINGS**

The Central Marine Fisheries Research Institute (CMFRI) in Kochi has collected data on fish landings and fishing effort from all landing centres in India since 1956, using stratified multistage random sampling. The CMFRI database shows that the annual average landings have increased from 584 235 mt



**Figure 5** Annual average landings (million mt) along the west coast of India during 1956–2011. Source: CMFRI



Figure 6 Contribution to west-coast landings (%). Source: CMFRI



Figure 7 Sharks landed at Azheekal beach, Cherthala, Kerala (India). © E Vivekanandan

during the period 1956–1959 to 1 862 375 mt during 2000–2011 (Figure 5), a threefold increase in 56 years. The contribution of the two major subregions has fluctuated over the years, but the overall contribution of each over the period was almost equal, with the southwestern coast contributing 53% of the total (Figure 6).

The two regions are rich in biodiversity. The CMFRI records show that at least 800 species, including elasmobranchs, teleosts, crustaceans, cephalopods, gastropods and bivalves, occur in the catches, with some 200 species contributing to a minor or major fishery in one subregion or the other. A single trawl haul, on average, lands ~40 species. In addition, several non-targeted groups, including jellyfish, echinoderms and other invertebrates, sea snakes, and protected animals such as turtles and dolphins, are also incidentally caught. CMFRI publishes the annual landing statistics by aggregating landings into 80 major species/groups. Analysis of data for the years 2004-2011 shows that the Indian oil sardine, Sardinella longiceps, tops the list and contributes 19% to the west-coast landings (Table 4). This is followed by another small pelagic species, the Indian mackerel, Rastrelliger kanagurta, at 7.4%. Penaeid and non-penaeid shrimps together contribute 13.3%.

 Table 4
 Annual average landings of top 10 species/groups on the west coast of India during 2004–2011.

	LANDINGS			
SPECIES/GROUP	mt			
Oil sardine	354 548	19.0		
Indian mackerel	137 213	7.4		
Penaeid shrimps	125 454	6.7		
Non-penaeid shrimps	122 618	6.6		
Ribbonfish	116 179	6.2		
Threadfin breams	107 470	5.8		
Sciaenids	93 641	5.0		
Bombay duck	71 596	3.8		
Cuttlefish	51 317	2.8		
Squids	49 546	2.7		
TOTAL	1 229 582	66.0		



Figure 8 Oil sardine haul, Tamil Nadu coast (India). © E Vivekanandan

Though the two subregions contribute equivalent quantities to fish landings, the composition of the landings differs substantially between them. Whereas small pelagic fishes, such as the oil sardine (34.5%), Indian mackerel (12%), whitebait (3.5%) and scads (3.1%), contributed 58.1% to the landings on the southwestern coast, small pelagics amounted to only 21% of northwestern-coast landings (Figure 9). The northwest subregion lands predominantly bottom-dwelling species, consisting of small demersal fishes (threadfin breams, small sciaenids, lizardfishes and soles) and larger fish species (catfish, pomfrets, large sciaenids, groupers and snappers), which together contributed 31.3% to the total landings. Crustaceans, such as non-penaeid shrimps (13.8%), penaeid shrimps (8%), crabs, lobsters and stomatopods, together contributed 24.7% to northwestern-coast landings.



Figure 9 Contribution (%) of fish groups to southwestern- and northwestern-coast catches. Source: CMFRI

The list of the five most-caught species differs between the southwestern and northwestern coasts (Table 5). The contribution of the top five species for the southwestern coast accounted for 62% of the total catch, whereas that of the northwestern coast totalled only 46%, revealing the dominance of only a few species/groups along the southwestern coast compared with along the northwestern coast. It is evident that each species/group has a strong habitat preference, determined mainly by the oceanographic features of each region (Vivekanandan & Krishnakumar 2010). The ecosystem of the southwestern coast is influenced by seasonal upwelling and high productivity, and thus is the domain of small pelagics with generally short lifespans and rapid generational turnover. By contrast, the northwesterncoast ecosystem is driven by the sinking of higher-density, nutrient-rich masses of water during the winter months, which promotes an abundance of demersal species.

The major small pelagic contributors to northwestern landings are Bombay duck (*Harpadon nehereus*, 8.2%), goldspotted grenadier anchovy (*Coilia dussumieri*, 2.4%) and Indian mackerel (*Rastrelliger kanagurta*, 2.1%). *Harpadon nehereus* forms the largest single-species fishery along the northwestern coast, and is dominant in the Bombay duck– *Coilia*–non-penaeid shrimp (mainly *Acetes indicus*) complex. This complex constitutes a unique predator–prey relationship.

Bombay duck inhabits waters to the 70 m isobath, and is inconspicuous or totally absent along the southwestern coast. However, the species forms a minor fishery at the same latitudes in the Bay of Bengal. Devaraj (1987) considered the high-tidal range, characteristic of the northwestern coast between 18° N and 23° N (as at the corresponding latitudes along the northeastern coast in the Bay of Bengal) as the major factor influencing the distribution of Bombay duck. The northwestern coast, especially the Gulf of Khambhat (22° N, 72° E) experiences a tidal range of ~10 m; the tidal range along the northeastern coast is also high, at ~6 m.

Table 5Top five species (%) in respective region's catch during2004–2011.

RANK	SW COAS	т	NW COAST		
	SPECIES/GROUP	Catch %	SPECIES/GROUP	Catch %	
1	Oil sardine	34.5	Non-penaeid shrimps	13.8	
2	Indian mackerel	12.0	Sciaenids	9.4	
3	Threadfin breams	6.5	Ribbonfish	9.2	
4	Penaeid shrimps	5.3	Bombay duck	8.2	
5	Whitebaits	3.7	Penaeid shrimps	5.4	
TOTAL		62.0		46.0	

Neither strictly pelagic nor demersal, Bombay duck effectively uses the high tidal range to conserve energy while foraging on non-penaeid shrimps and *Coilia*, which are abundant in the habitat. The species is harvested with a specialised dolnet, which is a fixed bag net that operates to a depth of 40 m, depending upon the turn of the tides since a strong tidal current is essential for operation of the net. The mouth of the net is always held in the direction of the tide so that the net functions as a filter to retain the fish, while the strong tidal current prevents the fish from escaping from the net.

To gain an understanding of trends in landings, catches of major species/groups were compared between 2004–2007 and 2008–2011. For this analysis, groups that showed <25% difference in landings for the two periods were considered as 'no change'. Of 57 species/groups that were investigated, 36 did not show any change (Table 6), 17 showed an increase in landings, and 4 (namely, hilsa shad *Tenualosa ilisha*, goldspotted grenadier anchovy *Coilia dussumieri*, adalah *Psettodes erumei*, and several species of gastropods) showed a decrease in landings over the total period.

**Table 6** Status of fish groups in the landings along the west coast of India; the values are percent difference in the landings during 2008–2011 from that of 2004–2007; difference between –25% and +25% is considered as 'no change'.

SPECIES/GROUP	NO CHANGE	INCREASE	DECREASE
Sharks	-3		
Skates	-10		
Rays	-12		
Eels	-22		
Wolf herring	13		
Oil sardine	4		
Other sardines	10		
Other shads	-16		
Thryssa	0		
Bombay duck	-11		
Halfbeaks	12		
Groupers	11		
Emperors	20		
Threadfins	-21		
Sciaenids	23		
Ribbonfish	-9		
Horse mackerel	9		
Scads	-2		

SPECIES/GROUP	NO CHANGE	INCREASE	DECREASE
Leather-jackets	9		
Other carangids	24		
Silverbellies	-18		
Black pomfret	3		
Silver pomfret	14		
Chinese pomfret	4		
Seerfish	-10		
Kawakawa	-10		
Mullets	-24		
Unicorn cod	-16		
Soles	6		
Penaeid prawns	0		
Non-penaeid prawns	24		
Lobsters	-2		
Stomatopods	5		
Bivalves	14		
Cuttlefish	20		
Octopus	20		
Catfish		69	
Whitebaits		115	
Other clupeids		57	
Lizardfish		65	
Flyingfish		77	
Snappers		76	
Threadfin breams		28	
Other perches		28	
Goatfish		56	
Whitefish		109	
Indian mackerel		56	
Skipjack tuna		286	
Longtail tuna		37	
Billfish		180	
Barracudas		26	
Crabs		34	
Squids		65	
Hilsa shad			-77
Coilia			-25
Indian halibut			-27
Gastropods			-50

#### LAKSHADWEEP

Lakshadweep is a group of scattered islands in the Laccadive Sea, between 08°00' N and 12°30' N and 71°00' E and 74°00' E, about 200–400 km off the southwestern coast of India. The archipelago consists of 11 inhabited and several uninhabited islands, islets and submerged reefs. The surface area of the islands is just 32 km<sup>2</sup>, but being oceanic their continentalshelf area is about 4 336 km<sup>2</sup>, and the lagoons have an area of 4 200 km<sup>2</sup>, offering rich biotic diversity and potential. Furthermore, the area of the territorial waters (or exclusive economic zone, EEZ) around the islands covers an area of some 400 000 km<sup>2</sup>, offering immense fisheries potential.

CMFRI reported 601 fish species in Lakshadweep waters in 2011. Of this total, more than 300 species belonging to 35 families are recognised as ornamental fishes. The major fisheries resources of the islands also include oceanic species such as tunas, billfish, pelagic sharks, marlins, sailfish, and other various groups such as barracuda, seerfish, snappers, breams, carangids, rays and octopus. Twenty-one species are utilised as bait (e.g., species of *Spratelloides, Apogon* and *Chromis*), and ornamental fishes are exclusive to the islands.

The live bait fishery is important for the capture of tunas in a traditional pole-and-line fishery in the islands. This major fishery accounts for more than 97% of the tuna landings, of which skipjack tuna (*Katsuwonus pelamis*) makes up 86%, yellowfin tuna (*Thunnus albacares*) 12%, frigate tuna (*Auxis thazard thazard*) 1%, and kawakawa (*Euthynnus affinis*) 1%. For the past 15 years, about 300 pole-and-line boats have operated in the islands, in addition to an estimated 1 401 nonmotorised, 1 249 motorised and 10 mechanised boats. The pole-and-line boats are 7.5–10.5 m in overall length and make one-day fishing trips.



**Figure 10** Pole-and-line fishing (Lakshadweep Archipelago). © E Vivekanandan

The annual potential yield of tunas is estimated at 50 000 mt, and potential yield of all other fish collectively at an additional 50 000 mt, as compared with the current yield of 15 000 mt, which amounts to only 15% of the potential yield. The current yield of tuna (about 10 000 mt) is small compared with that from the Maldives, at 148 000 mt, and from Sri Lanka, at 27 000 mt (CMFRI 2011).

## **CLIMATE CHANGE**

Global warming and the consequent changes in climatic patterns will have strong impacts on fisheries with farreaching consequences to the food and livelihood security of a sizeable portion of the global population. Any changes in sea temperatures, acidity, salinity and dissolved oxygen concentration would significantly influence the metabolism, growth rate, productivity and reproduction of aquatic organisms, as well as their susceptibility to diseases and toxins. Some species and ecosystems may gain, but others will lose. This in turn will impact ecosystem structures and functions.

Data on sea surface temperature (SST) collected from the International Comprehensive Ocean-Atmosphere Data Set (ICOADS) for the years 1961–2009 for the southwestern coast show that the number of decadal SST anomalous months increased from only 19 during 1960–1969 to 53 during 2000– 2009. Anomalies in the multivariate ENSO index for variables observed over the tropical Pacific also increased over these periods. The SRES A2 scenario prediction generated by the Intergovernmental Panel on Climate Change (IPCC) shows that the annual average SST along the southwestern coast may increase from 28.5 °C in 2000 to 31.5 °C by 2099.

The annual mean SST trend shows that the northwestern coast has also warmed, from 26.04 °C in 1985 to 26.1 °C in 2005, at the rate of 0.03 °C per decade. The SRES A2 scenario also indicates that the annual average SST along the northwestern coast may increase from 26.05 °C in 2000 to 29.5 °C in 2099. By the turn of this century, the annual average SST on the northwestern coast (between 15° and 24° N) will be higher than the current SST in the southern latitudes of 8°–14° N.

An interesting climate-driven change in the distributional boundary of the Indian oil sardine (*Sardinella longiceps*) has been observed in the last 20 years (Vivekanandan 2011). This sardine is a small pelagic fish of tropical coastlines and forms the basis of a massive fishery — perhaps the largest exploited stock in the Indian Ocean. It is a cheap source of protein and forms a staple, sustaining and nutritious food for millions of coastal inhabitants. The species has a rapid populationdoubling period of 15–24 months; it is known to have a restricted distribution between latitudes 8° and 14° N and longitudes 75° and 77° E (the Malabar upwelling zone), where



Figure 11 Oil sardine landings (mt) along the northwestern coast, 1981–2011. Source: CMFRI

the annual average SST is 27–29 °C; and, its distribution and abundance are governed by the vagaries of oceanic climate conditions. Until 1985, almost the entire catch of oil sardine was taken from the Malabar upwelling zone; north of 14° N the catch was either very low or there was no known catch at all (Figure 11). In the last two decades, however, the catches from latitudes 14°–23° N consistently increased, reaching an annual average of 14 152 mt during 2008–2011. The surface waters of the Indian seas have warmed by 0.03 °C per decade, and a warmer wedge (27–28.5 °C) of surface waters has expanded north of 14° N, enabling the oil sardine to extend its range further north.

It was also found that the catches of oil sardine from within its original home range (the Malabar upwelling zone) have not decreased, indicating a distributional extension and not a distributional shift. The annual average catch has increased over time and reached 354 548 mt during 2004-2011 in the upwelling zone off the southwestern coast. Time-series data on different climatic and oceanographic parameters gathered from different sources show that, corresponding to the annual SST, the annual average scalar wind speed increased from 3.58 m/s to 6.05 m/s during 1967-2007 along the southwestern coast (Manjusha et al. 2010). The zonal wind speed during the southwest monsoon season (June-September) increased from 3.34 m/s in 1967 to 5.52 m/s in 2007, with the speed exceeding 5 m/s in several years during 1992-2007. The monsoonal coastal upwelling index (CUI) nearly doubled, from 485 m³/s to 713 m³/s, during 1997-2007. This substantial increase in CUI elevated the chlorophyll a concentration, from 4.54 mg/m<sup>3</sup> in 1997 to 13.85 mg/m<sup>3</sup> in 2007, during the monsoon season. The increasing CUI and chlorophyll a concentration sustained an increasing catch of oil sardine, especially during the post-monsoon season (Vivekanandan 2011). The peak spawning activity of oil sardine is during the southwest monsoon. If wind speed and direction are ideal, the larvae are dispersed to favourable destinations where they find enough food and fewer predators. Egg development and

growth of transforming larvae are rapid, and the fish normally reach about 10 cm in length in 3 months. Individuals, which spawn during the southwest monsoon, are thus recruited to the fishery during the post-monsoon period.

This trend in oil sardine catches indicates that, in this instance, the current change in climate can be beneficial to production of small, pelagic, herbivorous fish (Vivekanandan 2011). However, the relationship between climatic and oceanographic variables, and fish distribution, abundance and biological characteristics has to be monitored continuously. If the SST increases in the southern latitudes beyond the physiological optimum of a fish species, and if other oceanographic variables likewise change unfavourably, it is possible that populations may not be able to adapt but instead will shift towards northern latitudes.

As the southern latitudes are warmer, the northwesterncoast fisheries may benefit from positive impacts for several years. Movement of fish from southern latitudes, as in the case of the oil sardine in the last two decades, would be an additional resource on the northwestern coast. Preliminary analysis using the Ecopath mass-balance trophodynamic model shows that incursion of oil sardine to the northwestern coastline would not immediately affect the abundance of other fish stocks in the area (CMFRI 2009). The ecosystem model also shows that the abundance of several other stocks would likewise increase. However, stress may be placed on the dominant species - Bombay duck Harpadon nehereus and goldspotted grenadier anchovy Coilia dussumieri. The distribution of these two species is restricted to northern latitudes of the Indian coast. As the northernmost boundary of the Indian coastal region is landlocked, these species cannot move further north. Because the region experiences a large tidal variation, up to 10 m in the Gulf of Khambhat (22° N), it is not clear how a rise in sea level will impact on tide levels, which may in turn affect the tide-dependent dolnet fisheries for Bombay duck. Moreover, the whale shark Rhincodon typus, which is a protected species, migrates from southern latitudes and aggregates off Saurashtra (in Gujarat) during December to March each year. Studies conducted elsewhere indicate that the migratory patterns of such large marine animals will be altered as a result of warming and acidification of the oceans.

In the long term, changes in ecosystem structures and functions and predator-prey relationships are likely. There is a need to assess the potential positive or negative impacts of additional species coming into the region on fisheries. Development of predictive models for fish abundance for different greenhouse-gas-emission scenarios is essential, because new fisheries will emerge depending on the adaptive capacity of species to different levels of these emission. Hence, it is important to consider additional post-harvest infrastructure requirements for species new to a locality, changes in craft-gear combinations, and increases in the entrepreneurship of fishermen. It is also imperative to monitor invasions of new fish parasites and diseases from southern latitudes to understand changes in host-parasite specificity.

### FISHERIES MANAGEMENT

The increase in marine fish production along the west coast of India (and along the east coast as well) is contrary to the trend in several other regions of the world, where catches are either decreasing or are stagnant. However, the increase in catches along the Indian coasts masks the following dynamics:

- The increase in catches is a result of increasing fishing efficiency and extension of fishing grounds. When all fishing grounds have become fully exploited, the stocks will decline and elevated catches cannot be sustained.
- 2. The current catches (annual average 1.86 million mt for 2008–2011) along the west coast are approaching the estimated potential yield of approximately 2.4 million mt (Ministry of Agriculture 2001), and hence any further increase in fishing effort and fish catches should be viewed with caution.
- 3. Evidence is accumulating that stocks of catfish, whitefish and sharks are declining, but such changes are not reflected in recorded catches.
- 4. Decline in the composition and quality of fish stocks has been observed, as is evident from the exploitation of juvenile fishes and a reduction in the average size of the fish caught.

In India, fisheries within 12 nautical miles off the coast are managed by the state governments, and offshore and oceanic fisheries beyond 12 nautical miles are managed by central government. In the past 50 years, marine fisheries operations in the region have developed from a subsistence activity, practised almost exclusively by traditional fishermen, to the status of a capital-intensive industry. In the early years, the emphasis of fisheries management was on increasing production by training fishermen in advanced fishing methods, and encouraging introduction of efficient fishing craft and gear by providing loans and subsidies. Support from research and development institutions, coupled with investments by private entrepreneurs in an export trade, supplemented government efforts to expand the growth of the fishery sector.

In recent years, the government has attempted to regulate the proliferation of fishing activities through various measures, such as limits on the number of boats, seasonal bans on fishing, mesh-size regulations, bans on destructive gear, declarations of Marine Protected Areas and protection of endangered or vulnerable species. As a result, India's marine fisheries are moving from an open-access system towards one of regulated access. However, implementation of such measures is challenging since the large population of coastal fishers is actually dependent on fishing, thus any proposal for regulatory measures tends to be resisted by the fishermen.

In addition to the overexploitation of resources, other key issues affecting fish stocks are habitat degradation, pollution and climate change. Critical habitats such as mangroves, coral reefs, and seagrass beds occur in the region, and these are affected by coast development and urbanisation. Some trade in products from these habitats also exists. The discharge of sewage, an increasing load of litter, and organic and inorganic pollutants further degrade coastal ecosystems. Fisheries sustainability must be addressed in an integrated way by considering the issues of all anthropogenic interferences especially increasing fishing intensity, damage to the physicalchemical and biological integrity of the ecosystems, and climate change. As India's fisheries are impacted by the developmental needs of several other prime sectors, such as agriculture, industries and power generation, it is not possible to find solutions to the issues of the fisheries sector alone. For instance, problems such as water contamination, the enforcement of standards for water discharge, maintaining the quality of river runoff, and reducing greenhouse gas emissions, need to be addressed by non-fisheries sectors. Considering the development needs of other important sectors, the challenges for the fisheries sector have increased as never before, and these demand adopting an ecosystem approach to fisheries management with the involvement of all stakeholders.

#### GLOSSARY

**decadal sea surface temperature (SST) anomalous months** – monthly sea surface temperatures are compared to average seasurface temperatures for 1971–2000, and plotted on a map. Where they are higher or lower than the average they are referred to as anomalous months.

**isobath** – a line on a map that joins all points of the same water depth in the sea.

**thermocline** – the layer where the temperature of the sea (and of lakes, where it is called the metalimnion) changes suddenly, with a layer of warm water lying over a deeper layer of colder water.

# THE COLLECTORS: ICHTHYOLOGICAL EXPLORATION OF THE WESTERN INDIAN OCEAN

Wouter Holleman and Barry C Russell, with Theodore W Pietsch



Figure 1 A Mosaics of fishes, Bardo Museum, Tunis. © W Holleman, NRF-SAIAB B Mural from Nebamun's tomb-chapel, c. 1350 BCE. © L Rich

The Mediterranean Sea and the rivers that flow into it were the source of fishes for the Egyptians, Phoenicians, Greeks, Romans and others who sailed, explored and fished it for food. Some of the fishes they caught are immortalised in mosaics, such as those in the Bardo Museum in Tunis and pictured on the walls of the tomb of Nebamun (c. 1350 BCE), near Thebes in Egypt. They gave names to the fishes they caught, some of which are still in common use today, such as kobios and thynnos (the Greek words for goby and tuna). Later, Roman and Arab traders – and probably Egyptians before them - ventured down the Red Sea and the Persian/ Arabian Gulf, and into the Indian Ocean to trade with India. Centuries later, Portuguese and Dutch explorers carved out sea routes around the Cape of Good Hope to the spice islands of the East Indies. All seas were naturally a source of food for people who lived along their shores, as shown by the many middens archaeologists have discovered along the coast.

The origins of our current knowledge of fishes were first recorded by *Georges Cuvier* in his *Historical Portrait of the Progress of Ichthyology, from Its Origins to Our Own Time,* first published in 1828. While the earliest collections and descriptions of Western Indian Ocean (WIO) fishes were made in the 1600s, it was only in the 19th century that scientific interest was focused on fishes, as epitomised in Cuvier and Valenciennes's *Histoire naturelle des poissons* published between 1828 and 1849. Later in this chapter we reveal more about these two remarkable scientists and their contributions to ichthyology.

The early WIO ichthyologists and collectors can be broadly grouped into three kinds: those who collected, named and described the fishes they found; those who collected fishes for others; and those who received, described and named specimens sent to them. In this chapter we trace these pioneers from the 17th century to the early 21st century. From these early foundations, their work has gradually and continuously built up the foundations on which current WIO ichthyological research is based. It is out of these threads that the story is woven, and, as you read and use this book you will come across some of their names, and so recognise their contribution to our knowledge of these fishes.

#### **BEGINNINGS**

The first person that we know of who collected and illustrated fishes from the WIO was *Isaac Johannes Lamotius*. He was born in Beverwijk, in Holland, in about 1646, joined the Dutch East India Company (VOC), and arrived in Mauritius as Governor in 1677. In 1682, Joan Huydecoper II, an administrator of the VOC and an amateur botanist, urged colonial administrators to undertake botanical research and make drawings of local plants. Lamotius, who was interested in natural history, became an expert on the fauna and flora of Mauritius, and over the next 15 years he collected plants and animals, describing and making drawings of them until his departure in 1692. They include about 250 drawings of fishes, now housed in Paris.

*Philibert Commerson* was the first of the explorer-collectors to visit islands in the Indian Ocean. He was born in Châtillon-les-Dombes, Ain, in France, in 1727, studied medicine, and was passionate about natural history. In 1766 he joined one of Louis Antoine de Bougainville's expeditions, visiting Brazil, the Falkland Islands, Tierra del Fuego and Tahiti. Along the way he collected plants, fishes and mammals. After visiting New Guinea and Java, Bougainville sailed to Mauritius, where Commerson stayed on. While on Mauritius, Commerson made a collecting trip to Madagascar. In all, he collected and made extensive notes on more than 160 species of fishes, and made fine drawings of many of them. Commerson did not live to publish any of his work. He died on Mauritius in 1773 and soon after his death, his papers and collections were sent to the Ministry of the Navy in France, which handed them over to Georges Buffon. Buffon used fragments of this material on birds, but ignored the rest. Later the material was deposited in the Jardin des Plantes (which became the Muséum national d'Histoire naturelle) in Paris. The fishes Commerson collected were unaccounted for until discovered by André Duméril, still in their original cases, in Buffon's attic some 20 years later. Pietsch writes:

Commerson's talent as an ichthyologist might have been lost if it were not for the fact that his manuscripts and drawings were later incorporated by *Bernard Germain Etienne de Lacépède* (1756–1825) in the latter's *Histoire Naturelle des Poissons*, originally published in five volumes between 1798 and 1803. Despite the fact that many authors have cited Lacépède as the original describer of these species (and have erroneously attributed the names to him), he provided no Latin binomials, referring to the species only in the French vernacular (Anon. 1910, 1925). Commerson had named all his fishes by lengthy Latin polynomials, but most of his species were described in great detail, and many were represented by illustrations that leave no doubt about their identity. (Pietsch & Grobecker 1987: 9)

The first of the men who never travelled or collected their own specimens, but worked solely on fishes lodged in collections and those sent to them, is *Marcus Elieser Bloch*, who is considered one of the most important ichthyologists of the 18th century. He was born at Ansbach, in what is now Germany, in 1723. His parents were very poor and he grew up barely literate – at the age of 19 he could not even read German. He did, however, know some Hebrew which enabled him to obtain a teacher's position in the house of a Jewish surgeon in Hamburg. There he became fluent in German and mastered some Latin. He also studied anatomy, which led to an interest in science. Bloch then went to Berlin, where he studied all branches of natural science and medicine. After taking the degree of MD. at Frankfurt-an-der-Oder in 1747, he settled in Berlin, where he became established as a physician.



**Figure 2** Philibert Commerson. Source: Wikimedia Commons



Figure 3 Bernard de Lacépède. Source: Wikimedia Commons





Figure 4 A Marcus Bloch; B Johann Schneider. Source: Wikimedia Commons



Between 1782 and 1795 Bloch produced twelve volumes on the general natural history of fishes. The last eight volumes dealt with fishes from other parts of the world and were titled *Naturgeschichte der ausländischen Fische*. Many of the fishes described in this work were supplied by Missionary John, a Danish missionary working at Tranquebar (today Tharangambadi), on the coast of Tamil Nadu in southern India; other fishes he bought from merchants or vendors. In 1801 Bloch's work was corrected and expanded by *Johann Gottlob Schneider* and published as *Systema ichthyologiae iconibus CX illustratum*, by Bloch & Schneider. Bloch described 290 fish species, as well as a further 425 species with Schneider. Bloch's collection of about 1 500 specimens is today preserved at the Museum of Natural History of the Humboldt University in Berlin.



Figure 5 A Title page of Systema Ichthyologiae iconibus CX illustratum by Bloch & Schneider 1801; B Scomberomorus guttatus; C Acanthurus lineatus. Source: Wikimedia Commons



Figure 6 Jean (Georges) Cuvier. Source: Wikimedia Commons



**Figure 7** Achille Valenciennes. © NRF-SAIAB (by R Palmer)

# THE 19TH CENTURY, THE AGE OF DISCOVERY

The most remarkable partnership in ichthyology was that of *Georges Cuvier* and *Achille Valenciennes*. Their work laid the foundations of the modern study of fishes and is encompassed in the 22 volumes of *Histoire naturelle des poissons*, published between 1828 and 1849. The volumes provided descriptions of 4 055 species of fishes, 2 311 of which were new to science. This monumental work could not have been carried out without the participation of many donors and collectors, several of whom we acknowledge in the following pages.

*Jean Léopold Nicolas Frédéric (known as Georges) Cuvier* was born in 1769 at Montbélliard, France. It soon became evident that young Georges was exceptionally intelligent, and was interested in the natural sciences. His family did not have the means to educate him adequately, but his talent was spotted by the sister-in-law of the Duke of Württemburg (who lived in the castle at Montbélliard). She gave him a stipend so that he could complete his schooling. Georges learned to speak German and the then universal language of academia, Latin. In 1788 he became a tutor in Normandy, where he met the famous botanist Abbé Alexander Henri Tessier, who, impressed by the young man's knowledge of natural history, recommended Georges to his colleagues at the newly established Muséum national d'Histoire naturelle in Paris.

Cuvier arrived in Paris in 1795, and in December of that year taught his first class in anatomy at the museum. In 1798 he published a summary of the classes he gave at the École central du Panthéon, and in the preface he wrote: "Linnaeus, M Bloch and Citizen Lacepède have been my principal guides for reptiles and fishes, …" (Bauchot *et al.*, in Pietsch & Anderson 1997, 29). With his rapidly growing and extensive knowledge he conceived of *Le Règne Animal distribue d'apres son organisation, pour servir de base à l'histoire naturelle des animaux et d'introduction à l'anatomie comparée* (The animal kingdom arranged according to its organisation, to be used as a basis for the natural history of animals and as an introduction to comparative anatomy). The first edition appeared at the end of 1816.

While gathering information for Le Règne Animal, Cuvier met a young zoologist, Achille Valenciennes, 25 years his junior. Valenciennes was born in Paris in 1794. His father was one of the first naturalist aides at the museum, but died leaving his widow and five children. Achille, the eldest at 18, was left to support the family. He became employed by the museum at the behest of the renowned zoologist Lacepède, a friend of an uncle, and was taken into the service of Lamarck, an early proponent of evolution, to work on invertebrate collections. It was in Lamarck's laboratory that he met Cuvier, and began to work with him on vertebrates.

Lamarck gradually became blind, and died in 1825. Valenciennes continued to work in the laboratory of his successor, André Marie Constant Duméril, until 1832, when Cuvier persuaded Lacepède to allow him to work in Cuvier's laboratory. It appears he had already been unofficially assisting Cuvier for about 17 years at that stage - in the 1827 prospectus announcing the publication *Histoire naturelle des poissons*, Cuvier wrote "Monsieur Valenciennes has not ceased to help me during the past twelve years in all my preparatory work" (Bauchot et al., in Pietsch & Anderson 1997, 31).

The first volume of *Histoire naturelle des poissons* was published in 1828, and on the title page was printed "by Monsieur le Baron Cuvier... and by Monsieur Valenciennes," in samesized font, an apparent acknowledgement that Cuvier's collaborator had become his equal in ichthyology. Together the two published five volumes of the Histoire naturelle, and when Cuvier died in 1832, Valenciennes continued the work, eventually publishing the 22 volumes on which the modern science of ichthyology has been built.

The first person to make extensive scientific collections in the Red Sea was *Peter (Pehr)* Simon Forsskål, who was born in Helsinki, Finland, in 1732. By the time he was 18 he had acquired a degree in theology from the University of Uppsala in Sweden, where he was one of Linnaeus's students (1756–1759) and qualified as a botanist. He was a bright young man and, amongst other subjects, studied oriental languages. With Linnaeus's approval Forsskål joined an expedition to Arabia in 1760. After a year in Egypt, where he studied Arabic dialects, the expedition travelled to South Yemen in 1762, where they were supposed to stay for 2-3 years. It seems all the expedition members picked up various diseases and, within a year, all the participants except Carsten Niebuhr, had died, Forsskål in 1763. Of the extensive collections of plants and animals, his fish collections coming mainly from Saudi Arabia and Yemen, little survived: most of the information we have is found in Niebuhr's copies and translations.

After Niebuhr's return to Denmark in 1767 he immediately started to publish the work of his colleagues, especially that of Forsskål - the fish specimens were described and published in 1775. Niebuhr's work was inconsistent and full of errors, but many of the fish species described are still considered valid today.

Exploration really gathered momentum in the middle of the 19th century. Two naturalists who collected but did not describe many fishes, were Wilhelm Friedrich Hemprich and Christian Gottfried Ehrenberg.

Wilhelm Hemprich was born in Glatz, Prussian Silesia, in 1795. His father was a doctor and he also studied medicine (at Breslau and Berlin). His first interest, however, was zoology, and he was appointed as a lecturer in natural science and physiology at the university in Berlin. He also became a collaborator at the Zoological Museum. Hemprich met Ehrenberg at university in Berlin and they became friends. Ehrenberg was born in Delitzsch in Germany in 1796 and first studied theology, followed by studies in medicine and the natural sciences in Berlin, where he met and became a friend of the explorer Alexander von Humboldt.

Sharing an interest in natural history, Hemprich and Ehrenberg were invited to participate in an expedition to Egypt in 1820. In 1821 they separated from the main expedition and spent the next two years collecting thousands of plant and animal specimens from Egypt, the Nile Valley, Libyan Desert and northern Red Sea. With renewed funding, the two set out in 1823 to investigate marine life in the Gulf of Suez, and along the Arabian coast, as far as Mohila. They collected many fishes, made drawings of unknown species and notes on anatomy. They also collected molluscs, crustaceans and various coelenterates.







Figure 8 A Cover of Volume 1 of Histoire naturelle des poissons by Cuvier & Valenciennes 1828; B Diploprion bifasciatum; C Uranoscopus lebeck. Source: www.biodiversitylibrary.org



Figure 9 Peter Forsskål. M Wallerstedt © Uppsala University Museum



Figure 10 Christian Ehrenberg. © Humbold University, Berlin



Figure 11 A Eduard Rüppell, as a young man of about 30 years old; B in 1869, at the age of 75. © Archiv Senckenberg Gesellschaft für Naturforschung, deposited in the Institut für Stadtgeschichte, Frankfurt am Main



Figure 12 Fishes named after Jean-Jacques Dussumier: **A** Hyporhampus dussumieri. PC Heemstra © NRF-SAIAB **B** Acanthurus dussumieri. © SV Bogorodsky

In 1824 the two men started their last expedition, to the southern Red Sea, travelling to the islands, mapping them, and collecting specimens along the way. After reaching the African coast, Hemprich undertook a trip to the Gedem Mountains whilst Ehrenberg remained in the camp due to illness. Hemprich's journey was catastrophic as he and most of his assistants died of fever in Massawa in 1825. Ehrenberg recovered from his illness and eventually returned to Germany at the end of that year.

When Valenciennes visited Berlin in 1827, Ehrenberg showed him his notes and drawings of fishes. In 1829 Ehrenberg travelled with Humboldt to the Ural and Altai mountains, and on his return donated specimens from the Red Sea, Nile River and Caspian Sea to the Paris museum.

*Wilhelm Peter Eduard Simon Rüppell*, the son of a prosperous banker, was born at Frankfurt-am-Main in 1794, and was destined to become a merchant. However, a visit to the Sinai Peninsula in 1817 awakened an interest in natural history. After attending lectures in botany and zoology at the universities of Pavia and Genoa, he taught himself systematics. He set off on his first expedition in 1821, travelled through the Sinai and reached the Gulf of Aqaba in 1822. In 1826 Rüppell began his work on Red Sea fauna, collecting large numbers of crustacea from the Gulf of Suez. This was followed by travels to the Dahlak Islands and Massawa coast, making diverse collections of fishes, crustaceans and other invertebrates that he sent back to the Senckenberg Museum. He also travelled up the Nile to Nubia in 1823 and collected specimens around Ambikul, now in northern Sudan. Between 1828 and 1840, he published the results of his travels, eventually describing 23 new genera and 221 new species of fishes (Klausewitz 2002).

Bordeaux ship-owner, merchant and natural history enthusiast *Jean-Jacques Dussumier* (1792–1883) made at least 11 commercial voyages between 1816 and 1840 in his own vessels, the *Buffon* and the *Georges Cuvier*. He sailed the Indian Ocean and made large collections of fishes, plants and other animals from the Seychelles, Réunion, Mauritius, India, and Sri Lanka to the South China Sea. On his return from a voyage, he gave the specimens to the Muséum national d'Histoire naturelle in Paris, together with descriptive notes and drawings he made of fresh specimens. Cuvier wrote in an 1824 report (as quoted by Lassius 1973: 390), "One awaits his returns and one notes them down in the fashion of the custom-house or the Stock Exchange." On many occasions Cuvier praised "this merchant, clever and enlightened in all branches of natural history, who has already given the most positive and incontestable proof of his unselfish motives in the matter of scientific research" (Cuvier, Archives Nationales, Paris, AJ 578, no. 26–33, as quoted by Laissus 1973: 399).

Often the fishes comprised the most interesting collections, particularly fishes from Indonesia and the Ganges from the third voyage (1821–1823), and fishes from the Seychelles gathered on the fourth voyage (1825–1826), "most being entirely new and others having been seen only by Commerson" (Cuvier, in *Rapport fait à l'Académie royale des sciences* [1830], as quoted by Laissus [1973: 391]). On the fifth voyage (1826–1827), Dussumier brought back from the coast of Malabar and from the Seychelles, according to Cuvier, "one of the finest and most abundant collections of fishes ever seen" (Cuvier, in *Rapport fait à l'Académie royale des sciences* [1830], in Lassius [1973: 392]), totalling 550 specimens representing 200 species. The collection included some freshwater species, many from the State of Mysore.

The most productive and extensive of his voyages was the sixth, starting late 1827 and lasting 33 months, 27 of which were spent at sea, with ports of call in the Seychelles, India (Mumbai, Puducherry, Mahé, the Ganges delta), Bengal, Réunion, Mauritius and St Helena. He returned with a total of 1 500 fishes representing 481 species. Cuvier described Dussumier's samples as follows:

It is especially the fishes in Monsieur Dussumier's gift that amaze naturalists and inspire in them the keenest gratitude... Not a ray, not a prickle is broken; their scales, even their colors, are usually preserved, and whenever the fluid has altered them, one finds in Monsieur Dussumier's descriptive notes whatever is necessary for completing the
description... And fortunately too, all these fishes, thanks to incisions made in their abdominal skins, have their viscera preserved in the best condition, so that they may be used in any anatomical research that one may desire. (Cuvier, in a report to the Royal Academy of Sciences: *Rapport fait à l'Académie royale des sciences* [1830], as quoted by Laissus [1973: 398])

Dussumier was named Correspondent of the Museum in 1827, made a Knight of the Legion of Honour in 1831, and an Officer in 1841. In 1865 he gave the museum the systematic catalogues of collections brought back from his voyages, as well as an album of watercolour drawings of the cyprinids of China, which he had made at Canton in 1820. He died in 1883 and 31 species of fishes today carry Dussumier's name.

Two further names linked with WIO fishes, are those of Albert Günther and Lambert Playfair. *Albert Charles Lewis Gotthilf Günther* was born in 1830 in Esslingen in Swabia. He graduated in medicine in 1852 and in the same year published a handbook on zoology for medical students. In 1855 he visited England, where he met John Edward Gray, Keeper of Zoology at the British Museum (Natural History), and palaeontologist Richard Owen at the Museum. In 1857 he was offered a position at the museum, his first job being to classify some 2 000 snakes. When Gray died in 1875, Günther was appointed Keeper of Zoology, a position he held for a further 20 years. His major work was the compilation of eight volumes of the *Catalogue of Fishes* (1859–1870). He also established the *Record of Zoological Literature* in 1864, and served as its editor for six years.

**Robert Lambert Playfair** was born in St Andrews, Scotland, in 1828. He became a military cadet and in 1846 joined the Madras Army and became an artillery officer. In 1854 Playfair was appointed Political Agent at Aden, and in 1862 he became Political Agent and then Consul of Zanzibar, a position he held until 1867. In that time he made extensive collections of fishes from Zanzibar, the Seychelles and Madagascar, which he sent to Albert Günther at the British Museum in London, and published his findings in the *Proceedings of the Zoological Society of London*. In 1866, in collaboration with Albert Günther, he published *Fishes of Zanzibar*, the first comprehensive account of the fishes of the east coast of Africa (see Myers, in Playfair & Günther 1971 reprint). Playfair described 36 species of fishes, of which 15 are still valid.

*François Paul Louis Pollen* was born in 1842 in Rotterdam and, at the age of only 20, financed his own expedition to Madagascar. In 1862 he moved to Leiden to study medicine but was encouraged by Hermann Schlegel to study zoology. After some training and scientific preparation, Pollen departed with his friend, *Douwe Casparus van Dam*, reaching Réunion in 1864. After much successful collecting in northwest Madagascar, Mayotte and Réunion, they returned via Mauritius in July 1866. Sponsored by Pollen, Van Dam again collected on Madagascar from 1868–1872 (or 1873). Later, Pollen engaged Joseph Peter Audebert to continue the explorations, which he did until 1882. Pollen's large collection of fishes (as well as birds, mammals and insects) were given to the Rijksmuseum in Amsterdam. The fishes collected by Pollen and Van Dam were the subject of several papers by Bleeker (see below) (Boeseman, in Pietsch & Anderson 1997: 81–100).

Traveller, collector and describer *Franz Steindachner* was born in Vienna in 1834. He began to study fossil fishes at the recommendation of a friend. His growing interest in natural history led to his appointment, at the age of only 26, as director of the fish collection at the Natural History Museum in Vienna. His reputation as an ichthyologist grew, and in 1868 Louis Agassiz invited him to accept an appointment at the Museum of Comparative Zoology at Harvard in the United States. He returned to Vienna in 1874 and, in 1887, was appointed director of the natural history department. From 1895 to 1896, Steindachner was in charge of the First Red Sea Expedition of His Majesty's Academy of Sciences. On the return journey, he travelled through Palestine, Syria, Anatolia, and northern Greece. In 1897, he took part in the Second



**Figure 13** Albert Günther. Source: Wikimedia Commons



**Figure 14** Plates from Playfair & Günther's *Fishes of Zanzibar*. Source: Playfair & Günther 1866 Public domain



Figure 15 Franz Steindachner. Source: Carl Leavitt Hubbs Papers. SMC 5. Special Collections and Archives, UC San Diego Library





Figure 16 A Carl Klunzinger. Source: Jahreshefte des Vereins für vaterländische Naturkunde in Württemberg, LXXI, 1915 B *Thalassoma rueppelli*, originally *T. klunzingeri*. © R Field

Red Sea Expedition but had to return early following year to take over the administration of the entire Kaiserlich-Königliches Naturhistorische Hof-Museum in Vienna. He also undertook trips to the Canary Is., Senegal and Latin America, described 1 034 species and published >200 papers on fishes.

*Carl Benjamin Klunzinger*, born in Güglingen, Germany, in 1834, also started his professional life as a physician. He later attended lectures in geology and zoology in Vienna and Prague. In 1862 Klunzinger travelled to Cairo and spent the next 18 months learning Arabic. He worked as a physician at Kosseir (now Al-Qusayr) on the Red Sea. Over the next five years, Klunzinger built up an enormous collection of Red Sea fishes and other marine animals, which were sent back to the Staatliches Museum für Naturkunde in Stuttgart. In 1869 he returned to Stuttgart and spent the next few years working on the collection. He published the two volumes of *Synopsis der Fische des Rothen Meeres* in 1870 and 1871. Two years later, he was back in Kosseir, collected more material, and returned to Stuttgart in 1875. In 1884 he published *Die Fisches des Rothen Meeres*, which was followed by books on the coral animals and the Cyclometopa crabs of the Red Sea before his death in Stuttgart in 1914.

The last of the 19th century explorers who contributed significantly to our knowledge of fishes of the northern WIO was Surgeon-Major *Atmaram Sadashiv Grandin Jayakar* (1844–1911). We know little of his early life, and presume he was born in India. He arrived in Oman in 1879 as successor to Surgeon-Major Mr Apothecary Gaspar de Rozario. Rozario had been appointed British Political Agent in Oman in 1861, responsible for medical services and recording the health of the population, as well as documenting the fauna and flora, socio-cultural practices and informing headquarters (in India) of political and military matters. During his 21-year tenure in Oman, Jayakar made extensive collections of marine fishes in the area, which he sent to the British Museum.

*George Albert Boulenger* was born in Brussels, Belgium, and joined the staff of the British Museum in 1880, was charged with the task of attending to Jayakar's donations. In 1887, he wrote of the first consignment:

The Natural History Museum has received a large and most valuable collection of Fishes, obtained at Muscat and presented by Mr. Jayakar, which I have been directed by Dr. Günther to examine. This collection, containing specimens of 172 species, many of which were unrepresented in the National Collection and 14 of which are apparently new to science, fills a gap in our knowledge of the distribution of the fishes of the Indian Ocean. Scarcely anything is known of the fauna of the Persian/Arabian Gulf and the neighbouring coasts, a district intermediate between two others, the fishes of which have been tolerably well investigated, viz. the North-eastern coast of Africa and the West coast of India. It therefore seems to me that, in addition to the description of the new species, the publication of a full list of the fishes obtained will be useful. With the exception of three (the names of which are preceded by an asterisk) all the species enumerated are marine. (Boulenger 1887: 653)

Two years later, further consignments arrived, and Boulenger wrote:

At the close of last year, the Natural History Museum received a second collection of Fishes from its generous correspondent at Muscat, Surgeon-Major A. S. G. Jayakar; this was shortly followed by a third, received a few days ago. Other examples of a number of the rare or new species enumerated in my preceding list having been sent again, I have, in some cases, been able to supplement my former descriptions; but all of the species enumerated hereafter, and to which numbers are attached, are new to the fauna of Muscat. Eight of the known species are altogether new to the Indian Ocean... Five species are considered to be new. The Sharks and Rays, which were hardly represented at

all in the first collection, have now reached us in numbers, and, from the size and beauty of most of the skins, will, to say nothing of the great zoo-geographical interest that attached to many of them, form a welcome addition to the Fish-Gallery of the Museum, where Mr. Jayakar's previous donations of large Sea-Perches, Scombroids, Sphyraenas, and Sword-fishes have already had conspicuous place. Thanks to the exertions of Mr. Jayakar, the number of Fishes recorded from Muscat now known amounts to 256. (Boulenger 1889: 236)

By the time of his retirement in 1820, Boulenger had described 1 137 species of fishes, many of them from the WIO.

There are other men whose interests lay elsewhere but who, during the 18th, 19th and early 20th centuries, contributed to our knowledge of WIO fishes by collecting specimens and sending them back to Europe: *Pierre Poivre* (1719–1786) who was General Commissioner of Mauritius and Réunion from 1767 to 1772, sent fish specimens from Mauritius; French naturalist *Jean-Baptiste Bory de Saint-Vincent* (1778–1846), also spent time on Mauritius and Réunion during his travels; and *Julien François Desjardin* (1799–1840), born on Mauritius, was a founding member of the Society of Natural History of Mauritius. The latter's specimens and notes were used by *Jean-Louis Hardouin Michelin de Choisy* for his *Essai d'une fauna de l'Ile de Maurice*, published in 1849. *Jean Baptiste Leschenault* (1773–1826) sent many fish specimens from India (Puducherry), Sri Lanka, Mauritius and Réunion to the Paris museum. Additionally, *Harald Blegvad* (1886–1951) was a fisheries biologist who spent two winters in the Persian/ Arabian Gulf in 1836 and 1938, where he greatly expanded our knowledge of the fish fauna with the publication in 1944 (with the assistance of Berndt Løppenth) of *Fishes of the Iranian Gulf*.

The British Empire controlled much of Africa's eastern coastline, from Egypt to Tanganyika (Tanzania) in the 19th and early 20th centuries, – and all of India. Captain *FrederickWilliam Townsend*, commander of the Indian cable ship *Patrick Stewart*, which worked in the Persian/ Arabian Gulf and Gulf of Oman from 1893 to1914, collected molluscs and deep-sea fishes, many of which he sent back the British Museum, and other museums in the United Kingdom.

In September 1933, an Anglo-Egyptian expedition was launched to investigate the fauna and oceanography of the northwestern Indian Ocean, an area that was relatively unknown. The expedition was financed from a bequest of Sir John Murray, one of the scientists on the *Challenger* expedition of 1872 to 1876, and was conducted using the Egyptian research trawler HEMS *Mabahiss*. The fishes (and other organisms) collected were lodged at the British Museum under the care of *John Roxborough Norman*.

John Norman, born in London in 1898, started his working life as a bank clerk. In 1921 he joined the staff of the British Museum where he worked for Charles Tate Regan. Norman published extensively on fishes and their classification – amongst his works are *A History of Fishes* (1931) and *A Draft Synopsis of the Orders, Families and Genera of Recent Fishes* (1957). He published accounts of nearly 250 species of fishes from the *Challenger* expedition (including deep-sea fishes), of which 29 were new to science (Norman 1939).

One other name stands out, that of *Pieter Bleeker*, who was stationed in the Dutch East Indies from 1842–1860. He was employed as a medical doctor with the Dutch East India Army, which is when he did most of his ichthyological work. He built up a network of contacts who sent him fishes from outlying government outposts on the many islands. In all, he collected some 12 000 specimens (which were eventually sent to the Natural History Museum in Leiden), and described more than 500 new genera and 1 932 new species. Of Bleeker it can be said that he described close to an entire fauna, something we cannot say of anyone else. While only some of the fishes were collected from the WIO (see under Castelnau below), many of the species in these volumes were named by Bleeker, and one of his many papers dealt specifically with fishes from the Cape (Bleeker 1860).



Figure 17 George Boulenger. © BMNH



Figure 18 Pieter Bleeker. © Rijksmuseum, Amsterdam



Figure 19 A Andrew Smith. © Iziko South African Museum B Rhincodon typus, the whale shark described by Smith in 1828. © M Fraser



Figure 20 A François Nompar de Caumont Laporte, Comte de Castelnau. Source: Joanne 1847 B Two clinids, drawn by Castelnau. © Aquarium-Muséum Liège – All rights reserved

# **EXPLORATION OF THE SEAS AROUND SOUTHERN AFRICA**

We next turn our attention to the south, where exploration of fishes developed in a very different way. Prior to the turn of the 19th century, few fishes had been collected from southern African waters. It was only with the establishment of the first museums in the Cape Colony, which was under British control from 1795–1803, and then from 1807 onwards, that collections were initiated.

Early in 1825, Governor Lord Charles Somerset travelled to Grahamstown to visit the military garrison. There he met a young medical officer, *Andrew Smith*, who showed him a small collection of reptile, bird and mammal skins he had made, and who raised the question of establishing a museum for the Colony. Just a few months later, Somerset transferred Smith to Cape Town and made him the first Superintendent of the South African Museum, which was founded in 1825. Smith thus became the first museum scientist in South Africa and his collections the first research collections at a South African institution/museum. The museum's first annual report (1830) described the fishes that had been collected to date: "... about 90, including several of great rarity and interest, particularly in the family of the sharks, of which the species are above 20." These were all specimens which had been donated to the museum. Over the next few years Smith also described 26 marine fish species from South African waters.

Smith was born in Kirkton, Scotland, in 1797. He studied medicine in Edinburgh and later joined the army as a hospital assistant before completing his medical degree in 1819. It was during this period that he became interested in natural history – as part of his military training – at McGrigor's museum at Fort Pitt, Chatham. He arrived in Cape Town in 1821 and was posted to Grahamstown a few weeks later. When Smith returned to England in 1837, he took his private collections with him. They had been on loan to the South African Museum for 12 years, and their removal depleted the museum's fish collection considerably.

The next person to collect fishes from around South Africa was Castelnau. *François Louis Nompar de Caumont Laporte, Comte de Castelnau*, was a widely travelled naturalist and diplomat. Born in London in 1802 (Evenhuis 2012), he later studied natural science in Paris under Georges Cuvier, Geoffroy Saint-Hilaire, and other notable French zoologists. From 1837 to 1841 he travelled in the United States and Canada, and from 1843 to 1847, he led an expedition to South America, crossing from the Mato Grosso to Peru and returning via the Amazon River. After the 1848 French Revolution, he took up a diplomatic career, and from 1848 to 1855 was French Consul at Salvador, Bahia, Brazil. In 1855, he returned to France and later travelled by steamer to the Cape of Good Hope, where he was French Consul from 1856 to1858. Following this posting, he took up the position of French Consul to Siam (Thailand) from 1858 to 1862. In 1863, Castelnau was appointed Consul-General for France, in Melbourne, Australia, where he remained after his retirement in 1877, until his death in 1880. He published several important papers on the Australian fish fauna during the 1870s.

Castelnau was a prolific ichthyologist. En route to the Cape Colony, he visited Île Bourbon (Réunion) where he collected and sketched a number of fishes. In South Africa, assisted by his son Ludovic, who visited Port Natal (now the city of Durban), and preparator Frédéric Daviaud, who visited Lake Ngami (Botswana), he collected fishes from Table Bay, Simons Bay (Kalk Bay) and Algoa Bay. Some of these specimens were sent to the famous Dutch ichthyologist, Pieter Bleeker, who in 1860 published a paper on the fishes of the Cape of Good Hope, describing several new species (Bleeker 1860). Castelnau himself published a monograph, *Mémoire sur les poissons de l'Afrique Australe* (Castelnau 1861) which included 71 new species. In 1865, Professor Jean-Theodore Lacordaire purchased most of Castelnau's specimens as well as a number of his sketchbooks, including unpublished watercolour paintings of many South African fishes, for the Zoological Museum, University of Liege, Belgium (Loneux 2006). Today this collection of Castelnau's specimens and paintings remains one of the earliest and most important records of South Africa's natural history heritage.

Several other museums were soon established in South Africa, in Grahamstown in 1855, in Port Elizabeth a year later, and in Durban in 1887. The fish specimens that found their way into collections at these museums and into the literature were all donated. That all changed in 1895. A growing concern that poor fishing practices would harm the fishing industry engendered an interest in the biology of fishes. The need to review the fishery industry led to the appointment of *John Dow Fisher Gilchrist* (1866–1926), a British marine biologist, to take charge of a survey of South African fishes and fish stocks.

Gilchrist was educated in Scotland, graduating from Edinburgh with a BSc and MSc. He pursued his studies on the feeding habits of fishes of Munich and Zurich Jena University, where he was awarded a PhD, and returned to Edinburgh in 1892 as assistant lecturer in zoology. In 1895, Gilchrist was appointed marine biologist in the Department of Agriculture of the Cape of Good Hope.

At Gilchrist's urging, the Government acquired a steam trawler in 1897, "to make a thorough investigation into what fishes are really in the sea and where they occur (Anonymous 1897). In the years that followed, the *Pieter Faure* trawled first in Cape waters, and then later off East London. Most of the trawling and dredging was from 40–200 m, and large quantities of marine organisms were collected. To identify the material, Gilchrist enlisted the help of Georges Boulenger of the British Museum, who was thus one of the first of those who named, but did not collect fishes from southern Africa.

Following the success of the *Pieter Faure's* survey in Cape waters, the Cape Government agreed to a similar survey off the Natal coast, and in 1901, the ship spent several months collecting material between Durban and Cape Vidal. A year later, Gilchrist produced the first, though preliminary, checklist of the marine and freshwater fishes of South Africa which included 55 families, 164 genera and 336 species. Then followed several years of personality clashes and arguments about who owned the collections made by the *Pieter Faure* and where they should be deposited. During this period few fishes were collected.

The next person to play a significant role in our understanding of WIO fishes (and crustacea) was *Keppel Harcourt Barnard* (1887–1964), an unusually gifted and versatile scholar. He was born in Lambeth in the United Kingdom, attended school in England, and then spent some time in Germany to polish his German, before entering Christ's College at Cambridge to read the Natural Sciences Tripos of Botany, Geology and Zoology. He added Anthropology, Ethnology and Geography to these, and then read law, becoming a barrister in 1911. In the same year he came to South Africa, joined the staff of the South African Museum, and was appointed Director in 1921.

In 1912, Barnard went on his first collecting trip, spending four months in Natal (now KwaZulu-Natal) and Mozambique. He also worked on the *Pieter Faure* collections, which had by then been lodged at the South African Museum. By the end of 1914, he reported that 670 species of marine fishes had been identified. The annual report for that year also listed HW Bell-Marley as a donor for the first time. Another notable collector during this period was JB Romer Robinson.

*Harold Walter Bell-Marley* was born in the United Kingdom in about 1872, and after the Second Anglo-Boer War he settled in Durban, where he was employed by a shipping company. In 1918 he was appointed Principal Fisheries Officer of the Natal Fisheries Board, a post he occupied for 19 years. Bell-Marley was an ardent naturalist with a rare talent for collecting. Primarily interested in insects, he also made extensive collections of plants and animals from all over South Africa, He was also the first to collect molluscs from the stomachs of deep-sea fishes. Like Robinson, he donated the specimens to overseas and South African museums and gave hundreds of his fish specimens to the Natal Museum.

*JB Romer Robinson* was the son of the Natal Prime Minister, Sir John Robinson, and also had a great interest in natural history, particularly fishes. He loved angling and was, for a long time, President of the Natal Coast Angling Association. Robinson understood the importance of research, collecting and donating many fish specimens to museums in South Africa





Figure 21 A John Gilchrist. Source: Ritchie 1918; courtesy of UCT B Paralichthodes algoensis described by Gilchrist. Source: CFSA



Figure 22 SS Pieter Faure. © NRF-SAIAB



Figure 23 Keppel Barnard. © Iziko South African Museum



Figure 24 Harold Bell-Marley. © NRF-SAIAB



Figure 25 JB Romer Robinson. Source: Natal Mercury



Figure 26 SS Pickle. © NRF-SAIAB



Figure 27 Cecil von Bonde. © NRF-SAIAB (by R Palmer)



Figure 28 JLB and Margaret Smith sorting through a local fish catch, East African coast, c. 1952. Source: NRF-SAIAB archives

(particularly the Natal Museum in Pietermaritzburg) and overseas, all at his own expense. He also photographed many of the fishes he collected, but in 1911, his relationship with the Natal Museum apparently ended, and none of his fish photographs have ever been found.

When Barnard was working on the flatfishes collected by the *Pieter Faure*, he formulated an idea, and in 1919 he wrote, "... the soles collected by the SS *Pieter Faure* have been sorted, classified and incorporated. I have completed a short, descriptive account of the S.A. representatives of this family, which it is hoped will eventually form part of a more comprehensive work embracing the whole of S.A. marine fishes" (South African Museum, 1920). Barnard's *Monograph of the Marine Fishes of South Africa* was published by the South African Museum in two parts, in 1925 and 1927.

In addition to Bell-Marley, another name that repeatedly appeared on the list of donors of fishes to the South African Museum collection during the 1920s and early 1930s – that of *JLB Smith*, of whom more will be written later in this chapter. In 1936, Barnard made an arrangement with the Cape Town-based commercial trawling company, Irvin & Johnson, that trawler skippers keep any unusual fishes for the museum. Accessions rocketed to thousands per year. Further impetus to ichthyological exploration was the replacement of the old *Pieter Faure* with a larger vessel, a 30-m whaler, the SS *Pickle*, modified for trawling, which worked Cape and Natal waters in 1921 and then again in 1925, travelling as far north on the west coast as Walvis Bay, and on the east coast to Lourenço Marques (now Maputo). The large amount of material collected from as deep as 540 m (and on occasion nets were lowered to 1 800 m) resulted in the description of many new species.

Probably due to personality issues between Gilchrist and Péringuey, Director of the South African Museum, the museum did not receive these collections. It also seems that neither Cecil von Bonde (Director of the Marine Biological Survey from 1928-1952), nor Barnard approached each other about the future of the Pickle collections. Correspondence between James Leonard Brierley Smith and Von Bonde suggests that the collections were stored in a building at Cape Town harbour. On a visit to Cape Town in 1931, JLB Smith was given permission by Von Bonde to take any specimens he wanted as a donation to the Albany Museum. Eventually, after about two years, the relationship between these two men also broke down and no records of donations during this period were kept. Later the relationship resumed and, in 1947, Von Bonde suggested donating the collection to Rhodes University, and wrote accordingly to the Registrar, who accepted his offer. Smith then arranged for a Mr Kock to estimate the size of the collection prior to making arrangements for its removal to Grahamstown. As a result of Kock's report, Smith wrote to Von Bonde: "I am rather puzzled by the report sent by Kock. He was surely not shown the collection I worked on in 1933 which contained many thousands of specimens, including numerous types. Perhaps you can give me more information as Kock estimates there not to be more than five hundred fish in all." To this Von Bonde replied: "Mr Kock was shown the same collection ... but owing to our having to use the containers ... it was necessary to get rid of many duplicates ... only about 500 are left now" (Gon & Skelton, in Pietsch & Anderson 1997: 147).

*JLB Smith* was born in Graaff Reinet, South Africa, in 1897. He obtained an MSc in Chemistry at Victoria College (Stellenbosch University), and served on the staff of the college for several years before going to Cambridge in 1919. In 1922 he was appointed as a lecturer in organic chemistry at Rhodes University, Grahamstown. He was also a keen angler and this interest in fishes grew over the years as Smith trained himself as an ichthyologist.

In 1938 Smith married *Mary Margaret McDonald*, thus initiating one of the most remarkable partnerships in biology, producing work that influenced ichthyology worldwide. Soon after their wedding they headed north to Mozambique on their first expedition, collecting at Inhaca Island, and as far north as Beira.

In December that same year, the first coelacanth was caught by a trawler near East London, and Smith spent the next six months describing *Latimeria chalumnae*, as it became named. It was, without doubt, the fish discovery of the century, and catapulted the Smiths to world fame.

Further expeditions followed, including another to Mozambique in 1946, and later that year, Smith joined an Irvin & Johnson trawler to collect deepwater fishes. These collections added 1 000 specimens to the collections, 17 of which were of undescribed species. In time, Smith became the pre-eminent collector and describer in the southern part of the WIO (and southeastern Atlantic).

The year 1946 also saw the establishment of a Department of Ichthyology at Rhodes University, with JLB Smith as Research Professor in Ichthyology, funded by the Council for Scientific and Industrial Research. With this security, the Smiths travelled to Bazaruto in 1948, and returned to Inhaca a year later. In 1949 Sea Fishes of Southern Africa was published, covering 1 249 species of marine fishes. In the same year Smith noted that he was planning a major treatise on the fishes of the WIO, a book that would be "at least twice the size of my current volume and would probably occupy six to ten years". In preparation for this work, the couple ventured further north, collecting fishes from Beira to Pinda reef in 1950, and Mozambique Island, Pinda, Ibo and the Kerimba (Quirimba) Islands a year later. In 1952 they travelled to Zanzibar, Pemba (Tanzania) and Kenya; and in 1953, went back to northern Mozambique, then to the Seychelles, Aldabra and Amirante Islands in 1954. They brought back large numbers of specimens, photographs and colour notes, and for the next ten years, JLB worked through the material family by family, which Margaret Mary (as she became known) illustrated. The result was a series of 32 major papers on the fishes of the WIO and Red Sea, and a book on the fishes of the Seychelles in 1963. By the end of his career – and life, in 1968 – Smith had described 397 new species, of which 71 are still valid today.



Figure 29 JLB and Margaret Smith with Fishes of the Seychelles. Source: NRF-SAIAB archives



Figure 30 Francis Day. Courtesy Australian Museum



Figure 31 Alfred Alcock. CC BY 4.0



Figure 32 AGK Menon. Courtesy of R Menon & U Menon

# **INDIA**

The first substantive work carried out on the fishes of Indian seas was by *Francis Day*, an army surgeon and naturalist based in Madras. He was born in Maresfield, England, in 1829, was schooled at Shrewsbury (the same school Charles Darwin attended). In 1852 he joined the British East India Company as an assistant surgeon in the Madras Army. In 1858 Day moved to Hyderabad and then to Cochin (Kochi) where he became interested in fishes. In 1865 he published *The Fishes of Malabar*, a catalogue of the freshwater fishes of the region. After a spell in England, Day was appointed Inspector-General of Fisheries for India in 1871. Over the years he made extensive collections of fishes and birds. His major work, two volumes on the *Fishes of India* was published in 1878 and 1879, and includes descriptions of more than 300 species. While writing these volumes in England, he offered his fish collections to the British Museum, but Albert Günther, a fierce critic of Day, refused to purchase them. Day eventually sold an extensive collection to the Australian Museum. He deposited his drawings of fishes with the Zoological Society of London. Day was made a Fellow of both the Linnean Society and Zoological Society and also received awards from by France, India and Italy for his work.

*Alfred William Alcock,* born in Bombay (Mumbai) in 1859, a physician and naturalist, spent four years with the Royal Indian Marine Survey Ship *Investigator*, surveying and describing the deep-sea fishes of the Arabian Sea.

The work of Day and Alcock paved the way for several generations of Indian ichthyologists. One of the most widely-acclaimed was *Ambat Gopalan Kutty Menon* (1921–2002), mentored by the ichthyologist and biogeographer *Sunder Lal Hora*, who was unusual in that he worked on both marine and freshwater fishes. His publications range from a monograph on the soles of the genus *Cynoglossus*, to a *Checklist of the Freshwater Fishes of India*. Another ichthyologist trained by SL Hora was *KS Misra*, who published extensively on elasmobranchs between 1947 and 1969, and was also the editor of *Fauna of India*, published in 1976.

# THE POST-WAR PERIOD

From the 1940s, the momentum to explore the fauna of the Red Sea and Indian Ocean grew rapidly. In October 1957, the yacht *Xarifa* set sail for the Red Sea and Indian Ocean. The expedition leader was Hans Hass, who pioneered scuba gear and diving, and took the first photographs of fishes under water. On board was a young *Wolfgang Klausewitz* from the Senckenberg Museum, charged with collecting fishes, at the Farso and Farasan islands in the Red Sea and then at the Maldives.

The establishment of ichthyology in Israel led to growing exploration of Red Sea fauna, with two large expeditions to the Dahlak Archipelago (southern Red Sea), in 1962, under the leadership of *Heinz Steinitz*, and again in 1965. It was also in those early days that the extent and significance of the migration of species from the Red Sea into the Mediterranean through the Suez Canal began to be understood. The canal, developed by the diplomat and entrepreneur Ferdinand de Lesseps, was opened in 1869, and such species are now known as Lessepsian migrants.

*John E Randall* collected fishes in the Gulf of Aqaba in 1972, and two years later again in the Gulf of Aqaba, and in the Red Sea. Further Red Sea, Sri Lanka and Maldives collections followed in 1975, and a long voyage in 1977 took him from the Red Sea to the Seychelles and South Africa. In 1977, as a consultant to the United Nations Food and Agriculture Organization, he made extensive collections in the Persian/Arabian Gulf and Gulf of Oman. He made further extensive collections in the WIO in 1979 and 1980, in the Persian/Arabian Gulf in 1984 and 1985, and in Oman in 1993. Randall has described 826 fish species and has added more to our knowledge of WIO fishes than any other 20th-century ichthyologist. Randall was accompanied on several WIO trips by his former student and Australian ichthyologist *Gerald R Allen*, who eventually published new species descriptions of more than 30 WIO coral reef fishes, mainly in the family Pomacentridae.

In March 1976 *Victor G Springer* and colleagues travelled by schooner from the Seychelles to St Brandon Shoals and the Agaléga Islands and made numerous collections of fishes, which were lodged in the National Museum of Natural History in Washington, DC.

The JLB Smith Institute for Ichthyology (JLBSI), now South African Institute for Aquatic Biodiversity (SAIAB), was established shortly after JLB Smith's death in 1968 and built on the foundations laid down by the Smiths. *Thomas H Fraser*, with an interest in coral-reef fishes, was appointed to the new institute in 1970, and soon went on exploratory expeditions to KwaZulu-Natal, Inhaca Island and Mauritius. Three years later *Richard Winterbottom* succeeded Fraser and led two collecting trips to northern KwaZulu-Natal. Soon after Winterbottom's return to Canada (in 1977) he participated in the 1978–1979



Figure 33 Wolfgang Klausewitz. © F Krupp



**Figure 34** Phillip Heemstra, John Randall and Gerald Allen in Grahamstown in 1979.

Joint Services Chagos Research Expedition and made extensive collections of fishes. This was followed in 1988 by another extensive expedition with the JLB Smith Institute to the Comoro Islands. Additional collecting expeditions to Indian Ocean localities followed: the joint Smithsonian Institution / JLB Smith Institute expedition with *Phillip C Heemstra* to Mauritius and Rodrigues in 1995.

While based at JLBSI/SAIAB, Heemstra also made collecting trips to Kenya in 1980, the Comoros in 1991 and 1993, Mozambique in 1992, 1994, 1995, 1997 and 2007, Yemen in 1998, Rodrigues in 2001, Seychelles in 2005, and Madagascar in 1995, 1999 and 2010. He also made at least 20 collecting trips along the east coast of South Africa with Rhodes University students.

The R/V *Dr Fridtjof Nansen*, manned and operated by staff of the Institute of Marine Research in Norway, has made several surveys of areas in the Indian Ocean. These trips were made in collaboration with various institutions and programmes, including the Agulhas and Somali Current Large Marine Ecosystems project (ASCLME), the Centre National de Recherches Océanographiques (CNRO), the Mozambique Fisheries Institute, the universities of Cape Town and Western Cape, the Oceanographic Research Institute (ORI), South Africa, and SAIAB. The first destination (in 2007) was Mozambique, followed in 2008 by a survey around Madagascar, Seychelles and Mauritius. In 2009, the focus was on the Comoros, northern Mozambique, and the southern seamounts. Fishes collected on these and subsequent trips are deposited at SAIAB.

Collections and general interest in Indian Ocean fishes have steadily grown. Consequently there is greater understanding of the faunal diversity, as well as a proliferation of new species descriptions. This high level of interest is reflected on a global scale with 4 064 new fishes having been described between 2003 and 2012, including 140 marine species from the WIO (Fishwisepro database). Certainly many more new discoveries can be expected from this diverse region.

# CONTRIBUTORS

Note: biographical information correct at time of writing.



#### **GERALD R ALLEN**

Gerald (Gerry) R Allen is an international authority on the classification of coral reef fishes. Born and raised in the USA, he received a PhD in Marine Zoology from the University of Hawaii in 1971. He served as senior curator of fishes at the Western Australian Museum in Perth from 1974 to 1997 and has subsequently worked as a private consultant, primarily involved with coral reef fish surveys in Southeast Asia for Conservation International. Gerry has logged well over 12 000 hours of scuba diving in all tropical seas, especially the vast Indo-Pacific region. Underwater photography is a consuming passion and hundreds of his photos have appeared in a variety of international publications. He is the author of more than 400 scientific articles and 35 books, is a past president of the Australian Society for Fish Biology, an honorary foreign member of the American Society of Ichthyology and Herpetology, and recipient of the K. Radway Allen Award for outstanding contributions to Australian ichthyological science.



#### **KUNIO AMAOKA**

Kunio Amaoka is an emeritus professor of Hokkaido University, honorary member of the Ichthyological Society of Japan and honorary foreign member of the American Society of Ichthyology and Herpetology. Before he retired in 2000, he was a professor of Hokkaido University for about 40 years, where he spent most of his career mentoring graduate students and teaching ichthyology to undergraduates. He led and participated in many projects on research of flatfishes, deep-sea fishes, fish diversity, and conservation of freshwater and marine fishes. Kunio has also led a number of expeditions to undeveloped islands, such as the Kurile Islands. He has published more than 120 papers and 20 books (as editor and author) on taxonomy and systematics of flatfishes and deep-sea marine fish, as well as numerous faunal works, and work on early life history stages.



#### **M ERIC ANDERSON**

M Eric Anderson began his studies on deep-sea fishes in the early 1970s in the USA. He obtained an MA on the mesopelagic fishes of the Monterey Submarine Canyon at the Moss Landing Marine Laboratories on Monterey Bay, California. A PhD on the systematics of the Zoarcidae from the Virginia Institute of Marine Science followed. Eric specialises in the cold-water, high-latitude suborder Zoarcoidei, but also studies many other deep-sea groups, especially lanternfishes, dragonfishes, batfishes, deep-sea anglerfishes, grenadiers and cuskeels. He has published 90 scientific papers, usually as sole author, chiefly on deep-sea fish systematics and ecology, but also on their development and zoogeography, and on his fossil fish discoveries in South Africa. Eric has been a co-investigator aboard 28 deep-sea biological voyages in the North Pacific, North and South Atlantic and Western Indian oceans, and has dived twice in deep-sea research submersibles (California and the Arctic). His museum work includes study or collection in 25 countries and he has named as new to science 73 species, 12 genera and one subfamily of fish.



## WILLIAM D ANDERSON Jr

William D Anderson Jr received his PhD degree at the University of South Carolina in 1960 and spent the summer of that year on the coast of the Gulf of Mexico at the University of Texas, Institute of Marine Science (Port Aransas). During the academic year 1960–1961 he taught biology at Susquehanna University, Selingsgrove, Pennsylvania, and from July 1961 to August 1965 he was a fishery biologist at the United States Fish & Wildlife Service's biological laboratory in Brunswick, Georgia. Following that, he held an interim appointment for one year in the Department of Biological Sciences, University of Florida (Gainesville), followed by three years as an associate professor of biology at the University of Chattanooga in Tennessee, moving in 1969 to Charleston, South Carolina, to teach and do research in the College of Charleston's Department of Biology and its Grice Marine Biological Laboratory (located on James Island near Charleston Harbor). Anderson retired in June 1996, becoming an emeritus professor and continuing to study systematic ichthyology, publishing on the families Callanthiidae, Lutjanidae, Serranidae (subfamily Anthiinae), and Symphysanodontidae.



## **RACHEL J ARNOLD**

Rachel J Arnold is a marine sciences faculty member at Northwest Indian College in Bellingham, Washington State. She received her PhD from the University of Washington where her research focused on the evolution of the anglerfishes (Order Lophiiformes). Her research resulted in the addition of six newly described species, new information about distinct life histories and reproduction, and a novel hypothesis of the evolutionary relationships of anglerfishes based on molecular data. She is currently studying the population genetics of forage fishes important to the Coast Salish peoples. As a graduate student, she enjoyed being involved in and supervising undergraduate research and education, and therefore encourages undergraduates to become involved in her research as much as possible.



## NEIL C ASCHLIMAN

Neil C Aschliman completed his PhD at Florida State University under the supervision of Gavin Naylor. For his dissertation he generated a taxon- and data-rich molecular phylogeny of skates, rays, and allies (batoid fishes), against which he reinterpreted their body plan evolution and fossil record. He had previously researched the morphological systematics of batoids with John McEachran and of needlefishes with Bruce Collette, as well as zebrafish development and elasmobranch endocrinology. He has completed fellowships at the University of Tokyo's Ocean Research Institute and the Smithsonian's National Museum of Natural History, and has visited collections on four continents. His other interests include elasmobranch conservation and descriptions of new species. He is currently an assistant professor in the Biology Department at St Ambrose University in Davenport, Iowa.

#### MEBRAHTU ATEWEBERHAN

Mebrahtu Ateweberhan is co-author of the introductory chapter "The origins and geology of reefs of the Western Indian Ocean" in this book. He is a visiting Research Fellow at the University of Warwick, United Kingdom. Ateweberhan is a coral reef ecologist and environmental consultant involved in environmental impact assessments, marine environmental surveys. His interests include climate change, adaptations to climate change, and its impacts.



## DAVID R BELLWOOD

David R Bellwood is a professor of marine biology based in the Australian Research Council Centre of Excellence for Coral Reef Studies, and College of Science and Engineering, at James Cook University (JCU), Townsville, Australia. Although he studies reef fish ecology, palaeontology, systematics, taxonomy, biomechanics and behavioural ecology, he is best known for his functional studies of reef fishes. David started keeping reef fish in aquaria in the early 1970s in the UK, before studying biology at the University of Bath, training in fisheries (at Lowestoft, UK) and in taxonomy and systematics at the British Museum (Natural History) as it was then (now the Natural History Museum, London). His taxonomic work started with crustacea before he moved on to fossil then extant fishes. His key mentors were Geoff Boxshall and Howard Choat. With over 270 publications, David is one of the most published workers in coral reef studies (with an h-index of 69, and over 20 000 citations). His research achievements were recognised by JCU in 2004 when he was awarded a Personal Professorial (Research) Chair in Marine Biology, and in 2015 he was awarded the title, Distinguished Professor of Marine Biology. In the same year he was recognised by the Australian Society for Fish Biology for his "outstanding contribution to fish or fisheries science" with the K. Radway Allen Award. He is an editor of *Journal of Biogeography and Ecology Letters*, continues to teach three courses at James Cook University on fishes and biogeography, has supervised over 35 Honours and 30 PhDs; and continues to lead an active research group looking at the evolution and ecology of reef fishes.



## EUGENIA B BÖHLKE

Eugenia (Genie) B Böhlke was born in Washington State in 1928. While studying for her MS degree at Stanford University, she met and married James (Jim) E Böhlke, a student of the noted ichthyologist George S Myers. After completing graduate school, Jim took a position at the Academy of Natural Sciences of Philadelphia, where he and Genie spent the rest of their careers. Genie joined Jim in his ichthyological work, and the two of them formed a productive scientific team. She became particularly interested in moray eels (Muraenidae). After Jim's death in 1982, Genie continued her ichthyological studies, and eventually became the world's leading authority on morays. Her accomplishments include editing the volume on eels for the *Fishes of the Western North Atlantic* series. She formally retired in 1994, but continued her studies until shortly before her death in 2001.



#### JOHN C BRIGGS

John (Jack) C Briggs graduated from Oregon State University in 1943. From 1943 to 1946, he served as a pilot in the US Army Air Corps. Support from the "G.I. Bill" made it possible to enter Stanford University where he was employed as a teaching assistant in the Biology Department and earned an MA degree in 1947, then a PhD in 1952. His early studies were devoted to fish life history and systematics which led to an interest in the evolutionary implications of biogeographic patterns. Work on contemporary patterns of distribution and biodiversity resulted in the study of palaeobiology and the historical development of such patterns. After two years of post-doctoral work at Stanford, he held faculty positions at the University of Florida, University of British Columbia, University of Texas, and University of South Florida (USF). In 1990 he retired from USF and became an emeritus professor. For the following ten years he did research at the Natural History Museum, University of Georgia, then returned to the west coast, where he once again became affiliated with Oregon State University. Jack has produced 164 publications, including six books or monographs. In 2005, he received the Alfred Russell Wallace Award from the International Biogeography Society for his lifetime contributions to biogeography. Jack died in May 2018.



## **KENT E CARPENTER**

Kent E Carpenter is a professor in biological sciences at Old Dominion University in Norfolk, Virginia where he has been in residence since 1996. He also manages the Marine Biodiversity Unit of the Global Species Programme of the International Union for Conservation of Nature. He completed undergraduate studies in biology at the Florida Institute of Technology and his graduate studies in zoology at the University of Hawaii. He was a US Peace Corps Volunteer and post-doctoral research associate in the Philippines, a marine scientist at the Kuwait Institute for Scientific Research, and a senior fisheries research officer of the Food and Agriculture Organization of the United Nations in Rome, Italy. His primary research interests are marine conservation biology, systematics and evolution of bony fishes, ecology of coral reefs, and marine biogeography and phylogeography of the Indo-Pacific.



# JOHN H CARUSO

John H Caruso's research has focused on the systematics and distribution of chaunacid and lophiid anglerfishes. He received his PhD from Tulane University in 1977, and has worked at several academic institutions, including Lafayette College in Easton Pennsylvania, the Louisiana Universities Marine Consortium (LUMCON) in Chauvin, Louisiana, the University of New Orleans and Tulane University in New Orleans, Louisiana. Teaching was always his primary responsibility at these institutions, and this continues to occupy most of his time. He has also served as Curator of Fishes and Reptiles at Audubon Zoological Garden in New Orleans, and Interim Curator of Fishes for the Tulane Museum of Natural History. Currently, John is Emeritus Senior Professor of Practice at Tulane University, New Orleans, Louisiana, USA.



# PROSANTA CHAKRABARTY

Prosanta Chakrabarty obtained his BSc from McGill University and PhD from the University of Michigan in 2006. He has been Curator of Fishes at the Louisiana Museum of Natural History since August 2008, and is an associate professor in the Department of Biological Sciences at Louisiana State University (LSU). He is a systematist and an ichthyologist studying the evolution and biogeography of both freshwater and marine fishes. His work includes studies of Neotropical (Central and South America, Caribbean) and Indo-Pacific fishes. Collecting efforts by his laboratory at LSU include local and international collecting trips to Japan, Australia, Taiwan, Honduras, Guatemala, Kuwait, amongst others. He has described over a dozen new fish species, including several anglerfish and cavefish taxa new to science.



# FRANÇOIS CHAPLEAU

François Chapleau did undergraduate studies in biology and completed a MSc (on darters) at Université de Montréal (Montréal, Canada), and his PhD at Queen's University, Kingston, Canada. François is currently a professor in the Department of Biology of the University of Ottawa, Canada. Teaching in French and in English, he has won multiple teaching awards and was invited to teach a biology course in China. Throughout his long academic career (including his PhD and post-doctorates at the Australian Museum, Sydney, Australia and Canadian Museum of Nature, Ottawa, Canada), he has studied the systematics, taxonomy and ecology of freshwater and marine fishes. His taxonomic and systematics research, carried out in collaboration with students and colleagues, includes species revisions, phylogenetic analyses and morphological studies of several groups of flatfishes (Order Pleuronectiformes).



# DANIEL M COHEN

Daniel M Cohen received his BA degree from Stanford University in 1952 and a PhD under George S Myers in 1958 on the deep-sea family Opisthoproctidae. In 1959 Dan was hired as a systematist at the US Bureau of Commercial Fisheries at the Smithsonian Institution, Washington, DC, a position he held until transferring to the Alaska Fisheries Science Center in Seattle, Washington, from 1981 to1982. During his time at the Smithsonian, Dan developed research programmes on ophidiiform fishes, often in conjunction with Danish colleague, Jørgen G Nielsen, and gadiforms, culminating in the organisation of the 1986 Workshop on Gadiform Systematics at the Los Angeles County Museum of Natural History (LACM). In 1982 Dan accepted a position at the LACM which he held until retirement in 1995. He was a sea-going ichthyologist, and participated in several international collecting expeditions in the Pacific, Atlantic and Indian oceans, and authored more than 120 research papers on systematics and the biology of deep-sea fishes. He received several professional honours and awards. Dan died in September 2016.



## **BRUCE B COLLETTE**

Bruce B Collette received his PhD from Cornell University for a taxonomic study of a small group of freshwater fishes called darters that live along the coastal plain from Maine to Florida. In 1960, he accepted a position as an ichthyologist at the National Systematics Laboratory in what is now the National Marine Fisheries Service of the National Oceanographic and Atmospheric Agency, housed in the National Museum of Natural History of the Smithsonian Institution in Washington, DC. Bruce's research focuses on the anatomy, systematics, evolution, and biogeography of tunas and their relatives, plus other fishes such as halfbeaks, needlefishes, and toadfishes. Results of his research have been published in over 270 papers in many scientific journals. He is also the author of two regional fish books, *The Fishes of Bermuda* and *Bigelow and Schroeder's Fishes of the Gulf of Maine*, and an ichthyology textbook, *The Diversity of Fishes: Biology, Evolution, and Ecology*, now in its second (2009) edition and the most widely-used college-level ichthyology text in the world. He is Chair of the IUCN Species Survival Tuna and Billfish Specialist Group. Bruce is a past president of the American Society of Ichthyologists and Herpetologists, and has earned many honours such as the first Robert H. Gibbs, Jr. Memorial Award for an outstanding body of published work in systematic ichthyology, the Joseph S. Nelson Lifetime Achievement Award in Ichthyology, and a gold medal from the US Department of Commerce for his leadership in using IUCN Red List standards to assess extinction risk of tunas, mackerels, and billfishes.



## MRINAL K DAS

Mrinal K Das received his BSc (Honours) and MSc in Zoology from the University of Calcutta, Kolkata, India. He then received a Government of India Fellowship and joined the Zoological Survey of India as a research fellow under the supervision of Dr KC Jayaram and worked on the systematics of the cyprinid genus *Labeo*. To continue his passion for fish systematics, Mrinal joined the laboratory of Dr Joe Nelson at the University of Alberta (U of A), Edmonton, Canada, and received his MSc and PhD in Zoology from that university. After graduating, he worked as a member of the teaching faculty at Augustana University, Camrose, Alberta, Bishop's University, Lennoxville, Quebec, and the U of A. He received a U of A Graduate Student Teaching Award in 1992 and a Teaching Excellence Award from the Alberta Chapter of the Delta Chi in 1999. In 2001, Mrinal joined Grant MacEwan University (then College) as an instructor of Biological Sciences. His research interests are the systematics and phylogeny of the fishes of the Suborder Trachinoidei and hybridisation in *Phoxinus eos* and *P. neogaeus*.



## MARCELO R DE CARVALHO

Marcelo R de Carvalho obtained his PhD in 1999 from the American Museum of Natural History/City University of New York joint programme in Evolutionary Biology, studying under Gareth Nelson and John Maisey. His greatest interests are in the systematic morphology and comparative anatomy of chondrichthyan fishes, both fossil and living. He has published widely on the systematics and morphology of sharks and rays, including the description of many new species of living electric rays and Neotropical freshwater stingrays, as well as new genera and species of fossil thornback rays, electric rays, and marine stingrays. As a professor in the Department of Zoology of the University of São Paulo, he taught vertebrate zoology, comparative anatomy, ichthyology, and systematics, and led a large laboratory with post-doctoral, graduate and undergraduate students, that focused on different aspects of chondrichthyan evolution. Major studies undertaken included taxonomic revisions of potamotrygonid stingrays and of many marine genera from the southwestern Atlantic Ocean and worldwide, as well as investigations in the comparative anatomy and development of sharks and rays.



## MARTINE DESOUTTER-MENIGER

Martine Desoutter-Meniger has worked at the Museum national d'Histoire naturelle in Paris for more than forty years, much of that time in the Collections of Fishes. She has worked as an alpha-systematist on the Order Pleuronectiformes, better known as flatfishes, and especially on the species belonging to the family Soleidae. She has produced many papers on the systematics of the Soleidae in collaboration with Dr François Chapleau of the University of Ottawa, Ontario, Canada. Identifying the numerous fishes collected by different oceanographic expeditions is one of her main activities. She uses morphological and osteological features to describe species and also collaborates with a team working on the phylogeny of acanthomorph fishes, using both morphological and molecular features.



# FABIO DI DARIO

Fabio Di Dario received his PhD in Zoology from the University of São Paulo (USP), Brazil, in 2005 after conducting studies in the Museum of Zoology (MZUSP) and the National Museum of Natural History, Smithsonian Institution. His research focuses on morphology, systematics (taxonomy and phylogeny) and conservation of fishes, especially clupeomorphs and basal teleosts. His passion for African fishes and the history of ichthyology in the continent began as a child when he first learned about the coelacanth discovery in South Africa. His experiences in African countries include Benin, Ethiopia and Burundi, where he attended the Third, Fourth and Fifth Meetings of the Pan African Fishes and Fisheries Association (PAFFA) respectively, in addition to academic visits to Mozambique and Tanzania, and a four-month residence at SAIAB between 2015 and 2016. Fabio is Professor of Zoology at the Federal University of Rio de Janeiro, in the city of Macaé, Rio de Janeiro, Brazil. He is also a member of the Brazilian Society of Ichthyology (SBI) and the Deep-Sea Fishes Research Group (GEOProf).



# RYU DOIUCHI

Ryu Doiuchi obtained a PhD in Agriculture in 2005 at Kyoto University, Japan, where he worked on the phylogeny of the Stromateoidei using both morphological and genetic techniques. While at Kyoto University he also investigated the taxonomy of Indo-Pacific species of the Sphyraenidae, including a description, with his supervisor Dr Tetsuji Nakabo, of a new species from Japan, *Sphyraena iburiensis*, which had been caught in abundance by local commercial fishermen but had been confused with similar species. Ryu is a researcher in the Marine Resources Department of Fisheries Experimental Station at Wakayama, Japan, and is working on the resource management of several fish species, such as *Trichiurus japonicus* (Trichiuridae), *Parapristipoma trilineatum* (Haemulidae), *Pagrus major* (Sparidae), and *Paralichthys olivaceus* (Paralichthyidae), all commercially important fishes in Japan. He has authored more than ten academic papers, reports and other publications on fish taxonomy and marine ecology, an article focusing on age and growth, reproductive biology, and other ecological traits of *P. trilineatum*. He is particularly interested in understanding the rich and diverse fish fauna in Wakayama, the coast swept by the Kuroshio Current.



## **TERRY J DONALDSON**

Terry J Donaldson is Professor of Ichthyology and Director of the University of Guam Marine Laboratory. He was educated at Michigan State University (BS Fisheries, 1977), the University of Guam Marine Laboratory (MS Biology, 1981) and the Louisiana State University Museum of Natural Science (PhD Systematics and Evolutionary Biology – Ichthyology, 1988). His graduate education was supplemented by work undertaken at the Marine Biological Laboratory, Woods Hole, the Laboratory of Ethology at Illinois State University, and the University of Arizona. He was a Japan Society for the Promotion of Science Fellow at Kyushu University (Japan, 1993–1994) and a US National Science Foundation, Japan Society for the Promotion of Science Research Fellow, also at Kyushu University (1997–1998). He is the author or co-author of several papers and book chapters in fish taxonomy, biogeography and biodiversity, behavioural ecology (especially mating systems and habitat utilisation), and conservation biology (extinction risk analyses). Terry has worked variously in government, in a conservation non-governmental organisation, and in academia. Most of his work has been undertaken in the Indo-Pacific region.



## JAMES K DOOLEY

James (Jim) K Dooley obtained a BS in Biology from the University of Miami (Miami, Florida), an MS in Zoology from the University of South Florida (Tampa), and a PhD in Zoology from the University of North Carolina (Chapel Hill). After receiving his bachelor's degree, he worked for two years at the University of Miami Rosenstiel School of Marine and Atmospheric Sciences; on projects for the United States Navy's (ASW) Bioacoustic Programme in the Bahamas; for juvenile shrimp and sports fisheries in the Everglades National Park; and at the National Oceanic & Atmospheric Administration (NOAA), Tropical Atlantic Biological Laboratory in the fish museum collections. For his MS thesis at the University South Florida, he worked on the pelagic sargassum community in the Florida Current, and in his early years at Adelphi University he was Principal Investigator for NOAA/ICAT Atlantic Bluefin Tuna sports fishing project, for New York State. Other early projects included research at Friday Harbor Laboratory, University of Washington, on homing behaviour of sculpins; and Principal Investigator for Earthwatch on a study of shore fishes of the Canary Islands, Spain, He is Professor of Biology at Adelphi University, Garden City, New York, and also served as Biology Department Chair for nine years and as head of the Science Division for Adelphi University. Jim has been a fisheries consultant for United Nations FAO and consultant for IUCN; board member of New York State Marine Sciences Consortium; and private consultant for numerous New York, Connecticut, Pennsylvania and Massachusetts environmental companies for freshwater environmental impact projects. He has published more than 50 papers, numerous book chapters, and is co-author of two books on biology laboratory procedures. His ongoing research is on the tilefish families Branchiostegidae and Malacanthidae, including 43 species worldwide, based on morphology and molecular characters, and describing 12 new species.



# DAVID A EBERT

David A Ebert earned his BA degree from Humboldt State University in northern California, a Masters from Moss Landing Marine Laboratories in central California, and his PhD at Rhodes University, Grahamstown, South Africa. He is Programme Director for the Pacific Shark Research Center, a research faculty member at Moss Landing Marine Laboratories, and an honorary research associate for the South African Institute for Aquatic Biodiversity and the California Academy of Sciences Department of Ichthyology. He serves as an advisor and consultant for the Food and Agriculture Organization of the United Nations on shark-related issues, and is the IUCN Shark Specialist Group Vice Chair for taxonomy, regional co-Chair of the IUCN Northeast Pacific Regional Shark Specialist Group, and a member of the American Elasmobranch Society, American Society of Ichthyologists and Herpetologists, and Oceania Chondrichthyan Society. Dave's research on sharks and their relatives (the rays, skates and ghost sharks) focuses on the biology, ecology and systematics of this enigmatic fish group. He has conducted research on six continents and in over 25 countries. He has authored 25 books, including the Sharks of the World, has published over 400 scientific papers and book chapters, and contributed over 120 IUCN Shark Specialist Group Red List species assessments. His research has been featured on Discovery Channel's Shark Week and on the acclaimed BBC series Shark. He has supervised more than 40 graduate students, and enjoys mentoring and helping develop aspiring marine biologists. His current research efforts are focused on finding, documenting, and raising awareness of the world's 'lost sharks', those little-known or unknown shark species that have largely been lost from the public and scientific awareness.



## THOMAS H FRASER

Thomas H Fraser received a PhD in 1970 from the University of Miami, Florida, for an osteological study of the family Apogonidae that initially set the modern classification for 19 genera and 14 subgenera. He spent three years at Rhodes University, South Africa, as a senior lecturer at the JLB Smith Institute of Ichthyology studying marine fishes. He has collected Indian Ocean fish in South Africa, Mozambigue, Mauritius, and twice on St Brandon Shoals (Cargados Carajos Bank). Another year was spent at the Smithsonian Institution as a post-doctoral fellow studying apogonids with Ernest A Lachner at the Division of Fishes. A private-sector position at Charlotte Harbor, Florida, allowed Tom to set in motion the descriptive research on water quality, primary production and fish communities in Florida's second largest estuary affected by freshwater inflow. He was appointed successively by two Governors of Florida to the Florida Marine Fisheries Commission, the rule-making body for living marine resources, and provided its recommendations for guiding state policies on marine natural resources to the governor and legislature. He served for eight years, two years as Vice-Chairman and four years as Chairman. Tom has published or co-authored at least 59 papers, many about the systematics and taxonomy of the Apogonidae, describing new subfamilies, tribes, genera (fossil and recent), and many new species. A research fellowship at the Australian Museum, Sydney, set in motion a renewed study of cardinalfish. He continues to be involved with the description of new species and genera of cardinalfishes as an adjunct scientist at Mote Marine Laboratory, Sarasota, Florida, and as a research associate at Florida State Museum, University of Florida, Gainesville, Florida. In the emerging field of molecular phylogeny he has joined Japanese colleagues in research concerning relationships within the Apogonidae as expressed by colour patterns, morphology and DNA. Tom has assessed apogonids in the Caribbean and Gulf of Mexico as part of the IUCN programmes.



## **RONALD FRICKE**

Ronald Fricke studied in Braunschweig, London, Hamburg and Freiburg, before being appointed Senior Curator of Fishes at the Staatliches Museum für Naturkunde Stuttgart, Germany. Apart from the collection of and research on freshwater fishes of central Europe that this position requires, his wider research interest focuses mainly on the taxonomy, zoogeography and evolution of marine fishes of the Indo-Pacific, eastern Atlantic and Mediterranean Sea. After initial revisionary studies on callionymid, draconettid and tripterygiid fishes, he expanded his taxonomic range to syngnathids, gobiesocids, blenniids, gobiids and other fish families, but specialised on small and sometimes cryptic fish species. In the WIO he focuses on the Red Sea, the Persian/Arabian Gulf and southern Arabia, and the Mascarenes and Esparses Islands. Outside the region, he works on the fish fauna of New Caledonia, New Guinea, the South China Sea, the Mediterranean, Canary Islands, Cape Verde Islands, Ascension and St Helena islands. He has collected fishes in 63 countries and participated in several research expeditions. In addition, he has been working in conservation of marine and freshwater habitats and species as an independent expert for the European Union for the NATURA 2000 protected area network, and has published several Red Lists of fishes in Europe and overseas. He also acted as national representative in the OSPAR, HELCOM and ICES organisations. Fricke has published the descriptions of eight new genera and 149 new species of fishes. He has authored 471 scientific papers, including nine books as a single author, five books with multiple authors, and 174 refereed papers in journals. He is a co-editor of Eschmeyer's Catalog of Fishes online database.



## JAVAD GHASEMZADEH

Javad Ghasemzadeh is an associate professor in the Department of Fisheries and Aquaculture, Faculty of Marine Sciences at Chabahar Maritime University, Iran. He completed his undergraduate studies in Fisheries and Natural Resources Management at Tehran University, and worked for a few years in the Iranian Fisheries Organization before moving to Australia to pursue his postgraduate studies in 1990. He did his MSc in Marine Ecology at the University of New South Wales, and his interest in ichthyology motivated him to work on the taxonomy and phylogeny of Indo-Pacific mullets (family Mugilidae), with special reference to the mullets of Australia, for his PhD at Macquarie University. For this project he collected mullets from all around Australia and many parts of the Indo-Pacific, and examined the extant type specimens of mugilids in Australia and most of the European museums. During his studies he collaborated closely with the Ichthyology Department of the Australian Museum, and after graduation, he examined all the mullet holdings of this museum through a visiting fellowship in 1998. Apart from teaching and guiding postgraduate students, he is actively involved in research and scientific collaboration with interested ichthyologists in resolving the very ambiguous and confusing systematics and phylogeny of mullets.



# ANTHONY C GILL

Anthony (Tony) C Gill is Natural History Curator in the Macleay Museum, University of Sydney, Australia. He previously held positions in the Natural History Museum, London, and in the School of Life Sciences and International Institute for Species Exploration, Arizona State University. Tony received his PhD from the University of New England, Australia, in 1991, based on research conducted at the Australian Museum, Sydney. He was later a Smithsonian post-doctoral fellow in the National Museum of Natural History, Washington, DC, and Lerner-Grey Research Fellow in the American Museum of Natural History. His research is primarily on the taxonomy and historical biogeography of Indo-Pacific shorefishes, and on the systematics and anatomy of acanthomorph fishes. Among other topics, he has published on the classification and taxonomy of the Indo-Pacific dottyback family Pseudochromidae, and on the classification of the perciform suborders Gobioidei and Percoidei.



# **ULISSES L GOMES**

Ulisses L Gomes was awarded his PhD by the Museu Nacional, Rio de Janeiro, Brazil in 2002. Since 1981 he has been a professor in the Zoology Department of the Universidade do Estado do Rio de Janeiro, Brazil. Dr Gomes has been conducting research and publishing papers on the anatomy and taxonomy of elasmobranchs, describing new species of Rajidae and Dasyatidae since the early 1980s. He is member of the Sociedade Brasileira de Ictiologia and the American Society of Ichthyologists and Herpetologists. Recently, he published as lead author an identification guide for sharks and rays known to occur in Rio de Janeiro state, Brazil.



# MARTIN F GOMON

Martin F Gomon is a long-standing senior curator of Ichthyology at Museum Victoria, the state museum of Victoria in Melbourne, Australia, and is a senior member of Australia's ichthyological community where he is regarded as a taxonomic authority on fishes in that country. Educated in the USA, he received degrees at Florida State University (BSc) and University of Miami (MSc, PhD), during which time he spent five years at the National Museum of Natural History, Smithsonian Institution, as a museum technician in the Fish Division and a finishing graduate student. Martin's research began with a focus on interrelationships within the Labridae, which provided an early advocacy for a wider view of the family incorporating the Scaridae and Odacidae, based on phylogenetic principles. Arriving in Australia, he turned his attention to other groups and has published taxonomic descriptions of species of a wide assortment of families from the Scyliorhinidae to the Soleidae, as well as labrids, with current work incorporating genetic and morphological methodologies investigating the Aulopidae, Chlorophthalmidae and Paraulopidae. Many of his studies arose from efforts to elucidate the taxonomy of temperate Australian species in preparation for what have become the most comprehensive field guides to southern Australian fishes published, Fishes of Australia's South Coast (Gomon, Glover & Kuiter 1994) and its successor Fishes of Australia's Southern Coast (Gomon, Brav & Kuiter 2008). A founding member of OzFishNet, an Australasia-based consortium of fish taxonomists organised to promote and disseminate information about Australia's fish diversity, he continues to work collaboratively and attract funding for projects, such as the consortium's Fishes of Australia website launched in 2012 (www.fishesofaustralia.net.au).



## OFER GON

Ofer Gon completed his MSc in oceanography and marine biology at the Hebrew University, Jerusalem, under the supervision of A Ben-Tuvia in 1979. His first research experience was at the Bernice Pauahi Bishop Museum, Honolulu, working on fishes of the tropical and subtropical Pacific Ocean under JE Randall. Professionally, this was a very important period as he acquired first-hand knowledge of many families, genera and species of fishes and published his first taxonomic papers on new species and records of fishes from Hawaii. In 1982 he was hired by Margaret Smith of the then JLB Smith Institute (now the South African Institute for Aquatic Biodiversity) in Grahamstown, South Africa, to assist with the preparation of *Smiths' Sea Fishes*. This was followed by a contract to produce *Fishes of the Southern Ocean* and a permanent research position at the Institute. In the course of his long career Ofer studied the taxonomy of mostly marine tropical and Antarctic fishes, but was also involved in some estuarine and freshwater fish studies while supervising students. He retired in 2014 and is now Curator Emeritus at SAIAB.



## WAYNE S GOSCHEN

Wayne S Goschen obtained his BSc (Hons) degree in physics from the University of Natal (now UKZN). He then chose to switch to physical oceanography where his talent for applied mathematics and physics could be put to practical use. This led to a study of ocean dynamics associated with prominent capes and bays off the south coast of South Africa and to a PhD in Oceanography from the University of Port Elizabeth (now NMMU) in 1991. Wayne's research included a study of features of the Agulhas Current in the southeastern region of the Western Indian Ocean and its influence on the adjacent coastal ocean. He then spent 15 years in the IT industry developing software, databases and websites (culminating in an MSc in IT from UCT) before returning to oceanography in 2008 with the South African Environmental Observation Network (NRF-SAEON). His interests are in the physical oceanography of nearshore, coastal and shelf waters, mostly along the east coast of South Africa and around Algoa Bay, but also including the Agulhas Current. His advances in research have been in the field of wind-driven circulation and coastal upwelling, Agulhas Current influences on shelf and coastal waters, coastal trapped waves and dynamics of large log-spiral bays.



#### GAVIN GOUWS

Gavin Gouws is a former senior scientist at the National Research Foundation – South African Institute for Aquatic Biodiversity (NRF-SAIAB) in Grahamstown, South Africa. He initially intended training as a broad zoologist but was bitten by the genetics bug during his postgraduate studies. He became excited by this developing field and the possibilities it opened up to address a range of questions around systematics and evolution, irrespective of the organism and the system. He subsequently chose to pursue studies on freshwater isopods, crabs and crayfish in South Africa and Australia, and was eventually employed by NRF-SAIAB as a fish taxonomist. Using genetics as an entry point, his research programmes have involved large-scale studies of reef fish biogeography, phylogeography and diversity across the Western Indian Ocean, more-focused studies on the systematics and taxonomy of specific groups (e.g., goatfishes, snappers, emperors and cardinalfishes), and population genetic studies of species of commercial and subsistence fisheries importance. His more recent or esoteric research interests include genomics, epigenetics, metagenomics, and the study of sexual selection and the evolutionary biology within certain fish families.



#### NICHOLAS AJ GRAHAM

Nicholas AJ Graham is a chief investigator and Principal Research Fellow in the ARC Centre of Excellence for Coral Reef Studies, James Cook University, Australia. He is funded through an Australian Research Council Fellowship. Prior to being based in Australia, Nick completed his PhD in the School of Marine Science and Technology, Newcastle University, UK. His research tackles large-scale ecological and social-ecological coastal issues under the overarching themes of climate change, human use and resilience. He has worked extensively on the ecological ramifications of fishing and closed area management. He has assessed the long-term impacts of climate impacts on coral reef fish assemblages, fisheries and ecosystem stability. He has studied the patterns and processes by which degraded coral reefs recover, and how this can be incorporated into, or influenced by, management action. Increasingly Nick works with social scientists and economists to assess methods of linking social-ecological systems for natural resource assessment and management. Much of his work is in the Western Indian Ocean, particularly Seychelles, Chagos, Maldives, Mauritius, Kenya and Tanzania.



#### DAVID W GREENFIELD

David W Greenfield is a research associate in the Department of Ichthyology at the California Academy of Sciences in San Francisco and an emeritus professor at the University of Hawaii, where he taught ichthyology. He is also a research associate at the Bernice Pauahi Bishop Museum in Honolulu, and a past president of the American Society of Ichthyologists and Herpetologists. David has published over 150 scientific papers on fishes and a book, *Fishes of the Continental Waters of Belize*, and has described over 100 species new to science, mainly from the families Batrachoididae, Holocentridae and Gobiidae. He has conducted extensive field work on both freshwater and marine fishes in North, Central and South America, as well as in Hawaii and Fiji. His latest research is on the systematics of Indo-Pacific coral-reef fishes, particularly the gobiid genus *Eviota*.



# **ELAINE HEEMSTRA**

Elaine Heemstra was introduced to fishes and fish illustration at the JLB Smith Institute (now SAIAB) where she has spent most of her career, and also where she met and married her ichthyologist husband, Phil Heemstra. Later she succumbed to the attraction of fishes and became fascinated by their beauty, form and variety. Phil introduced her to scuba diving and they dived together whenever possible. They also collaborated on numerous scientific papers as well as a book on the coastal fishes of southern Africa. In preparation for this WIO book, Elaine and Phil undertook several fish survey expeditions to learn more about the fishes of the region. Wherever they collected fishes they met fellow enthusiasts keen to share their local knowledge and photographs. This book is a record of an ongoing quest to learn more about WIO fishes and is also a celebration of the camaraderie Elaine and Phil encountered *en route*.



# PHILLIP C HEEMSTRA

Phillip (Phil) C Heemstra, an American ichthyologist, completed his MSc and PhD in marine biology at the University of Miami in the late 1960s and early 1970s. After some early work on taxonomy of sharks of the Gulf of Mexico and Caribbean, and various bony fishes in Miami, Phil accepted a position at the Academy of Natural Sciences in Philadelphia. After a year in Philadelphia, he took up a three-year position in the Australian CSIRO Fisheries Division at Cronulla. After the contract finished in 1978, Phil accepted an offer from Margaret Smith of the JLB Smith Institute of Ichthyology (now South African Institute for Aquatic Biodiversity) in Grahamstown, South Africa, to help her write a revision of her husband's book, the *Sea Fishes of Southern Africa*. This was a research post and Phil became Senior Curator of Fishes. Phil did much work in the WIO, collecting the fishes that laid the foundation for this book. He participated in R/V *Dr Fridtjof Nansen* research cruises off Kenya and Mozambique, and organised or participated in fish surveys of Mauritius, Rodrigues, Madagascar, Comoros, Seychelles and South Africa. He was honorary foreign member of the American Society of Ichthyologists and Herpetologists. Phil died in 2019 after a long illness.



# DANNIE A HENSLEY

Dannie A Hensley was a faculty member at the University of Puerto Rico, Mayagüez, for 28 years where he served as an assistant professor from 1980 to1984, an associate professor from 1984 to 1991, and was promoted to a full professor in 1991. His education in ichthyology started at the California State University, Fullerton, in 1970, where his advisor was David W Greenfield. From 1974 to 1978, Dannie was an ichthyologist with the Marine Research Laboratory of the Florida Department of Natural Resources in St Petersburg. For much of that time, he studied toward his PhD at the University of South Florida under John (Jack) C Briggs' guidance. It was during this time that he wrote his first paper on flatfishes, describing the larval development of *Engyophrys senta* (Bothidae). In it he discussed the systematic importance of intermuscular bones in flatfishes, and became the first author to use phylogenetic terminology when discussing relationships of flatfishes. Perhaps Dannie's most significant contribution to the classification of flatfishes was a chapter he wrote with Elbert Ahlstrom for Ahlstrom's 1984 volume on the development of flatfishes, and his synthesis is still widely cited. Dannie died in 2008. His final contributions to fish systematics were the accounts he wrote for this book, which were revised and edited by Kunio Amaoka.



# ERIC J HILTON

Eric J Hilton received his bachelor's degree in Wildlife Ecology and Management in 1996 and his PhD in Organismic and Evolutionary Biology in 2002, both from the University of Massachusetts, Amherst. Before coming to the Virginia Institute of Marine Science (VIMS) in 2007, he conducted post-doctoral research in the Geology Department at the Field Museum of Natural History, Chicago, and the Division of Fishes of the US National Museum of Natural History (Smithsonian Institution) in Washington, DC. He is a broadly trained vertebrate zoologist, with a primary research interest in the anatomy, evolution and diversification of the ray-finned fishes. He uses a multidisciplinary approach to study evolution, drawing on the data and techniques of palaeontology, comparative anatomy, developmental biology, biogeography, and histology. Other ongoing research projects include the ecology of larval fishes, the biology and management of Atlantic sturgeon, and monitoring the population of anadromous shad and herring in Virginian waters. As a faculty curator, he oversees the growth and use of the VIMS Nunnally Ichthyological Collection, in addition to teaching ichthyology and training graduate students and post-doctoral researchers.



# HSUAN-CHING HO

Hsuan-Ching (Hans) Ho is an associate research fellow of the National Museum of Marine Biology and Aquarium and an associate professor of the National Dong Hua University in Taiwan. He started to work on fish taxonomy as a graduate student in Academia Sinica in 2002 and obtained his PhD degree in 2010 with his thesis on the phylogenetic relationship and revision of Indo-Pacific genera of the batfish family Ogcocephalidae. He is interested in various fish groups, especially the anglerfishes, sand perches, lizardfishes and many other deep-sea fishes. He has published more than 80 scientific papers and described 45 new fish species. He has also spent a lot of time on the history of the work on Taiwanese fishes, and he hopes to review all Taiwanese fishes in the future.



# WOUTER HOLLEMAN

Wouter Holleman became involved in taxonomy as a postgraduate student at the request of Rick Winterbottom, then a lecturer at the JLB Smith Institute (now the South African Institute for Aquatic Biodiversity [SAIAB]), who needed some tripterygiids identified. In 1979 he moved to the Albany Museum and became involved in museum management for more than 20 years, whilst maintaining his interest in Tripterygiidae. After early retirement from the museum he moved back to SAIAB as a research associate. In the following ten years his research was interrupted by periods spent teaching English in Taiwan. It was during this time that he was asked to take on this WIO book project as an editor, and in between, he became involved in the taxonomy of South African clinids and some other groups of fishes. He is also on the team of fish editors for *Zootaxa*.



## **BRETT A HUMAN**

Brett A Human completed his PhD in 2003 in South Africa under the supervision of Dr Leonard Compagno at the University of Cape Town and South African Museum. For his thesis, he revised three African catshark genera, performed shape variation analyses on those genera, and published one of the first molecular phylogenetic studies on chondrichthyans. While in South Africa, Brett participated in the dissection of the first megamouth shark recorded from WIO. From 2006–2008 Brett was employed at the Marine Science and Fisheries Centre in the Sultanate of Oman. During this period he spent 112 days at sea where he collected over 2 000 specimens of marine fishes and established a fish collection there. Before his death in 2011, Brett lived in Perth, Australia, his country of origin, where he was continuing his work on elasmobranch taxonomy through the Western Australian Museum. Brett authored 11 chapters on various shark families for this book.



## **J BARRY HUTCHINS**

J Barry Hutchins was Curator of Fishes at the Western Australian Museum, Perth, until his retirement in 2007. His research was centred on the taxonomy and phylogeny of the Monacanthidae (filefishes) and Gobiesocidae (clingfishes), but also included studies of the biodiversity of the Western Australian reef fish fauna. After his retirement he maintained his connection with the museum as a research associate. Barry continues to work on the recruitment of tropical reef fishes at Rottnest Island, which is located off the Western Australian coast near Perth. This long-term investigation, started in 1976, has shown remarkable fluctuations in annual tropical recruitment that follow the El Niño/La Niña pattern of ocean circulation in the Australian region.



#### **HITOSHI IDA**

Hitoshi Ida is an emeritus professor of the School of Marine Biosciences, Kitasato University, Kanagawa, Japan. His research interests are ecology of the fishes of the family Salmonidae and he was the first to report the density dependence of *Oncorhynchus keta* in the northern Pacific in the early 1980s. He also cautioned against the practice of stock enhancement of the species, by the release of fry into the environment. He reviewed fishes of the family Ammodytidae and described two genera, viz. *Protammodytes* from continental shelves of both the tropical Pacific and Atlantic, and *Lepidammodytes* from Hawaiian waters. In this review he also described new species of *Protamodytes* and *Bleekeria* from the southern Indian Ocean with John Randall, with a focus on specialisations in the families of Perciformes, which accompany their sand-diving behaviour, such as elongation of the body, loss of scales, protrusion of lower jaw, degeneration of pelvic fins and specialisation of the olfactory organs. He retired from Kitasato University in 2007, but continued to focus on his interest in the fish diversity of Palauan waters. He contributed a description of the most primitive form of an anguilliform *Protanguilla palau* from an underwater cave in Palau. The taxon probably represents a new suborder of the order Anguilliformes.



## WALTER IVANTSOFF

Walter Ivantsoff was born in Shanghai, China, in 1938, to Russian parents whose own parents had left Russia at the time of the Russian Revolution in 1917. Walter attended several schools whilst in China, the most important of which was a school run by English and French Jesuit monks. In 1949, the United Nations Refugee Organization (IRO) was instrumental in moving Russian refugees out of China to a temporary location in the Philippines (a small island, Tubabao, off Samar Island) where he enjoyed living in a tent for 11 months with no school to go to. At the end of that year his family was accepted by Australia and he began his education which culminated in a PhD from Macquarie University, Sydney, where he remained as teacher and researcher for the rest of his working life. Walter published 60 research papers, mostly with his postgraduate students and colleagues, restricting his interests to families Atherinidae, Notocheiridae, Atherinonidae, and Pseudomugilidae, with some publications on melanotaeniids and telmatherinids. Walter died on 13 March 2019.



#### YUKIO IWATSUKI

Yukio (Yuk) Iwatsuki is a professor in the Department of Marine Biology & Environmental Sciences, Faculty of Agriculture, University of Miyazaki, Japan. His research interests are the taxonomy, speciation, biogeography and rich resources of coastal fishes (snapper, silverbiddy, seabream, grunt, and others). He served as an editor of the Ichthyological Society of Japan, and as a member of the IUCN Snapper, Seabream and Grunt Specialist Group.



#### **KELSEY C JAMES**

Kelsey C James is pursuing a PhD at the University of Rhode Island with Dr Lisa Natanson and Dr David Bengtson. Her research focus is on the age and growth of skates and rays. This complements her MSc in Marine Science from Moss Landing Marine Laboratories under Dr Gregor Cailliet and Dr David Ebert where she worked on the life history of the starry skate. She also described a new species of chimaera, *Hydrolagus melanophasma*, from the eastern North Pacific, and compiled and managed a global database on fisheries bycatch of marine megafauna (chondrichthyans, marine mammals, seabirds and sea turtles) at San Diego State University. She developed her passion for and skills in animal husbandry and scientific collecting during her BS at University of California, Santa Barbara.



# PATRICIA J KAILOLA

Patricia J Kailola is an Australian who has spent most of her working life in the Pacific region, with shorter periods in Australia and Southeast Asia. Her interest is in tropical fish systematics (emphasis on ariid catfishes), zoogeography, fisheries management and tropical aquatic biodiversity (particularly invasive fish species). Patricia co-authored *Continental shelf fishes of northern and north-western Australia: An illustrated guide* (1985, with K Sainsbury and G Leyland) and *Trawled fishes of southern Indonesia and north-western Australia* (1984, with Thomas Gloerfelt-Tarp). In Papua New Guinea, she maintained and added to the national fish collection for many years and later collated a substantial library of 'grey' fisheries literature (2003, *Aquatic resources bibliography of Papua New Guinea*). Patricia lives most of the time in the Republic of the Fiji Islands where she works with a human-rights orientated national non-governmental organisation and fulfils her obligations as an Honorary Fellow at the Institute of Applied Sciences, University of the South Pacific.



## ERI KATAYAMA

Eri Katayama received her BSc, MSc and PhD in Biology at the Kochi University in Japan from 2000 to 2011. She continued her studies for a further year in the Laboratory of Marine Biology under Dr Hiromitsu Endo and worked on a taxonomic review of the family Trichonotidae at the Laboratory of Marine Biology. She started to dive around Shikoku Island in southern Japan in 2004 and collected many shore fishes. She has participated in compiling several field-guide books and checklists of islands of southern Japan. In 2012 she moved to Tsukuba to work as a research assistant in the Department of Zoology, National Museum of Nature and Science, where she worked on building a database of SEM images. In 2017 she started to work at the Research Institute of Marine Invertebrates, where she is working to coordinate these supporting projects for Japanese researchers of the marine invertebrates. Her current research is a taxonomic study of the genus *Limnichthys* using morphology and molecular analysis. She is particularly interested in the systematics, biogeography and diversity of shore fishes, such as sanddivers, clingfishes and gobies.



#### **TOSHIO KAWAI**

Toshio Kawai obtained a PhD degree from the Graduate School of Fisheries Sciences, Hokkaido University, Hakodate, Japan, in 2006. He joined several deep-sea research cruises in Japanese, Indonesian and Peruvian waters during his school days. He worked in Bali, Indonesia, as a senior researcher at a private research laboratory sponsored by Kyowa Concrete Industry Co., Ltd, in July 2006 until March 2007, during which time he researched the marine environment around Bali by remotely operated vehicle (ROV). He returned to Japan in April 2007, and worked in the Fish Division of the National Museum of Nature and Science, Tokyo, as a research assistant for two years. Thereafter he became an assistant professor at the Hokkaido University Museum, and advises on the ichthyology course in the School of Fisheries Sciences and the Graduate School of Fisheries Sciences, Hokkaido University. In addition, Toshio is the collection manager for fishes in the Fisheries Science Center, which is a branch of the Hokkaido University Museum with more than 200 000 fish specimens that have been deposited in the branch. He also runs an annual parataxonomist training course in ichthyology. Toshio's research interests focus on the systematics, taxonomy and morphology of Peristediidae. He is dedicated to documenting marine fish fauna in northern Japan, including Hokkaido, which is the northernmost main island in Japan, where several species new to the area are discovered every year. He is also interested in marine fish fauna around Southeast Asia, especially the deep-sea fishes. He has authored more than 50 academic papers, reports and books.



#### JENNY M KEMPER

Jenny M Kemper received her BS in Biology from Florida State University in 2007 and her MS in Marine Biology with the Pacific Shark Research Center at Moss Landing Marine Laboratories in 2012. Her MS thesis research was on the feeding ecology of skates in Prince William Sound, Alaska. While working on her MS thesis, Jenny described two new species of chimaeroid fishes, one from South Africa, and the other from the Bahamas, which stimulated her interest in taxonomy and systematics. In 2012, she began her PhD at the Medical University of South Carolina in the Marine Biomedicine and Environmental Sciences Programme. Her dissertation research is on the phylogenetics of extant chimaeroid fishes (Chimaeriformes), an important yet understudied group of chondrichthyan fishes. Her goal is to use molecular data to estimate the evolutionary relationships of chimaeras, with comprehensive species sampling and the use of both nuclear and mitochondrial markers. She has further described another species of chimaera from New Zealand in 2015. She has co-authored several book chapters on chimaeras. Her main research interests are phylogenetics and systematics of chimaeroid fishes, cytogenetics, and understanding sex determination mechanisms and chromosome evolution in vertebrates.



## SEISHI KIMURA

Seishi Kimura received his BSc and MSc degrees in fisheries science from Mie University, Japan, and was then employed as an assistant professor at the Fisheries Research Laboratory, Mie University (FRLM), located in Shma, Mie Prefecture, in 1978. He studied fisheries ecology of some commercial fishes, especially the haemulid *Parapristipoma trilineatum*, and obtained a PhD degree from Kyoto University in 1987. Seishi is now a professor and director of FRLM, and his research interests are the taxonomy, phylogeny and zoogeography of Indo-Pacific fishes, especially Atherinidae, Carangidae and Leiognathidae. He is also interested in coastal fish fauna in Southeast Asia. He served as Editor-in-Chief (2004–2007) and President (2012–2013) of the Ichthyological Society of Japan. He has published more than 150 scientific papers and edited three field guides of fishes: *Fishes of Bitung, Northern Tip of Sulawesi, Indonesia; Fishes of Liobong Island, West Coast of Southern Thailand;* and *Fishes of Andaman Sea West Coast of Southern Thailand*. His latest work is on *Fishes of Ha Long Bay, Northern Vietnam*.



## **LESLIE W KNAPP**

Leslie W Knapp obtained his BS from Cornell University in 1952, an MA in Fisheries Biology in 1957, and a PhD in Vertebrate Zoology in 1964 from Cornell. From 1963 to 1992 he was Supervisor for Vertebrates at the Smithsonian Oceanographic Sorting Center and was its director from 1982 to 1988. In 1992 he joined the staff of the National Museum of Natural History, Smithsonian, as a marine biologist, and became a research associate in 1998. Les played an active role in the American Society of Ichthyologists and Herpetologists (Publications Secretary, 1968–1973; Board of Governors, 1973–1978); the Biological Society of Washington (Proceedings Vertebrate Editor, 1976–1978; Treasurer 1981–1986; President, 1990–1991); and the American Association of Zoological Nomenclature (President, 1996–1997). His field experience included collecting freshwater fishes in eastern US and marine fishes in the Red Sea, various Western Indian Ocean locales, coasts of Colombia, Peru and Chile and in the Philippines. Les published extensively on fishes of the families Percidae, Bembridae and Platycephalidae before he passed away in 2017.



# JAMES DS KNUCKEY

James DS Knuckey earned his BS in Marine Biology at San José State University, California and is working on his MSc at the Pacific Shark Research Center at the Moss Landing Marine Laboratories under the supervision of Dr David Ebert. During his undergraduate studies James, in collaboration with Dr Ebert, described a new species of lanternshark (*Etmopterus*) from the western North Pacific. His MSc thesis research project is on the phylogenetics of eastern North Pacific skates (Arhynchobatidae: *Bathyraja*), incorporating both traditional morphological and meristic methods, and newer innovative molecular techniques. James has also studied fish assemblages in San Francisco Bay, and researched the gill morphology of mosquitofish (*Gambusia affinis*). His main research interests are taxonomy, phylogenetics, parasitology of fish, and deep-sea assemblages.



#### **HELEN K LARSON**

Helen K Larson is Emeritus Curator of Fishes, Museum and Art Gallery of the Northern Territory, Darwin. She is also a research associate, Museum of Tropical Queensland, Townsville, and Adjunct Associate Professor, School of Marine and Tropical Biology, James Cook University, Townsville. Her research interests are the taxonomy, systematics, ecology and behaviour of Indo-Pacific mangrove and coral reef fishes, especially gobioid fishes (gobies, gudgeons, mudskippers). Much of her work is on the taxonomy of estuarine and coral reef gobiid fishes, preferably by way of generic revisions. The goby subfamily Gobionellinae is of particular interest to her. She has been actively publishing since 1975. Helen retired from the Museum and Art Gallery of the Northern Territory in late 2009 and moved to north Queensland where she works on gobies in association with the Museum of Tropical Queensland. Her expertise as a taxonomist/ systematist is regularly sought from around the world, whether as a reviewer of manuscripts or identifier of unknown fish species, and she has the honour of being the recipient of 13 eponyms to date. She is on the editorial boards of several journals and is an honorary foreign member of the American Society of Ichthyologists and Herpetologists.



# PETER R LAST

Peter R Last is the former director of the Australian National Fish Collection, Hobart, Australia, and now a retired honorary research fellow. Over the past 40 years he has specialised in the taxonomy and biogeography of Indo-Pacific fishes, with particular emphasis on shark and ray faunas across the region. He has more than 300 publications, including technical works, books and book chapters, aimed at both scientists and the general public.



## **JEFFREY M LEIS**

Jeffrey M Leis is a senior (retired) fellow in Ichthyology at the Australian Museum in Sydney, and an adjunct professor at the University of Tasmania. He has studied fish larvae in the warmer waters of the Indo-Pacific since the late 1970s. Jeff did his BSc in Zoology at the University of Arizona and began to study the porcupine fishes (Diodontidae) whilst a graduate student. He subsequently received his PhD in Biological Oceanography at the University of Hawaii. After 18 months in southern California, studying the impact of nuclear power stations on fish larvae, he moved to Australia. Jeff's research on fish larvae has provided a taxonomic base for their study in the Indo-Pacific. He has used them to reveal the relationships of fishes, and has shown their behaviour can strongly influence dispersal in the ocean. In 2014, he retired from the museum and took up an adjunct professorship with the University of Tasmania's Institute for Marine and Antarctic Studies, where he continues his ichthyological studies.



# JOHANN RE LUTJEHARMS

Johann RE Lutjeharms had an intense interest in the biogeography and life history of fishes and their relationship to the physical/ chemical marine environment. This outlook had its origin in the multi-disciplinary PhD education he received at the University of Washington after gaining his MSc degree at the University of Cape Town in 1971. His oceanographic interest extended geographically from the Southern Ocean near the continent of Antarctica to the tropical seas of the Indian and Atlantic oceans. His research activities in these regions resulted in approximately 500 publications, including the internationally acclaimed book, *The Agulhas Current*. He collaborated closely with colleagues from many countries, including the Netherlands, Germany, France, Italy, Russia, Brazil, Israel, Mozambique and Kenya. For his academic work he received numerous awards, amongst others, the Humboldt-Forschungspreis from Germany, the Fridtjof Nansen Medal of the European Geosciences Union, the Havengaprys for physical sciences of the Suid-Afrikaanse Akademie vir Wetenskap en Kuns and the John F. W. Herschel Medal of the Royal Society of South Africa. He received honorary doctorates from Rhodes University, the University of Pretoria and the University of Johannesburg and was a recipient of the Order of Mapungubwe, the highest civil honour the South African State bestows. Johann passed away in 2011.



## **B MABEL MANJAJI-MATSUMUTO**

B Mabel Manjaji-Matsumoto is a senior lecturer and head of the Endangered Marine Species Research Unit, Borneo Marine Research Institute, Universiti Malaysia Sabah. She holds a PhD in Zoology (University of Tasmania), and BSc (Hons) in Marine Science (Universiti Kebangsaan Malaysia), and has 20 years of work experience in the field of elasmobranch (sharks and rays) biodiversity. Her range of research interests includes ichthyology (fish studies), marine conservation, and sustainable fisheries.



# ANDREA D MARSHALL

Andrea D Marshall has been working in Mozambique since 2003. Educated in the United States and Australia, Andrea was the first person in the world to complete a PhD on manta rays. After completing her thesis, Andrea stayed on to spearhead the conservation efforts of manta rays in Mozambique. Her research examines aspects of the biology, reproductive ecology, habitat use, migrations and social behaviour of both species of *Manta*.



# **KEIICHI MATSUURA**

Keiichi Matsuura is Curator Emeritus of Department of Zoology at the National Museum of Nature and Science, a major natural history museum in Japan, where he has studied fishes since 1979. His research specialty is tetraodontiform fishes, particularly their taxonomy and phylogeny. His field work has taken him to many parts of Japan and its adjacent areas, and also to tropical regions in the Indo-Pacific. He served as the president of the Ichthyological Society of Japan and as a vice-chair of Global Biodiversity Information Facility (GBIF). He has published many papers and books on fishes, including field guides for tropical marine fishes in Southeast Asia.



# JOHN E MCCOSKER

John E McCosker is Senior Scientist and the first Chair of Aquatic Research at the California Academy of Sciences. From 1973 to 1994, he was Director of the Steinhart Aquarium in San Francisco. He received his PhD in Marine Biology from the Scripps Institution of Oceanography. His research activities around the world include field and laboratory studies ranging from microscopic bioluminescent bacteria to macroscopic man-eating elasmobranchs. He has also studied the taxonomy of snake eels and moray eels, the marine life of the Galápagos, and the biology of salmonid fishes. In addition he has researched dispersed and renewable energy sources as alternatives to national vulnerability and war, and contributed to the public understanding of sustainable seafoods. He is the author of more than 260 popular and scientific articles and books. His research on white shark behaviour and the deep-sea life of the Galápagos have been featured in numerous television documentaries.



## MICHAEL MAIA MINCARONE

Michael (Mike) Maia Mincarone is Professor of Zoology at Universidade Federal do Rio de Janeiro, and curator of the NUPEM Fish Collection. He obtained his Bachelor degree in Oceanography from the Universidade do Vale do Itajaí (Itajaí, SC) and his PhD in Zoology from the Pontifícia Universidade Católica do Rio Grande do Sul (Porto Alegre, RS). Part of his PhD was conducted at the National Museum of Natural History, Smithsonian Institution (Washington, DC), and California Academy of Sciences (San Francisco, CA). At the beginning of his career Mike focused on the systematics of hagfishes (Myxiniformes), but his current research includes the systematics and conservation of many other deep-sea fishes.



# RANDALL D MOOI

Randall D Mooi received his PhD in Zoology from the University of Toronto (Ontario, Canada) in 1991, working on the systematics and biogeography of percoids, particularly the Plesiopidae and putative relatives. Following a post-doctoral fellowship in the Division of Fishes of the Smithsonian Institution investigating systematics of deep-sea fishes (Chiasmodontidae, Champsodontidae), he was Curator of Fishes and Section Head of Vertebrate Zoology at the Milwaukee Public Museum in Wisconsin (USA) for 12 years. While there he was involved in several collaborative expeditions to Indo-Pacific coral reefs, surveying fishes in Thailand, French Polynesia, the Philippines, Solomon Is., Vanuatu and Australia. In 2004 he became Curator of Zoology at the Manitoba Museum (Winnipeg, Manitoba). This necessitated a broadening of research interests and field work to include freshwater fishes, birds, snakes and amphibians. He is an adjunct professor in the Department of Biological Sciences at the University of Manitoba and also serves on the Steering and Technical Committees of the Manitoba Breeding Bird Atlas. He continues to examine systematics of percomorph taxa, particularly gobioids, and ponders systematic methods.



# KATE A MOOTS

Kate A Moots completed a BSc in Zoology at the University of Alberta, followed by an MSc at the University of British Columbia in Vancouver. She completed her MPhil and PhD at the University of Kansas, working with Dr Edward O Wiley and Dr Frank B Cross, on the phylogenetic relationships of ~80 species of North American freshwater darters (Percidae). Both during and after completion of her PhD, Kate worked as Collections Manager for the University of Kansas Division of Ichthyology, where she was responsible for ~500 000 specimens. In 2000, Kate married Jeffrey Moots and moved to Saipan, where she worked as a fisheries biologist, an environmental consultant, and taught Developmental Mathematics and Marine Biology at Northern Marianas College. In 2004, she was awarded an NRF post-doctoral fellowship at the South African Institute for Aquatic Biodiversity, where she worked on the families Solenostomidae and Syngnathidae, and participated in field research in the Seychelles. This work served as the nucleus for her work on the chapters in this book and helped to develop her current research programme. Kate is now an associate professor at the University of Guam, where she teaches a variety of undergraduate biology lectures and labs, including Human Anatomy and Physiology, Comparative Vertebrate Anatomy, and Principles of Biology. During the summer, she travels, works on research, and scuba dives as much as possible.



# HIROYUKI MOTOMURA

Hiroyuki Motomura completed his PhD in 2001 at Miyazaki University, Japan. For his thesis, Dr Motomura revised the family Polynemidae (threadfins) and published part of the thesis as an issue of FAO Species Catalogue series (*Threadfins of the World*). He worked at the National Museum of Nature and Science, Tokyo, in 2002, and at the Australian Museum, Sydney, and CSIRO Marine & Atmospheric Research, Hobart, between 2003 and 2005. Hiroyuki has been employed at the Kagoshima University Museum, Japan, since October 2005, and has established a fish collection of 100 000 specimens collected mainly from southern Japan. He has published more than 350 papers and 40 books, and is currently working on a revision of the family Scorpaenidae in the Indo-Pacific. He has been a Professor of Ichthyology since 2010 and currently is the director of the Kagoshima University Museum.



# THOMAS A MUNROE

Thomas (Tom) A Munroe received his BS and MS degrees in Marine Biology from Southeastern Massachusetts University located in North Dartmouth, Massachusetts. He began his career at the Systematics Laboratory in 1987 after finishing a PhD degree at the Virginia Institute of Marine Science, College of William and Mary, and a postdoctoral fellowship in the Division of Fishes, Museum of Natural History, Smithsonian Institution where he is now employed as an ichthyologist at the National Systematics Laboratory (NOAA's National Marine Fisheries Service). Tom specialises in the systematics and taxonomy of the Pleuronectiformes, the flatfishes, with a special interest in the families Cynoglossidae (tonguefishes) and Soleidae (soles). He has conducted research on these and other flatfishes at sites located in different regions of the world. Past work has included systematic revisions of tonguefishes from both the eastern and western Atlantic Ocean and he has also described several species of tonguefishes from the eastern Pacific and Indo-Pacific oceans. Tom has contributed species assessments for tonguefishes and other flatfishes for several FAO regional species identification guides. His present research interests focus on continued efforts to discover and describe the biodiversity of flatfish species in the cynoglossid genera *Symphurus, Cynoglossus,* and *Paraplagusia*, as well as the soleid genus *Soleichthys*. Other research involves systematic studies of flatfishes from New Zealand, Australia, the eastern Pacific and Indo-Pacific regions, and the Americas.



## ΤΕΤΣΟΙΙ ΝΑΚΑΒΟ

Nakabo has published on fish systematics and taxonomy.

# JOSEPH S NELSON

Joseph S Nelson obtained his PhD in 1965 from the University of British Columbia under Dr CC Lindsey and then spent three years at Indiana University. A long career at the University of Alberta followed from 1968, where he taught courses and supervised many graduate students before his retirement in 2002. Joe passed away in 2011. He wrote research papers on a wide variety of topics, such as hybridisation, gasterosteids, *Antipodocottus*, psychrolutids, creediids, percophids, and trichonotids. He is probably best known as author of *Fishes of the World* (4th edition, 2006) and co-author of the *Fishes of Alberta* (2nd edition, 1992). He served as Chair of the American Fisheries Society/ American Society of Ichthyologists and Herpetologists, Committee on Names of Fishes (1991–2010), resulting in the 2004 book *Common and scientific names of fishes from the United States, Canada, and Mexico*, 6th edition, American Fisheries Society, Special Publication 29. He received many awards such as: the Robert H. Gibbs, Jr. Memorial Award, 2002; the Robert K. Johnson Award for Excellence in Service, 2010, American Society of Ichthyologists and Herpetologists; the J. Dewey Soper Award, 2003, Alberta Society of Professional Biologists; the Artedi Lecturer Diploma, 2008, Royal Swedish Academy of Sciences; and the Fry Medal, 2010, Canadian Society of Zoologists.

Tetsuji Nakabo is co-author of the account of the Sphyraenidae. He is a Japanese ichthyologist who was based at the Kyoto University Museum, and a professor at Kyoto University, Kyoto, Japan, until his retirement in 2015.



# JØRGEN G NIELSEN

Jørgen G Nielsen, Curator Emeritus of Fishes at the Zoological Museum, Natural History Museum of Denmark, worked at the museum as a curator from 1959 until he retired in 2002. In his first years, he continued his MSc interest in flatfishes, but soon changed to the study of deep-sea fishes, especially those belonging to the very diverse order, the Ophidiiformes. During his retirement he has continued revising ophidiiform genera, often in collaboration with his successor, Peter Rask Møller. He has participated in several deep-sea cruises and published about 160 scientific papers.



# LYNNE R PARENTI

Lynne R Parenti has been a curator of fishes and research scientist at the Smithsonian's National Museum of Natural History since 1990. She received a BS in 1975 from the State University of New York at Stony Brook and was awarded a PhD in 1980 from the joint graduate training programme in systematic biology between the American Museum of Natural History and the City University of New York. Her research focuses on the comparative anatomy, phylogenetic systematics and biogeography of freshwater and marine fishes, in particular atherinomorphs and gobioids; higher-level phylogenetic relationships of osteichthyans; historical biogeography; and the use of new or under-studied morphological characters in systematic ichthyology, such as those of reproductive morphology. Lynne is a co-editor of Interrelationships of Fishes and Ecology of the Marine Fishes of Cuba, which received an annual award from the Academia de Ciencias, Cuba, in 2002, for an outstanding scientific publication. She is also co-author of Cladistic Biogeography and Comparative Biogeography, which received the Smithsonian Secretary's Award for an outstanding research publication in 2010. Among her other honours, Lynne was elected President of the American Society of Ichthyologists and Herpetologists (ASIH), the first female ichthyologist to hold that office, and received the Robert H. Gibbs, Jr. Memorial Award for Excellence in Systematic Ichthyology for an outstanding body of published work in systematic ichthyology from ASIH in 2013. She is an elected Fellow of the American Association for the Advancement of Science and was an Artedi Lecturer, 2005, at the Royal Swedish Academy of Sciences. As an adjunct professor at the George Washington University, she directs the research of and advises graduate students in systematic ichthyology. Fieldwork has taken her around the globe to areas including New Guinea, Sulawesi, Borneo, the Malay Peninsula, Tasmania, China, Taiwan, Singapore, Surinam, Cuba and Hawaii.



# THEODORE W PIETSCH

Theodore (Ted) W Pietsch is Professor Emeritus at the School of Aquatic and Fishery Sciences, and Curator Emeritus of Fishes at the Burke Museum of Natural History and Culture, University of Washington, Seattle, where he has spent most of his career as a professor mentoring graduate students and teaching ichthyology to undergraduates. He is a fellow of the California Academy of Sciences, the Gilbert Ichthyological Society, the Linnean Society of London, the American Association for the Advancement of Science, the University of Washington Teaching Academy, an honorary member of the Ichthyological Society of Japan and of the Russian Hydrobiological Society, a member of the Washington State Academy of Sciences, and Correspondant du Museum, Museum national d'Histoire Naturelle, Paris. He is interested primarily in marine ichthyology, focusing on the relationships, evolutionary history, functional morphology, behaviour, and reproductive biology of teleosts, particularly deep-sea taxa. As former curator of the fish collection of the Burke Museum of Natural History and Culture, he is also interested in natural history collections and collection building, and in biotic survey and inventory, the latter best exemplified by a decade-long series of expeditions to collect plants and animals on the islands of the Kuril Archipelago in the Russian Far East. He has published extensively on the history of science, especially the history of ichthyology.



# STUART G POSS

Stuart G Poss received a BA in Zoology at UCLA (*cum laude*), a MS in Marine Biology at San Francisco State University, and a PhD in Evolutionary Biology at the University of Michigan. He was a post-doctoral fellow at the Academy of Natural Sciences, Philadelphia, worked as Collections Manager at the California Academy of Sciences, and was Senior Ichthyologist and Curator at the Gulf Coast Research Laboratory before retirement. He is currently a research associate at the California Academy of Sciences. He is interested in the biology of scorpaenoid fishes, particularly their taxonomy and systematics, with special emphasis on the study of morphometrics and variation in body morphology and its implications for other facets of their zoology. His work makes use of multivariate analyses to identify and characterise species and to study relative rates at which various morphological features change in both ontogeny and phylogeny. In addition, he is interested in the application of single-plane structured light range-sensing to morphometric studies and its potential for automated identification and feature extraction. He has also published on lancelets and the biology of endangered and non-indigenous species in the Gulf of Mexico.



## **HELEN A RANDALL**

Helen A Randall obtained her BA in Biology at Boston University (*cum laude*) in 1949, followed by an MA in 1950. She was a teaching assistant in zoology at the University of Hawaii from 1950 to 1953, and married Jack Randall in 1952. In 1955, with a fellowship from Yale University and the Bishop Museum, Jack, Helen, their two-and-a-halfyear-old daughter, Loreen, plus one crew member sailed their ketch to Tahiti to study the food habits of groupers and snappers being considered for introduction to Hawaiian waters. They spent an idyllic year doing their research in Moorea. Then followed almost ten years in the Caribbean, before returning to Hawaii, where, over the years, Helen was involved in a variety of projects. Since 1975, she has been a research assistant at the Bishop Museum in Honolulu, working with Jack on the systematics of Indo-Pacific fishes and as a reviewer of scientific manuscripts. She has also been Managing Editor for *Indo-Pacific Fishes* since 1982. Helen has published 11 papers.



## JOHN E RANDALL

John (Jack) E Randall began his ichthyological studies at UCLA in 1948 and obtained a BA in Zoology with honours. This was followed by employment as a teaching assistant and other work to pay off his debts before sailing his ketch, the *Nani*, to Hawaii in 1950 where he took up a post as a graduate assistant in zoology. His long career has included innumerable field trips, with thousands of hours underwater and five years as Professor of Zoology at the University of Puerto Rico (1961–1965). He is widely regarded as the undisputed master of Indo-Pacific fishes, and has more than 550 papers to his name. He wrote several regional taxonomic guide books, authored more than 500 new species of fishes, and had 39 species named after him. He was the first to explain the origin of ciguatera poisoning, and discovered that the broad band of bare sand around reefs, now known as the Randall Zone, was the result of overgrazing by herbivorous fishes. He developed the technique of taking photographs of fishes posed in specially designed photo tanks. In his nineties, Jack still continued to write and describe species. He died in April 2020 at the age of 95.



## WILLIAM J RICHARDS

William (Bill) J Richards received his BA degree from Wesleyan University in 1958, his MS from Syracuse University and SUNY College of Forestry in 1960, and his PhD from Cornell University in 1963. He has been employed by NOAA Fisheries (and predecessor agencies) in Washington, DC and Miami throughout his career. Bill has received numerous awards from the United States Department of Commerce, including its Silver Medal in 1985, Bronze Medal in 2007, and NOAA's Distinguished Career Award in 2003. The American Fisheries Society honoured him with the O.E. Sette Award in 2002 and the Elbert H. Ahlstrom Lifetime Achievement Award in 2009. In 1992 he was chosen as a Scholarin-Residence, Bellagio, Italy by the Rockefeller Foundation. He is a fellow of the American Institute of Fishery Research Biologists and a life member of the American Society of Ichthyologists and Herpetologists. He is also a member of Sigma Xi. He has presented aspects of his research at many scientific conferences in the US, and in Scotland, Japan, Guam, Italy, Spain, Canada, Mexico, Poland, France, Australia, South Africa, and Norway, thus earning wide international peer recognition. He served as editor of the Fishery Bulletin for three years, the Bulletin of Marine Science, and Studies in Tropical Oceanography for 23 years, as well as serving on the editorial boards of Copeia, Fishery Bulletin, and Fishes of the Western North Atlantic. In academia he served as a professor on the adjunct faculty of the University of Miami, has taught courses on the early life history of fish, and directed student research as chairman for seven PhD students and five MS students, plus served on committees of seven PhD and eleven MS students. His research interests have been broad, with over 300 published articles and three books on various aspects of ichthyology and marine ecology. including the ecology of early life history stages of fish, shrimp, and lobsters; the distribution and abundance of scombroid larvae; the relation of climate and environment to fisheries; and systematic studies of the trigloid fishes. He has participated in many research expeditions in the eastern Atlantic, Caribbean Sea, and Gulf of Mexico.



# **BERNHARD RIEGL**

Bernhard Riegl is a professor of marine science and the head of the Department of Marine and Environmental Sciences at Nova Southeastern University in Fort Lauderdale, Florida. He is also a director of the National Coral Reef Institute at his university. His PhD is from the University of Cape Town. Since then, he has worked on coral reefs, first in the Western Indian Ocean and Red Sea, and later, all around the world. Other lines of research are in carbonate sedimentology and acoustic mapping of marine and lagoonal systems. Work has progressed from empirical to increasingly quantitative/theoretical, with a heavy modelling component. He has been actively engaged in the conservation of coral reefs, having served with or advised the marine conservation services in several African, Middle Eastern and South American countries. He is also actively involved in conservation initiatives in his home-base, Florida.



# **TYSON R ROBERTS**

Tyson R Roberts is a research associate at the Smithsonian Tropical Research Institute, Panama, and advisor to the Institute of Molecular Biosciences, Mahidol University, Nakorn Pathom, Thailand. He studied at Stanford University where his greatest influences were ichthyologists George S Myers and Robert R Harry (name later changed to Robert Rofen). From there he travelled to Mexico, and to the Central Pacific on the Tethys Expedition of Scripps Institute of Oceanography in the summer of 1960. His undergraduate and graduate years at Stanford were interrupted by three years in tropical Africa: two years teaching at Achimota School in Ghana, during which time he collected fishes throughout the country, and he spent a year collecting fishes in the Congo including the rapids of the Lower Congo and the rain forest of the Cuvette Centrale. His preference for fieldwork led him to spend many years in the tropics with a particular focus on freshwater fish fauna. He has travelled to the Congo Basin and Lake Tanganyika, Cameroon, Thailand, Myanmar, India, Laos, Cambodia, Indonesia, Papua New Guinea, and most countries in South America.



# BARRY C RUSSELL

Barry C Russell is Curator Emeritus of Fishes at the Museum and Art Gallery of the Northern Territory, and is a research fellow at Charles Darwin University, Darwin, Australia. He also co-chairs the IUCN Snapper, Seabream and Grunt Specialist Group. He has more than 45 years' research experience on the systematics, ecology and behaviour of tropical demersal fishes of the Indo-Pacific, has published more than 150 papers on the taxonomy and ecology of fishes, and has contributed 270 IUCN Red List assessments of fishes. His current research interests include the taxonomy and phylogenetics of lizardfishes (Synodontidae) and threadfin breams (Nemipteridae). He is also interested in the history of ichthyology, and is presently researching the ichthyological contributions of Lt James Barker Emery (HMS *Beagle*), and the French explorer, diplomat and naturalist, FL de Castelnau.



## **HUGO RS SANTOS**

Hugo RS Santos is a biologist and curator of the Ichthyological Collections of the Zoology Department of the Universidade do Estado do Rio de Janeiro. He earned his PhD from the Museu Nacional in Rio de Janeiro, Brazil, in 2006. Hugo has been researching stingrays since 2004, and his PhD thesis includes taxonomic revision and morphology of western Atlantic dasyatid stingrays. He has described two new species of stingrays, and has published other papers, mainly on stingray taxonomy. He has also co-authored a popular identification guide to sharks and rays from Rio de Janeiro state, Brazil.



## KUNIO SASAKI

Kunio Sasaki received his PhD degree in 1989 from Hokkaido University in fishery sciences (systematic ichthyology). In the same year, he joined the Department of Biology, Kochi University, as an assistant professor in marine biology. Since April 2006 he has been a professor at the same university, responsible for classes in marine biology, vertebrate zoology, and systematic ichthyology. His areas of research interest are taxonomy (mostly of croakers or drums) and the anatomy of fishes, the latter including bones, muscles and nerves. Recent publications are mainly focused on neuroanatomical aspects of various fish groups such as cyprinids, gobies, and puffers (co-authored with his MS and PhD students), in an attempt to use the structure of the nervous system as a source of phylogenetic characters. He has served as an editor of *lchthyological Research* (published by the lchthyological Society of Japan) since 1996.



## **BERNARD SÉRET**

Bernard Séret is an ichthyologist specialising in the study of the chondrichthyan fishes (sharks, rays and chimaeras). He worked at the Institut de recherche pour le développement (IRD) for almost 40 years, and has been hosted by the Muséum national d'Histoire naturelle, Paris, for about 20 years. Retired since 2014, he continues his scientific work and consultancies. His research work concerns the biodiversity, fisheries and conservation of the chondrichthyan fishes. He has undertaken several scientific missions and participated in many exploratory cruises in the Atlantic, Southern Ocean and South Pacific. He is author/co-author of about 180 papers, with more than 120 on Chondrichthyes, including the description of several new species of sharks, rays and chimaeras; chapters on batoids in the FAO catalogue for ECA; and contributions to the Rays of the World book (Last et al. 2016). His interest in shark fisheries includes analysis of fishery data of French shark fisheries in European waters (he was the French representative at the ICES-WGEF); analysis of shark by-catches of French tuna fisheries (in collaboration with IRD fishery biologists); involvement in the programme MADE (measures to mitigate adverse impacts of fisheries targeting large pelagic fishes in the open ocean); and training fishery observers and officers to identify sharks and rays. His interest in shark conservation includes assessments made for both French and European bodies, and international organisations. He provided scientific advice on proposals for listing shark and ray species (CITES, IUCN Red List, OSPAR, etc.) and has been involved in the elaboration of several plans of action for the management and conservation of shark populations. He continues to be involved in the analysis of shark attack cases and provides advice on this matter. Bernard Séret is the former scientific head of the European Elasmobranch Association (EEA), and a member of the IUCN-SSG



# ILIA B SHAKHOVSKOY

Ilia B Shakhovskoy studied at Moscow State Institute of Food Industry from 1993 to 1998, majoring in Aquatic Bioresources and Aquaculture. He completed his postgraduate studies at the PP Shirshov Institute of Oceanology of Russian Academy of Sciences (IORAS) in 2001, under the supervision of Professor NV Parin. From 2001 to 2007 he was an engineer at the Laboratory of Ocean Ichthyofauna (IORAS). In 2011 he became a junior scientist, and in 2011 was appointed a scientific associate at IORAS. Ilia's primary interest is in systematics, distribution, biology, ecology and biogeography of flying fishes, and he worked on these fishes for 20 years before receiving his PhD in 2016. He has also worked on keeping and breeding hydrobionts in a closed system and on the marine biology of the White Sea. He has published 12 peer-reviewed papers.



# **CHARLES RC SHEPPARD**

Charles RC Sheppard is Professor Emeritus at the University of Warwick, UK. His research has been focused on marine habitats in the tropics, especially coral reefs and the effects of humans upon them. He has been longstanding editor of the journal *Marine Pollution Bulletin*, and has written numerous papers and articles, including about 12 books on marine science during this time. He is on advisory panels for EU, Arabian and UK grant bodies (marine science); served on the advisory panel for the establishment of the first University of the Republic of Seychelles; became a review member of the Intergovernmental Panel for Climate Change from 2004; and has lately been the advisor on environmental matters to the Commissioner of UK Overseas Territories. In 2014 he was awarded the OBE for his services as advisor.



# DAVID G SMITH

David G Smith received his undergraduate education at Cornell University where he was introduced to fishes by the noted ichthyologist, Edward C Raney. He subsequently attended the University of Miami, Florida, completing his MS and PhD degrees under the direction of C Richard Robins. He worked at the University of Texas Medical Branch at Galveston, and later, at the Museum of Comparative Zoology at Harvard University, before arriving at the Smithsonian Institution in 1989. David has specialised in the study of eels (Anguilliformes) and is considered one of the foremost authorities on the group. He officially retired in 2012, but continues his research. He is also interested in the history of ichthyology and has written several papers on this subject.



# W LEO SMITH

W Leo Smith works at the Kansas University Biodiversity Institute and Natural History Museum where he is an associate professor and associate curator. His research focus is the comparative evolutionary biology of fishes and he contributed the section on the classification of serranoid fishes.

## SHIRLEEN SMITH

Shirleen Smith is a museum specialist in the Division of Fishes, National Museum of Natural History, Smithsonian Institution, in Washington, DC. She has been with the museum since 1996.



#### WILLIAM F SMITH-VANIZ

William F Smith-Vaniz received his master's degree in Fisheries Biology from Auburn University and his PhD in Marine Sciences from the University of Miami. While in graduate school, he spent three months in Sri Lanka collecting specimens and data for a Smithsonian project on jacks and trevallies, which stimulated his interest in this family of approximately 150 species and led to his recognition as a global expert on the group. He has published descriptions of more than 80 new species in about a dozen families. He began his ichthyological career as a technician in the Smithsonian Fish Division before returning to graduate school; was a curator at the Academy of Natural Science of Philadelphia for 19 years; and then worked as a research scientist for the then-named Biological Resources Division of the US Geological Survey in Gainesville, Florida, before retiring in late 2006. He is a research associate of the Smithsonian Institution and the University of Florida, and also serves as consultant to the International Union for Conservation of Nature, participating in foreign workshops, and helping with IUCN Red List species assessments. He has authored two books, one on freshwater fishes of Alabama, the other on fishes of Bermuda, and over 100 scientific papers and book chapters. He was the recipient of the Robert H. Gibbs, Jr. Memorial Award presented by the American Society of Ichthyologists and Herpetologists (2011) for "an outstanding body of published work in systematic ichthyology."



## **MATEUS C SOARES**

Mateus C Soares completed his PhD at the Universidade de São Paulo in 2011, under the supervision of Marcelo de Carvalho. His thesis focused on the myology and phylogeny of Chondrichthyes, describing, comparing and raising new morphological characters to test the hypothesis of shark monophyly (i.e., without the rays). He published articles on mandibular and hyoid musculature of sharks and was co-author of an anatomical guide to musculature of sharks, rays and chimaeras. He is chiefly interested in the comparative anatomy and evolution of fishes, especially Chondrichthyes. Currently he is working as an environmental consultant and professor.



#### **JOHN S SPARKS**

John S Sparks, Curator-in-Charge and Professor, Department of Ichthyology, American Museum of Natural History, travels the world in search of bioluminescent and biofluorescent marine organisms. His research focuses on the systematics and biogeography of recent fishes, with a particular interest in the evolution and diversification of luminescent signalling systems and fluorescence in marine fishes. For these studies, John is heavily involved in the design and implementation of state-of-the-art underwater cameras and lighting systems. He has led numerous expeditions to Madagascar, where his work focuses on the diversity and biogeography of the island's endangered freshwater fishes, including cavefishes, and specialised hearing in cichlids. His recent fieldwork includes biotic surveys and inventories of freshwater, nearshore marine, and deep-sea fishes in Madagascar, the Indo-Pacific region, throughout Southeast Asia, South America, the western Atlantic, eastern Pacific, and the Caribbean. He is a professor in the Richard Gilder Graduate School at the Museum, an adjunct professor at CUNY and Columbia University, and a principal investigator with the museum's Sackler Institute for Comparative Genomics. He joined the museum in 2002 and served as lead curator of the museum's travelling exhibition Creatures of Light: Nature's Bioluminescence, and as co-lead curator of the new travelling exhibition Life at the Limits: Stories of Amazing Species. He also curated the temporary exhibit The Exosuit and produced two short films on marine biofluorescence, Reefs Illuminated and Capturing the Ocean's Glow. He received his PhD in Ecology and Evolutionary Biology from the University of Michigan in 2001.



# **VICTOR G SPRINGER**

Victor G Springer obtained a BA from Emory University in 1948; an MS (Marine Biology), University of Miami, Florida in 1954 (major professor, Luis R Rivas); and a PhD (Zoology) in 1957 with Clark Hubbs at the University of Texas. He served in the US Army Medical Corp from October 1950 to September 1952 (the second year in Korea). His professional career includes work as a freshwater fisheries biologist (1950) and then as marine ichthyologist for the State of Florida from 1957 to 1961 (primarily life-history, ecological studies of fishes). From 1961 to 1962 he was contracted by the Division of Fishes, US National Museum of Natural History (USNM), to do systematic studies on sharks, before being employed as Associate Curator, Curator and Senior Scientist in Systematics, Anatomy, and Biogeography of Fishes (1962–2005). From 2005 he has been Curator Emeritus at the USNM. He has participated in fish-collecting expeditions to Florida, Texas, northern Mexico, Bahamas, Dominica, US North Atlantic, Bermuda, Great Barrier Reef, Taiwan, Korea, Philippines, Indonesia, Israel, Eritrea, St Brandon Shoals, Agalega, Pohnpei, Fiji and Rotuma.



## **KENNETH A TIGHE**

Kenneth A Tighe graduated from Union College, Schenectady, New York, in 1970 after which he attended graduate school in the Department of Zoology, University of Rhode Island, completing his studies in 1975. Since 1982, he has worked at the National Museum of Natural History, Smithsonian Institution, where he has conducted research on the eel families Chlopsidae, Serrivomeridae and Synaphobranchidae.



## FRANZ UIBLEIN

Franz Uiblein is a principal scientist in fish taxonomy and marine biology at the Institute of Marine Research, Norway (IMR). He is also a research associate at the South African Institute of Biodiversity (SAIAB) and the Vietnam National Museum of Nature (VNMN). In addition he has served as Editor-in-Chief of the international journal *Marine Biology Research*. His main research interests are ecology, diversity and evolution of marine fishes, but he has also worked on freshwater fishes, ostracods and salamanders. Franz began his work on goatfish and deep-sea fish taxonomy in 1992 as a deputy head of the marine fish collection at the Senckenberg Museum, Frankfurt, Germany. Three years later he co-organised a deep-sea conference in Vienna, Austria, and subsequently participated in research cruises off the Canary Islands and to the Great Meteor Seamount (eastern Central Atlantic). His *in situ* studies of deep-sea fishes began in 2000 when he analysed video footage from submersible dives in the NE Atlantic, followed by dives with the *Johnson Sea Link* submersible in deep canyons of the Gulf of Maine (NW Atlantic). After writing a review article in 2007 emphasising the importance of goatfishes (family Mullidae) for coastal monitoring and management, he began working on taxonomic revisions of the two goatfish genera *Upeneus* and *Mulloidichthys*.



#### **RUDY VAN DER ELST**

Rudy van der Elst, a research associate at the Oceanographic Research Institute (ORI), Durban, was born in Utrecht, Holland, but completed schooling and studies in South Africa. He graduated from the University of South Africa (BSc) and the University of Natal with an MSc. Since 1969 he has been employed at the South African Association for Marine Biological Research (SAAMBR), serving as Head of Research at ORI (1992–2011) where he was responsible for research management, including the planning, funding and execution of marine research programmes. Rudy has been engaged in diverse marine resource-use and conservation studies, especially those relating to management problems in small-scale, recreational and artisanal fisheries. These have included some unpopular conservation plans, including the protection of the shad (Pomatomus saltatrix), reduced beach netting and the conservation of sharks. He has been involved in the development of long-term fisheries monitoring and assessment projects, including the National Marine Linefish catch and effort system and the nationwide ORI tagging project that both interact with the fishing public. He also played a role in the original development of marine protected areas in KwaZulu-Natal, including the application for World Heritage status of the iSimangaliso Wetland Park. He has played a role in new fisheries and coastal zone policies of South Africa as well as in diverse projects in the West Indian Ocean for a variety of international agencies, including the World Bank, UNDP, Norad, IUC, WWF and WIOMSA. Rudy has supervised postgraduate students, published a considerable number of scientific papers, and several popular books on the fishes of southern Africa.



#### **RICHARD P VARI**

Richard P Vari began his research career working on the Terapontidae, a widespread family in marine, estuarine and freshwaters of the Indo-Pacific region. His subsequent research involved western North Atlantic seahorses (Hippocampidae) and phylogenetic, revisionary and biogeographic studies of various families in the Characiformes of South America and Africa, Gymnotiformes in South America, and Siluriformes in South America, Africa and South Asia. Richard passed away in 2016.



## **BENJAMIN C VICTOR**

Benjamin C Victor is a research associate at the Guy Harvey Research Institute at Nova Southeastern University in Florida, USA, and founded the Ocean Science Foundation based in southern California, USA. Born and raised in Johannesburg, South Africa, Ben received a BA from Cornell University in New York, USA, and then moved to graduate school at the University of California, Santa Barbara (UCSB), where he received his PhD studying the population dynamics of bluehead wrasses in the Caribbean. He was part of the small group of pioneer researchers in the 1980s establishing larval recruitment as one of the most important factors controlling the populations of some coral reef fishes. As part of his studies, he was among the first researchers to use daily otolith increments to age reef fishes, and subsequently spent many years identifying larval reef fishes and exploring various aspects of the age and size of settling reef fish larvae. In addition, Ben surveyed fishes in the Galapagos Islands (where he discovered a new razorfish species, Xyrichtys victori), Baja California, Panama and Palau. After his graduate studies, Ben attended medical school at the University of California, Irvine (UCI), did an internship at Stanford University, and finished his residency in pathology (and an MBA) at UCI. He presently directs several medical laboratories in addition to continuing research on reef fishes. More recently, Ben has specialised in applying DNAbarcoding techniques to reef fish taxonomy, describing a number of new Caribbean gobies and blennioids. He has collaborated with taxonomists in resolving taxonomy and describing new species in the Indo-Pacific, mainly labrids, and more recently, the pempherids. He founded and edits the Journal of the Ocean Science Foundation as a vehicle for rapid, free, open-access publication of reef fish taxonomy.



#### ELAYAPERUMAL VIVEKANANDAN

Elayaperumal Vivekanandan (Vivek) acquired an MSc degree (Marine Biology, Annamalai University) in 1972 and his PhD degree (Fish Energetics, Madurai University) in 1976. He joined the Indian Agricultural Research Service as a scientist in the Central Marine Fisheries Research Institute (CMFRI) in 1976. Since then, he has served CMFRI as Senior Scientist and Principal Scientist, and in a variety of other capacities, including Head, Demersal Fisheries Division and Scientist-in-Charge, Madras Research Centre of CMFRI. He retired on superannuation as Principal Scientist and Scientist-in-Charge, Madras Research Centre of CMFRI in April, 2012, and rejoined the same centre as Emeritus Scientist and Consultant. Vivek's core areas of research are fish stock assessment, the valuation of coastal and marine biodiversity and ecosystems, coastal fisheries management, marine mammal conservation, and climate change. His contributions include fisheries management advisories, regional collaboration on marine fisheries in South and Southeast Asia with organisations such as FAO, Bay of Bengal Large Marine Ecosystem, Bay of Bengal Programme – Intergovernmental Organisation, and GiZ, Germany. On invitation, he served as Fisheries Team Leader in the UN Project "Oceanographic Survey in Support of Damage Assessment" at King Fahd University of Petroleum & Minerals, Dhahran, Saudi Arabia, for one year in 2003. FAO and BOBLME have recognised him as a Regional Trainer on Ecosystem Approach to Fisheries Management, and Science Communication. He has conducted a number of training programmes and trained about 200 young researchers on scientific paper writing and paper presentation in Phuket, Male, Colombo, Penang and Kochi. He was Chief Editor of the Journal of Marine Biological Association of India between 2008 and 2011. He has supervised PhD research programmes and five students acquired doctoral degrees from Madras University under his guidance. He has authored 157 publications, including 67 research papers in peer-reviewed journals, 15 books/chapters, and a number of national and international technical reports.



# JEFFREY T WILLIAMS

Jeffrey (Jeff) T Williams is an ichthyologist and until recently was Collections Manager of Fishes at the National Museum of Natural History (NMNH), Smithsonian Institution, where he managed the largest fish collection in the world, conducting research on the systematics, taxonomy and biogeography of marine fishes, and on the interrelationship between the tectonic history of the Indo-Pacific and the geographic distribution of fishes. He received his BS from the Florida State University, MS from the University of South Alabama and PhD from the University of Florida. He has collected and studied marine fishes around the world and is an authority on the cryptic marine fishes, particularly in the Indian and Pacific oceans. He has led and participated in fish-collecting expeditions to the Gulf of Mexico, western Atlantic Ocean, Caribbean Sea, Hawaiian Islands, Ryukyu Islands, Philippine Islands, Seychelles, Fiji Islands, Tonga Islands, New Caledonia, Loyalty Islands, Australia, Vanuatu, Solomon Islands, Wallis & Futuna, French Polynesia (Mururoa, Moorea, Rapa, Gambier Archipelago, Marquesas) and Palau. He was with the NMNH from 1983–2020. He continues to focus his research on discovering and describing tropical marine fishes from remote and poorly surveyed areas around the world.



# **RICHARD WINTERBOTTOM**

Richard (Rick) Winterbottom graduated with a BSc (Hons) from the University of Cape Town in 1967, and obtained a PhD from Queen's University, Ontario, in 1971. Post-doctorates at the Smithsonian Institution and the National Museum of Canada were followed by a three-year stint as Senior Lecturer at the JLB Smith Institute of Ichthyology at Rhodes University (now the South African Institute for Aquatic Biodiversity). He was appointed Assistant Curator of Fishes at the Royal Ontario Museum (ROM), Toronto, in 1978, as Associate Curator in 1979, and as Senior Curator in 1984. He was cross-appointed as Assistant Professor at the University of Toronto in 1980, Associate Professor in 1986, and Full Professor in 1990. He retired from the ROM in 2010, and was appointed Curator Emeritus. Rick's main research interests have involved the higher relationships of teleosts, which he investigated through studies of osteology and myology, and his 1974 paper on tetraodontiform comparative myology was one of the first to use Hennigian phylogenetic methodology for a group of fishes. His 1974 synonymy of teleost muscles is still frequently cited today. He has also been a proponent of historical biogeography, especially for the complex area known as the Coral Triangle. He has actively collected marine and mangrove fishes throughout the Indo-Pacific, and has been on or instigated about 20 major synoptic field trips. Thirteen species of fishes (mainly gobies) have been named in his honour. Current research interests devolve mostly on the systematics and biogeography of the Indo-Pacific goby genus *Trimma*, of which he has described or co-described 72 of the 106 currently recognised valid species.



#### **MEGAN V WINTON**

Megan V Winton grew up near the coast of Florida, USA, where she developed an early love for the ocean and a particular fascination with sharks. She has spent the past ten years researching elasmobranch populations on both coasts of the United States. Since 2019, Megan has been a research scientist at the Atlantic White Shark Conservancy, where she is working to improve scientific understanding of a new white shark aggregation site off Cape Cod, Massachusetts, USA. She is currently a PhD candidate at the University of Massachusetts Dartmouth's School for Marine Science and Technology and previously received her MSc in Marine Science from Moss Landing Marine Laboratories and her BSc in Biology from Emory University. Her research focuses on various aspects of fisheries biology and ecology, as well as the development of quantitative frameworks for interpreting and integrating tagging data into population assessments. Through her work on white sharks, Megan has become increasingly interested in the human dimensions of wildlife management and decision making.



# DAVID J WOODLAND

David J Woodland completed his BSc Honours degree at the University of Queensland, studying the fishery potential of the Heron Island inner reef flat, operating a 100-metre arrowhead trap. With Dick Slack-Smith, who was running poison stations on the outer reef, they collected over 400 species and published, in effect, the first checklist of Great Barrier Reef fishes in 1963. Following three years with Victoria Fisheries working with trout, David moved to the University of New England (UNE) where he completed his PhD with a study on the energy budget of a population of freshwater crayfish. After being appointed as a tenured lecturer in population ecology at UNE, sabbatical leaves afforded him an opportunity to work overseas. Encouraged by Jack Randall, he began a revision of the Siganidae, the rabbitfishes, collecting in locations as varied as the Seychelles, Andaman Islands, Tonga and Palau. FAO asked him to add the Gerreidae and Leiognathidae to his contributions to their species identification publications, which led to revisionary studies of these families as well. In addition to publications in the zoogeography of the Indo-Pacific region, his other research interests have ranged from insect physiology to evolution of alarm signals. As an Adjunct Research Fellow at UNE, he continues his studies on the Siganidae.



#### **LEANDRO YOKOTA**

Leandro Yokota is post-doctoral student at the Universidade de São Paulo, Brazil, where he is leading a worldwide taxonomic review of the family Gymnuridae (Myliobatiformes, Chondrichthyes). He earned his bachelor degree in biological sciences from Universidade Estadual Paulista in 2002. His master's degree in animal biology was obtained in 2005 at Universidade Federal de Pernambuco, and he completed his doctorate in zoology in 2010 at Universidade Estadual Paulista. His primary research interests focus on the biology and taxonomy of sharks and rays, but he is also interested in the ecology, conservation and fishery biology of the group. Leandro Yokota is a member of the Sociedade Brasileira para o Estudo dos Elasmobrânquios (SBEEL).



#### **UWE ZAJONZ**

Uwe Zajonz, an aquatic zoologist by qualification, is an ichthyologist, ecologist and conservationist specialising in tropical marine ecosystems, notably of the Indian Ocean and the Arabian region. As an ichthyologist he started his career at the Senckenberg Research Institute and Museum of Nature, Frankfurt am Main, Germany, studying deep-dwelling fishes of the Red Sea and Gulf of Aden. He worked up a number of important deep-sea fish collections, describing several new species, and planned to earn his PhD from these studies. However, his participation in a number of research expeditions excited his interest in reef fishes, particularly in the Socotra Archipelago. The scope of his studies has expanded to include fish faunistics, community ecology, biogeography, trophic ecology and fisheries. His work also covers little-studied adjacent coastal regions of the northern Indian Ocean. Uwe's varied career includes experience in marine resource use and coastal zone management, conservation at the interface to sustainable development, marine climate change research, and project management. Uwe has returned to the Senckenberg Research Institute to manage a large conservation project on Socotra for the United Nations Environment Programme. He has since also obtained his long overdue PhD in Biology/Zoology.
# **ABBREVIATIONS AND ACRONYMS**

aff. – abbreviation of the Latin affinis, meaning having a relationship with, but not identical to. BD/A – body depth at anus. BD/GO - body depth at gill opening. BL – body length. cf. - abbreviation of the Latin confer, conferatur (meaning to compare [with]). **CFSA** – Coastal Fishes of Southern Africa. DW – disc width. FL – fork length. **FSO** – Fishes of the Southern Ocean. **GR** – gill rakers. HL - head length. IO- infraorbital (bone). **IP** – initial phase. IUCN – International Union for the Conservation of Nature. LL – lateral line. LSS – lateral scale series. MVF – the number or mean number of predorsal, preanal and total vertebrae. **MVF H** – mean number of vertebrae when the first vertebra arises on the head (H). MYA – million years ago. NWIO - northwestern Indian Ocean. PALL – preanal lateral line. PCD – pharyngocutaneous duct. POM - preoperculomandibular. SFSA – The Sea Fishes of Southern Africa. SL - standard length. SO - supraorbital (bone). sp. (pl. spp.) - a single species; used when the genus is known, but not the species name. **SSF** – Smiths' Sea Fishes. ST - supratemporal (bone). SWIO – southwestern Indian Ocean. TL – total length. TP – terminal phase. TRB - transverse rows of scales backwards (see figure, Family Gobiidae, Volume 5). TRS (also TSR) - transverse rows of scales (or transverse scale rows). VF – vertebral formula.

WIO - Western Indian Ocean.

## GLOSSARY

For terms that are not included here, refer to the *Dictionary of Ichthyology* at http://www.briancoad.com/dictionary/complete%20 dictionary.htm

#### А

**abdominal** – on or pertaining to the belly or abdomen.

- acanthodian a class of early jawed vertebrates that resembled sharks in overall appearance, characterised by fin spines on their dorsal and paired fins (sometimes in between these), divided into three or four orders, and presently considered not to be monophyletic; acanthodians were entirely Palaeozoic ranging from the early Silurian to the Permian.
- acanthomorph / Acanthomorpha the formal name for the rayfinned fishes.

acinar glands – glands formed by a cluster of cells, looking a bit like a raspberry; found in Synanceiidae (stonefishes).

acoustico-lateralis system – the sensory system that includes the ears and lateral line.

acronurus – the specialised larva of a surgeonfish (Family Acanthuridae).

acuminate - tapering to a point.

- acuspidate without cusps.
- acute sharply pointed.
- adductor mandibulae main muscle that closes the jaw.
- **adelphophagy** the consumption of one embryo by another within the uterus.
- adenohypophysis the anterior of the pituitary gland.

adhesive filaments – filaments that attach the eggs to the substrate. adipose (tissue) – fatty.

- **adipose eyelid** more or less transparent tissue that partially covers the eye.
- adipose fin a small, fleshy fin behind the rayed dorsal fin, without bony elements.

adnate - joined from having grown together.

- adpressed flat against the body.
- advect to convey a fluid by horizontal mass movement.
- aeolian relating to the action of wind; wind-blown.
- **afferent circulation** taking blood to an organ.

air bladder - see swimbladder.

- **alar spines or thorns** very sharp spines near the lateral margins of the pectoral fins of male skates that serve to 'anchor' the male to the body of a female during copulation.
- **allele** a specific and unique form or variant of a particular gene or genetic marker; alleles typically refer to pairs of genes found on each of the two homologous chromosomes of a diploid organism.
- **allometric** differential changes with growth, e.g., eyes become larger relative to head length.

- **allopatric (distribution)** populations or species occupying mutually exclusive geographic areas.
- **allozyme** particular form of a given protein, coded by a particular allele at the gene responsible for encoding for that protein.
- **alveolar process of premaxilla** the thickened ridge on the premaxilla that contains the tooth sockets.
- **alveolar ramus** the tooth-bearing area of the lower jaw.
- **amphipods** a group of bilaterally compressed crustaceans.

**amphitemperate** – referring to a species that occurs in temperate water in both hemispheres, but is absent from the tropics.

- **ampullae of Lorenzini** jelly-filled tubes that open on a shark's skin and act as electroreceptors.
- **anadromous** fishes born in freshwater but spending most of their lives in the sea.
- **anal fin** the median, unpaired fin located on the ventral surface in front of the caudal fin.
- anal papilla see genital papilla.
- anastomosing joined together.
- **anchialine habitat** flooded inland marine caves with no direct connection to the sea.
- anterior on or towards the front (head) end of the fish.
- **anteromesial** toward the front and along the middle axis of the body.
- **anteroventral** relating to the underside of the front of the head; situated in front and below or toward the ventral side.

**antitropical (distribution)** – the distribution pattern where a group is found north and south of, but not in, the tropics (also anti-equatorial).

**antrorse** – pointing or curving anteriorly, specifically spines or fin elements.

anus - the exterior opening to the intestine; vent.

**apex predator (apex of pelagic food web)** – the predators that no other species tend to kill for food.

**aphakic space** – a space in the eye of fishes which is said to either enlarge the binocular field of vision or enhance the relative illumination of a portion of the retina of the fish.

**apomorph / apomorphy / apomorphic** – an evolutionarily derived or 'specialised' character or feature of an organism.

**aposematic** – denoting colouration or markings serving to warn or repel predators.

arcuate - curved like a bow.

**articular process** – the projection of a vertebra that articulates with the adjacent vertebra; also called a zygapophysis.

articulating - attached by means of a moveable joint.

- artisanal fishery a traditional fishery involving skilled but nonindustrialised fishers; typically a small-scale decentralised operation.
- ascending process a vertical extension of the anterior of the premaxilla.

**ascidians** – tunicate animals, including sea-squirts and red bait. **attenuate** – thin and stretched out.

autapomorphic characters – characters that are unique to a clade, and not shared with any other clade, such as the teeth fused into a beak and fused to the jaws.

autogenous - independently generated; without external influence.

- **autotomy or self-amputation** the ability of certain animals to deliberately cast off part of the body, most commonly known in lizards that 'lose' part of their tail when threatened.
- auxiliary scales small scales superimposed on the surface of larger scales.

**axil** – the inner or posterior part of the base of a fin.

axillary process (or scale) – enlarged, usually elongated scale or group of scales at the base of the pectoral or pelvic fins of some fishes.

## В

**bacterial luminescence** – the generation of light by the action of an enzyme on the light-producing molecule luciferin.

**barbel** – fleshy, tentacle-like process near the mouth of some fishes. **barotrauma** – gas expansion in the swimbladder leading to

haemorrhaging and death.

basal - at or towards the base.

**basibranchial** – median ventral cartilage or bony element in a branchial arch, behind the tongue.

**basidorsals** – two wedge-shaped, arched cartilages that form a continuous neural arch with the interdorsals to protect the spinal cord.

basipterygial process - processes on either side of the throat.

basipterygium – pelvic bone or cartilage to which the pelvic fin is attached.

bathyal – the benthic habitat from 200 m to ~1 000 m.

- **bathypelagic** the deep part of the oceanic zone, from 1 000 m to 4 000 m.
- **batoid** a flat elasmobranch fish, e.g., skate or ray, with the pectoral fins fused to the sides of the head and the gill openings on the ventral surface.

**batoids** – refers to the monophyletic group that contains all of the orders of rays.

bauplan – a body plan; a generalised set of morphological features shared by a group of organisms, especially at higher levels of classification, such as a phylum.

- benthic living on or close to the bottom.
- **benthopelagic** occurring near or just above the bottom of the sea; the depth zone about 100 m off the bottom at all depths below the edge of the continental shelf.

bicuspid - a tooth with two cusps or points.

bifurcate - split or divided into two parts; forked at tip.

bilobate - having two lobes.

**biogenic** – produced by living organisms.

**bioluminescence** – the production and emission of light by a living organism; the ability to produce light.

**bioluminescent oesophageal diverticula** – outgrowths of the oesophagus that contain bioluminescent bacteria.

**biserial** – arranged in two rows or series.

**body length** – the distance from the posterior opercular margin to caudal-fin base.

**bommie** – a large mound of corals some distance from the shore.

**bony contact organs** – in *Oryzias*, dermal bony outgrowths that project from a fin ray, surrounded by epidermis through which the bony outgrowths may protrude.

**brachyuran crabs** – typical crabs with the abdomen folded in under the thorax/carapace.

**brack / brackish** – with a salt concentration or salinity between fresh and sea water, i.e., 0.5–30 parts per thousand.

**branched ray** – a soft or segmented fin ray that is split into two or more parts distally.

branchial - pertaining to the gills.

branchiostegal membrane – the tissue supported by the branchiostegals and forming the lower wall of the gill cavity.

**branchiostegals** – bony 'rays' supporting the branchiostegal membrane on the underside of the head.

**buccal** – pertaining to the mouth cavity.

**buccopharyngeal pads** – pads at the back of the mouth and beginning of the gut.

**bucklers** – bony plates located at the base of certain fins or on the belly. **bulla (pl. bullae)** – a rounded structure.

**bycatch** – the other fish and organisms trapped by commercial fishing nets during fishing for a particular species.

#### С

caducous - easily shed (deciduous), usually scales.

**calanoid copepods** – small crustacea belonging to Order Calanoida, and found in the plankton.

**canine** – conical pointed tooth, usually larger than other teeth in the mouth.

**caniniform** – having the shape of a canine tooth.

**cardiform teeth** – numerous, pointed teeth arranged in distinct rows. **carinate** – with a keel or edge.

carnivorous - feeding on animals.

**cartilage** – skeletal tissue, usually soft and flexible, and in bony fishes it is mostly replaced by bone during the early growth stages.

**catadromous** – a term for fishes that are spawned in the sea and enter freshwaters as juveniles to grow to maturity, then return to the sea to spawn and die (opposite: anadromous).

**catadromy** – a life history strategy characterised by migration between marine habitats (used for reproduction) and freshwater habitats (used for growth).

cauda of sagittal sulcus - the posterior of the sulcus.

**caudal concavity** – difference between the longest (outer) and shortest (middle) rays of a lunate caudal fin.

caudal fin – the fin at the rear of the fish; tail fin.

**caudal peduncle** – the part of the body from the rear end of the anal fin base to the base of the caudal fin.

**centrum (pl. centra)** – one of the bones of the backbone. **cephalic** – pertaining to the head.

**cephalic tenacula** – appendage(s) on the head in male chimaeroids, both fossil and living, used to help grasp the female during copulation (at least in living species; function in fossils possibly the same).

- **cephalopod** a group of molluscs, including squid and octopus, which have a tubular siphon under the head and a group of muscular suckered arms around the mouth.
- **ceratobranchial** longest bones or cartilages of the branchial arches, situated immediately below the angle of the arch, between the epibranchials and the hypobranchials.
- ceratohyal bone, or cartilage, in ventral part of hyoid arch.
- **ceratotrichia** slender soft or stiff filaments of an elastic protein that resembles keratin.
- chaetognath torpedo-shaped, translucent, planktonic marine invertebrate organisms; arrow-worms.
- **character-based approach** an approach to defining a taxonomic group that includes all taxa that possess (at least ancestrally) a specified character or feature.
- **character reversal** the evolution of a character or feature of an organism to return to an ancestral condition.
- charge (of a molecule) molecules possess a net electrical charge, determined by the structure of the molecule (amino acids in the case of proteins), the atomic make-up of the molecule itself and the arrangement and nature of subatomic particles, under certain physical conditions (e.g., pH, chemical composition of the medium).
- **cheek** area between the eye and the edge of the preopercle bone. **chondrichthyans** – cartilaginous fishes (chimaeras, sharks, rays, etc.). **chorion** – the layer of tissue surrounding an oocyte, egg, or embryo. **chromatophore** – pigment cell in the skin responsible for skin colour.
- ciguatera poisoning (ciguatoxic) a type of food poisoning affecting the nervous system in humans caused by eating certain large reef fishes contaminated with ciguatoxin, a toxin produced by the dinoflagellate *Gambierdiscus toxicus*.
- **circuli** a concentric ridge on a scale of a fish.
- circumnarial around the nostrils.
- **circumoesophageal** around the oesophagus.
- circumpeduncular around the caudal peduncle.
- circumtropical distributed throughout the tropics.
- cirri (sing. cirrus) small, slender, filamentous appendages.
- **cladistics** a method for hypothesising relationships among organisms (phylogeny) by defining hierarchical groups defined by shared, derived features or characteristics (vs. defining groups based on overall similarity).
- **claspers** intromittent (copulatory) organs attached to the pelvic fins of male chondrichthyans.
- **cleave (DNA)** the breaking of the DNA molecule/strand into two (or more) fragments by breaking the phosphate bonds that link the individual nucleotides in the chain together. This can be done with the use of enzymes, chemically or by radiation.

**cleithral process** – a process on the major bone of the pectoral girdle.

- **cleithrum (pl. cleithra)** the major bone of the pectoral girdle, extending upward from the pectoral-fin base and forming the rear margin of the gill cavity.
- **cloaca** the cavity at the end of the digestive tract into which the genital tract also opens.
- cnidarians jellyfishes and relatives, of the phylum Cnidaria, characterised by the possession of special stinging cells called cnidocytes.
- coalesced fused or grown together.

- **coelenterate** radially symmetrical invertebrate animals including the corals, sea anemones and jelly fishes.
- **commensal** the relationship between two animals where one benefits and the other does not, nor is harmed.
- **common name** the informal vernacular name for a fish (or other organism), which may vary from place to place.

**compressed** – flattened from side to side.

**confluent** – joined together.

**congeneric** – of the same genus.

- **conspecific** of the same species.
- **continental shelf** the sea bottom from the shore out to a depth of 200 m.
- continental slope the sea bottom from 200 m to 2 000 m.
- **convective overturning** reversals of water masses with welldefined temperature and salinity regimes/characteristics.
- convex curved outward.
- **copepods** tiny planktonic crustaceans of major importance in marine food chains; some species are parasitic.
- **Coral Sea** the sea basin surrounded by northeastern Australia, New Guinea, the Solomon Islands and New Caledonia.
- **corselet** a densely scaled area in certain tuna-like fishes usually behind the pectoral fins.
- **cotylephore** a flap of skin that develops on the inner side of the pectoral fins of females, and in which the eggs are wrapped to provide them with nourishment.
- cranial fontanelle basically a 'hole in the skull'.
- craniates animals with a 'backbone' and a bony or cartilaginous skull.
- **cranium** the part of the skull containing the brain.
- crenate (cutting edge) a scalloped edge (see crenulate).
- **crenulate(d)** having a margin shaped into small, rounded scallops. **crepuscular** active at dawn and dusk.
- crown chondrichthyans present-day sharks, rays, skates and holocephalans and their immediate fossil relatives.
- **crown-group** a group comprising all extant members (and their extinct relatives) of a monophyletic group of organisms.
- crumenal see epibranchial.
- **cryptic** fishes with colour that conceals or camouflages them.
- ctenacanths a group of mostly Palaeozoic sharks characterised by specific dentitions and fin spines, usually phylogenetically placed on the chondrichthyan stem below the elasmobranch/ holocephalan divide.
- **ctenoid scale** a scale with minute teeth or spines (ctenii) on the rear margin, which gives the fish a rough feel when stroked towards the head.
- **ctenophores** comb jellies, of the phylum Ctenophora, that live in all oceans.
- **cuirass (cuirassed)** body armour, with breastplate and backplate fastened together.
- cultrate sharp-edged.
- cuneal wedge-shaped.
- cuneate (teeth) wedge-shaped teeth.
- **cusp** where two curves meet; top of.
- cutaneous pertaining to the skin, dermal.
- **cutaneous papillae (dermal papillae)** small papillae on the skin. **cycloid scale** thin, flexible scale with a smooth surface.

Cymodacea – a genus of seagrass. cyprinodontiforms – an order of small, mostly freshwater fishes.

#### D

decadal sea surface temperature (SST) anomalous months – monthly sea surface temperatures are compared to average sea surface temperatures for 1971–2000, and plotted on a map. Where they are higher or lower than the average they are referred to as anomalous months.

- deciduous easily shed or rubbed off.
- **demersal** living on or just above the bottom of the sea or a lake.

**demersal habitat** – on or near the sea floor.

- dendritic tree-like in shape or markings.
- **dentary** the anterior and largest of the bones making up the lower jaw that carry teeth.
- dentate bearing teeth or tooth-like projections.

denticle – the tooth-like scale of elasmobranchs; a tooth-like projection.

denticulate - tooth-like.

dentition – the characteristic arrangement and shape of the teeth.

depressed – flattened from top to bottom (dorsoventrally).

- dermal pertaining to the skin.
- dermal plicae folds in the skin.

diandric – protogynous species where some females change into (terminal) males.

diastema – the gap between two teeth.

diatom - unicellular alga with silica walls.

dichromatic - having two colour forms in the same species.

dignathic - teeth differing in shape in the upper and lower jaws.

- dimorphic having two forms.
- dioecious sexes are separate.
- **diphycercal** a type of caudal fin with a subtriangular shape in which both dorsal and ventral lobes are symmetrical and vertebrae continue towards the extremity of the fin.

**diploid** – having both chromosomes of the paired homologous chromosome set and, thus, both copies of each gene present in the cell nucleus; resulting from sexual reproduction.

**diplospondylous** – having two vertebrae in each body segment, as in the tail region of certain fishes.

**disc** – the combined head, trunk and pectoral fins of rays and skates. **distal** – furthest away from the point of attachment.

**distal-type teeth** – borne on distal extreme of long and closely packed fibrous strands, derived from jaw bones and supported by lip tissue.

**diurnal** – pertaining to the daytime; active during the day. **dorsal** – on or towards the back or upper part of a fish.

**dorsal-fin insertion** – the rear end of the fin, at the last ray; the anterior end of the fin is the origin.

**dorsomedian fontanelle (see also fontanelle)** – a gap between bones in the middle at the top of the skull, closed by a membrane.

dorsorostrally – upwards position of rostrum. dorsoventral – stretching from the dorsal to the ventral surface.

double emarginate – with a double-notched margin.

drumming muscles – the muscles that are attached to the swimbladder and produce drumming sounds when contracted.

**durophagous** – eating of hard-shelled (exoskeleton-bearing) organisms as a major part of the diet.

## Е

- ectoparasites parasites on outside of body.
- edentate without teeth.

efferent circulation – taking blood from an organ.

elasmobranchs – the group of fishes which includes the sharks, rays and skates.

elements – the spines or rays of a fin.

- emarginate with a slightly concave margin.
- **enameloid** an enamel-like tissue forming the outer layer of shark teeth and dermal denticles. Although the origin of enameloid is debated, it is probably more similar to dentine than true enamel.
- encoding (genes and proteins) the function and process whereby a particular gene produces a protein or enzyme; a gene responsible for a certain character, product, process, behaviour or phenotypic trait is said to *encode* for that feature.

endemic - native or restricted to a particular area.

- endochondral ossification a type of ossification (formation of bony tissue) in which osteoblasts replace hyaline cartilage with bone.
- endonuclease restriction enzymes bacterial enzymes which cleave (break) DNA molecules wherever a particular sequence of nucleotides (the restriction site) is recognised.
- **endopterygoids** paired dermal bones in the roof of the mouth of fishes.
- ENSO El Niño–Southern Oscillation; a recurring climate pattern involving changes in the temperature of waters in the central and eastern tropical Pacific Ocean.

ENSO teleconnections – the link between what happens in the Pacific Ocean during an El Niño event and what happens in the Indian Ocean.

entire - with a smooth or even margin.

- **Eocene** the period of geological time from 56–33.9 million years ago.
- epaxial muscles muscles that lie dorsal to a horizontal plane through the body of vertebrates (except humans).
- epibenthic living on the sediments on the bottom of the sea.
- **epibranchial** bone or cartilage forming the upper part of the gill arch, immediately above the angle of the arch.

epibranchial organ – a paired dorsal diverticulum at the back of the pharynx in some osteoglossiform, cypriniform, gonorhynchiform and clupeiform fishes.

- epioccipital ridges a ridge on a bone that lies above the eye.
- epiotic paired bones at the outer posterior margin of the skull.

epipelagic – the upper part of the oceanic zone from the surface to about 200 m.

**epithelium** – the thin tissue forming the outer layer of a body's surface and lining the alimentary canal and other hollow structures.

**epural** – an elongate detached bone above the urostyle and behind the last neural spine supporting caudal-fin rays.

erectile - can be raised or erected.

erisma (ventral arm of) – a thin bone with two 'arms' that lies in the middle between the skull and anteriormost dorsal interneural spines and below the rostral cartilage.

equatorial submergence – used for fishes and other marine creatures that move into deeper, cooler water when nearing the equator.

esca – the terminal lure or 'bait' on the angling apparatus (illicium) of anglerfishes (Lophiiformes).

ethmoid – the bone that separates the nasal cavity from the brain cavity.

ethmo-maxillary ligament – a ligament joining the ethmoid bone and maxilla that aids jaw protrusion in some fishes.

**euphausiids** – shrimp-like, planktonic crustaceans, which include krill. **euryhaline** – able to live in waters with a wide range of salinity.

evaginations – pouched-shaped structures.

eversible – can be turned inside out.

**excurrent** – liquid flowing outwards.

exoskeleton – hard outer parts that form an external skeleton to support and protect the body.

exserted – elongate; protruding.

extrinsic (musculature) - muscles 'outside' that act on a structure.

## F

facultatively monogamous – a mating system in which a male is not fully committed to one female, but chooses to stay with that female because there are no other mating opportunities available. falcate – sickle-shaped.

family – a category of organisms (taxon) which includes those genera that all share a common ancestor (see *genus*).

filamentous – thread-like.

**filter feeding** – to strain small food particles from the water by means of gill rakers.

fimbriate – with a fringed margin.

finlet – a small fin of one or two branched rays.

fish aggregating device (FAD) – man-made device used to attract pelagic fish species and used for fishing.

**fluorescence** – the property of absorbing light of short wavelength and emitting light of longer wavelength.

fontanelle – a gap between bones in the skull, closed by a membrane, where ossification of cartilage or connective tissue did not occur.

foramen – an opening through bone or tissue.

**foraminifera** – single-celled organisms with shells of various minerals, the shells with many tiny holes; may be planktonic or benthic.

**fork length** – the length from the tip of the upper jaw or snout to the end of the shortest ray of the caudal fin of a forked caudal fin.

**fossa** – a groove or pit.

frenum – a connecting or restraining membrane.

- frontoparietal fenestra a space between the frontal and parietal bones of the cranium through which a major nerve and blood vessel passes.
- **frontoparietal striae** the slight ridges or grooves on the bones above and behind the eyes.

**fulcral scale** – spine-like scales on the dorsal margin of the caudal fin. **fusiform** – tapering toward each end; spindle-shaped.

## G

Galaxea clavus & Galaxea fascicularis - species of coral.

galeomorphs – a diverse group of sharks that includes the orders Heterodontiformes (bullhead sharks), Orectolobiformes (carpet sharks), Lamniformes (mackerel sharks) and Carcharhiniformes (ground sharks). **ganoid scale** – a thick, bony scale covered by a layer of a hard tissue (ganoine); typically rhomboid or diamond shape; adjacent scales articulate by a peg-and-socket articulation.

gas bladder – see swimbladder.

**genital papilla** – a short, fleshy tube behind the anus from which the sperm or eggs are released; the sex of a fish can often be determined by the shape of its papilla.

**genotype** – the genetic make-up (profile) of the individual organism; the specific combination of genetic variants (e.g., a DNA sequence or a combination of alleles) for a genetic marker being studied or underpinning a character (phenotype) being considered.

**genus** – a category of organisms (taxon) which includes species more closely related to each other than to other, similar species, i.e., share a common ancestor.

geographically disjunct – two populations that occur far apart.

**ghost-lineage** – a branch of a phylogenetic tree that is inferred to have existed, but does not have a fossil record.

gibbous - convex or protruding.

**gill arch** – the bony or cartilaginous arches to which the gill filaments are attached.

gill chamber – the space on each side of the head containing the gills.

**gill cover** – the bones of the head that cover the gill chamber, comprising the preopercle, opercle, subopercle and interopercle; the operculum.

**gill filaments** – the soft, red, fleshy part of the gills, through which oxygen is taken into the blood from the water passing through the gills.

gill membrane – see branchiostegal membrane.

**gill opening** – the opening from the gill chamber to the outside; in elasmobranchs there are 5–7 openings on each side of the head; in bony fishes there is only one on each side (in some fishes the left and right gill openings may be continuous ventrally).

**gill rakers** – bony, finger-like (or lath-like) projections along the anterior edge of the gill arches. The number of gill rakers is often used to identify species.

gill slits – gill openings.

**gnathostomes** – vertebrates that have true jaws, or simply 'jawed vertebrates'; includes placoderms, acanthodians, chondrichthyans and osteichthyans.

gonad - the reproductive organ: ovary or testis.

gonadal sex reversal – when the gonads change from male (testicular) to female (ovarian) tissue, or the other way around.

**Gondwana / Gondwanan** – southern supercontinent following the break-up of Pangea, which included portions of modern South America, Africa, Arabia, Australia, Antarctica, and India, among other smaller regions; present both prior to the formation of Pangea, and following its break-up in the Jurassic.

gonochoristic - separate sexes, i.e., not hermaphroditic.

**gonopodium** – the modified anal-fin rays of males of certain fishes with internal fertilisation, and is used to transfer sperm bundles or spermatophores to females.

**gorgonian** – sea fans; a polyzoan coelenterate in which the polyps form upright branching colonies.

gravid - carrying unborn young; pregnant.

- **guanine** crystalline substance in dermis of fishes giving iridescent silvery or whitish colour.
- gular the area between the dentary bones; the throat.
- gular bones the median dermal bones that lie between the dentaries.
- gular flap in very large sharks the first gill slits are joined across the throat as a gular flap.

## Н

- habitat the place where a species normally lives.
- **haemal arch** the bony arch on the ventral side of a vertebra through which the dorsal aorta and caudal vein run.

haemal spine - spine on lower end of haemal arch in caudal vertebrae.

haploid – having only one chromosome (of the normally paired homologous chromosome set) and, thus, one copy of each gene present in the cell nucleus or cell organelle.

- haremic mating system a single male with a 'harem' of several females.
- **head length** the length of the head from the tip of the upper jaw or snout, where it projects over the upper jaw, to the posterior bony edge of the opercle.
- hemibranch a gill with filaments only on one side of gill arch.

herbivorous - feeding on plants.

- **hermaphrodite** having both sexes in the same individual (see also *protandrous, protogynous* and *synchronous hermaphrodite*).
- **heteracanth spines** asymmetrical dorsal-fin spines, with either the right or left side thickened.
- **heterocercal** caudal fin with the upper lobe larger than the lower.
- **heterodont** more than one kind of tooth in the same fish. **high aspect-ratio caudal fin** – a caudal fin of a fish that is tall and
- narrow, such as that of a tuna or billfish.
- **histotroph** uterine secretions that provide additional nutrition to embryos in some viviparous sharks and rays.
- holomorphic fossil a fossil that is more or less 'completely' preserved; fossils that include a good portion of the skeleton and body outline.
- **holothurian** sea cucumber, an elongate soft-bodied relative of the starfish.
- **holotype** a single specimen designated as the 'type' (i.e., 'name bearer') of a new species by the author of the original description.
- **homeothermic = endothermic** animals that regulate body temperature.

**homologous** – the same structure as; e.g., a bird's wing is homologous with a foreleg or arm or bat's wing.

homology - sharing the same evolutionary origins.

- **homonym** two organisms with the same name, of which the younger name is invalid.
- **homoplasy / homoplastic** a similar feature or characteristic that has evolved independently in different groups of organisms.

humeral – relating to the pectoral girdle or shoulder region. hyaline – translucent.

hybodonts – a diverse group of Palaeozoic and Mesozoic sharks that ranged significantly in size and morphology and that occupied many diverse habitats and niches, characterised by small cephalic spines in males, massive jaws and shoulder girdles, and characteristic fin spines and dentitions. Hybodonts are phylogenetically placed usually on the elasmobranch stem below neoselachians. hydrophiin sea snakes – sea snakes with a tail like a fin.

- **hypaxial myotomes** the blocks of muscles that lie ventral to the horizontal septum.
- **hypermineralised tooth plates** referring to the teeth of chimaeroids that undergo significant hardening by the excessive deposition of minerals to facilitate a durophagous diet.

 $\label{eq:hyperostosis} \textbf{hyperostosis} - excessive growth of bone.$ 

**hypersaline** – with salinities higher than that of sea water, i.e., more than 35 parts per thousand (ppt).

hypnosqualean hypothesis – a hypothesis that proposes that the batoids (rays and skates) are derived sharks, based on many morphological phylogenetic studies (cf. Shirai 1992, 1996; Carvalho 1996).

- hypobranchials paired bones on the lower part of the gill arch.
- **hypocercal** caudal fin with the lower lobe larger than the upper.
- **hypomaxilla** small, paired, tooth-bearing bone behind the premaxilla and below the maxilla.

hyposaline - having an abnormally low salinity.

**hypurals / hypural plate** – the fan-shaped series of bones (sometimes fused to one or two plates) to which the caudal-fin rays are attached.

#### I

illicium – the 'fishing pole' of anglerfishes, developed as a greatly modified dorsal-fin ray and located on top of the head.

imbricate - overlapping evenly.

*incertae sedis* – a qualification to indicate that a taxon's phylogenetic affinities are uncertain within a more inclusive group of organisms, and authors do not want to guess where a taxon belongs.

incised – notched

- incisor a tooth flattened at the tip like a chisel.
- Indonesian Throughflow an ocean current important for global climate, which moves warmer freshwater from the Pacific to the Indian Ocean, where most of the water flows from north of the Equator, past the south of the Philippines, between Borneo and Sulawesi and south between Bali and Lombok, Indonesia, into the eastern Indian Ocean.

Indo-Pacific – the tropical and subtropical waters of the area of ocean from the east coast of Africa, including the Red Sea and Persian/Arabian Gulf to Easter Island in the eastern Pacific, and in which the marine fauna are of a common origin. (Note: only broad distributions of species outside the Western Indian Ocean are given)

inferior – the position of the mouth when it is on the underside of the head.

infraorbital – below the eyes.

- **infrapharyngobranchial tooth plate** a tooth plate on bones in the pharynx (throat) of a fish, behind the gills.
- **insertion (of a fin)** the anterior end of the base of the paired fins; the posterior end of the base of a dorsal or anal fin.

interarcual cartilage - a cartilage in the pharyngeal arch.

**interbranchial septa** – the partitions separating the gill chambers. **interdorsal** – see *basidorsals*.

**interhaemal bone** – the bones between the haemal (ventral) spines of the vertebrae and the spines or rays of the anal fin.

internarial - between the nostrils.

**interneural bone** – the bones between the neural (dorsal) spines of the vertebrae and the spines and rays of the dorsal fin.

interorbital – the area between the eyes, on top of the head. interorbital commissure of supraorbitals – the suture of the supraorbital bones between the eyes.

**interpelvic process** – a narrow, pointed area between the pelvic fins. **inter-radial membrane** – the membrane between the rays of a fin. **interspecific** – between separate species.

interspinous membrane – the membrane between the spines of a fin.

**intertidal** – between the high and low tide marks on a shore. **intraspecific** – within one species.

**iodoform** – smelling of iodine, odour of flesh from a fish that feeds on sponges or tunicates rich in iodine compounds.

**iridescent** – with a wide range of often-changing, brilliant colours. **iris** – the black part of the eye, which is not the pupil (see *pupil*).

- **iris lappet** a fleshy flap-like structure in the eye, which can be short and rounded, simple or branched.
- **isobath** a line on a map that joins all points of the same water depth in the sea.
- isthmus the lower part of the head that separates the two gill chambers

isthmus muscle – the muscle that lies between the two gill chambers.

#### J

jugular – located in front of the pectoral fins.

junior synonym – a younger name for a species named earlier.

Κ

**keel** – a narrow ridge on the sides of the caudal peduncle or at the base of the caudal fin, typically found in fast-swimming fishes with narrow caudal peduncle. Term is also used for sharp blade on the peduncle plates of *Naso* and *Prionurus* species; a narrow ridge-like process.

**keeled scales** – scales with a narrow ridge along the middle. **keratin** – a protein that protects epithelial cells from damage.

keris larvae – the late post-larval stage of fishes of the genus Naso, which come inshore from the open ocean to metamorphose into juveniles.

kolachi vala nets - a kind of a drag net.

## L

labial – pertaining to the lips.

**labial ossicles** – small bones associated with the upper 'lips' of certain species of *Bleekeria*.

lachrymal - the anteriormost bone below the eye.

lacustrine – living in lakes.

lagerstätte – a sedimentary deposit that has extraordinarily preserved fossils, usually in abundance, and sometimes even with preserved soft tissues.

lamella (pl. lamellae) – plate-like fold of skin.

- lanceolate broad at the base and tapering to a point.
- **larva (pl. larvae)** the early life history stage between the time of hatching and transformation to a juvenile, the latter a miniature replica of the adult.

lateral – pertaining to, at, or towards the side.

lateral ethmoid – the bone forming the front of the eye socket.

- **lateral line** a system of tubes or canals containing cells sensitive to low-frequency sounds and vibrations produced by the movements of other animals in the vicinity.
- **lateral-line scales** the perforated or tubed scales on the body that are sensitive to changes to water pressure. These pored lateralline scales are often counted, with the count stopping at the caudal-fin base.
- **lateral scale series** this is generally the number of scales from the upper end of the gill opening, often lying just above the lateral line, to the base of the caudal fin. This may vary for different fishes as not all species have the same lateral-line configuration, and where different it is explained in the family account.

**Laurasia** – the northern supercontinent following the break-up of Pangea, which included portions of modern North America and Asia.

- **lecithotrophic** where embryos receive nutrition entirely from yolk reserves.
- **leptocephalus** the transparent, ribbon-like, pelagic larva of eels and other elopomorph fishes.
- **Lessepsian migrant** an organism that moves from the Red Sea to the Mediterranean Sea via the Suez Canal, and more rarely in the opposite direction (anti-Lessepsian).

lingual lure - a worm-like lure on the tongue.

- lipid fatty substances produced by fishes as an energy store and, in some species, to provide buoyancy; may be deposited in or around the swimbladder as fat, or as oil in the liver, skeleton or body muscles.
- **littoral** the intertidal zone of the marine environment, delimited by the tide marks of low and high water.
- lunate sickle- or moon-shaped.
- **lymphocytes** small white blood cells found in the lymphatic system.

## М

**malar thorns** – patch of thorns found on some mature male skates close to the edge of the upper surface of the disc opposite the eye.

 mandible – the lower jaw.
 Mascarene Plateau – an extensive WIO submarine plateau, extending from the Seychelles to Réunion in the west and to St

Brandon Shoals in the east.

Mascarenes (or Mascarene islands) – Réunion, Mauritius, Rodrigues and St Brandon Shoals.

**matrotrophic** – in viviparous species where nutrients are transferred to the developing embryos from the mother.

**maxilla (pl. maxillae)** – one of the two bones that comprise each half of the upper jaw.

medial – on or towards the middle of the body.

median – on the midline or axial plane of the body.

- median fins the dorsal and anal fins.
- **melanophore** a pigment cell containing melanin that gives skin a black, grey or brown colour.
- **membranous labyrinth** the fluid-filled, adjoining canals of the inner ear housed in the bony labyrinth of the skull and responsible for the sense of equilibrium.

Mendelian inheritance – the rules of genetic inheritance in diploid organisms, as deduced by Gregor Mendel's seminal 'genetic' hypotheses based on breeding, cross-breeding and observing the phenotypes of garden peas. This work suggested that phenotypes are controlled by two factors (alleles in a genetic context), one obtained from each parent; that these factors assort or act independently of factors for other phenotypes during cell division (meiosis) and fertilisation; that these factors can be dominant or recessive with respect to influencing the phenotype and that the phenotypic outcomes can be predicted numerically/statistically, based on the phenotypes present in the population and by tracing a genealogy.

mental – pertaining to the chin.

**meridional** – along a meridian or in a north–south direction.

meristic – countable features (e.g., lateral-line scales, fin rays or gill rakers).

**mesopelagic** – the region of the oceanic zone from 200 m to 1 000 m.

microphagous detritivores – animals that feed on small particles of detritus.

**mimic** – an organism that closely resembles another, usually unrelated, organism which is protected from predation for some reason.

**Miocene** – the period of geological time from 23.03–5.33 million years ago.

mitochondrial DNA – an extranuclear (i.e., occurring outside the cell nucleus), double-stranded, circular DNA molecule found in the mitochondria of animal cells; the molecule is maternally inherited and haploid.

molar - a blunt, rounded tooth adapted for crushing shellfish.

**molecular clock estimates** – the estimates of time of divergence between taxa based on mutation rates of DNA.

**monandric** – initial-phase fish all female, terminal phase all secondary males.

mongolepids – refers to an order of early fossil sharks known from isolated dermal denticles from the Ordovician or Silurian of Colorado (USA), Mongolia and China; includes the genera *Mongolepis*, *Solinalepis*, *Rongolepis*, *Teslepis*, *Shiqianolepis*, *Xinjianqichthys*, and *Sodolepis*.

monocuspidate - teeth ending in a single point.

**monophyletic** – a group of organisms (a clade) comprising the most recent common ancestor and all its descendants.

monophyly of genus – all species sharing a common ancestor.
 monospecific / monotypic – including only a single species.
 monospondylous – having a single vertebra in each body segment.
 morphology – the form and structure of organisms or the study of this subject.

mucus – a slimy substance secreted by the skin of fishes.

multicuspid – a tooth with many cusps.

multiserial – arranged in several rows or series.

myomere – a segment of the body (trunk) muscles, seen as V- or W-shaped muscle fibres, separated from its neighbours by a connective tissue septum (myosepta) and most easily seen in larval fishes; the number of myomeres is important in identifying larvae, since their number equals the number of vertebrae in the adult.

**myosepta** – the layer of connective tissue between myomeres. **mysids** – small, shrimp-like crustaceans found in all oceans and seas.

## Ν

**nape** – the part of the dorsum from the back of the head to the dorsal fin or vertical at the base of the pectoral fins.

naris (pl. nares) - nostril.

- **nasal fossa** the cavity or pit, containing nasal papillae, into which the nostrils open.
- **nasopharyngeal duct** the duct through which hagfishes take in water.
- **nasoral/oronasal groove** groove between the nostril and corner of mouth.

**nautiloid** – refers to the mostly extinct cephalopod molluscs of the group Nautiloidea, with shells that are coiled and with forward-facing concave septa, and that flourished in the Palaeozoic seas; a few genera, such as *Nautilus*, are extant.

**nekton** – organisms that actively swim in the water column. **neonate** – newborn.

neritic - the shallow pelagic zone over the continental shelf.

**neural arch** – the bony arch on the dorsum of a vertebra through which the spinal cord runs (see *haemal arch*).

**neural spine** – the spine on the neural arch of a vertebra.

**neurapophyses** – the structure that forms the neural arch of vertebrae.

**neurocranial** – the skeletal structure surrounding the brain and cranial sensory organs.

neurocranium – skull.

**neuston** – small, aquatic organisms that live in the surface of the water.

**niche** – the habitat and role (community relationships, food habits, etc.) of an organism.

nictitating membrane (nictitans) – a clear, moveable inner eyelid found in sharks.

**nocturnal** – active at night.

**node-based approach** – an approach to defining a taxonomic group that includes all taxa that share a most recent common ancestor.

**non-otolith fossil record** – the fossil record based on the remains of fishes other than their otoliths; otoliths are rarely found with the remains of the animal and are often of limited value.

**notochord** – the cartilaginous rod in the embryo that runs from head to tail and supports the nerve cord; in most fishes the notochord is replaced in the larval stage by the developing vertebrae, but in some primitive fishes (e.g., *Latimeria*, hagfishes, chimaeras) the notochord persists in the adult and serves as the vertebral column or 'backbone' of these species.

**nuchal** – pertaining to the nape.

nuchal crest – a fleshy crest on top of the middle of the head.

nuchal plate – a plate of bone at the neck.

**nyctoepipelagic** – migrating from deeper waters to the epipelagic zone at night.

## 0

**obligate inquiline** – an inquiline is an animal that exploits the living space of another (from the Latin *inquilinus*, a tenant).

obsolete - disappearing or scarcely evident.

obtuse - broadly rounded, having a blunt end.

occipital (pertaining to the occiput) – the upper rear part of the head.

occipital fossa – depression at the rear of the skull.

oceanic – pertaining to the open ocean beyond the continental shelf.
 ocellus (pl. ocelli) – pigment pattern on body or fins forming a false eve.

ocular - pertaining to the eyes.

- odontoid process a small, bony process on the dorsum of the second vertebra; also a small, tooth-like bony process on the dentary (part of the upper jaw).
- olfactory pertaining to the nasal organs or sense of smell.
- **oophagy / oophagous** where the embryos feed on unfertilised eggs in the uterus of viviparous species.
- **opercle** the large, posterior bone of the gill cover, the largest bone of the operculum.

opercular - pertaining to the operculum (gill cover).

- opisthotic a deep, endochondral bone forming the ventral element of the otic capsule, covering the ampulla of the posterior semicircular canal.
- orbit the dermal or bony eye socket.
- order the taxonomic rank that includes one or more families of organisms, all of which share a common ancestor.
- origin (of a fin) the anterior end of the base of a fin.
- ossicle bony plate.
- ossified become bone.
- ostraciids boxfishes.
- otolith(s) calcareous concretions in the ear capsules of bony fishes; also called 'ear bones' or 'ear stones'; there are three otoliths, the sagitta (largest), the astericus and the lapillus.
- otophysic connection an extension from the swimbladder to the inner ear.
- **oviparous** producing eggs that develop and hatch outside the body of the female.

**ovocyte** – a cell in the ovary that divides to form an ovum. **ovoviviparous** – see *yolk-sac viviparity.* 

## Ρ

Pacific Plate – that part of the Pacific Ocean floor bounded by the Aleutian Trench in the north, the trenches that extend from the Okhotsk Peninsula, past Japan, the Mariana Trench, to the Tonga and Kermadec trenches to cut through South Island, New Zealand, to the Antarctic Plate in the south. It is bounded on the east by mid-ocean ridges that extend past Easter Island to Baja California and along the west coast of North America to Alaska. It covers 103.3 million sg. km.

paired fins - the pectoral and pelvic fins.

- **palaeoclimatic evidence** what geological information can tell us about climates thousands to millions of years ago.
- **palate** the roof of the mouth.
- **palatines** elongate bone on each side of the palate.
- **palato-premaxillary** of, relating to, or connecting the palate and the premaxilla.
- **Pangea** the large supercontinental landmass that persisted from the late Palaeozoic to the early Mesozoic eras.

papilla (pl. papillae) – a small fleshy projection.

papillose - with many papillae.

- **parahypural** the last haemal spine that forms part of the hypural plate.
- **paralectotype** an additional specimen selected from a series of syntypes, after a lectotype has been designated.

- **paraphyletic** a taxonomic grouping that does not include all descendants of a most recent common ancestor.
- **parapophysis (pl. parapophyses)** a long, transverse process arising ventrally from the centrum of abdominal vertebrae; serves to support epipleural ribs when present and, in Gadidae, the gas bladder.
- parasphenoid a bone in the middle at the base of the skull.
- **paratype** a specimen, other than the holotype, on which the description of a new species is based.
- parsimony analysis a way of analysing data to give the most reasonable (or parsimonious) answer.

parturition - the process of giving birth.

- **pectoral-fin insertion** lower or rear end of the fin; the upper end of the fin is the origin.
- **pectoral fins** the anterior or uppermost of the paired fins, corresponding to the forelimbs of higher vertebrates.
- **pectoral girdle** the bony or cartilaginous skeletal arch supporting the pectoral fins.
- **pedicellate** with a stalk.
- pedicle a small, stalk-like structure connecting two structures.
- **pelagic** pertaining to the open ocean.
- pelvic claspers organs present in male sharks, rays and skates for introducing semen directly into the reproductive tracts of females.
- **pelvic fins** paired ventral fins usually located below or behind the pectoral fins.
- pelvic scute a long, pointed scale above the pelvic-fin base.
- percomorph fishes belonging to the Percomorpha, a large clade of bony fishes, mostly perciform, characterised by having bony fin spines, two dorsal fins, thoracic pelvic fins, a ductless swimbladder and ctenoid scales.
- peritoneal spots pigment spots on the peritoneum.
- peritoneum the membrane lining the abdominal cavity.
- **pharyngeal arch** a segment of the lateral wall of the pharynx that will form jaw and gill structures.
- **pharyngeal plates** the bones in the pharynx that carry teeth.
- pharyngeal sulcus a groove on either side of the
- pharyngobranchial chamber, part of the filter-feeding mechanism. **pharyngeal teeth** – teeth located on the pharyngeal bones
- (modified fifth gill arches) in the throat behind the gills.
- **pharyngobranchial tooth plates** tooth plates on bones in the pharynx (throat) of a fish, behind the gills.
- **pharyngocutaneous duct** a duct between the pharynx and the skin behind the last gill of hagfishes that clears large particles from the pharynx.
- **phenotype** physical features of the organism or aspects of the organism's biochemistry, physiology or behaviour that can be observed and are thought to reflect variation in the genotype or the interaction of the genotype and the environment.
- **philopatry** the habit of many animals to return to a particular site or place for various reasons, either to spawn at the same place they were spawned (e.g., salmon and anguillid eels), to breed (many sea birds), or to associate with their own species (raggedtooth sharks gathering by the dozen in shark gutters).
- **photic zone** the surface layer of the ocean that is reached by sunlight, and hence where photosynthesis can take place (roughly to 80+ m depth).

photophore - a light-emitting organ.

- **phylogeny / phylogenetic analysis** the hierarchical evolutionary relationships among fossil and living organisms, and the process for hypothesising such relationships.
- phylogenetically informative characters characters that reveal the historical patterns of phylogenetic relationships among organisms.
- **physoclistous** with a closed swimbladder (i.e., without any connection between swimbladder and gut).
- **phytoplankton** microscopic (unicellular) plant life that floats in the open ocean.
- phytoplankton bloom a large and sudden increase in microscopic algae caused by an increase in nutrient content in an area, common in the world's oceans, and often changing the local colour of the water.
- pinniform shaped like a fin.
- placentotrophic way of feeding embryos of viviparous species where nutrients are transferred across the mother's uterine epithelium which is in close contact with foetal tissue (a placenta).
- placoderms refers to a Palaeozoic (Silurian to Devonian) gnathostome group of armoured fishes with dermal bony plates on head and trunk, and which occupied numerous habitats and niches from large pelagic predators to small benthic forms; presently considered to not be monophyletic and placed on the stem below the divergence between osteichthyans and chondrichthyans.
- **placoid** plate-like scales of some extinct fishes, and of sharks and rays.
- planktivorous feeding on plankton.
- **plankton** small floating organisms that drift more or less passively with the ocean currents.
- plesiomorph / plesiomorphy / plesiomorphic an underived or 'primitive' character that cannot be used to define a group of organisms.
- pleural rib the ribs of the 'rib cage' around the viscera.
- **plicate** with pleat-like folds.
- **poikilothermic** = ectothermic animals that do not regulate body temperature, but have body temperature of their surroundings.
- polychaetes segmented marine worms with bristles.
- **polygynous** a male with many females.
- polyphagous feeding on different kinds of food.
- **polyphyly / polyphyletic** a group of organisms (a clade) that has more than one ancestor and is defined on the basis of convergent features or characteristics.
- **polytomy** unresolved branches within an otherwise hierarchical hypothesis of relationships among organisms.
- **population** a group of organisms belonging to the same species that co-occur in a distinct area.
- **postcleithrum** a bone behind the cleithrum of the pectoral girdle. **posterior** – at or towards the rear end of the fish.
- **post-interorbital knobs** bony knob posterior to the interorbital area.
- postlarva a larva after it has absorbed its yolk.
- post-maxillary process an extension posterior to the ascending process.
- **postocular, postorbital** that part of the head behind the eye.
- **post-pelvic scutes** keeled scales along the midline of the 'belly', from pelvic fin to anal fin.

- **precaudal pit** the notch just in front of the caudal fin in sharks. **prehensile** – able to grasp, and used as an anchor around a variety of
- objects in the wild and coiled ventrally in preservative.
- **prelingual filament** the worm-like filament on the floor of the mouth.
- **premaxillae (premaxillaries)** the main bones of the upper jaw that carry teeth; they lie in front of and below the maxillae.
- premaxillary diastema space between the premaxillary bones.
- **preopercle** the falcate bone before the opercle and behind and below the eye, and constituting the fore part of the operculum; its rear edge is usually free, and may be serrated or not.
- preorbital the bone or region before and below the eye.
- **pre-pelvic scutes** keeled scales along the midline of the 'chest', in front of the pelvic fins.
- preural vertebra the second-last vertebra of the spine.
- procumbent lying down and pointing forwards.
- **produced** elongated or projecting.
- prognathous jaw the lower jaw protruding in front of the upper jaw.
- **propagule** a vegetative/living structure that becomes detached from a parent to give rise to a new organism; an agent of reproduction, e.g., a sucker on a plant or the eggs fishes or corals shed into the water.
- **protandrous hermaphrodite** hermaphroditism that functions first as a male and then changes into a female.
- **protocols** a set of procedures or methodology to be followed in a laboratory.
- **protogynous hermaphrodite** an individual that functions first as a female and then changes to a male.
- protrusile, protrusible capable of being thrust out or extended forwards.
- **proximal** towards/closest to the body or base of attachment. **pseudobranch (pl. pseudobranchiae)** – a small, gill-like organ on
- the inner surface of the operculum; the reduced first gill arch.
- **pseudobranchial filaments** filaments of the small, gill-like organ on the inner surface of the operculum.
- **pseudoceanic** members of a basically oceanic group that are distributed over continental shelf and slope regions and in the neighbourhood of oceanic islands and that are associated with land-orientated food chains.
- **pseudoclasper** an accessory male copulatory organ, inserted into the female during spawning.
- **pseudocycloid scales** a few species of fishes fail to produce ctenii on their scales, and such 'secondary' cycloid scales are called pseudocycloid.
- pterotic a bone on the side of the cranium.
- **pterygiophore(s)** the three bones (distal, medial and proximal), sometimes some fused together, that support and articulate with the fin spines and rays.
- **pterygoid teeth** the teeth on the bones of the roof and walls of the mouth, behind the palatines.
- **pterygoids** bones of the roof and walls of the mouth in fishes, behind the palatines.
- pungent sharp-pointed.
- **pupil** the opening at the centre of the iris, is white or transparent, and is usually covered by the iris (see *iris*).

- **pyloric caecum (pl. caeca)** a variously shaped diverticulum from the junction of the stomach and intestine; the pyloric caeca may be long and numerous, or short and few, or profusely branched, or even absent.
- pyriform (scales) pear-shaped, with an extended middle posterior margin.
- **pyrosomes** large, free-floating colonial tunicates of the genus *Pyrosoma*.

## Q

**quincunx** – an arrangement of five objects in a rectangle, with one on each corner and one in the middle.

## R

- radial see pterygiophore.
- ramus a branch; one side of the lower jaw.
- raptorial adapted for seizing prey.
- recumbent spine a spine that is more or less parallel to the horizontal (lying down), rather than almost vertical.

recurved – curved backward or inward.

- replacement scales tilefishes inhabit mounds and burrows. As they move in and out of a burrow they apparently rub against the walls of the mound or burrow losing scales in the process. The scales that grow back lack the early annuli ('growth rings'), and are called replacement scales to differentiate them from the original scales. respiratory – associated with breathing.
- **rete mirabile** a network of small veins and arteries that lie very close together, where heat is transferred from outgoing venous blood to incoming arterial blood.
- reticulate resembling a network.
- **reticulum** a network of intercellular fibres in certain tissues, with a fine, net-like appearance.
- **retrorse (spines)** pointing or curved backward.
- rhipidion longitudinal, elongated flap of soft tissue on the distal opening of the claspers of chondrichthyans through which sperm is dispersed.
- rhomboidal diamond-shaped.
- rictus the corner or gape of the mouth.
- rocker bone a bean-shaped bone which is drummed against the anterior end of the gas bladder to produce sound.
- rostral pertaining to the snout or rostrum; towards snout (usually anteriorly).
- rostral cartilage the cartilage on the snout.
- rostral organ an organ on the snout.
- rostral process a beak-like structure on the rostrum or 'nose'.
- **rostrum** a projecting snout or beak; protracted anterior part of the skull in sharks and rays.
- ruga (pl. rugae) ridge or corrugation.
- rugose with a rough or wrinkled surface.
- rugosities wrinkles and lumps on a surface.

## S

saccus vasculosus – an organ associated with the hypothalamus of many jawed fishes, that seems to be able to sense the seasons.

**sagitta** – the largest of the otoliths or ear-bones.

Sahul Shelf – the section of continental crust which forms Australia and New Guinea.

**salinity** – the measure of dissolved salt content in water, usually expressed as a ratio in parts per thousand (ppt); average ocean salinity is ~35 ppt (or 3.5%, or 3.5 g of salt per litre of water).

salp (pl. salps) – a pelagic, barrel-shaped tunicate.

Sargassum – a genus of brown seaweed (macroalgae) common in temperate and tropical seas.

**scapula** – a bone of the pectoral girdle.

scientific name – the formal binominal name of an organism, which consists of the genus and specific names; a species has only one valid scientific name.

sclera (of eye) – the outer, protective layer of the eye; outside of the pupil and the iris; the transparent outer layer of the eye overlying the pupil (scleral cornea of pupil).

scleractinian corals – stony or hard corals that form coral reefs today.

- scute (pl. scutes) a modified (thickened) bony scale with a keel or spiny point.
- **scutella** the shield-shaped, bony elements between trunk and body rings; the rectangular, fused body scales of Syngnathidae.

**seamount** – a mountain rising from the ocean floor.

**segmented ray (soft-ray)** – a bilaterally paired fin ray that is marked by cross striations like the joints on a stalk of bamboo.

- **semicircular canal** fluid-filled canals embedded in the cranium and concerned with balance and hearing.
- sensu lato in a broad sense; the name of a taxonomic group used in such a way that it includes taxa that have been referred to a group, whether or not they have been robustly shown to share a mostrecent common ancestor with that group.

sensu stricto – in the strict sense; a strictly defined taxonomic group; usually used to exclude taxa that were historically allied to a particular group but are of questionable relation to that group.

serrae – saw-like teeth or serrations.

- serrate, serrated bearing small saw-like notches or spines (serrae).
- sessile an organism that lives attached to the substrate.

sessile teeth – directly attached to jaw bones.

setae – bristles.

- setiform bristle-like.
- **sexual dimorphism** males and females of the same species with different body form.
- **sexually dichromatic** males and females of the same species with different colour patterns.

shagreen-like – rough, like shark skin.

silica – silicon dioxide, a mineral used by marine organisms such as diatoms to build the cell wall.

simple - not divided into branches.

- **siphonophores** colonial, free-swimming or floating pelagic hydrozoans, mostly delicate and transparent with long filamentous tentacles.
- sipunculids a group of unsegmented marine worms that live in burrows or discarded shells; some bore into rock or coral. Also called peanut worms.
- **sister-group relationship** two taxa of equal rank that share a common ancestor.

somites - body muscle segments.

- **spawning-site fidelity** returning to the same site to spawn each season.
- **species** groups of actually (or potentially) interbreeding natural populations which are reproductively isolated from other such groups.

**spermatogonia** – cells produced early on in the formation of spermatozoa.

spine – a sharp, projecting point; a fin ray that is unpaired, unbranched, lacks segmentation and is usually stiff and sharppointed.

spinescent – with spines.

spinning down – losing energy, slowing down and dissipating.

**spinoid (scales)** – modified ctenoid scales with spines fused to the surface and/or posterior margin.

spinules - small, spine-like structures.

**spinulose** – with spines.

**spiracle** – an opening behind the eye of some elasmobranchs that leads into the mouth cavity and may serve as an incurrent opening for water entering the gill chambers.

spiral valve – a spiral structure in the intestines of some fishes.

**spur and groove zone** – a zone of finger-like projections of coral (the spurs) in a fringing reef that grow out towards oncoming waves, with 'valleys' or grooves in between, which together serve to break the energy of incoming waves.

squalene - oil produced in the liver of some sharks.

**standard length** – the length from the tip of the upper jaw or snout to the end of the hypural plate. This can usually be seen as a crease when the caudal fin is bent from side to side.

**stem chondrichthyan** – refers to a fossil shark that is placed very low on the chondrichthyan phylogeny, below the divergence of elasmobranchs and holocephalans.

stem elasmobranch – refers to a fossil shark that is placed very low on the elasmobranch phylogeny below the divergence of 'neoselachians', the level of more modern sharks and rays.

stem-group – the basal members of a taxon that are known exclusively as fossils, and that are hypothesised to lie outside of a crown-group taxon.

stem holocephalan – refers to a fossil holocephalan that is placed very low on the holocephalan phylogeny below the divergence of more modern chimaeroids.

stomatopods – mantis shrimps.

strangulate – close-set, dense, not sparse.

**stratophenetic** – relating to overall similarity in form and geologic age.

striate - marked by lines or grooves.

sub (as a prefix) - less than, not quite, approximately, below, under. subcutaneous - under the skin.

subequal – almost, but not exactly, equal.

subjective synonym – two species/names that do not have the same type specimen.

subocular (suborbital) - beneath the eye.

subopercle – the gill-cover bone that is ventral to the opercle.

**subspecies** – a taxonomically and geographically distinct subgroup within one species.

substrate – the material forming the floor/bottom of the sea.
sulcus – the indentation on the inner surface of the sagitta.
superior – above; dorsal to.

**supernumerary spine** – an additional spine (or spines); more than the normal single spine supported by a pterygiophore.

supracleithrum – the dorsalmost bone of the pectoral girdle in bony fishes, usually articulated with the post-temporal bone of the skull.

supramaxilla – a small bone along the upper rear edge of the maxilla.

supraneural bones – unpaired bones above the neural spines of the anterior vertebrae, between the skull and the origin of the dorsal fin.

**supraoccipital** – a median bone at the upper rear surface of the skull, frequently bearing an expanded crest (supraoccipital process).

**supraorbital crest** – narrow cartilaginous ridge from the chondrocranium extending above the eyes in catsharks of the family Scyliorhinidae. The crests can usually be felt by touch without needing any dissection.

suprascapular spines – spines on the posttemporal area.

**supratemporal bones** – a series of 2–8 small bones bordering the posterior margin of the roof of the skull.

**supratemporal commissure** – the lateral-line canal that crosses the back of the cranium.

**suspensorium** – the bones from which the lower jaw is suspended. **suture** – line where two bones join.

**swimbladder** – a membranous, usually gas-filled sac attached to the dorsal surface of the body cavity; in some fishes the swimbladder is filled with fat, and in a few species it is rudimentary or absent.

**symmoriiforms** – a diverse group of Palaeozoic sharks (or possibly basal holocephalans) known from many morphologically unique genera displaying strong sexual dimorphism.

**sympatric** – groups of organisms living together in the same spatial or geographic area.

symphyseal (or symphysial) – relating to a symphysis.

**symphyseal teeth** – teeth at the junction between the two halves of the lower jaw.

**symphysis** – the median point or junction of the two halves of the jaws; an immovable joint between two bones.

**symplectic** – a bone that lies between the hyomandibular and quadrate in the suspensorium.

**symplesiomorphic** – shared primitive characters, e.g., a genus of fishes where all but one species have palatine teeth; the loss of the teeth would be apomorphic (derived) and the retention of the teeth would be plesiomorphic, the original condition.

synapomorph – a shared, derived character, indicating common ancestry.

**synchronous hermaphrodite** – a hermaphrodite with both ovaries and testes that function at the same time.

**synechodontiforms** – an extinct order of sharks, mostly Mesozoic, which are phylogenetically close to living orders of elasmobranchs (sharks and rays) as they share, for example, calcified vertebral centra.

synonym – different names applied to the same taxon.

**synonymy** – chronological list of all scientific names that have been applied, correctly or incorrectly, to a given taxon.

**syntype** – one of the specimens on which a species description is based if no holotype was designated.

## Т

tanaids - a small order of shrimplike Crustacea, the Tanaidacea.

tandem articulation – two bones, one behind the other, that articulate with each other.

**taxon (pl. taxa)** – a group of organisms such as a species, genus or a family that share a common ancestor.

**taxonomic** – pertaining to the classification of organisms according to their natural (evolutionary) relationships; systematic.

tegument - the covering membrane of the body.

**teleost** – a 'modern' (advanced) bony fish (not including *Latimeria* and some other primitive fishes).

tenaculum (pl. tenacula) – an appendage on the forehead of male chimaeroid fishes, weakly differentiated to ridge-like in male juveniles (frontal tenaculum); additional small, retractable appendage on each side just in front of the pelvic fins with intromittent organs (claspers) of chimaeroid males, only partially differentiated in male juveniles, and hidden in a slit when retracted (pre-pelvic tenaculum).

**teratogenic pollution** – teratogens are substances that cause physical or functional defects in embryos.

**terminal** – pertaining to a mouth at the anteriormost part of the head.

tetrodotoxin – a potent neurotoxin found in the skin and some other organs, present in many tetraodontiforms, and also some gobiids.

**thermocline** – the distinct interface between warmer surface waters and cooler deeper waters.

tholichthys stage – the larval stage of a butterflyfish (Family Chaetodontidae).

thoracic – under the pectoral fins.

thorn – large denticle on the surface of a ray or skate.

time-calibrated – the use of minimum ages from the

palaeontological record to constrain molecular clock analyses. tooth file – see tooth series.

**tooth row** – in sharks, teeth arranged parallel to the jaw line that are at the same stage of development.

tooth series – in sharks, teeth at different stages of development, arranged from the front of the jaw inwards; also known as a file of teeth.

total length – the length from the tip of the snout to the end of the caudal fin.

transverse rows of scales (also transverse scale rows) – counted between a specified point along the dorsal-fin base, e.g., fin origin, and pelvic-fin insertion or anal-fin origin, depending on the fish; sometimes specified as two values—above and below the lateral line.

tricuspid - with three cusps or points.

tritor – grinding surface on a tooth, such as in Rhinochimaeridae (longnose chimaeras).

tropeic fold – ventral fold under the belly of some sharks.

truncate - having the end square-cut or even.

tunic ridges - ridges on the swimbladder.

tympanic spine - a spine near the 'ear'.

**tympanum** – an area of enlarged scales behind the gill opening of some Balistidae that can be vibrated to produce sound.

## U

uncinate processes – a hook-shaped projection.

unicuspid – teeth with a single-point cusp.

uniserial – arranged in a single row.

**urohyal** – bone at anterior floor of mouth, between the two sides of the hyoid arch.

urostylar centrum – the last vertebra, and part of the caudal skeleton.

## V

**vegetative (or vegetal) pole** – the side of an egg opposite to the 'animal' pole; cell growth is slower in the vegetal pole which contains more yolk.

**vent** – the external opening of the alimentary canal.

**venter** – relating to the underside, the abdomen.

ventral – on or towards the lower surface (underside) of the fish.

ventral fins – pelvic fins.

ventrorostrally - downwards position of rostrum.

**vermiculations** – a pattern of fine wavy lines; worm-like lines of colour.

vermiform - worm-like.

vertical fins – the dorsal, caudal and anal fins; median fins.

vestigial – the remaining part of an earlier, functional structure.

vicariance / vicariance biogeography – an approach to

understanding the distribution of organisms related to physical or biological separation between groups of organisms (in contrast to dispersal).

villi – fine bristles.

villiform – having the appearance of velvet or fine bristles of a brush.
 villose papillae – papillae (like taste buds) that look like fine bristles.
 viscera – the internal organs of the body cavity.

viviparous – bringing forth living (active, free-swimming) young. volume flux – a measure of water flow, where a particular

volume (m<sup>3</sup>) flows across a unit area (m<sup>2</sup>) in a unit of time. **vomer** – a median bone in the front of the palate that often

**romer** – a median bone in the front of the palate that often bears teeth.

## Х

xanthic – yellowish.

xenacanths – a group of fossil, mostly Palaeozoic, sharks phylogenetically positioned on the chondrichthyan stem according to many authors, known for their morphological peculiarity and living mostly in freshwater habitats.

## Y

**yolk-sac placenta** – a vascular placental-like structure in some sharks where the embryonic yolk-sac wall is closely associated with the maternal uterine or oviducal wall, providing nourishment to the embryo.

**yolk-sac viviparity** – producing eggs that hatch within the body of the mother, but the embryos lack a placental attachment to the oviduct.

## Ζ

zona pellucida – see chorion.

**zooplankton** – animals (mostly microscopic) which drift freely in the water column.

## **BIBLIOGRAPHY**

A major source for this bibliography is *Eschmeyer's Catalog* of *Fishes: Genera, species, references* (Fricke R, Eschmeyer WN & Van der Laan R) which is continually updated on a monthly basis. http://researcharchive.calacademy.org/research/ ichthyology/catalog/fishcatmain.asp

Note: Where references have been supplied with a DOI or URL, the latter have been retained as they may be useful to the reader. The absence of a DOI or URL does not imply that the publication is not available online.

## A

- Abbott CC. **1860**. Description of new species of apodal fishes in the museum of the Academy of Natural Sciences of Philadelphia. *Proceedings of the Academy of Natural Sciences of Philadelphia* 12: 475–479.
- Abbott CC. **1861**. Description of a new species of *Exocetus* from Chile. *Proceedings of the Academy of Natural Sciences* of Philadelphia 12 [for 1860]: 472–473.
- Abe T. **1949**. Taxonomic studies on the puffers (Tetraodontidae, Teleostei) from Japan and adjacent regions. V. Synopsis of the puffers from Japan and adjacent regions. *Bulletin of the Biogeographical Society of Japan* 14(13): 89–140.
- Abe T. **1953**. New, rare or uncommon fishes from Japanese waters. II. Records of rare fishes of the families Diretmidae, Luvaridae and Tetragonuridae, with an appendix (description of a new species, *Tetragonurus pacificus*, from off the Solomon Islands). *Japanese Journal of Ichthyology* 3(1): 39–47.
- Abe T. **1954**. *Cypselurus poecilopterus* (Valenciennes) (pp. 984–990). In: Tomiyama I, Abe T (eds) *Figures and descriptions of the fishes of Japan (a continuation of Dr. Shigeho Tanaka's work)* Vol. 50: 983–1011.
- Abe T. 1955. Exocoetus monocirrhus (Richardson), Parexocoetus brachypterus brachypterus (Richardson), Cypselurus cyanopterus (Valenciennes) (pp. 1013–1034).
  In: Tomiyama I, Abe T (eds) Figures and descriptions of the fishes of Japan (a continuation of Dr. Shigeho Tanaka's work) Vol. 51: 1012–1047.
- Abe T. **1955**. On a new Pacific flying fish, *Prognichthys sealei*, retaining five unbranched fin-rays above in the pectoral throughout life. *Records of Oceanographic Works in Japan* 2(1): 185–192.

Abe T. **1956**. *Cypselurus katoptron* (Bleeker), *Hirundichthys speculiger* (Valenciennes), *Hirundichthys oxycephalus* (Bleeker) (pp. 1049–1065). In: Tomiyama I, Abe T (eds) *Figures and descriptions of the fishes of Japan (a continuation of Dr. Shigeho Tanaka's work)* Vol. 52: 1048–1076.

- Abe T. **1956**. *Parexocoetus mento mento* (pp. 1115–1119). In: Tomiyama I, Abe T (eds) *Figures and descriptions of the fishes of Japan (a continuation of Dr. Shigeho Tanaka's work)* Vol. 55: 1115–1140.
- Abe T. **1957**. *Exocoetus volitans* Linnaeus (pp. 1141–1146). In: Tomiyama I, Abe T (eds) *Figures and descriptions of the fishes of Japan (a continuation of Dr. Shigeho Tanaka's work)* Vol. 56: 1141–1169.
- Abe T. **1958**. Two new subspecies of fishes from the path of the "Kuro-Shiwo." *Records of Oceanographic Works in Japan* Special No. 2: 175–180.
- Abe T. **1959**. New, rare or uncommon fishes from Japanese waters. VII. Description of a new species of *Beryx*. *Japanese Journal of Ichthyology* 7(5/6): 157–163.
- Abe T. **1961**. Notes on some fishes of the subfamily Braminae, with the introduction of a new genus, *Pseudotaractes*. *Japanese Journal of Ichthyology* 8(3/4): 92–99.
- Abe T. **1962**. Notes on some fishes of the subfamily Braminae, with the introduction of a new genus, *Pseudotaractes* (continued from 8(3/4): 99). *Japanese Journal of Ichthyology* 8(5/6): 101–114.
- Abe T. **1972**. Commercial fishery of berycoids and other fishes in previously unfished waters between Japan and the Midway Islands (pp. 525–526). In: *The Kuroshio II: Proceedings of the second Symposium on the Results of the Cooperative Study of the Kuroshio and Adjacent Regions [CSK], Tokyo, 28 September–1 October 1970*. Tokyo: Saikon Publishing Co. 562 pp.
- Abe T. **1975**. Notes on some fishes collected by the fisheries research vessel *Kaiyomaru* in the South China Sea. II. *Uo* (Japanese Society of Ichthyologists) 24: 1–6.
- Abe T. **1994**. *Figures and descriptions of the fishes of Japan (a continuation of Dr. Shigeho Tanaka's work)* 60: 1248–1283. Published by T. Abe.
- Abe T, Haneda Y. **1973**. Description of a new fish of the genus *Photoblepharon* (family Anomalopidae) from the Red Sea. *Bulletin of the Sea Fisheries Research Station, Haifa* 60: 57–62.

Abe T, Isokawa S, Misu T, Kishimoto T, Shimma Y, Shimma H. **1968**. Notes on some members of Osteodonti (Class Chondrichthyes). *Bulletin of the Tokai Regional Fisheries Research Laboratory* 56: 1–6.

Abe T, Kobata T. **1974**. Records of the young of the Louvar, *Luvarus imperialis* Rafinesque Schmaltz from Sagami Bay. *Uo* (Japanese Society of Ichthyologists) 20: 1–2.

Abe T, Maruyama K. **1971**. On a young example of *Beryx decadactylus* Cuvier from northern Japan. *Bulletin of the Tokai Regional Fisheries Research Laboratory* 65: 1–4.

Abe T, Shinohara S. **1962**. Description of a new lutianid fish from the Ryukyu Islands. *Japanese Journal of Ichthyology* 9(1–6): 163–171, Pl. 1.

Abe T, Tabeta O. 1983. Description of a new swellfish of the genus *Lagocephalus* (Tetraodontidae, Teleostei) from Japanese waters and the East China Sea. *Uo* (Japanese Society of Ichthyologists) 32: 1–8.

Abel EF. 1960. Zur Kenntnis des Verhaltens und der Ökologie von Fischen an Korrallenriffen bei Ghardaqa (Rotes Meer). Zeitschrift für Morphologie und Ökologie der Tiere 49(4): 430–503.

Able KW. **1973**. A new cyclopterid fish, *Liparis inquilinus*, associated with the sea scallop, *Placopecten magellanicus*, in the western North Atlantic, with notes on the *Liparis liparis* complex. *Copeia* 1973(4): 787–794.

Able KW, McAllister DE. **1980**. Revision of the snailfish genus *Liparis* from Arctic Canada. *Canadian Bulletin of Fisheries and Aquatic Sciences* 208: viii + 52 pp.

Aboim MA, Menezes G, Schlitt T, Rogers AD. **2005**. Genetic structure and history of populations of the deepsea fish *Helicolenus dactylopterus* (Delaroche, 1809) inferred from mtDNA sequence analysis. *Molecular Ecology* 14(5): 1343–1354.

Abou-Seedo F, Wright JM, Clayton DA. **1990**. Aspects of the biology of *Diplodus sargus kotschyi* (Sparidae) from Kuwait Bay. *Cybium* 14(3): 217–223.

Abraham KJ, Joshi KK, Murty VSR. **2011**. Taxonomy of the fishes of the family Leiognathidae (Pisces, Teleostei) from the west coast of India. *Zootaxa* 2886: 1–18. http://dx.doi. org/10.11646/zootaxa.2886.1.1

Abraham RK, Kelkar N, Kumar AB. **2011**. Reply to "Need for further research on the freshwater fish fauna of the Ashambu Hills landscape: a response to Abraham et al." *Journal of Threatened Taxa* 3(5): 1792–1797. http://dx.doi. org/10.11609/JoTT.o2797.1792-7

Abramov AA. **1992**. Species composition and distribution of *Epigonus* (Epigonidae) in the world ocean. *Voprosy Ikhtiologii* 32(2): 17–31. [In Russian. English translation in *Journal of Ichthyology* 32(5): 94–108] Abramov AA, Manilo LG. **1987**. *Epigonus angustifrons* sp. n., a new cardinal fish (Perciformes, Apogonidae) from submarine mountain ridges in the subtropical zone of the Indian Ocean. *Byulleten Moskovskogo Obshchestva Ispytatelei Prirody Otdel Biologicheskii [Bulletin of the Moscow Society of Naturalists Biological Series]* 92(2): 45–48. [In Russian, English summary]

Adam H, Strahan R. 1963. Systematics and geographical distribution of myxinoids (pp. 1–8). In: Brodal A, Fänge R (eds) *The biology of* Myxine. Oslo: Universiteitsforlaget. 588 pp.

Adam MS, Merrett NR, Anderson RC. **1998**. Additions to the fish fauna of the Maldive Islands. Part 1: an annotated checklist of the deep demersal fishes of the Maldive Islands. *Ichthyological Bulletin of the J.L.B. Smith Institute of Ichthyology* 67: 1–19.

Afonso P, Porteiro FM, Santos RS, Barreiros JP, Worms J, Wirtz P. **1999**. Coastal marine fishes of São Tomé Island (Gulf of Guinea). *Arquipélago: Boletim da Universidade dos Açores* 17A: 65–92.

Agafonova TB. **1988**. New data on the taxonomy and distribution of cigarfishes (*Cubiceps*, Nomeidae) of the Indian Ocean. *Journal of Ichthyology* 28(6): 46–61. [Original in Russian in *Voprosy Ikhtiologii* 28(4): 541–555 (Figures 1 & 4)]

Agafonova TB. **1994**. Systematics and distribution of *Cubiceps* (Nomeidae) of the world ocean. *Journal of Ichthyology* 34(5): 116–143. [Original in Russian in *Voprosy Ikhtiologii* 34(2): 161–179]

Agafonova TB, Piotrovskiy AS. **1990**. Validity of the genus *Parapsenes* and new information on the distribution of *P. rotundus* (Nomeidae). *Journal of Ichthyology* 30(4): 135–142.

Agassiz JLR. **1833–1843**. *Recherches sur les poissons fossiles*. Neuchâtel, Switzerland. 5 vols. with atlas.

Agassiz JLR. **1848**. Nomenclator zoologicus index universalis, continens nomina systematica classium, familiarum et generum animalium omnium, tam viventium quam fossilium, secundum ordinem alphabeticum unicum disposita, adjectis homonymus plantarum. Soloduri. x + 1135 pp.

Ahl E. **1923**. Zur Kenntniss der Knochenfischfamilie Chaetodontidae insbesondere der Unterfamilie Chaetodontinae. *Archiv für Naturgeschichte* 89(5): 1–205.

Ahl JN. **1789**. Dissertatio de *Muraena* et *Ophichtho*. In: Thunberg CP (praeses) *Dissertationes Academicae Upsaliae habitae* 3: 1–12.

Ahlstrom EH, Butler JL, Sumida BY. **1976**. Pelagic stromateoid fishes (Pisces, Perciformes) of the eastern Pacific: kinds, distributions, and early life histories and observations on five of these from the northwest Atlantic. *Bulletin of Marine Science* 26(3): 285–402.

Ahmed M, Qureshi MR. **1970**. Fishes of the order Scleroparei: Part II. Families: Synanceidae, Triglidae, Platycephalidae, Caracanthidae, and Dactylopteridae. *Pakistan Journal of Science* 22(3–4): 203–209.

Ajiad AM. **1987**. First record of *Thyrsoidea macrura* (Teleostei: Muraenidae) from the Red Sea. *Cybium* 11(1): 102–103.

Ajiad AM, El-Absy AH. **1986**. First record of *Lycodontis elegans* (Pisces: Muraenidae) from the Red Sea. *Cybium* 10(3): 297–298.

Ajiad AM, Mahasneh DM. **1986**. Redescription of *Ariomma brevimanus* (Klunzinger, 1884), a rare stromateoid from the Gulf of Aqaba (Red Sea). *Cybium* 10(2): 135–142.

Akazaki M. **1962**. Studies on the spariform fishes: anatomy, phylogeny, ecology and taxonomy. *Misaki Marine Biological Institute, Kyoto University, Special Report* 1(1): 1–368.

Akazaki M. 1972. A critical study of the serranid fishes of the genus *Chelidoperca* found in Japan. *Japanese Journal of Ichthyology* 19(4): 174–282.

Akazaki M. 1984. Family Sparidae (sea breams and porgies) (pp. 176–178). In: Masuda H, Amaoka K, Araga C, Uyeno T, Yoshino T (eds) *The fishes of the Japanese archipelago*. Tokyo: Tokai University Press. 437 pp.

Akazaki M. 1994. Comments on the taxonomic status of the sparid fish, Acanthopagrus bifasciatus (pp. 327–337). In: Systematics and evolution of Indo-Pacific fishes: Proceedings, Fourth Indo-Pacific Fish Conference, Bangkok, Thailand, 28 November-4 December 1993. Bangkok: Faculty of Fisheries, Kasetsart University. 502 pp.

Akhilesh KV, Bineesh KK, Shanis CPR, Human BA, Ganga U. **2011**. Rediscovery and description of the quagga shark, *Halaelurus quagga* (Alcock, 1899) (Chondrichthyes: Scyliorhinidae) from the southwest coast of India. *Zootaxa* 2781: 40–48. http://dx.doi.org/10.11646/zootaxa.2781.1.3

Akhilesh KV, Bineesh KK, White WT. **2012**. *Liopropoma randalli*, a new serranid (Teleostei: Perciformes) fish from the Indian Ocean. *Zootaxa* 3439: 43–50. http://dx.doi.org/10.11646/zootaxa.3439.1.2

Akhilesh KV, Bineesh KK, White WT, Shanis CPR, Hashim M, Ganga U, Pillai NGK. **2013**. Catch composition, reproductive biology and diet of the bramble shark *Echinorhinus brucus* (Squaliformes: Echinorhinidae) from the south-eastern Arabian Sea. *Journal of Fish Biology* 83(5): 1112–1127. https://doi.org/10.1111/jfb.12201

Akhilesh KV, Ganga U, Pillai NGK, Vivekanandan E, Bineesh KK, Shanis CPR, Hashim M. 2011. Deep sea fishing for chondrichthyan resources and sustainability concerns—a case study from southwest coast of India. *Indian Journal of Geo-Marine Sciences* 40(3): 347–355. http://eprints.cmfri.org.in/id/eprint/8801

Akhilesh KV, Pillai NGK, Ganga U, Bineesh KK, Rajool Shanis CP, Manjebrayakath H. **2009**. First record of the anthiine fish, *Meganthias filiferus* (Perciformes: Serranidae) from Indian waters. *Marine Biodiversity Records* 2(e113): 1–2. http://dx.doi.org/10.1017/S1755267209001201

Akihito. **1967**. On four species of the gobiid fishes of the genus *Eleotris* found in Japan. *Japanese Journal of Ichthyology* 14(4/6): 135–166.

Akihito. **1972**. On a specimen of "Matsugahaze", *Oxyurichthys ophthalmonema*, collected in Kanagawa Prefecture, Japan. *Japanese Journal of Ichthyology* 19(2): 103–110.

Akihito, Meguro K. 1974. On gobiid fishes Ophiocara porocephala and Ophieleotris aporos. Japanese Journal of Ichthyology 21(2): 72–84.

Akihito, Meguro K. **1975**. Description of a new gobiid fish, *Glossogobius aureus*, with notes on related species of the genus. *Japanese Journal of Ichthyology* 22(3): 127–142.

Akihito, Meguro K. **1975**. On a goby *Redigobius bikolanus*. *Japanese Journal of Ichthyology* 22(1): 49–52.

Akihito, Meguro K. **1977**. Five species of the genus *Callogobius* found in Japan and their relationships. *Japanese Journal of Ichthyology* 24(2): 113–127.

Akihito, Meguro K. **1980**. On the six species of the genus *Bathygobius* found in Japan. *Japanese Journal of Ichthyology* 27(3): 215–236.

Akihito, Meguro K. **1981**. A gobiid fish belonging to the genus *Hetereleotris* collected in Japan. *Japanese Journal of Ichthyology* 28(3): 329–339.

Al-Abdessalaam TZS. **1995**. *Marine species of the Sultanate of Oman: An identification guide*. Sultanate of Oman: Ministry of Agriculture and Fisheries. 412 pp.

Alabsi N, Komatsu T. 2014. Characterization of fisheries management in Yemen: a case study of a developing country's management regime. *Marine Policy* 50: 89–95. http://dx.doi.org/10.1016/j.marpol.2014.05.015

Al-Badri ME, Jawad LA. 2014. New fish records from the marine waters of Iraq. *Cahiers de Biologie Marine* 55(4): 431–436. http://dx.doi.org/10.21411/CBM.A.2A28F6B9

Al-Baharna WS. 1986. Fishes of Bahrain. Bahrain: Directorate of Fisheries, Ministry of Commerce and Agriculture. 294 pp.

Albert JS, Froese R, Bauchot R, Ito H. 1999. Diversity of brain size in fishes: preliminary analysis of a database including 1174 species in 45 orders (pp. 647–656). In: Séret B, Sire J-Y (eds) *Proceedings of the 5th Indo-Pacific Fish Conference, Nouméa, New Caledonia, 3–8 November 1997.* Paris: Société Française d'Ichtyologie. 866 pp.

Alcock AW. **1889**. Natural history notes from H.M. Indian marine survey steamer *Investigator*, Commander Alfred Carpenter, R.N., D.S.O., commanding. No. 10. List of the Pleuronectidae obtained in the Bay of Bengal in 1888 and 1889, with descriptions of new and rare species. *Journal of the Asiatic Society of Bengal* 58(2–3): 279–295.

Alcock AW. **1889**. Natural history notes from H.M. Indian marine survey steamer *Investigator*, Commander Alfred Carpenter, R.N., D.S.O., commanding. No. 12. Descriptions of some new and rare species of fishes from the Bay of Bengal, obtained during the season of 1888–89. *Journal of the Asiatic Society of Bengal* 58(2–3): 296–305.

Alcock AW. **1889**. Natural history notes from H.M. Indian marine survey steamer *Investigator*, Commander Alfred Carpenter, R.N., D.S.O., commanding. No. 13. On the bathybial fishes of the Bay of Bengal and neighbouring waters, obtained during the seasons 1885–1889. *Annals and Magazine of Natural History* (Series 6) 4(23): 376–399.

Alcock AW. 1890. Natural history notes from H.M. Indian marine survey steamer *Investigator*, Commander R.F. Hoskyn, R.N., commanding. No. 16. On the bathybial fishes collected in the Bay of Bengal during the season 1889–1890. *Annals and Magazine of Natural History* (Series 6) 6(33): 197–222.

Alcock AW. 1890. Natural history notes from H.M. Indian marine survey steamer *Investigator*, Commander R.F. Hoskyn, R.N., commanding. No. 20. On some undescribed shore-fishes from the Bay of Bengal. *Annals and Magazine of Natural History* (Series 6) 6(36): 425–443.

Alcock AW. 1891. Class Pisces. In: Wood-Mason J, Alcock AW. Natural history notes from H.M. Indian marine survey steamer *Investigator*, Commander R.F. Hoskyn, R.N., commanding. Series II, No. 1. On the results of deep-sea dredging during the season 1890–91. *Annals and Magazine* of *Natural History* (Series 6) 8(43; 44): 16–34; 119–138.

Alcock AW. 1892. Natural history notes from H.M. Indian marine survey steamer *Investigator*, Lieut. G.S. Gunn, R.N., commanding. Series II, No. 4. Some observations on the embryonic history of *Pteroplataea micrura*. *Annals and Magazine of Natural History* (Series 6) 10(55): 1–8.

Alcock AW. 1892. Natural history notes from H.M. Indian marine survey steamer *Investigator*, Lieut. G.S. Gunn, R.N., commanding. Series II, No. 6. A case of commensalism between a gymnoblastic anthomedusoid (*Stylactis minoi*), and a scorpaenid fish (*Minous inermis*). *Annals and Magazine of Natural History* (Series 6) 10(57): 207–214.

Alcock AW. 1892. Natural history notes from H.M. Indian marine survey steamer *Investigator*, Lieut. G.S. Gunn, R.N., commanding. Series II, No. 5. On the bathybial fishes collected during the season of 1891–92. *Annals and Magazine of Natural History* (Series 6) 10(59): 345–365.

Alcock AW. **1893**. New species of *Lophius*, *Physiculus*, *Neobythites*, *Odontostomus*, and *Congromuraena*. *Journal of the Asiatic Society of Bengal* 62(2): 177–184.

Alcock AW. 1894. Natural history notes from H.M. Indian marine survey steamer *Investigator*, Commander C.F.
Oldham, R.N., commanding. Series II, No. 9. An account of the deep sea collection made during the season of 1892–93. *Journal of the Asiatic Society of Bengal* 62(2): 169–184.

Alcock AW. 1894. Natural history notes from H.M. Indian marine survey steamer *Investigator*, Commander C.F.
Oldham, R.N., commanding. Series II, No. 11. An account of a recent collection of bathybial fishes from the Bay of Bengal and from the Laccadive Sea. *Journal of the Asiatic Society of Bengal* 63(2): 115–137.

Alcock AW. **1894**. Illustrations of the zoology of the Royal Indian marine surveying steamer *Investigator*, ... Fishes. *Illustrations of the Zoology of the Royal Indian Marine Surveying Steamer Investigator*. Part 2. Calcutta. Pls. 8–13.

Alcock AW. 1895. Natural history notes from H.M. Indian marine survey steamer *Investigator*, Commander C.F. Oldham, R.N., commanding. Series II, No. 18. On a new species of viviparous fish of the family Ophidiidæ. *Annals and Magazine of Natural History* (Series 6) 16(92): 144–146.

Alcock AW. **1895**. Illustrations of the zoology of the Royal Indian marine surveying steamer *Investigator*, ... Fishes. *Illustrations of the Zoology of the Royal Indian Marine Surveying Steamer Investigator*. Part 3. Calcutta. Pls. 14–16.

Alcock AW. **1896**. Natural history notes from H.M. Indian marine survey steamer *Investigator*, Commander C.F. Oldham, R.N., commanding. Series II, No. 23. A supplementary list of the marine fishes of India, with descriptions of two new genera and eight new species. *Journal of the Asiatic Society of Bengal* 65(2–3): 301–338.

Alcock AW. **1897**. Illustrations of the zoology of the Royal Indian marine surveying steamer *Investigator*, ... Fishes. *Illustrations of the Zoology of the Royal Indian Marine Surveying Steamer Investigator*. Part 4. Calcutta. Pl. 17.

Alcock AW. 1898. Natural history notes from H.M. Indian marine survey ship *Investigator*, Commander T.H. Heming, R.N., commanding. Series II, No. 25. A note on the deep-sea fishes, with descriptions of some new genera and species, including another probably viviparous ophidioid. *Annals* and Magazine of Natural History (Series 7) 2(8): 136–156.

Alcock AW. 1898. Illustrations of the zoology of H.M.
Indian marine surveying steamer *Investigator*, ... Fishes. *Illustrations of the Zoology of the Royal Indian Marine Surveying Steamer Investigator*. Part 5. Calcutta. Pls. 18–24
[also cited as Alcock & Anderson 1898].

Alcock AW. **1899**. *Halimochirurgus centriscoides*, a new deepsea fish from the Gulf of Manár. *Proceedings of the Asiatic Society of Bengal* 1899: 78. Alcock AW. **1899**. *A descriptive catalogue of the Indian deep-sea fishes in the Indian Museum. Being a revised account of the deep-sea fishes collected by the Royal Indian marine survey ship* Investigator. Calcutta: Trustees of the Indian Museum. *i*–*iii* + 1–211 + *i*–*viii* pp.

Alcock AW. **1899**. Illustrations of the zoology of the Royal Indian marine surveying steamer *Investigator*, ... Fishes. *Illustrations of the zoology of the Royal Indian marine surveying steamer* Investigator. Part 6. Calcutta. Pls. 25–26.

Alcock AW. **1900**. Illustrations of the zoology of the Royal Indian marine surveying steamer *Investigator*, ... Fishes. *Illustrations of the zoology of the Royal Indian marine surveying steamer* Investigator. Part 7. Calcutta. Pls. 27–35.

Alcock AW. **1902**. A naturalist in Indian seas, or, four years with the Royal Indian Marine Survey ship Investigator. London: John Murray. 328 pp.

Alcock AW, Anderson ARS. 1898. See Alcock AW. 1898.

Al-Hassan JM, Clayton DA, Thomson M, Criddle RS. 1987. Taxonomy and distribution of ariid catfishes from the Arabian Gulf. *Journal of Natural History* 22(2): 473–487.

Al-Hassan LAJ, Miller PJ. **1987**. *Rhinogobius brunneus* (Gobiidae) in the Persian Gulf. *Japanese Journal of Ichthyology* 33(4): 405–408.

Al-Hussaini AH. **1947**. The feeding habits and the morphology of the alimentary tract of some teleost living in the neighbourhood of the Marine Biological Station, Ghardaqa, Red Sea. *Publications of the Marine Biological Station Al-Ghardaqa* (Red Sea) 5: 1–61.

Ali A, Jawad LA, Sheikh A. **2009**. First record of *Neoharriotta pinnata* (Condrichthys: Rhinochimaeridae) and second record of *Satyrichthys adeni* (Osteichthys: Peristediidae) from the Gulf of Aden, Republic of Yemen. *Marine Biodiversity Records* 2(e170): 1–4. http://dx.doi.org/10.1017/ S1755267209990972

Ali AH, Abed JM, Taher MM. 2014. First record of saddleback silver-biddy *Gerres limbatus* Cuvier, 1830 (Pisces: Gerreidae) from Shatt Al-Arab River and marine territorial waters of Iraq. *International Journal of Marine Science* 4(No. 59): 1–5.

Al-Jufaili SM, Hermosa G, Al-Shuaily SS, Al Mujaini A. 2010. Oman fish biodiversity. *JKAU* [Journal of King Abdulaziz University] *Marine Sciences* 21(1): 3–51. http://dx.doi. org/10.4197/Mar.21-1.1

Allain V, Lorance P. 2000. Age estimation and growth of some deep-sea fish from the northeast Atlantic Ocean. *Cybium* 24(3) Supplement: 7–16.

Allen BR, Wintner SP. **2002**. Age and growth of the spinner shark *Carcharhinus brevipinna* (Müller and Henle, 1839) off the KwaZulu-Natal coast, South Africa. *South African Journal of Marine Science* 24: 1–8.

Allen GR. 1972. The anemonefishes. Their classification and biology. Neptune City, New Jersey: TFH Publications. 288 pp. Allen GR. **1973**. *Bodianus bimaculatus*, a new species of wrasse (Pisces: Labridae) from the Palau Archipelago. *Proceedings of the Biological Society of Washington* 86(32): 385–389.

Allen GR. **1975**. *Damselfishes of the South Seas*. Neptune City, New Jersey: TFH Publications. 240 pp.

Allen GR. **1975**. *The anemonefishes. Their classification and biology* (2<sup>nd</sup> edition). Neptune City, New Jersey: TFH Publications. 352 pp.

Allen GR. **1978**. A review of the archerfishes (family Toxotidae). *Records of the Western Australian Museum* 6(4): 355–378.

Allen GR. **1980**. *Butterfly and angelfishes of the world*. Vol. 2. New York: John Wiley & Sons. 208 pp (pp. 145–352). [also cited as 1979]

Allen GR. **1984**. A new genus and species of anthiid fish from Papua New Guinea. *Revue française d'Aquariologie Herpétologie* 11(2): 47–50.

Allen GR. **1984**. Lutjanidae. Snappers, jobfishes. In: Fischer W, Bianchi G (eds) *FAO species identification sheets for fishery purposes. Western Indian Ocean (Fishing Area 51)*. Vol. 3. Rome: FAO. [unpaginated]

Allen GR. **1985**. *FAO Species Catalogue. Snappers of the world. An annotated and illustrated catalogue of lutjanid species known to date.* Vol. 6. FAO Fisheries Synopsis. Rome: FAO. 208 pp.

Allen GR. **1986**. *Appendix: Butterfly and angelfishes of the world*. Vol. 2. Melle, Germany: Mergus Publishers. 7 pp.

Allen GR. **1987**. New Australian fishes. Part 2. Four new species of Apogonidae. *Memoirs of the Museum of Victoria* 48(1): 3–8.

Allen GR. **1991**. *Damselfishes of the world*. Melle, Germany: Mergus Publishers. 271 pp.

Allen GR. **1997**. *Marine fishes of tropical Australia and southeast Asia*. Perth: Western Australian Museum. 292 pp.

Allen GR. 1999. Ambassidae (= Chandidae). Perchlets, glassfishes (pp. 2433–2435). In: Carpenter K, Niem VH (eds) *FAO species identification guide for fishery purposes. The living marine resources of the western central Pacific.* Vol. 4. Bony fishes part 2 (pp. 2069–2790). Rome: FAO.

Allen GR. **2001**. Description of two new gobies (*Eviota*, Gobiidae) from Indonesian seas. *aqua, Journal of Ichthyology and Aquatic Biology* 4(4): 125–130.

Allen GR. 2002. Descriptions of two new species of damselfishes (Pomacentridae: *Pomacentrus*) from Madagascar. *aqua, Journal of Ichthyology and Aquatic Biology* 6(2): 45–52.

Allen GR. 2005. Reef fishes of northwest Madagascar (pp. 39–48). In: McKenna SA, Allen GR (eds) A rapid marine biodiversity assessment of the coral reefs of northwest Madagascar. *Bulletin of the Rapid Assessment Program* 31. Washington, DC: Conservation International. 124 pp.

Allen GR. **2018**. *Chromis torquata*, a new species of damselfish (Pomacentridae) from Mauritius and Réunion. *aqua*, *International Journal of Ichthyology* 24(1): 15–22.

Allen GR, Adrim M. **2003**. Coral reef fishes of Indonesia. *Zoological Studies* 42(1): 1–72.

Allen GR, Burgess WE. 1990. A review of the glassfishes (Chandidae) of Australia and New Guinea. *Records of the Western Australian Museum* Supplement No. 34: 139–206.

Allen GR, Cliff G. **2000**. Sharks caught in the protective gill nets off KwaZulu-Natal, South Africa. 9. The spinner shark *Carcharhinus brevipinna* (Müller and Henle). *South African Journal of Marine Science* 22: 199–215.

Allen GR, Cross NJ, Allen CJ, Gomon MF. 2006. Labridae.
Labrinae (pp. 1368–1418). In: Hoese DF, Bray DJ, Paxton JR, Allen GR (eds) *Zoological catalogue of Australia*. Vol. 35. Fishes. Part 2 (pp. 671–1472). Collingwood: CSIRO Publishing.

Allen GR, Dudgeon CL. **2010**. *Hemiscyllium michaeli*, a new species of bamboo shark (Hemiscyllidae) from Papua New Guinea. *aqua, International Journal of Ichthyology* 16(1): 19–30.

Allen GR, Emery AR. **1985**. A review of the pomacentrid fishes of the genus *Stegastes* from the Indo-Pacific, with descriptions of two new species. *Indo-Pacific Fishes* 3: 1–31.

Allen GR, Erdmann MV. **2005**. *Chromis xouthos*, a new species of damselfish (Pomacentridae) from the East Andaman Sea and central Indian Ocean. *aqua, Journal of Ichthyology and Aquatic Biology* 10(3): 89–94.

Allen GR, Erdmann MV. **2006**. *Pterocaesio flavifasciata*, a new species of fusilier (Telostei: Caesionidae) from Sumatra, Indonesia. *aqua, International Journal of Ichthyology* 12(1): 27–30.

Allen GR, Erdmann MV. **2008**. *Pterois andover*, a new species of scorpionfish (Pisces: Scorpaenidae) from Indonesia and Papua New Guinea. *aqua, International Journal of Ichthyology* 13(3–4): 127–138.

Allen GR, Erdmann MV. **2008**. Two new species of bamboo sharks (Orectolobiformes: Hemiscyllidae) from western New Guinea. *aqua, International Journal of Ichthyology* 13(3–4): 93–108.

Allen GR, Erdmann MV. **2012**. A new species of dragonet (*Synchiropus*: Callionymidae) from Indonesia. *aqua*, *International Journal of Ichthyology* 18(1): 9–14.

Allen GR, Erdmann MV. **2012**. *Reef fishes of the East Indies: Andaman Sea, Myanmar, Thailand, Indonesia, Christmas Island, Singapore, Malaysia, Brunei, Philippines, Papua New Guinea, Solomon Islands* (3 Vols). Perth, Australia: Tropical Reef Research. 1260 pp.

Allen GR, Erdmann MV. **2020**. *Chromis pacifica*, a new Pacific Ocean damselfish distinct from Indian Ocean *Chromis agilis* (Teleostei: Pomacentridae). *Journal of the Ocean Science Foundation* 35: 102–117. https://doi.org/10.5281/ zenodo.3988552 Allen GR, Hoese D, Paxton JR, Randall JE, Russell BC, Starck WA II, Talbot FH, Whitley GP. **1976**. Annotated checklist of the fishes of Lord Howe Island. *Records of the Australian Museum* 30(15): 365–454. https://doi.org/10.385 3/j.0067-1975.30.1976.287

Allen GR, Kuiter RH. **1976**. A review of the plesiopid fish genus *Assessor*, with descriptions of two new species. *Records of the Western Australian Museum* 4(3): 201–215.

Allen GR, Kuiter RH. **1978**. *Heniochus diphreutes* Jordan, a valid species of butterfly fish (Chaetodontidae) from the Indo-west Pacific. *Journal of the Royal Society of Western Australia* 61(1): 11–18.

Allen GR, Kuiter RH. **1994**. Descriptions of two news [sic] species of cardinalfishes (Apogonidae) from Malaysia and Indonesia. *Revue française d'Aquariologie Herpétologie* 21(1/2): 19–23.

 Allen GR, Kuiter RH, Randall JE. 1994. Descriptions of five new species of cardinalfishes (Apogonidae: *Apogon*) from Maumere Bay, Flores, Indonesia and surrounding regions. *Revue française d'Aquariologie Herpétologie* 21(1/2): 27–38.

Allen GR, Midgley SH, Allen M. 2002. Field guide to the freshwater fishes of Australia. Perth: Western Australian Museum. xiv + 394 pp.

Allen GR, Moyer JT. **1980**. *Ellerkeldia wilsoni*, a new species of serranid fish from southwestern Australia. *Japanese Journal of Ichthyology* 26(4): 329–333.

Allen GR, Rajasuriya A. **1995**. *Chrysiptera kuiteri*, a new species of damselfish (Pomacentridae) from Indonesia and Sri Lanka. *Records of the Western Australian Museum* 17(3): 283–286.

Allen GR, Randall JE. **1974**. Five new species and a new genus of damselfishes (family Pomacentridae) from the South Pacific Ocean. *Tropical Fish Hobbyist* 22(9): 36–46, 48–49.

Allen GR, Randall JE. **1977**. Review of the sharpnose pufferfishes (subfamily Canthigasterinae) of the Indo-Pacific. *Records of the Australian Museum* 30(17): 475–517.

Allen GR, Randall JE. **1981**. A review of the damselfishes (Teleostei: Pomacentridae) of the Red Sea. *Israel Journal of Zoology* 29(1/3) [for 1980]: 1–98.

Allen GR, Randall JE. **1994**. A new species of cardinalfish (*Apogon*: Apogonidae) from Arabian Seas. *Revue française d'Aquariologie Herpétologie* 21(1/2): 24–26.

Allen GR, Randall JE. **2002**. A review of the *leucogaster* species complex of the Indo-Pacific pomacentrid genus *Amblyglyphidodon*, with descriptions of two new species. *aqua, Journal of Ichthyology and Aquatic Biology* 5(4): 139–152.

Allen GR, Russell BC, Carlson BA, Starck WA II. **1975**. Mimicry in marine fishes. *Tropical Fish Hobbyist* 25: 47–56.

Allen GR, Smith-Vaniz WF. **1994**. Chapter 14. Fishes of the Cocos (Keeling) Islands. *Atoll Research Bulletin* 412: 1–21.

Allen GR, Starck WA II. **1973**. Notes on the ecology, zoogeography, and coloration of the gobiesocid clingfishes, *Lepadichthys caritus* Briggs and *Diademichthys lineatus* (Sauvage). *Proceedings of the Linnean Society of New South Wales* 98(2): 95–97.

Allen GR, Starck WA II. **1982**. The anthiid fishes of the Great Barrier Reef, Australia, with the description of a new species. *Revue française d'Aquariologie Herpétologie* 9(2):47–56.

Allen GR, Steene RC. **1987**. *Reef fishes of the Indian Ocean*. *Book 10. Pacific Marine Fishes*. Neptune City, New Jersey: TFH Publications. 240 pp.

Allen GR, Steene RC. 1988. Fishes of Christmas Island Indian Ocean. Christmas Island: Christmas Island Natural History Association. 197 pp.

Allen GR, Steene R. **1994**. *Indo-Pacific coral reef fish guide*. Singapore: Tropical Reef Research. v + 378 pp.

Allen GR, Steene R, Allen M. **1998**. A guide to angelfishes & butterflyfishes. Singapore: Tropical Reef Research. 250 pp.

Allen GR, Steene R, Humann P, DeLoach N. **2003**. *Reef fish identification: Tropical Pacific.* Jacksonville, Florida: New World Publications. 480 pp.

Allen GR, Swainston R. **1988**. *The marine fishes of northwestern Australia. A field guide for anglers and divers.* Perth: Western Australian Museum. vi + 201 pp.

Allen GR, Talbot FH. **1985**. Review of the snappers of the genus *Lutjanus* (Pisces: Lutjanidae) from the Indo-Pacific, with the description of a new species. *Indo-Pacific Fishes* 11: 1–87, Pls. 1–10.

Allen GR, White WT, Erdmann MV. **2013**. Two new species of snappers (Pisces: Lutjanidae: *Lutjanus*) from the Indo-West Pacific. *Journal of the Ocean Science Foundation* 6: 33–51.

Allen GR, Wright JE. **2003**. Description of a new species of damselfish (Pomacentridae: *Pomacentrus*) from Rodrigues Island, Indian Ocean. *aqua, Journal of Ichthyology and Aquatic Biology* 7(4): 133–138.

Alleyne HG, Macleay W. **1877**. The ichthyology of the *Chevert* Expedition. *Proceedings of the Linnean Society of New South Wales* 1(3–4): 261–281, 321–359.

Almeida AJ, Amoedo L, Saldanha L. 2001. Fish assemblages in the seagrass beds at Inhaca Island (Mozambique): cold season. *Boletim do Museu Municipal do Funchal*, Supplement. 6: 111–125.

Almeida AJ, Marques A, Saldanha L. **1999**. Some aspects of the biology of three fish species from the seagrass beds at Inhaca island, Mozambique. *Cybium* 23(4): 369–376.

Al Sakaff H, Esseen M. **1997**. *The artisanal fisheries of the Red Sea and the Gulf of Aden*. Fourth Fisheries Development Project. MEP-YE-105. Report to the European Commission (ALA/91/22) and to the Ministry of Fish Wealth, Government of the Republic of Yemen. Aden, Yemen: MacAlister, Elliott and Partners. Al Sakaff H, Esseen M. **1999**. Occurrence and distribution of fish species off Yemen (Gulf of Aden and Arabian Sea). *Naga, The ICLARM Quarterly* 22(1): 43–47.

Al-Shuaily S, Henderson AC. **2003**. Survey, status and utilization of the elasmobranch fisheries resources of the Sultanate of Oman. Species identification guide. Sultan Qaboos University, Muscat. 81 pp.

Alvarez MC, Thode G, Cano J. 1983. Somatic karyotypes of two Mediterranean teleost species: *Phycis phycis* (Gadidae) and *Epinephelus alexandrinus* (Serranidae). *Cytobios* 38(150): 91–95.

Amaoka K. **1963**. A revision of the flatfish referable to the genus *Psettina* found in the waters around Japan. *Bulletin of the Misaki Marine Biological Institute* (Kyoto University) 4: 53–62.

Amaoka K. **1964**. Development and growth of the sinistral flounder, *Bothus myriaster* (Temminck and Schlegel) found in the Indian and Pacific Oceans. *Bulletin of the Misaki Marine Biological Institute* (Kyoto University) 5: 11–29.

Amaoka K. **1969**. Studies on the sinistral flounders found in the waters around Japan — taxonomy, anatomy and phylogeny. *Journal of the Shimonoseki University of Fishes* 18(2): 65–340.

Amaoka K. 1970. Studies on the larvae and juveniles of the sinistral flounders. I. *Taeniopsetta ocellata* (Günther). *Japanese Journal of Ichthyology* 17(3): 95–104.

Amaoka K. **1971**. Studies on the larvae and juveniles of the sinistral flounders. II. *Chascanopseita lugubris*. *Japanese Journal of Ichthyology* 18(1): 25–32.

Amaoka K. **1972**. Osteology and relationships of the citharid flatfish *Brachypleura novaezeelandiae*. *Japanese Journal of Ichthyology* 19(4): 263–273.

Amaoka K. **1972**. Studies on the larvae and juveniles of the sinistral flounders. III. *Laeops kitaharae*. *Japanese Journal of Ichthyology* 19(3): 154–165.

Amaoka K. 1973. Studies on the larvae and juveniles of the sinistral flounders. IV. Arnoglossus japonicus. Japanese Journal of Ichthyology 20(3): 145–156.

Amaoka K. **2016**. *Flatfishes of Japan (Citharidae, Paralichthyidae, Bothidae, Pleuronectidae, Poecilopsettidae, Samaridae)*. Hiratsuka: Tokai University Press. 229 pp. [In Japanese]

Amaoka K, Ho H-C. **2018**. Review of the genus *Engyprosopon* Günther, 1862 (Pleuronectiformes: Bothidae) from waters off Taiwan, with descriptions of two new species. *Zootaxa* 4413(3): 449–481. http://dx.doi.org/10.11646/ zootaxa.4413.3.2

Amaoka K, Imamura H. 1990. Two new and one rare species of bothid flounders from Saya de Malha Bank, Indian Ocean (Teleostei: Pleuronectiformes). *Copeia* 1990(4): 1011–1019. Amaoka K, Kanayama T. 1981. Additional specimens of *Minous longimanus* from the western Indian Ocean, distinct from *M. inermis. Japanese Journal of Ichthyology* 27(4): 330–332.

Amaoka K, Last PR. **2014**. The Australian sinistral flounder *Arnoglossus aspilos praeteritus* (Actinopterygii, Pleuronectiformes: Bothidae) reassigned as a valid species of *Engyprosopon. Species Diversity* 19: 91–96. http://dx.doi. org/10.12782/sd.19.2.091

Amaoka K, Matsuura K, Inada T, Takeda M, Hatanaka H, Okada K (eds). **1990**. *Fishes collected by the R/V* Shinkai Maru *around New Zealand*. Japan Marine Fishery Resource Research Center. 410 pp.

Amaoka K, Mihara E, Rivaton J. 1993. Pisces,
Pleuronectiformes: flatfishes from the waters around New Caledonia. A revision of the genus *Engyprosopon*. In:
Crosnier A (ed) Résultats Campagnes MUSORSTOM,
Vol. 11. *Mémoires du Muséum National d'Histoire Naturelle*(Paris) (N.S., Série A, Zoologie) 158: 377–426.

Amaoka K, Mihara E, Rivaton J. 1997. Pisces,
Pleuronectiformes: flatfishes from the waters around New Caledonia. Six species of the bothid genera *Tosarhombus* and *Parabothus*. In: Séret B (ed) Résultats des Campagnes MUSORSTOM, Vol. 17. *Mémoires du Muséum National d'Histoire Naturelle* (N.S., Série A, Zoologie) 174: 143–172.

Amaoka K, Nishikawa S, Tanaka N. **1974**. Sexual dimorphism and an abnormal intersexual specimen in the bothid flounder *Bothus pantherinus*. *Japanese Journal of Ichthyology* 21: 16–20.

Amaoka K, Rivaton J. 1991. Pisces: Pleuronectiformes: a review of the genus *Tosarhombus* (Bothidae) with descriptions of two new species from Saya de Malha Bank (Indian Ocean) and the Chesterfield Islands (Coral Sea). In: Crosnier A (ed) Résultats Campagnes MUSORSTOM, Vol. 8. *Mémoires du Muséum National d'Histoire Naturelle* (Série A, Zoologie) 151: 449–466.

Amaoka K, Senou H, Ono A. **1994**. Record of the bothid flounder *Asterorhombus fijiensis* from the western Pacific, with observations of the use of the first dorsal-fin ray as a lure. *Japanese Journal of Ichthyology* 41(1): 23–28.

Amaoka K, Yamamoto E. **1984**. Review of the genus *Chascanopsetta*, with the description of a new species. *Bulletin of the Faculty of Fisheries Hokkaido University* 35(4): 201–224.

Amaoka K, Hatanaka H, Ida H, Ikeda I, Inada T, Iwai T, Kawahara S, Kono H, Kuronuma K, Matsuura K, Miyake S, Nakamura I, Ochiai A, Okamura O, Okutani T, Sasaki T, Sato T(Tetsuya), Sato T(Torao), Shimizu T, Takeda M, Taniuchi T, Tominaga Y, Uyeno T (eds). 1976. Colored illustrations of bottom fishes collected by Japanese trawlers. Vol. II. Tokyo: Japan Deep Sea Trawlers Association. 188 pp. Amir SA, Siddiqi PJA, Masroor R. **2014**. A new sparid fish of genus *Sparidentex* (Perciformes: Sparidae) from coastal waters of Pakistan (North Western Indian Ocean). *Pakistan Journal of Zoology* 46(2): 471–477.

Amir SA, Tanaka F, Siddiqui PJ, Iwatsuki Y. **2013**. First records of two sparid species, *Diplodus omanensis* and *Pagellus affinis* (Perciformes: Sparidae) from western coast of Pakistan. *Cybium* 37(3): 220–222. https://doi.org/10.26028/ cybium/2013-373-010

Amirthalingam C. **1969**. A new fish from the Red Sea. *Sudan Notes and Records* 50: 129–133, Pls. 1–3.

Anderson AW. **1954**. 152-year old lake sturgeon caught in Ontario. *Commercial Fisheries Review* 18: 28.

Anderson C. **1996**. *Common reef fishes of Sri Lanka*. Colombo: Wildlife Heritage Trust of Sri Lanka. 80 pp.

Anderson ME. 1992. A new sandperch, Parapercis maritzi (Teleostei: Pinguipedidae), from South Africa. South African Journal of Zoology 27(4): 151–155.

Anderson ME. **1994**. Systematics and osteology of the Zoarcidae (Teleostei: Perciformes). *Ichthyological Bulletin of the J.L.B. Smith Institute of Ichthyology* 60: 1–120.

Anderson ME. **2005**. Three new species of *Microbrotula* (Teleostei: Ophidiiformes; Bythitidae) from the Indo-West Pacific. *Zootaxa* 1006: 33–42.

Anderson ME, Heemstra PC. **2003**. Review of the glassfishes (Perciformes: Ambassidae) of the western Indian Ocean. *Cybium* 27(3): 199–209.

Anderson ME, Stein DL, Detrich HW. **2005**. Additions to the ichthyofauna of the Tristan da Cunha Group, South Atlantic Ocean. *Zootaxa* 1072: 27–33.

Anderson OF, Bagley NW, Hurst RJ, Francis MP, Clark MR, McMillan PJ. 1998. Atlas of New Zealand fish and squid distributions from research bottom trawls. *National Institute* of Water and Atmospheric Research (NIWA) Technical Report 42: 1–12.

Anderson RC. **2005**. *Reef fishes of the Maldives*. Malé, Maldives: Manta Marine. 130 pp.

Anderson RC, Ahmed H. **1993**. *The shark fisheries of the Maldives*. Malé, Maldives: Ministry of Fisheries and Agriculture, Maldives, and FAO. 76 pp.

Anderson RC, Randall JE, Kuiter RH. **1998**. Additions to the fish fauna of the Maldive Islands. Part 2: New records of fishes from the Maldive Islands, with notes on other species. *Ichthyological Bulletin of the J.L.B. Smith Institute of Ichthyology* 67: 20–32.

Anderson WD Jr. **1981**. A new species of Indo-West Pacific *Etelis* (Pisces: Lutjanidae), with comments on other species of the genus. *Copeia* 1981(4): 820–825.

Anderson WD Jr, Allen GR. 1986. Family No. 181: Lutjanidae.In: Smith MM, Heemstra PC (eds) *Smiths' sea fishes*.Johannesburg: Macmillan South Africa. 1047 pp.

Anderson WD Jr, Allen GR. **2001**. Family Lutjanidae. Snappers (jobfishes) (pp. 2840–2918). In: Carpenter KE, Niem VH (eds) *FAO* species identification guide for fishery purposes. *The living marine resources of the western central Pacific*. Vol. 5: Bony fishes part 3 (pp. 2791–3380). Rome: FAO.

Anderson WD Jr, Baranes A, Goren M. 2011.
Redescription of the perciform fish *Symphysanodon disii* (Symphysanodontidae) from the Gulf of Aqaba, Red
Sea, with comments on *S. pitondelafournaisei* and sexual dimorphism in the genus. *Zootaxa* 3027: 1–8. http://dx.doi. org/10.11646/zootaxa.3027.1.1

Anderson WD Jr, Bineesh KK. 2011. A new species of the perciform fish genus *Symphysanodon* (Symphysanodontidae) from the Arabian sea off the southwestern coast of India. *Zootaxa* 2966: 31–36. http://dx.doi.org/10.11646/zootaxa.2966.1.3

Anderson WD Jr, Chesalin MV, Jawad LA, Al Shajibi SR. **2015**. Redescription of the percoid fish *Symphysanodon andersoni* Kotthaus (Symphysanodontidae) from the northwestern Indian Ocean, based on the holotype and the second known specimen. *Zootaxa* 4021(3): 475–481. http://dx.doi. org/10.11646/zootaxa.4021.3.7

Anderson WD Jr, Heemstra PC. **1989**. *Ellerkeldia*, a junior synonym of *Hypoplectrodes*, with redescriptions of the type species of the genera (Pisces: Serranidae: Anthiinae). *Proceedings of the Biological Society of Washington* 102(4): 1001–1017.

 Anderson WD Jr, Johnson GD, Baldwin CC. 2015. Review of the splendid perches, *Callanthias* Lowe, 1839. *Transactions* of the American Philosophical Society 105(3). Philadelphia: American Philosophical Society Press. xxii + 126 pp.

Anderson WD Jr, Kailola PJ, Collette BB. **1992**. Two new snappers (Teleostei: Lutjanidae: Apsilinae): *Paracaesio paragrapsimodon* Anderson and Kailola from the western Pacific and *P. waltervadi* Anderson and Collette from the western Indian Ocean. *Proceedings of the Biological Society of Washington* 105(3): 443–461.

Anderson WD Jr, Springer VG. **2005**. Review of the perciform fish genus *Symphysanodon* Bleeker (Symphysanodontidae), with descriptions of three new species, *S. mona*, *S. parini*, and *S. rhax. Zootaxa* 996: 1–44.

Anderson WD Jr, Talwar PK, Johnson GD. 1977. A replacement name for *Tangia* Chan (Pisces: Perciformes: Lutjanidae) with redescriptions of the genus and typespecies. *Proceedings of the Biological Society of Washington* 89(44): 509–517.

Andreev PS, Coates MI, Karatajūtė-Talimaa V, Shelton RM, Cooper PR, Wang N-Z, Sansom IJ. **2016**. The systematics of the Mongolepidida (Chondrichthyes) and the Ordovician origin of the clade. *PeerJ* 4: e1850. http://dx.doi.org/10.7717/ peerj.1850 Andreev PS, Zhao W, Wang N-Z, Smith MM, Li Q, Cui X, Zhu M, Sansom IJ. **2020**. Early Silurian chondrichthyans from the Tarim Basin (Xinjiang, China). *PLoS ONE* 15(2): e0228589. https://doi.org/10.1371/journal.pone.0228589

Andrew T[G], Hecht T. **1992**. Feeding biology of *Acantholatris monodactylus* (Pisces) at Tristan da Cunha and Gough Island, South Atlantic. *South African Journal of Antarctic Research* 22: 41–49.

Andrew TG, Hecht T, Heemstra PC, Lutjeharms JRE. **1995**. Fishes of the Tristan da Cunha Group and Gough Island, south Atlantic Ocean. *Ichthyological Bulletin of the J.L.B. Smith Institute of Ichthyology* 63: 1–43.

Andriashev AP, Stein DL. 1998. Review of the snailfish genus *Careproctus* (Liparidae, Scorpaeniformes) in Antarctic and adjacent waters. *Contributions in Science* (Natural History Museum of Los Angeles County) 470: 1–63.

Angel F, Bertin L, Guibé J. **1946**. Note relative à la nomenclature d'un amphibien et d'un poisson. *Bulletin du Muséum National d'Histoire Naturelle* (Série 2) 18(6): 473–474.

Angot M. **1950**. Poissons Littoraux de Soalara. Carte des Fonds. — Systematique Observations Biologiques Possibilites de Pesche Europenne. *Memoires de l'Institut Scientifique de Madagascar*. Serie A, Tome IV, Fascicule 1: 175–196.

Annandale N. **1908**. A new sting ray of the genus *Trygon* from the Bay of Bengal. *Records of the Indian Museum* (Calcutta) 2(4): 393–394.

Annandale N. **1909**. Report on the fishes taken by the Bengal Fisheries Steamer *Golden Crown*. Part I. Batoidei. *Memoirs of the Indian Museum* 2(1): 1–58.

Annandale N. **1910**. Report on the fishes taken by the Bengal Fisheries Steamer *Golden Crown*. Part II. Additional notes on the Batoidei. *Memoirs of the Indian Museum* 3(1): 1–5.

Annandale N, Hora SL. **1925**. The freshwater fish from the Andaman Islands. *Records of the Indian Museum* (Calcutta) 27(2): 33–41.

Annandale N, Jenkins JT. **1910**. Report on the fishes taken by the Bengal Fisheries Steamer *Golden Crown*. Part III.
Plectognathi and Pediculati. *Memoirs of the Indian Museum* 3(1): 7–21.

Anonymous. **1798**. Latinisation de la "Raie Thouin" Lacepède, Histoire naturelle des Poissons, 1798. *Allgemeine Literatur-Zeitung* 3(287): 677 and 3(288): 685.

Anonymous. **1798**. Review of "Histoire naturelle des poissons" by La Cepède. *Allgemeine Literatur-Zeitung* 3(288): 681–682.

Anonymous. **1897**. *Report of the marine biologist for the year 1896*. Cape Town: WA Richardson and Sons.

Anonymous. **1955**. *Marine fishes of Karachi and the coasts of Sind and Makran*. Karachi, Pakistan: The Manager of Publications, Central Fisheries Department. 80 pp.

Anonymous (Bennett ET). 1830. Class Pisces (pp. 686–694). In: Raffles [Lady] S, Memoir of the life and public services of Sir Thomas Stamford Raffles. Particularly in the Government of Java, 1811–1816, and of Bencoolen and its dependencies, 1817–1824. London: John Murray. 694 pp.

Anthonipillai AM, see Mary AA.

Aoyagi H. **1943**. *Coral fishes. Part 1*. Tokyo: Maruzen Co. Ltd. viii + xii + 224 pp., Pls. 1–37.

Aoyagi H. 1954. Description of one new genus and three new species of Blenniidae from the Riu-Kiu Islands. *Dobutsugaku Zasshi* 63(5): 213–217. [In Japanese, English summary]

Arai M, Amaoka K. 1996. Arnoglossus macrolophus Alcock (Pleuronectiformes: Bothidae); a valid species distinct from A. tapeinosomus (Bleeker). Ichthyological Research 43(4): 359–365.

Arata GF Jr. 1954. A contribution to the life history of the swordfish, *Xiphias gladius* Linnaeus, from the South Atlantic coast of the United States and the Gulf of Mexico. *Bulletin of Marine Science* 4(3): 183–243.

Arndt E, Fricke R. **2019**. Intertidal fishes of Mauritius with special reference to shallow tidepools. *Biodiversity Data Journal* 7(e36754): 1–44. http://dx.doi.org/10.3897/ BDJ.7.e36754

Arnold DC. 1956. A systematic revision of the fishes of the teleost family Carapidae (Percomorphi, Blennioidea), with descriptions of two new species. *Bulletin of the British Museum (Natural History) Zoology* 4(6): 247–307.

Arnold RJ, Pietsch TW. 2012. Evolutionary history of frogfishes (Teleostei: Lophiiformes: Antennariidae): a molecular approach. *Molecular Phylogenetics and Evolution* 62: 117–129. http://dx.doi.org/10.1016/j.ympev.2011.09.012

Arnoult J. **1959**. *Faune de Madagascar. X. Poissons des eaux douces*. Publications de l'Institut de Recherche Scientifique Tananarive-Tsimbazaza. 163 pp., Pls. 1–23.

Arnoult J. 1986. Scatophagidae (p. 341). In: Daget J, Gosse
J-P, Thys van den Audenaerde DFE (eds) *Check-list of the freshwater fishes of Africa (CLOFFA)*. Vol. 2. Brussels: Institut Royal des Sciences Naturelles de Belgique. 521 pp.

Arnoult J, Bauchot-Boutin M-L, Roux-Estève R. 1958. Les poissons de l'île Aldabra. Campagne océanograhique de la *Calypso* (Mai–Juin 1954). *Annales de l'Institut Océanographique* (Monaco) (N.S.) 34: 47–90.

Arora HL, Banerji SK. **1957**. Flying-fish fishery along the Coromandel Coast. *Indian Journal of Fisheries* 4(1): 80–91.

Arratia G. **1981**. *Varasichthys ariasi* n. gen. et sp. from the Upper Jurassic of Chile (Pisces, Teleostei, Varasichthyidae n. fam.). *Palaentographica* 175: 107–139.

Arratia G. 1991. The caudal skeleton of Jurassic teleosts: a phylogenetic analysis (pp. 249–340). In: Chang M-M, Liu Y-H, Zhang G-R (eds) *Early vertebrates and related problems in evolutionary biology*. Beijing: Science Press. 514 pp.

Arratia G. 1996. Reassessment of the phylogenetic relationships of certain Jurassic teleosts and their implications on teleostean phylogeny (pp. 219–242). In: Arratia G, Viohl G (eds) *Mesozoic Fishes – Systematics and paleoecology. Proceedings of the 1st International Meeting on Mesozoic Fishes, Eichstätt, 1993.* Munich: Verlag Dr. Friedrich Pfeil. 576 pp.

Arratia G. **1997**. Basal teleosts and teleostean phylogeny. *Palaeo Ichthyologica* 7: 1–168.

 Arratia G. 1999. The monophyly of Teleostei and stem-group teleosts. Consensus and disagreements (pp. 265–334). In:
 Arratia G, Schultze H-P (eds) *Mesozoic Fishes 2 – Systematics and fossil record. Proceedings of the international meeting, Buckow, 1997.* Munich: Verlag Dr. Friedrich Pfeil. 604 pp.

Arratia G. **2000**. New teleostean fishes from the Jurassic of southern Germany and the systematic problems concerning the "Pholidophoriforms". *Paläontologische Zeitschrift* 74(1): 113–143.

Arratia G. 2000. Phylogenetic relationships of Teleostei. Past and present. *Estudios oceanológicos* 19: 19–51.

Arratia G. **2001**. The sister-group of Teleostei: consensus and disagreements. *Journal of Vertebrate Paleontology* 21(4): 767–773.

Arratia G. 2004. Mesozoic halecostomes and the early radiation of teleosts (pp. 279–315). In: Arratia G, Tintori A (eds) *Mesozoic Fishes 3 – Systematics, paleoenvironments and biodiversity. Proceedings of the international meeting, Serpianao, 2001.* Munich: Verlag Dr. Friedrich Pfeil. 649 pp.

Arratia G. **2008**. The vasarichthyid and other crossognathiform fishes, and the break-up of Pangea. In: Cavin L, Longbottom A, Richter M (eds) *Fishes and the break-up of Pangea. Geological Society of London Special Publication* 295: 71–92.

Arratia G. 2013. Morphology, taxonomy, and phylogeny of Triassic pholidophorid fishes (Actinopterygii, Teleostei). *Journal of Vertebrate Paleontology* 33 (Supplement 6, Mémoire 13): 1–138. http://dx.doi.org/10.1080/02724634.2 013.835642

Arratia G. 2015. Los peces osteíctios fósiles de Chile y su importancia en los contextos paleobiogeográfico y evolutivo. In: Rubilar-Rogers D, Otero R, Vargas A, Sallaberry M (eds) *Vertebrados Fósiles de Chile. Publicación Ocasional del Museo Nacional de Historia Natural* (Santiago, Chile) 63: 35–83.

Arratia G, Lambers P. 1996. The caudal skeleton of pachycormiforms: parallel evolution? (pp. 191–218) In: Arratia G, Viohl G (eds) *Mesozoic Fishes – Systematics and paleoecology. Proceedings of the 1st International Meeting on Mesozoic Fishes, Eichstätt, 1993.* Munich: Verlag Dr. Friedrich Pfeil. 576 pp. Arratia G, Schultze H-P. 2013. Outstanding features of a new Late Jurassic pachycormiform fish from the Kimmeridgian of Brunn, Germany and comments on current understanding of pachycormiforms (pp. 87–120).
In: Arratia G, Schultze H-P, Wilson MVH (eds) *Mesozoic Fishes 5 – Global diversity and evolution. Proceedings of the international meeting, Saltillo, 2010.* Munich: Verlag Dr. Friedrich Pfeil. 560 pp.

Arratia G, Tischlinger H. 2010. The first record of Late Jurassic crossognathiform fishes from Europe and their phylogenetic importance for teleostean phylogeny. *Fossil Record* 13(2): 317–341. http://dx.doi.org/10.1002/mmng.201000005
Artedi P. 1793. In: Röse AF.

Ascanius P. **1772**. *Icones rerum naturalium, ou figures enluminées d'histoire naturelle du Nord. Cahier 2*. Copenhagen: C.I. Philibert. 8 pp., Pls. 11–20.

Aschliman NC, Ebert DA, Compagno LJV. **2010**. A new legskate (Rajoidei: genus *Cruriraja*) from southern Africa. *Copeia* 2010(3): 364–372. http://dx.doi.org/10.1643/CI-09-215

Aschliman NC, Nishida M, Miya M, Inoue JG, Rosana KM, Naylor GJ. 2012. Body plan convergence in the evolution of skates and rays (Chondrichthyes: Batoidea). *Molecular Phylogenetics and Evolution* 63(1): 28–42. http://dx.doi. org/10.1016/j.ympev.2011.12.012

Aschliman NC, Tibbetts IR, Collette BB. **2005**. Relationships of sauries and needlefishes (Teleostei: Scomberesocoidea) to the internally fertilizing halfbeaks (Zenarchopteridae) based on the pharyngeal jaw apparatus. *Proceedings of the Biological Society of Washington* 118(2): 416–427.

Assadi H, Dehghani R, Jahanbakhsh M. **1997**. *Atlas of the Persian Gulf and the Sea of Oman fishes*. Iran: Iranian Fisheries Research and Training Organization. 226 pp.

Asso y del Rio IJ de. 1801. Introduccion a la ichthyologia oriental de España. Anales de Ciencias Naturales 4(10): 28–52.

Astakhov DA. **1978**. Materials on morphology, taxonomy and distribution of the species of the genus *Cubiceps* Lowe (Nomeidae, Osteichthyes). *Trudy Instituta Okeanologii* 111: 132–156. [In Russian]

Astakhov DA. **1996.** Discovery of *Amphiprion chrysogaster* (Pomacentridae) on the barrier reef of Tulear (southwestern Madagascar). *Voprosy Ikhtiologii* 36(6): 843–846. [In Russian. English translation in *Journal of Ichthyology* 36(9): 804–807]

Ateweberhan M, Van Reine P. **2005**. A taxonomic survey of seaweeds from Eritrea. *Blumea – Biodiversity, Evolution and Biogeography of Plants* 50: 65–111.

Atz JW. **1951**. Fishes that look like plants. *Animal Kingdom* 54(5): 130–136.

Au K-C. **1979**. Systematic study on the barracudas (Pisces: Sphyraenidae) from a northern sector of the South China Sea. *Journal of Natural History* 13(5): 619–647.

Ayres WO. **1855**. [Descriptions of new species of Californian fishes.] A number of short notices read before the Society at several meetings in 1855. *Proceedings of the California Academy of Sciences* Series 1(pt 1): 23–77.

Ayres WO. **1863**. Descriptions of fishes believed to be new. *Proceedings of the California Academy of Sciences* Series 1(pt 2): 209–211.

Azzurro E, Goren M, Diamant A, Galil B, Bernardi G. 2015. Establishing the identity and assessing the dynamics of invasion in the Mediterranean Sea by the dusky sweeper, *Pempheris rhomboidea* Kossmann & Räuber, 1877 (Pempheridae, Perciformes). *Biological Invasions* 17(3): 815–826. http://dx.doi.org/10.1007/s10530-014-0836-5

В

Baars M, Schaalk P, Veldhuis M. 1998. Seasonal fluctuations in plankton biomass and productivity in the ecosystems of the Somali Current, Gulf of Aden, and southern Red Sea (pp. 143–174). In: Sherman K, Okemwa E, Ntiba M (eds) *Large Marine Ecosystems of the Indian Ocean: assessment, sustainability and management. Parts I & II: Pelagic ecosystems*. Oxford: Blackwell Science. 420 pp.

Babu C, Ramachandran S, Varghese BC. 2011. New record of sixgill sting ray *Hexatrygon bickelli* Heemstra and Smith, 1980 from south-west coast of India. *Indian Journal of Fisheries* 58(2): 137–139.

Backus RH, Mead GW, Haedrich RL, Ebeling AW. **1965**. The mesopelagic fishes collected during cruise 17 of the R/V *Chain*, with a method for analyzing faunal transects. *Bulletin of the Museum of Comparative Zoology* 134(5): 139–158.

Backus RH, Springer S, Arnold EL Jr. **1956**. A contribution to the natural history of the white-tip shark, *Pterolamiops longimanus* (Poey). *Deep Sea Research* 3: 178–188.

Baelde P. 1996. Biology and dynamics of the reproduction of blue-eye trevalla, *Hyperoglyphe antarctica* (Centrolophidae), off Tasmania, southern Australia. *Fishery Bulletin* 94(2): 199–211.

Bagarinao T. **1994**. Systematics, distribution, genetics and life history of milkfish, *Chanos chanos. Environmental Biology of Fishes* 39(1): 23–41.

Bailly N, Hureau J-C. **1995**. Bases de données en biologie: quelques problèmes liés à la nomenclature et aux références bibliographiques. *Cybium* 19(4): 333–342.

Baird SF. **1872**. Venomous fish in the Mauritius. *Annual Record* of Science and Industry 1872–73: p. 263.

Baissac J de B. 1953. Contribution à l'étude des poissons de l'île Maurice. V. Proceedings of the Royal Society of Mauritius 1(3) [for 1952]: 185–240.

Baissac J de B. **1955**. *Annals of the fisheries report of Mauritius* (Appendix B).

Baissac J de B. **1956**. Description d'un nouveau serranide de l'île Maurice. *Proceedings of the Royal Society of Arts and Sciences of Mauritius* 1(4): 395–396.

Baissac J de B. **1958**. [Fishes]. *Annual Report of the Mauritius Institute* (Port Louis) 1957: 5–11.

Baissac J de B. **1962**. Contribution à l'étude des poissons de l'île Maurice. VIII. *Proceedings of the Royal Society of Arts and Sciences of Mauritius* 2(2): 162–189.

Baissac J de B. 1968. Some notes on the fish species of Rodrigues. Proceedings of the Royal Society of Arts and Sciences of Mauritius 3(1): 67–73.

Baissac J de B. 1976. Poissons de mer des eaux de l'île Maurice. Proceedings of the Royal Society of Arts and Sciences of Mauritius 3(2): 191–226.

Baissac J de B. **1990**. SWIOP Checklist of the marine fishes of Mauritius. Regional Project for the Development and Management of Fisheries in the Southwest Indian Ocean. RAF/87/008/WP/54/90, 42 pp.

Baker EA, Collette BB. **1998**. Mackerel from the northern Indian Ocean and the Red Sea are *Scomber australasicus*, not *Scomber japonicus*. *Ichthyological Research* 45(1): 29–33.

Bakus G, Arthur R, Ekaratne S, Jinendradasa SS. 2000. Chapter 10. India and Sri Lanka. (pp. 295–323). In: McClanahan TR, Sheppard CRC, Obura DO (eds) Coral reefs of the Indian Ocean: Their ecology and conservation. Oxford: Oxford University Press. xxiii + 525 pp.

Bal DV, Joshi MS. 1956. Studies on the biology of *Coilia dussumieri* (Cuv. and Val.). *Indian Journal of Fisheries* 3(1): 91–100.

Bal DV, Mohmed KH. **1957**. A systematic account of the eels of Bombay. *Journal of the Bombay Natural History Society* 54(3): 732–740.

Balasubrahmanyan K, Bhushana Rao KSP, Subba Raju RC. 1967. Larval and juvenile stages of the flying fish *Exocoetus volitans* Linn. from the Bay of Bengal. *Bulletin of the National Institute of Sciences of India* 38: 876–884.

Baldwin CC. **1990**. Morphology of the larvae of American Anthiinae (Teleostei: Serranidae) with comments on relationships within the subfamily. *Copeia* 1990(4): 913–955.

Baldwin CC, Johnson GD. 1993. Phylogeny of the Epinephelinae (Teleostei: Serranidae). Bulletin of Marine Science 52(1): 240–283.

Baldwin CC, Johnson GD. 1996. Interrelationships of Aulopiformes (pp. 355–404). In: Stiassny MLJ, Parenti LR, Johnson GD (eds) *Interrelationships of fishes*. San Diego: Academic Press. 496 pp.

Baldwin CC, Johnson GD. 1999. *Paxton concilians*: a new genus and species of pseudamine apogonid (Teleostei: Percoidei) from northwestern Australia: the sister group of the enigmatic *Gymnapogon. Copeia* 1999(4): 1050–1071.

Baldwin CC, Johnson GD, Paxton JR. 1997. Protoblepharon rosenblatti, a new genus and species of flashlight fish (Beryciformes: Anomalopidae) from the tropical South Pacific, with comments on anomalopid phylogeny. Proceedings of the Biological Society of Washington 110(3): 373–383.

Baldwin WJ. **1972**. A new genus and new species of Hawaiian gobiid fish. *Pacific Science* 26(1): 125–128.

Baldwin WJ. 1984. A note on the occurrence of the gold spot herring, *Herklotsichthys quadrimaculatus* (Rüppell) in Hawaii. *Pacific Science* 38(2): 123–126.

Baldwin ZH. **2005**. A new species of bullhead shark, genus *Heterodontus* (Heterodontiformes: Heterodontidae), from Oman. *Copeia* 2005(2): 262–264.

Baldwin ZH, Sparks JS. **2011**. A new species of *Secutor* (Teleostei: Leiognathidae) from the western Indian Ocean. *Zootaxa* 2998: 39–47. http://dx.doi.org/10.11646/zootaxa.2998.1.3

Balon EK, Bruton MN. **1994**. Fishes of the Tatinga River, Comoros, with comments on freshwater amphidromy in the goby *Sicyopterus lagocephalus*. *Ichthyological Exploration of Freshwaters* 5(1): 25–40.

Bamber RC. 1915. Reports on the marine biology of the Sudanese Red Sea, from collections made by Cyril Crossland, M.A., D.Sc., F.L.S. XXII. The fishes. *Journal of the Linnean Society of London* (Zoology) 31(210): 477–485.

Bancroft EN. **1829**. On the fish known in Jamaica as the seadevil. *Zoological Journal* 4(16): 444–457.

Banse K. **1959**. On upwelling and bottom trawling off the southwest coast of India. *Journal of the Marine Biological Association of India* 1: 33–49.

Bapat SV. **1970**. The Bombay duck, *Harpadon nehereus* (Ham.). Bulletin of the Central Marine Fish Research Institute 21: 1–66.

Baranes A. **2003**. Sharks from the Amirantes Islands, Seychelles, with a description of two new species of squaloids from the deep sea. *Israel Journal of Zoology* 49(1): 33–65.

Baranes A, Fricke R, Golani D, Appelbaum-Golani
B. 2017. Record of *Bodianus rubrisos* Gomon, 2006 from the northern Red Sea, previously misidentified as *B. leucosticticus* (non Bennett) or *B. trilineatus* (non Fowler) (Labridae). *Cybium* 40(4): 281–286. https://doi. org/10.26028/cybium/2016-404-003

Baranes A, Golani D. 1993. An annotated list of deep-sea fishes collected in the northern Red Sea, Gulf of Aqaba. *Israel Journal of Zoology* 39(4): 299–336.

Baranes A, Randall JE. 1989. Narcine bentuviai, a new torpedinoid ray from the northern Red Sea. Israel Journal of Zoology 36: 85–101. Barbosa du Bocage JV, De Brito Capello F. **1864**. Sur quelques espèces inédites de Squalidæ de la tribu Acanthiana, Gray, qui fréquentent les côtes du Portugal. *Proceedings of the Scientific Meetings of the Zoological Society of London* 1864 (pt 2): 260–263.

Barbour T. **1941**. Notes on pediculate fishes. *Proceedings of the New England Zoölogical Club* 19: 7–14, Pls. 2–7.

Bardack D. 1965. Anatomy and evolution of chirocentrid fishes. *The University of Kansas Paleontological Contributions: Vertebrata*, Article 10: 1–86, Pls. 1–2, Figs. 1–27.

Bariche M. **2010**. *Champsodon vorax* (Teleostei: Champsodontidae), a new alien fish in the Mediterranean. *aqua, International Journal of Ichthyology* 16(4): 197–200.

Bariche M, Heemstra P. **2012**. First record of the blacktip grouper *Epinephelus fasciatus* (Teleostei: Serranidae) in the Mediterranean Sea. *Marine Biodiversity Records* 5(e1): 1–3. https://doi.org/10.1017/S1755267211000509

Barlow G. **2002**. *The cichlid fishes: Nature's grand experiment in evolution*. Cambridge: Basic Books. 352 pp.

Barman RP, Mishra SS. **2006**. Review of the flying fish family Exocoetidae in the Indian waters. *Zoological Survey of India Occasional Paper* 256: 1–29.

Barnard KH. 1923. Diagnoses of new species of marine fishes from South African waters. Annals of the South African Museum 13(pt 8, no. 14): 439–445.

Barnard KH. **1925**. A monograph of the marine fishes of South Africa. Part I. *Annals of the South African Museum* 21(1): 1–418.

Barnard KH. 1925. Descriptions of new species of marine fishes from South Africa. Annals and Magazine of Natural History (Series 9) 15(88): 498–504.

Barnard KH. **1927**. A monograph of the marine fishes of South Africa. Part II. *Annals of the South African Museum* 21(2): 419–1065.

Barnard KH. 1927. Diagnoses of new genera and species of South African marine fishes. *Annals and Magazine of Natural History* (Series 9) 20(115): 66–79.

Barnard KH. 1934. New records and descriptions of two new species of South African marine fishes. Annals and Magazine of Natural History (Series 10) 13(74): 228–235.

Barnard KH. **1935**. Notes on South African marine fishes. *Annals of the South African Museum* 30(5): 645–658.

Barnard KH. **1937**. Further notes on South African marine fishes. *Annals of the South African Museum* 32(2–6): 41–67.

Barnard KH. 1943. Revision of the indigenous freshwater fishes of the S.W. Cape region. *Annals of the South African Museum* 36(2): 101–262.

Barnard KH. 1947. A pictorial guide to South African fishes marine and freshwater. Cape Town: Maskew Miller Limited. xvii + 226 pp. Barnard KH. **1948**. Further notes on South African marine fishes. *Annals of the South African Museum* 36(5): 341–406.

Barnard KH. **1950**. The date of issue of the "Illustrations of the Zoology of South Africa" and "The Marine Investigations in South Africa". *Journal of the Society for the Bibliography of Natural History* 2(6): 187–189.

Barnard KH. **1955**. South African parasitic Copepoda. *Annals* of the South African Museum 41: 223–312.

Barnard KH, Davies DH. 1947. Description of a new fish of the family Zeidae from the Cape. Annals and Magazine of Natural History (Series 11) 13(107) [for 1946]: 790–792.

Barsukov VV. 1973. The species composition of the genus *Helicolenus* (Sebastinae, Scorpaenidae, Pisces) and a description of a new species. *Voprosy Ikhtiologii* 13(2): 195–201. [In Russian. English translation in *Journal of Ichthyology* 13(2): 161–167]

Barsukov VV. 1979. Subspecies of the Atlantic Helicolenus dactylopterus (De la Roche, 1809). Voprosy Ikhtiologii 19(4): 579–595. [In Russian. English translation as Subspecies of the Atlantic blackbelly rosefish Helicolenus dactylopterus (De la Roche, 1809) in Journal of Ichthyology 19(4): 1–17]

Barsukov VV. **1989**. The upper Miocene rock fishes (Scorpaenidae, Sebastinae) from California. *Proceedings of the Zoological Institute of Leningrad* 201: 73–108. [In Russian]

Baschieri-Salvadori F. **1955**. Spedizione subacquea italiana nel Mar Rosso. Ricerche zoologiche. IX. Pomacentridi. *Rivista di Biologia Coloniale* 15: 57–68.

Bass AJ. **1973**. Analysis and description of variation in the proportional dimensions of scyliorhinid, carcharhinid and sphyrnid sharks. *Investigational Report. Oceanographic Research Institute* (Durban) 32: 1–28.

Bass AJ. 1978. Sharks (pp. 176–177). In: Heydorn AEF (ed) Ecology of the Agulhas Current region: an assessment of biological responses to environmental parameters in the South-west Indian Ocean. *Transactions of the Royal Society* of South Africa 43(2): 151–190.

Bass AJ, D'Aubrey JD, Kistnasamy N. **1973**. Sharks of the east coast of southern Africa. I. The genus *Carcharhinus* (Carcharhinidae). *Investigational Report*. *Oceanographic Research Institute* (Durban) 33: 1–168.

Bass AJ, D'Aubrey JD, Kistnasamy N. **1975**. Sharks of the east coast of southern Africa. II. The families Scyliorhinidae and Pseudotriakidae. *Investigational Report. Oceanographic Research Institute* (Durban) 37: 1–63.

Bass AJ, D'Aubrey JD, Kistnasamy N. **1975**. Sharks of the east coast of southern Africa. III. The families Carcharhinidae (excluding *Mustelus* and *Carcharhinus*) and Sphyrnidae. *Investigational Report. Oceanographic Research Institute* (Durban) 38: 1–100. Bass AJ, D'Aubrey JD, Kistnasamy N. **1975**. Sharks of the east coast of southern Africa. IV. The families Odontaspididae, Scapanorhynchidae, Isuridae, Cetorhinidae, Alopiidae, Orectolobidae and Rhiniodontidae. *Investigational Report*. *Oceanographic Research Institute* (Durban) 39: 1–102.

Bass AJ, D'Aubrey JD, Kistnasamy N. 1975. Sharks of the east coast of southern Africa. V. The families Hexanchidae, Chlamydoselachidae, Heterodontidae, Pristiophoridae and Squatinidae. *Investigational Report. Oceanographic Research Institute* (Durban) 43: 1–50.

 Bass AJ, D'Aubrey JD, Kistnasamy N. 1976. Sharks of the east coast of southern Africa. VI. The families Oxynotidae,
 Squalidae, Dalatiidae and Echinorhinidae. *Investigational Report. Oceanographic Research Institute* (Durban) 45: 1–103.

Basson PW, Burchard JE Jr, Hardy JT, Price ARG. 1977.
Biotypes of the Western Arabian Gulf: Marine life and environments of Saudi Arabia. Dhahran, Saudi Arabia: The Arabian American Oil Company, Department of Loss Prevention and Environmental Affairs. 284 pp.

Bath H. **1977**. Revision der Blenniini (Pisces: Blenniidae). *Senckenbergiana Biologica* 57(4/6): 167–234.

Bath H. 1983. Omobranchus hikkaduwensis n. sp. von Ceylon (Pisces: Blenniidae). Senckenbergiana Biologica 64(1/3): 25–30.

Bath H. 1983. Revision der Gattung Antennablennius Fowler 1931 mit Beschreibung einer neuen Art und Untersuchung der taxonomischen Stellung von Antennablennius anuchalis Springer & Spreitzer 1978. Senckenbergiana Biologica 64(1/3): 47–80.

Bath H. **1989**. Die Arten der Gattung *Parablennius* Ribeiro 1915 im Roten Meer, Indischen und NW des Pazifischen Ozeans. *Senckenbergiana Biologica* 69(4/6): 301–343.

Bath H. 1990. Taxonomie und Verbreitung von *Parablennius* Ribeiro 1915 an der W-Küste Afrikas und den Kapverdsischen Inseln mit Revalidation von *P. verryckeni* (Poll 1959) und Beschreibung drei neuer Arten (Pisces: Blenniidae). *Senckenbergiana Biologica* 70(1/3): 15–69.

Bath H. **1996**. Beitrag zur Osteologie der Arten der Tribus Parablenniini. Die Beziehungen der Knochen des Schädeldaches zum Seitenorgan-System und zu den Weichteilbildungen der Kopfoberseite sowie die systematische Bedeutung der Befunde nebst Bemerkungen zu *Lupinoblennius dispar* Herre 1942. (Pisces: Blenniidae). *Senckenbergiana Biologica* 76(1/2): 65–92. [In German, English summary]

Bauchot M-L. **1958**. In: Arnoult J, Bauchot-Boutin M-L, Roux-Estève R.

Bauchot M-L. 1963. Catalogue critique des types de poissons du Muséum National d'Histoire Naturelle. I. Famille des Labridae. *Publications diverses du Muséum National* d'Histoire Naturelle 20: 1–113; 180–195. Bauchot M-L. 1967. Catalogue critique des types de poissons du Muséum National d'Histoire Naturelle (suite). Sousordre des Blennioidei. Publications diverses du Muséum National d'Histoire Naturelle 21: 1–70.

Bauchot M-L. 1967. Poissons marins de l'Est Atlantique Tropical. Téléosteens Perciformes (2. Percodei, 3. Acanthuroidei, 4. Balistoidei). *Atlantide Report* 9: 7–43.

Bauchot M-L. 1967. Poissons marins de l'Est Atlantique Tropical. Téleosteens Perciformes (5. Blennioidei). *Atlantide Report* 9: 63–91.

Bauchot M-L. 1970. Catalogue critique des types des Poissons du Muséum National d'Histoire Naturelle (Lampridiformes, Stephanobéryciformes, Bericyformes, Zéiformes, Coryphaeniformes). Publications diverses du Muséum National d'Histoire Naturelle 24: 1–55.

Bauchot M-L. 1987. Poisson osseux (pp. 892–1422). In:
Fischer W, Schneider M, Bauchot M-L (eds) Fiches FAO d'identification des espèces pour les besoins de la pêche (Révision 1) : Méditerranée et Mer Noir (zone de pêche 37).
Vol. 2: Vertébrés. pp. 761–1530. Rome: FAO.

Bauchot M-L, Bianchi G. 1984. Diplodus cervinus omanensis, nouvelle sous-espèce de Diplodus cervinus (Lowe, 1841), capturée en mer d'Arabie (Pisces, Perciformes, Sparidae). Cybium 8(3): 103–105.

Bauchot M-L, Bianchi G. 1984. Guide des poissons commerciaux de Madagascar (espèces marines et d'eaux saumâtres). Fiches FAO d'identification sheets des espèces pour les besoins de la pêche. Rome: FAO. iv + 135 pp. [In French]

Bauchot M-L, Blache J. 1979. Présence d'Ariosoma balearicum (De la Roche, 1809) en Mer Rouge (Pisces, Teleosei, Congridae). Bulletin du Muséum National d'Histoire Naturelle (Série 4) 1A: 1131–1137.

Bauchot M-L, Blanc M. 1961. Catalogue des types de Scombroidei (Poissons Téléosteens Perciformes) des collections du Muséum National d'Histoire Naturelle de Paris. Bulletin du Muséum National d'Histoire Naturelle (Série 2) 33(4): 369–379.

Bauchot M-L, Blanc M. 1961. Poissons marins de l'EstAtlantique tropical. 1. Labroidei (Téléostéens Perciformes),Irepartie, 2. Percoidei. *Atlantide Report* 6: 65–100.

Bauchot M-L, Daget J. 1971. Les Diplodus (Pisces, Sparidae) de groupe cervinus-fasciatus. Cahiers ORSTOM (Série Océanographie) 9(3): 319–338.

Bauchot M-L, Daget J. 1972. Catalogue critique des types de poissons du Muséum National d'Histoire Naturelle. (Suite) (Famille des Sparidae). *Bulletin du Muséum National d'Histoire Naturelle* (Paris) (Série 3: Zoologie) 24 (Zoologie 18): 33–99.

Bauchot M-L, Desoutter M. **1989**. Catalogue critique des types de poissons du Muséum National d'Histoire Naturelle. (Suite) Sous-ordre des Percoidei. Familles des Aplodactylidae, Apolectidae, Arripidae, Cepolidae, Cheilodactylidae, Chironemidae, Cirrhitidae, Echeneidae, Enoplosidae, Embiotocidae, Gerreidae, Lactariidae, Latrididae, Leiognathidae, Lobotidae, Malacanthidae, Menidae, Nandidae, Oplegnathidae, Owstoniidae, Pomatomidae et Rachycentridae). *Bulletin du Muséum National d'Histoire Naturelle* (Paris) (Série 4) Section A 11(2) Supplement: 1–58.

Bauchot M-L, Desoutter M, Guézé P, Randall JE. 1985.
Catalogue critique des types de poissons de Muséum National d'Histoire Naturelle. (Suite) (Famille des Mullidae). Bulletin du Muséum National d'Histoire Naturelle (Paris) (Série 4) Section A 7(2) Supplement: 1–21.

Bauchot M-L, Desoutter M, Hoese DF, Larson HK. 1991.
Catalogue critique des types des poissons du Muséum National d'Histoire Naturelle. (Suite) Sous-ordre des Gobioidei. Bulletin du Muséum National d'Histoire Naturelle (Paris) (Série 4) 13(1–2): 1–82.

Bauchot M-L, Desoutter M, Randall JE. 1984. Catalogue critique des types de poissons du Muséum National d'Histoire Naturelle: Famille des Serranidae. *Bulletin du Muséum National d'Histoire Naturelle* (Paris) (Série 4) Section A 6(3) Supplement: 3–82.

Bauchot M-L, Guibé J. **1960**. Catalogue des types de poissons du Muséum National d'Histoire Naturelle: Famille des Scaridae. *Bulletin du Muséum National d'Histoire Naturelle* (Paris) (Série 2) 32(4): 290–300.

Bauchot M-L, Hureau J-C. 1990. Sparidae (pp. 790–812).
In: Quéro J-C, Hureau J-C, Karrer K, Post A, Saldanha L (eds) *Check-list of the fishes of the eastern tropical Atlantic* (*CLOFETA*). Vol. 2. Lisbon: Junta Nacional de Investigação Científica e Tecnológica.

Bauchot M-L, Hureau J-C, Miquel JC. 1981. Sparidae.
In: Fischer W, Bianchi G, Scott WB (eds) FAO species identification sheets for fishery purposes. Eastern central Atlantic (Fishing Area 34 and part of 47). Vol. 4. Rome: FAO. [unpaginated]

Bauchot M-L, Maugé AL. 1978. Première capture d'un *Thamnaconus* dans le golfe d'Aqaba: *Thamnaconus modestoides erythraeenis* n. ssp. (Pisces, Monacanthidae). No. 520. Bulletin du Muséum National d'Histoire Naturelle (Série 3: Zoologie) 356: 539–545.

Bauchot M-L, Maugé AL. 1980. Muraenichthys erythraeensis n. sp. de mer Rouge et première mention de Muraenichthys laticaudata (Ogilby, 1897) en mer Rouge (Pisces, Anguilliformes, Ophichthidae). Bulletin du Muséum National d'Histoire Naturelle (Série 4: Section A: Zoologie, Biologie et Écologie Animales) 2(3): 933–939.

Bauchot M-L, Randall JE. 1996. Catalogue critique des types de poissons du Muséum National d'Histoire Naturelle (suite). Families des Acanthuridae et des Zanclidae. *Cybium* 20(1): 55–74. Bauchot M-L, Skelton PH. 1986. Sparidae (pp. 331–332). In: Daget J, Gosse J-P, Thys van den Audenaerde DFE (eds) *Check-list of the freshwater fishes of Africa (CLOFFA)*. Vol. 2.
Brussels: Institut Royal des Sciences Naturelles de Belgique. 521 pp.

Bauchot M-L, Smith MM. 1984. Sparidae. Dentex, hottentots, pandoras, porgies, salemas, seabreams, stumpnoses. In:
Fischer W, Bianchi G (eds) FAO species identification sheets for fishery purposes. Western Indian Ocean (Fishing Area 51).
Vol. 4. Rome: FAO. [unpaginated]

Bauchot-Boutin M-L, see Bauchot M-L.

Bayoumi AR. **1972**. Recent biological investigations in the Red Sea along the A.R.E. coasts. 1. On some demersal fishes of economic importance from the Red Sea, with notes on migration of fish through the Suez Canal. *Bulletin of the Institute of Oceanography and Fisheries* (Egypt) 2: 157–183.

Bean BA, Weed AC. **1912**. Notes on a collection of fishes from Java, made by Owen Bryant and William Palmer in 1909, with description of a new species. *Proceedings of the United States National Museum* 42(1919): 587–611.

Beardsley GL Jr, Merrett NR, Richards WJ. 1975. Synopsis of the biology of the sailfish Istiophorus platypterus (Shaw and Nodder, 1791). NOAA Technical Report NMFS SSRF-675 (Part 3): 95–120.

Bebars MI. **1978**. *Scarus ghardaqensis*, n.s.p., a new parrotfish (Pisces, Scaridae) from the Red Sea, with a note on sexual dichromatism in the family. *Cybium* (Série 3) 2(3): 76–81.

Beckett JS. **1974**. Biology of swordfish, *Xiphias gladius* L., in the northwest Atlantic Ocean. *NOAA Technical Report* NMFS SSRF-675 (Part 2): 103–106.

Beckley LE. **1994**. Fish (pp. 211–285). In: Branch GM, Griffiths CL, Branch ML, Beckley LE (eds) *Two oceans: A guide to the marine life of South Africa.* Cape Town: David Philip Publishers. 360 pp.

Beckley LE, Cliff G, Smale MJ, Compagno LJV. **1997**. Recent strandings and sightings of whale sharks in South Africa. *Environmental Biology of Fishes* 50(3): 343–348.

Beebe W. **1933**. Deep-sea fishes of the Bermuda oceanographic expeditions. Family Alepocephalidae. *Zoologica* (N.Y.) 16(2): 15–93.

Beebe W, Tee-Van J. **1933**. Nomenclatural notes on the shore fishes of Bermuda. *Zoologica* (N.Y.) 13(7): 133–158.

Beebe W, Tee-Van J. 1941. Eastern Pacific expeditions of the New York Zoological Society. XXVIII Fishes from the tropical eastern Pacific. Part 3: Rays, mantas, and chimaeras. *Zoologica* (N.Y.) 26(pt 3, no. 16): 245–280.

Bell LJ, Colin PL. 1986. Mass spawning of *Caesio teres* (Pisces: Caesionidae) at Enewetak Atoll, Marshall Islands. *Environmental Biology of Fishes* 15(1): 69–74.

Bell MA, Foster SA (eds). 1994. The evolutionary biology of the threespine stickleback. Oxford: Oxford University Press. xii + 571 pp.

Belloc G. **1949**. Catalogue des types de poissons du Musée Océanographique de Monaco. *Bulletin de l'Institut Océanographique* 958: 1–23.

Bellotti C. **1874**. Sopra due specie di pesci raccolte in Egitto durante l'inverno del 1873–74. *Atti della Società Italiana di Scienze Naturali di Milano* 17(3): 262–265.

Bellwood DR. **1994**. A phylogenetic study of the parrotfishes Family Scaridae (Pisces: Labroidei), with a revision of genera. *Records of the Australian Museum, Supplement* 20: 1–86.

Bellwood DR, Randall JE. **2000**. *Pseudojuloides severnsi*, a new species of wrasse from Indonesia and Sri Lanka (Perciformes: Labridae). *Journal of South Asian Natural History* 5(1): 1–5.

Bellwood DR, Schultz O, Siqueira AC, Cowman PF. **2019**. A review of the fossil record of the Labridae. *Annalen des Naturhistorichen Museums in Wien*, Serie A 121: 125–193.

Belyanina TN. **1974**. Materials on development, taxonomy, and distribution of fishes of the family Bregmacerotidae. *Trudy Instituta Okeanologii* 96: 143–188. [In Russian, English summary]

Belyanina TN. **1993**. Early stages of development of the East Australian flying fishes (family Exocoetidae). *Trudy Instituta Okeanologii Akademii Nauk* 128: 108–146.

Belyanina TN. **1994**. Early stages of ontogeny of *Hirundichthys speculiger* and *H. affinis* (Exocoetidae) from the Atlantic Ocean. *Journal of Ichthyology* 34(5): 60–74. [Originally published in *Voprosy Ikhtiologii* 34(1): 98–107]

Bemert G, Ormond RFG. **1981**. *Red Sea coral reefs*. London: Kegan Paul International. 192 pp.

Bemis KE, Tyler JC, Psomadakis PN, Ferris LN, Kumar AB. 2020. Review of the Indian Ocean spikefish genus *Mephisto* (Tetraodontiformes: Triacanthodidae). *Zootaxa* 4802(1): 82–98. https://doi.org/10.11646/zootaxa.4802.1.5

Ben-Abdallah A, Al-Turky A, Nafti A, Shakman E. 2011. A new record of a Lessepsian fish, *Lagocephalus suezensis* (Actinopterygii: Tetraodontiformes: Tetraodontidae), in the south Mediterranean (Libyan coast). *Acta Ichthyologica et Piscatoria* 41(1): 71–72. http://dx.doi.org/10.3750/ AIP2011.41.1.11

Benbow B. **1974**. Records of five species of triacanthodid fishes from East Africa. *Journal of the Marine Biological Association of India* 16(3): 763–768.

Benham WB. **1904**. An apparently new species of *Regalecus* (*R. parkeri*). *Transactions of the New Zealand Institute* 36: 198–200.

Bennett BA. **1983**. *Clinus spatulatus*, a new species of clinid fish (Perciformes: Blennoidei) from South Africa, with a modified definition of the genus *Clinus*. *J.L.B. Smith Institute of Ichthyology Special Publication* 29: 1–9. Bennett ET. **1828**. Observations on the fishes contained in the collection of the Zoological Society. *Zoological Journal* 3(11): 371–378.

Bennett ET. **1828**. Observations on the fishes contained in the collection of the Zoological Society. On some fishes from the Sandwich Islands. *Zoological Journal* 4(13, art.3): 31–42.

Bennett ET. 1830. See Anonymous (Bennett ET).

Bennett ET. **1831**. Characters of a new species of *Pterois*. *Proceedings of the Zoological Society of London* 1830–1831(Part 1): 128. [Published on 25 Oct 1831; title in table of contents only]

Bennett ET. 1831–1832. Observations on an exhibition of the collection of fishes from the Mauritius, presented to the Society by Mr. Telfair. Proceedings of the Committee of Science and Correspondence of the Zoological Society of London 1830–1831(Part 1): 59–60, 61, 126–128, 165–169.

Bennett ET. **1833**. Characters of new species of fishes from Ceylon, presented to the Society by Dr. Sibbald. *Proceedings of the Zoological Society of London* 1832(Part 2): 182–184.

Bennett ET. **1833**. Characters of two new species of fishes, from the Mauritius, presented to the Society by Mr. Telfair. *Proceedings of the Committee of Science and Correspondence of the Zoological Society of London* 1832(Part 2): 184

Bennett ET. **1833**. Characters of new species from the Mauritius. *Proceedings of the Zoological Society of London* 1833(Part 1): 32.

Bennett ET. **1836**. Specimens were exhibited of various fishes, forming part of a collection from Mauritius, presented to the Society by M. Julien Desjardins. *Proceedings of the Zoological Society of London* 1835(Part 3): 206–208.

Bennett FD. 1840. Narrative of a whaling voyage round the globe, from the year 1833 to 1836. Vol. 2 [Fishes: pp. 255–289]. London: R. Bentley. vii + 395 pp.

Bennett JW. **1828–1830**. A selection from the most remarkable and interesting fishes found on the coast of Ceylon from drawings made in the southern part of that island from living specimens (1<sup>st</sup> edition). London: Longman, Rees, Orme, Brown and Green. viii + 30 pp.

Bennett JW. 1834. A selection from the most remarkable and interesting fishes found on the coast of Ceylon (2<sup>nd</sup> edition). London: Edward Bull.

Bennett JW. **1851**. A selection of rare and curious fishes found upon the coast of Ceylon: from drawings made in that island, and coloured from life, with letter-press descriptions. London: Printed for the author. 30 pp.

Ben-Tuvia A. **1953**. Mediterranean fishes of Israel. *Bulletin of the Sea Fisheries Research Station, Haifa* 8: 1–40.

Ben-Tuvia A. **1964**. Two siganid fishes of Red Sea origin in the eastern Mediterranean. *Bulletin of the Sea Fisheries Research Station, Haifa* 37(29): 1–9.

Ben-Tuvia A. **1966**. Red Sea fishes recently found in the Mediterranean. *Copeia* 1966(2): 254–275.

Ben-Tuvia A. **1968**. Report on the fisheries investigations of the Israel South Red Sea Expedition, 1962. *Bulletin of the Sea Fisheries Research Station, Haifa* 52: 21–55.

Ben-Tuvia A. 1976. Fish collections from the eastern Mediterranean, the Red Sea and inland waters of Israel. Jerusalem: Hebrew University. 32 pp.

Ben-Tuvia A. **1993**. A review of the Indo-west Pacific congrid fishes of genera *Rhynchoconger* and *Bathycongrus* with the description of three new species. *Israel Journal of Zoology* 39(4): 349–370.

Ben-Tuvia A, Golani D. 1989. A new species of goatfish (Mullidae) of the genus *Upeneus* from the Red Sea and the eastern Mediteranean. *Israel Journal of Zoology* 36(2): 103–112.

Ben-Tuvia A, Kissil GW. 1988. Fishes of the family Mullidae in the Red Sea, with a key to the species in the Red Sea and the eastern Mediterranean. *Ichthyological Bulletin of the J.L.B. Smith Institute of Ichthyology* 52: 1–16.

Ben-Tuvia A, Lourie A. 1969. A Red Sea grouper *Epinephelus tauvina* caught on the Mediterranean coast of Israel. *Israel Journal of Zoology* 18(2–3): 245–247.

Ben-Tuvia A, Steinitz H. 1952. Report on a collection of fishes from Eylat (Gulf of Aqaba). Bulletin of the Sea Fisheries Research Station, Haifa 2: 1–12.

Ben-Tuvia A, Trewavas E. **1987**. *Atrobucca geniae*, a new species of sciaenid fish from the Gulf of Elat (Gulf of Aqaba), Red Sea. *Israel Journal of Zoology* 34 [for 1986/87]: 15–21.

Berendzen PB, Dimmick WW. **2002**. Phylogenetic relationships of Pleuronectiformes based on molecular evidence. *Copeia* 2002(3): 642–652. https://doi.org/10.1643/0045-851 1(2002)002[0642:PROPBO]2.0.CO;2

Berg C. **1895**. Enumeración sistemática y sinonímica de los peces de las costas Argentina y Uruguaya. *Anales del Museo Nacional de Historia Natural de Buenos Aires* 4: 1–120.

Berg LS. 1940. Classification of fishes, both recent and fossil. *Trudy Zoologicheskogo Instituta, Akademii Nauk* (SSSR, Moskva, Leningrad) 5(2): 87–517. [In Russian, English subtitle]

Berg LS. **1949**. Freshwater fishes of Iran and of neighboring countries. *Trudy Zoologicheskogo Instituta, Akademii Nauk* (SSSR) 8(4): 783–858. [In Russian]

Bernadsky G, Goulet D. **1991**. A natural predator of the lionfish, *Pterois miles*. *Copeia* 1991(1): 230–231.

Bernardes RÁ, de Figueiredo JL, Rodrigues AR, Fischer LG, Vooren CM, Haimovici M, Rossi-Wongtschowski CLDB.
2005. Peixes de zona econômica exclusiva da região sudestesul do Brasil: Levantamento com armadilhas, pargueiras e rede de arrasto de fundo. São Paulo: Editora da Universidade de São Paulo. 295 pp. Bernardi G, Bucciarelli G, Costagliola D, Robertson DR, Heiser JB. 2004. Evolution of coral reef fish *Thalassoma* spp. (Labridae). 1. Molecular phylogeny and biogeography. *Marine Biology* 144: 369–375.

Berra TM. **2001**. *Freshwater fish distribution* (1<sup>st</sup> edition). San Diego, California: Academic Press. 604 pp.

Berry FH. **1958**. A new species of fish from the western North Atlantic, *Dikellorhynchus tropidolepis*, and relationships of the genera *Dikellorhynchus* and *Malacanthus*. *Copeia* 1958(2): 116–125.

Berry FH. **1959**. Boarfishes of the genus *Antigonia* of the western Atlantic. *Bulletin of the Florida State Museum* 4(7): 205–250.

Berry FH. **1968**. A new species of carangid fish (*Decapterus tabl*) from the western Atlantic. *Contributions in Marine Science* 13: 145–167.

Berry FH, Baldwin WJ. **1966**. Triggerfishes (Balistidae) of the eastern Pacific. *Proceedings of the California Academy of Sciences* (Series 4) 34(9): 429–474.

Berry FH, Cohen L. **1974**. Synopsis of the species of *Trachurus* (Pisces, Carangidae). *Quarterly Journal of the Florida Academy of Sciences* 35(4): 177–211.

Berry PF, van der Elst RP, Hanekom P, Joubert CSW, Smale MJ.
1982. Density and biomass of the ichthyofauna of a Natal littoral reef. *Marine Ecology Progress Series* 10: 49–55.

Bertin L. 1940. Catalogue des types de poissons du Muséum National d'Histoire Naturelle (2<sup>e</sup> partie). Dipneustes, Chondrostéens, Holostéens, Isospondyles. *Bulletin du Muséum National d'Histoire Naturelle* (Paris) (Série 2) 12(6): 244–372.

Bertin L. **1943**. Revue critique des Dussumierides actuels et fossiles. Description d'un genre nouveau. *Bulletin de l'Institut Océanographique* (Monaco) 853: 1–32.

Bertin L. 1946. In: Angel F, Bertin L, Guibé J.

Bertin L, Dollfus R-P. 1948. Revision des espèces du genre Decapterus (Téléostéens, Scombriformes). Mémoires du Muséum National d'Histoire Naturelle (Paris) (N.S.) 26(1): 1–29.

Bertin L, Estève R. 1950. Catalogue des types de poissons du Museum National d'Histoire Naturelle (6<sup>e</sup> partie). Haplomes, Hétéromes, Catostéomes. Paris: Imperial National. 60 pp.

Betancur-R R. **2009**. Molecular phylogenetics and evolutionary history of ariid catfishes revisited: a comprehensive sampling. *BMC Evolutionary Biology* 9 (art.175). http://dx.doi.org/10.1186/1471-2148-9-175

Betancur-R R, Broughton RE, Wiley EO, Carpenter K, López JA, Li C, Holcroft NI, Arcila D, Sanciangco M, Cureton JC II, Zhang F, Buser T, Campbell MA, Ballesteros JA, Roa-Varon A, Willis S, Borden WC, Rowley T, Reneau PC, Hough DJ, Lu G, Grande T, Arratia G, Orti G. **2013**. The tree of life and a new classification of bony fishes. *PLoS Currents Tree of Life* 5: 1–45. https://doi.org/10.1371/currents.tol.53ba26640df0ccaee75bb165c8c26288

Betancur-R R, Li C, Munroe TA, Ballesteros JA, Orti G. **2013**. Addressing gene tree discordance and non-stationarity to resolve a multi-locus phylogeny of the flatfishes (Teleostei: Pleuronectiformes). *Systematic Biology* 62(5): 763–785. http://dx.doi.org/10.1093/sysbio/syt039

Betancur-R R, Wiley EO, Arratia G, Acero A, Bailly N, Miya M, Lecointre G, Orti G. 2017. Phylogenetic classification of bony fishes. *BMC Evolutionary Biology* 17(art.162): 1–40. https://doi.org/10.1186/s12862-017-0958-3

Bhattathiri PMA, Pant A, Sawant S, Gauns M, Matondkar SGP, Mohanraju R. **1996**. Phytoplankton production and chlorophyll distribution in the eastern and central Arabian Sea in 1994–1995. *Current Science* 71: 857–862.

Bianchi G. **1985**. *Field guide to the commercial marine and brackish-water species of Pakistan*. Rome: FAO. 200 pp.

Bianchi G. **1985**. *Field guide to the commercial marine and brackish-water species of Tanzania*. Rome: FAO. 199 pp.

Bianchi G, Carpenter KE. **1993**. In: Bianchi *et al.* (eds). [see next entry]

Bianchi G, Carpenter KE, Roux J-P, Molloy FJ, Boyer D, Boyer HJ (eds). 1993. FAO species identification field guide for fishery purposes. The living marine resources of Namibia.
Rome: FAO. 250 pp., Pls. 8.

Bianchi G, Carpenter KE, Roux J-P, Molloy FJ, Boyer D, Boyer HJ (eds). 1999. FAO species identification guide for fishery purposes. Field guide to the living marine resources of Namibia. Rome: FAO. 265 pp., Pls. 11.

Bianconi GG. 1846. Lettera [sul Ostracion fornasini, n. sp. de pesce del Mosambico]. Nuovi Annali delle Scienze naturali Bologna (Series 2) 5: 113–115.

Bianconi GG. **1850–1862**. *Specimina zoologica mosambicana quibus vel novae vel minus notae animalium species illustrantur*. Bononiae. 363 pp. [Fasicles I (1850: pp. 1–28), X (1855: pp. 215–229), XII (1857: pp. 243–258), XIII (1859: pp. 259–270), XIV (1859: pp. 272–282), and XV (1862: pp. 283–293) involve fishes]

Bianconi GG. 1854. [Specimina zoologica mosambicana. Fasciculus VIII: Pisces novi Mozambicani.] Rendiconto delle Sessioni dell'Accademia delle Scienze dell'Istituto di Bologna 1853–1854: 68–69.

Bianconi GG. 1855. Specimina zoologica mosambicana. Fasciculus VIII. Memorie della Accademia delle Scienze dell'Istituto di Bologna 6: 139–151.

Bianconi GG. 1857. [Specimina zoologica mosambicana, Fasciculus X. Pisces novi Mozambicani.] *Rendiconto delle Sessioni dell'Accademia delle Scienze dell'Istituto di Bologna* 1856–1857: 99–102. Bianconi GG. **1858**. [Specimina zoologica mosambicana. Fasciculus XI. Pisces novi Mozambicani.] *Rendiconto delle Sessioni dell'Accademia delle Scienze dell'Istituto di Bologna* 1856–1857: 52–53.

Bianconi GG. **1858**. De Piscibus. In: Specimina zoologica mosambicana. Fasciculus XI. *Memorie della Accademia delle Scienze dell'Istituto di Bologna* 9: 435–444.

Bigelow HB, Schroeder WC. **1948**. Cyclostomes. Sharks. In: Fishes of the western North Atlantic. *Memoir of the Sears Foundation for Marine Research* 1(part 1): 29–576.

Bigelow HB, Schroeder WC. **1948**. New genera and species of batoid fishes. *Journal of Marine Research* 7: 543–566.

Bigelow HB, Schroeder WC. **1950**. New and little known cartilaginous fishes from the Atlantic. *Bulletin of the Museum of Comparative Zoology* 103(7): 385–408.

Bigelow HB, Schroeder WC. **1951**. Three new skates and a new chimaerid fish from the Gulf of Mexico. *Journal of the Washington Academy of Sciences* 41(12): 383–392.

Bigelow HB, Schroeder WC. 1953. Sawfishes, guitarfishes, skates and rays (pp. 1–514); Chimaeroids (pp. 515–562). In: Fishes of the western North Atlantic. *Memoir of the Sears Foundation for Marine Research* 1(part 2): 1–588.

Bigelow HB, Schroeder WC. **1957**. A study of the sharks of the suborder Squaloidea. *Bulletin of the Museum of Comparative Zoology* 117(1): 1–150.

Bigelow HB, Schroeder WC. **1965**. A further account of batoid fishes from the western Atlantic. *Bulletin of the Museum of Comparative Zoology* 132(5): 445–477.

Bigelow HB, Schroeder WC, Springer S. 1943. A new species of Carcharhinus from the western Atlantic. Proceedings of the New England Zoölogical Club 22: 69–74.

Bigelow HB, Schroeder WC, Springer S. 1953. New and little known sharks from the Atlantic and from the Gulf of Mexico. *Bulletin of the Museum of Comparative Zoology* 109(3): 213–276.

Bijukumar A, Abraham KM, Soumya D. 2008. Morphometry and meristics of longnose seahorse, *Hippocampus trimaculatus* (Actinopterygii: Syngnathidae), from Kerala, south-west coast of India. *Acta Ichthyologica et Piscatoria* 38(2): 149–155. http://dx.doi.org/10.3750/AIP2008.38.2.11

Bijukumar A, Deepthi GR. **2009**. Mean trophic index of fish fauna associated with trawl bycatch of Kerala, southwest coast of India. *Journal of the Marine Biological Association of India* 51(2): 145–157.

Bijukumar A, Sushama S. **2000**. Ichthyofauna of Ponnani estuary, Kerala. *Journal of the Marine Biological Association of India* 42(1–2): 182–189.

Bilecenoglu M. 2006. Status of the genus Macroramphosus (Syngnathiformes: Centriscidae) in the eastern Mediterranean Sea. Zootaxa 1273: 55–64.

Bilecenoglu M, Taskavak E, Mater S, Kaya M. **2002**. Checklist of the marine fishes of Turkey. *Zootaxa* 113: 1–194.

Bineesh KK, Akhilesh KV, Abdussamad EM, Pillai NGK. 2013. Chelidoperca maculicauda, a new species of perchlet (Teleostei: Serranidae) from the Arabian Sea. aqua, International Journal of Ichthyology 19(2): 71–78.

Bineesh KK, Akhilesh KV, Abdussamad EM, Pillai NKG, Thiel R, Jena JK, Gopalakrishnan A. 2014. Redescriptions of *Chelidoperca investigatoris* (Alcock, 1890) and *Chelidoperca occipitalis* Kotthaus, 1973 (Perciformes: Serranidae) from the south-west coast of India. *Indian Journal of Fisheries* 61(4): 117–122.

Bineesh KK, Akhilesh KV, Gomon M, Abdussamad EM, Pillai NGK, Gopalakrishnan A. 2014. Redescription of *Chlorophthalmus corniger*, a senior synonym of *Chlorophthalmus bicornis* (Family: Chlorophthalmidae). *Journal of Fish Biology* 84(2): 513–522. http://dx.doi. org/10.1111/jfb.12305

Bineesh KK, Akhilesh KV, Gopalakrishnan A, Jena JK. 2014. *Plectranthias alcocki*, a new anthiine fish species (Perciformes: Serranidae) from the Arabian Sea, off southwest India. *Zootaxa* 3785(3): 490–496. http://dx.doi.org/10.11646/zootaxa.3785.3.10

Bineesh KK, Akhilesh KV, Rajool Shanis CP, Abdussamad EM, Pillai NGK. **2012**. First report of longfin escolar, *Scombrolabrax heterolepis* (Perciformes: Scombrolabracidae) from Indian waters. *Marine Biodiversity Records* 5(e77): 1–3. http://dx.doi.org/10.1017/S1755267212000656

Bineesh KK, Greenfield DW. 2011. Redescription of the Indian toadfish *Perulibatrachus aquilonarius* Greenfield (Teleostei: Batrachoididae). *Zootaxa* 2886: 63–68. http://dx.doi. org/10.11646/zootaxa.2886.1.5

Bineesh KK, Manju S, Akhilesh KV, Pillai NGK. **2012**. A preliminary study on the length-weight relationship of *Sacura boulengeri* (Heemstra, 1973) from Indian waters. *Turkish Journal of Zoology* 36(2): 267–270. http://dx.doi. org/10.3906/zoo-0907-91

Bineesh KK, Sajeela KA, Akhilesh KV, Pillai NGK, Abdussamad EM. 2011. Redescription of Sphenanthias whiteheadi Talwar (Perciformes: Cepolidae) with DNA barcodes from the southern coasts of India. Zootaxa 3098: 64–68. http://dx.doi.org/10.11646/zootaxa.3098.1.7

Bini G. 1969. Atlante dei pesci delle coste italiane. Vol. 7: Perciformi (Ofidioidei ... Dactilopteroidei). Milano: Mondo Sommerso Editrice. 196 pp.

Birstein VJ. **1993**. Sturgeons and paddlefishes: threatened fishes in need of conservation. *Conservation Biology* 7: 773–787.

Blaber SJM. 1977. The feeding ecology and relative abundance of mullet (Mugilidae) in Natal and Pondoland estuaries.*Biological Journal of the Linnean Society* 9(3): 259–275.

Blaber SJM. **1978**. Fishes of the Kosi system. *Lammergeyer* 24: 28–41.

Blaber SJM. **1979**. The biology of filter feeding teleosts in Lake St Lucia, Zululand. *Journal of Fish Biology* 15(1): 37–59.

Blaber SJM, Cyrus DP, Whitfield AK. **1981**. The influence of zooplankton food resources on the morphology of the estuarine clupeid *Gilchristella aestuarius* (Gilchrist, 1914). *Environmental Biology of Fishes* 6: 351–355.

Blaber SJM, Whitfield AK. 1977. The feeding ecology of juvenile mullet (Mugilidae) in south-east African estuaries. *Biological Journal of the Linnean Society* 9(3): 277–284.

Blache J. **1967**. Contribution à la connaissance des poissons Anguilliformes de la côte occidentale d'Afrique. 4e note: le genre *Lycodontis* McClelland. 1844. *Bulletin de l'Institut Fondamental d'Afrique Noire* (Série A) 29(3): 1122–1187.

Blache J. **1967**. Contribution à la connaissance des poissons Anguilliformes de la côte occidentale d'Afrique. 5e note: le genre *Gymnothorax* Bloch 1795. *Bulletin de l'Institut Fondamental d'Afrique Noire* (Série A) 29(4): 1695–1705.

Blache J. **1975**. Contribution à la connaissance des poissons Anguilliformes de la côte occidentale d'Afrique. 15e note. *Bulletin de l'Institut Fondamental d'Afrique Noire* (Série A) 37(3): 708–740.

Blache J, Cadenat J, Stauch A. **1970**. Clés de détermination des poissons de mer signalés dans l'Atlantique oriental (entre le 20e parallele N. et Ie 15e parallele S.). *Faune tropicale* 18: 1–479.

Blache J, Saldanha L. 1972. Contribution à la connaissance des Poissons Anguilliformes de la côte occidentale d'Afrique. 12e note: les genres *Pisodonophis, Ophichthus, Brachysomophis* et *Ophisurus* (Fam. des Ophichthidae). *Bulletin de l'Institut Fondamental d'Afrique Noire* (Série A) 34(1): 127–159.

Blainville H de. **1816**. Prodrome d'une nouvelle distribution systématique du règne animal. *Bulletin de la Société Philomathique de Paris* 8: 105–112 [sic for 113–120] + 121–124.

Blainville H de. 1825. Vertébrés. Class V. Poissons. In: Vieillot L et al. (eds) Faune Française; ou, histoire naturelle, générale et particulière des animaux qui se trouvent en France. Livr. 13 & 14. Paris. 96 pp., Pls. 1–24.

Blanc M, Hureau J-C. 1968. Catalogue critique des types de poissons du Muséum National d'Histoire Naturelle (Poissons à joues cuirassées). *Publications diverses du Muséum National d'Histoire Naturelle* No. 23: 1–71.

Blanc M, Hureau J-C. 1972. Catalogue critique des types de poissons du Muséum National d'Histoire Naturelle. (Suite) (Mugiliformes et Polynémiformes). *Bulletin du Muséum National d'Histoire Naturelle* (Série 3) 15 [for 1971]: 673–734.

Blanc M, Paulian P. 1957. Poissons des iles Saint-Paul et Amsterdam. Mémoires de l'Institut Scientifique de Madagascar (Série F: Océanographie) 1: 325–335.

Blanc M, Postel E. 1958. Sur une petite collection de poissons de la Réunion. Mémoires de l'Institut Scientifique de Madagascar (Série F: Océanographie) 1: 367–376. Bleeker P. **1845**. Bijdragen tot de geneeskundige Topographie van Batavia. Generisch overzicht der Fauna. *Natuur-en Geneeskundig Archief voor Neêrlandsch Indië* 2(3): 505–528.

Bleeker P. 1846. Overzigt der Siluroïden, welke te Batavia voorkomen. Natuur-en Geneeskundig Archief voor Neêrlandsch Indië 3(2): 135–184.

Bleeker P. 1846. Siluroideorum bataviensium species nuperrime detectae. Natuur-en Geneeskundig Archief voor Neêrlandsch Indië 3(2): 284–293.

Bleeker P. **1847**. Overzigt der te Batavia voorkomende Gladschubbige Labroïeden, met beschrijving van 11 nieuwe species. *Verhandelingen van het Bataviaasch Genootschap van Kunsten en Wetenschappen* 22(art.3): 1–64.

Bleeker P. **1848**. A contribution to the knowledge of the ichthyological fauna of Sumbawa. *Journal of the Indian Archipelago and Eastern Asia* (Singapore) 2(9): 632–639.

Bleeker P. **1849**. A contribution to the knowledge of the ichthyological fauna of Celebes. *Journal of the Indian Archipelago and Eastern Asia* (Singapore) 3(1): 65–74.

Bleeker P. 1849. Bijdrage tot de kennis der Scleroparei van den Soenda-Molukschen Archipel. Verhandelingen van het Bataviaasch Genootschap van Kusten en Wetenschappen 22(art. 5): 1–10.

Bleeker P. 1849. Bijdrage tot de kennis der Blennioïden en Gobioïden van der Soenda-Molukschen Archipel, met beschrijving van 42 nieuwe soorten. Verhandelingen van het Bataviaasch Genootschap van Kunsten en Wetenschappen 22(art.6): 1–40.

Bleeker P. **1849**. Bijdrage tot de kennis der ichthyologische fauna van het eiland Bali, met beschrijving van eenige nieuwe species. *Verhandelingen van het Bataviaasch Genootschap van Kunsten en Wetenschappen* 22(art.7): 1–11.

Bleeker P. **1849**. Bijdrage tot de kennis der ichthyologische fauna van het eiland Madura, met beschrijving van eenige neiuwe soorten. *Verhandelingen van het Bataviaasch Genootschap van Kunsten en Wetenschappen* 22(art.8): 1–16.

Bleeker P. 1849. Bijdrage tot de kennis der Percoïden van den Malayo-Molukschen Archipel, met beschrijving van 22 nieuwe soorten. Verhandelingen van het Bataviaasch Genootschap van Kunsten en Wetenschappen 22(art.13): 1–64.

Bleeker P. 1850. Bijdrage tot de kennis der Maenoïden van den Soenda-Molukschen Archipel. Verhandelingen van het Bataviaasch Genootschap van Kunsten en Wetenschappen 23(art.7): 1–13.

Bleeker P. **1850**. Bijdrage tot de kennis der Blootkakige visschen van den Soenda-Molukschen Archipel. *Verhandelingen van het Bataviaasch Genootschap van Kunsten en Wetenschappen* 24(art.10): 1–26.

Bleeker P. **1850**. Over eenige nieuwe soorten van *Belone* en *Hemiramphus* van Java. *Natuurkundig Tijdschrift voor Nederlandsch Indië* 1: 93–95. Bleeker P. **1850**. Over drie nieuwe soorten van *Tetraödon* van den Indischen Archipel. *Natuurkundig Tijdschrift voor Nederlandsch Indië* 1: 96–97.

Bleeker P. **1851**. Over eenige nieuwe soorten van Blennioïden en Gobioïden van den Indischen Archipel. *Natuurkundig Tijdschrift voor Nederlandsch Indië* 1: 236–258.

Bleeker P. **1851**. Over eenige nieuwe geslachten en soorten van Makreelachtige visschen van der Indischen Archipel. *Natuurkundig Tijdschrift voor Nederlandsch Indië* 1: 341–372.

Bleeker P. **1851**. Over eenige nieuwe soorten van Pleuronectoïden van den Indischen Archipel. *Natuurkundig Tijdschrift voor Nederlandsch Indië* 1: 401–416.

Bleeker P. 1851. Nieuwe bijdrage tot de kennis der Percoïdei, Scleroparei, Sciaenoïdei, Sparoidei, Maenoïdei, Chaetodontoïdei en Scomberoïdei van den Soenda-Molukschen Archipel. Natuurkundig Tijdschrift voor Nederlandsch Indië 2: 163–179.

Bleeker P. **1851**. Nieuwe bijdrage tot de kennis der ichthyologische fauna van Celebes. *Natuurkundig Tijdschrift voor Nederlandsch Indië* 2: 209–224.

Bleeker P. **1851**. Bijdrage tot de kennis der ichthyologische fauna van de Banda-eilanden. *Natuurkundig Tijdschrift voor Nederlandsch Indië* 2: 225–261.

Bleeker P. **1851**. Vijfde bijdrage tot de kennis der ichthyologische fauna van Borneo, met beschrijving van eenige nieuwe soorten van zoetwatervisschen. *Natuurkundig Tijdschrift voor Nederlandsch Indië* 2: 415–442.

Bleeker P. **1851**. Bijdrage tot de kennis der ichthyologische fauna van Riouw. *Natuurkundig Tijdschrift voor Nederlandsch Indië* 2: 469–497.

Bleeker P. **1851**. Bijdrage tot de kennis der Makreelachtige visschen van den Soenda-Molukschen Archipel. *Verhandelingen van het Bataviaasch Genootschap van Kunsten en Wetenschappen* 24(art.5): 1–93. [preprint in 1851, published 1852]

Bleeker P. 1851. Bijdrage tot de kennis der Balistini en Ostraciones van den Indischen Archipel. Verhandelingen van het Bataviaasch Genootschap van Kunsten en Wetenschappen 24(art.11): 1–38.

Bleeker P. **1852**. Bijdrage tot de kennis der Chirocentroïdei, Lutodeiri, Butirini, Elopes, Notopteri, Salmones, Echeneoïdei, en Ophidini van den Soenda-Molukschen Archipel. *Verhandeling van het Bataviaasch Genootschap van Kunsten en Wetenschappen* 24(art.12): 1–24.

Bleeker P. **1852**. Bijdrage tot de kennis der ichthyologische fauna van Singapore. *Natuurkundig Tijdschrift voor Nederlandsch Indië* 3: 51–86.

Bleeker P. 1852. Bijdrage tot de kennis der ichthyologische fauna van de Moluksche Eilanden. Visschen van Amboina en Ceram. Natuurkundig Tijdschrift voor Nederlandsch Indië 3: 229–309. Bleeker P. **1852**. Bijdrage tot de kennis der ichthyologische fauna van het eiland Banka. *Natuurkundig Tijdschrift voor Nederlandsch Indië* 3: 443–460.

Bleeker P. **1852**. Nieuwe bijdrage tot de kennis der ichthyologische fauna van Amboina. *Natuurkundig Tijdschrift voor Nederlandsch Indië* 3: 545–568.

Bleeker P. 1852. Diagnostische beschrijvingen van nieuwe of weinig bekende vischsoorten van Sumatra. Tiental I–IV. Natuurkundig Tijdschrift voor Nederlandsch Indië 3: 569–608.

Bleeker P. **1852**. Nieuwe visschen van Banda Neira. Natuurkundig Tijdschrift voor Nederlandsch Indië 3: 643–646.

Bleeker P. 1852. Bijdrage tot de kennis der Plagiostomen van den Indischen Archipel. Verhandelingen van het Bataviaasch Genootschap van Kunsten en Wetenschappen 24(art.12): 1–92.

Bleeker P. 1853. Bijdrage tot de kennis der Muraenoïden en Symbranchoïden van den Indischen Archipel. Verhandeling van het Bataviaasch Genootschap van Kunsten en Wetenschappen 25(art.5): 1–76.

Bleeker P. **1853**. Bijdrage tot de kennis der Troskieuwige visschen van der Indischen Archipel. *Verhandelingen van het Bataviaasch Genootschap van Kunsten en Wetenschappen* 25(art.6): 1–38.

Bleeker P. **1853**. Nieuwe bijdrage tot de kennis der ichthyologische fauna van Ceram. *Natuurkundig Tijdschrift voor Nederlandsch Indië* 3(5): 689–714.

Bleeker P. **1853**. Nieuwe bijdrage tot de kennis der ichthyologische fauna van het eiland Banka. *Natuurkundig Tijdschrift voor Nederlandsch Indië* 3(5): 715–738.

Bleeker P. **1853**. Derde bijdrage tot de kennis der ichthyologische fauna van Celebes. *Natuurkundig Tijdschrift voor Nederlandsch Indië* 3(5): 739–782.

Bleeker P. **1853**. Derde bijdrage tot de kennis der ichthyologische fauna van Amboina. *Natuurkundig Tijdschrift voor Nederlandsch Indië* 4: 91–130.

Bleeker P. 1853. Exocoetus hexazona, eene nieuwe soort van Banka. Natuurkundig Tijdschrift voor Nederlandsch Indië 4: 206–207.

Bleeker P. 1853. Diagnostische beschrijvingen van nieuwe of weinig bekende vischoorten van Sumatra. Tiental V–X. *Natuurkundig Tijdschrift voor Nederlandsch Indië* 4: 243–302.

Bleeker P. 1853. Diagnostische beschrijvingen van nieuwe of weinig bekende vischsoorten van Batavia. Tiental I–VI. Natuurkundig Tijdschrift voor Nederlandsch Indië 4: 451–516.

Bleeker P. 1853. Nieuwe bijdrage tot de kennis der ichthyologische fauna van Ternate en Halmaheira (Gilolo). Natuurkundig Tijdschrift voor Nederlandsch Indië 4: 595–610.

Bleeker P. **1853**. Bijdrage tot de kennis der ichthyologische fauna van Solor. *Natuurkundig Tijdschrift voor Nederlandsch Indië* 5: 67–96. Bleeker P. **1853**. Derde bijdrage tot de kennis der ichthyologische fauna van Ceram. *Natuurkundig Tijdschrift voor Nederlandsch Indië* 5: 233–248.

Bleeker P. **1853**. Vierde bijdrage tot de kennis der ichthyologische fauna van Amboina. *Natuurkundig Tijdschrift voor Nederlandsch Indië* 5: 317–352. [also cited as 1854]

Bleeker P. **1853**. Nieuwe tientallen diagnostische beschrijvingen van nieuwe of weinig bekende vischsoorten van Sumatra. *Natuurkundig Tijdschrift voor Nederlandsch Indië* 5: 495–534.

Bleeker P. **1853**. Nalezingen op de ichthyologie van Japan. *Verhandeling van het Bataviaasch Genootschap van Kunsten en Wetenschappen* 25(art.7): 1–56.

Bleeker P. **1853**. Nalezingen op de ichthyologische fauna van Bengalen en Hindostan. *Verhandeling van het Bataviaasch Genootschap van Kunsten en Wetenschappen* 25(art.8): 1–166.

Bleeker P. **1854**. Derde bijdrage tot de kennis der ichthyologische fauna van de Banda-eilanden. *Natuurkundig Tijdschrift voor Nederlandsch Indië* 6: 89–114.

Bleeker P. **1854**. Species piscium bataviensium novae vel minus cognitae. *Natuurkundig Tijdschrift voor Nederlandsch Indië* 6: 191–202.

Bleeker P. **1854**. Nieuwe bijdrage tot de kennis der ichthyologische fauna van Timor. *Natuurkundig Tijdschrift voor Nederlandsch Indië* 6: 203–214.

Bleeker P. **1854**. Bijdrage tot de kennis der ichthyologische fauna van het eiland Flores. *Natuurkundig Tijdschrift voor Nederlandsch Indië* 6: 311–338.

Bleeker P. **1854**. Faunae ichthyologicae japonicae species novae. Species Novae. *Natuurkundig Tijdschrift voor Nederlandsch Indië* 6: 395–426.

Bleeker P. **1854**. Vijfde bijdrage tot de kennis der ichthyologische fauna van Amboina. *Natuurkundig Tijdschrift voor Nederlandsch Indië* 6: 455–508.

Bleeker P. **1854**. Bijdrage tot de kennis der ichthyologische fauna van de Kokos-eilanden. *Natuurkundig Tijdschrift voor Nederlandsch Indië* 7: 37–48.

Bleeker P. **1854**. Ichthyologische waarnemingen, gedaan op verschillende reizen in de residentie Banten. *Natuurkundig Tijdschrift voor Nederlandisch Indië* 7: 309–326.

Bleeker P. **1854**. Specierum piscium javanensium novarum vel minus cognitarum diagnoses adumbratae. *Natuurkundig Tijdschrift voor Nederlandsch Indië* 7: 415–448.

Bleeker P. **1854**. Zesde bijdrage tot de kennis der ichthyologische fauna van Celebes. *Natuurkundig Tijdschrift voor Nederlandsch Indië* 7: 449–452.

Bleeker P. **1854**. Bijdrage tot de kennis der Troskieuwige visschen van den Indischen Archipel. *Verhandelingen van het Bataviaasch Genootschap van Kunsten en Wettenschappen* 25(art.6): 1–30. Bleeker P. **1854**. Nalezingen op de ichthyologische fauna van Bengalen en Hindostan. *Verhandelingen van het Bataviaasch Genootschap van Kunsten en Wetenschappen* 25(art.8): 1–164.

Bleeker P. 1854. Bijdrage tot de kennis der Sphyraenoïden van den Indischen Archipel. Verhandelingen van het Bataviaasch Genootschap van Kunsten en Wetenschappen 26(art.3): 1–22.

Bleeker P. **1855**. Derde bijdrage tot de kennis der ichthyologische fauna van de Kokos-eilanden. *Natuurkundig Tijdschrift voor Nederlandsch Indië* 8: 169–180.

Bleeker P. **1855**. Bijdrage tot de kennis der ichthyologische fauna van de Batoe eilanden. *Natuurkundig Tijdschrift voor Nederlandsch Indië* 8: 305–328.

Bleeker P. 1855. Visschen van de Duizendeilanden. Natuurkundig Tijdschrift voor Nederlandsch Indië 8: 344.

Bleeker P. **1855**. Zesde bijdrage tot de kennis der ichthyologische fauna van Amboina. *Natuurkundig Tijdschrift voor Nederlandsch Indië* 8: 391–434.

Bleeker P. **1855**. Negende bijdrage tot de kennis der ichthyologische fauna van Borneo. Zoetwatervisschen van Pontianak en Bandjermasin. *Natuurkundig Tijdschrift voor Nederlandsch Indië* 9: 415–430.

Bleeker P. **1855**. Bijdrage tot de kennis der ichthyologische fauna van het eiland Groot-obij. *Natuurkundig Tijdschrift voor Nederlandsch Indië* 9: 431–438.

Bleeker P. **1855**. Derde bijdrage tot de kennis der ichthyologische fauna van Batjan. *Natuurkundig Tijdschrift voor Nederlandsch Indië* 9: 491–504.

Bleeker P. 1855. Over eenige visschen van Van Diemensland. Verhandelingen der Koninklijke Akadamie van Wetenschappen (Amsterdam) 2(art.7): 1–31.

Bleeker P. 1856. Beschrijvingen van nieuwe en weinig bekende vischsoorten van Amboina, verzameld op eene reis door den Molukschen Archipel gedaan in het gevolg van dan Gouverneur Generaal Duymaer Van Twist, in September en Oktober 1855. Acta Societatis Regiae Scientarum Indo-Neêrlandicae 1: 1–76.

Bleeker P. 1856. Beschrijvingen van nieuwe of weinig bekende vischsoorten van Manado en Makassar, grootendeels verzameld op eene reisnaar den Molukschen Archipel in het gevolg van den Gouverneur Generaal Duymaer van Twist. Acta Societatis Regiae Scientiarum Indo-Neêrlandicae 1(6): 1–80.

Bleeker P. **1856**. Tweede bijdrage tot de kennis der ichthyologische fauna van het eiland Bintang. *Natuurkundig Tijdschrift voor Nederlandsch Indië* 10: 345–356.

Bleeker P. 1856. Zevende bijdrage tot de kennis der ichthyologische fauna van Ternate. Natuurkundig Tijdschrift voor Nederlandsch Indië 10: 357–386.

Bleeker P. 1856. Carcharias (Prionodon) amblyrhynchus, eene nieuwe haaisoort, gevangen nabij bet eiland Solombo. Natuurkundig Tijdschrift voor Nederlandsch Indië 10: 467–468. Bleeker P. **1856**. Verslag omtrent eenige vischsoorten gevangen aan de Zuidkust van Malang in Oost-Java. *Natuurkundig Tijdschrift voor Nederlandsch Indië* 11: 81–92.

Bleeker P. **1856**. Vijfde bijdrage tot de kennis der ichthyologische fauna van de Banda-eilanden. *Natuurkundig Tijdschrift voor Nederlandsch Indië* 11: 93–110.

Bleeker P. **1856**. Bijdrage tot de kennis der ichthyologische fauna van het eiland Boeroe. *Natuurkundig Tijdschrift voor Nederlandsch Indië* 11: 383–414.

Bleeker P. **1856**. Achtste bijdrage tot de kennis der ichthyologische fauna van Ternate. *Natuurkundig Tijdschrift voor Nederlandsch Indië* 12: 191–210.

Bleeker P. **1857**. Achtste bijdrage tot de kennis der vischfauna van Amboina. *Acta Societatis Regiae Scientarum Indo-Neêrlandicae* 2(7): 1–102.

Bleeker P. **1857**. Vierde bijdrage tot de kennis der ichthyologische fauna van Japan. *Acta Societatis Regiae Scientiarum Indo-Neêrlandicae* 3(art.10): 1–46.

Bleeker P. **1857**. Tweede bijdrage tot de kennis der ichthyologische fauna van Boeroe. *Natuurkundig Tijdschrift voor Nederlandsch Indië* 13: 55–82.

Bleeker P. 1857. Bijdrage tot de kennis ichthyologische fauna van de Sangi-eilanden. Natuurkundig Tijdschrift voor Nederlandsch Indië 13: 369–380.

Bleeker P. **1857**. Index descriptionum specierum piscium Bleekerianarum in voluminibus I ad XIV diarii societatis scientiarum indo-Batavae. Natuurkundig Tijdschrift voor Nederlandsch Indië 14: 447–486.

Bleeker P. **1858**. Zesde bijdrage tot de kennis der vischfauna van Sumatra. Visschen van Padang, Troessan, Priaman, Sibogha en Palembang. *Acta Societatis Regiae Scientiarum Indo-Neêrlandicae* 3(art.9): 1–50.

Bleeker P. **1858**. Tiende bijdrage tot de kennis der vischfauna van Celebes. *Acta Societas Scientifica Indo-Neêrlandicae* 3(art.11): 1–16.

Bleeker P. **1858**. Negende bijdrage tot de kennis der vischfauna van Amboina. *Acta Societatis Regiae Scientiarum Indo-Neêrlandicae* 3(art.14): 1–6.

Bleeker P. **1858**. Twaalfde bijdrage tot de kennis der vischfauna van Borneo. Visschen van Sinkawang. *Acta Societatis Regiae Scientiarum Indo-Neêrlandicae* 5(art.7): 1–10.

Bleeker P. **1858**. Vijfde bijdrage tot de kennis der ichthyologische fauna van Japan. *Acta Societatis Regiae Scientiarum Indo-Neêrlandicae* 5(art.9): 1–12.

Bleeker P. **1858**. De visschen van den Indischen Archipel. Beschreven en toegelicht. Siluri. *Acta Societatis Regiae Scientiarum Indo-Neêrlandicae* 4: i–xii + 1–370. [Also: *Ichthyologiae Archipelagi Indici Prodromus*, Vol. 1. Siluri. Lange & Co., Batavia. xii + 370.]

Bleeker P. **1858**. Bijdrage tot de kennis der vischfauna van den Goram-Archipel. *Natuurkundig Tijdschrift voor Nederlandsch Indië* 15: 197–218.
Bleeker P. **1858**. Vijfde bijdrage tot de kennis der ichthyologische fauna van de Kokos-eilanden. *Natuurkundig Tijdschrift voor Nederlandsch Indië* 15: 457–468.

Bleeker P. **1858**. Derde bijdrage tot de kennis der ichthyologische fauna van Bali. *Natuurkundig Tijdschrift voor Nederlandsch Indië* 17: 141–175.

Bleeker P. 1859. Bijdrage tot de kennis der vischfauna van Nieuw-Guinea. Acta Societatis Regiae Scientarum Indo-Neêrlandicae 6: 1–24.

Bleeker P. 1859. Enumeratio specierum piscium hucusque in Archipelago indico observatarum, adjectis habitationibus citationibusque, ubi descriptiones earum recentiores reperiuntur, nec non speciebus Musei Bleekeriani Bengalensibus, Japonicis, Capensibus Tasmanicisque. Acta Societatis Regiae Scientiarum Indo-Neêrlandicae 6: xxxvi + 276.

Bleeker P. **1859**. Over eenige vischsoorten van de Zuidkustwateren van Java. *Natuurkundig Tijdschrift voor Nederlandsch Indië* 19: 329–352. [also seen as 1860]

Bleeker P. 1860. Achtste bijdrage tot de kennis der vischfauna van Sumatra (Visschen van Benkoelen, Priaman, Tandjong, Palembang en Djambi). Acta Societatis Regiae Scientiarum Indo-Neêrlandicae 8 (art.2): 1–88.

Bleeker P. **1860**. Dertiende bijdrage tot de kennis der vischfauna van Borneo. *Acta Societatis Regiae Scientarum Indo-Neêrlandicae* 8(art.4): 1–64.

Bleeker P. 1860. Dertiende bijdrage tot de kennis der vischfauna van Celebes (Visschen van Bonthain, Badjoa, Sindjai, Lagoesi en Pompenoea). Acta Societatis Regiae Scientarum Indo-Neêrlandicae 8(art.7): 1–60.

Bleeker P. 1860. Ichthyologiae Archipelagi Indici Prodromus (Vol. II: Cyprini. Ordo Cyprini. Karpers). Batavia: Lange & Co.

Bleeker P. **1860**. Over eenige vischsoorten van de Kaap de Goede Hoop. *Natuurkundig Tijdschrift voor Nederlandsch Indië* 21: 49–80.

Bleeker P. 1861. Iets over de geslachten der Scaroïden en hunne Indisch-archipelagische soorten. Verslagen en Mededeelingen der Koninklijke Akademie van Wetenschappen (Afdeling Natuurkunde) 12: 228–244.

Bleeker P. **1862**. Conspectus generum Labroideorum analyticus. *Proceedings of the Zoological Society of London* 1861(3): 408–418. [also cited as 1861]

Bleeker P. 1862. Sur quelques genres de la famille des Pleuronectoïdes. Verslagen en Mededeelingen der Koninklijke Akademie van Wetenschappen (Afdeling Natuurkunde) 13: 422–429.

Bleeker P. 1862. Sixième mémoire sur la faune ichthyologique de l'île de Batjan. Verslagen en Mededeelingen der Koninklijke Akademie van Wetenschappen (Afdeling Natuurkunde) 14: 99–112. Bleeker P. **1862–1878**. Atlas ichthyologique des Indes Orientales Néêrlandaises, publié sous les auspices du Gouvernement colonial Néêrlandais (9 vols). Amsterdam: Frédéric Muller.

Bleeker P. **1863**. Onzième notice sur la faune ichthyologique de l'île de Ternate. *Nederlandsch Tijdschrift voor de Dierkunde* 1: 228–238.

Bleeker P. 1863. Mémoire sur les poissons de la côte de Guinée. Natuurkundige Verhandelingen van de Hollandsche Maatschappij der Wetenschappen te Haarlem (Series 2) (18): 1–136.

Bleeker P. **1863**. Sur une nouvelle espèce de *Synaptura* du Cap de Bonne Espérance. *Verslagen en Mededeelingen der Koninklijke Akademie van Wetenschappen* (Afdeling Natuurkunde) 13: 422–429.

Bleeker P. **1863**. Treizième mémoire sur la faune ichthyologique de l'île d'Amboine. *Verslagen en Mededeelingen der Koninklijke Akademie van Wetenschappen* (Afdeling Natuurkunde) 15: 19–29.

Bleeker P. **1864**. Poissons inédits indo-archipélagiques de l'ordre des Murènes. *Nederlandsch Tijdschrift voor de Dierkunde* 2: 38–54.

Bleeker P. **1864**. Sixième notice sur la faune ichthyologique de Siam. *Nederlandsch Tijdschrift voor de Dierkunde* 2: 171–176.

Bleeker P. **1864**. Description de quelques espèces inédites de poissons de l'Archipel des Moluques. *Nederlandsch Tijdschrift voor de Dierkunde* 2: 177–181.

Bleeker P. **1865**. Description de quelques espèces inédites des genres *Pseudorhombus* et *Platophrys* de l'Inde Archipélagique. *Nederlandsch Tijdschrift voor de Dierkunde* 3: 43–50.

Bleeker P. **1865**. Sur les espèces d'exocet de l'Inde Archipélagique. *Nederlandsch Tijdschrift voor de Dierkunde* 3: 105–129.

Bleeker P. **1865**. Description d'une espèce inédite d'exocet découverte par M. François Pollen. *Nederlandsch Tijdschrift voor de Dierkunde* 3: 130–133.

Bleeker P. **1865**. Description du *Narcacion Polleni*, espèce inédite des mers de l'île de la Réunion. *Nederlandsch Tijdschrift voor de Dierkunde* 3: 171–173.

Bleeker P. **1866**. Notice sur le *Cirrhites punctatus* CV. *Nederlandsch Tijdschrift voor de Dierkunde* 3: 174–177.

Bleeker P. **1866**. Description de quelques espèces inédites ou peu connues de Clupéoïdes de l'Inde Archipélagique. *Nederlandsch Tijdschrift voor de Dierkunde* 3: 293–308.

Bleeker P. **1867**. Description de quelques espèces nouvelles de *Gobius* de Madagascar. *Archives Néerlandaises des Sciences Exactes et Naturelles* 2: 403–420.

Bleeker P. 1868. Description de deux espèces nouvelles de Blennioïdes de l'Inde archipélagique. Verslagen en Mededeelingen der Koninklijke Akademie van Wetenschappen (Afdeling Natuurkunde) (Series 2) 2: 278–280. Bleeker P. 1868. Description de deux espèces inédites d'Epinephelus rapportées de l'île de la Réunion par M.M. Pollen et van Dam. Verslagen en Mededeelingen der Koninklijke Akademie van Wetenschappen (Afdeling Natuurkunde) (Series 2) 2: 336–341.

Bleeker P. **1868**. Description de deux espèces inédites de *Choerops. Archives Néerlandaises des Sciences Exactes et Naturelles* 3: 273–277.

Bleeker P. 1869. Description d'une espèce inédite de Caesio de l'île de Nossibé. Verslagen en Mededeelingen der Koninklijke Akademie van Wetenschappen (Afdeling Natuurkunde) (Series 2) 3: 78–79.

Bleeker P. 1869. Description d'une espèce inédite de Chaetopterus de l'île d'Amboine. Verslagen en Mededeelingen der Koninklijke Akademie van Wetenschappen (Afdeling Natuurkunde) (Series 2) 3: 80–85.

Bleeker P. **1869**. Description de deux espèces inédites d'Alticus de Madagascar. *Verslagen en Mededeelingen der Koninklijke Akademie van Wetenschappen* (Afdeling Natuurkunde) (Series 2) 3: 234–236.

Bleeker P. **1869**. Description et figure d'une espèce inédite de Platycéphale. *Verslagen en Mededeelingen der Koninklijke Akademie van Wetenschappen* (Afdeling Natuurkunde) (Series 2) 3: 253–254.

Bleeker P. **1871**. Description de quelques espèces de poissons de l'île de la Réunion et de Madagascar. *Nederlandsch Tijdschrift voor de Dierkunde* 4: 92–105.

Bleeker P. **1872**. Mémoire sur la faune ichthyologique de Chine. *Nederlandsch Tijdschrift voor de Dierkunde* 4: 113–154.

Bleeker P. **1872**. Révision des espèces indo-archipélagiques du groupe des Anthianini. *Nederlandsch Tijdschrift voor de Dierkunde* 4: 155–169.

Bleeker P. 1873. Mededeelinge omtrent eene herziening der Indisch-Archipelagische soorten van Epinephelus, Lutjanus, Dentex en verwante geslachten. Verslagen en Mededeelingen der Koninklijke Akademie van Wetenschappen (Afdeling Natuurkunde) (Series 2) 7: 40–46.

Bleeker P. 1873. Révision des espèces insulindiennes des genres Diapterus et Pentaprion. Verslagen en Mededeelingen der Koninklijke Akademie van Wetenschappen (Afdeling Natuurkunde) (Series 2) 7: 233–255.

Bleeker P. **1873**. Sur les especès indo-archipélagiques d'*Odontanthias* et de *Pseudopriacanthus*. *Nederlandsch Tijdschrift voor de Dierkunde* 4: 235–240.

Bleeker P. 1874. Typi nonnuli generici piscium neglecti.
Verslagen en Mededeelingen der Koninklijke Akademie van Wetenschappen (Afdeling Natuurkunde) (Series 2)
8: 367–371.

Bleeker P. 1874. Notice sur les genres Amblyeleotris, Valenciennesia et Brachyeleotris. Verslagen en Mededeelingen der Koninklijke Akademie van Wetenschappen (Afdeling Natuurkunde) (Series 2) 8: 372–376. Bleeker P. **1874**. Esquisse d'un système naturel des Gobioïdes. *Archives Néerlandaises des sciences exactes et naturelles* 9: 289–331.

Bleeker P. **1874**. Poissons de Madagascar et de l'île de la Réunion des collections de MM. Pollen et van Dam. In: *Recherches sur la faune de Madagascar et de ses dépendences, d'après les découvertes de François P. L. Pollen et D. C. van Dam* (pt 4). Leyden: E.J. Brill. 104 pp.

Bleeker P. **1874**. Révision des espèces indo-archipélagiques du groupe des Apogonini. *Natuurkundige Verhandelingen van de Hollandsche Maatschappij der Wetenschappen te Haarlem* (Series 3) 2(1): 1–82.

Bleeker P. **1874**. Révision des espèces d'*Ambassis* et de Parambassis de l'Inde Archipélagigue. Natuurkundige Verhandelingen van de Hollandsche Maatschappij der Wetenschappen te Haarlem (Series 3) 2(2): 83–106.

Bleeker P. **1874**. Révision des espèces insulindiennes de la famille des Synancéoïdes. *Natuurkundige Verhandelingen van de Hollandsche Maatschappij der Wetenschappen te Haarlem* (Series 3) 2(3): 1–22.

Bleeker P. 1874. Révision des espèces indo-archipelagiques du groupe des Epinephelini et de quelques genres voisins. *Verhandelingen der Koninklijke Akademie van Wetenschappen* (Afdeling Natuurkunde) (Amsterdam) 14: 1–134.

Bleeker P. **1874**. Sur les espèces insulindiennes de la famille des Cirrhitéoïdes. *Verhandelingen der Koninklijke Akademie van Wetenschappen* (Afdeling Natuurkunde) (Amsterdam) 15: 1–20.

Bleeker P. 1875–1878. Atlas ichthyologique des Indes Orientales Néêrlandaises, publié sous les auspices du Gouvernement colonial néêrlandais. Tome IX. Percoïdes III. Amsterdam: Fréderic Muller et Co. pp. 1–80, Pls. 355–360, 363–420.
[Reprinted, Smithsonian Institution Press, Washington, DC and London, 1989]

Bleeker P. **1876**. Mémoire sur les espèces insulindiennes de la famille des Scorpénoïdes. *Verhandelingen der Koninklijke Akademie van Wetenschappen* (Afdeling Natuurkunde) (Amsterdam) 16: 1–100.

Bleeker P. 1876. Genera familiae Scorpaenoideorum conspectus analyticus. Verslagen en Mededeelingen der Koninklije Akademie van Wetenschappen (Afdeling Natuurkunde) (Series 2) 9: 294–300.

Bleeker P. **1876**. Description de quelques espèces inédites de Pomacentroïdes de l'Inde Archipélagique. *Verslagen en Mededeelingen der Koninklijke Akademie van Wetenschappen* (Afdeling Natuurkunde) (Series 2) 10: 384–391.

Bleeker P. 1876. Systema Percarum revisum. Archives Néerlandaises des Sciences Exactes et Naturelles 11(1&2): 247–340. Bleeker P. **1877**. Mémoire sur les chromides marins ou pomacentroïdes de l'Inde archipélagique. *Natuurkundige Verhandelingen van de Hollandsche Maatschappij der Wetenschappen te Haarlem* (Series 3) 2(6): 1–166.

Bleeker P. **1877**. Revision des espèces de *Pempheris* de l'Inde Archipélagique. *Archives Néerlandaises de Sciences Exactes et Naturelles* 12: 42–54.

Bleeker P. 1878. Quatrième mémoire sur la faune ichthyologique de la Nouvelle-Guinée. Archives Néerlandaises des Sciences Exactes et Naturelles 13(3): 35–66.

Bleeker P. **1879**. Contribution à la faune ichthyologique de l'île Maurice. *Verhandelingen der Koninklijke Akademie van Wetenschappen* (Afdeling Natuurkunde) (Amsterdam) 18(art.3): 1–23.

Bleeker P. 1879. Enumération des espèces de poissons actuellement connues du Japon et description de trois espèces inédites. Verhandelingen der Koninklijke Akademie van Wetenschappen (Afdeling Natuurkunde) (Amsterdam) 18(art.6): 1–33.

Bleeker P. 1879. Révision des espèces insulindiennes du genre Platycephalus. Verhandelingen der Koninklijke Akademie van Wetenschappen (Afdeling Natuurkunde) (Amsterdam) 19: 1–31.

Blegvad H, Løppenthin B. 1944. Fishes of the Iranian Gulf. Danish Scientific Investigations in Iran. Part 3. Copenhagen: Einar Munksgaard. 247 pp.

Bliss R. 1883. Descriptions of new species of Mauritian fishes. Transactions of the Royal Society of Arts and Sciences of Mauritius (Série Nouvelle) 13: 45–63.

Bloch ME. **1785–1795**. *Naturgeschichte der ausländischen Fische* (9 vols). Berlin: Auf Kosten des Verfassers, und in Commission in der Buchhandlung der Realschule.

Bloch ME. **1789**. Tvä Utlandfka Fiskar. *Kongliga Vetenskaps Academiens nya Handlingar* (Stockholm) 10: 234–236.

Bloch ME. **1801**. *Histoire naturelle des poissons: avec les figures dessinées d'après nature par Bloch, ouvrage classé par ordres, genres et espèces, d'après le systeme de Linné; avec les caractères génériques par René-Richard Castel (10 vols). Paris.* 

Bloch ME, Schneider JG. 1801. M.E. Blochii, Systema Ichthyologiae iconibus cx illustratum. Post obitum auctoris opus inchoatum absolvit, correxit, interpolavit Jo. Gottlob Schneider, Saxo. Berlin: Berolini. Sumtibus Auctoris Impressum et Bibliopolio Sanderiano Commissum. 584 pp.

Blot J. 1980. La faune ichthyologique des gisements du Monte Bolca (Province de Vérone, Italie). Catalogue systématique présentant l'état actuel des recherches concernant cette faune. Bulletin du Muséum National d'Histoire Naturelle, Paris (Série 4), sect. C 2(4): 339–396.

Blum SD. **1988**. The osteology and phylogeny of the Chaetodontidae (Pisces: Perciformes). PhD dissertation, University of Hawaii, Honolulu. 365 pp. Blum SD. **1989**. Biogeography of the Chaetodontidae: an analysis of allopatry among closely related species. *Environmental Biology of Fishes* 25(1–3): 9–31.

Blyth E. 1852. Report on Ceylon mammals, birds, reptiles and fishes (in Appendix, pp. 37–50). In: Kelaart EF (ed) *Prodromus faunae Zeylanicae, being contributions to the zoology of Ceylon*. Colombo, Ceylon. viii + xxxiii + 197 + 54 pp [Appendix].

Blyth E. **1858**. Report of Curator, Zoological Department, for May, 1858. *Journal and Proceedings of the Asiatic Society of Bengal* 27(3): 267–290.

Blyth E. **1860**. The cartilaginous fishes of lower Bengal. *Journal and Proceedings of the Asiatic Society of Bengal* 29(1): 35–45.

Blyth E. 1860. Report on some fishes received chiefly from the Sitang River and its tributary streams, Tenasserim Provinces. *Journal and Proceedings of the Asiatic Society of Bengal* 29(2): 138–174.

Bocage JVB, see Barbosa du Bocage JV.

Bock KR. **1972**. Preliminary checklist of lagoonal fishes of Diani, Kenya. *Journal of the East African Natural History Society and National Museum* 137: 1–6.

Bock KR. 1996. Checklist of reef fishes of Diani and Galu, Kenya. Journal of East African Natural History 85(1&2): 5–21.

Boddaert P. **1772**. Epistola ... de Chaetodonte diacantho descripto. [In Dutch and Latin] Amstelodamae (Amsterdam, apud M. Magerum).

Boddaert P. **1781**. Beschreibung zweyer merkwürdiger Fische (Sparus palpebratus und Muraena colubrina). Neue Nordische Beyträge zur physikalischen und geographischen Erd- und Völkerbeschreibung, Naturgeschichte und Oekonomie 2: 55–57.

Bogorodsky SV, Alpermann TJ, Mal AO, Gabr MH. **2014**. Survey of demersal fishes from southern Saudi Arabia, with five new records for the Red Sea. *Zootaxa* 3852(4): 401–437. http://dx.doi.org/10.11646/zootaxa.3852.4.1

Bogorodsky S[V], Kovačić M, Larson HK. **2010**. The first records of four gobies (Pisces: Gobiidae) in the Red Sea. *aqua, International Journal of Ichthyology* 16(3): 117–128.

Bogorodsky SV, Kovačić M, Randall JE. **2011**. A new species and three new records of gobiid fishes from the Red Sea. *Cybium* 35(3): 213–222.

https://doi.org/10.26028/cybium/2011-353-005

Bogorodsky SV, Randall JE, Golani D. **2011**. Four new records of shore fishes for the Red Sea, with notes on *Parupeneus heptacanthus* and *Diodon liturosus*. *Zootaxa* 3057: 49–60. http://dx.doi.org/10.11646/zootaxa.3057.1.2

Bogorodsky SV, Randall JE, Krupp F. **In press**. *Fauna of Arabia* 26: *Coastal fishes of the Red Sea*. Springer.

Böhlke EB. **1984**. Catalog of type specimens in the ichthyological collection of the Academy of Natural Sciences of Philadelphia. *Academy of Natural Sciences of Philadelphia Special Publication* 14: 1–246. Böhlke EB. **1989**. Methods and terminology. Anguilliformes and Saccopharyngiformes. In: Fishes of the western North Atlantic. *Memoir of the Sears Foundation for Marine Research* 1(part 9): 1–7.

Böhlke EB. 1997. *Gymnothorax robinsi* (Anguilliformes, Muraenidae), a new dwarf moray with sexually dimorphic dentition from the Indo-Pacific. *Bulletin of Marine Science* 60(3): 648–655.

Böhlke EB. 2000. Notes on the identity of small, brown, unpatterned Indo-Pacific moray eels, with descriptions of three new species (Anguilliformes: Muraenidae). *Pacific Science* 54(4): 395–416.

Böhlke EB, McCosker JE. **1982**. *Monopenchelys*, a new genus, and redescription of the type species, *Uropterygius acutus* Parr (Pisces: Muraenidae). *Proceedings of the Academy of Natural Sciences of Philadelphia* 134: 127–134.

Böhlke EB, Randall JE. 1995. Gymnothorax megaspilus, a new species of moray eel (Anguilliformes: Muraenidae) from southern Oman and Somalia. Notulae Naturae (Academy of Natural Sciences of Philadelphia) 472: 1–5.

Böhlke EB, Randall JE. **1996**. *Siderea flavocula*, a new species of moray eel (Anguilliformes: Muraenidae) from Oman. *Journal of South Asian Natural History* 2(1): 95–101.

Böhlke EB, Randall JE. **2000**. A review of the moray eels (Anguilliformes: Muraenidae) of the Hawaiian Islands, with descriptions of two new species. *Proceedings of the Academy of Natural Sciences of Philadelphia* 150: 203–278.

Böhlke EB, Smith DG. 2002. Muraenidae. Moray eels (pp. 700–718). In: Carpenter KE (ed) FAO species identification guide for fishery purposes. The living marine resources of the western central Atlantic. Vol. 2. Bony fishes part 1 (pp. 601–1374). Rome: FAO. [also cited as 2003]

Böhlke JE. **1953**. A catalogue of the type specimens of recent fishes in the natural history museum of Stanford University. *Stanford Ichthyological Bulletin* 5(art.1): 1–168.

Böhlke JE. **1956**. A synopsis of the eels of the family Xenocongridae (including the Chlopsidae and Chilorhinidae). *Proceedings of the Academy of Natural Sciences of Philadelphia* 108: 61–95.

Böhlke JE. **1967**. The descriptions of three new eels from the tropical west Atlantic. *Proceedings of the Academy of Natural Sciences of Philadelphia* 118(4): 91–108.

Böhlke JE. 1981. Muraenidae [and] Ophichthidae. In: Fischer
W, Bianchi G, Scott WB (eds) FAO species identification
sheets for fishery purposes. Eastern central Atlantic (Fishing
Area 34 and part of 47). Vol 3. Rome: FAO. [unpaginated]

Böhlke JE, Briggs JC. **1953**. The rare cirrhitid fish genus *Oxycirrhites* in American waters. *California Fish and Game* 39(3): 375–380.

Böhlke JE, Randall JE. 1981. Four new garden eels (Congridae, Heterocongrinae) from the Pacific and Indian oceans. Bulletin of Marine Science 31(2): 366–382. Böhlke JE, Smith DG. **1967**. A new xenocongrid eel from the western Indian and western Atlantic Ocean. *Notulae Naturae* (Academy of Natural Sciences of Philadelphia) 408: 1–6.

Böhlke JE, Smith DG. **1968**. A new xenocongrid eel from the Bahamas, with notes on other species in the family. *Proceedings of the Academy of Natural Sciences of Philadelphia* 120(2): 25–43.

Bonaparte CL. **1831**. Saggio di una distribuzione metodica degli animati vertebrati. Prospetto del sistema generale d'Ittiologia. *Giornale Arcadico di Scienze, Lettere, ed Arti* 52: 155–189.

Bonaparte CL. **1832**. *Saggio di una distribuzione metodica degli animali vertebrati a sangue freddo*. Rome: Antonio Boulzaler. 86 pp.

Bonaparte CL. 1832–1841. Inconografia della fauna italica per le quattro classi degli animali vertebrati, Tomo 3, Pesci, Roma, without pagination, 75 puntate, in 30 fasc. 1832:
Fasc. 1 (puntata 1–6); 1833: Fasc. 2–5 (puntata 7–28); 1834:
Fasc.6–11 (puntata 29–58); 1835: Fasc.12–14 (puntata 59– 79); 1836: Fasc. 15–18 (puntata 80–93); 1837: Fasc. 19–21 (puntata 94–103, 105–9); 1838: Fasc. 22–23 (puntata 104, 110–120); 1839: Fasc. 24–26 (puntata 121–135); 1840: Fasc. 27–29 (puntata 136–154); 1841: Fasc. 30 (puntata 155–160).

Bonaparte CL. **1838**. Selachorum tabula analytica. *Nuovi* annali delle scienze naturali e rendiconto dei lavori dell'Accademia della Scienze dell'Instituto di Bologna con appendice agraria 2: 195–214.

Bonaparte CL. **1846**. Sur deux espèces de poissons (*Cubiceps bipinnatus* et *Scarus siculus*). Actes do 7. Congrès de Naples, p. 715.

Bond CE, Uyeno T. **1981**. Remarkable changes in the vertebrae of perciform fish *Scombrolabrax* with notes on its anatomy and systematics. *Japanese Journal of Ichthyology* 28(3): 259–262.

Bonelli FA. **1820**. Description d'une nouvelle espèce de poisson de la Méditerranée appartenant au genre Trachyptère (*T. cristatus*) avec des observations sur les caractères de çe même genre. *Memorie della Reale Accademia delle Scienze di Torino* 24: 485–494.

Bonfiglioli A, Hariri K. **2004**. Small-scale fisheries in Yemen: Social assessment and development prospects. FAO & World Bank. 113 pp.

Bonfil R, Abdallah M. **2004**. *Field identification guide to the sharks and rays of the Red Sea and Gulf of Aden. FAO species identification guide for fishery purposes.* Rome: FAO. 71 pp.

Bonfil R, Meÿer M, Scholl MC, Johnson R, O'Brien S, Oosthuizen H, Swanson S, Kotze D, Paterson M. 2005.
Transoceanic migration, spatial dynamics, and population linkages of white sharks. *Science* 310(5475): 100–103. https://www.science.org/doi/10.1126/science.1114898 Bonnaterre JP. **1788**. *Tableau encyclopédique et méthodique des trois règnes de la nature. Ichthyologie*. Paris: Panckoucke. lvi + 215 pp.

Booth AJ, Walmsley-Hart SA. 2000. Biology of the redspotted tonguesole *Cynoglossus zanzibarensis* (Pleuronectiformes: Cynoglossidae) on the Agulhas Bank, South Africa. *South African Journal of Marine Science* 22(1): 185–197.

Borden WC. 1998. Phylogeny of the unicornfishes (*Naso*, Acanthuridae) based on soft anatomy. *Copeia* 1998(1): 104–113.

Borkenhagen K. **2011**. Range extension of the saddleback silver-biddy *Gerres limbatus* Cuvier, 1830 (Teleostei: Gerreidae) with a new record from an Iranian tributary to the Persian Gulf. *Journal of Applied Ichthyology* 27(1): 150– 152. http://dx.doi.org/10.1111/j.1439-0426.2010.01614.x

Borodin NA. **1934**. About the types of fishes from Mauritius Island, Indian Ocean, in the Museum of Comparative Zoology. *Copeia* 1934(1): 44.

Borsieri C. **1904**. Contribuzione alla conoscenza della fauna ittiologica della colonia Eritrea. *Annali del Museo Civico di Storia Naturale "Giacomo Doria" di Genova* (Série 3) 1(41): 187–220.

Bos AR, Ogwang J, Bariche M, Horoiwa M, Megahed M, Ouf A, Yasuda N. 2020. Anti-Lessepsian migration rectified: the comber *Serranus cabrilla* (L. 1758) existed in the Red Sea prior to the Suez Canal opening. *Marine Biology* 167(art. 126): 1–10. https://doi.org/10.1007/s00227-020-03748-0

Bosc LAG. 1816–1819. [Pisces accounts]. Nouveau Dictionnaire d'Histoire Naturelle. Vols. 1–36 [Vol. 7 (1817)]. Paris.

Bosworth W, Huchon P, McClay K. **2005**. The Red Sea and Gulf of Aden basins. *Journal of African Earth Sciences* 43: 334–378.

Botha L. **1969**. The growth of the Cape hake, *Merluccius capensis*. Department of Industries, Division of Sea Fisheries, Republic of South Africa. [unpaginated]

Botha L. **1971**. Growth and otolith morphology of the Cape hakes *Merluccius capensis* Cast. and *M. paradoxus* Franca. *Investigational Report: Division of Sea Fisheries, South Africa* 97: 32 pp.

Botha L. **1973**. Migrations and spawning behaviour of the Cape hakes. *South African Shipping News and Fishing Industry Reviews* 28: 62–67.

Bottard A. **1887**. Poissons dangereux particuliers aux côtes de la Réunion. *Sciences et Arts*: 163–171.

Boughton DA, Collette BB, McCune AR. **1991**. Heterochrony in jaw morphology of needlefishes (Teleostei: Belonidae). *Systematic Zoology* 40(3): 329–354.

Bouhlel M. 1988. Poissons de Djijbouti. Dubai Printing Press, United Arab Emirates. 416 pp. [In French, reprinted in English by RDA International, Inc. Placerville, California] Boulenger GA. **1888**. An account of the fishes obtained by Surgeon-Major A.S.G. Jayakar at Muscat, east coast of Arabia. *Proceedings of the Zoological Society of London* 1887(pt 4): 653–667.

Boulenger GA. **1889**. Second account of the fishes obtained by Surgeon-Major A.S.G. Jayakar at Muscat, east coast of Arabia. *Proceedings of the Zoological Society of London* 1889(pt 2): 236–246.

Boulenger GA. **1892**. Third account of the fishes obtained by Surgeon-Major A.S.G. Jayakar at Muscat, east coast of Arabia. *Proceedings of the Zoological Society of London* 1892(pt 1): 134–136.

Boulenger GA. **1895**. *Catalogue of the perciform fishes in the British Museum* (2<sup>nd</sup> edition, Vol. 1). London. xix + 394 pp.

Boulenger GA. **1895**. Description of a new eagle-ray from Muscat. *Annals and Magazine of Natural History* (Series 6) 15(86, art.15): 141.

Boulenger GA. **1897**. Descriptions of new fishes from the Mekran coast, Persia. *Annals and Magazine of Natural History* (Series 6) 20(119, art.49): 420–422.

Boulenger GA. **1898**. Descriptions of two new fishes from the coast of Sind. *Annals and Magazine of Natural History* (Series 7) 2(8, art.20): 133–134.

Boulenger GA. **1898**. The flat fishes of Cape Colony. *Marine Investigations in South Africa* no. 1: 1–4. [Published separately in 1898, then in Vol. 1 in 1902]

Boulenger GA. **1898**. Descriptions of two new gobiiform fishes from the Cape of Good Hope. *Marine Investigations in South Africa* no. 2: 3–4. [Published separately in 1898, then as pp. 8–9 in Vol. 1 in 1902]

Boulenger GA. **1899**. Description of a new genus of perciform fishes from the Cape of Good Hope. *Annals of the South African Museum* 1(art.10): 379–380.

Boulenger GA. **1900**. Description of a new sea-horse (*Hippocampus*) from Muscat. *Annals and Magazine of Natural History* (Series 7) 6(31): 51–52.

Boulenger GA. **1900**. Descriptions of new fishes from the Cape of Good Hope. *Marine Investigations in South Africa* no. 8: 10–12. [Published separately in 1900, then as pp. 10–12 in Vol. 1 in 1902]

Boulenger GA. 1900. On a specimen of *Lophotes cepedianus* from the Cape of Good Hope. *Marine Investigations in South Africa* no. 8: 13. [Published separately in 1900, then as p. 13 in Vol. 1 in 1902]

Boulenger GA. **1901**. On some deep-sea fishes collected by Mr. F.W. Townsend in the sea of Oman. *Annals and Magazine of Natural History* (Series 7) 7(39): 261–263.

Boulenger GA. 1901. Description of a new fish of the genus Gobius obtained by Mr. A. Blayney Percival in South Arabia. Proceedings of the Zoological Society of London 1901 1(part 1): 152–154. Boulenger GA. **1902**. Notes on the classification of teleostean fishes. II. On the Berycidae. *Annals and Magazine of Natural History* (Series 7) 9(51): 197–204.

Boulenger GA. **1902**. Description of a new South-African galeid selachian. *Annals and Magazine of Natural History* (Series 7) 10(55): 51–52.

Boulenger GA. **1903**. Descriptions of six new perciform fishes from the coast of Natal. *Annals of the South African Museum* 3(pt 3): 63–67.

Boulenger GA. 1909–1916. Catalogue of the fresh-water fishes of Africa in the British Museum (Natural History) (Vols 1–4).
London: British Museum (Natural History). Vol. 1: 1909, 373 pp.; Vol. 2: 1911, 529 pp.; Vol. 3: 1915, 526 pp.; Vol. 4: 1916, 392 pp. [1964 reprint Codicote: Wheldon & Wesley Ltd]

Bowen BW, Bass AL, Rocha LA, Grant WS, Robertson DR.
2001. Phylogeography of the trumpetfishes (*Aulostomus*): ring species complex on a global scale. *Evolution* 55(5) 1029–1039.

Bradbury MG. **1967**. The genera of batfishes (family Ogcocephalidae). *Copeia* 1967(2): 399–422.

Bradbury MG. **1980**. A revision of the fish genus *Ogcocephalus* with descriptions of new species from the western Atlantic Ocean (Ogcocephalidae; Lophiiformes). *Proceedings of the California Academy of Sciences* (Series 4) 42(7): 229–285.

Braithwaite CJR. 1987. Geology and palaeogeography of the Red Sea region (pp. 22–44). In: Edwards AJ, Heads SM (eds) *Red Sea*. Oxford: Pergamon Press. 441 pp.

Branch GM, Griffiths CL, Branch ML, Beckley LE. **1994**. *Two oceans: A guide to the marine life of southern Africa*. Cape Town: David Philip Publishers. 360 pp.

Branch GM, Griffiths CL, Branch ML, Beckley LE. 1999. *Two oceans: A guide to the marine life of southern Africa* (4<sup>th</sup> impression). Cape Town: David Philip Publishers.
360 pp.

Branch GM, Griffiths CL, Branch ML, Beckley LE. 2005. *Two oceans: A guide to the marine life of southern Africa* (2<sup>nd</sup> edition). Cape Town: David Philip Publishers. 456 pp.

Branch GM, Griffiths CL, Branch ML, Beckley LE. **2010**. *Two oceans: A guide to the marine life of southern Africa* (revised edition). Cape Town: Struik Nature. 456 pp.

Brauer A. **1901**. Über einige von der *Valdivia*-Expedition gesammelte Tiefseefische und ihre Augen. *Sitzungsberichte der Gesellschaft zur Beförderung der Gesamten Naturwissenschaften zu Marburg* 8: 115–130.

Brauer A. 1906. Die Tiefsee-Fische. I. Systematischer Teil. In: Chun C (ed) Wissenschaftliche Ergebnisse der Deutschen Tiefsee-Expedition, auf dem Dampfer Valdivia, 1898–99.
Vol. 15(1). Jena: Fischer. 432 pp., Pls. 16.

Bray DJ. **1994**. Order Zeiformes (pp. 416–427). In: Gomon MF, Glover CJM, Kuiter RH (eds) *The fishes of Australia's south coast*. Adelaide: State Print. 992 pp. Bray DJ. **2015**. Families Zeidae (pp. 1026–1028), Parazenidae (pp. 1033–1034), Cyttidae (pp. 1035–1037), Zeniontidae (pp. 1038–1040), Oreosomatidae (pp. 1041–1048). In: Roberts CD, Stewart AL, Struthers CD (eds) *The fishes of New Zealand*. Volume Three (pp. 581–1152). Wellington: Te Papa Press.

Bray DJ, Hoese DF. 2006. Family Uranoscopidae. Monkfish, stargazers, stonelifters (pp. 1509–1515). In: Hoese DF, Bray DJ, Paxton J, Allen G (eds) *Zoological catalogue of Australia*. Vol. 35. Fishes. Part 3 (pp. 1473–2173). Collingwood: CSIRO Publishing.

Bray DJ, Hoese DF, Paxton JR. **2006**. Family Sparidae. Breams, porgies, seabreams (pp. 1225–1230). In: Hoese DF, Paxton J, Allen G (eds) *Zoological catalogue of Australia*. Vol. 35. Fishes. Part 2 (pp. 671–1472). Collingwood: CSIRO Publishing.

Brazeau MD, Friedman M. **2015**. The origin and early phylogenetic history of jawed vertebrates. *Nature* 520: 490– 497. http://dx.doi.org/10.1038/nature14438

Breder CM Jr. 1928. Scientific results of the second oceanographic expedition of the *Pawnee* 1926.
Nematognathii, Apodes, Isospondyli, Synentognathi, and Thoraeostroci from Panama to lower California with a generic analysis of the Exocoetidae. *Bulletin of the Bingham Oceanographic Collection* (Yale University) 2(art.2): 1–25.

Breder CM Jr. **1938**. A contribution to the life histories of Atlantic Ocean flyingfishes. *Bulletin of the Bingham Oceanographic Collection* (Yale University) 6(art.5): 1–126.

Breder CM Jr. **1946**. An analysis of the deceptive resemblances of fishes to plant parts, with critical remarks on protective coloration, mimicry, and adaptation. *Bulletin of the Bingham Oceanographic Collection* (Yale University) 10(art.2): 1–49.

Breder CM Jr, Rosen DE. **1966**. *Modes of reproduction in fishes*. New York: Natural History Press. 941 pp.

Brevoort JC. **1856**. Notes on some figures of Japanese fish taken from recent specimens by the artists of the U.S. Japan Expedition (pp. 253–288). In: Perry MC (ed) *Narrative of the expedition of an American Squadron to the China Seas and Japan, performed in the years 1852, 1853, and 1854 under the command of Commodore M.C. Perry, United States Navy, by order of the Government of the United States* (Vol. 2). U.S. Senate Ex. Doc. No. 79, 33<sup>rd</sup> Congress, 2<sup>nd</sup> Session. Washington, DC: Beverley Tucker. 414 pp.

Brewster B. **1987**. Eye migration and cranial development during flatfish metamorphosis: a reappraisal (Teleostei: Pleuronectiformes). *Journal of Fish Biology* 31(6): 805–833.

Briggs JC. **1955**. A monograph of the clingfishes (Order Xenopterygii). *Stanford Ichthyological Bulletin* 6: i–iv + 1–224.

Briggs JC. **1962**. A new clingfish of the genus *Lepadichthys* from the New Hebrides. *Copeia* 1962(2): 424–425.

Briggs JC. **1966**. A new clingfish of the genus *Lepadichthys* from the Red Sea. *Bulletin of the Sea Fisheries Research Station, Haifa* 42: 37–40.

Briggs JC. **1969**. A new species of *Lepadichthys* (Gobiesocidae) from the Seychelles, Indian Ocean. *Copeia* 1969(3): 464–466.

Briggs JC. **1974**. *Marine zoogeography*. New York: McGraw-Hill. 480 pp.

Briggs JC. **1976**. A new genus and species of clingfish from the Western Pacific. *Copeia* 1976(2): 339–341.

Briggs JC, Berry FH. **1959**. The Draconettidae – a review of the family with the description of a new species. *Copeia* 1959(2): 123–133.

Briggs JC, Bowen BW. 2012. A realignment of marine biogeographic provinces with particular reference to fish distributions. *Journal of Biogeography* 39(1): 12–30. http://dx.doi.org/10.1111/j.1365-2699.2011.02613.x

Briggs JC, Link G. **1963**. New clingfishes of the genus *Lepadichthys* from the northern Indian Ocean and the Red Sea (Pisces, Gobiesocidae). *Senckenbergiana Biologica* 44(2): 101–105.

Brignon A. **2015**. Le "Pygopterus" bonnardi Agassiz 1833 (Xenacanthidae) de l'Autunien de Muse (Saône-et-Loire) : premier requin paléozoïque découvert en France. *Fossiles*, *Revue française de Paléontologie* 23: 15–20.

Brisout de Barneville CNF. 1846. Note sur les Diodoniens.*Revue Zoologique par la Société Cuvierienne* (Paris)9: 136–143.

Brisout de Barneville CNF. **1847**. Note sur un nouveau genre d'Anguilliformes. *Revue Zoologique par la Société Cuvierienne* (Paris) 10: 219–220.

Brito A. **1991**. *Catalogo de los pesces de las Islas Canarias*. La Laguna: Francisco Lemus. 230 pp.

Brito PM. **1997**. Révision des Aspidorhynchidae (Pisces, Actinopterygii) du Mésozoïque: ostéologie, relations phylogénétiques, données environnementales et biogéographiques. *Geodiversitas* 19(4): 681–772.

Brito PM. **1999**. The caudal skeleton of aspidorhynchids (Actinopterygii, Halecostomi): phylogenetic implications (pp. 249–264). In: Arratia G, Schultze H-P (eds) *Mesozoic Fishes 2 – Systematics and fossil record. Proceedings of the international meeting, Buckow, 1997.* Munich: Verlag Dr. Friedrich Pfeil. 604 pp.

Britz R, Johnson GD. 2002. "Paradox Lost": Skeletal ontogeny of *Indostomus paradoxus* and its significance for the phylogenetic relationships of the Indostomidae (Teleostei, Gasterosteiformes). *American Museum Novitates* 3383: 1–43.

Brooks RD, Maclennan B. **2002**. *The nature of diversity*. Chicago University Press. 676 pp.

Broughton RE. 2010. Phylogeny of teleosts based on mitochondrial genome sequences (pp. 61–76). In: Nelson JS, Schultze H-P, Wilson MVH (eds) Origin and phylogenetic interrelationships of teleosts. Munich: Verlag Dr. Friedrich Pfeil. 482 pp.

Broughton RE, Betancur-R R, Li C, Arratia G, Orti G. **2013**. Multi-locus phylogenetic analysis reveals the pattern and tempo of bony fish evolution. *PLoS Currents Tree of Life*. April 16. Edition 1. https://doi.org/10.1371/currents. tol.2ca8041495ffafd0c92756e75247483e

Broussonet PMA. **1782**. *Ichthyologia sistens piscium descriptions et icones*. London: Emsley.

Broussonet PMA. **1787**. Observations sur les écailles de plusieurs espèces de poissons qu'on croit communément dépourvus de ces parties. *Journal de Physique* 3(1): 12–19.

Brownell CL. **1979**. Stages in the early development of 40 marine fish species with pelagic eggs from the Cape of Good Hope. *Ichthyological Bulletin of the J.L.B. Smith Institute of Ichthyology* 40: 1–84.

Bruce RW. **1980**. On the occurrence of very small terminal phase parrotfishes. *Copeia* 1980(4): 887–889.

Bruce RW, Randall JE. **1985**. Revision of the Indo-Pacific parrotfish genera *Calotomus* and *Leptoscarus*. *Indo-Pacific Fishes* 5: 1–32, Pls. 1–3.

Bruner JC, Arnam S. 1979. Chromis woodsi, a new species of damselfish (Pomacentridae) from the western Indian Ocean with a redescription of Chrornis axillaris (Bennett) 1831. Fieldiana Zoology 73(3): 49–63.

Brünnich MT. **1788**. Om en ny fiskart, den draabeplettede pladefisk fanget ved Helsingör i Nordsöen 1786. *Kongelige Danske Videnskabernes Selskabs Skrivter, Nye Samling af det* 3: 398–407.

Brüss R. **1987**. Tiefenwasser- und Tiefseefische aus dem Roten Meer. XIII. *Uranoscopus marisrubri* n. sp. aus dem zentralen und nördlichen Roten Meer (Pisces: Perciformes: Uranoscopidae). *Senckenbergiana Biologica* 68(1/3): 39–48.

Brüss R. 1987. Two new species of Uranoscopus Linnaeus, 1758, from the Red Sea: Uranoscopus dollfusi n. sp. and U. bauchotae n. sp. Bulletin du Muséum National d'Histoire Naturelle (Paris) (Série 4, Section A: Zoologie Biologie et Ecologie Animales) 8(4): 955–967.

Bruton MN. **1997**. Threatened fishes of the world: *Silhouettea sibayi* Farquharson, 1970 (Gobiidae). *Environmental Biology of Fishes* 49(1): 44.

Bruton MN, Cooper KH. 1980. Studies on the ecology of Maputaland. Grahamstown: Rhodes University, and Durban: Natal Branch of the Wildlife Society of Southern Africa. 560 pp.

Bruun AF. **1933**. On the value of the number of vertebrae in the classification of the Exocoetidae. *Videnskabelige Meddelelser fra Dansk Naturhistorisk Forening* (Kjøbenhavn) 94: 375–384. Bruun AF. **1935**. Flying-fishes (Exocoetidae) of the Atlantic. Systematic and biological studies. *Dana Report* 6: 1–108.

Bryan PG. **1975**. Food habits, functional digestive morphology, and assimilation efficiency of the rabbitfish *Siganus spinus* (Pisces, Siganidae) on Guam. *Pacific Science* 29(3): 269–277.

Bucciarelli G, Golani D, Bernardi G. **2002**. Genetic cryptic species as biological invaders: the case of a Lessepsian fish migrant, the hardyhead silverside *Atherinomorus lacunosus*. *Journal of Experimental Marine Biology and Ecology* 273(2): 143–149.

Budker P. **1935**. Description d'un genre nouveau de la famille des Carcharinidés. *Bulletin du Muséum National d'Histoire Naturelle* (Paris) (Série 2) 7(2): 107–112.

Budker P. 1935. Sélaciens capturés dans la région de Dakar (mai-août 1934). Bulletin du Muséum National d'Histoire Naturelle (Paris) (Série 2) 7(2): 183–189.

Burgess WE. **1978**. *Butterfly fishes of the world: A monograph of the family Chaetodontidae*. Neptune City, New Jersey: TFH Publications. 832 pp.

Burgess WE. 1989. An atlas of freshwater and marine catfishes. A preliminary survey of the Siluriformes. Neptune City, New Jersey: TFH Publications. 784 pp., Pls. 1–285.

Burgess W[E], Axelrod HR. 1973. Pacific marine fishes. Book 3: Fishes of Sri Lanka, (Ceylon), the Maldives Islands and Mombasa. Neptune City, New Jersey: TFH Publications. pp. 561–839.

Burgess W[E], Axelrod HR. 1974. Pacific marine fishes. Book 4: Fishes of Taiwan and adjacent waters. Neptune City, New Jersey: TFH Publications. pp. 841–1110.

Burgess W[E], Axelrod HR. 1975. Pacific marine fishes. Book 6: Fishes of Melanesia. Neptune City, New Jersey: TFH Publications. pp. 1383-1654.

Burgess WE, Axelrod HR, Hunziker RE III. **1988**. *Dr. Burgess's atlas of marine aquarium fishes*. Neptune City, New Jersey: TFH Publications. 736 pp.

Burhanuddin AI, Iwatsuki Y, Yoshino T, Kimura S. 2002. Small and valid species of *Trichiurus brevis* Wang and You, 1992 and *T. russelli* Dutt and Thankam, 1966, defined as the "*T. russelli* complex" (Perciformes: Trichiuridae). *Ichthyological Research* 49(3): 211–223.

Burke CV. **1912**. A new genus and six new species of fishes of the family Cyclogasteridae. *Proceedings of the United States National Museum* **43**(1941): 567–574.

Burridge CP. 1999. Suggestion of synonymy for Nemadactylus and Acantholatris (Perciformes: Cirrhitoidea) (pp. 413–416).
In: Séret B, Sire J-Y (eds) Proceedings of the 5<sup>th</sup> Indo-Pacific Fish Conference, Noumea, New Caledonia, 3–8 November 1997. Paris: Société Française d'Ichtyologie. 866 pp.

Burridge CP, Smolenski AJ. **2004**. Molecular phylogeny of the Cheilodactylidae and Latridae (Perciformes: Cirrhitoidea) with notes on taxonomy and biogeography. *Molecular Phylogenetics and Evolution* 30: 118–127. Burt JA, Feary DA, Bauman AG, Usseglio P, Cavalcante GH, Sale PF. 2011. Biogeographic patterns of reef fish community structure in the northeastern Arabian Peninsula. *ICES Journal of Marine Science* 68(9): 1875–1883. http://dx.doi.org/10.1093/icesjms/fsr129

Burton E. **1835**. Description of a species of *Agriopus*, Cuv. *Proceedings of the Zoological Society of London* 3: 116–118.

Busakhin SV. **1981**.*Trachichthodes druzhinini* Busakhin, a new species of the family Berycidae (Osteichthyes) from the Indian Ocean. *Zoologicheskii Zhurnal* 60(11): 1728–1731.

Busakhin SV. **1982**. Taxonomy and distribution of the family Berycidae (Osteichthyes) in the world ocean. *Voprosy Ikhtiologii* 22(6): 904–921. [In Russian. English translation in *Journal of Ichthyology* 22(6): 1–21]

Busby MS, Orr JW. **1999**. A pelagic basslet *Howella sherborni* (Family Acropomatidae) off of the Aleutian Islands. *Alaska Fishery Research Bulletin* 6(1): 49–53.

Bussing WA. **1965**. Studies of the midwater fishes of the Peru-Chile Trench. In: Llano GA (ed) Biology of the Antarctic Seas II. *Antarctic Research Series* 5: 185–227.

Butler JL. **1979**. The nomeid genus *Cubiceps* (Pisces) with a description of a new species. *Bulletin of Marine Science* 29(2): 226–241.

Buxton CD, Smale MJ, Wallace JH, Cockcroft VG. 1984. Inshore small-mesh trawling survey of the Cape south coast. Part 4. Contributions to the biology of some Teleostei and Chondrichthyes. *South African Journal of Zoology* 19(3): 180–188.

## C

Cadenat J. **1937**. Recherches systématiques sur les poissons littoraux de la côte occidentale d'Afrique. Liste des poissons littoraux récoltés par le navire *Président Théodore-Tissier* au cours de sa cinquième croisière (1936). *Revue des Travaux de l'Institut des Pêches Maritimes* 10(fasc.4:40): 425–562.

Cadenat J. **1959**. Notes d'Ichtyologie ouest-Africaine. XX. *Galeus polli* espèce nouvelle ovovivipare de Scylliorhinidae. *Bulletin de l'Institut Français de l'Afrique Noire* (Série A) 21(1): 395–409.

Cadenat J. **1960**. Notes d'Ichtyologie ouest-Africaine. XXVII. *Raja doutrei*, espèce nouvelle des eaux profondes des côtes du Sénégal. *Bulletin de l'Institut Français de l'Afrique Noire* (Série A) 22: 294–330.

Cadenat J. **1963**. Notes d'Ichtyologie ouest-Africaine. XXXIX. Notes sur les requins de la famille des Carchariidae et formes apparentées de l'Atlantique ouest-Africaine (avec la description d'une espèce nouvelle: *Pseudocarcharias pelagicus*, classée dans un sous-genre nouveau). *Bulletin de l'Institut Français de l'Afrique Noire* (Série A) 25(2): 536–543. Cadenat J. **1964**. Notes d'Ichtyologie ouest-Africaine. XLI. Les Sphyraenidae de la côte occidentale d'Afrique. *Bulletin de l'Institut Français de l'Afrique Noire* (Série A) 26(2): 659–685.

Cakić P, Lenhardt M, Mićković D, Sekulić N, Budakov LJ.
2002. Biometric analysis of *Syngnathus abaster* populations. *Journal of Fish Biology* 60(6): 1562–1569.

Campana SE, Jones C, McFarlane GA, Myklevoll S. **2006**. Bomb dating and age validation using the spines of spiny dogfish (*Squalus acanthias*). *Evironmental Biology of Fishes* 77: 327–336.

Campbell MA, Chanet B, Chen J-N, Lee M-Y, Chen W-J. **2019**. Origins and relationships of the Pleuronectoidei: molecular and morphological analysis of living and fossil taxa. *Zoologica Scripta* 48(5): 640–656. https://doi.org/10.1111/ zsc.12372

Campbell MA, Chen W-J, López JA. **2013**. Are flatfishes (Pleuronectiformes) monophyletic? *Molecular Phylogenetics and Evolution* 69(3): 664–673. https://doi.org/10.1016/j. ympev.2013.07.011

Campbell MA, López JA, Satoh TP, Chen W-J, Miya M. **2014**. Mitochondrial genomic investigation of flatfish monophyly. *Gene* 551(2): 176–182. https://doi.org/10.1016/j. gene.2014.08.053

Campos PN, Bonecker ACT, de Castro MS, Anderson WD Jr. **2009**. First record of the fish genus *Symphysanodon* (Teleostei: Perciformes: Symphysanodontidae) from the western South Atlantic Ocean. *Zootaxa* 2270: 63–68. http://dx.doi.org/10.11646/zootaxa.2270.1.4

Cantor TE. **1849**. Catalogue of Malayan fishes. *Journal of the Asiatic Society of Bengal* 18(2): 983–1443, Pls. 1–14. [also referenced as 1850]

Cantwell GE. **1964**. A revision of the genus *Parapercis*, family Mugiloididae. *Pacific Science* 18(3): 239–280.

Capapé C. 1980. Nouvelle description de Heptranchias perlo (Bonnaterre, 1788) (Pisces: Pleurotremata, Hexanchidae).
Données sur la biologie de la reproduction et le régime alimentaire des specimens des côtes Tunisiennes. Bulletin de l'Office National des Pêches/Republic Tunisienne 4(2): 231–264.

Capapé C. **2008**. Diet of the angular rough shark *Oxynotus centrina* (Chondrichthyes: Oxynotidae) off the Languedocian coast (southern France, north-western Mediterranean). *Vie et Milieu* 58(1): 57–61.

Capapé C, Desoutter M. 1979. Nouvelle description de Aetomylaeus nichofii (Bloch et Schneider, 1801) (Pisces, Myliobatidae). Premières observations biologiques. Cahiers de l'Indo-Pacifique 1(3): 305–322.

Capapé C, Seck AA, Quignard J-P. **1999**. Observations on the reproductive biology of the angular rough shark, *Oxynotus centrina* (Oxynotidae). *Cybium* 23(3): 259–271.

Cappetta H. **2012**. *Chondrichthyes. Mesozoic and Cenozoic Elasmobranchii: teeth*. In: Schultze H-P (ed) *Handbook of Paleoichthyology*. Vol. 3E. Munich: Verlag Dr. Friedrich Pfeil. 512 pp.

Carbone F, Accordi G. **2000**. The Indian Ocean coast of Somalia. *Marine Pollution Bulletin* 41: 141–159.

Carbone F, Matteucci R, Pignatti JS, Russo A. **2003**. Facies analysis and biostratigrafia of the Auradu limestone formation in the Berbera-Sheikh area, northwestern Somalia. *Geologica Romana* 29: 213–235.

Carcasson RH. **1977**. *A field guide to the coral reef fishes of the Indian and west Pacific oceans*. London: Collins. 320 pp.

Cárdenas L, Hernández CE, Poulin E, Magoulas A, Kornfield I, Ojeda FP. **2005**. Origin, diversification, and historical biogeography of the genus *Trachurus* (Perciformes: Carangidae). *Molecular Phylogenetics and Evolution* 35(2): 496–507.

Carlson BA. **1981**. A new Indo-Pacific fish of the genus *Cirripectes* (Blenniidae, Salariini). *Pacific Science* 34(4): 407–414.

Carlson BA, Randall JE, Dawson MN. **2008**. A new species of *Epibulus* (Perciformes: Labridae) from the west Pacific. *Copeia* 2008(2): 476–483. http://dx.doi.org/10.1643/CI-07-085

Carmichael D. **1819**. Some accounts of the Island of Tristan da Cunha and of its natural productions. *Transactions of the Linnean Society of London* 12(2, art.29): 483–513.

Carpenter KE. **1987**. Revision of the Indo-Pacific fish family Caesionidae (Lutjanoidea), with descriptions of five new species. *Indo-Pacific Fishes* 15: 1–56, Pls. 1–7.

Carpenter KE. **1988**. FAO species catalogue. Vol. 8. Fusilier fishes of the world. An annotated and illustrated catalogue of caesionid species known to date. *FAO Fisheries Synopsis No. 125* (Vol. 8). Rome: FAO. iv + 75 pp., Pls. 1–5.

Carpenter KE. **2000**. Family Sparidae (porgies and seabreams) (p. 619). In: Randall JE, Lim KKP (eds) A checklist of the fishes of the South China Sea. *Raffles Bulletin of Zoology* Supplement 8: 569–667.

Carpenter KE. **2001**. Sparidae. Porgies (seabreams) (pp. 2990–3003). In: Carpenter KE, Niem VH (eds) *FAO species identification guide for fishery purposes. The living marine resources of the western central Pacific*. Vol. 5. Bony fishes part 3 (pp. 2791–3380). Rome: FAO.

Carpenter KE. **2002**. Sparidae. Porgies (pp. 1554–1577). In: Carpenter KE (ed) *FAO species identification guide for fishery purposes. The living marine resources of the western central Atlantic.* Vol. 3. Bony fishes part 2 (pp. 1375–2127). Rome: FAO.

Carpenter KE, Allen GR. 1989. FAO species catalogue. Vol. 9.
Emperor fishes and large-eye breams of the world (family Lethrinidae). An annotated and illustrated catalogue of lethrinid species known to date. *FAO Fisheries Synopsis No. 125* (Vol. 9). Rome: FAO. v + 118 pp., Pls. 1–8.

Carpenter KE, Collette BB, Russo JL. **1995**. Unstable and stable classifications of scombroid fishes. *Bulletin of Marine Science* 56(2): 379–405.

Carpenter KE, De Angelis N (eds). **2016**. *FAO species identification guide for fishery purposes. The living marine resources of the eastern central Atlantic.* Vol. 4. Bony fishes part 2 (Perciformes to Tetraodontiformes) and sea turtles. Rome: FAO. i–xiii + 2343–3124 pp.

Carpenter KE, Harrison PL, Hodgson G, Alsaffar AH, Alhazeem SH. 1997. The corals and coral reef fishes of Kuwait. Kuwait Institute for Scientific Research. 166 pp.

Carpenter KE, Johnson GD. **2002**. A phylogeny of sparoid fishes (Perciformes, Percoidei) based on morphology. *Ichthyological Research* 49(2): 114–127.

Carpenter KE, Krupp F, Jones DA, Zajonz U. **1997**. FAO species identification guide for fishery purposes. The living marine resources of Kuwait, eastern Saudi Arabia, Bahrain, Qatar, and the United Arab Emirates. Rome: FAO. 293 pp., Pls. 17.

Carpenter KE, Niem VH (eds). **1999**. *FAO species identification guide for fishery purposes. The living marine resources of the western central Pacific.* Vol. 3. Batoid fishes, chimaeras and bony fishes part 1 (Elopidae to Linophrynidae). FAO: Rome. iii–vi + 1397–2068 pp.; Pls. I–IV.

Carpenter K, Niem VH (eds). 1999. FAO species identification guide for fishery purposes. The living marine resources of the western central Pacific. Vol. 4. Bony fishes part 2 (Mugilidae to Carangidae). Rome: FAO. iii–v, 2069–2790 pp.; Pls. I–VII.

Carpenter KE, Niem VH (eds). **2001**. *FAO species identification guide for fishery purposes. The living marine resources of the western central Pacific.* Vol. 5. Bony fishes part 3 (Menidae to Pomacentridae). Rome: FAO. iii–iv, 2791–3380 pp.; Pls. I–XXVII.

Carpenter KE, Niem VH (eds). **2001**. *FAO species identification guide for fishery purposes. The living marine resources of the western central Pacific.* Vol. 6. Bony fishes part 4 (Labridae to Latimeriidae), estuarine crocodiles, sea turtles, sea snakes and marine mammals. Rome: FAO. iii–v, 3381–4218 pp.; Pls. I–XIX.

Carrier JC, Musick JA, Heithaus MR (eds). **2004**. *Biology of sharks and their relatives*. Boca Raton, Florida: CRC Press. 596 pp.

Carrier JC, Musick JA, Heithaus MR (eds). **2010**. *Sharks and their relatives II: Biodiversity, adaptive physiology, and conservation*. Boca Raton, Florida: CRC Press. 746 pp. https://doi.org/10.1201/9781420080483

Carrier JC, Musick JA, Heithaus MR (eds). **2012**. *Biology of sharks and their relatives* (2<sup>nd</sup> edition). Boca Raton, Florida: CRC Press. 672 pp. https://doi.org/10.1201/b11867

Caruso JH. **1977**. The systematics of the fish family Lophiidae. PhD dissertation, Tulane University, New Orleans.

Caruso JH. **1981**. The systematics and distribution of the lophiid anglerfishes. I. A revision of the genus *Lophiodes* 

with the description of two new species. *Copeia* 1981(3): 522–549.

Caruso JH. **1983**. The systematics and distribution of the lophiid anglerfishes. II. Revisions of the genera *Lophiomus* and *Lophius*. *Copeia* 1983(1): 11–30.

Caruso JH. **1989**. A review of the Indo-Pacific members of the deep-water chaunacid anglerfish genus *Bathychaunax*, with the description of a new species from the eastern Indian Ocean (Pisces: Lophiiformes). *Bulletin of Marine Science* 45(3): 574–579.

Caruso JH. **1989**. Systematics and distribution of Atlantic chaunacid anglerfishes (Pisces: Lophiiformes). *Copeia* 1989(1): 153–165.

Caruso JH. **1999**. Family Lophiidae. Anglerfishes (monkfishes) (pp. 2004–2012). In: Carpenter KE, Niem VH (eds) *FAO species identification guide for fishery purposes. The living marine resources of the western central Pacific*. Vol. 3. Batoid fishes, chimaeras and bony fishes part 1 (pp. 1397–2068). Rome: FAO.

Caruso JH, Bullis HR Jr. **1976**. A review of the lophiid angler fish genus *Sladenia* with a description of a new species from the Caribbean Sea. *Bulletin of Marine Science* 26(1): 59–64.

Caruso JH, Ho H-C, Pietsch TW. **2006**. *Chaunacops* Garman, 1899, a senior objective synonym of *Bathychaunax* Caruso, 1989 (Lophiiformes: Chaunacoidei: Chaunacidae). *Copeia* 2006(1): 120–121.

Carvalho MR de, *see* De Carvalho MR.

Castelnau FL. **1861**. *Mémoire sur les poissons de l'Afrique australe*. Paris: J-B Baillière et fils. 78 pp.

Castelnau FL. **1872**. Contribution to the ichthyology of Australia. *Proceedings of the Zoological and Acclimatisation Society of Victoria* (Melbourne) 1: 29–242.

Castelnau FL. **1873**. Contribution to the ichthyology of Australia. Nos. III thru IX. *Proceedings of the Zoological and Acclimatisation Society of Victoria* (Melbourne) 2: 37–158.

Castelnau FL. **1875**. Researches on the fishes of Australia. Philadelphia Centennial Expedition of 1876. *Intercolonial Exhibition Essays* 1875–6(art. 2): 1–52.

Castelnau FL. **1878**. Australian fishes. New or little known species. *Proceedings of the Linnean Society of New South Wales* 2(3): 225–248.

Castelnau FL. **1879**. Essay on the ichthyology of Port Jackson. *Proceedings of the Linnean Society of New South Wales* 3(4): 347–402.

Castle PHJ. **1959**. A large leptocephalid (Teleostei, Apodes) from off South Westland, New Zealand. *Transactions of the Royal Society of New Zealand* 87(1/2): 179–184.

Castle PHJ. **1964**. Congrid leptocephali in Australasian waters with descriptions of *Conger wilsoni* (Bl. & Schn.) and *C. verreauxi* Kaup. *Zoological Publications from Victoria University of Wellington* 37: 1–45. Castle PHJ. **1967**. Taxonomic notes on the eel, *Muraenesox cinereus* (Forsskål, 1775), in the western Indian Ocean. *Department of Ichthyology, Rhodes University, Special Publication* 2: 1–10.

Castle PHJ. **1968**. A contribution to a revision of the moringuid eels. *Department of Ichthyology, Rhodes University, Special Publication* 3: 1–29.

Castle PHJ. **1968**. The congrid eels of the western Indian Ocean and the Red Sea. *Ichthyological Bulletin, Department of Ichthyology, Rhodes University* 33: 685–726.

Castle PHJ. **1969**. The eel genera *Congrina* and *Coloconger* off southern Mozambique and their larval forms. *J.L.B. Smith Institute of Ichthyology, Rhodes University, Special Publication* 6: 1–10.

Castle PHJ. **1969**. An index and bibliography of eel larvae. *J.L.B. Smith Institute of Ichthyology, Rhodes University, Special Publication* 7: 1–121.

Castle PHJ. **1972**. The eel genus *Benthenchelys* (Fam. Ophichthidae) in the Indo-Pacific. *Dana Report* 82: 1–32.

Castle PHJ. **1982**. Tiefenwasser- und Tiefseefische aus dem Roten Meer. III. A new species of *Uroconger* from Red Sea benthos (Pisces: Teleostei: Congridae). *Senckenbergiana Biologica* 62(4/6): 205–209.

Castle PHJ. **1995**. Alcock's congrid eels from the *Investigator* collection in Indian seas 1888–1894. *Copeia* 1995(3): 706–718.

Castle PJH, McCosker JE. 1999. A new genus and two new species of myrophine worm-eels, with comments on *Muraenichthys* and *Scolecenchelys* (Anguilliformes: Ophichthidae). *Records of the Australian Museum* 51(2–3): 113–122. https://doi.org/10.3853/j.0067-1975.51.1999.1300

Castle PJH, Randall JE. **1999**. Revision of Indo-Pacific garden eels (Congridae: Heterocongrinae), with descriptions of five new species. *Indo-Pacific Fishes* 30: 1–52.

Castle PJH, Williamson GR. **1975**. Systematics and distribution of eels of the *Muraenesox* group (Anguilliformes, Muraenesocidae). A preliminary report and key. *J.L.B. Smith Institute of Ichthyology Special Publication* 15: 1–9.

Castro JI. **2011**. *The sharks of North America*. Oxford: Oxford University Press. xiii + 613 pp.

Castro-Aguirre JL, Arvizu-Martinez J, Alarcón-Gonzalez C. **1991**. Una especie nueva de *Regalecus* (Pisces: Lampridiformes) hallada en la Bahía de la Paz, Baja California Sur, Mexico, con notas y observaciones taxonómicas y biogeográficas de la familia Regalecidae. *Anales de la Escuela Nacional de Ciencias Biológicas, Mexico* 34: 159–171.

Catesby M. 1771. *The natural history of Carolina, Florida and the Bahama Islands*; containing the figures of birds, beasts, fishes, serpents ... with their descriptions in English and French. Third edition, 2 vols. London.

Cavin L. **2001**. Osteology and phylogenetic relationships of the teleost *Goulmimichthys arambourgi* Cavin, 1995, from the Upper Cretaceous of Goulmima, Morocco. *Eclogae Geologicae Helvetiae* 94: 509–535.

Cavin L, Forey PL, Giersch S. **2013**. Osteology of *Eubiodectes libanicus* (Pictet & Humbert, 1866) and some other ichthyodectiformes (Teleostei): phylogenetic implications. *Journal of Systematic Palaeontology* 11(2): 115–177. https://doi.org/10.1080/14772019.2012.691559

Cavin L, Forey PL, Lécuyer C. **2007**. Correlation between environment and Late Mesozoic ray-finned fish evolution. *Palaeogeography, Palaeoclimatology, Palaeoecology* 245(3–4): 335–367.

Cavin L, Grigorescu D. **2005**. A new *Crossognathus* (Actinopterygii, Teleostei) from the Lower Cretaceous of Romania with comments on Crossognathidae relationships. *Geodiversitas* 27(1): 5–16.

CFSA (Coastal Fishes of Southern Africa), *see* Heemstra PC, Heemstra E. 2004.

Chabanaud P. 1923. Description de deux Plagiostomiens nouveaux d'Indo-Chine, appartenant au genre Dasybatus (Trygon). Bulletin du Muséum National d'Histoire Naturelle, Paris (Série 1) 29(1): 45–50.

Chabanaud P. **1925**. Lepadogaster (Mirbelia) bimaculatus Penn., microcephalus Brook, et Pellegrini, nov. sp. (Pisces: Gobiesocidae). Bulletin du Muséum National d'Histoire Naturelle, Paris (Série 1) 31(4): 283–287.

Chabanaud P. **1927**. Les soles de l'Atlantic oriental nord et des mers adjacents. *Bulletin de l'Institut Océanographique* (Monaco) 488: 1–67.

Chabanaud P. **1927**. Sur diverses espèces du genre *Taenioides* Lacep. [Poissons Gobiformes]. *Bulletin de la Société Zoologique de France* 52: 404–415.

Chabanaud P. **1929**. Description d'un nouvel élasmobranche batoïde de Madagascar. *Bulletin du Muséum National d'Histoire Naturelle, Paris* (Série 2) 1(6): 365–369.

Chabanaud P. **1929**. Observations sur la taxonomie, la morphologie et la bionomie des Soléidés du genre *Pegusa*. *Annales de l'Institut Océanographique* (Monaco) 7: 215–261.

Chabanaud P. **1929**. Remarques sur divers poissons de la famille des Syngnathidae et description de deux espèces nouvelles de l'Inde Archipelagique. *Bulletin de la Société Zoologique de France* 54: 165–172.

Chabanaud P. **1930**. Description d'un nouveau *Cubiceps* (Pisces, Stromateidae) de la mer Rouge. *Bulletin du Muséum National d'Histoire Naturelle, Paris* (Série 2) 2(5): 519–523.

Chabanaud P. **1931**. Sur divers poissons soléiformes de la région Indo-Pacifique. *Bulletin de la Société Zoologique de France* 56: 291–305.

Chabanaud P. **1932**. Poissons recueillis dans le Grand lac Amer (isthme de Suez) par M. le Professeur A. Gruvel, en 1932.

*Bulletin du Muséum National d'Histoire Naturelle, Paris* (Série 2) 4: 822–835.

Chabanaud P. **1933**. Sur divers poissons de la Mer Rouge et du canal de Suez. Description de deux espèces nouvelles. *Bulletin de l'Institut Océanographique* (Monaco) 627: 1–12.

Chabanaud P. **1934**. Les soléides du group *Zebrias*. Définition d'un sous-genre nouveau et description d'une sous-espèce nouvelle. *Bulletin de la Société Zoologique de France* 59: 420–436.

Chabanaud P. **1934**. Poissons recueillis dans le lac Timsah (isthme de Suez) par M. le Professeur A. Gruvel, en 1933. *Bulletin du Muséum National d'Histoire Naturelle, Paris* (Série 2) 6(1): 156–160.

Chabanaud P. **1937**. II. Poissons. In: Gruvel A, Chabanaud P (eds) Missions A. Gruvel dans le canal de Suez. *Mémoires de l'Institut d'Égypte* N.S. 35: 1–31.

Chabanaud P. **1938**. Contribution a la morphologie et a la systématique des téléostéens dyssymétriques. *Archives du Museum National d'Histoire Naturelle* (Série 6) 15: 59–139, Pls. 1–9.

Chabanaud P. **1943**. Notules ichthyologiques. XVII. Additions à la faune de la mer Rouge. *Bulletin du Muséum National d'Histoire Naturelle, Paris* (Série 2) 14(6) [for 1942]: 396–402.

Chabanaud P. **1947**. Notules Ichthyologiques. XXX. Additions à la faune de la Mer Rouge. *Bulletin du Muséum National d'Histoire Naturelle, Paris* (Série 2) 19(2): 156–157.

Chabanaud P. **1948**. Description d'un nouveau *Cynoglossus* de l'Inde. *Annals and Magazine of Natural History* (Series 11) 14(119) [for 1947]: 813–815.

Chabanaud P. **1949**. Description d'un nouveau *Cynoglossus* de la Mer Rouge. *Bulletin de la Société Zoologique de France* 74: 146–148.

Chabanaud P. **1951**. Sur divers *Cynoglossus* de la région Indo-Pacific. *Annals and Magazine of Natural History* (Series 12) 4(39, art.26): 268–273.

Chabanaud P. **1954**. Description d'un nouvel *Aseraggodes* de la côte orientale de l'Afrique. In: Morrow JE (ed) Fishes from East Africa, with new records and descriptions of two new species. *Annals and Magazine of Natural History* (Series 12) 7(83): 797–820.

Chabanaud P. **1954**. Notules Ichthyologiques (suite). Notule 46. Description d'un nouveau *Symphurus* de la côte sud de l'Arabie (pp. 464–465). *Bulletin du Muséum National d'Histoire Naturelle, Paris* (Série 2) 26(4): 464–467.

Chabanaud P. **1954**. Notules Ichthyologiques (suite). Notule 47. Présence inédite d'un *Cynoglossus* dans la Méditerranée orientale (pp. 465–466). *Bulletin du Muséum National d'Histoire Naturelle, Paris* (Série 2) 26(4): 464–467.

Chabanaud P. **1955**. Flatfishes of the genus *Symphurus* from the U.S.S. *Albatross* Expedition to the Philippines, 1907–1910. *Journal of the Washington Academy of Sciences* 45(1): 30–32.

Chabanaud P. **1955**. Revision des *Symphurus* du *Siboga*. *Beaufortia* 5(46): 43–45.

Chabanaud P. **1955**. Sur cinq espèces du genre *Symphurus*, dont trois sont inédites. *Bulletin du Muséum National d'Histoire Naturelle, Paris* (Série 2) 27(5): 368–370.

Chabanaud P. **1956**. Les *Symphurus* marbrés du complexe Indo-Pacifique tropical. *Archives du Museum National d'Histoire Naturelle, Paris* (Série 7) 4: 79–100.

Chabanaud P. 1968. Description d'un nouveau Laeops (Pleuronectiformes Bothidae) de la Mer Rouge (Mission R. Ph. Dollfus). Bulletin du Muséum National d'Histoire Naturelle, Paris (Série 2) 39(5) [for 1967]: 838–845, Pls. 1–2.

Chabanet P. **1994**. Étude des relations entre les peuplements benthiques et les peuplements ichtyologiques sur le complexe récifal de Saint-Gilles – La Saline (Île de la Réunion). Thèse de Doctorat, Université d'Aix-Marseille. 263 pp.

Chabanet P. **2002**. Coral reef fish communities of Mayotte (western Indian Ocean) two years after the impact of the 1998 bleaching event. *Marine and Freshwater Research* 53: 107–113.

Chabanet P, Dufour V, Galzin R. **1995**. Disturbance impact on reef fish communities in Reunion Island (Indian Ocean). *Journal of Experimental Marine Biology and Ecology* 188(1): 29–48. https://doi.org/10.1016/0022-0981(94)00184-F.

Chabanet P, Letourneur Y. **1995**. Spatial pattern of size distribution of four fish species on Reunion coral reef flats. *Hydrobiologia* 300–301(1): 299–308. https://doi.org/10.1007/BF00024470

Chabanet P, Ralambondrainy H, Amanieu M, Faure G, Galzin R. **1997**. Relationships between coral reef substrata and fish. *Coral Reefs* 16: 93–102.

Chakrabarty P, Amarasinghe T, Sparks JS. **2009**. Redescription of ponyfishes (Teleostel: Leiognathidae) of Sri Lanka and the status of *Aurigequula* Fowler 1918. *Ceylon Journal of Science* (Biological Sciences) 37(2): 143–161.

Chakrabarty P, Davis MP, Smith WL, Baldwin ZH, Sparks JS. **2011**. Is sexual selection driving diversification of the bioluminescent ponyfishes (Teleostei: Leiognathidae)? *Molecular Ecology* 20: 2818–2834.

https://doi.org/10.1111/j.1365-294X.2011.05112.x Chakrabarty P, Davis MP, Smith WL, Berquist R, Gledhill KM,

Frank LR, Sparks JS. 2011. Evolution of the light organ system in ponyfishes (Teleostei: Leiognathidae). *Journal* of Morphology 272: 704–721. https://doi.org/10.1002/ jmor.10941

Chakrabarty P, Sparks JS. **2007**. Phylogeny and taxonomic revision of *Nuchequula* Whitley 1932 (Teleostei: Leiognathidae), with the description of a new species. *American Museum Novitates* 3588: 1–25. http://dx.doi. org/10.1206/0003-0082(2007)3588[1:PATRON]2.0.CO;2

Chakrabarty P, Sparks JS. **2008**. Diagnoses for *Leiognathus* Lacepède 1802, *Equula* Cuvier 1815, *Equulites* Fowler 1904, *Eubleekeria* Fowler 1904, and a new ponyfish genus (Teleostei: Leiognathidae). *American Museum Novitates* 3623: 1–11. http://dx.doi.org/10.1206/618.1

Chakrabarty P, Sparks JS, Ho H-C. **2010**. Taxonomic review of the ponyfishes (Perciformes: Leiognathidae) of Taiwan. *Marine Biodiversity* 40(2): 107–121. https://doi.org/10.1007/ s12526-010-0037-0

Chakraborty A, Aranishi F, Iwatsuki Y. **2006**. Genetic differences among three species of the genus *Trichiurus* (Perciformes: Trichiuridae) based on mitochondrial DNA analysis. *Ichthyological Research* 53(1): 93–96.

Chakraborty A, Iwatsuki Y. **2006**. Genetic variation at the mitochondrial 16S rRNA gene among *Trichiurus lepturus* (Teleostei: Trichiuridae) from various localities: preliminary evidence of a new species from West coast of Africa. *Hydrobiologia* 563: 501–513.

Chakraborty A, van Oijen MJP, Lim KKP, Iwatsuki Y. **2006**. *Lepturacanthus roelandti* (Bleeker, 1860), a valid species of hairtail (Perciformes: Trichiuridae). *Ichthyological Research* 53(1): 41–46.

Chakraborty A, Yoshino T, Iwatsuki Y. **2006**. A new species of scabbardfish, *Evoxymetopon macrophthalmus* (Scombroidei: Trichiuridae), from Okinawa, Japan. *Ichthyological Research* 53(2): 137–142.

Chan WL. **1967**. A new species of congrid eel from the South China Sea. *Journal of Natural History* 1(1): 97–112.

Chan WL. **1970**. A new genus and two new species of commercial snappers from Hong Kong. *Hong Kong Fisheries Bulletin* (Fisheries Branch of the Agriculture and Fisheries Department, Hong Kong) 1: 19–38.

Chan WL, Chilvers RM. **1974**. A revision of the Indo-Pacific spariform percoids of the Monotaxinae, with the description of a new genus *Wattsia*. *Hong Kong Fisheries Bulletin* (Fisheries Branch of the Agriculture and Fisheries Department, Hong Kong) 4: 85–95.

Chandy M. **1954**. A key for the identification of the catfishes of the genus *Tachysurus* La Cepède, with a catalogue of the specimens in the collection of the Indian Museum. *Records of the Indian Museum* (Calcutta) 51: 1–18.

Chandy M. **1954**. Notes on the Indian flying fishes of the genus *Cypselurus* Swainson. *Records of the Indian Museum* (Calcutta) 52: 177–184.

Chandy M. **1957**. *Memoirs on Indian animal types*. I. *Dasyatis* (the sting ray). Lucknow, India: Maxwell Company. ix + 123 pp.

Chanet B. **2003**. Interrelationships of scophthalmid fishes (Pleuronectiformes: Scophthalmidae). *Cybium* 27(4): 275–286.

Chanet B, Chapleau F, Desoutter M. **2004**. Os et ligaments intermusculaires chez les poissons plats [Teleostei: Pleuronectiformes]: interpretations phylogenetiques. *Cybium* 28(1): 9–14.

Chang M-M, Jin F. 1996. Mesozoic fish faunas of China (pp. 461–478). In: Arratia G, Viohl G (eds) Mesozoic Fishes
Systematics and paleoecology. Proceedings of the 1st International Meeting on Mesozoic Fishes, Eichstätt, 1993. Munich: Verlag Dr. Friedrich Pfeil. 576 pp.

Chang M-M, Miao D. 2004. An overview of Mesozoic fishes in Asia (pp. 535–563). In: Arratia G, Tintori A (eds) *Mesozoic Fishes 3 – Systematics, paleoenvironments and biodiversity. Proceedings of the international meeting, Serpianao, 2001.* Munich: Verlag Dr. Friedrich Pfeil. 649 pp.

Chao LN. **1986**. Sciaenidae (pp. 865–874). In: Whitehead PJP, Bauchot M-L, Hureau J-C, Nielsen JG, Tortonese E (eds) *Fishes of the north-eastern Atlantic and the Mediterranean*. Vol. II. Paris: UNESCO.

Chapleau F. **1988**. Comparative osteology and intergeneric relationships of the tongue soles (Pisces; Pleuronectiformes; Cynoglossidae). *Canadian Journal of Zoology* 66(5): 1214–1232.

Chapleau F. **1993**. Pleuronectiform relationships: a cladistic reassessment. *Bulletin of Marine Science* 52(1): 516–540.

Chapleau F, Amaoka K. **1998**. Flatfishes (pp. 223–226). In: Paxton JR, Eschmeyer WN (eds) *Encyclopedia of fishes*. *A comprehensive Illustrated guide by international experts* (2<sup>nd</sup> edition). San Diego: Academic Press. 240 pp.

Chapleau F, Keast A. **1988**. A phylogenetic reassessment of the monophyletic status of the family Soleidae, with comments on the suborder Soleoidei (Pisces: Pleuronectiformes). *Canadian Journal of Zoology* 66(12): 2797–2810.

Chapleau F, Renaud CB. **1993**. *Paraplagusia sinerama* (Pleuronectiformes: Cynoglossidae), a new Indo-Pacific tongue sole with a revised key to species of the genus. *Copeia* 1993(3): 798–807.

Chapleau F, Renaud CB, Kailola PJ. **1991**. *Paraplagusia longirostris*, a new flatfish (Cynoglossidae) from Australia and Papua New Guinea. *Japanese Journal of Ichthyology* 38(3): 239–244.

Chapman WM. **1948**. The osteology and relationships of the round herring *Etrumeus micropus* Temminck and Schlegel. *Proceedings of the California Academy of Sciences* (Series 4) 26(2): 25–41.

Chapman WM, Schultz LP. **1952**. Review of the fishes of the blennioid genus *Ecsenius*, with descriptions of five new species. *Proceedings of the United States National Museum* 102(3310): 507–528.

Chaudhuri BL. **1908**. Description of a new species of saw-fish captured off the Burma coast by the government of Bengal's steam trawler *Golden Crown*. *Records of the Indian Museum* (Calcutta) 2(4): 391–392.

Chaudhuri BL. **1909**. Description of two new species of *Caranx* from the Bay of Bengal. *Records of the Indian Museum* (Calcutta) 3(2): 141–143.

Chaudhuri BL. **1916**. Fauna of the Chilka Lake: Fish. Part 1. *Memoirs of the Indian Museum* 5(4): 403–439.

Chaudhuri BL. **1916**. Fauna of the Chilka Lake: Fish. Part 2. *Memoirs of the Indian Museum* 5(5): 441–458.

Chaudhuri BL. **1917**. Fauna of the Chilka Lake: Fish. Part 3. *Memoirs of the Indian Museum* 5(6): 491–508.

Chaudhuri BL. **1923**. Fauna of the Chilka Lake: Fish. Part 4. *Memoirs of the Indian Museum* 5(11): 711–769.

Chave EH, Mundy BC. **1994**. Deep-sea benthic fish of the Hawaiian Archipelago, Cross Seamount, and Johnston Atoll. *Pacific Science* 48: 367–409.

Chávez Ramos H, Galván Magaña F, Torres Villegas JR.
1985. Primer registro de *Regalecus ruselli* (Shaw) (Pisces: Regalecidae) de aguas mexicanas. *Investigaciones Marinas* CICIMAR 2(2): 105–112.

Chen C-H. **1987**. Studies of the early life history of flying fishes (family Exocoetidae) in the northwestern Pacific. *Taiwan Museum Special Publication* Series 7: 1–203.

Chen H-M, Shao K-T. **1995**. New eel genus, *Cirrimaxilla*, and description of the type species, *Cirrimaxilla formosa* (Pisces: Muraenidae) from southern Taiwan. *Bulletin of Marine Science* 57(2): 328–332.

Chen J-P, Jan R-Q, Shao K-T. **1997**. Checklist of reef fishes from Taiping Island (Itu Aba Island), Spratly Islands, South China Sea. *Pacific Science* 51(2): 143–166.

Chen J-P, Shao K-T. **1993**. New species of cardinalfish, *Archamia goni* (Pisces: Apogonidae), from Taiwan. *Copeia* 1993(3): 781–784.

Chen JTF. **1963**. A review of the sharks of Taiwan. *Biological Bulletin Tunghai University Ichthyology* (Series 1) 19: 1–102.

Chen JTF, Weng HTC. **1965**. A review of the flatfishes of Taiwan. *Biological Bulletin Tunghai University Ichthyology* (Series 5) 25–27: 1–103.

Chen JTF, Weng HTC. **1967**. A review of the apodal fishes of Taiwan. *Biological Bulletin Tunghai University Ichthyology* (Series 6) 32: 135–220.

Chen L-C. **1981**. Scorpaenid fishes of Taiwan. *Quarterly Journal of the Taiwan Museum* 34(1/2): 1–60.

Chen L-C, Liu W-Y. **1984**. *Pteroidichthys amboinensis*, a scorpaenid fish new to Taiwan, with a description of the species and a discussion of its validity. *Journal of Taiwan Museum* 37(2): 101–103.

Chen Q-C, Cai Y-Z, Ma X-M. **1997**. *Fishes from Nansha Islands to South China coastal waters 1*. Peking: Science Press. 202 pp.

Chen W-J, Bonillo C, Lecointre G. **2003**. Repeatability of clades as a criterion of reliability: a case study for molecular phylogeny of Acanthomorpha (Teleostei) with larger number of taxa. *Molecular Phylogenetics and Evolution* 26(2): 262–288. https://doi.org/10.1016/S1055-7903(02)00371-8

Chen W-J, Borsa P. **2020**. Diversity, phylogeny, and historical biogeography of large-eye seabreams (Teleostei: Lethrinidae). *Molecular Phylogenetics and Evolution* 151(art. 106902). https://doi.org/10.1016/j.ympev.2020.106902

Chen W-J, Santini F, Carnevale G, Chen J-N, Liu S-H, Lavoué S, Mayden RL. **2014**. New insights on early evolution of spiny-rayed fishes (Teleostei: Acanthomorpha). *Frontiers in Marine Science* 1(art. 53): 1–17. https://doi.org/10.3389/fmars.2014.00053

Chernova NV. **2008**. Systematics and phylogeny of fish of the genus *Liparis* (Liparidae, Scorpaeniformes). *Journal of Ichthyology* 48(10): 831–852. http://dx.doi.org/10.1134/ S0032945208100020

Cheung C, DeVantier L (eds). **2006**. *Socotra: A natural history of the islands and their people*. Hong Kong: Odyssey Books & Guides. 386 pp.

Chevey P. **1927**. Sur un genre nouveau de Scorpaenidae du Tonkin. *Bulletin du Muséum National d'Histoire Naturelle* (Paris) (Série 1) 33(3): 222–223.

Chiang W-C, Sun C-L, Yeh S-Z, Su W-C, Liu D-C, Chen W-Y.
2006. Sex ratios, size at sexual maturity, and spawning seasonality of sailfish *Istiophorus platypterus* from eastern Taiwan. *Bulletin of Marine Science* 79(3): 727–737.

Chiba SN, Iwatsuki Y, Yoshino T, Hanzawa N. **2009**. Comprehensive phylogeny of the family Sparidae (Perciformes: Teleostei) inferred from mitochondrial gene analyses. *Genes & Genetic Systems* (Shizuoka) 84(2): 153–170. http://dx.doi.org/10.1266/ggs.84.153

Chitravadivelu K, Sivapalan A. **1984**. Food and feeding of *Siganus lineatus* from waters around northern Sri Lanka. *Journal of the National Science Council of Sri Lanka* 12(1): 129–139.

Chlupaty P. **1986**. The batfishes. *Tropical Fish Hobbyist* 34(8#362): 24–27.

Choat JH, Davies CR, Ackerman JL, Mapstone BD. **2006**. Age structure and growth in a large teleost, *Cheilinus undulatus*, with a review of size distribution in labrid fishes. *Marine Ecology Progress Series* 318: 237–246.

Choat JH, Klanten OS, Van Herwerden L, Robertson DR, Clements KD. **2012**. Patterns and processes in the evolutionary history of parrotfishes (Family Labridae). *Biological Journal of the Linnean Society* 107(3): 529–557. http://dx.doi.org/10.1111/j.1095-8312.2012.01959.x

Choo CK, Liew HC. **2003**. Spatial distribution, substrate assemblages and size composition of sea horses (Family Syngnathidae) in the coastal waters of Peninsular Malaysia. *Journal of the Marine Biological Association of the UK* 83(2): 271–276.

Christensen MS. **1977**. A redescription of and new locality record for *Charibarbitus celetus* Smith 1963 (Pisces, Callionymidae), with two further range extensions of the South African ichthyofauna. *South African Journal of Science* 73: 145–146.

Christensen MS. **1978**. *Pavoclinus myae*, a new species of clinid fish (Perciformes Blennoidei) from South Africa, with a note on the identity of *P. graminis* and *P. laurentii*, and a key to the known species of *Pavoclinus*. *J.L.B. Smith Institute of Ichthyology Special Publication* 18: 1–16.

Chu Y-T, Lo Y-L, Wu H-L. **1963**. *A study of the classification of the sciaenoid fishes of China, with descriptions of new genera and species*. Shanghai Fisheries Institute. [In Chinese, English resumé pp. 83–94]

Chu Y-T, Meng C-W, Liu J-X. **1981**. Description of a new genus and a new species of Squalidae of China. *Acta Zootaxonomica Sinica* 6(1): 100–103. [In Chinese, English summary]

Chu Y-T, Meng Q-W. 1982. In: Chu et al. [see next entry]

Chu Y-T, Meng Q-W, Hu A-S, Li S. **1982**. Five new species of elasmobranchiate fishes from the deep waters of South China Sea. *Oceanologia et Limnologia Sinica* 13(4): 301–311.

Chu Y-T, Wu H-L. **1962**. Description of a new genus and a new species of a trichiurid fish off China. *Acta Zoologica Sinica* 14(2): 219–223.

Chu Y-T, Wu H-L, Jin X-B. **1981**. Four new species of the families Ophichthyidae and Neenchelidae. *Journal of Fisheries of China* 5(1): 21–27.

Clark E. **1966**. Pipefishes of the genus *Siokunichthys* Herald in the Red Sea with description of a new species. Israel South Red Sea Expedition, 1962, Report No. 18. *Bulletin of the Sea Fisheries Research Station, Haifa* 41: 3–6.

Clark E. **1968**. Eleotrid gobies collected during the Israel South Red Sea Expedition (1962), with a key to Red Sea species. *Bulletin of the Sea Fisheries Research Station, Haifa* 49: 3–7.

Clark E. **1980**. Red Sea fishes of the family Tripterygiidae with descriptions of eight new species. *Israel Journal of Zoology* 28(2–3) [for 1979]: 65–113.

Clark E, Ben-Tuvia A. **1973**. Red Sea fishes of the family Branchiostegidae with a description of a new genus and species *Asymmetrurus oreni*. Contributions to the knowledge of the Red Sea No. 51. *Bulletin of the Sea Fisheries Research Station, Haifa* 60: 63–74.

Clark E, Ben-Tuvia A, Steinitz H. **1968**. Observations on a coastal fish community, Dahlak Archipelago, Red Sea. *Bulletin of the Sea Fisheries Research Station, Haifa* 49: 15–31.

Clark E, Chao S. **1973**. A toxic secretion from the Red Sea flatfish *Pardachirus marmoratus* (Lacépède). *Bulletin of the Sea Fisheries Research Station, Haifa* 60: 53–56.

Clark E, George A. **1979**. Toxic soles, *Pardachirus marmoratus* from the Red Sea and *P. pavoninus* from Japan, with notes on other species. *Environmental Biology of Fishes* 4(2): 103–123.

Clark E, Gohar HAF. **1953**. The fishes of the Red Sea: Order Plectognathi. *Publications of the Marine Biological Station Al-Ghardaqa* (Red Sea) 8: 1–80.

Clark E, Petzold R. **1998**. Spawning behavior of the collared knifefish, *Cymolutes torquatus* (Labridae) in Papua New Guinea. *Environmental Biology of Fishes* 53: 459–464.

Clark E, Pohle JF. **1992**. Monogamy in the tilefish, *Malacanthus latovitiatus*, compared with polygamy in related species. *National Geographic Research and Exploration* 8: 276–295.

Clark E, Pohle JF, Halstead B. **1998**. Ecology and behavior of tilefishes, *Hoplolatilus starcki*, *H. fronticinctus* and related species (Malacanthidae): non-mound and mound builders. *Environmental Biology of Fishes* 52(4): 395–417. https://doi.org/10.1023/A:1007440719123

Clark E, Pohle M, Rabin J. **1991**. Spotted sandperch dynamics. *National Geographic Research and Exploration* 7(2): 138–155.

Clark E, Von Schmidt K. **1966**. A new species of *Trichonotus* (Pisces: Trichonotidae) from the Red Sea. *Bulletin of the Sea Fisheries Research Station, Haifa* 42: 29–36.

Clark HW. **1937**. New fishes from the Templeton Crocker Expedition of 1934–35. *Copeia* 1937(2): 88–91.

Clarke FE. **1879**. On some new fishes. *Transactions and Proceedings of the New Zealand Institute* 11(25) [for 1878]: 291–295, Pls. 14–15.

Clarke TA. **1984**. Diet and morphological variation in snipefishes, presently recognized as *Macrorhamphosus scolopax*, from southeast Australia: evidence for two sexually dimorphic species. *Copeia* 1984(3): 595–608.

Clayton DA, Vaughan TC. **1982**. Pentagonal territories of the mudskipper *Boleophthalmus boddarti* (Pisces: Gobidae). *Copeia* 1982(1): 232–234.

Cliff G. **1995**. Sharks caught in the protective gill nets off Natal, South Africa. 8. The great hammerhead *Sphyrna mokarran* (Rüppell). *South African Journal of Marine Science* 15(1): 105–114.

Cliff G, Compagno LJV, Smale MJ, van der Elst RP, Wintner SP. 2000. First records of white sharks *Carcharodon carcharias*, from Mauritius, Zanzibar, Madagascar and Kenya. *South African Journal of Science* 96(7): 365–366.

Cliff G, Dudley SFJ, Davis B. **1990**. Sharks caught in the protective gill nets off Natal, South Africa. 3. The shortfin mako, *Isurus oxyrinchus* (Rafinesque). *South African Journal of Marine Science* 9: 115–126.

Cliff G, Wilson RB. **1994**. *Natal Sharks Board's field guide to sharks and other marine animals* (revised edition). Umhlanga: Natal Sharks Board. 57 pp. Cloquet H. **1816**. Alutere. *Dictionaire des sciences naturelles* (Vol. 1, Supplement, pp. 135–136). Paris: F.G. Levrault.

Cloquet H. **1819**. Eptatrème ou Eptatrète. *Dictionaire des sciences naturelles* (Vol. 15: pp. 134–136). Paris: F.G. Levrault.

CMFRI. **2006**. *Marine fisheries census 2005*. Part 1. Kochi: Central Marine Fisheries Research Institute.

CMFRI. **2009**. *Annual report 2008–2009*. Kochi: Central Marine Fisheries Research Institute.

CMFRI. **2012**. *Marine fisheries census 2010*. Part 1. Kochi: Central Marine Fisheries Research Institute.

Coad BW. **1991**. Fishes of the Tigris-Euphrates Basin: a critical checklist. *Syllogeus* 68: 1–49.

Coad BW. **1998**. Systematic biodiversity in the freshwater fishes of Iran. *Italian Journal of Zoology* 65(Suppl.): 101–108.

Coad BW. **2010**. *Freshwater fishes of Iraq*. Sofia and Moscow: Pensoft Publishers. 294 pp.

Coates MI, Finarelli JA, Sansom IJ, Andreev PS, Criswell KE, Tietjen K, Rivers ML, La Riviere PJ. **2018**. An early chondrichthyan and the evolutionary assembly of a shark body plan. *Proceedings of the Royal Society* B 285: 2017.2418. http://dx.doi.org/10.1098/rspb.2017.2418

Coates MI, Gess RW. **2007**. A new reconstruction of *Onychoselache traquairi*, comments on early chondrichthyan pectoral girdles and hybodontiform phylogeny. *Palaeontology* 50(6): 1421–1446. http://dx.doi.org/10.1111/j.1475-4983.2007.00719.x

Coates MI, Gess RW, Finarelli JA, Criswell KE, Tietjen K. 2017. A symmoriiform chondrichthyan braincase and the origin of chimaeroid fishes. *Nature* 541: 209–211. http://dx.doi.org/10.1038/nature20806

Coates MI, Sequeira SEK. **2001**. A new stethacanthid chondrichthyan from the lower Carboniferous of Bearsden, Scotland. *Journal of Vertebrate Paleontology* 21(3): 438–459. http://dx.doi.org/10.1671/0272-4634(2001)021%5B0438:AN SCFT%5D2.0.CO;2

Cocco A. **1829**. Su di alcuni nuovi pesci de' mari di Messina. *Giornale di Scienze Lettere e Arti per La Sicilia Anno 7* 26(77): 138–147.

Cocco A. **1833**. Su di alcuni pesci de' mari di Messina. *Giornale di Scienze Lettere e Arti per La Sicilia* 42: 9–21.

Cocco A. **1839**. Ittiologia. [Sopra un nuovo genere di pesci della famiglia de' Centrolofini e di una nuova specie di *Trachurus*.] L' Innominato. *Giornale di Amena Letteratura e Belle Arti Anno* 3(7): 56–59.

Cocco A. **1840**. Su di alcuni nuovi pesci del mare di Messina. Il Maurolico. *Giornale di Scienze Lettere e Arti* 4(6): 236–244.

Cockcroft VG. 1991. Incidence of shark bites on Indian Ocean hump-backed dolphins (*Sousa plumbea*) off Natal, South Africa (pp. 277–282). In: Leatherwood S, Donovan GP (eds) *Cetaceans and cetacean research in the Indian Ocean Sanctuary*. UNEP Marine Mammal Technical Report 3. vii + 287 pp. Coetzee DJ. **1982**. Stomach content analyses of *Gilchristella aestuarius* and *Hepsetia breviceps* from the Swartvlei system and Groenvlei, southern Cape. *South African Journal of Zoology* 17(2): 59–66.

Coetzee DJ, Pool RC. **1985**. Stomach content analysis of the sea barbel, *Galeichthys feliceps* (Valenciennes in C & V), from the Swartvlei system, southern Cape. *South African Journal of Zoology* 20(1): 33–37.

Cohen DM. **1961**. On the identity of the species of the fish genus *Argentina* in the Indian Ocean. *Galathea Report* 5: 19–21.

Cohen DM. **1964**. A review of the ophidioid fish genus *Oligopus* with the description of a new species from West Africa. *Proceedings of the United States National Museum* 116(3494): 1–22.

Cohen DM. **1966**. A new tribe and a new species of ophidioid fish. *Proceedings of the Biological Society of Washington* 79: 183–204.

Cohen DM. **1981**. New and rare ophidiiform fishes from the eastern Atlantic: Canary Islands to the Cape of Good Hope. *Proceedings of the Biological Society of Washington* 94(4): 1085–1103.

Cohen DM. 1984. Gadiformes: overview. In: Moser HG, Richards WJ, Cohen DM, Fahay MP, Kendall SW Jr, Richardson SL (eds) Ontogeny and systematics of fishes. *American Society of Ichthyologists and Herpetologists Special Publication* 1: 259–264.

Cohen DM. **1990**. Family Bregmacerotidae (pp. 16–17). In: Cohen DM, Inada T, Iwamoto T, Scialabba N (eds) FAO species catalogue. Vol. 10. Gadiform fishes of the world (Order Gadiformes). An annotated and illustrated catalogue of cods, hakes, grenadiers and other gadiform fishes known to date. *FAO Fisheries Synopsis No. 125* (Vol. 10). Rome: FAO. 442 pp.

Cohen DM, Atsaides SP. **1969**. Additions to a revision of Argentinine fishes. *United States Fish and Wildlife Service Fishery Bulletin* 68(1): 13–36.

Cohen DM, Davis WP. **1969**. Vertical orientation in a new gobioid fish from New Britain. *Pacific Science* 23(3): 317–324.

Cohen DM, Nielsen JG. **1978**. Guide to the identification of genera of the fish order Ophidiiformes with a tentative classification of the order. *NOAA Technical Report* NMFS Circular No. 417: 1–72.

Cohen DM, Nielsen JG. **1982**. *Spottobrotula amaculata*, a new ophidiid fish from the Philippines. *Copeia* 1982(3): 497–500.

Cohen DM, Robins CR. **1986**. A review of the ophidiid fish genus *Sirembo* with a new species from Australia. *Memoirs of the Queensland Museum* 22(2): 252–263.

Cohen DM, Russo JL. **1979**. Variation in the fourbeard rockling, *Enchelyopus cimbrius*, a north Atlantic gadid fish, with comments on the genera of rocklings. *Fishery Bulletin* 77(1): 91–104.

Colborn J, Crabtree RE, Shaklee JB, Pfeiler E, Bowen BW. **2001**. The evolutionary enigma of bonefishes (*Albula* spp.): cryptic species and ancient separations in a globally distributed shorefish. *Evolution* 55(4): 807–820.

Cole KS. **2003**. Hermaphroditic characteristics of gonad morphology and inferences regarding reproductive biology in *Caracanthus* (Teleostei, Scorpaeniformes). *Copeia* 2003(1): 68–80.

Collett R. **1889**. Diagnoses de poissons nouveaux provenant des campagnes de *l'Hirondelle*. *Bulletin de la Société Zoologique de France* 14: 291–293.

Collett R. 1896. Poissons provenant des campagnes du yacht l'Hirondelle (1885–1888). Résultats des campagnes scientifiques accomplies sur son yacht par Albert 1<sup>er</sup> Monaco. Résultats des campagnes scientifiques du Prince de Monaco Fasc. 10: viii + 198 pp.

Collett R. **1905**. On some fishes from the sea off the Azores. *Zoologischer Anzeiger* 28(21/22): 723–730.

Collette BB. **1965**. Hemirhamphidae (Pisces: Synentognathi) from tropical West Africa. *Atlantide Report* 8: 217–235.

Collette BB. **1966**. A review of the venomous toadfishes, subfamily Thalassophryninae. *Copeia* 1966(4): 846–864.

Collette BB. **1966**. Revue critique des types de Scombridae des collections du Muséum National d'Histoire naturelle de Paris. *Bulletin du Muséum National d'Histoire Naturelle, Paris* (Série 2) 38(4): 362–375.

Collette BB. **1970**. *Rastrelliger kanagurta*, another Red Sea immigrant into the Mediterranean Sea, with a key to the Mediterranean species of Scombridae. *Bulletin of the Sea Fisheries Research Station, Haifa* 54: 3–6.

Collette BB. **1974**. The garfishes (Hemiramphidae) of Australia and New Zealand. *Records of the Australian Museum* 29(2): 11–105.

Collette BB. **1976**. Indo-west Pacific halfbeaks (Hemiramphidae) of the genus *Rhynchorhamphus* with descriptions of two new species. *Bulletin of Marine Science* 26(1): 72–98.

Collette BB. **1979**. Adaptations and systematics of the mackerels and tunas (pp. 7–39). In: Sharp GD, Dizon AE (eds) *The physiological ecology of tunas*. London: Academic Press. 485 pp.

Collette BB. **1982**. Status of two names for South Airican halfbeaks, *Hyporhamphus delagoae* (Barnard) and *Hyporhamphus improvisus* (Smith). *Copeia* 1982(3): 719–721.

Collette BB. **1982**. Rediscovery of *Hyporhamphus xanthopterus*, a half beak endemic to Vembanad Lake, Kerala, southern India. *Matsya* 7: 29–40.

Collette BB. **1983**. Recognition of two species of double-lined mackerels (*Grammatorcynus*: Scombridae). *Proceedings of the Biological Society of Washington* 96(4): 715–718.

Collette BB. **1984**. Belonidae. Needlefishes. In: Fischer W, Bianchi G (eds) *FAO species identification sheets for fishery purposes. Western Indian Ocean (Fishing Area 51)*. Vol. 1. Rome: FAO. [unpaginated]

Collette BB. **1984**. Coryphaenidae. Dolphinfishes, "dolphins". In: Fischer W, Bianchi G (eds) *FAO species identification sheets for fishery purposes. Western Indian Ocean (Fishing Area 51)*. Vol. 2. Rome: FAO. [unpaginated]

Collette BB. **1984**. Hemiramphidae. Halfbeaks. In: Fischer W, Bianchi G (eds) *FAO species identification sheets for fishery purposes. Western Indian Ocean (Fishing Area 51).* Vol. 2. Rome: FAO. [unpaginated]

Collette BB. **1984**. Scombridae. Albacores, bonitos, kawakawas, mackerels, seerfishes, tunas and wahoos. In: Fischer W, Bianchi G (eds) *FAO species identification sheets for fishery purposes. Western Indian Ocean (Fishing Area 51).* Vol. 4. Rome: FAO. [unpaginated]

Collette BB. **1999**. Hemiramphidae. Halfbeaks (pp. 2180–2196). In: Carpenter KE, Niem VH (eds) *FAO species identification guide for fishery purposes. The living marine resources of the western central Pacific.* Vol. 4. Bony fishes part 2 (pp. 2069–2790). Rome: FAO.

Collette BB. **1999**. Mackerels, molecules, and morphology (pp. 149–164). In: Séret B, Sire J-Y (eds) *Proceedings of the* 5<sup>th</sup> Indo-Pacific Fish Conference, Nouméa, New Caledonia, 3–8 November 1997. Paris: Société Française d'Ichtyologie. 866 pp.

Collette BB. 2001. Scombridae. Tunas (also, albacore, bonitos, mackerels, seerfishes, and wahoo) (pp. 3721–3756). In:
Carpenter KE, Niem VH (eds) FAO species identification guide for fishery purposes. The living marine resources of the western central Pacific. Vol. 6. Bony fishes part 4 (pp. 3381–4218). Rome: FAO.

Collette BB. **2002**. Mackerels, family Scombridae (pp. 516–536). In: Collette BB, Klein-MacPhee G (eds) *Bigelow and Schroeder's fishes of the Gulf of Maine* (3<sup>rd</sup> edition). Washington, DC: Smithsonian Institution Press. 882 pp.

Collette BB. **2003**. Family Scombridae Rafinesque 1815. Mackerels, tunas, and bonitos. *California Academy of Sciences Annotated Checklists of Fishes* No. 19. 28 pp.

Collette BB. **2004**. Family Hemiramphidae Gill 1859. Halfbeaks. *California Academy of Sciences Annotated Checklists of Fishes* No. 22. 35 pp.

Collette BB, Aadland CR. **1996**. Revision of the frigate tunas (Scombridae, *Auxis*), with descriptions of two new subspecies from the eastern Pacific. *Fishery Bulletin* 94(3): 423–441.

Collette BB, Amorim AF, Boustany A, Carpenter KE, De Oliveira Leite Jr N, Di Natale A, Die D, Fox W, Fredou FL, Graves J, Viera Hazin FH, Hinton M, Juan Jorda M, Kada O, Minte Vera C, Miyabe N, Nelson R, Oxenford H, Pollard D, Restrepo V, Schratwieser J, Teixeira Lessa RP, Pires Ferreira Travassos PE, Uozumi Y. **2011**. *Thunnus thynnus*. In: IUCN Red List of Threatened Species. Version 2012.2. www.iucnredlist.org

Collette BB, Berry FH. **1965**. Recent studies on the needlefishes (Belonidae): an evaluation. *Copeia* 1965(3): 386–392.

Collette BB, Carpenter KE, Polidoro BA, Juan-Jorda MJ, Boustany A, Die DJ, Elfes C, Fox W, Graves J, Harrison LR, McManus R, Minte-Vera CV, Nelson R, Restrepo V, Schratwieser J, Sun C-L, Amorim A, Brick Peres M, Canales C, Cardenas G, Chang S-K, Chiang W-C, De Oliveira Leite N Jr, Harwell H, Lessa R, Fredou FL, Oxenford HA, Serra R, Shao K-T, Sumaila R, Wang S-P, Watson R, Yáñez E.
2011. High value and long life—double jeopardy for tunas and billfishes. *Science* 333(6040): 291–292. http://dx.doi. org/10.1126/science.1208730

Collette B[B], Chang S-K, Di Natale A, Fox W, Juan Jorda M, Miyabe N, Nelson R, Uozumi Y, Wang S. **2011**. *Thunnus maccoyii*. In: IUCN Red List of Threatened Species. Version 2012.2. www.iucnredlist.org

Collette BB, Chao LN. **1975**. Systematics and morphology of the bonitos (*Sarda*) and their relatives (Scombridae, Sardini). *Fishery Bulletin* 73(3): 516–625.

Collette BB, Gillis GB. **1992**. Morphology, systematics, and biology of the double-lined mackerels (*Grammatorcynus*, Scombridae). *Fishery Bulletin* 90: 13–53.

Collette BB, Graves J, Kells V. **2019**. *Tunas and billfishes of the world*. Baltimore, Maryland: Johns Hopkins University Press. 351 pp.

Collette BB, McDowell JR, Graves JE. **2006**. Phylogeny of recent billfishes (Xiphioidei). *Bulletin of Marine Science* 79(3): 455–468.

Collette BB, McGowen GE, Parin NV, Mito S. 1984.
Beloniformes: development and relationships. In: Moser
HG, Richards WJ, Cohen DM, Fahay MP, Kendall SW Jr,
Richardson SL (eds) Ontogeny and systematics of fishes.
American Society of Ichthyologists and Herpetologists Special
Publication 1: 334–354.

Collette BB, Nauen CE. **1983**. FAO species catalogue. Vol. 2. Scombrids of the world. An annotated and illustrated catalogue of tunas, mackerels, bonitos and related species known to date. *FAO Fisheries Synopsis No. 125* (Vol. 2). Rome: FAO. 137 pp.

Collette BB, Parin NV. **1970**. Needlefishes (Belonidae) of the eastern Atlantic Ocean. *Atlantide Report* 11: 7–60.

Collette BB, Parin NV. **1978**. Five new species of halfbeaks (Hemiramphidae) from the Indo-West Pacific. *Proceedings of the Biological Society of Washington* 19(3): 731–747.

Collette BB, Parin NV. **1991**. Shallow-water fishes of Walters Shoals, Madagascar Ridge. *Bulletin of Marine Science* 48(1): 1–22. Collette BB, Potthoff T, Richards WJ, Ueyanagi S, Russo JL, Nishikawa Y. **1984**. Scombroidei: development and relationships. In: Moser HG, Richards WJ, Cohen DM, Fahay MP, Kendall SW Jr, Richardson SL (eds) Ontogeny and systematics of fishes. *American Society of Ichthyologists and Herpetologists Special Publication* 1: 591–620.

Collette BB, Reeb C, Block BA. **2001**. Systematics of the tunas and mackerels (Scombridae) (pp. 1–33). In: Block BA, Stevens ED (eds) *Tuna: Physiology, ecology, and evolution*. San Diego: Academic Press. 468 pp.

Collette BB, Russo JL. **1979**. An introduction to the Spanish mackerels, genus *Scomberomorus* (pp. 3–16). In: Nakamura EL, Bullis HR Jr (eds) *Proceedings: Colloquium on the Spanish and King Mackerel resources of the Gulf of Mexico*. Gulf States Marine Fisheries Commission Publication No. 4. vii + 117 pp.

Collette BB, Russo JL. **1985**. Morphology, systematics, and biology of the Spanish mackerels (*Scomberomorus*, Scombridae). *Fishery Bulletin* 82(4): 545–692. [1984 on article but actually published Oct. 1985 (BB Collette, pers. comm., 11/2002)]

Collette BB, Smith DG, Böhlke EB. **1991**. *Gymnothorax parini*, a new species of moray eel (Teleostei: Muraenidae) from Walters Shoals, Madagascar Ridge. *Proceedings of the Biological Society of Washington* 104(2): 344–350.

Collette BB, Su J-X. **1986**. The halfbeaks (Pisces, Beloniformes, Hemiramphidae) of the Far East. *Proceedings of the Academy of Natural Sciences of Philadelphia* 138(1): 250–302.

Collette BB, Uyeno T. **1972**. *Pontinus niger*, a synonym of the scorpionfish *Ectreposebastes imus*, with extension of its range to Japan. *Japanese Journal of Ichthyology* 19(1): 26–28.

Colwell RK, Chao A, Gotelli NJ, Lin S, Mao CX, Chazdon RL, Longino JT. 2012. Models and estimators linking individualbased and sample-based rarefaction, extrapolation and comparison of assemblages. *Journal of Plant Ecology* 5(1): 3–21. http://dx.doi.org/10.1093/jpe/rtr044

Compagno LJV. **1973**. *Ctenacis* and *Gollum*, two new genera of sharks (Selachii; Carcharhinidae). *Proceedings of the California Academy of Sciences* (Series 4) 39(14): 257–272.

Compagno LJV. **1973**. *Gogolia filewoodi*, a new genus and species of shark from New Guinea (Carcharhiniformes: Triakidae), with a redefinition of the family Triakidae and a key to the genera. *Proceedings of the California Academy of Sciences* (Series 4) 39(19): 383–410.

Compagno LJV. **1973**. Interrelationships of living elasmobranchs (pp. 15–61). In: Greenwood PH, Miles RS, Patterson C (eds) *Interrelationships of fishes*. London: Academic Press. 536 pp.

Compagno LJV. **1977**. Phyletic relationships of living sharks and rays. *American Zoologist* 17(2): 303–322.

Compagno LJV. **1984**. Chimaeras. In: Fischer W, Bianchi G (eds) *FAO species identification sheets for fishery purposes. Western Indian Ocean (Fishing Area 51)*. Vol. 5. Rome: FAO. [unpaginated]

Compagno LJV. 1984. Sharks. In: Fischer W, Bianchi G
(eds) FAO species identification sheets for fishery purposes.
Western Indian Ocean (Fishing Area 51). Vol. 5. Rome: FAO.
[unpaginated]

Compagno LJV. **1984**. FAO species catalogue. Vol. 4. Sharks of the world. An annotated and illustrated catalogue of shark species known to date. Part 1. Hexanchiformes to Lamniformes. *FAO Fisheries Synopsis No. 125* (Vol. 4, part 1, pp. 1–249). Rome: FAO.

Compagno LJV. **1984**. FAO species catalogue. Vol. 4. Sharks of the world. An annotated and illustrated catalogue of shark species known to date. Part 2. Carcharhiniformes. *FAO Fisheries Synopsis No. 125* (Vol. 4, part 2, pp. 251–655). Rome: FAO.

Compagno LJV. **1986**. Sharks of the Eastern Cape coast. *Ichthos Field Guide* No. 1. Grahamstown: J.L.B. Smith Institute of Ichthyology. 27 pp.

Compagno, LJV. 1988. Scyliorhinus comoroensis sp. n., a new catshark from the Comoro Islands, western Indian Ocean (Carcharhiniformes, Scyliorhinidae). Bulletin du Muséum National d'Histoire Naturelle, Paris (Série 4) 10(3): 603–625.

Compagno LJV. **1988**. *Sharks of the Order Carcharhiniformes*. Princeton, New Jersey: Princeton University Press. xxii + 572 pp.

Compagno LJV. **1990**. Alternate life-history styles of cartilaginous fishes in time and space. *Environmental Biology of Fishes* 28: 33–75.

Compagno LJV. **1990**. Relationships of the megamouth shark, *Megachasma pelagios* (Lamniformes: Megachasmidae), with comments on its feeding habits (pp. 357–379). In: Pratt HL Jr, Gruber SH, Taniuchi T (eds) Elasmobranchs as living resources: Advances in the biology, ecology, systematics, and the status of the fisheries. *NOAA Technical Report* NMFS 90: viii + 526 pp.

Compagno LJV. **1990**. Sharks. Lamnidae and Squalidae (pp. 81–85). In: Gon O, Heemstra PC (eds) *Fishes of the Southern Ocean*. Grahamstown: J.L.B. Smith Institute of Ichthyology. 462 pp.

Compagno LJV. **1999**. An overview of chondrichthyan systematics and biodiversity in southern Africa. *Transactions of the Royal Society of South Africa* 54(1): 75–120.

Compagno LJV. **1999**. Checklist of living elasmobranchs (pp. 471–498). In: Hamlett WC (ed) *Sharks, skates, and rays: the biology of elasmobranch fishes*. Baltimore, Maryland: Johns Hopkins University Press. 528 pp. Compagno LJV. **1999**. Chimaeras (pp. 1532–1537). In: Carpenter KE, Niem VH (eds) *Species identification guide for fisheries purposes. The living marine resources of the western central Pacific.* Vol. 3. Batoid fishes, chimaeras and bony fishes part 1 (pp. 1397–2068). Rome: FAO.

Compagno LJV. **2001**. Sharks of the world. An annotated and illustrated catalogue of shark species known to date. Vol. 2: Bullhead, mackerel and carpet sharks (Heterodontiformes, Lamniformes and Orectolobiformes). *FAO Species Catalogue for Fishery Purposes* No. 1 (Vol. 2). Rome: FAO. 269 pp.

Compagno LJV. **2003**. *Sharks of the Order Carcharhiniformes*. 1<sup>st</sup> reprint. Caldwell, New Jersey: The Blackburn Press. xii + 572 pp.

Compagno LJV. **2005**. Checklist of living Chondrichthyes (pp. 503–548). In: Hamlett WC (ed) *Reproductive biology and phylogeny of Chondrichthyes: Sharks, batoids and chimaeras.* Vol. 3 of Series: Reproductive biology and phylogeny. Boca Raton, Florida: CRC Press. 572 pp.

Compagno LJV, Dagit DD. **2006**. *Hydrolagus africanus*. In: IUCN Red List of Threatened Species. Version 2010.1. www.iucnredlist.org

Compagno LJV, Dando M, Fowler S. **2005**. *A field guide to the sharks of the world*. London: Harper-Collins. 368 pp. + 128 pages of plates.

Compagno LJV, Dando M, Fowler S. **2005**. *Sharks of the world*. Princeton Field Guides. Princeton, New Jersey: Princeton University Press. 368 pp. + 127 pages of plates.

Compagno LJV, Ebert DA. **2007**. Southern African skate biodiversity and distribution. *Environmental Biology of Fishes* 80(2): 125–145.

Compagno LJV, Ebert DA, Cowley PD. **1991**. Distribution of offshore demersal cartilaginous fishes (Class Chondrichthyes) off the west coast of southern Africa, with notes on their systematics. *South African Journal of Marine Science* 11: 43–139.

Compagno LJV, Ebert DA, Smale MJ. 1989. Guide to the sharks and rays of southern Africa. Cape Town: Struik Publishers.
160 pp. [Also published in 1989 New Holland Publishers Ltd, London]

Compagno LJV, Follett WI. **1986**. *Carcharias* Rafinesque, 1810 (Chondrichthyes, Lamniformes): proposed conservation by the use of the relative precedence procedure. *Bulletin of Zoological Nomenclature* 43(1): 89–92.

Compagno LJV, Heemstra PC. **1984**. *Himantura draco*, a new species of stingray (Myliobatiformes: Dasyatidae) from South Africa, with a key to the Dasyatidae and the first record of *Dasyatis kuhlii* (Müller & Henle, 1841) from Southern Africa. *J.L.B. Smith Institute of Ichthyology Special Publication* 33: 1–17.

Compagno LJV, Heemstra PC. **2007**. *Electrolux addisoni*, a new genus and species of electric ray from the east coast of South Africa (Rajiformes: Torpedinoidei: Narkidae), with a review of torpedinoid taxonomy. *Smithiana* Bulletin 7: 15–49.

Compagno LJV, Krupp F, Carpenter KE. **1996**. A new weasel shark of the genus *Paragaleus* from the northwestern Indian Ocean and the Arabian Gulf (Carcharhiniformes: Hemigaleidae). *Fauna of Saudi Arabia* 15: 391–401.

Compagno LJV, Last PR. 1999. Pristidae. Sawfishes (pp. 1410–1417); Rhinidae (= Rhynchobatidae). Wedgefishes (pp. 1418–1422); Rhinobatidae. Guitarfishes (pp. 1423–1429); Narkidae. Sleeper rays (pp. 1443–1446). In: Carpenter KE, Niem VH (eds) FAO species identification guide for fishery purposes. The living marine resources of the western central Pacific. Vol. 3. Batoid fishes, chimaeras and bony fishes part 1 (pp. 1397–2068). Rome: FAO.

Compagno LJV, Last PR. **1999**. Gymnuridae. Butterfly rays (pp. 1506–1510); Myliobatidae. Eagle rays (pp. 1511–1519); Rhinopteridae. Cownose rays (pp. 1520–1523); Devil rays (pp. 1524–1529). In: Carpenter KE, Niem VH (eds) *FAO species identification guide for fishery purposes. The living marine resources of the western central Pacific.* Vol. 3. Batoid fishes, chimaeras and bony fishes part 1 (pp. 1397–2068). Rome: FAO.

Compagno LJV, Last PR. 2008. A new species of wedgefish, *Rhynchobatus palpebratus* sp. nov. (Rhynchobatoidei: Rhynchobatidae), from the Indo-West Pacific (pp. 227–240).
In: Last PR, White WT, Pogonoski JJ (eds) *Descriptions of new Australian chondrichthyans*. Hobart, Australia: CSIRO Marine and Atmospheric Research. 358 pp.

Compagno LJV, Randall JE. **1987**. *Rhinobatos punctifer*, a new species of guitarfish (Rhinobatiformes: Rhinobatidae) from the Red Sea, with notes on the Red Sea batoid fauna. *Proceedings of the California Academy of Sciences* (Series 4) 44(14): 335–342.

Compagno LJV, Roberts TR. **1982**. Freshwater stingrays (Dasyatidae) of southeast Asia and New Guinea, with description of a new species of *Himantura* and reports of unidentified species. *Environmental Biology of Fishes* 7(4): 321–339.

Compagno LJV, Smale MJ. **1985**. *Paragaleus leucolomatus*, a new shark from South Africa, with notes on the systematics of hemigaleid sharks (Carcharhiniformes: Hemigaleidae). *J.L.B. Smith Institute of Ichthyology Special Publication* 37: 1–21.

Compagno LJV, Smale MJ. **1986**. Recent records of four warmwater elasmobranchs from the eastern Cape Province, South Africa. *South African Journal of Marine Science* 4(1): 11–15.

Compagno LJV, Springer S. **1971**. *lago*, a new genus of carcharhinid sharks with a redescription of *I. omanensis*. *Fishery Bulletin* 69(3): 615–626.

Compagno LJV, Stehmann M, Ebert DA. **1990**. *Rhinochimaera africana*, a new longnose chimaera from southern Africa, with comments on the systematics and distribution of the genus *Rhinochimaera* Garman, 1901 (Chondrichthyes, Chimaeriformes, Rhinochimaeridae). *South African Journal of Marine Science* 9(1): 201–222.

Compagno LJV, Stevens JD. **1993**. *Atelomycterus fasciatus* n.sp., a new catshark (Chondrichthyes: Carcharhiniformes: Scyliorhinidae) from tropical Australia. *Records of the Australian Museum* 45(2): 147–169.

Compagno LJV, Stevens JD. **1993**. *Galeus gracilis* n.sp., a new sawtail catshark from Australia, with comments on the systematics of the genus *Galeus* Rafinesque, 1810 (Carcharhiniformes: Scyliorhinidae). *Records of the Australian Museum* 45(2): 171–194.

Compagno LJV, Talwar PK. **1985**. Generic relationship and status of the scyliorhinid shark, *Scyliorhinus* (*Halaelurus*) *silasi* Talwar, 1974 (Chondrichthyes: Selachii, Scyliorhinidae). *Bulletin of the Zoological Survey of India* 7(1): 37–39.

Condé, B. **1977**. Le Pélor à filaments, poisson-pierre de l'île Maurice (Scorpenides, Inimicines). *Revue française d'Aquariologie Herpétologie* 4(1): 15–18.

Condé B. **1977**. Nouvelles observations sur les scorpénidés du *Rhinopias* à Maurice. *Revue française d'Aquariologie Herpétologie* 4(1): 19–20.

Condé B. **1983**. Redécouverte de *Calloplesiops argus* Fowler et Bean, 1930 (Perciformes: Plesiopidae): Note préliminaire. *Revue française d'Aquariologie Herpétologie* 10(3): 87–92.

Condé B, Jauffret LP. **1978**. Quelques poissons intéressants de l'île Maurice. *Revue française d'Aquariologie Herpétologie* 4: 107–114.

Connell AD. **1993**. New records of fish species for South African waters. *African Wildlife* 47(5): 199–203.

Connell AD. **1996**. Sea fishes spawning pelagic eggs in the St Lucia estuary. *South African Journal of Zoology* 31(1): 37–41.

Connell AD, Victor BC, Randall JE. **2015**. A new species of *Pseudojuloides* (Perciformes: Labridae) from the southwestern Indian Ocean. *Journal of the Ocean Science Foundation* 14: 49–56.

http://dx.doi.org/10.5281/zenodo.1037600

Cooke A, Lutjeharms JRE, Vasseur P. **2004**. Marine and coastal ecosystems (pp. 179–209). In: Goodman SM, Benstead JP (eds) *The natural history of Madagascar*. Chicago: The University of Chicago Press. 1709 pp.

Cooper JA, Chapleau F. **1998**. Phylogenetic status of *Paralichthodes algoenis* (Pleuronectiformes: Paralichthodidae). *Copeia* 1998(2): 477–481.

Cope ED. **1871**. Contribution to the ichthyology of the Lesser Antilles. *Transactions of the American Philosophical Society* 14(3): 445–483.

Cope ED. **1889**. Synopsis of the families of Vertebrata. *American Naturalist* 23(274): 849–877.

Cornic A. **1987**. *Poissons de l'île Maurice*. Stanley, Rose Hill, Ile Maurice: Editions de l'Océan Indien. 335 pp.

Corsini M, Margies P, Kondilatos G, Economidis PS. **2006**. Three new exotic fish records from the SE Aegean Greek waters. *Scientia Marina* 70(2): 319–323.

Courtenay WR, McKittrick FA. **1970**. Sound producing mechanisms in carapid fishes, with notes on phlyogenetic implications. *Marine Biology* 7(2): 131–137.

Cowley PD, Compagno LJV. **1993**. A taxonomic re-evaluation of the blue stingray from southern Africa (Myliobatiformes: Dasyatidae). *South African Journal of Marine Science* 13: 135–149.

Cowman PF, Bellwood DR. **2011**. Coral reefs as drivers of cladogenesis: expanding coral reefs, cryptic extinction events, and the development of biodiversity hotspots. *Journal of Evolutionary Biology* 24(12): 2543–2562. http://dx.doi.org/10.1111/j.1420-9101.2011.02391.x

Cowman PF, Bellwood DR, van Herwerden L. **2009**. Dating the evolutionary origins of wrasse lineages (Labridae) and the rise of trophic novelty on coral reefs. *Molecular Phylogenetics and Evolution* 52(3): 621–631. http://dx.doi.org/10.1016/j.ympev.2009.05.015

Cozzi J, Clark E. **1995**. Darting behavior of a sandburrower fish, *Limnichthys nitidus* (Creediidae), in the Red Sea. *Environmental Biology of Fishes* 44(4): 327–336.

Craig MT. **2007**. Preliminary observations on the life history of the white-streaked grouper, *Epinephelus ongus*, from Okinawa, Japan. *Ichthyological Research* 54: 81–84.

Craig MT, Bogorodsky SV, Randall JE, Mal AO. **2015**. *Lepadichthys bilineatus*, a new species of clingfish from Oman (Teleostei: Gobiesocidae), with a redescription of *Lepadichthys erythraeus* Briggs and link from the Red Sea. *Zootaxa* 3990(1): 113–122. https://doi.org/10.11646/ zootaxa.3990.1.6

Craig MT, Hastings PA. **2007**. A molecular phylogeny of the groupers of the subfamily Epinephelinae (Serranidae) with a revised classification of the Epinephelini. *Ichthyological Research* 54(1): 1–17.

Craig MT, Hastings PA, Pondella DJ. 2004. Notes on the systematics of the crestfish genus *Lophotus* (Lampridiformes: Lophotidae), with a new record from California. *Bulletin of the Southern California Academy of Sciences* 103(2): 57–65.

Craig MT, Randall JE. **2008**. Two new species of the Indo-Pacific clingfish genus *Discotrema* (Gobiesocidae). *Copeia* 2008(1): 68–74. http://dx.doi.org/10.1643/CI-07-025

Craig MT, Randall JE. **2009**. *Briggsia hastingsi*, a new genus and species of clingfish from Oman. *Zootaxa* 2271: 64–68. http://dx.doi.org/10.11646/zootaxa.2271.1.6 Craig MT, Sadovy de Mitcheson YJ, Heemstra PC (eds). **2011**. *Groupers of the world: A field and market guide.* Grahamstown, South Africa: NISC (Pty) Ltd. xx + 356 + A:47.

Crame JA. **1980**. Succession and diversity in the Pleistocene coral reefs of the Kenya coast. *Palaeontology* 23: 1–37.

Crame JA. **1981**. Ecological stratification in the Pleistocene coral reefs of the Kenya coast. *Palaeontology* 24: 609–646.

Crame JA. **1986**. Late Pleistocene molluscan assemblages from the coral reefs of the Kenya coast. *Coral Reefs* 4: 183–196.

Cramer F. **1895**. On the cranial characters of the genus Sebastodes (rock-fish). Proceedings of the California Academy of Sciences (Series 2) 5: 573–610. [Reprinted as Contributions of the Biological Hopkins Seaside Laboratory (Leland Stanford Jr. University Publications) No. 2: 573–610]

Crawford RJM. **1981**. Distribution, availability and movements of pilchard *Sardinops ocellata* off South Africa, 1964–1976. *Fisheries Bulletin* (South Africa) 14: 1–46.

Crawford RJM. **1981**. Distribution, availability and movements of anchovy *Engraulis capensis* off South Africa, 1964–1976. *Fisheries Bulletin* (South Africa) 14: 51–94.

Cressey RF. **1981**. Revision of Indo-West Pacific lizardfishes of the genus *Synodus* (Pisces: Synodontidae). *Smithsonian Contributions to Zoology* 342: iii + 53.

Cressey RF, Collette BB. **1971**. Copepods and needlefishes: a study in host-parasite relationships. *Fishery Bulletin* 68(3): 347–432.

Cressey RF, Cressey HB. **1979**. The parasitic copepods of Indo-West Pacific lizardfishes (Synodontidae). *Smithsonian Contributions to Zoology* 296: 1–71.

Cressey RF, Lachner EA. **1970**. The parasitic copepod diet and life history of diskfishes (Echeneidae). *Copeia* 1970(2): 310–318.

Cressey RF, Waples RS. **1984**. Synodontidae. Lizardfishes. In: Fischer W, Bianchi G (eds) *FAO species identification sheets for fishery purposes. Western Indian Ocean (Fishing Area 51).* Vol. 4. Rome: FAO.

Crowley LELM, Ivantsoff W. **1988**. A new species of Australian *Craterocephalus* (Pisces: Atherinidae) and redescription of four other species. *Records of the Western Australian Museum* 14(2): 151–169.

Cumbaa SL, Murray AM. **2008**. New Late Cretaceous pachyrhizodontid and enchodontoid fishes and associated ichthyofauna from the Northwest Territories, Canada (pp. 229–256). In: Arratia G, Schultze H-P, Wilson MVH (eds) *Mesozoic Fishes 4 – Homology and phylogeny. Proceedings of the international meeting, Miraflores de la Sierra, 2005*. Munich: Verlag Dr. Friedrich Pfeil. 502 pp.

Cunningham JT. **1910**. On the marine fishes and invertebrates of St. Helena. *Proceedings of the Zoological Society of London* 1910(pt 1): 86–131.

Curtiss A. **1944**. Further notes on the zoology of Tahiti. *Zoology of Tahiti* (Supplement): 1–30.

Cushing D. **1971**. Survey of resources in the Indian Ocean and Indonesian area. FAO/UNDP Indian Ocean Programme. IOFC/DEV/71/2.

Cuvier G. **1814**. Observations et recherches critiques sur différens poissons de la Méditerranée et, à leur occasion, sur des poissons des autres mers plus ou moins liés avec eux [in a report by Desmarest AG]. *Bulletin des Sciences, par la Société Philomathique de Paris* (Série 3) 1: 80–92.

Cuvier G. 1816. Le règne animal distribué d'après son organisation pour servir de base à l'histoire naturelle des animaux et d'introduction à l'anatomie comparée. Les reptiles, les poissons, les mollusques et les annélides. (1<sup>st</sup> edition, 4 volumes; Poissons in Vol. II, pp. 104–351; Pls. 9–10 in Vol. IV). Paris: Deterville. [Date of 1817 on publication]

Cuvier G. **1817**. Sur le genre Chironectes Cuv. (Antennarius Commers.). *Mémoires du Muséum National d'Histoire Naturelle* (Paris) 3: 418–435.

Cuvier G. **1818**. Sur les Diodons, vulgairement orbes-épineux. *Mémoires du Muséum National d'Histoire Naturelle* (Paris) 4: 121–138.

Cuvier G. **1826–1836**. *Histoire des progrès des sciences naturelles depuis 1789 jusqu'à ce jour* (5 volumes).

Cuvier G. **1828**. *Historical portrait of the progress of ichthyology, from its origins to our own time.* **See** Pietsch TW. 1995.

Cuvier G. **1829–1830**. Le règne animal, distribué d'après son organisation, pour servir de base à l'histoire naturelle des animaux et d'introduction à l'anatomie comparée (nouvelle édition, 5 volumes; Poissons in Vol. II [1829], pp. 122–406). Paris: Déterville.

Cuvier G. 1830. In: Cuvier G, Valenciennes A.

Cuvier G. 1836. Le règne animal distribué d'après son organisation, pour servir de base à l'histoire naturelle des animaux et d'introduction à l'anatomie comparée ("Disciples" [Troisième] edition). Les poissons avec un Atlas par M.A. Valenciennes. Paris: Fortin, Masson. 392 pp.

Cuvier G. **1837**. *The animal kingdom, arranged according to its organization, serving as a foundation for the natural history of animals and an introduction to comparative anatomy* (4 volumes; Vol. II. Reptiles–Fishes). London: G Henderson. [English translation of *Le règne animal*]

Cuvier G, Valenciennes A. **1828–1850**. *Histoire naturelle des poissons* (22 volumes). Paris: F.G. Levrault.

## D

Da Franca P. **1960**. Nova contribuição para o conhecimento do género *Merluccius* no Atlântico oriental ao sul do Equador. *Memórias da Junta de Investigações Científicas do Ultramar* (Série 2a) 18: 57–101.

Da Franca P. **1962**. Considerations sur la taxonomie des *Merluccius* de l'Atlantique oriental. *Memórias da Junta de Investigações Científicas do Ultramar* (Series 2) 36: 7–48.

Da Franca P, da Franca MLP. **1969**. On the occurrence of a species of *Cottunculus* (Pisces: Perciformes, Cortunculidae) in Angola waters. *Notas do Centro Biologica Aquatica Tropical* (Lisboa) 13: 1–15. [In Portuguese]

Dafni J, Diamant A. **1984**. School-oriented mimicry, a new type of mimicry in fishes. *Marine Ecology Progress Series* 20: 45–50.

Daget J, Gosse J-P, Thys van den Audenaerde DFE (eds). 1986. Check-list of the freshwater fishes of Africa (CLOFFA) (3 volumes). Brussels: Institut Royal des Sciences Naturelles de Belgique. MRAC Tervuren, ORSTOM, Paris. Vol. 1. 429 pp.; Vol. 2. 521 pp.; Vol. 3. 278 pp.

Dagit DD, Compagno LJV. **2006**. *Chimaera jordani*. In: IUCN Red List of Threatened Species. Version 2010.1. www.iucnredlist.org

D'Ancona U. **1928**. Murenoidi (Apodes) del Mar Rosso e del Golfo di Aden. Materiali raccolti dal Prof. Luigi Sanzo nella Campagna della R.N. *Ammiraglio Magnaghi* 1923–24. *Memoria, Comitato Talassografico Italiano* 146: 1–146.

D'Ancona U, Cavinato G. **1965**. The fishes of the family Bregmacerotidae. *Dana Report* 64: 1–91.

Darling ES, Green SJ, O'Leary JK, Côté IM. **2011**. Indo-Pacific lionfish are larger and more abundant on invaded reefs: a comparison of Kenyan and Bahamian lionfish populations. *Biological Invasions* 13(9): 2045–2051. http://dx.doi. org/10.1007/s10530-011-0020-0

Darwall WRT, Guard M. **2000**. Chapter 5. Southern Tanzania (pp. 131–165). In: McClanahan TR, Sheppard CRC, Obura DO (eds) *Coral reefs of the Indian Ocean: Their ecology and conservation*. Oxford: Oxford University Press. xxiii + 525 pp.

Darwin C. **1842**. *The structure and distribution of coral reefs. Being the first part of the geology of the voyage of the* Beagle, *under the command of Capt, Fitzroy, R.N. during the years 1832 to 1836*. London: Smith Elder & Co. 217 pp.

Darwin C. **1859**. *On the origin of species by means of natural selection, or the preservation of favoured races in the struggle for life*. London: John Murray, Albemarle Street. 502 pp.

Das A. **2003**. A catalogue of new taxa described by scientists of the zoological survey of India during 1916–1991. *Records of the Zoological Survey of India, Occasional Paper* 208: 1–530.

Das MK, Nelson JS. **1996**. Revision of the percophid genus *Bembrops* (Actinopterygii: Perciformes). *Bulletin of Marine Science* 59(1): 9–44.

Dasilao JC Jr, Sasaki K, Okamura K. **1997**. The hemiramphid, *Oxyporhamphus*, is a flyingfish (Exocoetidae). *Ichthyological Research* 44: 101–107.

Datta NC, Chaudhuri S. **1996**. A monograph on the taxonomy of the ambassid fishes of India. *Journal of the Asiatic Society* (Calcutta) 38(4): 1–30.

D'Aubrey JD. **1964**. Preliminary guide to the sharks found off the east coast of South Africa. *Investigational Report*. *Oceanographic Research Institute* (Durban) 8: 1–95.

D'Aubrey JD. **1964**. A carchariid shark new to South African waters. *Investigational Report. Oceanographic Research Institute* (Durban) 9: 1–16.

D'Aubrey JD. **1969**. Two species of shark new to South African waters. *Bulletin of the South African Association for Marine Biological Research* 7: 30–37.

D'Aubrey JD. **1971**. The taxonomy of two shark species of the genus *Carcharhinus*. MSc thesis, University of Natal, Durban. 171 pp.

Daudin FM. **1816**. Antennaire. *Dictionaire des sciences naturelles* (2<sup>nd</sup> edition, Vol. 2, p. 193). Paris: F.G. Levrault.

David JHM. **1987**. Diet of the South African fur seal (1974–1985) and an assessment of competition with fisheries in southern Africa. *South African Journal of Marine Science* 5(1): 693–713.

David LR. **1973**. Miocene fishes of Southern California. *Geological Society of America Special Papers* 43: xiii–xiv.

Davies DH. **1948**. A new goby from the Knysna River. *Annals and Magazine of Natural History* (Series 12) 1(5): 375–376.

Davies DH. 1949. Preliminary investigations on the foods of South African fishes (with notes on the general fauna of the area surveyed). *Investigational Report* (Fisheries and Marine Biological Survey Division, Union of South Africa) 11: 1–36. [Reprint from "Commerce & Industry", Jan. 1949.]

Davies DH. **1950**. A new species of *Macruronus* from South Africa. *Annals and Magazine of Natural History* (Series 12) 3(30): 512–515.

Davis MP. **2010**. Evolutionary relationships of the Aulopiformes (Euteleostei: Cyclosquamata): a molecular and total evidence approach (pp. 431–470). In: Nelson JS, Schultze H-P, Wilson MVH (eds) *Origin and phylogenetic interrelationships of teleosts*. Munich: Verlag Dr. Friedrich Pfeil. 482 pp.

Davis WP, Cohen DM. **1969**. A gobiid fish and a palaemonid shrimp living on an antipatharian sea whip in the tropical Pacific. *Bulletin of Marine Science* **18**(4): 749–761.

Davis WP, Fricke R. 1990. Callionymidae (pp. 921–924).
In: Quéro J-C, Hureau J-C, Karrer K, Post A, Saldanha L (eds) *Check-list of the fishes of the eastern tropical Atlantic* (*CLOFETA*). Vol. 2. Lisbon: Junta Nacional de Investigação Científica e Tecnológica.

Dawson CE. **1967**. *Paragunnellichthys seychellensis*, a new genus and species of gobioid fish (Microdesmidae) from the western Indian Ocean. *Proceedings of the Biological Society of Washington* 80(13): 73–81.

Dawson CE. 1968. Two new wormfishes (Gobioidea: Microdesmidae) from the Indian Ocean. Proceedings of the Biological Society of Washington 81(8): 53–68. Dawson CE. **1969**. *Paragunnellichthys fehlmanni*, a new gobioid fish (Microdesmidae) from the Indian Ocean. *Proceedings of the Biological Society of Washington* 82: 373–380.

Dawson CE. 1970. A new wormfish (Gobioidae: Microdesmidae) from the northern Red Sea. Proceedings of the Biological Society of Washington 83(25): 267–272.

Dawson CE. **1973**. Indo-Pacific distribution of microdesmid fishes (Gobioidea). *Journal of the Marine Biological Association of India* 15(1): 318–322.

Dawson CE. **1976**. Review of the Indo-Pacific pipefish genus *Choeroichthys* (Pisces: Syngnathidae), with descriptions of two new species. *Proceedings of the Biological Society of Washington* 89(3): 39–66.

Dawson CE. **1977**. Review of the Indo-Pacific pipefish genus *Lissocampus* (Syngnathidae). *Proceedings of the Biological Society of Washington* 89(53): 599–620.

Dawson CE. **1977**. Review of the pipefish genus *Corythoichthys* with description of three new species. *Copeia* 1977(2): 295–338.

Dawson CE. **1977**. Synopsis of syngnathine pipefishes usually referred to the genus *Ichthyocampus* Kaup, with description of new genera and species. *Bulletin of Marine Science* 27(4): 595–650.

Dawson CE. **1978**. Review of the Indo-Pacific pipefish genus *Hippichthys* (Syngnathidae). *Proceedings of the Biological Society of Washington* 91(1): 132–157.

Dawson CE. 1979. The Indo-Pacific pipefish genera Notiocampus gen. nov. and Nannocampus Günther. Proceedings of the Biological Society of Washington 92(3): 482–493.

Dawson CE. 1979. Notes on western Atlantic pipefishes with description of Syngnathus caribbaeus n. sp. and Cosmocampus n. gen. Proceedings of the Biological Society of Washington 92(4): 671–676.

Dawson CE. **1979**. Review of the polytypic doryrhamphine pipefish *Oostethus brachyurus* (Bleeker). *Bulletin of Marine Science* 29(4): 465–480.

Dawson CE. **1980**. Notes on some *Siboga* Expedition pipefishes previously referred to the genus *Syngnathus*. *Bijdragen tot de Dierkunde* 50(1): 221–226.

Dawson CE. **1981**. Notes on four pipefishes (Syngnathidae) from the Persian Gulf. *Copeia* 1981(1): 87–95.

Dawson CE. **1981**. Review of the Indo-Pacific pipefish genus *Doryrhampus* Kaup (Pisces: Syngnathidae), with descriptions of a new species and a new subspecies. *Ichthyological Bulletin of the J.L.B. Smith Institute of Ichthyology* 44: 1–27.

Dawson CE. 1982. Family Syngnathidae: the pipefishes.
Subfamilies Doryrhamphinae and Syngnathinae. In:
Fishes of the western North Atlantic. *Memoir of the Sears Foundation for Marine Research* 1(part 8): 1–172. Dawson CE. **1982**. Review of the genus *Micrognathus* Duncker (Pisces; Syngnathidae), with description of *M. natans*, n. sp. *Proceedings of the Biological Society of Washington* 95(4): 657–687.

Dawson CE. **1982**. Synopsis of the Indo-Pacific genus *Solegnathus* (Pisces: Syngnathidae). *Japanese Journal of Ichthyology* 29(2): 139–161.

Dawson CE. **1983**. Synopsis of the Indo-Pacific pipefish genus *Siokunichthys* (Syngnathidae), with description of *S. nigrolineatus* n. sp. *Pacific Science* 37(1): 49–63.

Dawson CE. **1984**. A new pipehorse (Syngnathidae) from Western Australia, with remarks on the subgenera of *Acentronura. Japanese Journal of Ichthyology* 31(2): 156–160.

Dawson CE. **1984**. *Bulbonaricus* Herald (Pisces: Syngnathidae), a senior synonym of *Enchelyocampus* Dawson and Allen, with description of *Bulbonaricus brucei* n. sp. from eastern Africa. *Copeia* 1984(3): 565–571.

Dawson CE. **1984**. *Halicampus zavorensis* and *H. inarquesensis*, new species of pipefishes (Syngnathidae) from Mozambique and the Marquesas Islands. *J.L.B. Smith Institute of Ichthyology Special Publication* 34: 1–7.

Dawson CE. **1984**. Review of the Indo-Pacific pipefish genus *Trachyrhamphus* (Syngnathidae). *Micronesica* 18(2): 163–191.

Dawson CE. **1984**. Revision of the genus *Microphis* Kaup (Pisces: Syngnathidae). *Bulletin of Marine Science* 35(2): 117–181.

Dawson CE. **1985**. *Indo-Pacific pipefishes (Red Sea to the Americas)*. Ocean Springs, Mississippi: Gulf Coast Research Laboratory. 230 pp.

Dawson CE. **1986**. Syngnathidae (pp. 628–639). In: Whitehead PJP, Bauchot M-L, Hureau J-C, Nielsen J, Tortonese E (eds) *Fishes of the north-eastern Atlantic and the Mediterranean*. Vol. II. Paris: UNESCO.

Dawson CE, Allen GR. **1978**. Synopsis of the 'finless' pipefish genera (*Penetopteryx*, *Apterygocampus* and *Enchelyocampus*, gen. nov.). *Records of the Western Australian Museum* 6(4): 391–411.

Dawson CE, Allen GR. **1981**. *Micrognathus spinirostris*, a new Indo-Pacific pipefish (Syngnathidae). *Journal of the Royal Society of Western Australia* 64(pt 2): 65–68.

Dawson CE, Randall JE. **1975**. Notes on Indo-Pacific pipefishes (Pisces: Syngnathidae) with description of two new species. *Proceedings of the Biological Society of Washington* 88(25): 263–280.

Dawson CE, Yasuda F, Imai C. **1979**. Elongate dermal appendages in species of *Yozia* (Syngnathidae) with remarks on *Trachyrhamphus*. *Japanese Journal of Ichthyology* 25(4): 244–250.

Day F. **1865**. *The fishes of Malabar*. London: Bernard Quaritch. 293 pp.

Day F. **1865**. On the fishes of Cochin, on the Malabar coast of India. Part I. Acanthopterygii. *Proceedings of the Zoological Society of London* 1865 (part 1): 2–40.

Day F. **1865**. On the fishes of Cochin, on the Malabar coast of India. Part II. Anacanthini. *Proceedings of the Zoological Society of London* 1865 (part 1): 286–318.

Day F. **1867**. On some new or imperfectly known fishes of Madras. *Proceedings of the Zoological Society of London* 1867 (part 2): 935–942.

Day F. **1867**. On some new or imperfectly known fishes of India. *Proceedings of the Zoological Society of London* 1867 (part 3): 699–707.

Day F. **1868**. On some new or imperfectly known fishes of India. *Proceedings of the Zoological Society of London* 1868 (part 1): 149–156.

Day F. **1868**. On some new fishes from Madras. *Proceedings of the Zoological Society of London* 1868 (part 1): 192–199.

Day F. 1870. Remarks on some of the fishes in the Calcutta Museum. Part I. Proceedings of the Zoological Society of London 1869 (part 3) [for 11 November 1869]: 511–527.

Day F. **1870**. Remarks on some of the fishes in the Calcutta Museum. Part II. *Proceedings of the Zoological Society of London* 1869 (part 3) [for 25 November 1869]: 548–560.

Day F. **1870**. On some of the fishes in the Calcutta Museum. Part III. *Proceedings of the Zoological Society of London* 1869 (part 3) [for 9 December 1869]: 611–614.

Day F. **1871**. On the fishes of the Andaman Islands. *Proceedings* of the Zoological Society of London 1870 (part 3): 677–705.

Day F. 1873. On some new fishes of India. Journal of the Linnean Society of London (Zoology) 11(56): 524–530.

Day F. **1873**. On some new or imperfectly known fishes of India and Burma. *Proceedings of the Zoological Society of London* 1873 (part 1): 107–112.

Day F. **1873**. On some new or little-known fishes of India. *Proceedings of the Zoological Society of London* 1873 (part 3): 704–710.

Day F. **1873**. *Report on the freshwater fish and fisheries of India and Burma*. Calcutta: Office of the Superintendent of Government Printing.

Day F. 1875-1878. The fishes of India; being a natural history of the fishes known to inhabit the seas and fresh water of India, Burma, and Ceylon (Parts 1-4). London: Bernard Quaritch / Williams & Norgate. [Part 1 (1875): 1-168, Pls. 1-40; Part 2 (1876): 169-368, Pls. 41-78; Part 3 (1877): 369-552, Pls. 79-138; Part 4 (1878): xx + 553-778, Pls. 139-195]

Day F. **1881**. *The fishes of Great Britain and Ireland*. Volume 1. Creative Media Partners, LLC (2019 edition). pp. 65–240, Pls. 28–68.

Day F. **1888**. Observations on the fishes of India. Part 1. *Proceedings of the Scientific Meetings of the Zoological Society of London* 1888: 258–265. Day F. **1888**. *The fishes of India; being a natural history of the fishes known to inhabit the seas and fresh waters of India, Burma, and Ceylon* (Supplement). London: Williams & Norgate. pp. 779–816.

Day F. **1889**. Fishes. Vol. 1. In: WT Blanford (ed) *The fauna of British India, including Ceylon and Burma.* London: Taylor & Francis. xviii + 548 pp.

Day F. **1889**. Fishes. Vol. 2. In: Blanford WT (ed) *The fauna of British India, including Ceylon and Burma*. London: Taylor & Francis. xiv + 509 pp.

Day JH. **1969**. *A guide to marine life on South African shores*. Cape Town: AA Balkema. 300 pp.

Day JH. **1974**. *A guide to marine life on South African shores* (2<sup>nd</sup> [revised] edition). Cape Town: AA Balkema. 300 pp.

Day JH. **1974**. The ecology of Morrumbene Estuary, Moçambique. *Transactions of the Royal Society of South Africa* 41(1): 43–97.

Day JH, Field JG, Penrith MJ. **1970**. The benthic fauna and fishes of False Bay, South Africa. *Transactions of the Royal Society of South Africa* 39(1): 1–108.

Day JJ. **2002**. Phylogenetic relationships of the Sparidae (Teleostei: Percoidei) and implications for convergent trophic evolution. *Biological Journal of the Linnean Society* 76(2): 269–301.

Dean B. 1904. Notes on Japanese myxinoids. A new genus *Paramyxine* and a new species *Homea okinoseana*.
Reference also to their eggs. *Journal of the College of Science*, *Imperial University* (Tokyo) 19(art.2): 1–25.

Dean MN, Summers AP. **2006**. Mineralized cartilage in the skeleton of chondrichthyan fishes. *Zoology* 109(2): 164–168. http://dx.doi.org/10.1016/j.zool.2006.03.002

De Beaufort LF. **1912**. On some new Gobiidae from Ceram and Waigen. *Zoologischer Anzeiger* **39**(3): 136–143.

De Beaufort LF. **1913**. Fishes of the eastern part of the Indo-Australian archipelago, with remarks on its zoogeography. *Bijdragen tot de Dierkunde* 19: 95–163.

De Beaufort LF. **1940**. The fishes of the Indo-Australian archipelago. VIII. Percomorphi (continued). Cirrhitoidea, Labriformes, Pomacentriformes. In: Weber M, De Beaufort LF (eds) *Fishes of the Indo-Australian Archipelago*. Vol. 8. Leiden: E.J. Brill. xv + 508 pp.

De Beaufort LF. **1948**. On a new genus of fishes of the family Creediidae from South Africa, with remarks on its geographical distribution. *Transactions of the Royal Society of South Africa* 31(5): 475–478.

De Beaufort LF. **1957**. On a new species of *Platycephalus*. *Proceedings of the National Institute of Sciences of India* 22(B:2): 83–85.

De Beaufort LF. 1962. In: De Beaufort LF, Briggs JC.

De Beaufort LF, Briggs JC. **1962**. Scleroparei (pp. 1–177), Hypostomides, Pediculati, Plectognathi, Opisthomi, Discocephali, Xenopterygii. In: Weber M, De Beaufort LF (eds) *The fishes of the Indo-Australian Archipelago*. Vol. 11. Leiden: E.J. Brill. xi + 481 pp.

De Beaufort LF, Chapman WM. **1951**. IX. Percomorphi (concluded), Blennoidea. In: Weber M, De Beaufort LF (eds) *The fishes of the Indo-Australian archipelago*. Vol. 9. A.J. Reprints Agency. 484 pp.

Debelius H. **1987**. *Underwater guide: Red Sea fishes*. Stuttgart: Verlag Stephanie Naglschmid. 167 pp.

Debelius H. **1993**. Indian Ocean tropical fish guide: Maledives, Sri Lanka, Mauritius, Madagascar, East Africa, Seychelles, Arabian Sea, Red Sea. Frankfurt: IKAN-Unterwasser Archive. 321 pp.

Debelius H. **1993**. *Fischführer Indischer Ozean*. Melle: Tetra Verlag. 321 pp.

Debelius H. **1998**. *Red Sea reef guide: Egypt, Israel, Jordan, Sudan, Saudi Arabia, Yemen, Arabian Peninsula (Oman, UAE, Bahrain)*. Frankfurt: IKAN. 321 pp.

Debelius H. **1999**. *Indian Ocean reef guide: Maldives, Sri Lanka, Thailand, South Africa, Mauritius, Madagascar, East Africa, Seychelles.* Frankfurt: IKAN. 321 pp.

Debelius H. **2007**. *Red Sea reef guide: Egypt, Israel, Jordan, Sudan, Saudi Arabia, Yemen, Arabian Peninsula (Oman, UAE, Bahrain)* (4<sup>th</sup> edn). Frankfurt: IKAN. 321 pp.

Debelius H, Tanaka H, Kuiter RH. **2003**. *Angelfishes: A comprehensive guide to Pomacanthidae*. Chorleywood, UK: TMC Publishing. 208 pp.

De Brito Capello F. **1868**. Descripção de dois peixes novos provenientes dos mares de Portugal. *Jornal de Sciências Mathemáticas, Physicas e Naturaes*, Lisboa 1(4) [for 1867]: 314–317.

De Brito Capello F. **1868**. Description de trois nouveaux poissons des mers du Portugal. *Jornal de Sciências Mathemáticas, Physicas e Naturaes*, Lisboa 1(4): 318–322.

De Brito Capello F. **1871**. Primeira lista dos peixes da ilha da Madeira, Açores e das possessões portuguezas d'Africa, que existem no Museum de Lisboa. *Jornal de Sciências Mathemáticas, Physicas e Naturaes*, Lisboa 3(11): 194–202.

De Bruin GHP, Russell BC, Bogusch A. **1994**. FAO species identification guide for fishery purposes. The marine fishery resources of Sri Lanka. Rome: FAO. 400 pp. [sometimes cited as 1995]

De Buen F. **1930**. Sur une collection de Gobiinae provenant du Maroc. Essai de synopsis des espèces de l'Europe. *Bulletin de la Société des sciences naturelles du Maroc* 10: 120–147.

De Buen F. **1960**. Tiburones, rayas y quimeras en la estacion de Biologia Marina de Montemar, Chile. *Revista de Biologia Marina y Oceanografía* (Valparaiso) 10(1–3): 1–50.

De Carvalho MR. **1996**. Higher-level elasmobranch phylogeny, basal squaleans, and paraphyly (pp. 35–62). In: Stiassny MLJ, Parenti LR, Johnson GD (eds) *Interrelationships of fishes*. San Diego: Academic Press. 496 pp. De Carvalho MR. 1999. A synopsis of the deep-sea genus Benthobatis Alcock, with a redescription of the type species Benthobatis moresbyi Alcock, 1898 (Chondrichthyes, Torpediniformes, Narcinidae) (pp. 231–255). In: Séret B, Sire J-Y (eds) Proceedings of the 5<sup>th</sup> Indo-Pacific Fish Conference, Nouméa, New Caledonia, 3–8 November 1997. Paris: Société Française d'Ichtyologie. 866 pp.

De Carvalho MR. **2016**. Sleeper rays. Family Narkidae (pp. 170–181). In: Last PR, White WT, Carvalho MR de, Séret B, Stehmann MFW, Naylor GJP (eds) *Rays of the world*. CSIRO Publishing & Cornell University Press. 790 pp.

De Carvalho MR, Compagno LJV, Ebert DA. **2003**. *Benthobatis yangi*, a new species of blind electric ray from Taiwan (Chondrichthyes: Torpediniformes: Narcinidae). *Bulletin of Marine Science* 72(3): 923–939.

De Carvalho MR, Compagno LJV, Last PR. 1999. Narcinidae. Numbfishes (pp. 1433–1442). In: Carpenter K, Niem VH (eds) FAO species identification guide for fishery purposes. The living marine resources of the western central Pacific. Vol. 3. Batoid fishes, chimaeras and bony fishes part 1 (pp. 1397–2068). Rome: FAO.

De Carvalho MR, Compagno LJV, Mee JKL. **2002**. *Narcine oculifera*: a new species of electric ray from the gulfs of Oman and Aden (Chondrichthyes: Torpediniformes: Narcinidae). *Copeia* 2002(1): 137–145.

De Carvalho MR, Last PR. **2016**. Numbfishes. Family Narcinidae (pp. 137–169). In: Last PR, White WT, Carvalho MR de, Séret B, Stehmann MFW, Naylor GJP (eds) *Rays of the world*. CSIRO Publishing & Cornell University Press. 790 pp.

De Carvalho MR, Last PR, Séret B. **2016**. Torpedo rays. Family Torpedinidae (pp.185–203). In: Last PR, White WT, Carvalho MR de, Séret B, Stehmann MFW, Naylor GJP (eds) *Rays of the world*. CSIRO Publishing & Cornell University Press. 790 pp.

De Carvalho MR, Maisey JG. 1996. The phylogenetic relationship of the Late Jurassic shark *Protospinax* Woodward, 1919 (Chondrichthyes: Elasmobranchii) (pp. 9–46). In: Arratia G, Viohl G (eds) *Mesozoic Fishes – Systematics and paleoecology. Proceedings of the 1st international meeting on Mesozoic fishes, Eichstätt, 1993.* Munich: Verlag Dr. Friedrich Pfeil. 576 pp.

De Carvalho MR, Maisey JG, Grande L. **2004**. Freshwater stingrays of the Green River Formation of Wyoming (Early Eocene), with the description of a new genus and species and an analysis of its phylogenetic relationships (Chondrichthyes: Myliobatiformes). *Bulletin of the American Museum of Natural History* 284: 1–136.

De Carvalho MR, Randall JE. **2003**. Numbfishes from the Arabian Sea and surrounding gulfs, with the description of a new species from Oman (Chondrichthyes:

Torpediniformes: Narcinidae). *Ichthyological Research* 50(1): 59–66.

De Carvalho MR, Séret B, Compagno LJV. **2002**. A new species of electric ray of the genus *Narcine* Henle, 1834 from the south-western Indian Ocean (Chondrichthyes: Torpediniformes: Narcinidae). *South African Journal of Marine Science* 24(1): 135–149.

De Carvalho MR, Séret B, Schelly RC. **2007**. Pristidae (Vol. 1, pp. 148–153). In: Stiassny MLJ, Teugels GG, Hopkins CD (eds) *The fresh and brackish water fishes of Lower Guinea, West-Central Africa.* [Poissons d'eaux douces et saumâtres de basse Guinée, ouest de l'Afrique centrale.] Vols. 1 & 2. Paris: Muséum National d'Histoire Naturelle. 1456 pp.

De Carvalho MR, Stehmann M, Manilo LG. **2002**. *Torpedo adenensis*, a new species of electric ray from the Gulf of Aden, with comments on nominal species of *Torpedo* from the western Indian Ocean, Arabian Sea, and adjacent areas (Chondrichthyes: Torpediniformes: Torpedinidae). *American Museum Novitates* 3369: 1–34.

De Filippi F. **1853**. Nouvelles espèces de poissons. *Revue et Magasin de Zoologie* (Séries 2) 5: 164–171.

Dekay JE. **1842**. Zoology of New-York; or the New-York fauna; comprising detailed descriptions of all the animals hitherto observed within the State of New-York, with brief notices of those occasionally found near its borders, and accompanied by appropriate illustrations. Part 4. Fishes. Albany: W. & A. White & J. Visscher.

De la Paz R, Aragones N. **1986**. Mangrove fishes of Pagbilao (Quezon Province, Luzon Island), with notes on their abundance and seasonality. *Natural and Applied Science Bulletin* (Manila) 37(2) [for 1985]: 171–190.

De la Pylaie M. **1835**. Recherches en France sur les poissons de l'océan pendant les années 1832–1833. *Congrès Scientifique de France (Seconde Session) tenue à Poitiers Septembre 1834* (art. 5): 524–534.

Delarbre CC, Gallut C, Barriel V, Janvier P, Gachelin G. **2002**. Complete mitochondrial DNA of the hagfish, *Eptatretus burgeri*: the comparative analysis of mitochondrial DNA sequences strongly supports the cyclostome monophyly. *Molecular Phylogenetics and Evolution* 22: 184–192.

Delaroche FE. **1809**. Suite du mémoire sur les espèces de poissons observées à Iviça. Observations sur quelquesuns des poissons indiqués dans le précédent tableau et descriptions des espèces nouvelles ou peu connues. *Annales du Muséum d'Histoire Naturelle* (Paris) 13: 313–361.

Del Cerro L, Lloris D. 1997. Gurnard fishes (Scorpaeniformes, Triglidae) from off New Caledonia, with description of five new species. No. 6. In: Séret B (ed) Résultats des Campagnes MUSORSTOM, Vol. 17. Mémoires du Muséum National d'Histoire Naturelle (Paris) (N.S., Série A, Zoologie) 174: 91–124. Delrieu-Trottin E, Williams JT, Bacchet P, Kulbicki M, Mourier J, Galzin R, Lison de Loma T, Mou-Tham G, Siu G, Planes S. **2015**. Shore fishes of the Marquesas Islands, an updated checklist with new records and new percentage of endemic species. *Check List* 11(5): art. 1758. http://dx.doi. org/10.15560/11.5.1758

Delventhal NR, Mooi RD. **2013**. *Callogobius winterbottomi*, a new species of goby (Teleostei: Gobiidae) from the Western Indian Ocean. *Zootaxa* 3630(1): 155–164. https://doi.org/10.11646/zootaxa.3630.1.6

Delventhal NR, Mooi RD, Bogorodsky SV, Mal AO. **2016**. A review of the *Callogobius* (Teleostei: Gobiidae) from the Red Sea with the description of a new species. *Zootaxa* 4129(2): 225–243. http://dx.doi.org/10.11646/zootaxa.4179.2.3

Demidov VF, Viskrebentsev BF. **1970**. The distribution and some biological features of the main commercial ichthyofauna in the north-west part of the Red Sea. *Trudy Azovo-Cernomorskij Naucno-Issledovateľskij Institut Morskogo Rybnogo Chozjajstva i Okeanografii* 30: 30–113.

 De Miranda Ribeiro A. 1915. Fauna brasiliense. Peixes. Tomo
 V. (Eleutherobranchios aspirophoros) – Physoclisti. Archivos do Museu Nacional do Rio de Janeiro 17: 1–679.

De Moussac G. **1986**. Mise en évidence de l'hermaphrodisme protogyne d'*Epinephelus chlorostigma* (Valenciennes, 1828) aux Seychelles (Pisces, Serranidae). *Cybium* 10(3): 249–262.

De Pinna MCC. **1996**. Teleostean monophyly (pp. 147–162). In: Stiassny MLJ, Parenti LR, Johnson GD (eds) *Interrelationships of fishes*. San Diego: Academic Press. 496 pp.

Deraniyagala PEP. 1929. Ceylon sardines. Ceylon Journal of Science (Section B – Zoology & Geology) 15(1): 31–47.

Deraniyagala PEP. **1933**. Names of some fishes from Ceylon. *Ceylon Journal of Science* (Section C) 5: 79–111.

Deraniyagala PEP. **1936**. Two new fishes from Ceylon. *Ceylon Journal of Science* (Section B) 19(3): 219–224.

Deraniyagala PEP. **1952**. A colored atlas of some vertebrates from Ceylon (Vol. 1). Fishes. Ceylon National Museum Publication. 149 pp.

De Silva PHDH. **1956**. The order Thoracostei off Ceylon, with a list of the specimens in the Colombo Museum. *Spolia Zeylanica* 28: 35–45.

Desjardins J. **1832**. Deux nouvelles espèces de Chétodons (Troisième). In: Desjardins J (ed) (Troisième) *Rapport Annuel sur les Travaux de la Société d'Histoire Naturelle de l'île Maurice* 3: 106–109.

Desjardins J. **1834**. Extrait du Troisième Rapport sur les Travaux de la Société d'Histoire Naturelle de l'Ile Maurice [presented to Zoological Society, Nov. 12, 1833]. *Proceedings* [of the General Meetings for Scientific Business] *of the Zoological Society of London* 1833(1): 117.

Desjardins J. **1836**. Poissons. In: *Rapport Annuel sur les Travaux de la Société d'Histoire Naturelle de l'île Maurice* 7: 1–64. Desjardins J. **1840**. Description d'une nouvelle espèce de l'île Maurice, appartenant à la famille des pectorales pédiculées et au genre *Chironecte. Magasin de Zoologie, d'Anatomie Comparée et de Palæontologie* (Série 2) 2 [for 1839]: 1–4.

Desmarest E. **1874**. Reptiles et poissons. In: Chenu JG Encyclopédie d'histoire naturelle; ou, Traité complet de cette science d'après les travaux des naturalistes les plus éminents de toutes les époques, etc...) (2<sup>nd</sup> edition; 2 volumes). Paris: E. Girard. 360 pp.

Desoutter M. 1990. Monodactylidae (p. 830), Kyphosidae (pp. 831–833), Ephippidae (pp. 834–836), Acanthuridae (pp. 962–964), Soleidae (pp. 1037–1049), Cynoglossidae (pp. 1050–1054). In: Quéro J-C, Hureau J-C, Karrer C, Post A, Saldanha L (eds) *Check-list of the fishes of the eastern tropical Atlantic (CLOFETA)*. Vol. 2. Lisbon: Junta Nacional de Investigação Científica e Tecnológica.

Desoutter M. **1994**. Révision des genres *Microchirus*, *Dicologlossa* et *Vanstraelenia* (Pleuronectiformes, Soleidae). *Cybium* 18(3): 215–249.

Desoutter M, Chapleau F. **1997**. Taxonomic status of *Bathysolea profundicola* and *B. polli* (Soleidae; Pleuronectiformes) with notes on the genus. *Ichthyological Research* 44(4): 399–412.

Desoutter M, Chapleau F, Munroe TA, Chanet B, Beaunier M. 2001. Catalogue critique des types de poissons du Muséum National d'Histoire Naturelle (suite) Ordre des Pleuronectiformes. *Cybium* 25(4): 299–368.

De Sylva DP. **1963**. Systematics and life history of the great barracuda, *Sphyraena barracuda* (Walbaum). *Studies in Tropical Oceanography* 1. Institute of Marine Science, University of Miami Press. viii + 179 pp.

De Sylva DP. **1975**. Barracudas (Pisces: Sphyraenidae) of the Indian Ocean and adjacent seas – a preliminary review of their systematics and ecology. *Journal of the Marine Biological Association of India* 15(1): 74–94.

De Sylva DP. **1981**. Sphyraenidae. Barracudas. In: Fischer W, Bianchi G, Scott WB (eds) *FAO species identification sheets for fishery purposes. Eastern central Atlantic (Fishing Area 34 and part of 47*). Vol. 4. Rome: FAO. [unpaginated]

De Troch M, Mees J, Papadopoulos I, Wakwabi EO. **1996**. Fish communities in a tropical bay (Gazi Bay, Kenya): seagrass beds vs. unvegetated areas. *Netherlands Journal of Zoology* 46(3): 236–252.

Dettaï A, Lecointre G. **2008**. New insights into the organization and evolution of vertebrate IRBP genes and utility of IRBP gene sequences for the phylogenetic study of the Acanthomorpha (Actinopterygii: Teleostei). *Molecular Phylogenetics and Evolution* 48(1): 258–269. http://dx.doi. org/10.1016/j.ympev.2008.04.003

Devanesan DW. **1937**. On the spawning habits and hatching experiments with the eggs of *Hemirhamphus georgii* Cuv. et Val. *Proceedings of the 12<sup>th</sup> International Congress of Zoology* (Lisbon 1935) 3: 2175–2181.

Devanesan DW, Chidambaram K. **1953**. *The common foodfishes of the Madras State*. Madras: Government Press. 79 pp.

DeVantier L, De'Ath G, Klaus R, Al-Moghrabi S, Abdulaziz M, Reinicke GB, Cheung C. **2004**. Reef-building corals and coral communities of the Socotra Archipelago, a zoogeographic 'crossroads' in the Arabian Sea. *Fauna of Arabia* 20: 117–168.

Devaraj M. **1975**. Osteology and relationships of the Spanish mackerels and seerfishes of the tribe Scomberomorini. *Indian Journal of Fisheries* 22(1&2): 1–67.

Devaraj M. **1976**. Discovery of the scombrid *Scomberomorus koreanus* (Kishinouye) in India, with taxonomic discussion on the species. *Japanese Journal of Ichthyology* 23(2): 79–87.

Devaraj M. **1982**. Age and growth of three species of seerfishes *Scomberomorus commerson, S. guttatus* and *S. lineolatus. Indian Journal of Fisheries* 28(1&2): 104–127.

Devaraj M. **1983**. Maturity, spawning and fecundity of the king seer, *Scomberomorus commerson* (Lacepede), in the seas around the Indian Peninsula. *Indian Journal of Fisheries* 30(2): 203–230.

Devaraj M. **1987**. Maturity, spawning and fecundity of the streaked seer, *Scomberomorus lineolatus* (Cuvier & Valenciennes), in the Gulf of Mannar and Palk Bay. *Indian Journal of Fisheries* 33(3) [for 1986]: 293–319.

Devaraj M. **1987**. Maturity, spawning and fecundity of the spotted seer, *Scomberomorus guttatus*, in the Gulf of Mannar and Palk Bay. *Indian Journal of Fisheries* 34(1): 48–77.

Devaraj M. **1987**. The natural cause and effect theory of fish abundance as the basis of human intervention in marine fisheries. In: *Advances in Aquatic Biology and Fisheries* Prof. N.B. Nair Felicitation Volume: 251–256.

Devaraj M. **1999**. Food and feeding habits of the kingseer, *Scomberomorus commerson* (Lacepede), in the seas around the Indian Peninsula. *Journal of the Marine Biological Association of India* 40(1/2): 69–90.

Devaraj M. **1999**. Food and feeding habits of the streaked seer, *Scomberomorus lineolatus* (Cuvier and Valenciennes), in the Gulf of Mannar and Palk Bay. *Journal of the Marine Biological Association of India* 40(1/2): 91–104.

Devaraj M. **1999**. Food and feeding habits of the spotted seer, *Scomberomorus guttatus* (Bloch and Schneider), in the Gulf of Mannar and Palk Bay. *Journal of the Marine Biological Association of India* 40(1/2): 105–124.

Deva-Sundaram MP. **1951**. Systematics of Chilka mullets with a key to their identification. *Journal of the Zoological Society of India* 3(1): 19–25.

Devi CB. **1969**. Occurrence of larvae of *Pseudorhombus elevatus* Ogilby (Heterosomata Pisces) along the south-west coast of India. *Proceedings of the Indian Academy of Sciences* (Section B) 70(4): 178–186. Devi K, Rao DV. **2003**. Poisonous and venomous fishes of Andaman Islands, Bay of Bengal. *Records of the Zoological Survey of India* 211: 1–71.

De Vis CW. **1882**. Descriptions of some new Queensland fishes. *Proceedings of the Linnean Society of New South Wales* 7(3): 367–371.

De Vis CW. **1884**. Fishes from South Sea Islands. *Proceedings of the Linnean Society of New South Wales* 8(4): 445–457.

De Vis CW. **1885**. New Australian fishes in the Queensland museum. *Proceedings of the Linnean Society of New South Wales* 9(2): 389–400.

Dhanze JR, Jayaram KC. 1982. Some biometric studies of certain closely related species of the genus *Arius* (Pisces: Siluriformes: Ariidae). *Proceedings of the Indian Academy of Sciences* (Animal Sciences) 91(1): 79–98.

Díaz de Astarloa JM, Figueroa DE, Lucifora L, Menni RC, Prenski BL, Chiaramonte G. **1999**. New records of the Pacific sleeper shark, *Somniosus pacificus* (Chondrichthyes: Squalidae) from the southwest Atlantic. *Ichthyological Research* 46(3): 303–308.

Díaz de Astarloa JM, Figueroa DE, Reta R. **2003**. First documented occurrence of the starry toadfish *Arothron firmamentum* (Teleostei: Tetraodontidae) in the south-west Atlantic. *Journal of the Marine Biological Association of the UK* 83(4): 879–880.

DiBattista JD, Choat HJ, Gaither MR, Hobbs J-PA, Lozano-Cortés DF, Myers RF, Paulay G, Rocha LA, Toonen RJ, Westneat MW, Berumen ML. **2015**. On the origin of endemic species in the Red Sea. *Journal of Biogeography* 43(1) [2016]: 13–30. https://doi.org/10.1111/jbi.12631

DiBattista JD, Randall JE, Bowen BW. **2012**. Review of the round herrings of the genus *Etrumeus* (Clupeidae: Dussumieriinae) of Africa, with descriptions of two new species. *Cybium* 36(3): 447–460. https://doi.org/10.26028/ cvbium/2012-363-004

DiBattista JD, Roberts MB, Bouwmeester J, Bowen BW, Coker DJ, Lozano-Cortés DF, Choat JH, Gaither MR, Hobbs J-PA, Khalil MT, Kochzius M, Myers RF, Paulay G, Robitzch VSN, Saenz-Agudelo P, Salas E, Sinclair-Taylor TH, Toonen RJ, Westneat MW, Williams ST, Berumen ML. **2015**. A review of contemporary patterns of endemism for shallow water reef fauna in the Red Sea. In: *Journal of Biogeography* 43(3): 423–439. https://doi.org/10.1111/jbi.12649

DiBattista JD, Rocha LA, Hobbs JA, He S, Priest MA, Sinclair-Taylor TH, Bowen BW, Berumen ML. **2015**. When biogeographical provinces collide: hybridization of reef fishes at the crossroads of marine biogeographical provinces in the Arabian Sea. *Journal of Biogeography* 42(9): 1601–1614. https://doi.org/10.1111/jbi.12526

Dicken ML, Booth AJ, Smale MJ, Cliff G. **2007**. Spatial and seasonal distribution patterns of juvenile and adult raggedtooth sharks (*Carcharias taurus*) tagged off the east coast of South Africa. *Marine and Freshwater Research* (Chondrichthyan Special Issue) 58(1): 127–134.

Didier DA. **1995**. Phylogenetic systematics of extant chimaeroid fishes (Holocephali, Chimaeroidei). *American Museum Novitates* 3119: 1–86.

Didier DA. **2004**. Phylogeny and classification of extant Holocephali (pp. 115–135). In: Carrier JC, Musick JA, Heithaus MR (eds) *Biology of sharks and their relatives*. Boca Raton, Florida: CRC Press. 596 pp.

Didier DA, Kemper JM, Ebert DA. **2012**. Phylogeny, biology, and classification of extant holocephalans (pp. 97–122). In: Carrier JC, Musick JA, Heithaus MR (eds) *Biology of sharks and their relatives* (2<sup>nd</sup> edition). Boca Raton, Florida: CRC Press. 672 pp.

Didier DA, Nakaya K. 1999. Redescription of *Rhinochimaera pacifica* (Mitsukuri) and first record of *R. africana* Compagno, Stehmann & Ebert from Japan (Chimaeriformes: Rhinochimaeridae). *Ichthyological Research* 46(2): 139–152.

Didier DA, Stehmann M. **1996**. *Neoharriotta pumila*, a new species of longnose chimaera from the northwestern Indian Ocean (Pisces, Holocephali, Rhinochimaeridae). *Copeia* 1996(4): 955–965.

Dieuzeide R. **1950**. Sur un *Epigonus* nouveau de la Méditerranée (*Epigonus denticulatus* nov. sp.). *Bulletin des travaux publiés par la station d'agriculture et de pêche de Castiglione* (N.S.) 2: 87–105.

Dingerkus G. 1986. Interrelationships of the Orectolobiform sharks (Chondrichthyes: Seachii) (pp. 227–245). In: Uyeno T, Arai R, Taniuchi T, Matsuura K (eds) Indo-Pacific fish biology: Proceedings of the Second International Conference on Indo-Pacific Fishes. Tokyo: Ichthyological Society of Japan. 985 pp.

Dingerkus G, DeFino TC. 1983. A revision of the orectolobiform shark family Hemiscyllidae
(Chondrichthyes, Selachii). Bulletin of the American Museum of Natural History 176(1): 1–94.

Dmitrenko EM. 1974. Arius deya – Arius dayi sp. n.
(Cypriniformes, Ariidae) iz Arabiiskogo Morya. [Arius dayi sp. n. (Cypriniformes, Ariidae) from the Arabian Sea.]
Vestnik Zoologii, 1974(3): 37–41. [In Russian]

Doiuchi R, Nakabo T. **2005**. The *Sphyraena obtusata* group (Perciformes: Sphyraenidae) with a description of a new species from southern Japan. *Ichthyological Research* 52(2): 132–151.

Doiuchi R, Sato T, Nakabo T. **2004**. Phylogenetic relationships of the stromateoid fishes (Perciformes). *Ichthyological Research* 51(3): 202–212.

Dollfus RP, Petit G. **1938**. Les syngnathidae de la Mer Rouge. Liste des espèces avec la description d'une sous-espèce nouvelle. *Bulletin du Muséum d'Histoire Naturelle* (Série 2) 10(5): 496–506. Donaldson TJ. **1986**. Courtship and spawning of the hawkfish *Cirrhitichthys falco* at Miyake-jima, Japan. *Japanese Journal of Ichthyology* 33(3): 329–333.

Donaldson TJ. 1986. Distribution and species richness patterns of Indo-West Pacific Cirrhitidae: support for Woodland's hypothesis (pp. 623–628). In: Uyeno T, Arai R, Taniuchi T, Matsuura K (eds) Indo-Pacific fish biology: Proceedings of the Second International Conference on Indo-Pacific Fishes. Tokyo: The Ichthyological Society of Japan. 985 pp.

Donaldson TJ. **1989**. Facultative monogamy in obligate coraldwelling hawkfishes (Cirrhitidae). *Environmental Biology of Fishes* 26(4): 295–302.

Donaldson TJ. **1990**. Reproductive behavior and social organization of some Pacific hawkfishes (Cirrhitidae). *Japanese Journal of Ichthyology* 36(4): 439–458.

Donaldson TJ. **1995**. Courtship and spawning of nine species of wrasses (Labridae) from the western Pacific. *Japanese Journal of Ichthyology* 42(3/4): 311–319.

Donaldson TJ, Colin PL. **1989**. Pelagic spawning of the hawkfish *Oxycirrhites typus*. *Environmental Biology of Fishes* 24(4): 295–300.

Donovan E. **1802–1808.** The natural history of British fishes, including scientific and general descriptions of the most interesting species, and an extensive selection of accurately finished coloured plates, taken entirely from original drawings, purposely made from the specimens in a recent state, and for the most part whilst living . 5 Volumes. London. 516 pp in total.

Dooley JK. **1978**. Systematics and biology of the tilefishes (Perciformes; Branchiostegidae and Malacanthidae) with descriptions of two new species. *NOAA Technical Report* NMFS Circular 411: v + 78 pp.

Dooley JK. **1984**. Branchiostegidae. Tilefishes. In: Fischer W, Bianchi G (eds) *FAO species identification sheets for fishery purposes. Western Indian Ocean (Fishing Area 51).* Vol. 1. Rome: FAO. [unpaginated]

Dooley JK. **1984**. Malacanthidae. Blanquillos, sand tilefishes. In: Fischer W, Bianchi G (eds) *FAO species identification sheets for fishery purposes. Western Indian Ocean (Fishing Area 51).* Vol. 3. Rome: FAO. [unpaginated]

Dooley JK. **1999**. Branchiostegidae (= Malacanthidae). Tilefishes (also, quakerfish, blanquillos, burrowfishes, amadais, horseheads, and sand tilefishes) (pp. 2630– 2648). In: Carpenter KE, Niem VH (eds) *FAO species identification guide for fishery purposes. The living marine resources of the western central Pacific.* Vol. 4. Bony fishes part 2 (pp. 2069–2790). Rome: FAO.

Dooley JK. 2000. Family Malacanthidae (tilefishes) (p. 615). In: Randall JE, Lim KKP (eds) A checklist of the fishes of the South China Sea. *Raffles Bulletin of Zoology* Supplement 8: 569–667. Dooley JK, Jimenez L. 2008. *Hoplolatilus luteus* Allen & Kuiter, 1989, a junior synonym of *H. fourmanoiri* Smith,1964 (Perciformes: Malacanthidae), based on morphological and molecular data. *aqua, International Journal of Ichthyology* 14(2): 77–84.

Dooley JK, Kailola PJ. **1988**. Four new tilefishes from the northeast Indian Ocean, with a review of the genus *Branchiostegus. Japanese Journal of Ichthyology* 35(3): 247–260.

Dooley JK, Paxton JR. **1975**. A new species of tilefish (family Branchiostegidae) from eastern Australia. *Proceedings of the Linnean Society of New South Wales* 99: 151–156.

Dooley JK, Rau N. **1982**. A remarkable tilefish record and comments on the Philippine tilefishes. *Japanese Journal of Ichthyology* 28(4): 450–452.

Dor M. **1965**. *Zoological lexicon, Vertebrata*. Tel-Aviv: Dvir. 474 pp. [In Hebrew]

Dor M. **1970**. Contributions to the knowledge of the Red Sea No. 44. Nouveaux poissons pour la faune de la Mer Rouge. *Bulletin of the Sea Fisheries Research Station, Haifa* 54: 7–28.

Dor M. **1984**. *Checklist of the Fishes of the Red Sea (CLOFRES)*. Jerusalem: Israel Academy of Sciences and Humanities. 437 pp.

Dor M, Allen GR. **1977**. *Neopomacentrus miryae*, a new species of pomacentrid fish from the Red Sea. *Proceedings of the Biological Society of Washington* 90(1): 183–188.

Dor M, Fraser-Brunner A. **1977**. Record of *Hemipteronotus melanopus* (Teleostei: Labridae) from the Red Sea. *Israel Journal of Zoology* 26: 135–136.

Douady CJ, Delsuc F, Boucher Y, Doolittle WF, Douzery EJP. 2003. Comparison of Bayesian and maximum likelihood bootstrap measures of phylogenetic reliability. *Molecular Biology and Evolution* 20(2): 248–254. http://dx.doi. org/10.1093/molbev/msg042

Douady CJ, Dosay M, Shivji MS, Stanhope MJ. **2003**. Molecular phylogenetic evidence refuting the hypothesis of Batoidea (rays and skates) as derived sharks. *Molecular Phylogenetics and Evolution* 26(2): 215–221. http://dx.doi. org/10.1016/S1055-7903(02)00333-0

Doubilet D, Ghisotti A. **1993**. *The Red Sea.* Israel: Steimatzky Ltd. 160 pp. [also published in 1994 by Smithmark Publishers and Swan Hill Press, Shrewsbury]

Duarte-Bello PP. **1959**. Catalogo de peces Cubanos. Laboratorio de Biologia Marina, Universidad Catolica de Santo Tomas de Villanueva Monografia 6: 1–208.

Dubochkin AS, Kotlyar AN. **1989**. On the feeding of alfoncino (*Beryx splendens*). *Journal of Ichthyology* 29(5): 1–8.

Dudley SFJ, Anderson-Reade MD, Thompson GS, McMullen PB. **2000**. Concurrent scavenging off a whale carcass by great white sharks, *Carcharodon carcharias*, and tiger sharks, *Galeocerdo cuvier. Fishery Bulletin* 98(3): 646–649. Dudley SFJ, Cliff G. **1993**. Sharks caught in the protective gill nets off Natal, South Africa. 7. The blacktip shark *Carcharhinus limbatus* (Valenciennes). *South African Journal of Marine Science* **13**(1): 237–254.

Duhamel G. **1989**. Ichtyofaune des îles Saint-Paul et Amsterdam (Océan Indien Sud). *Mésogée* 9: 21–47.

Duhamel G. **1995**. Révision des genres *Centriscops* et *Notopogon*, Macroramphosidae des zones subtropicale et tempérée de l'hémisphère sud. *Cybium* 19(3): 261–303, Pl. 1–2.

Duhamel G. **1999**. Deep-sea demersal ichthyology off the St-Paul and Amsterdam Islands (central southern India Ocean) (pp. 185–194). In: Séret B, Sire J-Y (eds) *Proceedings of the* 5<sup>th</sup> *Indo-Pacific Fish Conference, Nouméa, New Caledonia,* 3–8 *November* 1997. Paris: Société Française d'Ichtyologie. 866 pp.

Duhamel G, Compagno LJV. **1985**. Sharks (pp. 209–216). In: Fischer W, Hureau J-C (eds) *FAO species identification sheets for fishery purposes. Southern Ocean: Fishing Areas* 48, 58 *and* 88 (CCAMLR Convention Area). Vol. 1. Rome: FAO. 232 pp.

Duhamel G, Hureau J-C. **1982**. Données complémentaires sur l'ichtyofaune des îles australes françaises. *Cybium* (Série 3) 6(1): 65–80.

Dulčić J, Soldo A, Jardas I. **2005**. Review of Croatian selected scientific literature on species mostly exploited by the national small-scale fisheries. Report of the AdriaMed Technical Consultation on Adriatic Sea Small-Scale Fisheries, Split, Croatia, 14<sup>th</sup>–15<sup>th</sup> October 2003. *AdriaMed Technical Documents* 15: 126–179.

Dumbraveanu D, Sheppard CRC. **1999**. Areas of substrate at different depths in the Chagos Archipelago. In: Sheppard CRC, Seaward MRD (eds) *Ecology of the Chagos Archipelago*. *Occasional Publications of the Linnean Society of London* 2: 35–44.

Duméril AHA. 1852. Monographie de la famille des torpédiniens, ou poissons plagiostomes électriques, comprenant la description d'un genre nouveau, de 3 espèces nouvelles, et de 2 espèces nommées dans le Musée de Paris, mais non encore décrites. *Revue et Magasin de Zoologie* (Série 2) 4: 176–189, 227–244, 270–285.

Duméril AHA. **1855**. Note sur un travail inédit de Bibron relatif aux poissons Plectognathes Gymnodontes (Diodons et Tétrodons). *Revue et Magasin de Zoologie* (Série 2) 7: 274–282.

Duméril AHA. **1865**. *Histoire naturelle des poissons ou ichthyologie générale (Vol. I, 2 parts). Elasmobranches: Plagiostomes et Holocéphales ou Chimères.* Paris: Librarie Encylopedique de Roret. 720 pp.

Duméril AHA. 1870. Histoire naturelle des poissons, ou ichthyologie générale (Vol. II). Ganoïdes, Dipnés, Lophobranches. Paris: Librairie Encyclopédique de Roret. 624 pp. Duméril AMC. **1806**. *Zoologie analytique, ou méthode naturelle de classification des animaux*. Paris: Libraire Allais. xxxiii + 344 pp. [also seen as 1805]

Duncker G. **1910**. On some syngnathids ("pipe fish") from Ceylon. *Spolia Zeylanica* 7(25): 25–34.

Duncker G. **1912**. Die Gattungen der Syngnathidae. *Mitteilungen aus dem Naturhistorischen (Zoologischen) Museum in Hamburg* 29: 219–240.

Duncker G. 1912. Die Süßwasserfische Ceylons. Mitteilungen aus dem Naturhistorischen (Zoologischen) Museum in Hamburg 29: 241–272.

Duncker G. **1914**. Description of a new species of *Hippocampus. Records of the Indian Museum* (Calcutta) 10(pt 5, art.18): 295.

Duncker G. **1915**. Revision der Syngnathidae. Erster Teil. *Mitteilungen aus dem Naturhistorischen (Zoologischen) Museum in Hamburg* 32: 9–120.

Duncker G. **1925**. Description of a new species of *Hippocampus* from the Andamans. *Records of the Indian Museum* (Calcutta) 27(6): 475–476.

Duncker G. 1940. Ueber einige Syngnathidae aus dem Roten Meer. Publications of the Marine Biological Station Al-Ghardaga (Red Sea) 3: 83–88.

Duncker G, Mohr E. 1929. Die Fische der Südsee-Expedition der Hamburgischen Wissenschaftlichen Stiftung 1908– 1909. 3. Teil. Acanthopteri sens. ampl., Physoclisti Malacopterygii, Physostomi, Plagiostomi. *Mitteilungen aus dem Zoologischen Staatsinstitut und Zoologischen Museum in Hamburg* 44: 57–84.

Duncker G, Mohr EW. **1935**. Die nordeuropäischen Ammodytes-Arten des Hamburger Zoologischen Museums. Zoologischer Anzeiger 110(7/8): 216–220.

Duncker G, Mohr E. 1939. Revision der Ammodytidae. Mitteilungen aus dem Zoologischen Museum in Berlin 24(1): 8–31.

Durand J-D, Borsa P. 2015. Mitochondrial phylogeny of grey mullets (Acanthopterygii: Mugilidae) suggests high proportion of cryptic species. *Comptes Rendus Biologies* 338(4): 266–277. https://doi.org/10.1016/j.crvi.2015.01.007

Durand J-D, Chen W-J, Shen K-N, Fu C, Borsa P. **2012**. Genuslevel taxonomic changes implied by the mitochondrial phylogeny of grey mullets (Teleostei: Mugilidae). *Comptes Rendus Biologies* 335(10–11): 687–697. https://doi.org/10.1016/j.crvi.2012.09.005

Durville P, Chabanet P, Quod JP. **2003**. Visual census of the reef fishes in the natural reserve of the Glorieuses Islands (Western Indian Ocean). *Western Indian Ocean Journal of Marine Science* 2(2): 95–104.

Durville P, Mulochau T, Diringer A. 2011. Poissons profonds de l'océan Indien. Réunion : ORPHIE. 208 pp.

Dutt S, Radhakrishna Rao MPR. **1980**. *Apogon andhrae*, a new species of apogonid fish from the Bay of Bengal. *Journal of Natural History* 14(5): 743–748.

Dutt S, Rao KH. **1965**. A new bothid flatfish *Cephalopsetta ventrocellatus* gen. et sp. nov. from Bay of Bengal. *Proceedings of the Indian Academy of Sciences* (Section B) 62(4): 180–187.

Dutt S, Sagar JV. 1981. Saurida pseudotumbil – a new species of lizardfish (Teleostei: Synodidae) from Indian coastal waters. Proceedings of the Indian National Science Academy (Part B Biological Sciences) 47(6): 845–851.

Dutt S, Thankam V. **1967**. Two new species of trichiurid fish from Waltair. *Journal of the Bombay Natural History Society* 63(3, art.8) [for 1966]: 755–758.

Dyer BS, Chernoff B. **1996**. Phylogenetic relationships among atheriniform fishes (Teleostei: Atherinomorpha). *Zoological Journal of the Linnean Society* 117: 1–69.

## Ε

Ebert DA. **1990**. The taxonomy, biogeography and biology of cow and frilled sharks (Chondrichthyes: Hexanchiformes). PhD thesis, Rhodes University, Grahamstown. 308 pp.

Ebert DA. **1991**. Observations on the predatory behaviour of the sevengill shark *Notorynchus cepedianus*. *South African Journal of Marine Science* 11(1): 455–465.

Ebert DA. **1991**. Diet of the sevengill shark *Notorynchus cepedianus* in the temperate coastal waters of southern Africa. *South African Journal of Marine Science* 11(1): 565–572.

Ebert DA. **1994**. Diet of the sixgill shark *Hexanchus griseus* off southern Africa. *South African Journal of Marine Science* 14(1): 213–218.

Ebert DA. **1996**. Biology of the sevengill shark, *Notorynchus cepedianus*, in the temperate coastal waters of southern Africa. *South African Journal of Marine Science* 17(1): 93–103.

Ebert DA. **2002**. Ontogenetic changes in the diet of the sevengill shark (*Notorynchus cepedianus*). *Marine and Freshwater Research* 53(2): 517–523.

Ebert DA. **2002**. Some observations on the reproductive biology of the sixgill shark *Hexanchus griseus* (Bonnaterre, 1788) from southern African waters. *South African Journal of Marine Science* 24(1): 359–363.

Ebert DA. **2003**. *Sharks, rays, and chimaeras of California*. Berkeley, California: University of California Press. xiii + 284 pp.

Ebert DA. **2013**. Deep-sea sharks of the Indian Ocean. Vol. 1. Sharks. *FAO Species Catalogue for Fishery Purposes* No. 8 (Vol. 1). Rome: FAO. 268 pp. Ebert DA. **2014**. Deep-sea cartilaginous fishes of the Indian Ocean. Vol. 2. Batoids and Chimaeras. *FAO Species Catalogue for Fishery Purposes* No. 8 (Vol. 2). Rome: FAO. 129 pp.

Ebert DA, Cailliet GM. **2011**. *Pristiophorus nancyae*, a new species of sawshark (Chondrichthyes: Pristiophoridae) from Southern Africa. *Bulletin of Marine Science* 87(3): 501–512. http://dx.doi.org/10.5343/bms.2010.1108

Ebert DA, Clerkin PJ. **2015**. A new species of deep-sea catshark (Scyliorhinidae: *Bythaelurus*) from the southwestern Indian Ocean. *Journal of the Ocean Science Foundation* 15: 53–63. http://dx.doi.org/10.5281/zenodo.1001009

Ebert DA, Compagno LJV. **2009**. *Chlamydoselachus africana*, a new species of frilled shark from southern Africa (Chondrichthyes, Hexanchiformes, Chlamydoselachidae). *Zootaxa* 2173: 1–18. http://dx.doi.org/10.11646/ zootaxa.2173.1.1

Ebert DA, Compagno LJV, Cowley PD. **1992**. A preliminary investigation of the feeding ecology of squaloid sharks off the west coast of southern Africa. *South African Journal of Marine Science* 12(1): 601–609.

Ebert DA, Compagno LJV, Cowley PD. **2006**. Reproductive biology of catsharks (Chondrichthyes: Scyliorhinidae) off the west coast of southern Africa. *ICES Journal of Marine Science* 63(6): 1053–1065.

Ebert DA, Compagno LJV, Cowley PD. **2008**. Aspects of the reproductive biology of skates (Chondrichthyes: Rajiformes: Rajoidei) from southern Africa. *ICES Journal of Marine Science* 65(1): 81–102. http://dx.doi.org/10.1093/icesjms/ fsm169

Ebert DA, Cowley PD. **2003**. Diet, feeding behaviour and habitat utilisation of the blue stingray *Dasyatis chrysonota* (Smith, 1828) in South African waters. *Marine and Freshwater Research* 54(8): 957–965.

Ebert DA, Cowley PD. **2009**. Reproduction and development of the blue stingray, *Dasyatis chrysonota*, in southern African waters. *Journal of the Marine Biological Association of the UK* 89(4): 809–815. http://dx.doi.org/10.1017/ S0025315408002907

Ebert DA, Cowley PD, Compagno LJV. **1991**. A preliminary investigation of the feeding ecology of skates (Batoidea: Rajidae) off the west coast of southern Africa. *South African Journal of Marine Science* 10(1): 71–81.

Ebert DA, Cowley PD, Compagno LJV. **1996**. A preliminary investigation of the feeding ecology of catsharks (Scyliorhinidae) off the west coast of southern Africa. *South African Journal of Marine Science* 17(1): 233–240.

Ebert DA, Cowley PD, Compagno LJV. **2002**. First records of the longnose spiny dogfish *Squalus blainvillei* (Squalidae) and the deep-water stingray *Plesiobatis daviesi* (Urolophidae) from South African waters. *South African Journal of Marine Science* 24(1): 355–357. Ebert DA, Dando M. **2021**. *Field guide to sharks, rays & chimaeras of Europe and the Mediterranean*. Princeton, New Jersey: Princeton University Press. 383 pp.

Ebert DA, Dando M, Fowler S. **2021**. *Sharks of the world* (2<sup>nd</sup> edition). Princeton, New Jersey: Princeton University Press. 607 pp.

Ebert DA, Fowler S, Compagno LJV. **2013**. *Sharks of the world. A fully illustrated guide*. Plymouth, England: Wild Nature Press. 528 pp.

Ebert DA, Gon O. **2017**. *Rhinobatos austini* n. sp., a new species of guitarfish (Rhinopristiformes: Rhinobatidae) from the southwestern Indian Ocean. *Zootaxa* 4276(2): 204–214. http://dx.doi.org/10.11646/zootaxa.4276.2.3

Ebert DA, Haas DL, Carvalho MR de. **2015**. *Tetronarce cowleyi*, sp. nov., a new species of electric ray from southern Africa (Chondrichthyes: Torpediniformes: Torpedinidae). *Zootaxa* 3936(2): 237–250. http://dx.doi.org/10.11646/ zootaxa.3936.2.4

Ebert DA, Stehmann MFW. **2013**. Sharks, batoids, and chimaeras of the North Atlantic. *FAO Species Catalogue for Fishery Purposes* No. 7. Rome: FAO. 523 pp.

Ebert DA, White WT, Goldman KJ, Compagno LJV, Daly-Engel TS, Ward RD. **2010**. Resurrection and redescription of *Squalus suckleyi* (Girard, 1854) from the North Pacific, with comments on the *Squalus acanthias* subgroup (Squaliformes: Squalidae). *Zootaxa* 2612: 22–40. http://dx.doi.org/10.11646/zootaxa.2612.1.2

Ebert DA, Wintner SP, Kyne PM. **2021**. An annotated checklist of the chondrichthyans of South Africa. *Zootaxa* 4947(1): 1–127. https://doi.org/10.11646/zootaxa.4947.1.1

Ebert DA, Winton MV. **2010**. Chondrichthyans of high latitude seas (pp. 115–158). In: Carrier JC, Musick JA, Heithaus MR (eds) *Sharks and their relatives II: Biodiversity, adaptive physiology, and conservation*. Boca Raton, Florida: CRC Press. 746 pp.

Eccles DH. **1992**. *FAO species identification sheets for fishery purposes. Field guide to the freshwater fishes of Tanzania.* Rome: FAO. 145 pp.

Edwards AJ. **1987**. Climate and oceanography (pp. 45–69). In: Edwards AJ, Head SM (eds) *Red Sea*. Key Environments Series. Oxford: Pergamon Press in collaboration with IUCN. 441 pp.

Edwards AJ, Randall JE. **1983**. A new dottyback of the genus *Pseudoplesiops* (Teleostei: Perciformes: Pseudochromidae) from the Red Sea. *Revue française d'Aquariologie Herpétologie* 9(4): 111–114.

Edwards AJ, Shepherd AD. **1992**. Environmental implications of aquarium-fish collection in the Maldives, with proposals for regulation. *Environmental Conservation* 19(1): 61–72.

Edwards G. **1743–1751**. A natural history of uncommon birds, and of some other rare and undescribed animals, quadrupedes, fishes, reptiles, insects, &c. (in 4 parts). London: College of Physicians. 249 pp.

Edwards G. 1771. Appendix. In: Catesby M (ed) *The natural history of Carolina, Florida and the Bahama Islands; containing the figures of birds, beasts, fishes, serpents, insects, and plants; particularly those not hitherto described, or incorrectly figured by authors, with their descriptions in English and French* (3<sup>rd</sup> edition). 2 vols. London: Benjamin White.

Edwards MS. **1971**. Fish and fishing (pp. 24–32). In: The Swartkops Estuary: an ecological survey. The Zwartkops Trust, Port Elizabeth. 42 pp.

Ege V. **1939**. A revision of the genus *Anguilla* Shaw. A systematic, phylogenetic and geographical study. *Dana Report* 16: 1–257.

Eggert B. **1935**. Beitrag zur Systematik, Biologie und geographischen Verbreitung der Periophthalminae. Ergebnisse einer durch die Notgemeinschaft der Deutschen Wissenschaft ermöglichten Reise nach Niederländisch-Indien ... der Deutschen Wissenschaft 1929–1930. Zoologische Jahrbücher Abteilung für Systematik, Geographie und Biologie der Tiere (Jena) 67: 29–116.

Ehrenberg CG. 1829. De percoïdes à dorsale unique à sept rayons branchiaux et à dents en velours ou en cardes. In: Cuvier G, Valenciennes A *Histoire naturelle des poissons*.
Vol. 3. Suite du livre troisième. Des percoïdes àdorsale unique àsept rayons branchiaux et àdents en velours ou en cardes. Paris: F.G. Levrault. xxvii + 500 pp.

Ehrenberg CG, Hemprich FG. **1899**. Symbolae physicae, seu icones adhuc ineditae corporum naturalium novorum aut minus cognitorum quae ex itineribus per Libyam, Aegyptiam, Nubiam, Dongolam, Syriam, Arabiam et Habessiniam publico institutis sumptu ... Studio annis MDCCCXX-MDCCCXXV redierunt. Berlin: Pars Zoologica.

Eibl-Eibesfeldt I. 1964. Land of a thousand atolls: a study of marine life in the Maldive and Nicobar islands. London: MacGibbon & Kee. 195 pp.

Eichler D, Lieske E. **1994**. *Korallenfische Indischer Ozean*. Hamburg: Jahr Verlag. 368 pp.

Eichler D, Myers RF. **1997**. *Korallenfische zentraler Indopazifik*. Hamburg: Jahr-Verlag. 489 pp.

Eigenmann CH, Beeson CH. **1893**. Preliminary note on the relationship of the species usually united under the generic name *Sebastodes*. *American Naturalist* 27(319): 668–671.

Engelhardt R. **1912**. Über einige neue Selachier-Formen. *Zoologischer Anzeiger* 39(21/22): 643–648.

Erdmann M. 2008. Latimeria menadoensis. The IUCN Red List of Threatened Species 2008. e.T135484A4129545. https://doi. org/10.2305/IUCN.UK.2008.RLTS.T135484A4129545.en Eryilmaz L, Dalyan C. **2006**. First record of *Apogon queketti* Gilchrist (Osteichthyes: Apogonidae) in the Mediterranean Sea. *Journal of Fish Biology* 69(4): 1251–1254.

Eschmeyer WN. **1965**. Three new scorpionfishes of the genera *Pontinus, Phenacoscorpius* and *Idiastion* from the western Atlantic Ocean. *Bulletin of Marine Science* 15(3): 521–534.

Eschmeyer WN. **1969**. A new scorpionfish of the genus *Scorpaenodes* and *S. muciparus* (Alcock) from the Indian Ocean, with comments on the limits of the genus. *Occasional Papers of the California Academy of Sciences* 76: 1–11.

Eschmeyer WN. **1969**. A systematic review of the scorpionfishes of the Atlantic Ocean (Pisces: Scorpaenidae). *Occasional Papers of the California Academy of Sciences* 79: 1–143.

Eschmeyer WN. 1981. Scorpaenidae. Scorpionfishes, rockfishes, rosefishes. In: Fischer W, Bianchi G, Scott WB (eds) FAO species identification sheets for fishery purposes. Eastern central Atlantic (Fishing Area 34 and part of 47). Vol. 4. Rome: FAO. [unpaginated]

Eschmeyer WN. **1983**. A new species of the fish genus *Pontinus* (Scorpaeniformes: Scorpaenidae) from off Natal, South Africa. *J.L.B. Smith Institute of Ichthyology Special Publication* 28: 1–4.

Eschmeyer WN. **1986**. Family No. 149: Scorpaenidae. In: Smith MM, Heemstra PC (eds) *Smiths' sea fishes*. Johannesburg: Macmillan South Africa. 1047 pp.

Eschmeyer WN. **1990**. *Catalog of the genera of recent fishes*. San Francisco: California Academy of Sciences. 697 pp.

Eschmeyer WN. **1997**. A new species Dactylopteridae (Pisces) from the Philippines and Australia, with a brief synopsis of the family. *Bulletin of Marine Science* 60(3): 727–738.

Eschmeyer WN (ed). **1998**. *Catalog of fishes* (3 volumes). San Francisco: California Academy of Sciences. 2905 pp.

Eschmeyer WN (ed). **2010**. *Catalog of fishes* [electronic version accessed 12 July 2010]. San Francisco: California Academy of Sciences. http://research.calacademy.org/ichthyology/ catalog/fishcatmain.asp

Eschmeyer WN, Collette BB. **1966**. The scorpionfish subfamily Setarchinae, including the genus *Ectreposebastes*. *Bulletin of Marine Science* 16(2): 349–375.

Eschmeyer WN, Dempster LJ. **1990**. Scorpaenidae (pp. 665– 679). In: Quéro J-C, Hureau J-C, Karrer K, Post A, Saldanha L (eds) *Check-list of the fishes of the eastern tropical Atlantic (CLOFETA)*. Vol. 2. Lisbon: Junta Nacional de Investigação Científica e Tecnológica.

Eschmeyer WN, Dor M. **1978**. *Cocotropus steinitzi*, a new species of the fish family Aploactinidae (Pisces: Scorpaeniformes) from the Red Sea and Andaman Islands. *Israel Journal of Zoology* 27(4): 165–168.

Eschmeyer WN, Fricke R, Fong JD, Polack DA. **2010**. Marine fish diversity: history of knowledge and discovery (Pisces). *Zootaxa* 2525: 19–50. http://dx.doi.org/10.11646/ zootaxa.2525.1.2

Eschmeyer WN, Fricke R, van der Laan R (eds). **2018**. *Catalog* of Fishes [electronic version accessed 2 August 2018]. San Francisco: California Academy of Sciences. http://research. calacademy.org/ichthyology/catalog/fishcatmain.asp

Eschmeyer WN, Hallacher LE, Ramo-Rao KV. **1979**. The scorpionfish genus *Minous* (Scorpaenidae, Minoinae) including a new species from the Indian Ocean. *Proceedings of the California Academy of Sciences* (Series 4) 41(20): 453–473.

Eschmeyer WN, Hirosaki Y, Abe T. **1973**. Two new species of the scorpionfish genus *Rhinopias*, with comments on related genera and species. *Proceedings of the California Academy of Sciences* (Series 4) 39(16): 285–310.

Eschmeyer WN, Hureau J-C. **1971**. *Sebastes mouchezi*, a senior synonym of *Helicolenus tristanensis*, with comments on *Sebastes capensis* and zoogeographical considerations. *Copeia* 1971(3): 576–579.

Eschmeyer WN, Rama-Rao KV. **1972**. Two new scorpionfishes (Genus *Scorpaenodes*) from the Indo-West Pacific, with comments on *Scorpaenodes muciparus* (Alcock). *Proceedings of the California Academy of Sciences* (Series 4) 39(5): 55–64.

Eschmeyer WN, Rama-Rao KV. **1973**. Two new stonefishes (Pisces, Scorpaenidae) from the Indo-West Pacific, with a synopsis of the subfamily Synanceiinae. *Proceedings of the California Academy of Sciences* (Series 4) 39(18): 337–382.

Eschmeyer WN, Rama-Rao KV. **1978**. A new scorpionfish, *Ebosia falcata* (Scorpaenidae, Pteroinae) from the western Indian Ocean, with comments on the genus. *Matsya* 3 [for 1977]: 64–71.

Eschmeyer WN, Rama-Rao KV, Hallacher LE. **1979**. Fishes of the scorpionfish subfamily Choridactylinae from the western Pacific and the Indian Ocean. *Proceedings of the California Academy of Sciences* (Series 4) 41(21): 475–500.

Eschmeyer WN, Randall JE. **1975**. The scorpaenid fishes of the Hawaiian Islands, including new species and new records (Pisces: Scorpaenidae). *Proceedings of the California Academy of Sciences* (Series 4) 41(11): 265–333.

Esmark L. **1862**. Beskrivelse over en ny Fiskeart, *Brama Raschii* Esm. *Forhandlinger i Videnskabs-selskabet i Christiania* [for 1861]: 238–247.

Esseen M, Al-Saqaf H. 2002. Review of the Fisheries Management Plan for the Socotra Island Group (pp. 437– 479). In: Apel M, Hariri K, Krupp F (eds) Conservation and sustainable use of biodiversity of Socotra Archipelago. Marine habitat, biodiversity and fisheries surveys and management. Final Report of Phase III. Senckenberg Research Institute, Frankfurt a.M., Germany. Esseen M, Khanbash M. 1999. The rock lobster fishery of Socotra – current status and managament options (pp. 181–190). In: Krupp F, Hariri K (eds) *Conservation and sustainable use of biodiversity of Socotra Archipelago. Marine habitat, biodiversity and fisheries surveys and management.* Report of Phase I. Senckenberg Research Institute, Frankfurt a.M., Germany.

Euphrasen BA. **1788**. Beskrifning på 3:ne fiskar. *Kongliga Vetenskaps Akademiens nya Handlingar* (Stockholm) 9: 51–55.

Euphrasen BA. **1790**. *Raja* (Narinari). *Kongliga Vetenskaps Akademiens nya Handlingar* (Stockholm) 11: 217–219.

Euphrasen BA. **1791**. *Scomber* (atun) och *Echeneis* (tropica.). *Kongliga Vetenskaps Akademiens nya Handlingar* (Stockholm) 12: 315–318.

Evenhuis NL. **2003**. Publication and dating of the journals forming the *Annals and Magazine of Natural History* and the *Journal of Natural History*. *Zootaxa* 385: 1–68 + Errata.

Evenhuis NL. **2012**. François-Louis Comte de Castelnau (1802–1880) and the mysterious disappearance of his original insect collection. *Zootaxa* 3168: 53–63. http://dx.doi.org/10.11646/zootaxa.3168.1.4

Everett BI, Cliff G, Dudley SFJ, Wintner SP, van der Elst RP. 2015. Do sawfish *Pristis* spp. represent South Africa's first local extirpation of marine elasmobranchs in the modern era? *African Journal of Marine Science* 37(2): 275–284. https://doi.org/10.2989/1814232X.2015.1027269

Everhart MJ. **2007**. Remains of a pycnodont fish (Actinopterygii: Pycnodontiformes) in a coprolite; an uppermost record of *Micropycnodon kansasensis* in the Smoky Hill Chalk, western Kansas. *Transactions of the Kansas Academy of Science* 110(1/2): 35–43.

Evermann BW, Seale A. **1907**. Fishes of the Philippine Islands. *Bulletin of the Bureau of Fisheries* 26(Doc 607): 49–110.

Eydoux JFT, Souleyet FA. **1850**. Poissons (pp. 155–216). In: *Voyage autour du monde exécuté pendant les années 1836 et 1837 sur la corvette* La Bonite, *commandée par M. Vaillant. Zoologie*. Vol. 1. Part 2. Paris. 334 pp.

## F

Fabricius JC. 1775. In: Niebuhr C. 1775. Descriptiones animalium avium, amphibiorum, piscium, insectorum, vermium; quae in itinere orientali observavit Petrus Forskål. Post mortem auctoris edidit Carsten Niebuhr. Hauniae. 20 + xxxiv + 164 pp.

Fang P-W. **1942**. Poissons de Chine de M. Ho: description de cinq espèces et deux sous-espèces nouvelles. *Bulletin de la Société Zoologique de France* 67: 79–85.

FAO. **2014**. *The state of world fisheries and aquaculture*. Rome: FAO. 243 pp.

Faria VV, McDavitt MT, Charvet P, Wiley TR, Simpfendorfer CA, Naylor GJP. 2013. Species delineation and global population structure of critically endangered sawfishes (Pristidae). *Zoological Journal of the Linnean Society* 167(1): 136–164. http://dx.doi.org/10.1111/j.1096-3642.2012.00872.x

Farquhar GB. **1963**. Sharks of the Family Lamnidae. *Technical Report TR-157*. US Naval Oceanographic Office, Stennis Space Center, Mississippi. 22 pp.

Farquharson FL. 1970. A new freshwater goby (Pisces: Gobiidae) from Lake Sibayi, Zululand, South Africa. Annals of the Cape Provincial Museums (Natural History) 8(10): 85–87.

Fautin DG, Allen GR. **1992**. *Field guide to anemonefishes and their host sea anemones*. Perth: Western Australian Museum. 160 pp.

Fedorov VV, Foroshchuk VP. 1988. A new flounder, *Psettina multisquamea* sp. nova, (Bothidae) from Saya-de-Mal'ya Bank, Indian Ocean. *Voprosy Ikhtiologii* 28(4): 531–540. [In Russian. English translation in *Journal of Ichthyology* 28(6), 1989: 36–45]

Fedoryako BI. 1976. Materials on the systematics and distribution of the "oceanic Cheilodipteridae". *Trudy Instituta Okeanologii Imeni P.P. Shirshova* 104: 156–190. [In Russian]

Fedoryako BI. **1979**. Triggerfishes of the genera *Canthidermis* and *Xanthichthys* (Balistidae, Tetraodontiformes) from the Indian and Pacific Oceans. *Voprosy Ikhtiologii* 19(6, art.119): 983–995.

Felis T, Lohmann G, Kuhnert H, Lorenz SJ, Scholz D, Pätzold J, Al-Rousan SA, Al-Moghrabi SM. **2004**. Increased seasonality in Middle East temperatures during the last interglacial period. *Nature* 429: 164–168.

Feltes RM. **1991**. Revision of the polynemid fish genus *Filimanus*, with the description of two new species. *Copeia* 1991(2): 302–322.

Fennessy ST. **2000**. Comparative life histories and stock assessments of rockcods (Family Serranidae) from the east coast of South Africa. PhD thesis, University of Natal, Durban. 165 pp.

Fennessy ST. **2004**. Reproductive biology and growth of the yellowbelly rockcod *Epinephelus marginatus* (Serranidae) from South-East Africa. *African Journal of Marine Science* 28(1): 1–11.

Fergusson IK, Graham KJ, Compagno LJV. 2008. Distribution, abundance and biology of the smalltooth sandtiger shark *Odontaspis ferox* (Risso, 1810) (Lamniformes: Odontaspididae). *Environmental Biology of Fishes* 81(2): 207–228. http://dx.doi.org/10.1007/s10641-007-9193-x

Fermor X. 1913. Wissenschaftliche Ergebnisse einer Reise von S. Aweninzew in die Tropen Africas 1. Einige Befunde zur Kenntnis von Ariodes polystaphylodon. Zoologischer Anzeiger 42: 196–199. Fernandes CA, Rohling EJ, Siddall M. **2006**. Absence of post-Miocene Red Sea land bridges: biogeographic implications. *Journal of Biogeography* 33: 961–966.

Fernandez-Silva I, Randall JE, Golani D, Bogorodsky SV. 2016. Mulloidichthys flavolineatus flavicaudus Fernández-Silva & Randall (Perciformes, Mullidae), a new subspecies of goatfish from the Red Sea and Arabian Sea. Zookeys 605: 131–157. http://dx.doi.org/10.3897/zookeys.605.8060

Fernando EFW. **1968**. Species composition of fish captured by trawlers in the Wadge Bank. *Proceedings of the Indo-Pacific Fisheries Council* 13(111): 521–531.

Fernholm B. **1974**. Diurnal variations in the behaviour of the hagfish *Eptatretus burgeri*. *Marine Biology* 27(4): 351–356.

Fernholm B. 1981. A new species of hagfish of the genus Myxine, with notes on other eastern Atlantic myxinids. Journal of Fish Biology 19(1): 73–82.

Fernholm B, Quattrini AM. **2008**. A new species of hagfish (Myxinidae: *Eptatretus*) associated with deep-sea coral habitat in the western North Atlantic. *Copeia* 2008(1): 126–132.

Fernholm B, Wheeler A. **1983**. Linnean fish specimens in the Swedish Museum of Natural History, Stockholm. *Zoological Journal of the Linnean Society* 78(3): 199–286.

Ferraris CJ Jr. **2007**. Checklist of catfishes, recent and fossil (Osteichthyes: Siluriformes), and catalogue of siluriform primary types. *Zootaxa* 1418: 1–628.

Ferraris CJ Jr, McGrouther MA, Parkinson KL. **2000**. A critical review of the types and putative types of southern Asian marine and freshwater fish species in the Australian Museum named by Francis Day. *Records of the Australian Museum* 52: 289–306.

Fiedler K. **1967**. Das Fortpflanzungsverhalten von *Syngnathus djarong*, einer Süsswassernadel aus Ceylon (Syngnathidae: Teleostei). *Natur und Museum* 97(7): 259–269.

Field RF. **2005**. *Reef fishes: UAE and Gulf of Oman*. Dubai, UAE: Motivate Publishing. 143 pp.

Field R[F]. **2013**. *Reef fishes of Oman*. Published by Richard & Mary Field. 97 pp.

Field R[F], Field M. **1998**. *Reef fishes of the Red Sea. A guide to identification*. London: Kegan Paul International. 192 pp.

Fierstine HL. **2006**. Fossil history of billfishes (Xiphioidei). *Bulletin of Marine Science* 79(3): 433–453.

Filleul A, Lavoué S. 2001. Basal teleosts and the question of elopomorph monophyly. Morphological and molecular approaches. *Comptes Rendus de l'Académie des Sciences* (Series 3 – Sciences de la Vie) 324(4): 393–399.

Fink SV, Fink WL. **1981**. Interrelationships of the ostariophysan fishes (Teleostei). *Zoological Journal of the Linnean Society* 72(4): 297–353.

Finn D. **1983**. Land use and abuse in the East African region. *Ambio* 12: 296–301.

Fischer G. 1813. Zoognosia, tabulis synopticis illustrata, in usum praelectionum Academiae Imperialis Medico-Chirurgicae Mosquensis. Vol. 1 (3<sup>rd</sup> edition). Moscow: Vsevolosky. i–xii + 1–466, Pls. 8. [author sometimes cited as Fischer von Waldheim]

Fischer J, Schott F, Stramma L. **1996**. Currents and transports of the Great Whirl-Socotra Gyre system during the summer monsoon, August 1993. *Journal of Geophysical Research* 101(C2): 3573–3587.

Fischer JG. **1885**. II. Über einige afrikanische Fische des Naturhistorischen Museums in Hamburg (pp. 66–77). In: *Ichthyologische und herpetologische Bemerkungen. Jahrbuch der Hamburgischen Wissenschaftlichen Anstalten* 2: 49–121.

Fischer W, Bianchi G (eds). 1984. FAO species identification sheets for fishery purposes. Western Indian Ocean (Fishing Area 51) (6 volumes). Rome: FAO. [unpaginated] [sometimes cited as 1983]

Fischer W, Sousa I, Silva C, de Freitas A, Poutiers JM,
Schneider W, Borges TC, Féral JP, Massinga A. 1990. Fichas
FAO de identificaçío de espécies para actividades de pesca.
Guía de campo das espécies comerciais marinhas e De águas
salobras de Moçambique. NORAD, Rome: FAO. 424 pp.
[In Portugese]

Fischer W, Whitehead PJP (eds). 1974. FAO species identification sheets for fishery purposes. Eastern Indian Ocean (Fishing Area 57) and western central Pacific (Fishing Area 71). Bony fishes (Vols. 1–4). Rome: FAO. [unpaginated]

Fishelson L. 1966. Solenostomus cyanopterus Blecker [sic] (Teleostei, Solenostomidae) in Elat (Gulf of Akaba). Israel Journal of Zoology 15(3–4): 95–103.

Fishelson L. 1975. Ethology and reproduction of pteroid fishes found in the Gulf of Aqaba (Red Sea) especially *Dendrochirus brachypterus* (Cuvier) (Pteroidae Teleostei). *Publicatione Stazione Zoologica Napoli* 39(Supplement): 635–656.

Fishelson L. **1977**. Ultrastructure of the epithelium from the ovary wall of *Dendrochirus brachypterus* (Pteroidae, Teleostei). *Cell and Tissue Research* 177: 375–381.

Fishelson L. **1978**. Oogenesis and spawn-formation in the pigmy lion fish *Dendrochirus brachypterus* (Pteroidae). *Marine Biology* 46(4): 341–348.

Fishelson L. **1981**. *Eilat coral reef fish: Biology and taxonomy*. Tel Aviv: Hakibbutz Hameuchad. 141 pp. [In Hebrew]

Fishelson L. **1993**. Israeli ichthyology in the Red Sea (1951–1992) — a personal perspective. *Israel Journal of Zoology* 39(4): 287–291.

Fishelson L. 1996. Lexicon of plants and animals of the land of Israel. Vol. 3: Aquatic Life. Jerusalem: Ministry of Defence Publishing House. 316 pp. [In Hebrew]  Fishelson L. 1997. Experiments and observations on food consumption, growth and starvation in *Dendrochirus brachypterus* and *Pterois volitans* (Pteroinae: Scorpaenidae). *Environmental Biology of Fishes* 50(4): 391–403.

Fishelson L. **2003**. Coral and fish biocoenosis: ecological cells gradually maturing in complexity, species composition and energy turnover. *Environmental Biology of Fishes* 68(4): 391–405.

Fishelson L. **2006**. Evolution in action – peacock-feather like supraocular tentacles of the lionfish, *Pterois volitans* – the distribution of a new signal. *Environmental Biology of Fishes* 75(3): 343–348.

Fishelson L, Golani D, Galil B, Goren M. 2010. Comparison of taste bud form, number and distribution in the oropharyngeal cavity of lizardfishes (Aulopiformes, Synodontidae). *Cybium* 34(3): 269–277. https://doi.org/10.26028/cybium/2010-343-005

Fishelson L, Golani D, Russell B, Galil B, Goren M.
2012. Comparative morphology and cytology of the alimentary tract in lizardfishes (Teleostei, Aulopiformes, Synodontidae). *Acta Zoologica* (Stockholm) 93(3): 308–318. http://dx.doi.org/10.1111/j.1463-6395.2011.00504.x

Fishelson L, Golani D, Russell B, Galil B, Goren M. 2012. Melanization of the alimentary tract in lizardfishes (Teleostei, Aulopiformes, Synodontidae). *Environmental Biology of Fish*es 95(2): 195–200. http://dx.doi.org/10.1007/s10641-012-9982-8

Fishelson L, Gon O, Goren M, Ben-David-Zaslow R. **2005**. The oral cavity and bioluminescent organs of the cardinal fish species *Siphamia permutata* and *S. cephalotes* (Perciformes, Apogonidae). *Marine Biology* 147(3): 603–609.

Fishelson L, Popper D, Gunderman N. **1971**. Diurnal cyclic behaviour of *Pempheris oualensis* Cuv. & Val. (Pempheridae, Teleostei). *Journal of Natural History* 5: 503–506.

Fishelson L, Russell B, Golani D, Goren M. 2011. Rodlet cells in the alimentary tract of three genera of lizardfishes (Synodontidae, Aulopiformes): more on these enigmatic "gate-guards" of fishes. *Cybium* 35(2): 121–129. https://doi.org/10.26028/cybium/2011-352-006

Fitch JE. **1951**. Notes on the squaretail, *Tetragonurus cuvieri*. *California Fish & Game* 37(1): 55–59.

Fitch JE. **1952**. Toxicity and taxonomic notes on the squaretail, *Tetragonurus cuvieri. California Fish & Game* 38(2): 251–252.

Fitch JE. **1964**. The ribbonfishes (family Trachipteridae) of the eastern Pacific Ocean, with a description of a new species. *California Fish & Game* 50(4): 228–240.

Fitch JE, Crooke SJ. **1984**. Revision of eastern Pacific catalufas (Pisces: Priacanthidae) with description of a new genus and discussion of the fossil record. *Proceedings of the California Academy of Sciences* (Series 4) 43(19): 301–315.
Fitch JE, Roedel PM. **1963**. A review of the frigate mackerels (genus *Auxis*) of the world. *FAO Fisheries Report* 6(3): 1329–1342.

Fleitmann D, Burns SJ, Mangini A, Mudelsee M, Kramers J, Villa I, Neff U, Al-Subbary AA, Buettner A, Hippler D, Matter A. 2007. Holocene ITCZ and Indian monsoon dynamics recorded in stalagmites from Oman and Yemen (Socotra). *Quaternary Science Reviews* 26(1–2): 170–188.

Fleitmann D, Matter A, Burns S, Al-Subbary A, Al-Aowah A. 2004. Geology and Quaternary climate history of Socotra. *Fauna of Arabia* (20): 27–32.

Follett WI, Dempster LJ. **1963**. Relationships of the percoid fish *Pentaceros richardsoni* Smith, with description of a specimen from the coast of California. *Proceedings of the California Academy of Sciences* (Series 4) 32(10): 315–338.

Forbes HO. **1890**. On a new genus of fishes of the family Percidae, from New Zealand. *Transactions and Proceedings of the New Zealand Institute* 22(30) [for 1889]: 273–275.

Forey PL. **1973**. A revision of the elopiform fishes, fossil and recent. *Bulletin of the British Museum (Natural History) Geology* Supplement 10. 222 pp.

Forey PL. 1973. Relationships of elopomorphs (pp. 351–368). In: Greenwood PH, Miles RS, Patterson C (eds) *Interrelationships of fishes*. London: Academic Press. 536 pp.

Forey PL. 1977. The osteology of Notelops Woodward, Rhacolepis Agassiz and Pachyrhizodus Dixon (Pisces: Teleostei). Bulletin of the British Museum (Natural History) Geology 28(2): 126–204.

Forey PL. **1984**. Yet more reflections on agnathan– gnathostome relationships. *Journal of Vertebrate Paleontology* 4(3): 330–343.

Forey PL. **1998**. *History of the coelacanth fishes*. London: Chapman & Hall. xii + 419 pp.

Forey PL, Hilton EJ. 2010. Two new Tertiary osteoglossid fishes (Teleostei: Osteoglossomorpha) with notes on the history of the family (pp. 215–246). In: Elliot D, Yu X, Maisey J, Miao D (eds) *Fossil fishes and related biota: Morphology, phylogeny and paleobiogeography*. Munich: Verlag Dr. Friedrich Pfeil. 472 pp.

Forey PL, Littlewood DTJ, Ritchie P, Meyer A. 1996.
Interrelationships of elopomorph fishes (pp. 175–191). In: Stiassny MLJ, Parenti LR, Johnson GD (eds) *Interrelationships* of fishes. San Diego: Academic Press. 496 pp.

Foroshchuk VP. 1991. A new species of the genus *Chascanopsetta* from the Saya-de-Malha Bank (Indian Ocean). *Voprosy Ikhtiologii* 31(1): 3–8. [In Russian. English translation in *Journal of Ichthyology* 31 (3): 75–81]

Foroshchuk VP, Fedorov VV. 1992. Poecilopsetta normani—A new species of flounder (Pleuronectidae) from Saya de Malha Bank, Indian Ocean. Journal of Ichthyology 32(7): 37–44. [English edition published March 1993] Forsskål P. 1775. *Descriptiones animalium avium*, *amphibiorum, piscium, insectorum, vermium; quae in itinere orientali observavit*. Post mortem auctoris edidit Carsten Niebuhr. Hauniae. 20 + xxxiv + 164 pp.

Forster GR, Badcock JR, Longbottom MR, Merrett NR, Thompson KS. 1970. Results of the Royal Society Indian Ocean deep slope fishing expedition, 1969. *Proceedings of the Royal Society of London* B 175: 367–404.

Forster JR. **1781**. *Zoologia Indica Selecta Tabulis XV aeneis illustrata*. Johann Jacob Gebauer, Halle. i-iv + 1-42, Pls. 15.

 Forster JR. 1795. Zoologia Indica, sistens I. Descriptiones animalium selectorum tabulis aeneis delineatorum (2<sup>nd</sup> edition). Zweyte sehr vermehrte Auflage. Halle. iv + 180 pp., Pls. 15. [In German and Latin]

Forster JR. 1801. In: Bloch ME, Schneider JG.

Fouda MM. **1995**. Life history strategies of four small-size fishes in the Suez Canal, Egypt. *Journal of Fish Biology* 46(4): 687–702.

Fourmanoir P. **1955**. Ichthyologie et pêche aux Comores. *Mémoires de l'Institut Scientifique de Madagascar* (Série A: Biologie Animale) 9: 187–239. [also cited as 1954]

Fourmanoir P. **1957**. Poissons téléostéens des eaux malgaches du canal de Mozambique. *Mémoires de l'Institut Scientifique de Madagascar* (Série F: Océanographie) 1: 1–316.

Fourmanoir P. **1959**. *Lutjanus guilcheri*, nouvelle espèce de Lutjanide capturée dans le Nord-Ouest de Madagascar. *Naturaliste Malgache* 10(1–2) (1958): 129–130, Pl. 3.

Fourmanoir P. **1961**. Requins de la côte ouest de Madagascar. *Mémoires de l'Institut Scientifique de Madagascar* (Série F: Océanographie) 4: 1–81.

Fourmanoir P. **1961**. Liste complémentaire des poissons du Canal de Mozambique. *Mémoires de l'Institut Scientifique de Madagascar* (Série F: Océanographie) 4: 83–107.

Fourmanoir P. **1963**. *Epinephelus fuscus*, nouvelle espèce de Serranidé trouvée dans les eaux Malgaches. *Bulletin du Muséum National d'Histoire Naturelle, Paris* (Série 2) 35(2): 140–142.

Fourmanoir P. **1963**. Raies et requins-scie de la côte ouest de Madagascar (ordre des *Batoidei*). *Cahiers ORSTOM* (Série Océanographie) 6: 33–58.

Fourmanoir P. **1966**. Trois espèces nouvelles de poissons perciformes du Viet-Nam et de la Réunion. *Bulletin du Muséum National d'Histoire Naturelle, Paris* (Série 2) 38(3): 217–222.

Fourmanoir P. **1966**. Nouvelle dénomination proposée pour un Scombridae de Canal de Mozambique: *Scomberomorus plurilineatus* nov. sp. *Bulletin du Muséum National d'Histoire Naturelle, Paris* (Série 2) 38(3): 223–226.

Fourmanoir P. **1967**. Nouvelle détermination proposée pour un Apogonidae de Mer Rouge et de l'Océan Indien. *Bulletin du Muséum National d'Histoire Naturelle, Paris* (Série 2) 39(2): 265–266. Fourmanoir P. **1970**. Notes ichtyologiques (I). *Cahiers ORSTOM* (Série Océanographie) 8(2): 19–33.

Fourmanoir P. **1971**. Notes ichtyologiques (II). *Cahiers ORSTOM* (Série Océanographie) 8(3) [for 1970]: 35–46.

Fourmanoir P. **1971**. Notes ichtyologiques (IV). *Cahiers ORSTOM* (Série Océanographie) 9(4): 491–500.

Fourmanoir P. **1977**. Description de deux nouvelles espèces d'Anthiinae (famille Serranidae). *Cahiers du Pacifique* 20: 267–270.

 Fourmanoir P, Crosnier A. 1964. Deuxième liste complémentaire des poissons du canal de Mozambique.
 Diagnoses préliminaires de 11 espèces nouvelles. *Cahiers* ORSTOM (Série Océanographie) 6(1963): 1–32.

Fourmanoir P, Do-Thi N-N. **1965**. Liste complémentaire des poissons marins de Nha-Trang. *Cahiers ORSTOM* (Série Océanographie) No. spécial: 1–114.

Fourmanoir P, Guézé P. **1961**. Poissons de la Réunion. II. Families des Labridés, Callodontides, Malacanthidae. *Publications de l'Institut de Recherche Scientifique de Madagascar*: 1–21.

Fourmanoir P, Guézé P. 1962. Les poissons de la Réunion.
5. Familles des Bothides, Soleides, Polymixiides, Uranoscopides, Priacanthides, Holocentrides, Acanthurides, Duleides, Pentapodides, Lutianides, Pomadasyides, Plectorrhynchides, Apogonides. *Publications de l'Institut de Recherche Scientifique de Madagascar* 5: 1–10.

Fourmanoir P, Guézé P. 1963. Les Poissons de la Réunion. 6. Familles des Istiophorides, Scombrides, Scomberomorides, Lethrinides, Sphyraenides, Polynemides, et familles de moindre importance économique. *Publications de l'Institut de Recherche Scientifique de Madagascar* 6: 5–24.

Fourmanoir P, Guézé P. 1966. Peloropsis frondosa. In: Young DR (ed) Proceedings of the California Academy of Sciences 1972–1974 (Fourth Series) 39(16): 285–310.

Fourmanoir P, Guézé P. **1967**. Poissons nouveaux ou peu connus provenant de la Réunion et de Madagascar. *Cahiers ORSTOM* (Série Océanographie) 5(1): 47–58.

Fourmanoir P, Guézé P. 1976. Pseudupeneus forsskali nom. nov. (= Mullus auriflamma Forsskål 1775). Travaux et Documents de l'ORSTOM 47: 45–48.

Fourmanoir P, Laboute P. 1976. Poissons de Nouvelles Caledonie et des Nouvelles Hebrides. Papeete: Les Editions du Pacifique. 376 pp.

Fourmanoir P, Rivaton J. **1979**. Poissons de la pente récifale externe de Nouvelle-Calédonie et des Nouvelles-Hébrides. *Cahiers de l'Indo-Pacifique* 1(4): 405–443.

Fowler HW. **1900**. Observations on fishes from the Caroline Islands. *Proceedings of the Academy of Natural Sciences of Philadelphia* 51 [for 1899]: 482–496.

Fowler HW. **1903**. Descriptions of several fishes from Zanzibar Island, two of which are new. *Proceedings of the Academy of Natural Sciences of Philadelphia* 55: 161–176. Fowler HW. **1903**. Descriptions of new, little known and typical Atherinidae. *Proceedings of the Academy of Natural Sciences of Philadelphia* 55: 727–742.

Fowler HW. **1904**. A collection of fishes from Sumatra. *Journal of the Academy of Natural Sciences of Philadelphia* (Second Series) 12(pt 4): 495–560.

Fowler HW. **1904**. New and little known Mugilidae and Sphyraenidae. *Proceedings of the Academy of Natural Sciences of Philadelphia* 55(3) [for 1903]: 743–752.

Fowler HW. 1905. New, rare or little known scombroids. No. 1. Proceedings of the Academy of Natural Sciences of Philadelphia 56 [for 1904]: 757–771.

Fowler HW. **1910**. Notes on batoid fishes. *Proceedings of the Academy of Natural Sciences of Philadelphia* 62: 468–475.

Fowler HW. **1911**. Some fishes from Venezuela. *Proceedings of the Academy of Natural Sciences of Philadelphia* 63: 419–437.

Fowler HW. **1912**. Notes on salmonoid and related fishes. *Proceedings of the Academy of Natural Sciences of Philadelphia* 63 [for 1911]: 551–571.

Fowler HW. **1918**. A list of Philippine fishes. *Copeia* 1918(58): 62–65.

Fowler HW. **1919**. Notes on synentognathous fishes. Proceedings of the Academy of Natural Sciences of Philadelphia 71: 2–15.

Fowler HW. **1922**. Notes on hemibranchiate and lophobranchiate fishes. *Proceedings of the Academy of Natural Sciences of Philadelphia* 73 [for 1921]: 437–448.

Fowler HW. 1923. Fishes from Madeira, Syria, Madagascar and Victoria, Australia. Proceedings of the Academy of Natural Sciences of Philadelphia 75: 33–45.

Fowler HW. **1924**. Notes and descriptions of Indian fishes. *Journal of the Bombay Natural History Society* 30(1): 36–41.

Fowler HW. **1925**. Descriptions of three new marine fishes from the Natal coast. *Annals of the Natal Museum* 5(2): 195–200.

Fowler HW. **1925**. Fishes from Natal, Zululand, and Portuguese East Africa. *Proceedings of the Academy of Natural Sciences of Philadelphia* 77: 187–268.

Fowler HW. **1925**. Notes and descriptions of Indian fishes. *Journal of the Bombay Natural History Society* 30(2): 314–321.

Fowler HW. **1926**. Descriptions of three unusual fishes from the Natal coast. *Annals of the Natal Museum* 5(3): 399–402.

Fowler HW. **1927**. Fishes of the tropical central Pacific. *Bulletin of the Bernice Pauahi Bishop Museum* **38**: 1–32.

Fowler HW. **1928**. Further notes and descriptions of Bombay shore fishes. *Journal of the Bombay Natural History Society* 33(1): 100–119.

Fowler HW. **1928**. Notes and descriptions of fishes from Ceylon. *Journal of the Bombay Natural History Society* 32(4): 704–710.

Fowler HW. **1928**. The fishes of Oceania. *Memoirs of the Bernice Pauahi Bishop Museum* 10: i–iii +1–540.

Fowler HW. **1929**. New and little-known fishes from the Natal coast. *Annals of the Natal Museum* 6(2): 245–264.

Fowler HW. **1931**. A small collection of fishes from Singapore. *Proceedings of the Academy of Natural Sciences of Philadelphia* 83: 443–448.

Fowler HW. **1931**. Contributions to the biology of the Philippine Archipelago and adjacent regions. The fishes of the families Pseudochromidae, Lobotidae, Pempheridae, Priacanthidae, Lutjanidae, Pomadasyidae, and Teraponidae, collected by the United States Bureau of Fisheries steamer *Albatross*, chiefly in Philippine Seas and adjacent waters. *Bulletin of the United States National Museum* 100(11): i–ix + 1–388.

Fowler HW. 1931. The fishes of Oceania. Supplement I. Memoirs of the Bernice Pauahi Bishop Museum 11(5): 313–381.

Fowler HW. 1932. Fishes obtained at Samoa in 1929. Occasional Papers of the Bernice Pauahi Bishop Museum of Polynesian Ethnology and Natural History 9(18): 1–16.

Fowler HW. **1932**. The fishes obtained by Lieut. H.C. Kellers of the United States Naval Eclipse Expedition of 1930, at Niuafoou Island, Tonga Group, in Oceania. *Proceedings of the United States National Museum* 81(2931): 1–9.

Fowler HW. **1933**. Contributions to the biology of the Philippine Archipelago and adjacent regions. The fishes of the families Banjosidae, Lethrinidae, Sparidae, Girellidae, Kyphosidae, Oplegnathidae, Gerridae, Mullidae, Emmelichthyidae, Sciaenidae, Sillaginidae, Arripidae, and Enoplosidae collected by the United States Bureau of Fisheries steamer *Albatross*, chiefly in Philippine seas and adjacent waters. *Bulletin of the United States National Museum* 100(12): 1–145.

Fowler HW. **1934**. Zoological results of the third De Schaunesee Siamese expedition. Part 1. Fishes. *Proceedings of the Academy of Natural Sciences of Philadelphia* 86: 67–163.

Fowler HW. **1934**. Descriptions of new fishes obtained 1907 to 1910, chiefly in the Philippine Islands and adjacent seas. *Proceedings of the Academy of Natural Sciences of Philadelphia* 85: 233–367.

Fowler HW. **1934**. The buckler dory and descriptions of three new fishes from off New Jersey and Florida. *Proceedings of the Academy of Natural Sciences of Philadelphia* 86: 353–361.

Fowler HW. **1934**. Fishes obtained by Mr. H.W. Bell-Marley chiefly in Natal and Zululand in 1929 to 1932. *Proceedings of the Academy of Natural Sciences of Philadelphia* 86: 405–514.

Fowler HW. **1934**. Natal fishes obtained by Mr. H.W. Bell-Marley. *Annals of the Natal Museum* 7(3): 403–433.

Fowler HW. **1934**. The fishes of Oceania. Supplement II. *Memoirs of the Bernice Pauahi Bishop Museum* 11(6): 385–466.

Fowler HW. **1935**. A synopsis of the fishes of China. Part V continued. The cods, opahs, flounders, soles, john dories,

berycoids, pipe fishes, silversides, mullets, barracudas and thread fishes. *Hong Kong Naturalist* 6(1): 62–77.

Fowler HW. **1935**. Description of a new scorpaenoid fish (*Neomerinthe hemingwayi*) from off New Jersey. *Proceedings* of the Academy of Natural Sciences of Philadelphia 87: 41–43.

Fowler HW. **1935**. South African fishes received from Mr. H.W. Bell-Marley in 1935. *Proceedings of the Academy of Natural Sciences of Philadelphia* 87: 361–408.

Fowler HW. 1936. The marine fishes of West Africa based on the collection of the American Museum Congo Expedition, 1909–1915. Part I. Bulletin of the American Museum of Natural History 70(1): 1–605.

Fowler HW. **1936**. The marine fishes of West Africa based on the collection of the American Museum Congo Expedition, 1909–1915. Part II. *Bulletin of the American Museum of Natural History* 70(2): 607–1493.

Fowler HW. **1936**. A synopsis of the fishes of China. Part VI continued. The mackerels and related fishes. *Hong Kong Naturalist* 7(2): 186–202.

Fowler HW. **1938**. Descriptions of new fishes obtained by the United States Bureau of Fisheries steamer *Albatross*, chiefly in Philippine seas and adjacent waters. *Proceedings of the United States National Museum* 85(3032): 31–135.

Fowler HW. **1938**. The fishes of the George Vanderbilt South Pacific Expedition, 1937. *Monographs of the Academy of Natural Sciences of Philadelphia* 2: i–iv + 1–349.

Fowler HW. **1939**. Ichthyological notes. No. 1. *Notulae Naturae* (Academy of Natural Sciences of Philadelphia) 3: 1–2.

Fowler HW. **1939**. New subfamilies, genera and subgenera of fishes. *Notulae Naturae* (Academy of Natural Sciences of Philadelphia) 26: 1–2.

Fowler HW. **1941**. Contributions to the biology of the Philippine archipelago and adjacent regions. The fishes of the groups Elasmobranchii, Holocephali, Isospondyli, and Ostarophysi obtained by the United States Bureau of Fisheries steamer *Albatross* in 1907 to 1910, chiefly in the Philippine Islands and adjacent seas. *Bulletin of the United States National Museum* 100(13): i–x + 1–879.

Fowler HW. **1941**. The George Vanderbilt Oahu survey: the fishes. *Proceedings of the Academy of Natural Sciences of Philadelphia* 93: 247–279.

Fowler HW. **1943**. Contributions to the biology of the Philippine Archipelago and adjacent regions. Descriptions and figures of new fishes obtained in Philippine seas and adjacent waters by the United States Bureau of Fisheries steamer *Albatross. Bulletin of the United States National Museum* 100(14, pt 2): i–iii + 53–91.

Fowler HW. **1944**. Fishes obtained in the New Hebrides by Dr. Edward L. Jackson. *Proceedings of the Academy of Natural Sciences of Philadelphia* 96: 155–199. Fowler HW. **1944**. Results of the fifth George Vanderbilt Expedition (1941) (Bahamas, Caribbean Sea, Panama, Galápagos Archipelago and Mexican Pacific islands). The fishes. *Monographs of the Academy of Natural Sciences of Philadelphia* 6: 57–529.

Fowler HW. **1945**. Fishes from Saipan Island, Micronesia. Proceedings of the Academy of Natural Sciences of Philadelphia 97: 59–74.

Fowler HW. **1945**. Fishes of Chile. Systematic catalog. *Revista Chilena de Historia Natural* (Santiago) 15–17: 1–171.

Fowler HW. **1946**. A collection of fishes obtained in the Riu Kiu Islands by Captain Ernest R. Tinkham A.U.S. *Proceedings of the Academy of Natural Sciences of Philadelphia* 98: 123–218.

Fowler HW. **1949**. The fishes of Oceania. Supplement III. Memoirs of the Bernice Pauahi Bishop Museum of Polynesian Ethnology and Natural History 12(2): 1–152.

Fowler HW. 1949. Results of the two Carpenter African Expeditions, 1946–1948. Part II – The fishes. Proceedings of the Academy of Natural Sciences of Philadelphia 101: 233–275.

Fowler HW. 1953. On a collection of fishes made by Dr. Marshall Laird at Norfolk Island. *Transactions of the Royal Society of New Zealand* 81(2): 257–267.

Fowler HW. **1956**. *Fishes of the Red Sea and southern Arabia*. *I. Branchiostomida to Polynemida*. Jerusalem: Weizmann Science Press. 240 pp.

Fowler HW. **1964**. A catalog of world fishes. *Quarterly Journal of the Taiwan Museum* (Taipei) 17(3/4): 1–62.

Fowler HW, Ball SC. **1924**. Descriptions of new fishes obtained by the *Tanager* expedition of 1923 in the Pacific Islands west of Hawaii. *Proceedings of the Academy of Natural Sciences of Philadelphia* 76: 269–274.

Fowler HW, Ball SC. **1925**. Fishes of Hawaii, Johnston Island, and Wake Island. *Bulletin of the Bernice Pauahi Bishop Museum* 26: 1–31.

Fowler HW, Bean BA. **1928**. Contributions to the biology of the Philippine Archipelago and adjacent regions. The fishes of the families Pomacentridae, Labridae, and Callyodontidae collected by the United States Bureau of Fisheries steamer *Albatross* chiefly in Philippine seas and adjacent waters. *Bulletin of the United States National Museum* 100(7): i–viii + 1–525.

Fowler HW, Bean BA. **1929**. Contributions to the biology of the Philippine Archipelago and adjacent waters. The fishes of the series Capriformes, Ephippiformes, and Squamipennes, collected by the United States Bureau of Fisheries steamer *Albatross* chiefly in Philippine Seas and adjacent waters. *Bulletin of the United States National Museum* 100(8): i–xi + 1–352. Fowler HW, Bean BA. **1930**. Contributions to the biology of the Philippine Archipelago and adjacent regions. The fishes of the families Amiidae, Chandidae, Duleidae, and Serranidae, obtained by the United States Bureau of Fisheries steamer *Albatross*, in 1907 to 1910, chiefly in the Philippine Islands and adjacent seas. *Bulletin of the United States National Museum* 100(10): i–xi + 1–334.

Fowler HW, Steinitz H. **1956**. Fishes from Cyprus, Iran, Iraq, Israel and Oman. *Bulletin of the Research Council of Israel* 5B(3–4): 260–292.

Frable BW, Tea Y-K. **2019**. A new species of damselfish (Teleostei: Pomacentridae: *Pomacentrus*) from Nosy Faho, Madagascar. *Copeia* 107(2): 323–331. http://dx.doi. org/10.1643/CI-19-221

Franca P da, see Da Franca P.

Francis MP, Stevens JD, Last PR. **1988**. New records of *Somniosus* (Elasmobranchii: Squalidae) from Australasia, with comments on the taxonomy of the genus. *New Zealand Journal of Marine and Freshwater Research* 22(3): 401–409.

Francis TJ, Shor GG. **1966**. Seismic refraction measurements in the northwest Indian Ocean. *Journal of Geophysical Research* 71(2): 427–449. https://doi.org/10.1029/JZ071i002p00427

Franz V. 1910. Die japanischen Knochenfische der Sammlungen Haberer und Doflein. Beiträge zur Naturgeschicthe Ostasiens. Abhandlungen der Mathematisch-Physikisch Klasse, Beiträge der Akademie Wissenschaften (Series 2) 4(Suppl. 1): 1–135.

Fraser TH. **1972**. A new species of the klipfish genus Springeratus (Clinidae) from the Indian Ocean. J.L.B. Smith Institute of Ichthyology Special Publication 9: 1–14.

Fraser TH. **1972**. Comparative osteology of the shallow water cardinal fishes (Perciformes: Apogonidae) with reference to the systematics and evolution of the family. *Ichthyological Bulletin of the J.L.B. Smith Institute of Ichthyology* 34: 1–105.

Fraser TH. **1973**. The fish *Elops machnata* in South Africa. *J.L.B. Smith Institute of Ichthyology Special Publication* 11: 1–6.

Fraser TH. **1998**. A new species of cardinalfish (Apogonidae) from the Philippines, with comments on species of *Apogon* with six first dorsal spines. *Proceedings of the Biological Society of Washington* 111(4): 986–991.

Fraser TH. **1999**. A new species of cardinalfish (Perciformes: Apogonidae) from the Bay of Bengal, Indian Ocean. *Proceedings of the Biological Society of Washington* 112(1): 40–44.

Fraser TH. 2000. A new species of *Apogon* (Perciformes: Apogonidae) from the Saya de Malha Bank, Indian Ocean, with redescriptions of *Apogon regani* Whitley, 1951, *A. gardineri* Regan, 1908, and *A. heraldi* (Herre, 1943). *Proceedings of the Biological Society of Washington* 113(1): 249–263. Fraser TH. **2005**. A review of the species in the *Apogon fasciatus* group with a description of a new species of cardinalfish from the Indo-West Pacific (Perciformes: Apogonidae). *Zootaxa* 924: 1–30. https://doi.org/10.11646/ zootaxa.924.1.1

Fraser TH. **2008**. Cardinalfishes of the genus *Nectamia* (Apogonidae, Perciformes) from the Indo-Pacific region with descriptions of four new species. *Zootaxa* 1691: 1–52. https://doi.org/10.11646/zootaxa.1691.1.1

Fraser TH. 2010. A new deep-water cardinalfish (*Neamia*: Apogonidae) from Kiritimati Atoll, Kiribati. *Proceedings of the Biological Society of Washington* 123(2): 153–158. http://dx.doi.org/10.2988/10-03.1

Fraser TH. **2012**. A new species of deeper dwelling West Pacific cardinalfish (Percomorpha: Apogonidae) with a redescription of *Ostorhinchus atrogaster*. *Zootaxa* 3492: 77– 84. https://doi.org/10.11646/zootaxa.3492.1.5

Fraser TH. 2013. Family status of *Amioides* and the senior synonym of *Coranthus* (Percomorpha: Apogonidae). *Copeia* 2013(1): 23–30. https://doi.org/10.1643/CI-12-060

Fraser TH. **2013**. A new genus of cardinalfish (Apogonidae: Percomorpha), redescription of *Archamia* and resemblances and relationships with *Kurtus* (Kurtidae: Percomorpha). *Zootaxa* 3714(1): 1–63. https://doi.org/10.11646/ zootaxa.4144.2.5

Fraser TH. 2018. Redescription of the cardinalfish Apogonichthyoides maculipinnis (Percomorpha: Apogonidae: Apogoninae) from the Maldives, last collected 118 years ago. Journal of the Ocean Science Foundation 30: 100–104. http://dx.doi.org/10.5281/zenodo.1409246

Fraser TH, Allen GR. **2001**. A new species of cardinalfish in *Neamia* (Apogonidae, Perciformes) from Mauritius, Indian Ocean, with a review of *Neamia octospina*. *Records of the Western Australian Museum* 20: 159–165.

Fraser TH, Allen GR. **2010**. Cardinalfish of the genus *Apogonichthyoides* Smith, 1949 (Apogonidae) with a description of a new species from the West-Pacific region. *Zootaxa* 2348: 40–56. http://dx.doi.org/10.11646/ zootaxa.2348.1.3

Fraser TH, Allen GR. **2011**. A new cardinalfish of the genus *Apogonichthyoides* (Perciformes, Apogonidae) from Raja Ampat Islands, with a key to species. *Zootaxa* 3095: 63–68. https://doi.org/10.11646/zootaxa.3095.1.6

Fraser TH, Fourmanoir P. **1971**. The deepwater fish *Scombrosphyraena oceanica* from the Caribbean Sea with comments on its possible relationships. *J.L.B. Smith Institute of Ichthyology Special Publication* 8: 1–7.

Fraser TH, Lachner EA. **1984**. An unusual Indo-West Pacific cardinalfish of the genus *Apogon* (Teleostei: Apogonidae). *Proceedings of the Biological Society of Washington* 97(3): 632–636. Fraser TH, Lachner EA. 1985. A revision of the cardinalfish subgenera *Pristiapogon* and *Zoramia* (genus *Apogon*) of the Indo-Pacific region (Teleostei: Apogonidae). *Smithsonian Contributions to Zoology* No. 412: i–iii + 1–47.

Fraser TH, Mabuchi K. **2014**. In: Mabuchi K, Fraser TH, Song H, Azuma Y, Nishida M.

Fraser TH, Prokofiev AM. **2016**. A new genus and species of cardinalfish (Percomorpha, Apogonidae, Sphaeramiini) from the coastal waters of Vietnam: luminescent or not? *Zootaxa* 4144(2): 227–242. http://dx.doi.org/10.11646/ zootaxa.4144.2.5

Fraser TH, Randall JE. **2002**. *Apogon dianthus*, a new species of cardinalfish (Perciformes: Apogonidae) from Palau, western Pacific Ocean with comments on other species of the subgenus *Apogon*. *Proceedings of the Biological Society of Washington* 115(1): 25–31.

Fraser TH, Randall JE. **2011**. Two new species of *Foa* (Apogonidae) from the Pacific Plate, with redescriptions of *Foa brachygramma* and *Foa fo. Zootaxa* 2988: 1–27. https://doi.org/10.5281/zenodo.278368

Fraser TH, Randall JE, Allen GR. **2002**. Clarification of the cardinalfishes (Apogonidae) previously confused with *Apogon moluccensis* Valenciennes, with a description of a related new species. *Raffles Bulletin of Zoology* 50(1): 175–184.

Fraser TH, Randall JE, Lachner EA. **1999**. A review of the Red Sea cardinalfishes of the *Apogon bandanensis* complex, with a description of a new species. *J.L.B. Smith Institute of Ichthyology Special Publication* 63: 1–13.

Fraser TH, Smith MM. **1974**. An exterilium larval fish from South Africa with comments on its classification. *Copeia* 1974(4): 886–892.

Fraser TH, Struhsaker PJ. **1991**. A new genus and species of cardinalfish (Apogonidae) from the Indo-West Pacific, with a key to apogonine genera. *Copeia* 1991(3): 718–722.

Fraser-Brunner A. 1931. Some interesting West African fishes, with descriptions of a new genus and two new species. *Annals and Magazine of Natural History* (Series 10) 8(45): 217–235.

Fraser-Brunner A. **1933**. A revision of the chaetodont fishes of the subfamily Pomacanthinae. *Proceedings of the Zoological Society of London* pt 3(30): 543–599.

Fraser-Brunner A. **1935**. Notes on the plectognath fishes. I. A synopsis of the genera of the family Balistidae. *Annals and Magazine of Natural History* (Series 10) 15(90): 658–663.

Fraser-Brunner A. **1935**. Notes on the plectognath fishes. II. A synopsis of the genera of the family Ostraciontidae. *Annals and Magazine of Natural History* (Series 10) 16(92): 313–320.

Fraser-Brunner A. **1938**. Notes on the classification of certain British fishes. *Annals and Magazine of Natural History* (Series 11) 2(11): 410–416. Fraser-Brunner A. **1940**. Notes on the plectognath fishes. III. On *Monacanthus setifer* Bennett and related species, with a key to the genus *Stephanolepis* and descriptions of four new species. *Annals and Magazine of Natural History* (Series 11) 5(30): 518–535.

Fraser-Brunner A. **1941**. Notes on the plectognath fishes. V. The families of triacanthiform fishes, with a synopsis of the genera and description of a new species. *Annals and Magazine of Natural History* (Series 11) 7(41): 420–430.

Fraser-Brunner A. 1941. Notes on the plectognath fishes. VI. A synopsis of the genera of the family Aluteridae, and descriptions of seven new species. *Annals and Magazine of Natural History* (Series 11) 8(45): 176–199.

Fraser-Brunner A. **1941**. Notes on the plectognath fishes. VII. The Aracanidae, a distinct family of ostraciontoid fishes, with descriptions of two new species. *Annals and Magazine of Natural History* (Series 11) 8(46): 306–313.

Fraser-Brunner A. **1943**. Notes on the plectognath fishes. VIII. The classification of the suborder Tetraodontoidea, with a synopsis of the genera. *Annals and Magazine of Natural History* (Series 11) 10(61): 1–18.

Fraser-Brunner A. **1949**. *Holacanthus xanthotis*, sp. n., and other chaetodont fishes from the Gulf of Aden. *Proceedings of the Zoological Society of London* 120(1): 43–48.

Fraser-Brunner A. **1949**. On the fishes of the genus *Euthynnus*. *Annals and Magazine of Natural History* (Series 12) 2(20): 622–627.

Fraser-Brunner A. **1949**. Note on the electric rays of the genus *Torpedo. Annals and Magazine of Natural History* (Series 12) 2(24): 943–947.

Fraser-Brunner A. 1950. Studies in plectognath fishes from the *Dana*-Expeditions. I. An interesting new genus of triacanthodid fishes from the Celebes Sea. *Dana Report* 35: 1–8.

Fraser-Brunner A. **1950**. The fishes of the family Scombridae. *Annals and Magazine of Natural History* (Series 12) 3(26): 131–163.

Fraser-Brunner A. **1951**. Some new blennioid fishes, with a key to the genus *Antennablennius*. *Annals and Magazine of Natural History* (Series 12) 4(39): 213–220.

Fraser-Brunner A. **1951**. The ocean sunfishes (family Molidae). Bulletin of the British Museum (Natural History) Zoology 1(6): 89–121.

Fraser-Brunner A. **1955**. A synopsis of the centropomid fishes of the subfamily Chandinae, with descriptions of a new genus and two new species. *Bulletin of the Raffles Museum* 25: 185–213.

Fraser-Brunner A, Whitley GP. **1949**. A new pipefish from Queensland. *Records of the Australian Museum* 22(2): 148–150. https://doi.org/10.3853/j.0067-1975.22.1949.595 Fratantoni DM, Bower AS, Johns WE, Peters H. **2006**. Somali Current rings in the eastern Gulf of Aden. *Journal of Geophysical Research* 111(C9): C09039.

Freer DWL, Griffiths CL. **1993**. The fishery for, and general biology of, the St Joseph *Callorhinchus capensis* (Dumeril) off the south-western Cape, South Africa. *South African Journal of Marine Science* 13(1): 63–74.

Freihofer WC. **1963**. Patterns of the ramus lateralis accessorius and their systematic significance in teleostean fishes. *Stanford Ichthyological Bulletin* 8(2): 80–189.

Freinschlag M, Patzner RA. 2012. Shrimp-gobies in the southern Gulf of Aqaba (Red Sea) (Osteichthyes: Gobiidae). *Zoology in the Middle East* 55: 41–46. http://dx.doi.org/10.1080/09397140.2012.10648916

Frey L, Coates MI, Tietjen K, Rücklin M, Klug C. 2020. A symmoriiform from the Late Devonian of Morocco demonstrates a derived jaw function in ancient chondrichthyans. *Communications Biology* 3(681): 1–10. http://dx.doi.org/10.1038/s42003-020-01394-2

Fricke H, Fricke S. **1977**. Monogamy and sex change by aggressive dominance in coral reef fish. *Nature* (London) 266: 830–832.

Fricke H, Kacher H. 1982. A mound-building deep water sand tilefish of the Red Sea: *Hoplolatilus geo* n. sp. (Perciformes: Branchiostegidae). Observations from a research submersible. *Senckenbergiana Maritima* 14(5–6): 245–259.

Fricke R. 1980. Neue Fundorte und noch nicht beschriebene Geschlechtsunterschiede einiger Arten der Gattung *Callionymus* (Pisces, Perciformes, Callionymidae), mit Bemerkungen zur Systematik innerhalb dieser Gattung und Beschreibung einer neuen Untergattung und einer neuen Art. Annali del Museo Civico di Storia Naturale "Giacomo Doria" 83: 57–105.

Fricke R. **1981**. Revision of the genus *Synchiropus* (Teleostei: Callionymidae). Braunschweig (J Cramer). 194 pp.

Fricke R. **1981**. *Diplogrammus (Climacogrammus) pygmaeus* sp. nov., a new callionymid fish (Pisces, Perciformes, Callionymoidei) from the South Arabian coast, northwestern Indian Ocean. *Journal of Natural History* 15(4): 685–692.

Fricke R. **1981**. On a new species of the family Callionymidae (Pisces, Perciformes, Callionymoidei), *Callionymus stigmatopareius* sp. nov. from Mozambique. *Journal of Natural History* 15(1): 161–167.

Fricke R. **1981**. The kaianus-group of the genus *Callionymus* (Pisces, Callionymidae), with descriptions of six new species. *Proceedings of the California Academy of Sciences* (Series 4) 42(14): 349–377.

Fricke R. **1981**. Two new and a rare species of the genus *Callionymus* (Teleostei: Callionymidae). *Annali del Museo Civico di Storia Naturale "Giacomo Doria"* 83: 387–400. Fricke R. **1982**. Nominal genera and species of dragonets (Teleostei: Callionymidae, Draconettidae). *Annali del Museo Civico di Storia Naturale "Giacomo Doria"* 84: 53–92.

Fricke R. **1983**. *Revision of the Indo-Pacific genera and species of the dragonet family Callionymidae (Teleostei)*. Theses Zoologicae 3. Braunschweig: J. Cramer. x +774 pp.

Fricke R. 1984. Callionymidae. Dragonets. In: Fischer W, Bianchi G (eds) FAO species identification sheets for fishery purposes. Western Indian Ocean (Fishing Area 51). Vol. 1. Rome: FAO. [unpaginated]

Fricke R. **1988**. Systematik und historische Zoogeographie der Callionymidae (Teleostei) des Indischen Ozeans. Inaugural Dissertation, Alberts-Ludwigs-Universität, Freiburg. 612 pp.

Fricke R. **1989**. New species and new records of *Callionymus* from the Pacific Ocean (Teleostei: Callionymidae). *Hydrobiologia* 183(1): 47–57.

Fricke R. **1990**. A new and a rare species of dragonet (Teleostei: Callionymidae) from New Guinea and the Solomon Islands. *Stuttgarter Beiträge zur Naturkunde* (Serie A) (Biologie) 446: 1–13.

Fricke R. **1992**. Revision of the family Draconettidae (Teleostei), with descriptions of two new species and a new subspecies. *Journal of Natural History* 26(1): 165–195.

Fricke R. 1992. Types in the fish collection of the Staatliches Museum f
ür Naturkunde in Stuttgart. Part 2. The Klunzinger collection. *Stuttgarter Beiträge zur Naturkunde* (Serie A) (Biologie) 473: 1–25.

Fricke R. 1994. Tripterygiid fishes of Australia, New Zealand and the southwest Pacific Ocean with descriptions of 2 new genera and 16 new species (Teleostei). Theses Zoologicae 24. Königstein: Koeltz Scientific Books. ix + 585 pp.

Fricke R. 1997. Tripterygiid fishes of the western and central Pacific, with descriptions of 15 new species, including an annotated checklist of world Tripterygiidae (Teleostei). Theses Zoologicae 29. Königstein: Koeltz Scientific Books. 607 pp.

Fricke R. 1999. Fishes of the Mascarene Islands (Réunion, Mauritius, Rodriguez): an annotated checklist, with descriptions of new species. Theses Zoologicae 31. Koenigstein: Koeltz Scientific Books. viii + 759 pp.

Fricke R. **2000**. Callionymidae of New Caledonia, with remarks on related species and descriptions of 10 new species from New Caledonia, Australia, New Guinea, and Hawaii (Teleostei). *Stuttgarter Beiträge zur Naturkunde* Serie A (Biologie) 617: 1–81.

Fricke R. 2002. Annotated checklist of the dragonet families Callionymidae and Draconettidae (Teleostei: Callionymoidei), with comments on callionymid fish classification. *Stuttgarter Beiträge zur Naturkunde* Serie A (Biologie) 645: 1–103. Fricke R. **2004**. *Cocotropus richeri*, a new species of velvetfish (Teleostei: Aploactinidae) from Lifou, Loyalty Islands. *Stuttgarter Beiträge zur Naturkunde* Serie A (Biologie) 660: 1–7.

Fricke R. 2005. Types in the fish collection of the Staatliches Museum für Naturkunde in Stuttgart, described in 1845– 2004. *Stuttgarter Beiträge zur Naturkunde* Serie A (Biologie) 684: 1–95.

Fricke R. **2008**. Authorship, availability and validity of fish names described by Peter (Pehr) Simon Forsskål and Johann Christian Fabricius in the 'Descriptiones animalium' by Carsten Niebuhr in 1775 (Pisces). *Stuttgarter Beiträge zur Naturkunde A, Neue Serie* 1: 1–76.

Fricke R. 2009. Systematics of the Tripterygiidae (triplefins) (pp. 31–67). In: Patzner RA, Gonçalves EJ, Hastings PA, Kapoor BG (eds) *The biology of blennies*. Science Publishers, Enfield, New Hampshire. xi + 482 pp.

Fricke R, Al-Hassan LAJ. 1995. Raja pita, a new species of skate from the Arabian/Persian Gulf (Elasmobranchii: Rajiformes). Stuttgarter Beiträge zur Naturkunde Serie A (Biologie) 529: 1–8.

Fricke R, Bilecenoglu M, Sari HM. **2007**. Annotated checklist of fish and lamprey species (Gnathostomata and Petromyzontomorphi) of Turkey, including a Red List of threatened and declining species. *Stuttgarter Beiträge zur Naturkunde* Serie A (Biologie) 706: 1–174.

Fricke R, Bogorodsky SV, Mal AO. **2014**. Review of the genus *Diplogrammus* (Teleostei: Callionymidae) of the Red Sea, with description of a new species from Saudi Arabia. *Journal of Natural History* 48(39–40): 2419–2448. https://doi.org/10.1080/00222933.2014.925598

Fricke R, Cadet C, Mulochau T. 2016. First record of the sawspine dragonet, *Diplogrammus infulatus* Smith 1963 (Actinopterygii: Perciformes: Callionymidae), from La Réunion, south-western Indian Ocean. *Acta Ichthyologica et Piscatoria* 45(4): 407–410. http://dx.doi.org/10.3750/ AIP2015.45.4.09

Fricke R, Durville P, Bernardi G, Borsa P, Mou-Tham G, Chabanet P. 2013. Checklist of the shore fishes of Europa Island, Mozambique Channel, southwestern Indian Ocean, including 302 new records. *Stuttgarter Beiträge zur Naturkunde A, Neue Serie* 6: 247–276.

Fricke R, Durville P, Mulochau T. 2013. Scorpaenopsis rubrimarginatus, a new species from Réunion, southwestern Indian Ocean (Teleostei: Scorpaenidae). Cybium 37(3): 207–215. https://doi.org/10.26028/cybium/2013-373-008

Fricke R, Golani D. **2012**. *Limnichthys marisrubri*, a new species of sand diver (Teleostei: Creediidae) from the Red Sea. *Stuttgarter Beiträge zur Naturkunde A, Neue Serie* 5: 287–292.

Fricke R, Golani D. **2013**. *Callionymus profundus* n. sp., a new species of dragonet from the Gulf of Aqaba (Gulf of Eilat), Red Sea (Teleostei: Callionymidae). *Stuttgarter Beiträge zur Naturkunde A, Neue Serie* 6: 277–285.

Fricke R, Golani D, Appelbaum-Golani B. 2014. Emmelichthys marisrubri, a new rover from the southern Red Sea (Teleostei: Emmelichthyidae). Cybium 38(2): 83–87. https://doi.org/10.26028/cybium/2014-382-001

Fricke R, Golani D, Appelbaum-Golani B. 2014. Evoxymetopon moricheni, a new cutlassfish from the northern Red Sea (Teleostei: Trichiuridae). Ichthyological Research 61(3): 293–297. http://dx.doi.org/10.1007/s10228-014-0394-y

Fricke R, Golani D, Appelbaum-Golani B. 2015. Confirmed record of the Ambon scorpionfish *Pteroidichthys amboinensis* (Teleostei: Scorpaenidae) from the Red Sea. *Marine Biodiversity Records* 8(e113): 1–3. http://dx.doi.org/10.1017/S1755267215000949

Fricke R, Jawad LA, Al-Mamry JM. 2014. Callionymus omanensis, a new species of dragonet from Oman, northwestern Indian Ocean (Teleostei: Callionymidae). Journal of Fish Biology 85(5): 1303–1319. http://dx.doi.org/10.1111/jfb.12460

Fricke R, Koch I. **1990**. A new species of the lantern shark genus *Etmopterus* from southern Africa (Elasmobranchii: Squalidae). *Stuttgarter Beiträge zur Naturkunde* Serie A (Biologie) 450: 1–9.

Fricke R, Kulbicki M. 2006. Checklist of the shore fishes of New Caledonia. First edition. Compendium of marine species from New Caledonia. *Documents Scientifiques et Techniques* II(7) (vol. spécial): 313–357, Pls 15/1–15/2.

Fricke R, Kulbicki M, Wantiez L. **2011**. Checklist of the fishes of New Caledonia, and their distribution in the Southwest Pacific Ocean (Pisces). *Stuttgarter Beiträge zur Naturkunde A, Neue Serie* 4: 341–463.

Fricke R, Mahafina J, Behivoke F, Jaonalison H, Léopold M, Ponton D. 2018. Annotated checklist of the fishes of Madagascar, southwestern Indian Ocean, with 158 new records. *FishTaxa* 3(1): 1–432.

Fricke R, Mulochau T, Durville P, Chabanet P, Tessier E, Letourneur Y. **2009**. Annotated checklist of the fish species (Pisces) of La Réunion, including a Red List of threatened and declining species. *Stuttgarter Beiträge zur Naturkunde A*, *Neue Serie* 2: 1–168.

Fricke R, Randall JE. **1992**. Tripterygiid fishes of the Maldives Islands, with descriptions of two new species (Teleostei: Blennioidei). *Stuttgarter Beiträge zur Naturkunde* Serie A (Biologie) 484: 1–13.

Fridman D. **1972**. Notes on the ecology of a new species of *Photoblepharon* (Pisces, Anomalopidae) from the Gulf of Elat (Aqaba). *Scientific Newsletter, H. Steinitz Marine Biology Laboratory* 2: 1–2.

Fridman D, Malmqvist T. **1994**. *Wonders of the Red Sea*. Herzliya, Israel: Palphot Ltd. [unpaginated]

Friedman M. 2010. Explosive morphological diversification of spiny-finned teleost fishes in the aftermath of the end-Cretaceous extinction. *Proceedings of the Royal Society* B 277(1688): 1675–1683. http://dx.doi.org/10.1098/rspb.2009.2177

Friedman M, Shimada K, Martin LD, Everhart MJ, Liston J, Maltese A, Triebold M. 2010. 100-million-year dynasty of giant planktivorous bony fishes in the Mesozoic seas. *Science* 327(5968): 990–993. http://dx.doi.org/10.1126/ science.1184743

Fries BF. **1837**. *Pterycombus*, ett nytt fisk-slägte från Ishafvet. *Kungliga Svenska Vetenskapsakademiens handlingar* 1837: 14–22.

Fritzsche RA. **1976**. A review of the cornetfishes, genus *Fistularia* (Fistulariidae), with a discussion of intrageneric relationships and zoogeography. *Bulletin of Marine Science* 26(2): 196–204.

Fritzsche RA. **1980**. Revision of the eastern Pacific Syngnathidae (Pisces: Syngnathiformes), including both recent and fossil forms. *Proceedings of the California Academy of Sciences* (Series 4) 42(6): 181–227.

Froese R, Pauly D (eds). **2008**, **2014**, *et al. FishBase* (online). https://www.fishbase.in/search.php

Frøiland Ø. 1972. The scorpaenids of the Red Sea (Pisces: Scorpaenidae): a taxonomical and zoogeographical study. Thesis, University of Bergen. v + 159 pp.

Frøiland Ø. 1976. Pterois zebra Quoy and Gaimard, 1825. (Pisces: Scorpaenidae): request for suppression, using plenary powers. Z.N.(S.) 2113. Bulletin of Zoological Nomenclature 32(4): 250–251.

Fry GC, Brewer DT, Venables WN. 2006. Vulnerability of demersal fishes to commercial fishing: evidence from a study around a tropical seamount in Papua New Guinea. *Fisheries Research* 81: 126–141.

FSO (Fishes of the Southern Ocean), *see* Gon O, Heemstra PC (eds). 1990.

Fukuhara T, Chakraborty A, Iwatsuki Y. **2006**. *Gerres infasciatus* (Perciformes: Gerreidae) from the Indian Ocean. *Ichthyological Research* 53(2): 197–199.

Fukui A, Ozawa T. **1990**. Early ontogeny of two bothid species, *Psettina iijimae* and *Laeops kitaharae*. *Japanese Journal of Ichthyology* 37(2): 127–132.

Fukui Y, Motomura H. **2016**. *Terelabrus flavocephalus* sp. nov., a new hogfish (Perciformes: Labridae) from the Maldives, Indian Ocean. *Ichthyological Research* 63(4): 529–535. http://dx.doi.org/10.1007/s10228-016-0523-x

- Gabie V. **1960**. A new record from Mozambique belonging to the family Syngnathidae. *Revista de Biologia Lisboa* 2(2): 73–74.
- Gabrié C, Vasseur P, Randriamiarana H, Maharavo J, Mara E.
  2000. Chapter 14. The reefs of Madagascar (pp. 411–444).
  In: McClanahan TR, Sheppard CRC, Obura DO (eds) Coral reefs of the Indian Ocean: Their ecology and conservation.
  Oxford: Oxford University Press. xxiii + 525 pp.
- Gaither MR, Randall JE. **2012**. On the validity of the cirrhitid fish genus *Itycirrhitus. aqua*, *International Journal of Ichthyology* 18(4): 219–226.
- Gaither MR, Randall JE. **2013**. Reclassification of the Indo-Pacific hawkfish *Cirrhitus pinnulatus* (Forster). *Zootaxa* 3599(2): 189–196. http://dx.doi.org/10.11646/ zootaxa.3599.2.5
- Gallup CD, Edwards RL, Johnson RG. 1994. The timing of high sea levels over the past 200,000 years. *Science* 263: 796–800.
- Gardiner BG. **1960**. A revision of certain actinopterygian and coelacanth fishes, chiefly from the Lower Lias. *Bulletin of the British Museum (Natural History) Geology* 4(7): 239–384.
- Gardiner BG, Maisey JG, Littlewood DTJ. **1996**. Interrelationships of basal neopterygians (pp. 117–146). In: Stiassny MLJ, Parenti LR, Johnson GD (eds) *Interrelationships of fishes*. San Diego: Academic Press. 496 pp.
- Gardiner JS. **1936**. The reefs of the Western Indian Ocean. I. Chagos Archipelago. II. The Mascarene region. *Transactions* of *Linnean Society, London* 19: 393–436.
- Garman S. **1880**. New species of selachians in the museum collection. *Bulletin of the Museum of Comparative Zoology* 6 (11): 167–172.
- Garman S. **1884**. New sharks. *Chlamydoselachus anguineus*. *Heptranchias pectorosus*. *Bulletin of the Essex Institute* 16: 3–15.
- Garman S. **1888**. On an eel from the Marshall Islands. [Also titled: An eel (*Rhinomuraena quaesita*) from the Marshall Islands.] *Bulletin of the Essex Institute* 20: 114–116.
- Garman S. 1899. XXVI The fishes. In: Reports on an exploration off the west coasts of Mexico, Central and South America, and off the Galapagos Islands, in charge of Alexander Agassiz, by the the U.S. Fish Commission steamer *Albatross*, during 1891, Lieut. Commander Z.L. Tanner, U.S.N., commanding. *Memoirs of the Museum of Comparative Zoology* 24: 431 pp + 97 Pls.
- Garman S. **1901**. Genera and families of the chimaeroids. *Proceedings of the New England Zoölogical Club* 2: 75–77.
- Garman S. **1903**. Some fishes from Australasia. *Bulletin of the Museum of Comparative Zoology* **39**(8): 299–241.
- Garman S. **1906**. New Plagiostomia. *Bulletin of the Museum of Comparative Zoology* 46(11): 203–208.

- Garman S. **1908**. New Plagiostomia and Chismopnea. *Bulletin* of the Museum of Comparative Zoology 51(9): 249–256.
- Garman S. **1913**. The Plagiostomia (sharks, skates, and rays). *Memoirs of the Museum of Comparative Zoology* 36: xiii + 515 pp.
- Garpe KC, Öhman MC. **2003**. Coral and fish distribution patterns in Mafia Island Marine Park, Tanzania: fish-habitat interactions. *Hydrobiologia* 498: 191–211.
- Garrett A. **1863**. Descriptions of new species of fishes. *Proceedings of the California Academy of Sciences* (Series 1) 3: 63–67.
- Garrett A. 1864. Descriptions of new species of fishes No. II. *Proceedings of the California Academy of Sciences* (Series 1) 3: 103–107.
- Garrick JAF. **1957**. Studies on New Zealand Elasmobranchii. Part VI. Two new species of *Etmopterus* from New Zealand. *Bulletin of the Museum of Comparative Zoology* 116(3): 169–190.
- Garrick JAF. **1959**. Studies on New Zealand Elasmobranchii. Part VIII. Two northern hemisphere species of *Centroscymnus* in New Zealand waters. *Transactions of the Royal Society of New Zealand* 87(1): 75–89.
- Garrick JAF. **1960**. Studies on New Zealand Elasmobranchii. Part X. The genus *Echinorhinus*, with an account of a second species *E. cookei* Pietschmann, 1928, from New Zealand waters. *Transactions of the Royal Society of New Zealand* 88(1): 105–117.
- Garrick JAF. **1960**. Studies on New Zealand Elasmobranchii. Part XI. Squaloids of the genera *Deania*, *Etmopterus*, *Oxynotus* and *Dalatias* in New Zealand waters. *Transactions of the Royal Society of New Zealand* 88(3): 489–517.
- Garrick JAF. **1960**. Studies on New Zealand Elasmobranchii. Part XII. The species of *Squalus* from New Zealand and Australia; and a general account and key to the New Zealand Squaloidea. *Transactions of the Royal Society of New Zealand* 88(3): 519–557.
- Garrick JAF. **1967**. Revision of sharks of genus *Isurus* with description of a new species (Galeoidea, Lamnidae). *Proceedings of the United States National Museum* 118(3537): 663–690.
- Garrick JAF. **1971**. *Harriotta raleighana*, a long-nosed chimaera (family Rhinochimaeridae), in New Zealand waters. *Journal of the Royal Society of New Zealand* 1(3/4): 203–213.
- Garrick JAF. **1974**. First record of an odontaspidid shark in New Zealand waters. *New Zealand Journal of Marine and Freshwater Research* 8(4): 621–630.
- Garrick JAF. **1982**. Sharks of the genus *Carcharhinus*. *NOAA Technical Report* NMFS Circular 445: 1–194.

Garrick JAF. **1985**. Additions to a revision of the shark genus *Carcharhinus*: synonymy of *Aprionodon* and *Hypoprion*, and description of a new species of *Carcharhinus*. *NOAA Technical Report* NMFS 34: 1–26.

Garrick JAF, Inada T. **1975**. Dimensions of long-nosed chimaera *Harriotta raleighana* from New Zealand. *New Zealand Journal of Marine and Freshwater Research* 9(2): 159–167.

Garrick JAF, Paul LJ. **1971**. *Heptranchias dakini* Whitley, 1931, a synonym of *H. perlo* (Bonnaterre, 1788), the sharpsnouted sevengill or perlon shark, with notes on sexual dimorphism in this species. *Zoological Publications from Victoria University of Wellington* 153: 1–14.

Garrick JAF, Springer VG. **1964**. *Isistius plutodus*, a new squaloid shark from the Gulf of Mexico. *Copeia* 1964(4): 678–682.

Garstang W. **1931**. The phyletic classification of Teleostei. *Proceedings of the Leeds Philosophical and Literary Society* (Scientific Section) 2: 240–260.

Gaudant M. **1978**. Contribution à une révision des poissons crétacés du Jebel Tselfat (Rides prérifaines, Maroc). *Notes du Service Geologique du Maroc* 39: 79–124.

Geevarghese C, John PA. **1983**. A new goby, *Glossogobius minutus* (Teleostei: Gobiidae) from the south-west coast of India. *Journal of Fish Biology* 22(2): 231–240.

Gehringer JW. **1956**. Observations on the development of the Atlantic sailfish *Istiophorus americanus* (Cuvier) with notes on an unidentified species of istiophorid. *Fishery Bulletin* 57(110): 139–171.

Gehringer JW. **1970**. Young of the Atlantic sailfish *Istiophorus platypterus*. *Fishery Bulletin* 68(2): 177–189.

Gell FR, Whittington MW. **2002**. Diversity of fishes in seagrass beds in the Quirimba Archipelago, northern Mozambique. *Marine and Freshwater Research* 53(2): 115–121.

Geoffroy Saint Hilaire E. 1809, 1817. Poissons du Nil, de la mer Rouge et de la Méditerranée. In: Description de l'Egypte ou recueil des observations et des recherches qui ont été faites en Égypte pendant l'expedition de l'Armée français (Imprimerie Impériale). Paris. Histoire Naturelle v. 1 (part 1): 1–52, 265–310; 311–340; Poissons Pls. 1–17 [1809]; Pls. 18–27 [1817].

George CJ. **1972**. Notes on the breeding and movements of the rabbitfishes, *Siganus rivulatus* Forsskål and S. *luridus* Rüppell, in the coastal waters of Lebanon. *Annali del Museo Civico di Storia Naturale Genova* 79: 32–44.

George KC. **1970**. Notes on two species of flat-heads (Platycephalidae: Pisces) from the trawl grounds of the south-west coast of India. *Journal of the Marine Biological Association of India* 10(2): 354–356.

Gerlach J. **2001**. A first Seychelles record of the ghost-pipefish *Solenostomus cyanopterus*. *Phelsuma* (Mahé) 9: 75–76.

Gerlach T, Sprenger D, Michiels NK. **2014**. Fairy wrasses perceive and respond to their deep red fluorescent coloration. *Proceedings of the Royal Society* B 281: 20140787. https://doi.org/10.1098/rspb.2014.0787

Gess RW, Coates MI, Rubidge BS. **2006**. A lamprey from the Devonian period of South Africa. *Nature* 443: 981–984.

Ghanbarifardi M, Malek M. **2009**. Distribution, diversity, and abundance of rocky intertidal fishes in the Persian Gulf and Gulf of Oman, Iran. *Marine Biology Research* 5(5): 496–502. http://dx.doi.org/10.1080/17451000802441293

Ghasemzadeh J. **1998**. Phylogeny and systematics of Indo-Pacific mullets (Teleostei: Mugilidae) with special reference to the mullets of Australia. PhD thesis, Macquarie University, Sydney, Australia. 397 pp.

Ghasemzadeh J, Ivantsoff W, Aarn. **2004**. Historical overview of mugilid systematics, with description of *Paramugil* (Teleostei: Mugiligormes: Mugilidae), new genus. *aqua*, *Journal of Ichthyology and Aquatic Biology* 8(1): 9–22.

Gibbs RH Jr. **1960**. *Alepisaurus brevirostris*, a new species of lancet fish from the western North Atlantic. *Breviora* 123: 1–14.

Gibbs RH Jr. **1981**. Exocoetidae. Flyingfishes. In: Fischer W, Bianchi G, Scott WB (eds) *FAO species identification sheets for fishery purposes. Eastern central Atlantic (Fishing Area 34 and part of 47)*. Vol. 2. Rome: FAO. [unpaginated]

Gibbs RH Jr, Collette BB. **1959**. On the identification, distribution and biology of the dolphins, *Coryphaena hippurus* and *C. equiselis*. *Bulletin of Marine Science of the Gulf and Caribbean* 9(2): 117–152.

Gibbs RH Jr, Collette BB. **1967**. Comparative anatomy and systematics of the tunas, genus *Thunnus*. *Fishery Bulletin* 66(1): 65–130.

Gibbs RH Jr, Staiger JC. **1970**. Eastern tropical Atlantic flyingfishes of the genus *Cypselurus* (Exocoetidae). *Studies in Tropical Oceanography* 4(2): 432–466.

Gibbs RH Jr, Wilimovsky NJ. **1966**. Family Alepisauridae. In: Fishes of the western North Atlantic. *Memoir of the Sears Foundation for Marine Research* 1(part 5): 482–497.

Gibson RN. **2005**. The behaviour of flatfishes (pp. 213–239). In: Gibson RN (ed) *Flatfishes: Biology and exploitation*. Oxford: Blackwell Science. xxiv + 391 pp.

Gibson RN (ed). **2005**. *Flatfishes: Biology and exploitation*. Oxford: Blackwell Science. xxiv + 391 pp.

Gibson RN, Nash RDM, Geffen AJ, van der Veer HW (eds). 2015. Flatfishes: Biology and exploitation (2<sup>nd</sup> edition). Oxford: Wiley-Blackwell. 576 pp. https://doi. org/10.1002/9781118501153

Giglioli EH. **1880**. *Elenco dei Mammiferi, degli Uccelli e dei Rettili ittiofagi appartenenti alla Fauna Italiana, e Catalogo degli Anfibie dei Pesci Italiani in Catalogo Sezione italiana* (Pisces: pp. 18–55). Firenze: Stamperia Reale. 55 pp. Giglioli EH. **1882**. New and very rare fish from the Mediterranean. *Nature* (London) 25: 535.

Giglioli EH. **1883**. Zoology at the Fisheries Exhibition. II. Notes on the Vertebrata. *Nature* (London) 28: 313–316.

Giglioli EH. **1889**. Note intorno agli animali vertebrati raccolti dal Conte Augusto Boutourline e dal D. Leopoldo Traversi ad Assab e nello Scioa negli anni 1884–87. *Annali del Museo Civico di Storia Naturale di Genova* (Serie 2) 6: 1–73.

Gilbert CH. **1890**. A preliminary report on the fishes collected by the steamer *Albatross* on the Pacific coast of North America during the year 1889, with descriptions of twelve new genera and ninety-two new species. *Proceedings of the United States National Museum* 13(797): 49–126.

Gilbert CH. **1892**. Descriptions of apodal fishes from the tropical Pacific. In: Scientific results of explorations by the U.S. Fish Commission steamer *Albatross. Proceedings of the United States National Museum* 14(856): 347–352.

Gilbert CH. **1905**. The deep-sea fishes of the Hawaiian Islands. In: The aquatic resources of the Hawaiian Islands. *Bulletin of the U.S. Fish Commission* 23(pt 2) [for 1903]: 577–713.

Gilbert CH, Cramer F. **1897**. Report on the fishes dredged in deep water near the Hawaiian Islands, with descriptions and figures of twenty-three new species. *Proceedings of the United States National Museum* 19(1114): 403–435, Pls. 36–48.

Gilbert CR. **1967**. A revision of the hammerhead sharks (family Sphyrnidae). *Proceedings of the United States National Museum* 119(3539): 1–88.

Gilchrist JDF. **1902**. Catalogue of fishes recorded from South Africa. *Marine Investigations in South Africa* 1: 97–179.

Gilchrist JDF. **1903**. South African fishes. *Marine Investigations in South Africa* 2: 101–113.

Gilchrist JDF. **1903**. Descriptions of new South African fishes. *Marine Investigations in South Africa* 2[1904]: 203–211.

Gilchrist JDF. **1904**. Descriptions of new South African fishes. *Marine Investigations in South Africa* 3[1905]: 1–16.

Gilchrist JDF. **1906**. Descriptions of fifteen new South African fishes, with notes on other species. *Marine Investigations in South Africa* 4[1908]: 143–171, Pls. 37–51.

Gilchrist JDF. **1909**. Descriptions of fishes from the coast of Natal (part 2). *Annals of the South African Museum* 6(3): 15–279.

Gilchrist JDF. **1913**. Description of a new species of stingray (*Trygon*) from South Africa. *Transactions of the Royal Society of South Africa* 3(1): 33–34.

Gilchrist JDF. **1913**. Review of the South African Clupeidae (herrings) and allied families of fishes. *Marine Biological Report* 1: 46–66.

Gilchrist JDF. **1914**. Observations on the habits of some South African fishes. *Marine Biological Report* 2: 90–115.

Gilchrist JDF. **1914**. The snoek and allied fishes in South Africa. *Marine Biological Report* 2: 116–127.

Gilchrist JDF. **1914**. Description of three new South African fishes. *Marine Biological Report* 2: 128–131.

Gilchrist JDF. **1916**. Eggs and larvae of Cape fishes. *Marine Biological Report* 3: 1–26.

Gilchrist JDF. **1916**. Note on protective resemblance, etc., in post-larval stages of some Cape fishes. *Transactions of the Royal Society of South Africa* 6(3): 205–208.

Gilchrist JDF. **1918**. Note on eggs and embryos of the South African myxinoid, *Bdellostoma (Heptatretus) hexatrema*, Müll. *Journal of Cell Science* s2-63(249): 141–159.

Gilchrist JDF. **1918**. The eggs and spawning habits of the pilot fish (*Naucrates ductor*). *Annals and Magazine of Natural History* (Series 9) 2(7): 114–118.

Gilchrist JDF. **1922**. Deep-sea fishes procured by the S.S. *Pickle* (Part I). *Union of South Africa Fisheries and Marine Biological Survey Report* 2(art.3): 41–79. [Also as separate, pp. 1–39]

Gilchrist JDF. **1922**. Observations on living fishes brought by H.M.S. *Challenger* from tropical East Africa to Cape waters. *Transactions of the Royal Society of South Africa* 10(1): 23–26.

Gilchrist JDF, Hunter H. **1919**. Reproduction of fishes in Table Bay. *Transactions of the Royal Society of South Africa* 8: 1–16.

Gilchrist JDF, Thompson WW. **1908**. The Blenniidae of South Africa. *Annals of the South African Museum* 6(pt 2): 97–143.

Gilchrist JDF, Thompson WW. **1908**. Descriptions of fishes from the coast of Natal. *Annals of the South African Museum* 6(pt 2): 145–206.

Gilchrist JDF, Thompson WW. **1909**. Descriptions of fishes from the coast of Natal (Part 2). *Annals of the South African Museum* 6(pt 3): 213–279.

Gilchrist JDF, Thompson WW. **1911**. Description of fishes from the coast of Natal (Part 3). *Annals of the South African Museum* 11(pt 2): 29–58.

Gilchrist JDF, Thompson WW. **1911**. Descriptions of three new species of freshwater fishes from South Africa. *Annals and Magazine of Natural History* (Series 8) 7(41): 477–478.

Gilchrist JDF, Thompson WW. **1914**. Descriptions of fishes from the coast of Natal (Part 4). *Annals of the South African Museum* 13(pt 3): 65–95.

Gilchrist JDF, Thompson WW. **1916**. A catalogue of the sea fishes recorded from Natal, Part I. *Annals of the Durban Museum* 1(3): 255–290.

Gilchrist JDF, Thompson WW. **1916**. Description of four new South African fishes. *Marine Biological Report* 3: 56–61.

Gilchrist JDF, Thompson WW. **1917**. A catalogue of the sea fishes recorded from Natal, Part 2. *Annals of the Durban Museum* 1(4): 291–431.

Gilchrist JDF, Thompson WW. **1917**. The freshwater fishes of South Africa. *Annals of the South African Museum* 11(pt 6): 465–575.

Gilchrist JDF, Von Bonde C. **1923**. The Stromateidae (butterfishes) collected by the S.S. *Pickle. Union of South Africa Fisheries and Marine Biological Survey Special Report* 4: 1–12.

Gilchrist JDF, Von Bonde C. **1924**. Deep-sea fishes procured by the S.S. *Pickle* (Part II). *Union of South Africa Fisheries and Marine Biological Survey Report* 3(7): 1–24, Pls. 1–6.

Gill AC. **1998**. *Hetereleotris georgegilli*, a new species of gobiid fish, with notes on other Mauritian *Hetereleotris* species. *Bulletin of the Natural History Museum London (Zoology)* 64(1): 91–95.

Gill AC. **1999**. Subspecies, geographic forms and widespread Indo-Pacific coral-reef fish species: a call for change in taxonomic practice (pp. 79–87). In: Séret B, Sire J-Y (eds) *Proceedings of the 5<sup>th</sup> Indo-Pacific Fish Conference, Nouméa, New Caledonia, 3–8 November 1997*. Paris: Société Française d'Ichtyologie. 866 pp.

Gill AC. **2004**. Revision of the Indo-Pacific dottyback fish subfamily Pseudochrominae (Perciformes: Pseudochromidae). *Smithiana* Monograph 1: 1–214.

Gill AC, Bogorodsky SV, Mal AO. **2013**. *Acanthoplesiops cappuccino*, a new species of acanthoclinine fish from the Red Sea (Teleostei: Plesiopidae). *Zootaxa* 3750(3): 216–222. https://doi.org/10.11646/zootaxa.3750.3.2

Gill AC, Bogorodsky SV, Mal AO. **2014**. *Gymnoxenisthmus tigrellus*, new genus and species of gobioid fish from the Red Sea (Gobioidei: Xenisthmidae). *Zootaxa* 3755(5): 491–495. http://dx.doi.org/10.11646/zootaxa.3755.5.9

Gill AC, Edwards AJ. **1999**. Monophyly, interrelationships and description of three new genera in the dottyback fish subfamily Pseudoplesiopinae (Teleostei: Perciformes: Pseudochromidae). *Records of the Australian Museum* 51(2–3): 141–160.

Gill AC, Edwards AJ. **2002**. Two new species of the Indo-Pacific fish genus *Pseudoplesiops* (Perciformes, Pseudochromidae, Pseudoplesiopinae). *Bulletin of the Natural History Museum London (Zoology)* 68(1): 19–26.

Gill AC, Edwards AJ. 2004. Revision of the Indian Ocean dottyback fish genera *Chlidichthys* and *Pectinochromis* (Perciformes: Pseudochromidae: Pseudoplesiopinae). *Smithiana, Publications in Aquatic Biodiversity* Bulletin 3: 1–47.

Gill AC, Fricke R. 2001. Revision of the Western Indian Ocean fish subfamily Anisochrominae (Perciformes, Pseudochromidae). *Bulletin of the Natural History Museum London (Zoology)* 67(2): 191–207.

Gill AC, Mee JKL. **1993**. Notes on dottyback fishes of the genus *Pseudochromis* of Oman, with description of a new species (Perciformes: Pseudochromidae). *Revue française d'Aquariologie Herpétologie* 20(2): 53–60.

Gill AC, Mooi RD. **1993**. Monophyly of the Grammatidae and of the Notograptidae, with evidence for their phylogenetic positions among perciforms. *Bulletin of Marine Science* 52: 327–350.

Gill AC, Randall JE. **1994**. *Chlidichthys cacatuoides*, a new species of pseudoplesiopine dottyback from southern Oman, with a diagnosis of the genus *Chlidichthys* Smith, and new record of *Pseudochromis punctatus* Kotthaus from Oman (Teleostei: Perciformes: Pseudochromidae). *Revue française d'Aquariologie Herpétologie* 21(1–2): 11–18.

Gill AC, Randall JE. **1994**. *Xenisthmus balius*, a new species of fish from the Persian Gulf (Gobioidei: Xenisthmidae). *Proceedings of the Biological Society of Washington* 107(3): 445–450.

Gill AC, Randall JE. 1997. Redescription of and lectotype designation for *Balistes macrolepis* Boulenger, 1887, a senior synonym of *Canthidermis longirostris* Tortonese, 1954 and *C. villosus* Fedoryako, 1979 (Teleostei, Tetraodontoformes, Balistidae). *Bulletin of the Natural History Museum London* (*Zoology*) 63(1): 27–31.

Gill AC, Zajonz U. **2003**. *Halidesmus socotraensis* new species and *Haliophis guttatus* (Forsskål), new records of congrogadine fishes from the Socotra Archipelago (Perciformes: Pseudochromidae). *Proceedings of the Biological Society of Washington* 116(1): 52–60.

Gill AC, Zajonz U. **2011**. Pseudochromine and pseudoplesiopine dottyback fishes from the Socotra Archipelago, Indian Ocean, with descriptions of two new species of *Pseudochromis* Rüppell (Perciformes: Pseudochromidae). *Zootaxa* 3106: 1–23. http://dx.doi.org/10.11646/zootaxa.3106.1.1

Gill TN. **1859**. Description of a new generic form of Gobinae from the Amazon River. *Annals of the Lycium of Natural History of New York* 7(1–3): 45–48.

Gill TN. **1859**. Description of *Hyporhamphus*, a new genus of fishes allied to *Hemirhamphus*, Cuv. *Proceedings of the Academy of Natural Sciences of Philadelphia* 11: 131.

Gill TN. **1859**. Notes on a collection of Japanese fishes, made by Dr. J. Morrow. *Proceedings of the Academy of Natural Sciences of Philadelphia* 11: 144–150.

Gill TN. **1859**. Description of a third genus of Hemirhamphinae. *Proceedings of the Academy of Natural Sciences of Philadelphia* 11: 155–157.

Gill TN. **1859**. Description of a new genus of Salarianae, from the West Indies. *Proceedings of the Academy of Natural Sciences of Philadelphia* 11: 168.

Gill TN. 1860. Conspectus piscium in expeditione ad oceanum Pacificum septentrionalem, C. Ringold et J. Rodgers ducibus, a Gulielmo Stimpson collectorum. Sicydianae. *Proceedings of the Academy of Natural Sciences of Philadelphia* 12: 100–102.

Gill TN. **1860**. Monograph of the genus *Labrosomus* Sw. *Proceedings of the Academy of Natural Sciences of Philadelphia* 12: 102–108.

Gill TN. **1861**. Synopsis generum Rhyptici et affinium. *Proceedings of the Academy of Natural Sciences of Philadelphia* 13: 52–54.

Gill TN. **1861**. [Descriptions of two new species of marine fishes.] *Proceedings of the Academy of Natural Sciences of Philadelphia* 13: 98–99.

Gill TN. **1861**. Notes on some genera of fishes of the western coast of North America. *Proceedings of the Academy of Natural Sciences of Philadelphia* 13: 164–168.

Gill TN. **1861**. Catalogue of the fishes of the eastern coast of North America from Greenland to Georgia. *Proceedings of the Academy of Natural Sciences of Philadelphia* 13(Suppl.): 1–63.

Gill TN. **1862**. Analytical synopsis of the Order of Squali; and revision of the nomenclature of the genera. *Annals of the Lyceum of Natural History of New York* 7(art.32): 367–413.

Gill TN. **1862**. Synopsis of the family of cirrhitoids. *Proceedings of the Academy of Natural Sciences of Philadelphia* 14: 102–124.

Gill TN. **1862**. On the limits and arrangement of the family of scombroids. *Proceedings of the Academy of Natural Sciences of Philadelphia* 14: 124–127.

Gill TN. **1862**. Catalogue of the fishes of Lower California in the Smithsonian Institution, collected by Mr. J. Xantus. *Proceedings of the Academy of Natural Sciences of Philadelphia* 14: 140–151.

Gill TN. **1862**. Remarks on the relations of the genera and other groups of Cuban fishes. *Proceedings of the Academy of Natural Sciences of Philadelphia* 14: 235–242.

Gill TN. **1862**. Notice of a collection of the fishes of California presented to the Smithsonian Institution by Mr. Samuel Hubbard. *Proceedings of the Academy of Natural Sciences of Philadelphia* 14: 274–282.

Gill TN. **1862**. Note on some genera of fishes of western North America. *Proceedings of the Academy of Natural Sciences of Philadelphia* 14: 329–332.

Gill TN. **1862**. Synopsis of carangoids of the eastern coast of North America. *Proceedings of the Academy of Natural Sciences of Philadelphia* 14: 430–443.

Gill TN. **1863**. On an unnamed generic type allied to Sebastes. Proceedings of the Academy of Natural Sciences of Philadelphia 15: 207–209.

Gill TN. **1863**. Descriptions of the genera of gadoid and brotuloid fishes of western North America. *Proceedings of the Academy of Natural Sciences of Philadelphia* 15: 242–254.

Gill TN. **1863**. Descriptions of the gobioid genera of the western coast of temperate North America. *Proceedings of the Academy of Natural Sciences of Philadelphia* 15: 262–267.

Gill TN. **1863**. On the gobioids of the eastern coast of the United States. *Proceedings of the Academy of Natural Sciences of Philadelphia* 15: 267–271.

Gill TN. **1863**. Note on the genera of Hemirhamphinae. *Proceedings of the Academy of Natural Sciences of Philadelphia*elphia 15: 272–273.

Gill TN. **1864**. Critical remarks on the genera of *Sebastes* and *Sebastodes* of Ayres. *Proceedings of the Academy of Natural Sciences of Philadelphia* 16: 145–147.

Gill TN. **1865**. On a new genus of Serraninae. *Proceedings of the Academy of Natural Sciences of Philadelphia* 17: 104–106.

Gill TN. **1865**. On a remarkable new type of fishes allied to *Nemophis. Annals of the Lyceum of Natural History of New York* 8: 138–141.

Gill TN. **1865**. On a new family type of fishes related to the blennioids. *Annals of the Lyceum of Natural History of New York* 8: 141–144

Gill TN. 1877. In: Streets TH.

Gill TN. **1879**. On the proper specific name of the common pelagic antennariid *Pterophryne*. *Proceedings of the United States National Museum* 1(32): 223–226.

Gill TN. **1883**. Supplementary note on the Pediculati. *Proceedings of the United States National Museum* 5(316): 551–556.

Gill TN. **1884**. Three new families of fishes added to the deepsea fauna in a year. *American Naturalist* **18**(4): 433.

Gill TN. **1884**. Synopsis of the genera of the superfamily Teuthidoidea (families Teuthididae and Siganidae). *Proceedings of the United States National Museum* 7(26–27): 275–281.

Gill TN. **1884**. Synopsis of the plectognath fishes. *Proceedings* of the United States National Museum 7(26–27): 411–427.

Gill TN. **1888**. Note on the genus *Dipterodon*. *Proceedings of the United States National Museum* 11(684): 67–68.

Gill TN. **1889**. On the classification of the mail-cheeked fishes. *Proceedings of the United States National Museum* 11(756): 567–592.

Gill TN. **1890**. Osteological characteristics of the family Muraenesocidae. *Proceedings of the United States National Museum* 13(815): 231–234.

Gill TN. **1892**. On the genera *Labrichthys* and *Pseudolabrus*. *Proceedings of the United States National Museum* 14(1981): 395–404.

Gill TN. **1893**. A comparison of antipodal faunas. *Memoirs of the National Academy of Sciences* 6 (5<sup>th</sup> memoir): 91–124.

Gill TN. **1905**. Note on the genera of Synanceine and Pelorine fishes. *Proceedings of the United States National Museum* 28(1394): 221–225.

Gill TN, Ryder JA. **1883**. Diagnoses of new genera of Nemichthyoid eels. *Proceedings of the United States National Museum* 6(381): 260–262. Gillibrand CJ, Harris AR, Mara E. **2007**. Inventory and spatial assemblage study of reef fish in the area of Andavadoaka, south-west Madagascar (Western Indian Ocean). *Western Indian Ocean Journal of Marine Science* 6(2): 183–197.

Gilmore RG, Dodrill JW, Linley PA. **1983**. Reproduction and embryonic development of the sand tiger shark, *Odontaspis taurus* (Rafinesque). *Fishery Bulletin* 81(2): 201–225.

Giltay L. **1928**. Notes ichthyologiques. II. Une espèce nouvelle de *Rhinobatus* du Congo Belge (*Rhinobatus congolensis*, nov. sp.). *Annales de la Société Royale Zoologique de Belgique* 59: 21–27.

Giltay L. **1933**. Résultats scientifiques du voyage aux Indes Orientales Néerlandaises de LL. AA. RR. le Prince et la Princesse Léopold de Belgique. Poissons. *Mémoires du Musée Royal d'Histoire Naturelle de Belgique* 5(3): 1–129.

Giltay L. **1934**. Notes ichthyologiques. VIII. Les larves de Schindler sont-elles des Hemirhamphidae? *Bulletin du Musée Royal d'Histoire Naturelle de Belgique* 10(13): 1–10.

Ginsberg I. **1953**. Western Atlantic scorpion-fishes. Smithsonian Miscellaneous Collections 121(8): 1–103.

Giorna ME. **1809**. Mémoire sur des poissons d'espèces nouvelles et du genres nouveaux. *Mémoires de l'Académie Imperiale de Torino* 2: 177–180.

Girard C. **1854**. Descriptions of new fishes, collected by Dr. A.L. Heermann, naturalist attached to the survey of the Pacific Railroad Route, under Lieut. R.S. Williamson, U.S.A. *Proceedings of the Academy of Natural Sciences of Philadelphia* 7: 129–165.

Girard CF. **1855**. Characteristics of some cartilaginous fishes of the Pacific coast of North America. *Proceedings of the Academy of Natural Sciences of Philadelphia* 7(6): 196–197.

Girard CF. **1859**. Ichthyological notices. *Proceedings of the Academy of Natural Sciences of Philadelphia* 11: 56–68.

Gistel J. **1848**. *Naturgeschichte des Tierreichs für höhere Schulen*. Stuttgart. xvi + 216 pp.

Gloerfelt-Tarp T, Kailola PJ. **1984**. *Trawled fishes of southern Indonesia and northwestern Australia*. The Australian Development Assistance Bureau, Directorate General of Fishes (Indonesia), German Agency for Technical Cooperation. xiii + 406 pp.

Glover CN, Bucking C, Wood CM. 2011. Adaptations to *in situ* feeding: novel nutrient acquisition pathways in an ancient vertebrate. *Proceedings of the Royal Society* B 278: 3096–3101. https://doi.org/10.1098/rspb.2010.2784

Gmelin JF. 1789. Pisces (1126–1516). In: Linné C (ed) Systema Naturae per regna tria naturae, secumdum classes, ordines, genera, species; cum characteribus, differentiis, synonymis, locis. Editio decima tertia, aucta, reformata 1(part 3) (pp. 1033–1516). Lipsiae: Georg Emanuel Beer.

Godkin C, Winterbottom R. **1985**. Phylogeny of the family Congrogadidae (Pisces; Perciformes) and its placement as a subfamily of the Pseudochromidae. *Bulletin of Marine*  *Science* 36(3): 633–671.

Gohar HAF, Mazhar FM. **1964**. The elasmobranchs of the north-western Red Sea. *Publications of the Marine Biological Station Al-Ghardaqa* (Red Sea) 13: 1–144.

Golani D. **1984**. *Sargocentron inacrosquamis* a new squirrelfish from the Amirantes Islands and the Red Sea (Holocentridae, Pisces). *Cybium* 8(2): 39–43.

Golani D. **1984**. The squirrelfish *Ostichthys hypsipterygion sufensis* a new subspecies from the Red Sea (Holocentridae: Pisces). *Cybium* 8(3): 97–102.

Golani D. **1993**. The biology of the Red Sea migrant, *Saurida undosquamis*, in the Mediterranean and comparison with the indigenous confamilial *Synodus saurus* (Teleostei: Synodontidae). *Hydrobiologia* 271: 109–117.

Golani D. **1996**. The marine ichthyofauna of the eastern Levant – history, inventory, and characterization. *Israel Journal of Zoology* 42(1): 15–55.

Golani D. **1998**. Distribution of Lessepsian migrant fish in the Mediterranean. *Italian Journal of Zoology* 65(S1): 95–99.

Golani D. **2001**. *Upeneus davidaromi*, a new deepwater goatfish (Osteichthyes, Mullidae) from the Red Sea. *Israel Journal of Zoology* 47(2): 111–121.

Golani D. **2002**. The Indo-Pacific striped eel catfish, *Plotosis lineatus* (Thunberg, 1787), (Osteichthyes: Siluriformes), a new record from the Mediterranean. *Scientia Marina* 66(3): 321–323.

Golani D. **2006**. An annotated list of types in the Hebrew University fish collection. *Haasiana* (newsletter of the biological collections of the Hebrew University) 2006(3): 20–40.

Golani D, Appelbaum-Golani B, Gon O. 2008. Apogon smithi (Kotthaus, 1970) (Teleostei: Apogonidae), a Red Sea cardinalfish colonizing the Mediterranean Sea. Journal of Fish Biology 72(6): 1534–1538. http://dx.doi.org/10.1111/j.1095-8649.2008.01812.x

Golani D, Ben-Tuvia A. **1990**. Two Red Sea flatheads (Platycephalidae) immigrants in the Mediterranean. *Cybium* 14(1): 57–61.

Golani D, Bernardi G. **2012**. Differential invading potential among cryptic species of a Lessepsian bioinvader, the blotchfin dragonet *Callionymus filamentosus*. *Marine Ecology Progress Series* 450: 159–166. http://dx.doi. org/10.3354/meps09575

Golani D, Bogorodsky SV. **2010**. The fishes of the Red Sea – reappraisal and updated checklist. *Zootaxa* 2463: 1–135. http://dx.doi.org/10.11646/zootaxa.2463.1.1

Golani D, Diamant A. **1991**. Biology of the sweeper, *Pempheris vanicolensis* Cuvier and Valenciennes, a Lessepsian migrant in the eastern Mediterranean, with a comparison with the original Red Sea population. *Journal of Fish Biology* 38(6): 819–827.

Golani D, Fine M. **2002**. On the occurrence of *Hippocampus fuscus* in the eastern Mediterranean. *Journal of Fish Biology* 60(3): 764–766.

Golani D, Fricke R. **2018**. Checklist of the Red Sea fishes with delineation of the Gulf of Suez, Gulf of Aqaba, endemism and Lessepsian migrants. *Zootaxa* 4509(1): 1–215. http://dx.doi.org/10.11646/zootaxa.4509.1.1

Golani D, Fricke R, Appelbaum-Golani B. 2011. First record of the Indo-Pacific slender ponyfish *Equulites elongatus* (Günther, 1874) (Perciformes: Leiognathidae) in the Mediterranean. *Aquatic Invasions* 6(Suppl. 1): S75–S77. http://dx.doi.org/10.3391/ai.2011.6.S1.017

Golani D, Fricke R, Tikochinski Y. **2013**. *Sillago suezensis*, a new whiting from the northern Red Sea, and status of *Sillago erythraea* Cuvier (Teleostei: Sillaginidae). *Journal of Natural History* 2013: 413–468. https://doi.org/10.1080.0022 2933.2013.800609

Golani D, Sonin O. 1992. New records of the Red Sea fishes, *Pterois miles* (Scorpaenidae) and *Pteragogus pelycus* (Labridae) from the eastern Mediterranean Sea. *Japanese Journal of Ichthyology* 39(2): 167–169.

Golovan GA. **1976**. Rare and first finds of cartilaginous (Chondrichthyes) and bony (Osteichthyes) fishes from the continental slope off West Africa. *Transactions of the P.P. Shirshov Institute of Oceanology* 104: 277–317.

Gomon JR. 1984. Plotosidae. Stinging catfishes, coral reef catfishes, eel catfishes, barbel eels. In: Fischer W, Bianchi G (eds) FAO species identification sheets for fishery purposes. Western Indian Ocean (Fishing Area 51). Vol. 3. Rome: FAO. [unpaginated]

Gomon JR, Taylor WR. **1982**. *Plotosus nkunga*, a new species of catfish from South Africa, with a redescription of *Plotosus limbatus* Valenciennes and key to the species of *Plotosus* (Siluriformes: Plotosidae). *J.L.B. Smith Institute of Ichthyology Special Publication* 22: 1–16.

Gomon MF. **1997**. Relationships of fishes of the labrid tribe Hypsigenyini. *Bulletin of Marine Science* 60(3): 789–871.

Gomon MF. **2006**. A revision of the labrid fish genus *Bodianus* with descriptions of eight new species. *Records of the Australian Museum, Supplement* 30: 1–133.

Gomon MF. **2007**. A new genus and miniature species of pipehorse (Syngnathidae) from Indonesia. *aqua*, *International Journal of Ichthyology* 13(1): 25–30.

Gomon MF. 2008. Family Rhinochimaeridae. Longnose chimaeras (pp. 148–149); Family Uranoscopidae. Stargazers (pp. 678–683). In: Gomon MF, Bray DJ, Kuiter RH (eds) *Fishes of Australia's southern coast*. Sydney, Australia: New Holland Publishers. 928 pp.

Gomon MF, Bray DJ, Kuiter RH (eds). **2008**. *Fishes of Australia's southern coast*. Sydney, Australia: New Holland Publishers. 928 pp. Gomon MF, Glover JCM, Kuiter RH (eds). **1994**. *The fishes of Australia's south coast*. Adelaide: State Print. 992 pp.

Gomon MF, Ho H-C. **2008**. Family Chaunacidae. Coffinfishes, sea toads (pp. 377–379). In: Gomon MF, Bray DJ, Kuiter RH (eds) *Fishes of Australia's southern coast*. Sydney, Australia: New Holland Publishers. 928 pp.

Gomon MF, Johnson JW. **1999**. A new fringed stargazer (Uranoscopidae: *Ichthyscopus*) with descriptions of the other Australian species. *Memoirs of the Queensland Museum* 43(2): 597–619.

Gomon MF, Kuiter RH. **2009**. Two new pygmy seahorses (Teleostei: Syngnathidae: *Hippocampus*) from the Indo-West Pacific. *aqua, International Journal of Ichthyology* 15(1): 37–44.

Gomon MF, Madden WD. **1981**. Comments on the labrid fish subgenus *Bodianus* (*Trochocopus*) with a description of a new species from the Indian and Pacific oceans. *Revue française d'Aquariologie Herpétologie* 7(4): 121–126.

Gon O. **1986**. *Apogon bifasciatus* Rüppell 1838, a junior synonym of *Apogon taeniatus* Ehrenberg 1828, and description of *Apogon pseudotaeniatus* n. sp. (Pisces: Apogonidae). *Senckenbergiana Biologica* 67(1/3): 5–17.

Gon O. **1987**. Redescription of *Apogon (Ostorhinchus) fleurieu* (Lacepède, 1802) with notes on its synonymy. *Japanese Journal of Ichthyology* 34(2): 138–145.

Gon O. **1993**. Revision of the cardinalfish genus *Cheilodipterus* (Perciformes: Apogonidae), with description of five new species. *Indo-Pacific Fishes* 22: 1–59.

Gon O. **1995**. Revision of the cardinalfish subgenus *Lepidamia* (Perciformes, Apogonidae, *Apogon*). *Israel Journal of Zoology* 41: 1–22.

Gon O. **1997**. Revision of the cardinalfish subgenus *Jaydia* (Perciformes, Apogonidae, *Apogon*). *Transactions of the Royal Society of South Africa* 51(1996): 147–194.

Gon O. **2000**. The taxonomic status of the cardinalfish species *Apogon niger, A. nigripinnis, A. pharaonis, A. sialis,* and related species (Perciformes: Apogonidae). *J.L.B. Smith Institute of Ichthyology Special Publication* 65: 1–20.

Gon O, Allen GR. **2012**. Revision of the Indo-Pacific cardinalfish genus *Siphamia* (Perciformes: Apogonidae). *Zootaxa* 3294: 1–84. http://dx.doi.org/10.11646/ zootaxa.3294.1.1

Gon O, Bogorodsky SV. **2010**. The cardinalfish *Fowleria isostigma* in the Red Sea and the validity of *F. punctulata* (Perciformes: Apogonidae). *Zootaxa* 2677: 27–37. http://dx.doi.org/10.11646/zootaxa.2677.1.3

Gon O, Bogorodsky SV, Mal AO. **2013**. Description of a new species of the cardinalfish genus *Pseudamiops* (Perciformes, Apogonidae) from the Red Sea. *Zootaxa* 3701(1): 93–100. https://doi.org/10.11646/zootaxa.3701.1.8

Gon O, Golani D. **2002**. A new species of the cardinalfish genus *Gymnapogon* (Perciformes, Apogonidae) from the Red Sea. *Ichthyological Research* 49(4): 346–349.

Gon O, Gouws G, Mwaluma J, Mwale M. 2013. Re-description of two species of the cardinalfish genus *Archamia* (Teleostei: Apogonidae) from the Red Sea and Western Indian Ocean. *Zootaxa* 3608(7): 587–594. http://dx.doi.org/10.11646/ zootaxa.3608.7.5

Gon O, Heemstra PC (eds). **1990**. *Fishes of the Southern Ocean*. Grahamstown: J.L.B. Smith Institute of Ichthyology. xviii + 462 pp.

Gon O, Klages NTW. **1988**. The marine fishes fauna of the sub-Antarctic Prince Edward Islands. *South African Journal of Antarctic Research* 8(2): 32–54.

Gon O, Randall JE. **1995**. Descriptions of three new species of the cardinalfish genus *Archamia* (Perciformes; Apogonidae). *Israel Journal of Zoology* 41: 539–550.

Gon O, Randall JE. **2003**. A review of the cardinalfishes (Perciformes: Apogonidae) of the Rea [sic] Sea. *Smithiana*, *Publications in Aquatic Biodiversity* Bulletin 1: 1–46.

Gonzales BJ, Okamura O, Nakamura K, Miyahara H. **1994**. New record of the annular sole, *Synaptura annularis* (Soleidae, Pleuronectiformes) from Japan. *Japanese Journal of Ichthyology* 40(4): 491–494.

Goode GB. **1881**. Fishes from the deep water on the south coast of New England obtained by the United States Fish Commission in the summer of 1880. *Proceedings of the United States National Museum* 3(177): 467–486.

Goode GB, Bean TH. **1882**. *Benthodesmus*, a new genus of deep-sea fishes, allied to *Lepidopus*. *Proceedings of the United States National Museum* 4(241): 379–383.

Goode GB, Bean TH. 1883. Reports on the results of dredging under the supervision of Alexander Agassiz, on the east coast of the United States, during the summer of 1880, by the U.S. Coast Survey steamer *Blake*, Commander J.R. Bartlett, U.S.N., commanding. XIX: Report on the Fishes. *Bulletin of the Museum of Comparative Zoology* 10(5): 183–226.

Goode GB, Bean TH. **1885**. Descriptions of new fishes obtained by the United States Fish Commission mainly from deep water off the Atlantic and Gulf coasts. *Proceedings of the United States National Museum* 8(543): 589–605.

Goode GB, Bean TH. 1886. Reports on the results of dredging, under the supervision of Alexander Agassiz, in the Gulf of Mexico (1877–78) and in the Caribbean Sea (1879–80), by the U.S. Coast Survey Steamer *Blake*, Lieut.-Commander C.D. Sigsbee, U.S.N., and Commander J.R. Bartlett, U.S.N., commanding. XXVIII. Description of thirteen species and two genera of fishes from the *Blake* collection. *Bulletin of the Museum of Comparative Zoology* 12(5): 153–170.

Goode GB, Bean TH. **1895**. On *Harriotta*, a new type of chimaeroid fish from the deeper waters of the northwestern Atlantic. In: Scientific results of exploration by the U.S. Fish Commission Steamer *Albatross. Proceedings of the United States National Museum* 17(1014): 471–473.

Goode GB, Bean TH. **1896**. Oceanic ichthyology, a treatise on the deep-sea and pelagic fishes of the world, based chiefly upon the collections made by steamers *Blake*, *Albatross* and *Fish Hawk* in the northwestern Atlantic. *Special Bulletin of the United States National Museum* 2: xxxv + 553 pp. [also cited as 1895]

Gorbman A. 1983. Reproduction in cyclostome fishes and its regulation (pp. 1–29). In: Hoar WS, Randall DJ, Donaldson EM (eds) *Fish Physiology* (Series). Vol. 9. Reproduction. Part A. Endocrine tissues and hormones. New York: Academic Press. 483 pp.

Gorbman A. **1997**. Hagfish development. *Zoological Science* 14(3): 375–390.

Gorbunova NN. **1976**. The classification and distribution of the larvae of Indo-Pacific species of billfishes from the family Istiophoridae. *Journal of Ichthyology* 16(3): 437–451.

Gorbunova NN, Parin NV. **1963**. Development of eggs and larvae of the flying fish, *Cheilopogon (Ptenichthys) unicolor* (Cuv. et Val.) (Pisces, Exocoetidae). *Trudy Instituta Okeanologii* 62: 62–67. [In Russian, English summary]

Gordon AL, Lutjeharms JR, Gründlingh ML. 1987.
Stratification and circulation at the Agulhas Retroflection.
Deep Sea Research Part A. Oceanographic Research Papers 34(4): 565–599.

Goreau T, McClanahan T, Hayes R, Strong A. **2000**. Conservation of coral reefs after the 1998 global bleaching event. *Conservation Biology* 14: 5–15.

Goren M. **1978**. Comparative study of *Bathygobius fuscus* (Rüppell) and related species of the Red Sea, including *B. fishelsoni* n. sp. (Pisces: Gobiidae). *Senckenbergiana Biologica* 58(5/6): 267–273.

Goren M. **1978**. A new gobiid genus and seven new species from Sinai coasts (Pisces; Gobiidae). *Senckenbergiana Biologica* 59(3/4): 191–203.

Goren M. **1979**. A new gobioid species *Coryogalops sufensis* from the Red Sea (Pisces, Gobiidae). *Cybium* (Série 3) 3(6): 91–95.

Goren M. **1979**. On the occurrence of *Acentrogobius belissimus* (Pisces: Gobiidae) in the Red Sea. *Israel Journal of Zoology* 28(2–3): 160–162.

Goren M. **1979**. Red Sea fishes assigned to the genus *Callogobius* Bleeker with a description of a new species (Teleostei: Gobiidae). *Israel Journal of Zoology* 28(4): 209–217.

Goren M. **1979**. The Gobiinae of the Red Sea (Pisces: Gobiidae). *Senckenbergiana Biologica* 60(1/2): 13–64.

Goren M. **1980**. Red Sea fishes assigned to the genus *Callogobius* Bleeker with a description of a new species (Teleostei: Gobiidae). *Israel Journal of Zoology* 28(4): 209–217.

Goren M. **1981**. Three new species and three new records of gobies from New Caledonia. *Cybium* (Série 3) 5(3): 93–101.

Goren M. **1982**. *Quisquilius flavicaudatus*, a new gobioid fish from the coral reefs of the Red Sea. *Zoologische Mededelingen* 56(11): 139–142.

Goren M. **1984**. A new species of *Oplopomops* Smith 1959 from Elat, northern Red Sea (Pisces: Gobiidae). *Senckenbergiana Biologica* 65(1/2): 19–23.

Goren M. **1984**. Three new species and two new records for the Red Sea of invertebrate associated gobies (Gobiidae, Pisces). *Cybium* 8(1): 71–82.

Goren M. **1985**. A review of the gobiid fish genus *Monishia* Smith, 1949, from the Western Indian Ocean and Red Sea, with description of a new species. *Contributions in Science* (Los Angeles) 360: 1–9.

Goren M. **1985**. *Trimma fishelsoni*, a new gobiid fish from the Gulf of Elat, northern Red Sea. *Israel Journal of Zoology* 33(1–2): 63–67.

Goren M. 1986. A suggested model for the recolonization process of the Red Sea at the post glacial period (pp. 648–654). In: Uyeno T, Arai R, Taniuchi T, Matsuura K (eds) *Indo-Pacific fish biology: Proceedings of the Second International Conference on Indo-Pacific Fishes*. Tokyo: Ichthyological Society of Japan. 985 pp.

Goren M. **1987**. *Kraemeria nudum* (Regan) – first record of the family Kraemeriidae in the Red Sea. *Israel Journal of Zoology* 34(3–4): 149–153.

Goren M. **1991**. A new *Coryogalops* species (Pisces: Gobiidae) from South Africa. J.L.B. Smith Institute of Ichthyology Special Publication 52: 1–7.

Goren M. **1992**. *Obliquogobius turkayi*, a new species of gobiid fish from the deep water of the central Red Sea. *Senckenbergiana Maritima* 22(3): 265–270.

Goren M. **1996**. A review of the southern African gobiid fish genus *Caffrogobius* Smitt, 1900. *J.L.B. Smith Institute of Ichthyology Special Publication* 57: 1–28.

Goren M, Baranes A. **1995**. *Priolepis goldschmidtae* (Gobiidae), a new species from the deep water of the northern Gulf of Aqaba, Red Sea. *Cybium* 19(4): 343–347.

Goren M, Dor M. **1994**. *An updated checklist of the fishes of the Red Sea (CLOFRES II).* Jerusalem: Israel Academy of Sciences and Humanities, and Elat: Interuniversity Institute for Marine Sciences. xii + 120 pp.

Goren M, Karplus I. **1980**. *Fowleria abocellata*, a new cardinal fish from the Gulf of Elat - Red Sea (Pisces, Apogonidae). *Zoologische Mededelingen* (Leiden) 55(20): 231–234.

Goren M, Karplus I. **1983**. *Tomiyamichthys randalli* n. sp., a gobiid associated with a shrimp, from the Red Sea (Pisces: Gobiidae). *Senckenbergiana Biologica* 63(1/2): 27–31.

Goren M, Klausewitz W. **1978**. Two Mediterranean gobiid fishes new in the Red Sea (Pisces: Gobiidae). *Senckenbergiana Biologica* 59(1/2): 19–24.

Goren M, Miroz A, Baranes A. **1991**. *Callogobius amikami* a new species of goby (Gobiidae) from the Red Sea. *Cybium* 15(4): 299–302.

Goren M, Spanier E. **1985**. The communities of benthic fish in Foul Bay (Tiran Island, Red Sea). *Oceanologia Acta* 8(4): 471–478.

Goren M, Stern N, Galil B, Diamant A. 2011. On the occurrence of the Indo-Pacific *Champsodon nudivittis* (Ogilby, 1895) (Perciformes, Champsodontidae) from the Mediterranean coast of Israel, and the presence of the species in the Red Sea. *Aquatic Invasions* 6(Suppl. 1): S115–S117. https://doi.org/10.3391/ai.2011.6.S1.026

Goren M, Voldarsky Z. **1980**. *Paragobiodon xanthosoma* (Bleeker) new for the Red Sea (Pisces: Gobiidae). *Israel Journal of Zoology* 29(1–3): 150–152.

Gorlick DL. **1980**. Ingestion of host fish surface mucus by the Hawaiian cleaning wrasse, *Labroides phthirophagus* (Labridae), and its effect on host species preference. *Copeia* 1980(4): 863–868.

Gosline WA. **1951**. The osteology and classification of the ophichthid eels of the Hawaiian Islands. *Pacific Science* 5(4): 298–320.

Gosline WA. **1953**. Hawaiian shallow-water fishes of the family Brotulidae, with the description of a new genus and notes on brotulid anatomy. *Copeia* 1953(4): 215–225.

Gosline WA. **1958**. Central Pacific eels of the genus *Uropterygius*, with descriptions of two new species. *Pacific Science* 12(3): 221–228.

Gosline WA. **1959**. Four new species, a new genus, and a new suborder of Hawaiian fishes. *Pacific Science* 13(1): 67–77.

Gosline WA. **1960**. Contributions toward a classification of modern isospondylous fishes. *Bulletin of the British Museum* (*Natural History*) *Zoology* 6(6): 325–365.

Gosline WA. **1966**. The limits of the fish family Serranidae, with notes on other lower percoids. *Proceedings of the California Academy of Sciences* (Series 4) 33(6): 91–111.

Gosline WA. **1968**. The suborders of perciform fishes. *Proceedings of the United States National Museum* 124(3647): 1–78.

Gosline WA. **1971**. *Functional morphology and classification of teleostean fishes*. Honolulu: University Press of Hawaii. 208 pp.

Gosline WA, Brock VE. **1960**. *Handbook of Hawaiian fishes*. Honolulu: University of Hawaii Press. ix + 372 pp. Goto T. **2001**. Comparative anatomy, phylogeny and cladistic classification of the Order Orectolobiformes (Chondrichthyes, Elasmobranchii). *Memoirs of the Graduate School of Fisheries Sciences, Hokkaido University* 48(1): 1–100.

Gotshall DW, Fitch JE. **1968**. The louvar, *Luvarus imperialis*, in the eastern Pacific, with notes on its life history. *Copeia* 1968(1): 181–183.

Goüan A. 1770. Historia piscium, sisteas ipsorum anatomen externam, internam, atque genera in classes et ordines redacta. Strasbourg: Amand König, Argentorati. xviii + 252 pp. [Text in Latin and French]

Gouiric-Cavalli S. **2015**. *Jonoichthys challwa* gen. et sp. nov., a new Aspidorhynchiform (Osteichthyes, Neopterygii, Teleosteomorpha) from the marine Upper Jurassic sediments of Argentina, with comments about paleobiogeography of Jurassic aspidorhynchids. *Comptes Rendus Palevol* 14(4): 291–304. http://dx.doi.org/10.1016/j. crpv.2015.03.007

Govoni JJ, Olney JE, Markle DF, Curtsinger WR.
1984. Observations on structures and evaluation of possible functions of the vexillum in larval carapidae (Ophidiiformes). *Bulletin of Marine Science* 34(1): 60–70.

Govoni JJ, West MA, Zivotofsky D, Zivotofsky AZ, Bowser PR, Collette BB. **2004**. Ontogeny of squamation in swordfish, *Xiphias gladius. Copeia* 2004(2): 391–396.

Graham JB, Dickson KA. 2000. The evolution of thunniform locomotion and heat conservation in scombrid fishes. New insights based on the morphology of *Allothunnus fallai*. *Zoological Journal of the Linnean Society* 129(4): 419–466.

Graham JB, Dickson KA. **2001**. Anatomical and physiological specializations for endothermy (pp. 121–165). In: Block BA, Stevens ED (eds) *Tuna: Physiology, ecology, and evolution*. San Diego: Academic Press. 468 pp.

Graham NAJ, Wilson SK, Jennings S, Polunin NVC, Bijoux JP, Robinson J. **2006**. Dynamic fragility of oceanic coral reef ecosystems. *Proceedings of the National Acadamy of Sciences* 103(22): 8425–8429. https://doi.org/10.1073/ pnas.0600693103

Grande L. **2010**. An empirical synthetic pattern study of gars (Lepisosteiformes) and closely related species, based mostly on skeletal anatomy. The resurrection of Holostei. *American Society of Ichthyologists and Herpetologists Special Publication* 6: x + 871 pp. [Supplementary issue to *Copeia* 2010.]

Grande L, Bemis WE. **1998**. A comprehensive phylogenetic study of amiid fishes (Amiidae) based on comparative skeletal anatomy. An empirical search for interconnected patterns of natural history. *Journal of Vertebrate Paleontology* 18(Supplement 1, Memoir 4): 1–696.

Grande TC. **1999**. Revision of the genus *Gonorynchus* Scopoli, 1777 (Teleostei: Ostariophysi). *Copeia* 1999(2): 453–469.

Grant WS, Clark A-M, Bowen BW. **1998**. Why restriction length polymorphism analysis of mitochondrial DNA failed to resolve sardine (*Sardinops*) biogeography: insights from mitochondrial DNA cyctochrome b sequences. *Canadian Journal of Fisheries and Aquatic Sciences* 55: 2539–2547.

Gray JE. 1830–1835. Illustrations of Indian zoology; chiefly selected from the collection of Major-General Hardwicke, F.R.S. 20 parts in 2 Vols. Pls. 1–202 (Fishes on Pls. 84–99 in Vol. 1 and 88–102 in Vol. 2). London: Treuttel, Wurtz, Treuttel jun. & Richter.

Gray JE. **1831**. Description of twelve new genera of fish, discovered by Gen. Hardwicke, in India, the greater part in the British Museum. *Zoological Miscellany* 1831: 7–9.

Gray JE. **1831**. Description of a genus (*Micropus*) of percoid fish, discovered by Samuel Stutchbury in the Pacific sea, and now in the British Museum. *Zoological Miscellany* 1831: 20.

Gray JE. **1838**. Description of a new species of *Tetrapturus* from the Cape of Good Hope. *Annals and Magazine of Natural History* (Series 1) 1(4): 313.

Gray JE. **1851**. *List of the specimens of fish in the collection of the British Museum. Part I. Chondropterygii*. London: British Museum. x + 160 pp.

Gray JE (ed). **1854**. *Catalogue of fish collected and described by Laurence Theodore Gronow, now in the British Museum*. London: British Museum. vii + 196 pp.

Gray KN, McDowell J, Collette B, Graves JE. **2009**. A molecular phylogeny of the remoras and their relatives. *Bulletin of Marine Science* 84(2): 183–197.

Greenfield DW. **1974**. A revision of the squirrelfish genus *Myripristis* Cuvier (Pisces: Holocentridae). *Science Bulletin, Natural History Museum of Los Angeles County* 19: 1–54.

Greenfield DW. **1996**. *Perulibatrachus kilburni*, a new toadfish from east Africa (Teleostei: Batrachoididae). *Copeia* 1996(4): 901–904.

Greenfield DW. **1997**. *Allenbatrachus*, a new genus of Indo-Pacific toadfish (Batrachoididae). *Pacific Science* 51(3): 306–313.

Greenfield DW. **2001**. Revision of the *Apogon erythrinus* complex (Teleostei: Apogonidae). *Copeia* 2001(2): 459–472.

Greenfield DW. 2005. Perulibatrachus aquilonarius, a new toadfish species from India (Teleostei: Batrachoididae).
Proceedings of the California Academy of Sciences (Series 4) 56(7): 76–79.

Greenfield DW. **2006**. Two new toadfish genera (Teleostei: Batrachoididae). *Proceedings of the California Academy of Sciences* (Series 4) 57(32): 945–954.

Greenfield DW. **2007**. Geographic variation in a cardinalfish, *Apogon dianthus* (Teleostei: Apogonidae). *Proceedings of the California Academy of Sciences* (Series 4) 58(30): 601–605. Greenfield DW. **2012**. *Colletteichthys occidentalis*, a new toadfish species from the Arabian Peninsula and northern Arabian Sea (Teleostei: Batrachoididae). *Zootaxa* 3165: 64–68. http://dx.doi.org/10.11646/zootaxa.3165.1.4

Greenfield DW. **2012**. *Austrobatrachus iselesele*, a new toadfish species from South Africa (Teleostei: Batrachoididae). *Zootaxa* 3400: 58–63. http://dx.doi.org/10.11646/ zootaxa.3400.1.4

Greenfield DW. **2014**. A new toadfish species from Somalia (Teleostei: Batrachoididae). *Copeia* 2014(4): 668–672. http://dx.doi.org/10.1643/CI-14-113

Greenfield DW, Bineesh KK, Akhilesh KV. **2012**. *Colletteichthys flavipinnis*, a new toadfish species from Sri Lanka and India (Teleostei: Batrachoididae). *Zootaxa* 3437: 24–31. http://dx.doi.org/10.11646/zootaxa.3437.1.2

Greenfield DW, Bogorodsky SV, Mal AO. **2014**. Two new Red Sea dwarfgobies (Teleostei, Gobiidae, *Eviota*). *Journal of the Ocean Science Foundation* 10: 1–10. http://dx.doi.org/10.5281/zenodo.1042767

Greenfield DW, Jewett SL. **2012**. Two new gobiid fishes of the genus *Eviota* from the Indian Ocean (Teleostei: Gobiidae). *Zootaxa* 3515: 67–74. http://dx.doi.org/10.11646/ zootaxa.3515.1.5

Greenfield DW, Mee JKL, Randall JE. **1994**. *Bifax lacinia*, a new genus and species of toadfish (Batrachoididae) from the south coast of Oman. *Fauna of Saudi Arabia* 14: 276–281.

Greenfield DW, Smith WL. **2004**. Allenbatrachus meridionalis, a new toadfish (Batrachoididae) from Madagascar and Reunion. Prococeedings of the California Academy of Sciences (Series 4) 55(30): 568–572.

Greenfield DW, Winterbottom R, Collette BB. **2008**. Review of the toadfish genera (Teleostei: Batrachoididae). *Prococeedings of the California Academy of Sciences* (Series 4) 59(15): 665–710.

Greenwood PH, Miles RS, Patterson C (eds). **1973**. *Interrelationships of fishes*. Supplement No. 1 to the Zoological Journal of the Linnean Society 53. London: Academic Press. 536 pp.

Greenwood PH, Rosen DE, Weitzman SH, Myers GS. **1966**. Phyletic studies of teleostean fishes, with a provisional classification of living forms. *Bulletin of the American Museum of Natural History* 131(art.4): 341–455.

Gregory WK. **1935**. Winged sharks. *Bulletin of the New York Zoological Society* 38: 129–133.

Grey M. **1953**. Fishes of the family Gempylidae, with records of *Nesiarchus and Epinnula* from the western Atlantic and description of two new subspecies of *Epinnula orientalis*. *Copeia* 1953(3): 135–141.

Grey M. **1955**. The fishes of the genus *Tetragonurus* Risso. *Dana Report* 41: 1–75.

Grey M. **1955**. Notes on a collection of Bermuda deep-sea fishes. *Fieldiana Zoology* **37**: 265–302.

Grey M. **1960**. Description of a western Atlantic specimen of *Scombrolabrax heterolepis* Roule and notes on fishes of the family Gempylidae. *Copeia* 1960(3): 210–215.

Grey M. **1964**. Family Gonostomatidae. In: Fishes of the western North Atlantic. *Memoir of the Sears Foundation for Marine Research* 1(part 4): 78–240.

Grier HJ, Collette BB. **1987**. Unique spermatozeugmata in testes of halfbeaks of the genus *Zenarchopterus* (Teleostei: Hemiramphidae). *Copeia* 1987(2): 300–311.

Grier HJ, Linton JR, Leatherland JF, de Vlaming VL. 1980. Structural evidence for two different testicular types in teleost fishes. *American Journal of Anatomy* 159(3): 331–345.

Griffin LT. **1926**. Descriptions of New Zealand fishes. *Transactions of the New Zealand Institute* 56: 538–546, Pls. 93–98.

Griffin LT. **1928**. Studies in New Zealand fishes. *Transactions* and Proceedings of the New Zealand Institute 59(2): 374–388.

Griffith E, Smith CH. **1834**. *The class Pisces, arranged by the Baron Cuvier, with supplementary additions*. London: Whitaker. 680 pp.

Griffiths MH, Heemstra PC. **1995**. A contribution to the taxonomy of the marine fish genus *Argyrosomus* (Perciformes: Sciaenidae), with description of two new species from southern Africa. *Ichthyological Bulletin of the J.L.B. Smith Institute of Ichthyology* 65: 1–40.

Grindley JR, Penrith MJ. **1965**. Notes on the bathypelagic fauna of the seas around South Africa. *Zoologica Africana* 1(2): 275–295.

Grobecker DB. **1981**. Steady as a rock, fast as lightning. *Natural History* 90(5): 50–53.

Grogan ED, Lund R. **2004**. The origin and relationships of early Chondrichthyes (pp. 3–31). In: Carrier JC, Musick JA, Heithaus MR (eds) *Biology of sharks and their relatives*. Boca Raton: CRC Press. 596 pp.

Grogan ED, Lund R, Greenfest-Allen E. **2012**. The origin and relationships of early chondrichthyans (pp. 3–29). In: Carrier JC, Musick JA, Heithaus MR (eds) *Biology of sharks and their relatives* (2<sup>nd</sup> edition). Boca Raton: CRC Press. 672 pp.

Gronow LT. **1763**. Zoophylacium Gronovianum, exhibens animalia quadrupeda, amphibia, pisces, insecta, vermes, mollusca, testacea, et xoophyta, quae in museo suo adservavit, examine subjecit, systematice disposuit, atque descripsit (fasciculus primus). Leiden: Theodorus Haak & Samuel et Johanness Luchtmans. 136 pp.

Gronow LT. **1772**. Animalium rariorum fasciculus. Pisces. *Acta Helvetica, Physico-Mathematico-Anatomico-Medica* (Basileae) 7: 43–52. Gronow LT. 1854. Catalogue of fish collected and described by Laurence Theodore Gronow, now in the British Museum.
Edited from the manuscript by JE Gray. London: British Museum. vii + 196 pp.

Grove JS, Lavenberg RJ. **1997**. *The fishes of the Galápagos Islands*. Stanford, California: Stanford University Press. xiv + 683 pp.

Gruber SH, Compagno LJV. **1981**. Taxonomic status and biology of the bigeye thresher, *Alopias superciliosus*. *Fishery Bulletin* 79(4): 617–640.

Grudtsev ME, Salekhova LP, Lushchina VG. **1986**. Distribution, ecology, and intraspecies variation of flying fishes of the genus *Exocoetus* from the Atlantic Ocean. *Voprosy Ikhtiologii* 26(6): 919–929.

Gruvel A. **1936**. Contribution à l'étude de la bionomie générale et de l'exploration de la faune du canal de Suez. *Mémoires présentées à l'Institut d'Égypte N.S.* 29: 1–255.

Gruvel A, Chabanaud P. **1937**. Missions A. Gruvel dans le Canal de Suez. II. Poissons. *Mémoires de l'Institut d'Égypte N.S.* 35: 1–31.

Gubanov EP, Schleib NA (eds). 1980. Sharks of the Arabian Gulf. Kuwait Ministry of Public Works, Agriculture Department, Fisheries Division. 69 pp.

Gubanov YP. **1972**. On the biology of the thresher shark *Alopias vulpinus* (Bonnaterre) in the northwest Indian Ocean. *Journal of Ichthyology* 12: 591–600.

Gubanov YP. **1978**. The reproduction of some species of pelagic sharks from the equatorial zone of the Indian Ocean. *Voprosy Ikhtiologii* 18: 781–792.

Gubanov YP. **1985**. Presence of the sharp tooth sand shark, *Odontaspis ferox* (Odontaspididae), in the open waters of the Indian Ocean. *Journal of Ichthyology* 25(2): 156–158.

Gubanov YP, Kondyuran VV, Myagkov NA. 1986. Sharks of the world ocean: Identification handbook [=Akuly Mirovogo okeana: Spravochnik-opredelitel']. Moscow, Agropromizdat. 272 pp. [In Russian]

Gubanov YP, Grigor'yev VN. 1975. Distribution and biology of the blue shark *Prionace glauca* (Carcharhinidae) of the Indian Ocean. *Journal of Ichthyology* 15(1): 37–43.
[Translated from *Voprosy Ikhtiologii*]

Gudger EW. **1929**. Nicolas Pike and his unpublished paintings of the fishes of Mauritius, western Indian Ocean, with an index to the fishes. *Bulletin of the American Museum of Natural History* 58(art.9): 489–530.

Guérin-Méneville FE. **1829–1838**. Iconographie du Règne animal de G. Cuvier ... (1829–44). I. Planches des Animaux Vertébrés. IV. Poissons (pp. 1–44, Pls. 1–70). Paris.

Guerrero RD, Shelton WL. **1974**. An aceto-carmine squash method for sexing juvenile fishes. *The Progressive Fish-Culturist* 36(1): 56.

Guézé P. **1976**. *Upeneus niebuhri*, espèce nouvelle de Mullidae de la Mer Rouge (Pisces, Perciformes). *Revue des Travaux de l'Institut des Pêches Maritimes* 40(3–4): 596.

Guichenot A. **1847**. Description de deux nouvelles espèces de *Cossyphes. Revue Zoologique par la Société Cuvierienne* (Paris) 10: 282–284.

Guichenot A. **1848**. Sur une nouvelle espèce de malacanthe (*Malacanthus brevirostris*). *Revue Zoologique par la Société Cuvierienne* (Paris) 11: 14–15.

Guichenot A. **1853**. Poissons. In: de la Sagra R (ed) *Histoire physique, politique et naturelle de l'Ile de Cuba*. Vol. 2. Paris. pp. 1–206, Pls. 1–5.

Guichenot A. 1863. Faune ichthyologique. Part 2. Annexe
C: C1–C32. In: Maillard L Notes sur l'île de la Réunion (Bourbon) (2 volumes in 1). Paris: Dentu. 22 pp.
[Date uncertain; cited as 1862 to 1865]

Guichenot A. **1866**. Catalogue des poissons de Madagascar de la collection du Musée de Paris avec la description de plusiers espèces nouvelles. *Mémoires de la Société Impériale des Sciences Naturelles de Cherbourg* 12 (= Series 2, vol. 2): 129–148.

Guichenot A. **1867**. Ichthyologie. III. L'Argentine léioglosse, nouveau genre de Salmonoïdes. *Annales de la Société Linnéenne du Département de Maine-et-Loire* 9: 15–17.

Guichenot A. **1868**. Index generum ac specierum Anthiadidorum hucusque in Museo Parisiensi observatorum. *Annales de la Société Linnéenne du Département de Maine-et-Loire* 10: 80–87.

Guichenot A. **1869**. Notice sur quelques poissons inédits de Madagascar et de la Chine. *Nouvelles Archives du Muséum d'Histoire Naturelle, Paris* 5(fasc. 3): 193–206.

Guilcher A. **1988**. A heretofore neglected type of coral reef: the ridge reef. Morphology and origin. *Proceedings of 6th International Coral Reef Symposium, Australia* 3: 399–402.

Guitart Manday D. **1966**. Nuevo nombre para una especie de tiburón del género *Isurus* (Elasmobranchii: Isuridae) de aguas Cubanas. *Poeyana* (Series A) (15): 1–9.

Gunderman N, Popper DM, Lichatowich T. **1983**. Biology and life cycle of *Siganus vermiculatus* (Siganidae, Pisces). *Pacific Science* 37(2): 165–180.

Gunnerus JE. **1765**. Von der Seekatze. Der Drontheimischen Gesellschaft Schriften as dem Dänischen überfest 2: 284–290.

Gunnerus JE. 1765. Beschreibung des Brugden (Selache maxima). Der Drontheimischen Gesellschaft Schriften 3: 28–43.

Günther A. **1859**. *Catalogue of the fishes in the British Museum*. Vol. 1. *Catalogue of the acanthopterygian fishes in the collection of the British Museum*. *Gasterosteidae, Berycidae, Percidae, Aphredoderidae, Pristipomatidae, Mullidae, Sparidae*. London: Trustees of the British Museum. xxxii + 524 pp. Günther A. **1860**. *Catalogue of the fishes in the British Museum*. Vol. 2. *Catalogue of the acanthopterygian fishes in the collection of the British Museum*. *Squamipinnes, Ciirrhitidae, Triglidae, Polynemidae, Sphyraenidae, Trichiuridae, Scombridae, Carangidae, Xiphiidae*. London: British Museum. xxi + 548 pp.

Günther A. **1860**. On the history of *Echeneis*. *Annals and Magazine of Natural History* (Series 3) 5(29): 386–402.

Günther A. **1861**. On a new species of fish of the genus *Gerres. Proceedings of the Zoological Society of London* 1861(pt 1): 142–143.

Günther A. **1861**. On the British species of *Mugil*, or grey mullets. *Annals and Magazine of Natural History* (Series 3) 7(41): 345–352.

Günther A. **1861**. On a new species of fish of the genus *Gerres. Annals and Magazine of Natural History* (Series 3) 8(44): 189.

Günther A. **1861**. A preliminary synopsis of the labroid genera. *Annals and Magazine of Natural History* (Series 3) 8(47): 382–389.

Günther A. 1861. Catalogue of the fishes in the British Museum.
Vol. 3. Catalogue of the acanthopterygian fishes in the
British Museum. Gobiidae, Discoboli, Pediculati, Blenniidae,
Labyrinthici, Mugilidae, Notacanthi. London: British
Museum. xxv + 586 pp.

Günther A. **1862**. *Catalogue of the fishes in the British Museum*. *Vol. 4. Catalogue of the Acanthopterygii, Pharyngognathi and Anacanthini in the collection of the British Museum*. London: British Museum. xxi + 534 pp.

Günther A. **1864**. *Catalogue of the fishes in the British Museum*. Vol. 5. *Catalogue of the Physostomi, containing the families Siluridae, Characinidae, Haplochitonidae, Sternoptychidae, Scopelidae, Stomiatidae in the collection of the British Museum*. London: British Museum. xxii + 455 pp.

Günther A. **1864**. On a new genus of pediculate fish from the Sea of Madeira. *Proceedings of the Zoological Society of London* 1864(pt 2): 301–303.

Günther A. **1866**. *Catalogue of the fishes in the British Museum. Vol. 6. Catalogue of the Physostomi, containing the families Salmonidae, Percopsidae, Galaxidae, Mormyridae, Gymnarchidae, Esocidae, Umbridae, Scombresocidae, Cyprinodontidae, in the collection of the British Museum.* London: British Museum. xv + 368 pp.

Günther A. **1867**. Descriptions of some new or little-known species of fishes in the collection of the British Museum. *Proceedings of the Zoological Society of London* 1867(1): 99–104.

Günther A. **1868**. Additions to the ichthyological fauna of Zanzibar. *Annals and Magazine of Natural History* (Series 4) 1(6): 457–459.

Günther A. **1868**. *Catalogue of the fishes in the British Museum*. Vol. 7. *Catalogue of the Physostomi, containing the families Heteropygii, Cyprinidae, Gonorhynchidae, Hyodontidae, Osteoglossidae, Clupeidae,... [through]... Halosauridae, in the collection of the British Museum*. London: British Museum. xx + 512 pp.

Günther A. **1868**. Diagnoses of some new freshwater fishes from Surinami and Brazil, in the collection of the British Museum. *Annals and Magazine of Natural History* (Series 4) 1(6): 475–481.

Günther A. **1869**. Descriptions of two new species of fishes discovered by the Marquis J. Doria. *Annals and Magazine of Natural History* (Series 4) 3(18): 444–445.

Günther A. **1870**. *Catalogue of the fishes in the British Museum*. Vol. 8. *Catalogue of the Physostomi, containing the families Gymnotidae, Symbranchidae, Muraenidae, Pegasidae, and of the Lophobranchii, Plectognathi, Dipnoi, ...[through] ... Leptocardii, in the British Museum*. London: British Museum. xxv + 549 pp.

Günther A. **1872**. On some new species of reptiles and fishes collected by J. Brenchley, Esq. *Annals and Magazine of Natural History* (Series 4) 10(60): 418–426.

Günther A. **1872**. Report on several collections of fishes recently obtained for the British Museum. *Proceedings of the Zoological Society London* 1871(pt 3) [for 1871]: 652–675.

Günther A. **1873–1910**. Andrew Garrett's Fische der Südsee, beschrieben und redigirt von A.C.L.G. Günther (3 vols). [refer separate entries 1873–1875; 1876–1881; 1909–1910]

Günther A. **1873**. Erster ichthyologischer Beitrag nach Exemplaren aus dem Museum Godeffroy. *Journal des Museum Godeffroy* 1(2): 97–103.

Günther A. **1873**. Zweiter ichthyologischer Beitrag nach Exemplaren aus dem Museum Godeffroy. *Journal des Museum Godeffroy* 1(4): 89–92.

Günther A. **1873–1875**. Andrew Garrett's Fische der Südsee. *Journal des Museum Godeffroy* 2(Hefts 3, 5, 7, 9): 1–128.

Günther A. **1874**. Third notice of a collection of fishes made by Mr. Swinhoe in China. *Annals and Magazine of Natural History* (Series 4) 13(74): 154–159.

Günther A. **1874**. Descriptions of new species of fishes in the British Museum. *Annals and Magazine of Natural History* (Series 4) 14(83): 368–371.

Günther A. **1874**. Descriptions of new species of fishes in the British Museum. [Continued from p. 371] *Annals and Magazine of Natural History* (Series 4) 14(84): 453–455.

Günther A. **1876–1881**. Andrew Garrett's Fische der Südsee. *Journal des Museum Godeffroy* 4(Hefts 11, 13, 15): 129–256.

Günther A. 1876. Remarks on fishes, with descriptions of new species in the British Museum, chiefly from Southern Seas. Annals and Magazine of Natural History (Series 4) 17(101): 389–402. Günther A. **1877**. Preliminary notes on new fishes collected in Japan during the expedition of H M S *Challenger*. *Annals and Magazine of Natural History* (Series 4) 20(119): 433–446.

Günther A. **1878**. Preliminary notices of deep-sea fishes collected during the voyage of H.M.S. *Challenger. Annals and Magazine of Natural History* (Series 5) 2(8): 179–187.

Günther A. 1880. Report on the shore fishes procured during the voyage of H.M.S. *Challenger* during the years 1873–1876. *Report on the scientific results of the voyage of H.M.S.* Challenger during the years 1873–76. Zoology 1(pt 6): 1–82.

Günther A. **1881**. Account of the zoological collections made during the survey of H.M.S. *Alert* in the Straits of Magellan and on the coast of Patagonia. *Proceedings of the Zoological Society of London* 4(1181 pt 1): 2–141.

Günther A. **1884**. Reptilia, Batrachia, and Pisces (pp. 29–33). In: *Report on the zoological collections made in the Indo-Pacific Ocean during the voyage of HMS* Alert, *1881–2*. London: British Museum. 684 pp.

Günther A. **1886**. Note on *Pachymetopon* and the Australian species of *Pimelepterus*. *Annals and Magazine of Natural History* (Series 5) 18(107): 367–368.

Günther A. **1887**. Descriptions of two new species of fishes from Mauritius (*Latilus fronticinctus, Platycephalus subfasciatus*). *Proceedings of the Zoological Society of London* 1887(pt 3): 550–551.

Günther A. **1887**. Report on the deep-sea fishes collected by H.M.S. *Challenger* during the years 1873–76. *Report on the Scientific Results of the Voyage of H.M.S.* Challenger 22(pt 57): 1–268.

Günther A. 1889. Complete catalogue of Linné's private collection of fishes, now in possession of the Linnean Society. *Proceedings of the Linnean Society of London* 111<sup>th</sup> Session: 20–38.

Günther A. **1889**. Report on the pelagic fishes collected by H.M.S. *Challenger* during the years 1873–76. *Report on the Scientific Results of the Voyage of H.M.S.* Challenger 31(pt 78): 1–47.

Günther A. **1890**. Description of a new species of deep-sea fish from the Cape (*Lophotes fiski*). *Proceedings of the Zoological Society of London* 1890(pt 2): 244–247.

Günther A. **1892**. Description of a remarkable fish from Mauritius, belonging to the genus *Scorpaena*. *Proceedings of the Zoological Society of London* 1891(pt 4): 482–483.

Günther A. **1909–1910**. Andrew Garrett's Fische der Südsee. *Journal des Museum Godeffroy* 6(Hefts 16–17): 261–498.

Gupta MV. 1966. Two new species of ribbonfishes of the genus *Trichiurus* Linnaeus (Pisces: Trichiuridae) from the Hooghly estuarine system. *Proceedings of the Zoological Society of Calcutta* 19(2): 169–171. Gushchin AV, Sukhovershin, Konovalenko II, Sukhorukova VS. **1986**. A polar shark of the genus *Somniosus* (Squalidae) caught in the Southern Hemisphere. *Voprosy Ikhtiologii* 26: 514–515. [In Russian]

Gushiken S. **1972**. *Fishes of the Okinawa Islands*. Urasoe: Tiger Press. 251 pp.

Gushiken S. **1973**. *Fishes of the Okinawa Islands* (2<sup>nd</sup> edition). Naha: Ryukyu Fisheries Association. 251 pp. [In Japanese]

## Н

Haas DL, Ebert DA. **2006**. *Torpedo formosa* sp. nov., a new species of electric ray (Chondrichthyes: Torpediniformes: Torpedinidae) from Taiwan. *Zootaxa* 1320: 1–14.

Haas G, Steinitz H. **1947**. Erythrean fishes on the Mediterranean coast of Palestine. *Nature* 160: 28.

Haedrich RL. **1967**. A new species of *Psenopsis* (Stromateoidei, Centrolophidae) from Indo-Malayan seas. *Japanese Journal of Ichthyology* 14(4/6): 187–196.

Haedrich RL. **1967**. The stromateoid fishes; systematics and a classification. *Bulletin of the Museum of Comparative Zoology* 135(2): 31–139.

Haedrich RL. **1968**. The stromateoid fish *Psenopsis obscura* from off Mozambique. *Copeia* 1968(4): 873–874.

Haedrich RL. **1969**. A new family of aberrant stromateoid fishes from the equatorial Indo-Pacific. *Dana Report* 76: 1–14.

Haedrich RL. **1972**. Ergebnisse der Forschungsreisen des FFS *Walther Herwig* nach Südamerika. XXIII. Fishes of the family Nomeidae (Perciformes, Stromateoidei). *Archiv für Fischereiwissenschaft* 23(2): 73–88.

Haedrich RL. 1986. Centrolophidae (pp. 1177–1182). In: Whitehead PJP, Bauchot M-L, Hureau J-C, Nielsen J, Tortonese E (eds) *Fishes of the north-eastern Atlantic and the Mediterranean*. Vol. III. Paris: UNESCO.

Haedrich RL. 1990. Centrolophidae (pp. 1010–1013);
Nomeidae (pp. 1014–1018); Stromateidae (p. 1021);
Tetragonuridae (p. 1022). In: Quéro J-C, Hureau J-C, Karrer K, Post A, Saldanha L (eds) *Check-list of the fishes of the eastern tropical Atlantic (CLOFETA)*. Vol. 2. Lisbon: Junta Nacional de Investigação Científica e Tecnológica.

Haedrich RL, Cervigón F. **1969**. Distribution of the centrolophid fish *Schedophilus pemarco*, with notes on its biology. *Breviora* 340: 1–9.

Haedrich RL, Horn MH. **1972**. A key to the stromateoid fishes. *Woods Hole Oceanographic Institution Technical Report* 72-15: 1–47.

Haigh EH. **1972**. Larval development of three species of economically important South African fishes. *Annals of the South African Museum* 59(3): 1–70.

Haigh EH, Whitfield AK. 1993. Larval development of Gilchristella aestuaria (Gilchrist, 1914) (Pisces: Clupeidae) from southern Africa. South African Journal of Zoology 28(4): 168–172.

Hallacher LE. 1974. The comparative morphology of extrinsic gasbladder musculature in the scorpionfish genus Sebastes (Pisces: Scorpaenidae). Proceedings of the California Academy of Sciences (Series 4) 40(3): 59–86.

Halstead BW. 1965. Poisonous and venomous marine animals of the world. Vol. I. Invertebrates. Washington, DC: U.S. Government Printing Office. xxxv + 994 pp.

Halstead BW. **1967**. *Poisonous and venomous marine animals of the world*. Vol. II. Vertebrates. Washington, DC: U.S. Government Printing Office. xxxi + 1070 pp.

Halstead BW. 1970. Poisonous and venomous marine animals of the world. Vol. III. Vertebrates (continued). Washington, DC: U.S. Government Printing Office. xxvi + 1006 pp.

Halstead BW. **1978**. *Poisonous and venomous marine animals of the world* (revised edition). Princeton, New Jersey: Darwin Press. xvi + 1043 pp.

Halstead BW. **1988**. *Poisonous and venomous marine animals of the world* (2<sup>nd</sup> edition). Princeton, New Jersey: Darwin Press. 1456 pp.

Halstead BW, Auerbach PS, Campbell DR. **1990**. *A color atlas of dangerous marine animals*. London: Wolfe Medical Publications. 192 pp.

Halstead BW, Danielson DD, Baldwin WJ, Engen PC. **1972**. Morphology of the venom apparatus of the leatherback fish *Scomberoides sanctipetri* (Cuvier). *Toxicon* 10(3): 249–250.

Haly A. 1875. Descriptions of new species of fish in the collection of the British Museum. *Annals and Magazine of Natural History* (Series 4) 15(88) (art.35): 268–270.

Haly A. **1886**. Note on the collection of fish in the Colombo Museum. *The Taprobanian* 1: 165–167.

Hamady LL, Natanson LJ, Skomal GB, Therold SR. **2014**. Vertebral bomb radiocarbon suggests extreme longevity in white sharks. *PLoS ONE* 9(1): e84006.

https://doi.org/10.1371/journal.pone.0084006

Hamaguchi S. **1996**. Bilateral asymmetrical testis in fishes of the genus *Oryzias*. *Zoological Science* 13(5): 757–763.

Hamilton F. **1822**. *An account of the fishes found in the river Ganges and its branches*. Edinburgh & London: Archibald Constable & Co. vii + 405 pp.

Hamilton-Buchanan F, see Hamilton F.

Hamlett WC (ed). **1999**. *Sharks, skates, and rays: the biology of elasmobranch fishes*. Baltimore, Maryland: Johns Hopkins University Press. 528 pp.

Hamlett WC (ed). **2005**. *Reproductive biology and phylogeny of Chondrichthyes: sharks, batoids and chimaeras*. Enfield, New Hampshire: Science Publishers, Inc. 572 pp. Hamner RM, Freshwater DW, Whitfield PE. 2007.
Mitochondrial cytochrome *b* analysis reveals two invasive lionfish species with strong founder effects in the western Atlantic. *Journal of Fish Biology* 71(Suppl. B): 214–222.

Haneda Y. **1958**. Studies on the luminous organisms found in waters adjacent to the Pacific coasts of Japan. Report II. *Records of Oceanographic Works in Japan* (New series) Special No. 2: 171–174.

Haneda Y, Johnson FH. **1958**. The luciferin-luciferase reaction in a fish, *Parapriacanthus beryciformis*, of newly discovered luminescence. *Proceedings of the National Academy of Sciences* 44(2): 127–129. https://doi.org/10.1073/pnas.44.2.127

Haneda Y, Johnson FH. **1962**. The photogenic organs of *Parapriacanthus beryciformes* Franz and other fish with the indirect type of luminescent system. *Journal of Morphology* 110(2): 187–198.

Haneda Y, Tsuji FI, Sugiyama N. **1969**. Luminescent systems in apogonid fishes from the Philippines. Science 165(3889): 188–190.

Hanel R, Tsigenopoulos CS. **2011**. Phylogeny, evolution and taxonomy of sparids with some notes on their ecology and biology (pp. 51–73). In: Pavlidis M, Mylonas CC (eds) *Sparidae: Biology and aquaculture of gilthead sea bream and other species*. Wiley-Blackwell, Oxford. xvii + 390 pp. http://dx.doi.org/10.1002/9781444392210.ch2

Hansen PEH. **1986**. Revision of the tripterygiid fish genus *Helcogramma*, including descriptions of four new species. *Bulletin of Marine Science* 38(2): 313–354.

Hardenberg JDF. **1933**. Notes on some genera of the Engraulidae. *Natuurkundig Tijdschrift voor Nederlandsch Indië* 93(2): 230–256.

Hardisty MW. **1979**. *Biology of the cyclostomes*. London: Chapman & Hall. 428 pp.

Hardy GS. **1980**. A redescription of the antitropical pufferfish *Arothron firmamentum* (Plectognathi: Tetraodontidae). *New Zealand Journal of Zoology* 7(1): 115–125.

Hardy GS. **1981**. New records of pufferfishes (family Tetraodontidae) from Australia and New Zealand, with notes on *Sphoeroides pachygaster* (Muller and Troschel) and *Lagocephalus sceleratus* (Gmelin). *Records of the National Museum of New Zealand* 1(20): 311–316.

Hardy GS. **1982**. First Pacific records of *Pelagocephalus marki* Heemstra and Smith (Tetraodontiformes: Tetraodontidae), and first male specimen of the genus. *New Zealand Journal of Zoology* 9(3): 377–380.

Hardy GS. **1983**. A revision of the fishes of the family Pentacerotidae (Perciformes). *New Zealand Journal of Zoology* 10(2): 177–220.

Hardy GS. **1983**. The status of *Torquigener hypselogeneion* (Bleeker) (Tetraodontiformes: Tetraodontidae) and some related species, including a new species from Hawaii. *Pacific Science* 37(1): 65–74.

Hardy GS. **1984**. *Tylerius*, a new generic name for the Indo-Pacific pufferfish, *Spheroides spinosissimus* Regan, 1908 (Tetraodontiformes: Tetraodontidae) and comparisons with *Amblyrhynchotes* (Bibron) Duméril. *Bulletin of Marine Science* 35(1): 32–37.

Hardy GS. **1985**. A new species of catshark in the genus *Parmaturus* Garman (Scyliorhinidae), from New Zealand. *New Zealand Journal of Zoology* 12(1): 119–124.

Hardy GS. **1989**. Description of a new species of *Torquigener* Whitley (Pisces: Tetraodontidae) from South Africa, with a key to the genus. *National Museum of New Zealand Records* 3(11): 119–123.

Hardy GS, Randall JE. **1983**. Description of a new species of pufferfish (Tetraodontiformes: Tetraodontidae) from the Red Sea and adjacent waters. *Israel Journal of Zoology* 32(1): 13–20.

Hariri K. 2002. Notes on the implementation of the Fisheries Management Plan for the Socotra Island Group (pp. 481–485). In: Apel M, Hariri K, Krupp F (eds) *Conservation and sustainable use of biodiversity of Socotra Archipelago. Marine habitat, biodiversity and fisheries surveys and management*. Final Report of Phase III. Senckenberg Research Institute, Frankfurt a.M., Germany.

Hariri K, Nichols P, Krupp F, Mishrighi S, Barrania A, Ali A, Kedidi S. 2002. Status of living marine resources in the Red Sea and Gulf of Aden and their management. The Regional Organization for the Conservation of the Red Sea and Gulf of Aden (PERSGA). Technical Series 4: 56 pp. Jedda.

Hariri K, Schotah J. **1999**. Review of fisheries legislation in Yemen (pp. 141–160). In: Krupp F, Hariri K (eds) *Conservation and sustainable use of biodiversity of Socotra Archipelago. Marine habitat, biodiversity and fisheries surveys and management*. Report of Phase I. Senckenberg Research Institute, Frankfurt a.M., Germany.

Hariri K, Yusif M. 1999. Fishing communities and status of the fisheries sector in the Socotra Archipelago (pp. 161–179).
In: Krupp F, Hariri K (eds) *Conservation and sustainable use of biodiversity of Socotra Archipelago. Marine habitat, biodiversity and fisheries surveys and management.* Report of Phase I. Senckenberg Research Institute, Frankfurt a.M., Germany.

Harman MAJ, Blaber SJM, Cyrus DP. **1982**. The biology and taxonomic status of an estuarine population of *Pranesus pinguis* (Lacépède) (Teleostei: Atherinidae) in south east Africa. *South African Journal of Zoology* 17(1): 15–23.

Harmelin JG, Harmelin-Vivien ML. **1976**. Observations "in situ" des aires de ponte de *Spicara amaris* (L.) (Pisces, Perciformes, Centracanthidae) dans les eaux de Port-Cros. *Scientific Reports of Port-Cros National Park* 2: 115–120.

Harmelin-Vivien ML. **1974**. Ichtyofaune des herbiers de phanérogames marines du Grand Récif de Tuléar (Malagasy). 1: Les peuplements et leur distribution écologique. *Tethys* 5(2–3): 425–436.

Harmelin-Vivien ML. **1976**. Ichtyofaune de quleques récifs corallines des îles Maurice et la Réunion (Archipel des Mascareignes, Océan Indien). *The Mauritius Institute Bulletin* 8(2): 69–104.

Harmelin-Vivien ML. 1977. Ecological distribution of the fishes on the outer slope of Tulear reef (Madagascar) (pp. 289–295). In: Taylor DL (ed) *Proceedings of the 3<sup>rd</sup> International Coral Reef Symposium, Miami*. Vol. 1. Miami, Florida: Rosenstiel School of Marine and Atmospheric Science.

Harmelin-Vivien ML. **1979**. *Ichtyofaune des récifs coralliens en France Outre-Mer. ICRI*. Secrétariat d'Etat à l'OutreMer et Ministère de l'Aménagement du Territoire et de l'Environment. 136pp.

Harmelin-Vivien ML, Bouchon C. 1976. Feeding behavior of some carnivorous fishes (Serranidae and Scorpaenidae) from Tuléar (Madagascar). *Marine Biology* 37(4): 329–340.

Harris SA, Cyrus DP. **1995**. Occurrence of larval fishes in the St. Lucia Estuary, KwaZulu-Natal, South Africa. *South African Journal of Marine Science* 16(1): 333–350.

Harris SA, Cyrus DP. **1996**. Occurrence of Schindler's fishes, genus *Schindleria* (Teleostei: Gobioidei), at a small reef in the mouth of the Kosi Estuary, KwaZulu Natal: a first record for southern Africa. *Bulletin of Marine Science* 59(1): 228–234.

Harris SA, Cyrus DP. **1997**. Composition, abundance and seasonality of larval fish in Richards Bay Harbour, KwaZulu-Natal, South Africa. *South African Journal of Aquatic Sciences* 23(1): 56–78.

Harris TFW. **1978**. Review of coastal currents in southern African waters. *National Scientific Programmes Unit: CSIR, SANSP Report* 30: 1–109.

Harrison CMH, Palmer G. **1968**. On the neotype of *Radiicephalus elongatus* Osório with remarks on its biology. *Bulletin of the British Museum (Natural History) Zoology* 16(5): 185–208.

Harrison TD. **2005**. Ichthyofauna of South African estuaries in relation to the zoogeography of the region. *Smithiana, Publications in Aquatic Biodiversity* Bulletin 6: 1–27.

Harrison TD, Whitfield AK. **1995**. Fish community structure in three temporarily open/closed estuaries on the Natal coast. *Ichthyological Bulletin of the J.L.B. Smith Institute of Ichthyology* 64: 1–80.

Hastings MC, Popper AN, Finneran JJ, Lanford PJ. 1996.
Effects of low-frequency underwater sound on hair cells of the inner ear and lateral line of the teleost fish *Astronotus ocellatus. Journal of the Acoustical Society of America* 99(3): 1759–1766. Hatooka K. **1984**. *Uropterygius nagoensis*, a new muraenid eel from Okinawa, Japan. *Japanese Journal of Ichthyology* 31(1): 20–22.

Hatooka K. **1988**. New record of the moray *Gymnothorax pindae* from the Amami Islands, Japan. *Japanese Journal of Ichthyology* 35(1): 87–89.

Hatooka K, Iwata A. **1993**. First record of *Gymnothorax zonipectis* from Japan (Pisces: Muraenidae). *Bulletin of the Osaka Museum of Natural History* 47: 19–24.

Hatooka K, Randall JE. **1992**. A new moray eel (*Gymnothorax*: Muraenidae) from Japan and Hawaii. *Japanese Journal of Ichthyology* 39(3): 183–190.

Hatooka K, Yoshino T, Ono A. **1995**. A rare ophichthid eel, *Ichthyapus vulturis*, from Izu Peninsula, Japan (Pisces: Ophichthidae). *Bulletin of the Osaka Museum of Natural History* 49: 19–22.

Hayashi M. **1990**. Two new cardinalfish (Apogonidae: genus *Apogon*) from the Indo-west Pacific. *Science Report of the Yokosuka City Museum* 38: 7–18.

Hayashi M, Hagiwara K, Hayashi H. **1986**. Osteology of the clingfishes of Japan. *Science Report of the Yokosuka City Museum* 34: 39–66.

Hayashi M, Hayashi H. **1985**. Two new records of gobiesocid fishes from Japan and a morphological study of their key characters. *Science Report of the Yokosuka City Museum* 33: 49–67.

Hayashi M, Suzuki T, Itoh T, Senou H. **1981**. Gobiid fishes of the Ryukyu Islands, southern Japan (III). Suborder Gobioidei. *Science Report of the Yokosuka City Museum* 28: 1–25. [In Japanese]

Hayden BP, Ray GC, Dolan R. 1984. Classification of coastal and marine environments. *Environmental Conservation* 11(3): 199–207. https://doi.org/10.1017/ S0376892900014211

Hays GC, Farquhar MR, Luschi P, Teo SLH, Thys TM. **2009**. Vertical niche overlap by two ocean giants with similar diets: ocean sunfish and leatherback turtles. *Journal of Experimental Marine Biology and Ecology* 370(1–2): 134–143.

Healey AJE, Gouws G, Fennesy ST, Kuguru B, Sauer WHH, Shaw PW, McKeown NJ. **2018**. Genetic analysis reveals harvested *Lethrinus nebulosus* in the Southwest Indian Ocean comprise two cryptic species. *ICES Journal of Marine Science* 75(4): 1465–1472. https://doi.org/10.1093/icesjms/ fsx245

Hecht T. **1977**. Contributions to the biology of the Cape gurnard, *Trigla capensis* (Pisces: Triglidae): age, growth and reproduction. *Zoologica Africana* 12(2): 373–382.

Hecht T. 1990. On the life history of Cape horse mackerel *Trachurus trachurus capensis* off the south-east coast of South Africa. South African Journal of Marine Science 9(1): 317–326. https://doi.org/10.2989/025776190784378907 Hecht T, Hecht A. 1978. A descriptive systematic study of the otoliths of the neopterygian marine fishes of South Africa.II. The delimitation of teleost orders, some systematic notes and a provisional new phyletic order sequence. *Transactions of the Royal Society of South Africa* 43(2): 199–218.

Heckel JJ. 1837. Ichthyologische Beiträge zu den Familien der Cottoiden, Scorpaenoiden, Gobioiden und Cyprinoiden. Annalen des Wiener Museums der Naturgeschichte 2(1): 143–164.

Heckel JJ. 1843. Ichthyologie von Syrien. In: Von Russegger
J (ed) Reisen in Europa, Asien und Afrika: mit besonderer
Rücksicht auf die naturwissenschaftlichen Verhältnisse der
betreffenden Länder unternommen in den Jahren 1835
bis 1841. Vol 1. Part 2. E. Stuttgart: Schweizerbart'sche
Verlagshandlung. 642 pp.

Heckel JJ. **1846**. Abbildungen und Beschreibungen der Fische Syriens (pp. 209–254). In: Von Russegger J (ed) *Reisen in Europa, Asien und Afrika: mit besonderer Rücksicht auf die naturwissenschaftlichen Verhältnisse der betreffenden Länder unternommen in den Jahren 1835 bis 1841*. Stuttgart: Schweizerbart'sche Verlagshandlung. 642 pp.

Hector J. **1874**. Notice of *Motella novae-zealandiae*, sp. nov. *Transactions and Proceedings of the New Zealand Institute* 6: 107–108.

Hector J. 1875. Descriptions of five new species of fishes obtained in New Zealand seas by H.M.S. *Challenger* expedition, July 1874. *Annals and Magazine of Natural History* (Series 4) 15(85): 78–82.

Heemstra E, Heemstra PC, Smale MJ, Hooper T, Pelicier D. **2004**. Preliminary checklist of coastal fishes from the Mauritian island of Rodrigues. *Journal of Natural History* 38(23–24): 3315–3350.

Heemstra PC. **1973**. *Anthias conspicuus* sp. nova (Perciformes: Serranidae) from the Indian Ocean, with comments on related species. *Copeia* 1973(2): 200–210.

Heemstra PC. **1980**. A revision of the zeid fishes (Zeiformes: Zeidae) of South Africa. *Ichthyological Bulletin of the J.L.B. Smith Institute of Ichthyology* 41: 1–18.

Heemstra PC. **1982**. Taxonomic notes on some triglid and peristediid fishes (Pisces: Scorpaeniformes) from southern Africa. *Copeia* 1982(2): 291–295.

Heemstra PC. **1984**. *Apolemichthys kingi*, a new species of angelfish (Pomacanthidae) from South Africa, with comments on the classification of angelfishes and a checklist of the pomacanthids of the western Indian Ocean. J.L.B. Smith Institute of Ichthyology Special Publication 35: 1–17.

Heemstra PC. **1984**. Mugiloididae. Sandsmelts, sandperches, grubfishes. In: Fischer W, Bianchi G (eds) *FAO species identification sheets for fishery purposes. Western Indian Ocean (Fishing Area 51)*. Vol. 3. Rome: FAO. [unpaginated]

Heemstra PC. **1986**. Family No. 162: Caproidae. In: Smith MM, Heemstra PC (eds) *Smiths' sea fishes*. Johannesburg: Macmillan South Africa. 1047 pp.

Heemstra PC. **1995**. Additions and corrections for the 1995 impression (pp. v–xv). In: Smith MM, Heemstra PC (eds) *Smiths' sea fishes* (3<sup>rd</sup> impression). Johannesburg: Southern Book Publishers.

Heemstra PC. **1995**. Family No. 97: Carapidae. In: Smith MM, Heemstra PC (eds) *Smiths' sea fishes* (3<sup>rd</sup> impression). Johannesburg: Southern Book Publishers. 1047 pp.

Heemstra PC. **1996**. A review of species of the the anthiine fish genus *Plectranthias* (Perciformes: Serranidae) from Mauritius. *Transactions of the Royal Society of South Africa* 51(1): 139–146.

Heemstra PC. **1999**. Order Zeiformes (pp. 2257–2261). In: Carpenter KE, Niem VH (eds) FAO species identification guide for fishery purposes. *The living marine resources of the western central Pacific.* Vol. 4. Bony fishes part 2 (pp. 2069–2790). Rome: FAO.

Heemstra PC. 2001. Suborder Acanthuroidei. Ephippidae.
Spadefishes (batfishes) (pp. 3611–3622). In: Carpenter
KE, Niem VH (eds) FAO species identification guide for
fishery purposes. The living marine resources of the western
central Pacific. Vol. 6. Bony fishes part 4 (pp. 3381–4218).
Rome: FAO.

Heemstra PC. **2004**. *Gymnothorax hansi*, a new species of moray eel (Teleostei: Anguilliformes: Muraenidae) from the Comoro Islands, Western Indian Ocean. *Zootaxa* 515: 1–7.

Heemstra PC. **2010**. Taxonomic review of the perciform fish genus *Acanthistius* from the east coast of southern Africa, with description of a new species and designation of a neotype for *Serranus sebastoides* Castelnau, 1861. *Zootaxa* 2352: 59–68. http://dx.doi.org/10.11646/zootaxa.2352.1.4

Heemstra PC, Akhilesh KV. **2012**. A review of the anthiine fish genus *Pseudanthias* (Perciformes: Serranidae) of the western Indian Ocean, with description of a new species and a key to the species. *aqua, International Journal of Ichthyology* 18(3): 121–164.

Heemstra PC, Fricke H, Hissman K, Schauer J, Smale MJ, Sink K. 2006. Interactions of fishes with particular reference to coelacanths in the canyons at Sodwana Bay and the St Lucia marine protected area of South Africa: coelacanth research. *South African Journal of Science* 102(9&10): 461–465.

Heemstra PC, Golani D. 1993. Clarification of the Indo-Pacific groupers (Pisces: Serranidae) in the Mediterranean Sea. *Israel Journal of Zoology* 39: 381–390.

Heemstra PC, Hecht T. **1986**. Dinopercidae, a new family for the percoid marine fish genera *Dinoperca* Boulenger and *Centrarchops* Fowler (Pisces: Perciformes). *Ichthyological Bulletin of the J.L.B. Smith Institute of Ichthyology* 51: 1–20.

Heemstra PC, Heemstra E. **2004**. *Coastal fishes of southern Africa*. Grahamstown: NISC and SAIAB. xxiv + 488 pp. Heemstra PC, Hissmann K, Fricke H, Smale MJ, Schauer
J. 2006. Fishes of the deep demersal habitat at Ngazidja (Grand Comoro) Island, Western Indian Ocean: coelacanth research. *South African Journal of Science* 102(9&10): 444–460.

Heemstra PC, Kannemeyer SX. **1984**. The families Trachipteridae and Radiicephalidae (Pisces, Lampriformes) and a new species of *Zu* from South Africa. *Annals of the South African Museum* 94(2): 13–39.

Heemstra PC, Nelson JS. **1984**. Percophidae. Duckbills. In: Fischer W, Bianchi G (eds) *FAO species identification sheets for fishery purposes. Western Indian Ocean (Fishing Area 51)*. Vol. 3. Rome: FAO. [unpaginated]

Heemstra PC, Parin NV. **1986**. Family No. 116: Exocoetidae. In: Smith MM, Heemstra PC (eds) *Smiths' sea fishes*. Johannesburg: Macmillan South Africa. 1047 pp.

Heemstra PC, Randall JE. **1977**. A revision of the Emmelichthyidae (Pisces: Perciformes). *Australian Journal of Marine and Freshwater Research* 28(3): 361–396.

Heemstra PC, Randall JE. **1979**. A revision of the anthiine fish genus *Sacura* (Perciformes: Serranidae) with descriptions of two new species. *J.L.B. Smith Institute of Ichthyology Special Publication* 20: 1–13.

Heemstra PC, Randall JE. 1984. Serranidae. Groupers, seabasses, rockcods, hinds, combers, coral trouts, lyretails.
In: Fischer W, Bianchi G (eds) *FAO species identification* sheets for fishery purposes. Western Indian Ocean (Fishing Area 51). Vol. 4. Rome: FAO. [unpaginated]

Heemstra PC, Randall JE. **1986**. Family No. 166: Serranidae.In: Smith MM, Heemstra PC (eds) *Smiths' sea fishes*.Johannesburg: Macmillan South Africa. 1047 pp.

Heemstra PC, Randall JE. **1993**. *Groupers of the world (family Serranidae, subfamily Epinephelinae): An annotated and illustrated catalogue of the grouper, rockcod, hind, coral grouper and lyretail species known to date.* Rome: FAO. viii + 382 pp.

Heemstra PC, Randall JE. **2009**. A review of the anthiine fish genus *Plectranthias* (Perciformes: Serranidae) of the western Indian Ocean, with description of a new species, and a key to the species. *Smithiana* Bulletin 10: 3–17, Pl. 1.

Heemstra PC, Smith MM. **1980**. Hexatrygonidae, a new family of stingrays (Myliobatiformes: Batoidea) from South Africa, with comments on the classification of batoid fishes. *Ichthyological Bulletin of the J.L.B. Smith Institute of Ichthyology* **43**: 1–17.

Heemstra PC, Smith MM. **1981**. *Pelagocephalus marki*, a new species of pufferfish (Tetraodontidae) from South Africa. *Bulletin of Marine Science* 31(4): 911–915.

Heemstra PC, Smith MM. 1983. A new species of the triggerfish genus *Xenobalistes* Matsuura (Tetraodontiformes: Balistidae) from South Africa. *J.L.B. Smith Institute of Ichthyology Special Publication* 26: 1–5.

Heemstra PC, Wright JE. **1986**. Two new species of clinid fishes (Perciformes: Clinidae) from South Africa. *J.L.B. Smith Institute of Ichthyology Special Publication* 40: 1–11.

Heimberg AM, Cowper-Sal-lari R, Sémon M, Donoghue PCJ, Peterson KJ. **2010**. microRNAs reveal the interrelationships of hagfish, lampreys, and gnathostomes and the nature of the ancestral vertebrate. *Proceedings of the National Academy of Sciences* 107(45): 19379–19383. http://dx.doi.org/10.1073/pnas.1010350107

Heithaus MR, Marshall GJ, Buhleier BM, Dill LM. 2001.Employing Crittercam to study habitat use and behavior of large sharks. *Marine Ecology Progress Series* 209: 307–310.

Helfman GS, Collette BB, Facey DE, Bowen BW. 2009. The diversity of fishes: biology, evolution, and ecology (2<sup>nd</sup> edition). Chichester: Wiley-Blackwell. 720 pp.

Hemprich FG, Ehrenberg CG. 1899. Symbolae physicae, seu icones adhue ineditae corporum naturalium novorum aut minus cognitorum quae ex itineribus per Libyam, Aegyptiam, Nubiam, Dongolam, Syriam, Arabiam et Habessiniam publico institutis sumptu (studio annis MDCCCXX – MDCCCXXV redierunt). Zoologica (Berlin) 1–17.

Henderson AC, Al-Oufi H, McIlwain JL. 2008. Survey, status and utilisation of the elasmobranch fisheries resources of the Sultanate of Oman. Final report (September 2004). Muscat, Oman: Sultan Qaboos University. 136 pp.

Henderson AC, McIlwain JL, Al-Oufi HS, Al-Sheili S. **2007**. The Sultanate of Oman shark fishery: species composition, seasonality and diversity. *Fisheries Research* 86(2–3): 159–168.

Henderson AC, Reeve AJ. 2011. Noteworthy elasmobranch records from Oman. *African Journal of Marine Science* 33(1): 171–175. http://dx.doi.org/10.2989/181423 2X.2011.572380

Henderson AC, Reeve AJ, Jabado RW, Naylor GJP. **2016**. Taxonomic assessment of sharks, rays and guitarfishes (Chondrichthyes: Elasmobranchii) from south-eastern Arabia, using the NADH dehydrogenase subunit 2 (NADH2) gene. *Zoological Journal of the Linnean Society* 176(2): 399–442. http://dx.doi.org/10.1111/zoj.12309

Henle FGJ. 1834. Ueber Narcine, eine neue Gattung electrischer Rochen nebst einer Synopsis der electrischen Rochen. Berlin. 44 pp.

Hennemann RM. **2001**. *Sharks and rays: Elasmobranch guide of the world*. Frankfurt: Ikan. 304 pp.

Hennig W. **1966**. *Phylogenetic systematics*. Urbana, Illinois: University of Illinois Press. 263 pp.

Hensley DA. 1986. Family No. 259: Bothidae. In: Smith MM, Heemstra PC (eds) *Smiths' sea fishes*. Johannesburg: Macmillan South Africa. 1047 pp.

Hensley DA. **1997**. An overview of the systematics and biogeography of the flatfishes. *Journal of Sea Research* 37(3–4): 187–194.

Hensley DA. **2001**. Family Pleuronectidae. Righteye flounders (pp. 3863–3877). In: Carpenter KE, Niem VH (eds) *FAO species identification guide for fishery purposes. The living marine resources of the western central Pacific.* Vol. 6. Bony fishes part 4 (pp. 3381–4218). Rome: FAO.

Hensley DA. **2003**. On the status of *Rhombus cocosensis* Bleeker, and description of a new species of *Engyprosopon* based on misidentification of this species (Pleuronectiformes: Bothidae). *Copeia* 2003(4): 833–837.

Hensley DA. **2005**. Revision of the genus *Asterorhombus* (Pleuronectiformes: Bothidae). *Copeia* 2005(3): 445–460.

Hensley DA, Ahlstrom EH. **1984**. Pleuronectiformes: relationships. In: Moser HG, Richards WJ, Cohen DM, Fahay MP, Kendall AW and Richardson SL (eds) Ontogeny and systematics of fishes. *American Society of Ichthyology* and Herpetology Special Publication 1: 670–687.

Hensley DA, Amaoka K. 1989. A redescription of *Pseudorhombus megalops*, with comments on *Cephalopsetta ventrocellata* (Osteichthyes: Pleuronectiformes: Paralichthyidae). *Proceedings of the Biological Society of Washington* 102(3): 577–585.

Hensley DA, Amaoka K. 2001. Family Bothidae. Lefteye flounders (pp. 3799–3841). In: Carpenter KE, Niem VH (eds) FAO species identification guide for fishery purposes. The living marine resources of the western central Pacific. Vol. 6. Bony fishes part 4 (pp. 3381–4218). Rome: FAO.

Hensley DA, Randall JE. **1983**. A new species of *Abudefduf* (Pisces: Pomacentridae) from the east coast of South Africa. *Occasional Papers of the Bernice Pauahi Bishop Museum of Polynesian Ethnology and Natural History* 25(6): 1–10.

Hensley DA, Randall JE. **1990**. A redescription of *Engyprosopon macrolepis* (Teleostei: Bothidae). *Copeia* 1990(3): 674–680.

Hensley DA, Randall JE. **2003**. A new flatfish of the Indo-Pacific genus *Asterorhombus* (Pleuronectiformes: Bothidae). *Smithiana* Special Publication 2: 1–9.

Hensley DA, Smale MJ. **1998**. A new species of the flatfish genus *Chascanopsetta* (Pleuronectiformes: Bothidae), from the coasts of Kenya and Somalia with comments on *C. lugubris. J.L.B. Smith Institute of Ichthyology Special Publication* 59: 1–16. [Apparently not published until 1998, Dec. 1997 on cover.]

Herald ES. **1940**. A key to the pipefishes of the Pacific American coasts with descriptions of new genera and species. *Report of the Allan Hancock Pacific Expeditions 1932–1938* 9(3): 51–64.

Herald ES. 1953. Family Syngnathidae: pipefishes (pp. 231–278). In: Schultz LP, Herald ES, Lachner EA, Welander AD, Woods LP (eds) *Fishes of the Marshall and Marianas Islands. Vol. 1. Families from Asymmetrontidae through Siganidae. Bulletin of the United States National Museum* 202: i–xxxii + 1–685, Pls. 1–74.

Herald ES. **1959**. From pipefish to seahorse—a study of phylogenetic relationships. *Proceedings of the California Academy of Sciences* (Series 4) 24(13): 465–473.

Herald ES, Randall JE. **1972**. Five new Indo-Pacific pipefishes. *Proceedings of the California Academy of Sciences* (Series 4) 39(11): 121–140.

Herler J, Bogorodsky SV, Suzuki T. 2013. Four new species of coral gobies (Teleostei: Gobiidae: Gobiodon), with comments on their relationships within the genus. Zootaxa 3709(4): 301–329. https://doi.org/10.11646/zootaxa.3709.4.1

Herler J, Hilgers H. **2005**. A synopsis of coral and coral-rock associated gobies (Pisces: Gobiidae) from the Gulf of Aqaba, northern Red Sea. a*qua, Journal of Ichthyology and Aquatic Biology* 10(3): 103–132.

Hermann J. 1783. Tabula affinitatum animalium olim academico specimine edita, nunc uberiore commentario illustrata com annotationibus ad historiam naturalem animalium augendam facientibus. Argentorati: J.G. Treuttel. 370 pp.

Herold D, Clark E. 1993. Monogamy, spawning, and skinshedding of the seamoth, *Eurypegasus draconis* (Pisces: Pegasidae). *Environmental Biology of Fishes* 37: 219–236.

Herre AWCT. **1923**. A review of the eels of the Philippine Archipelago. *Philippine Journal of Science* 23(2): 123–236, Pls. 1–11.

Herre AWCT. **1926**. Four new Philippine fishes. *Philippine Journal of Science* 31(4): 533–543.

Herre AWCT. **1927**. A new genus and three new species of Philippine fishes. *Philippine Journal of Science* 32(3): 413–415.

Herre AWCT. **1927**. Four new fishes from Lake Taal (Bombon). *Philippine Journal of Science* 34(3): 273–279.

Herre AWCT. **1927**. Gobies of the Philippines and the China Sea. *Monographs of the Bureau of Science, Manila, Philippine Islands* 23: 1–352.

Herre AWCT. **1932**. Five new Philippine fishes. *Copeia* 1932(3): 139–142.

Herre AWCT. **1933**. Twelve new Philippine fishes. *Copeia* 1933(1): 17–25.

Herre AWCT. **1934**. *Notes on fishes in the Zoological Museum of Stanford University.* 1. *The fishes of the Herre Philippine expedition of 1931 with descriptions of 17 new species.* Hong Kong: Newspaper Enterprise Ltd. 106 pp.

Herre AWCT. **1935**. New fishes obtained by the Crane Pacific Expedition. *Field Museum of Natural History* (Zoological Series) 18(12): 383–438.

Herre AWCT. **1936**. Fishes of the Crane Pacific Expedition. *Field Museum of Natural History* (Zoological Series) 21(353): 1–472.

Herre AWCT. **1936**. Fishes in the Zoölogical Museum of Stanford University, III. New genera and species of gobies

and blennies and a new *Myxus* from the Pelew Islands and Celebes. *Philippine Journal of Science* 59(2): 275–287.

Herre AWCT. **1939**. On a collection of fishes from Nanyo, the Japanese mandated islands. *Annotationes Zoologicae Japonenses* 18(4): 298–307.

Herre AWCT. **1939**. On a collection of littoral and freshwater fishes from the Andaman Islands. *Records of the Indian Museum* (Calcutta) 41(4): 327–372.

Herre AWCT. **1939**. The Philippine blennies. *Philippine Journal* of Science 70(4): 315–373, Pls. 1–5.

Herre AWCT. **1940**. Notes on fishes in the Zoölogical Museum of Stanford University, VII. New and rare Philippine gobies from the Herre 1936–37 Oriental Expedition, and in the collections of the Bureau of Science. *Philippine Journal of Science* 72(4): 357–369.

Herre AWCT. **1941**. A list of fishes known from the Andaman Islands. *Memoirs of the Indian Museum* 13(3): 331–403.

Herre AWCT. **1942**. A new genus and species of Gobiesocidae from the Philippines. *Stanford Ichthyological Bulletin* 2(4): 120–122.

Herre AWCT. **1944**. Notes on fishes in the Zoological Museum of Stanford University. XVII. New fishes from Johore and India. *Proceedings of the Biological Society of Washington* 57: 45–51.

Herre AWCT. **1945**. Notes on fishes in the Zoological Museum of Stanford University. XIV. A new genus and three new species of gobies from the Philippines. *Proceedings of the Biological Society of Washington* 58: 11–15.

Herre AWCT. **1945**. Notes on fishes in the Zoological Museum of Stanford University. XIX. Two new Philippine gobies, with key to the genera of gobies with vomerine teeth. *Proceedings of the Biological Society of Washington* **58**: 77–81.

Herre AWCT. **1945**. Notes on fishes in the Zoological Museum of Stanford University: XX, New fishes from China and India, a new genus, and a new Indian record. *Journal of the Washington Academy of Sciences* **35**(12): 399–404.

Herre AWCT. **1945**. Two new genera and four new gobies from the Philippines and India. *Copeia* 1945(1): 1–6.

Herre AWCT. **1951**. Six additions to the Philippine fish fauna including two new species. *Philippine Journal of Science* 79(3): 341–346.

Herre AWCT. **1952**. A review of the scorpaenoid fishes of the Philippines and adjacent seas. *Philippine Journal of Science* 80(4) [for December 1951]: 381–482.

Herre AWCT. **1953**. Check-list of Philippine fishes. U.S. Fish and Wildlife Service Research Report 20: 1–977.

Herre AWCT. **1955**. Remarks on the fish genus *Mirolabrichthys*, with description of a new species. *Copeia* 1955(3): 223–225.

Herring PJ. **1977**. Bioluminescence of marine organisms. *Nature* (Ocean Sciences Supplement) 267: 788–793. Herring PJ. **1992**. Bioluminescence of the oceanic apogonid fishes *Howella brodiei* and *Florenciella lugubris*. *Journal of the Marine Biological Association of the UK* 72(1): 139–148.

Hiatt RW, Strasberg DW. **1960**. Ecological relationships of the fish fauna on coral reefs of the Marshall Islands. *Ecological Monographs* 30(1): 65–127.

Hibino Y, Kimura S, Golani D. **2014**. A new ophichthid species from the Red Sea of the genus *Mixomyrophis*, formerly known as Atlantic genus. *Ichthyological Research* 62(2): 184–188. http://dx.doi.org/10.1007/s10228-014-0422-y

Hibino Y, Kimura S, Hoshino K, Hatooka K, McCosker JE.
2012. Validity of *Scolecenchelys aoki*, with a redescription of *Scolecenchelys gymnota* (Anguilliformes: Ophichthidae). *Ichthyological Research* 59(2): 179–188. http://dx.doi. org/10.1007/s10228-011-0269-4

Hibino Y, McCosker JE, Kimura S. **2013**. Redescription of a rare worm eel, *Muraenichthys macrostomus* Bleeker 1864, a senior synonym of *Skythrenchelys lentiginosa* Castle and McCosker 1999 (Anguilliformes: Ophichthidae, Myrophinae). *Ichthyological Research* 60(3): 227–231. http://dx.doi.org/10.1007/s10228-013-0337-z

Hidaka K, Iwatsuki Y, Randall JE. **2008**. A review of the Indo-Pacific bonefishes of the *Albula argentea* complex, with a description of a new species. *Ichthyological Research* 55(1): 53–64. http://dx.doi.org/10.1007/s10228-007-0010-5

Hidaka K, Kishimoto H, Iwatsuki Y. **2004**. A record of an albulid fish, *Albula glossodonta*, from Japan (Albuliformes: Albulidae). *Japanese Journal of Ichthyology* 51(1): 61–66.

Higgins BE, Mori K, Uyeno T. 1970. Distribution of Scornbrotabrax heterolepis Roule (Order Perciformes) in the Pacific and Indian Oceans. Japanese Journal of Ichthyology 17(1): 51–53.

Hilgendorf FM. **1878**. Über das Vorkommen einer *Brama*art und einer neuen Fischgattung *Centropholis* aus der Nachbarschaft des genus *Brama* in den japanischen Meeren. *Sitzungs-berichte der Gesellschaft Naturforschender Freunde zu Berlin* 1878: 1–2.

Hilgendorf FM. **1880**. Üebersicht über die japanischen Sebastes-Arten. Sitzungs-berichte der Gesellschaft Naturforschender Freunde zu Berlin 1880(1): 166–172. [In German]

Hill BJ. **1966**. A contribution to the ecology of the Umlazi Estuary. *Zoologica Africana* 2(1): 1–24.

Hilton EJ. **2001**. Tongue-bite apparatus of osteoglossomorph fishes: variation of a character complex. *Copeia* 2001(2): 372–382.

Hilton EJ. 2002. Osteology of the extant North American fishes of the genus *Hiodon* Lesueur 1818 (Teleostei: Osteoglossomorpha: Hiodontiformes). *Fieldiana Zoology* (New Series) 100: 1–142.

Hilton EJ. **2003**. Comparative osteology and phylogenetic systematics of fossil and living bony-tongue fishes

(Actinopterygii, Teleostei, Osteoglossomorpha). Zoological Journal of the Linnean Society 137(1): 1–100.

Hilton EJ. **2011**. Bony fish skeleton (pp. 434–448). In: Farrell AP (ed) *Encyclopedia of fish physiology: from genome to environment*. Volume 1: The senses, supporting tissues, reproduction, and behavior. San Diego: Academic Press. 2163 pp.

Hipeau-Jacquotte R. 1967. Notes de faunistique et biologie marines de Madagascar. IV. Observation sur le comportement du poisson Carapidae: *Carapus hornei* (Richardson, 1844) de Madagascar. *Recueils des Travaux de la Station Marine d'Endoume* Fasc. 6: 141–151.

Hirasaka K, Nakamura H. **1947**. On the Formosan spearfishes. *Bulletin of the Oceanographic Institute Taiwan* 3: 9–24, Pls. 1–3.

Hiyama Y. **1940**. Descriptions of two new species of fish, *Raja tobitukai* and *Chlorophthalmus acutifrons*. *Japanese Journal of Zoology* 9(1): 169–173.

Ho H-C. **2013**. Redescription of *Parapercis punctata* (Cuvier, 1829) and status of *Neosillago* Castelnau, 1875 and its type species *Neosillago marmorata* Castelnau, 1875 (Perciformes: Pinguipedidae). *Zootaxa* 3736(3): 291–299. https://doi.org/10.11646/zootaxa.3736.3.7

Ho H-C, Heemstra PC, Imamura H. 2014. A new species of the sandperch genus *Parapercis* from the western Indian Ocean (Perciformes: Pinguipedidae). *Zootaxa* 3802(3): 335–345. http://dx.doi.org/10.11646/zootaxa.3802.3.3

Ho H-C, Johnson GD. **2012**. *Protoblepharon mccoskeri*, a new flashlight fish from eastern Taiwan (Teleostei: Anomalopidae). *Zootaxa* 3479: 77–87. http://dx.doi. org/10.11646/zootaxa.3479.1.5

Ho H-C, Last PR. **2013**. Two new species of the coffinfish genus *Chaunax* (Lophiiformes: Chaunacidae) from the Indian Ocean. *Zootaxa* 3710(5): 436–448. http://dx.doi.org/10.11646/zootaxa.3710.5.3

Ho H-C, Ma WC. **2016**. Revision of the southern African species of the anglerfish genus *Chaunax* (Lophiiformes: Chaunacidae), with descriptions of three new species. *Zootaxa* 4144(2): 175–194. http://dx.doi.org/10.11646/ zootaxa.4144.2.2

Ho H-C, McCosker JE, Smith DG. 2013. Revision of the worm eel genus *Neenchelys* (Ophichthidae; Myrophinae), with descriptions of three new species from the western Pacific Ocean. *Zoological Studies* 52: 1–20. http://dx.doi. org/10.1186/1810-522X-52-58

Ho H-C, Prokofiev AM, Shao K-T. **2010**. *Synodus cresseyi* Prokofiev, 2008, an unnecessary replacement for *S. macrocephalus* Cressey, 1981, with description of a new species from the Western Indian Ocean (Teleostei: Synodontidae). *Zootaxa* 2419: 63–68. http://dx.doi. org/10.11646/zootaxa.2419.1.3 Ho H-C, Prokofiev AM, Shao K-T. **2009**. A new species of the batfish genus *Malthopsis* (Lophiiformes: Ogcocephalidae) from the northwestern Indian Ocean. *Zoological Studies* 48(3): 394–401.

Ho H-C, Shao K-T. **2010**. A review of *Malthopsis jordani* Gilbert, 1905, with description of a new batfish from the Indo-Pacific Ocean (Lophiiformes: Ogcocephalidae). *Bulletin of the National Museum of Nature and Science* (Tokyo) (Series A) Supplement No. 4: 9–19.

Hobson E. **1974**. Feeding relationships of teleostean fishes on coral reefs in Kona, Hawaii. *Fishery Bulletin* 72: 915–1031.

Hoda SMS. **1980**. A contribution to the gobioid fishes of Pakistan. *Proceedings of the 1st Pakistan Congress of Zoology* B: 469–482.

Hoda SMS. **1983**. A new species of gobiid fish *Monishia bulejienis* [sic] (family: Gobiidae) from the coast of Pakistan. *Biologia* (Lahore) 29(1): 111–114.

Hoda SMS. **1983**. *Enneapterygius nasimae* a new species of tripterygiid fish from Karachi coast, northern Arabian Sea. *Indian Journal of Fisheries* 30(1): 116–123.

Hoda SMS, Goren M. 1990. Bathygobius karachiensis (Gobiidae), a new species from Pakistan. Cybium 14(2): 143–150.

Hodge JR, Bellwood DR. 2016. The geography of speciation in coral reef fishes: the relative importance of biogeographical barriers in separating sister-species. *Journal of Biogeography* 43(7): 1324–1335. http://dx.doi.org/10.1111/jbi.12729

Hodnett J-PM, Grogan ED, Lund R, Lucas SG, Suazo T, Elliott DK, Pruitt J. 2021. Ctenacanthiform sharks from the Late Pennsylvanian (Missourian) Tinajas member of the Atrasado Formation, Central New Mexico. In: Lucas SG, DiMichele WA, Allen BD (eds) *Kinney Brick Quarry Lagerstätte. New Mexico Museum of Natural History and Science Bulletin* 84: 391–424.

Hoese DF. **1986**. Descriptions of two new species of *Hetereleotris* (Pisces: Gobiidae) from the western Indian Ocean, with discussion of related species. *J.L.B. Smith Institute of Ichthyology Special Publication* **41**: 1–25.

Hoese DF, Bogorodsky SV, Mal AO. 2015. Description of a new species of *Trimma* (Perciformes: Gobiidae) from the Red Sea, with a discussion of the generic separation of *Trimma* and *Priolepis*, with discussion of sensory papillae terminology. *Zootaxa* 4027(4): 538–550. https://doi.org/10.11646/zootaxa.4027.4.4

Hoese DF, Bray DJ, Paxton JR, Allen GR. **2006**. *Zoological catalogue of Australia*. Volume 35. Fishes. Parts 1–3. Collingwood: CSIRO Publishing. [Part 1, pp. i–xxiv + 1–670; Part 2, pp. i–xxi + 671–1472; Part 3, pp. i–xxi + 1473–2173.]

Hoese DF, Fourmanoir P. 1978. Discordipinna griessingeri, a new genus and species of gobiid fish from the tropical Indo-West Pacific. Japanese Journal of Ichthyology 25(1): 19–24. Hoese DF, Larson HK. **1994**. Revision of the Indo-Pacific gobiid fish genus *Valenciennea*, with descriptions of seven new species. *Indo-Pacific Fishes* 23: 1–71.

Hoese DF, Larson HK. 2006. Gobiidae (pp. 1612–1697). In: Hoese DF, Bray DJ, Paxton JR, Allen GR (eds) *Zoological catalogue of Australia*. Vol. 35. Fishes. Part 3 (pp. 1473–2173). Collingwood: CSIRO Publishing.

Hoese DF, Randall JE. **1982**. Revision of the gobiid fish genus *Stonogobiops. Indo-Pacific Fishes* 1: 1–18.

Hoese DF, Reader S. **1985**. A new gobiid fish, *Fusigobius duospilus*, from the tropical Indo-Pacific. *J.L.B. Smith Institute of Ichthyology Special Publication* 36: 1–9.

Hoese DF, Winterbottom R. **1979**. A new species of *Lioteres* (Pisces, Gobiidae) from Kwazulu, with a revised checklist of South African gobies and comments on the generic relationships and endemism of western Indian Ocean gobioids. *Royal Ontario Museum Life Sciences Occasional Paper* 31: 1–13.

Hollard HLGM. **1854**. Monographie de la famille des Balistides. Suite 2. *Annales des Sciences Naturelles, Paris* (Zoologie) (Série 4) 1: 39–72, 303–339.

Holleman W. 1982. Three new species and a new genus of tripterygiid fishes (Blennioidei) from the Indo-West Pacific Ocean. *Annals of the Cape Provincial Museums* (Natural History) 14(4): 109–137.

Holleman W. 1991. A revision of the tripterygiid fish genus Norfolkia Fowler, 1953 (Perciformes: Blennioidei).
Annals of the Cape Provincial Museums (Natural History) 18(11): 227–243.

Holleman W. 2005. A review of the tripplefin fish genus Enneapterygius (Blennioidei: Tripterygiinae) in the western Indian Ocean, with descriptions of four new species. Smithiana, Publications in Aquatic Biodiversity Bulletin 5: 1–25.

Holleman W. **2006**. Fishes of the *Helcogramma steinitzi* species group (Blennioidei: Tripterygiidae) from the Indian Ocean, with descriptions of two new species. *aqua, Journal of Ichthyology and Aquatic Biology* 11(3): 89–104.

Holleman W. **2007**. Fishes of the genus *Helcogramma* (Blennioidei: Tripterygiidae) in the western Indian Ocean, including Sri Lanka, with descriptions of four new species. *Smithiana* Bulletin 7: 51–81.

Holleman W, Bogorodsky SV. **2012**. A review of the blennioid fish family Tripterygiidae (Perciformes) in the Red Sea, with description of *Enneapterygius qirmiz*, and reinstatement of *Enneapterygius altipinnis* Clark, 1980. *Zootaxa* 3152: 36–60. http://dx.doi.org/10.11646/zootaxa.3152.1.2

Holleman W, Buxton CD. **1993**. *Acanthanectes*, a new genus of triplefin with two new species from the southern coast of South Africa (Blennioidei: Tripterygiidae). *Cybium* 17(4): 327–342.

Holleman W, Connell AD, Carpenter KE. 2013. Caesio xanthalytos, a new species of fusilier (Perciformes: Caesionidae) from the Western Indian Ocean, with records of range extensions for several species of Caesionidae. Zootaxa 3702(3): 262–272. https://doi.org/10.11646/ zootaxa.3702.3.4

Holleman W, Von der Heyden S, Zsilavecz G. 2012.
Delineating the fishes of the *Clinus superciliosus* species complex in southern African waters (Blennioidei: Clinidae: Clinini), with the validation of *Clinus arborescens* Gilchrist & Thompson, 1908 and *Clinus ornatus* Gilchrist & Thompson, 1908, and with descriptions of two new species. *Zoological Journal of the Linnean Society* 166(4): 827–853. http://dx.doi.org/10.1111/j.1096-3642.2012.00865.x

Hollister G. **1937**. Caudal skeleton of Bermuda shallow water fishes. II. Order Percamorphi, suborder Percesoces: Atherinidae, Mugilidae, Sphyraenidae. *Zoologica* (N.Y.) 22(3): 265–279.

Holly M. **1929**. Drei neue Fischformen aus Persien. *Anzeiger der Akademie der Wissenschaften in Wien* 66(7): 62–64.

Holly M. **1933**. *Cyclostomata*. Das Tierreich 59. Berlin: De Gruyter. xiv + 62 pp.

Holt EWL, Byrne LW. **1908**. New deep-sea fishes from the south-west coast of Ireland. *Annals and Magazine of Natural History* (Series 8) 1(1): 86–95.

Holt EWL, Byrne LW. **1909**. Preliminary note on some fishes from the Irish Atlantic slope. *Annals and Magazine of Natural History* (Series 8) 3(15): 279–280.

Holthuis LB, Pietsch TW. 2006. Les Planches inédites de Poissons et autres Animaux marins de l'Indo-Ouest Pacifique d'Isaac Johannes Lamotius [Isaac Johannes Lamotius (1646– c. 1718) and His Paintings of Indo-Pacific Fishes and Other Marine Animals]. Paris: Publications Scientifiques du Muséum, Muséum National d'Histoire naturelle. 292 pp.

Hopley D. **1982**. *The geomorphology of the Great Barrier Reef: Quaternary development of coral reefs*. New York: Wiley-Interscience. 453 pp.

Hora SL. **1925**. Notes on fishes in the Indian Museum. VII. On a new genus of "Globe-fishes" (fam. Tetraodontidae). *Records of the Indian Museum* (Calcutta) 26(6): 579–582.

Hora SL. **1926**. Notes on fishes in the Indian Museum. IX–XIV. *Records of the Indian Museum* (Calcutta) 27(6): 453–469.

Hora SL. **1926**. On a new species of the genus *Ctenotrypauchen* Steindachner. *Records of the Indian Museum* (Calcutta) 28(4): 221–223.

Hora SL, Misra KS. **1943**. On a small collection of fishes from Iraq. *Journal of the Asiatic Society of Bengal* 9(1): 1–15.

Hora SL, Mukerji DD. **1936**. Notes on the fishes in the Indian Museum. XXVII. On two collections of fish from Maungmagan, Tavoy District, Lower Burma. *Records of the Indian Museum* (Calcutta) 38(1): 15–39. Horn MH. **1972**. Systematic status and aspects of the ecology of the elongate ariommid fishes (suborder Stromateoidei) in the Atlantic. *Bulletin of Marine Science* 22(3): 537–558.

Horn MH. **1973**. Systematic comparison of the stromateid fishes *Stromateus brasilienses* Fowler and *Stromateus stellatus* Cuvier from coastal South America with a review of the genus. *Bulletin of the British Museum (Natural History) Zoology* 24(7): 319–339.

Horn MH. **1984**. Stromateoidei: development and relationships. In: Moser HG, Richards WJ, Cohen DM, Fahay MP, Kendall SW Jr, Richardson SL (eds) Ontogeny and systematics of fishes. *American Society of Ichthyologists and Herpetologists Special Publication* 1: 620–628.

Hornell J. **1923**. The flying-fish fishery of the Coromandel coast and the spawning habits of *Cypsilurus*. *Madras Fisheries Bulletin* 15(4): 99–109.

Hornell J, Fowler HW. **1922**. Description of a new gobioid fish from Tuticorin. *Journal of the Bombay Natural History Society* 28(4): 924–925.

Horrill JC, Kamukuru AT, Mgaya YD, Risk M. 2000. Chapter
6. Northern Tanzania, Zanzibar and Pemba (pp. 167–198).
In: McClanahan TR, Sheppard CRC, Obura DO (eds) Coral reefs of the Indian Ocean: Their ecology and conservation.
Oxford: Oxford University Press. xxiii + 525 pp.

Hoshino K. **2001**. Monophyly of the Citharidae (Pleuronectoidei: Pleuronectiformes: Teleostei) with considerations of pleuronectoid phylogeny. *Ichthyological Research* 48(4): 391–404.

Hoshino K, Amaoka K. 1998. Description of a juvenile specimen of the rarely-caught *Samaris macrolepis* (Pleuronectiformes, Samaridae), from off northwest Australia. *Ichthyological Research* 45(4): 401–403.

Hoshino K, Amaoka K, Last PR. 2000. Description of transformation larvae and a juvenile of the dextral flounder, *Poecilopsetta praelonga* (Pleuronectidae, Poecilopsettinae) from the northwestern waters of Australia. *Ichthyological Research* 47(3): 263–270.

Houde ED. **1981**. Distribution and abundance of four types of codlet (Pisces: Bregmacerotidae) larvae from the eastern Gulf of Mexico. *Biological Oceanography* 1(1): 81–104.

Houde ED. **1984**. Bregmacerotidae: development and relationships. In: Moser HG, Richards WJ, Cohen DM, Fahay MP, Kendall SW Jr, Richardson SL (eds) Ontogeny and systematics of fishes. *American Society of Ichthyologists and Herpetologists Special Publication* 1: 300–308.

Houttuyn M. 1764–1765. Natuurlyke historie of uitvoerige beschryving der dieren, planten en mineraalen, volgens het samenstel van den Heer Linnaeus (met naauwkeurige afbeeldingen). 3 Vols. in 37 parts; Fishes, Parts 7 (1764; vi + 446 pp) and 8 (1765; viii + 525 + 67 (unnumbered) + 30 pp). Amsterdam. Houttuyn M. **1782**. Beschryving van eenige Japanese visschen, en andere zee-schepzelen. *Verhandelingen der Hollandsche Maatschappij der Wetenschappen* (Haarlem) 20(pt 2): 311–350.

 Howe JC, Springer VG. 1993. Catalog of type specimens of recent fishes in the National Museum of Natural History, Smithsonian Institution. 5: Sharks (Chondrichthyes: Selachii). Smithsonian Contributions to Zoology 540: 1–19.

Howell BR, Yamashita Y. 2005. Aquaculture and stock enhancement (pp. 347–372). In: Gibson RN (ed) *Flatfishes: Biology and exploitation*. Oxford: Blackwell Science. xxiv + 391 pp.

Hubbs CL. **1915**. Flounders and soles from Japan collected by the United States Bureau of Fisheries steamer *Albatross* in 1906. *Proceedings of the United States National Museum* 48(2082): 449–496, Pls. 25–27.

Hubbs CL. **1929**. *Oostethus*: a new generic name for a doryrhamphine pipefish. *Occasional Papers of the Museum of Zoology University of Michigan* 199: 1–4.

Hubbs CL. **1941**. A new family of fishes. *Journal of the Bombay Natural History Society* **42**: 446–447.

Hubbs CL. **1944**. Species of the circumtropical fish genus *Brotula*. *Copeia* 1944(3): 162–178.

Hubbs CL. **1945**. Phylogenetic position of the Citharidae, a family of flatfishes. *Miscellaneous Publications of the Museum of Zoology, University of Michigan* 63: 1–38.

Hubbs CL. 1951. Allosebastes, new subgenus for Sebastodes sinensis, scorpaenid fish of the Gulf of California.
Proceedings of the Biological Society of Washington 64: 129–130

Hubbs CL. **1952**. A contribution to the classification of blennioid fishes of the family Clinidae, with a partial revision of the Eastern Pacific forms. *Stanford Ichthyological Bulletin* 4(2): 41–165.

Hubbs CL. **1958**. *Dikellorhynchus* and *Kanazawaichthys*: Nominal fish genera interpreted as based on prejuveniles of *Malacanthus* and *Antennarius*, respectively. *Copeia* 1958(4): 282–285.

Hubbs CL, Hubbs LC. **1945**. Bilateral asymmetry and bilateral variation in fishes. *Papers of the Michigan Academy of Science, Arts and Letters* 30: 229–310.

Hubbs CL, Ishiyama R. **1968**. Methods for the taxonomic study and description of skates (Rajidae). *Copeia* 1968(3): 483–491.

Hubbs CL, Iwai T, Matsubara K. **1967**. External and internal characters, horizontal and vertical distribution, luminescence, and food of the dwarf pelagic shark, *Euprotomicrus bispinatus. Bulletin of the Scripps Institution of Oceanography* 10: 1–64.

Hubbs CL, Lagler KF. **1964**. *Fishes of the Great Lakes Region*. Ann Arbor, Michigan: University of Michigan Press. 213 pp. Hubbs CL, Wisner RL. **1980**. Revision of the sauries (Pisces: Scomberesocidae) with descriptions of two new genera and one new species. *Fishery Bulletin* 77(3): 521–566.

Huber JH. 1996. Killi-Data 1996. Updated checklist of taxonomic names, collecting localities and bibliographic references of oviparous Cyprinodont fishes (Atherinomorpha, Pisces). Paris: Société Française d'Ichtyologie, Muséum National d'Histoire Naturelle. 399 pp.

Hubrecht AAW. **1879**. *Catalogue des collections formées et laissées par M.-P. Bleeker*. Leiden: De Breuk & Smits. iv + 71 pp.

Hulley PA. **1966**. The validity of *Raja rhizacanthus* Regan and *Raja pullopunctata* Smith, based on a study of the clasper. *Annals of the South African Museum* 48(20): 497–514.

Hulley PA. **1969**. The relationship between *Raja miraletus* Linnaeus and *Raja ocellifera* Regan based on a study of the clasper. *Annals of the South African Museum* 52(6): 137–147.

Hulley PA. **1970**. An investigation of the Rajidae of the west and south coasts of southern Africa. *Annals of the South African Museum* 55(4): 151–220.

Hulley PA. **1971**. *Centrophorus squamosus* (Bonnaterre) (Chondrichthyes, Squalidae) in the eastern south Atlantic. *Annals of the South African Museum* 57(11): 265–270.

Hulley PA. 1972. The origin, interrelationships and distribution of southern African Rajidae (Chondrichthyes, Batoidei). *Annals of the South African Museum* 60(1): 1–103.

Hulley PA. 1972. The rare plectognath fish, Macrorhamphosodes uradoi (Kamohara) (Triacanthodidae) in South African waters. Annals of the South African Museum 60(5): 191–195.

Hulley PA. **1972**. A report on the mesopelagic fishes collected during the deep-sea cruises of R.S. *Africana II*, 1961–1966. *Annals of the South African Museum* 60(6): 197–236.

Hulley PA. **1972**. Mesopelagic fishes from Vema Seamount (1K Station 52). *Annals of the South African Museum* 60(7): 237–244.

Hulley PA. **1972**. A new species of southern African brevirajid skate (Chondrichthyes, Batoidei, Rajidae). *Annals of the South African Museum* 60(9): 253–263.

Hulley PA. 1973. Interrelationships within the Anacanthobatidae (Chondrichthyes, Rajoidea) with a description of the lectotype of *Anacanthobatis marmoratus*, von Bonde & Swart, 1923. *Annals of the South African Museum* 62(4): 131–158.

Hulley PA, Rau RE. **1969**. A female *Regalecus glesne* from Cape Province, South Africa. *Copeia* 1969(4): 835–839.

Hulley PA, Stehmann M. **1977**. The validity of *Malacoraja* Stehmann, 1970 (Chondrichthyes, Batoidei, Rajiidae) and its phylogenetic significance. *Annals of the South African Museum* 72(12): 227–237. Human BA. **2006**. A taxonomic revision of the catshark genus *Poroderma* Smith, 1837 (Chondrichthyes: Carcharhiniformes: Scyliorhinidae). *Zootaxa* 1229: 1–32.

Human BA. 2006. A taxonomic revision of the catshark genus *Holohalaelurus* Fowler 1934 (Chondrichthyes: Carcharhiniformes: Scyliorhinidae), with descriptions of two new species. *Zootaxa* 1315: 1–56.

Human BA. **2006**. Size-corrected shape variation analysis and quantitative species discrimination in a morphologically conservative catshark genus, *Poroderma* Smith, 1837 (Chondrichthyes: Carcharhiniformes: Scyliorhinidae). *African Natural History* 2: 1–15.

Human BA. **2007**. A taxonomic revision of the catshark genus *Haploblepharus* Garman 1913 (Chondrichthyes: Carcharhiniformes: Scyliorhinidae). *Zootaxa* 1451: 1–40.

Human BA. **2007**. Size corrected shape variation analysis and quantitative species discrimination in a morphologically conserved catshark genus, *Haploblepharus* Garman 1913 (Chondrichthyes: Carcharhiniformes: Scyliorhinidae). *African Natural History* 3: 59–73.

Human BA. **2007**. Size-corrected shape variation analysis and quantitative species discrimination in a morphologically conserved catshark genus, *Holohalaelurus* Fowler 1934 (Chondrichthyes: Carcharhiniformes: Scyliorhinidae). *African Natural History* 3: 75–88.

Human BA. **2010**. Range extension and a further female specimen of the grinning izak (*Holohalaelurus grennian* Human 2006; Scyliorhinidae; Chondrichthyes). *Smithiana* Bulletin 11: 25–33.

Human BA, Compagno LJV. 2006. Description of *Haploblepharus kistnasamyi*, a new catshark (Chondrichthyes: Scyliorhinidae) from South Africa. *Zootaxa* 1318: 41–58.

Human BA, Owen EP, Compagno LJV, Harley EH. **2006**. Testing morphologically based phylogenetic theories within the cartilaginous fishes with molecular data, with special reference to the catshark family (Chondrichthyes; Scyliorhinidae) and the interrelationships within them. *Molecular Phylogenetics and Evolution* **39**(2): **384–391**.

Hureau J-C. **1969**. Remarques sur les poissons des iles Saint-Paul et Amsterdam. *Bulletin du Muséum National d'Histoire Naturelle* (Paris) (Série 2) 40(6) [for 1968]: 1150–1161.

Hureau J-C, Litvinenko NI. **1986**. Scorpaenidae (pp. 1211– 1229). In: Whitehead PJP, Bauchot M-L, Hureau J-C, Nielsen J, Tortonese E (eds) *Fishes of the north-eastern Atlantic and the Mediterranean*. Vol. III. Paris: UNESCO.

Hureau J-C, Monod T (eds). 1973. Check-list of the fishes of the north-eastern Atlantic and of the Mediterranean (CLOFNAM). 2 volumes. Paris: UNESCO. Vol. I: xxii + 683 pp.; Vol. II: 331 pp. Hureau J-C, Nielsen JG. **1981**. Les poissons Ophidiiformes des campagnes du N.O. *Jean Charcot* dans l'Atlantique et la Mediterranée. *Cybium* (Série 3) 5(3): 3–27.

Hurley IA, Mueller RL, Dunn KA, Schmidt EJ, Friedman M, Ho RK, Prince VE, Yang Z, Thomas MG, Coates MI. 2007.A new time-scale for ray-finned fish evolution. *Proceedings* of the Royal Society B 274(1609): 489–498.

Hussain S, Jawad LA. 2014. First records of *Opisthognathus* muscatensis Boulenger, 1888 (Opisthognathidae), Trachinotus baillonii (Lacepède, 1801) (Carangidae), and Atrobucca nibe (Jordan & Thompson, 1911) (Sciaenidae) off the Iraq Coast, Arabian Gulf. International Journal of Marine Science 4(No. 28): 253–258.

Hutchings K, Griffiths MH. **2005**. Identity and distribution of southern African sciaenid fish species of the genus *Umbrina*. *African Journal of Marine Science* 27(1): 1–21.

Hutchins JB. **1977**. Descriptions of three new genera and eight new species of monacanthid fishes from Australia. *Records of the Western Australian Museum* 5(1): 3–58.

Hutchins JB. **1981**. Description of a new species of serranid fish from Western Australia, with a key to the Australian species of *Acanthistius*. *Records of the Western Australian Museum* 8(4): 491–499.

Hutchins JB. **1981**. Nomenclatural status of the toadfishes of India. *Copeia* 1981(2): 336–341.

Hutchins JB. **1984**. Batrachoididae. Toadfishes. In: Fischer W, Bianchi G (eds) *FAO species identification sheets for fishery purposes. Western Indian Ocean (Fishing Area 51).* Vol. 1. Rome: FAO. [unpaginated]

Hutchins JB. **1986**. Review of the monacanthid fish genus *Pervagor*, with descriptions of two new species. *Indo-Pacific Fishes* 12: 37 pp.

Hutchins JB. **1988**. The comparative morphology and phylogeny of the monacanthid fishes. Unpublished PhD thesis, Murdoch University, Western Australia.

Hutchins JB. **1994**. Description of a new genus and species of monacanthid fish from India. *Records of the Western Australian Museum* 16(4): 567–574.

Hutchins JB. **1997**. Review of the monacanthid fish genus *Paramonacanthus*, with descriptions of three new species. *Records of the Western Australian Museum* Supplement No. 54: 1–57.

Hutchins JB. **2001**. Checklist of the fishes of Western Australia. *Records of the Western Australian Museum* Supplement No. 63: 9–50.

Hutchins JB. **2002**. Description of a new genus and species of miniature monacanthid fish from the Seychelles and Marshall Islands. *Records of the Western Australian Museum* 21(2): 213–219.

Hutchins JB, Kuiter RH. 1982. A new species of *Acanthistius* (Pisces: Serranidae) from eastern Australia. *Records of the Western Australian Museum* 10(2): 127–131. Hutchins JB, Matsuura K. 1984. Description of a new monacanthid fish of the genus *Thamnaconus* from Fiji. *Records of the Western Australian Museum* 11(4): 387–391.

Hutchins JB, Randall JE. **1982**. *Cantherhines longicaudus*, a new filefish from Oceania, with a review of the species of the *C. fronticinctus* complex. *Pacific Science* 36(2): 175–185.

Hutchins JB, Swainston R. 1985. Revision of the monacanthid fish genus Brachaluteres. Records of the Western Australian Museum 12(1): 57–78.

Hutton FW. **1872**. *Fishes of New Zealand. Catalogue with diagnoses of the species*. Colonial Museum and Geological Survey Department, Wellington. 192 pp.

Hutton FW. **1873**. Contributions to the ichthyology of New Zealand. *Transactions and Proceedings of the New Zealand Institute* 1872 5(art.28): 259–272.

Hutton FW. **1875**. Descriptions of new species of New Zealand fish. *Annals and Magazine of Natural History* (Series 4) 16(95): 313–317.

Hutton FW, Hector J. 1872. *Fishes of New Zealand*. Wellington: Colonial Museum and Geological Survey Department. 133 pp.

Hyde JR, Humphreys R Jr, Musyl M, Lynn E, Vetter R. **2006**. A central North Pacific spawning ground for striped marlin, *Tetrapturus audax*. *Bulletin of Marine Science* 79(3): 683–690.

Hyde JR, Vetter RD. 2006. The origin, evolution, and diversification of rockfishes of the genus *Sebastes* (Cuvier). *Molecular Phylogenetics and Evolution* 44(2): 790–811.

## I

Ibarra M, Stewart DJ. **1987**. Catalog of type specimens of recent fishes in Field Museum of Natural History. *Fieldiana Zoology* (New Series) 35: 1–112.

Ida H. **1973**. Extra ossicles in the oral region on three species of *Bleekeria* (Ammodytidae). *Japanese Journal of Ichthyology* 20(2): 67–72.

Ida H, Sirimontaporn P, Monkolprasit S. **1994**. Comparative morphology of the fishes of the family Ammodytidae, with a description of two new genera and two new species. *Zoological Studies* 33(4): 251–277.

IFAD (International Fund for Agricultural Development).2010. Fisheries Investment Project, Republic of Yemen.Project final design report. [unpaginated]

IGFA (International Game Fish Association). 2009. World record game fishes. Dania Beach, Florida. 408 pp.

Iglésias SP. **2013**. *Apristurus nakayai* sp. nov., a new species of deepwater catshark (Chondrichthyes: Pentanchidae) from New Caledonia. *Cybium* 36(4) [for 2012]: 511–519. https://doi.org/10.26028/cybium/2012-364-003

Iglésias SP, Lecointre G, Sellos DY. **2005**. Extensive paraphylies within the sharks of the order Carcharhiniformes inferred from nuclear and mitochondrial genes. *Molecular Phylogenetics and Evolution* 34(3): 569–583. https://doi. org/10.1016/j.ympev.2004.10.022

Iglésias SP, Nakaya K, Stehmann M. **2004**. *Apristurus melanoasper*, a new species of deep-water catshark from the North Atlantic (Chondrichthyes: Carcharhiniformes: Scyliorhinidae). *Cybium* 28(4): 345–356.

Imai S. **1954**. On two flying-fishes of the genus *Hirundichthys* Breder and their juveniles from Japan. *Kagoshimadaigaku Suisangakabu Kiyo* 3(2): 62–72. [In Japanese]

Imai S. **1955**. On the life history of *Cypselurus naresii* Günther obtained in Japan. *Memoirs of the Faculty of Fisheries, Kagoshima University* 4: 97–104.

Imai S. 1959. Studies on the life histories of the flying fishes found in the adjacent waters of Japan. *Memoirs of the Faculty of Fisheries, Kagoshima University* 7(part 1): 1–85.

Imamura H. **1996**. Phylogeny of the family Platycephalidae and related taxa (Pisces: Scorpaeniformes). *Species Diversity* 1(2): 123–233.

Imamura H. **1997**. *Platycephalus americanus* Sauvage, 1878, a junior synonym of *Grammoplites scaber* (Linnaeus, 1758) (Scorpaeniformes: Platycephalidae). *Ichthyological Research* 44(4): 425–428.

Imamura H. **2003**. *Sunagocia*, a replacement name for the platycephalid genus *Eurycephalus* (Actinopterygii), with taxonomic comments on the species of the genus. *Species Diversity* 8(3): 301–306.

Imamura H. 2004. Phylogenetic relationships and new classification of the superfamily Scorpaenoidea (Actinopterygii: Perciformes). Species Diversity 9(1): 1–36.

Imamura H. 2009. Family accounts: Muraenidae (pp. 17–21), Ophichthidae (p. 21), Congridae (pp. 22–23), Aploactinidae (p. 71), Platycephalidae (pp. 74–77), Pempheridae (p. 177), Pinguipedidae (pp. 240–242), Tripterygiidae (pp. 243–244), Acanthuridae (pp. 301–307). In: Kimura S, Satapoomin U, Matsuura K (eds) *Fishes of Andaman Sea, west coast of southern Thailand*. i–vi + 346 pp.

Imamura H. 2015. Taxonomic revision of the flathead fish genus *Platycephalus* Bloch, 1785 (Teleostei: Platycephalidae) from Australia, with description of a new species. *Zootaxa* 3904(2): 151–207. https://doi.org/10.11646/zootaxa.3904.2.1

Imamura H. **2016**. Rediagnosis of *Onigocia grandisquama* (Actinopterygii: Perciformes: Platycephalidae) and comparison with congeners. *Species Diversity* 21(2): 151–159. http://dx.doi.org/10.12782/sd.21.2.151

Imamura H, Amaoka K. **1994**. A new species of flathead, *Grammoplites knappi* (Scorpaeniformes: Platycephalidae) from the South China Sea. *Japanese Journal of Ichthyology* 41(2): 173–179. Imamura H, Amaoka K. **1996**. *Rogadius serratus* (Cuvier, 1829), a senior synonym of *R. polijodon* (Bleeker, 1853) (Scorpaeniformes: Platycephalidae). *Ichthyological Research* 43(1): 97–100.

Imamura H, Ida H, Moyer JT. **1995**. Redescription of a flathead, *Thysanophrys otaitensis* (Scorpaeniformes, Platycephalidae). *Japanese Journal of Ichthyology* 42(3/4): 277–283.

Imamura H, Knapp LW. **1997**. A new species of deepwater flathead, *Bembras adenensis* (Scorpaeniformes: Bembridae) from the western Indian Ocean. *Ichthyological Research* 44(1): 9–14.

Imamura H, Knapp LW. **1998**. Review of the genus *Bembras* Cuvier, 1829 (Scorpaeniformes: Bembridae) with description of three new species collected from Australia and Indonesia. *Ichthyological Research* 45(2): 165–178.

Imamura H, Knapp LW. **1999**. *Thysanophrys papillaris*, a new species of flathead from the Andaman Sea and northern Australia (Scorpaeniformes: Platycephalidae). *Ichthyological Research* 46(2): 179–183.

Imamura H, Matsuura K. **2003**. Record of a sandperch, *Parapercis xanthozona* (Actinopterygii: Pinguipedidae), from Japan, with comments on its synonymy. *Species Diversity* 8(1): 27–33.

Imamura H, Psomadakis P, Thein H. **2018**. Two new species of deepwater flathead *Bembras* Cuvier, 1829 from the Andaman Sea, eastern Indian Ocean. *Ichthyological Research* 65: 488–495. https://doi.org/10.1007/s10228-018-0639-2

Imamura H, Sakashita M. 1997. Redescription of a rare flathead, *Onigocia grandisquamma* (Regan, 1908) (Scorpaeniformes: Platycephalidae). *Ichthyological Research* 44: 119–124.

Imamura H, Shinohara G. 1997. Phylogenetic studies of the order Scorpaeniformes (Pisces: Acanthopterygii): progress, present condition and problems. *Japanese Journal* of *Ichthyology* 44(2): 77–95. [In Japanese with English summary]

Imamura H, Shinohara G. 1998. Scorpaeniform fish phylogeny: an overview. Bulletin of the National Science Museum (Tokyo) (Series A) 24(3): 185–212.

Imamura H, Shinohara G. **2004**. A new species of *Cocotropus* (Actinopterygii: Teleostei: Aploactinidae) from South Africa, western Indian Ocean. *Species Diversity* 9(3): 201–205.

 Imamura H, Shinohara G. 2019. First record of *Onigocia* grandisquama (Scorpaeniformes: Platycephalidae) from Japan. Japanese Journal of Ichthyology 66(2): 155–160.
 [In Japanese] https://doi.org/10.11369/jji.18-050

Imamura H, Shirai SM, Yabe M. **2005**. Phylogenetic position of the family Trichodontidae (Teleostei: Perciformes), with a revised classification of the perciform suborder Cottoidei. *Ichthyological Research* 52(3): 264–274. Imamura H, Yabe M. 2002. Demise of the Scorpaeniformes (Actinopterygii: Percomorpha): an alternative phylogenetic hypothesis. *Bulletin of Fisheries Sciences, Hokkaido* University 53(3): 107–128.

Imamura H, Yoshino T. 2007. Three new species of the genus *Parapercis* from the western Pacific, with redescription of *Parapercis hexophtalma* (Perciformes: Pinguipedidae). *Bulletin of the National Museum of Nature and Science* (Tokyo) (Series A) Supplement No.1: 81–100.

Inada T. **1981**. Studies on the merlucciid fishes. *Bulletin of the Far Seas Fisheries Research Laboratory* 18: 1–172.

Inger RF. **1955**. A revision of the fishes of the genus *Plesiops* Cuvier. *Pacific Science* 9(3): 259–276.

Inoue JG, Miya M, Tsukamoto K, Nishida M. **2003**. Basal actinopterygian relationships: a mitogenomic perspective on the phylogeny of the "ancient fish". *Molecular Phylogenetics and Evolution* 26(1): 110–120.

Inoue JG, Miya M, Venkatesh B, Nishida M. **2005**. The mitochondrial genome of Indonesian coelacanth *Latimeria menadoensis* (Sarcopterygii: Coelacanthiformes) and divergence time estimation between the two coelacanths. *Gene* 349: 227–235.

Inoue T, Nakabo T. **2006**. The *Saurida undosquamis* group (Aulopiformes: Synodontidae), with description of a new species from southern Japan. *Ichthyological Research* 53: 379–397.

International Commission on Zoological Nomenclature.
1987. Opinion 1439. *Cephalopholis argus* Schneider,
1801 and *Serranus sexmaculatus* (currently *Cephalopholis sexmaculata*) Rüppell, 1830 (Osteichthyes): conserved. *Bulletin of Zoological Nomenclature* 44(2): 142–143.

International Commission on Zoological Nomenclature. **2003**. Opinion 2056 (Case 3186). *Squalus edwardsii* (currently *Haploblepharus edwardsii*; Chondrichthyes, Carcharhiniformes): attributed to Schinz, 1822 and *edwardsii* conserved as the correct original spelling of the specific name. *Bulletin of Zoological Nomenclature* 60(3): 250.

Irisson J-O, Paris CB, Guigand C, Planes S. **2010**. Vertical distribution and ontogenetic 'migration' in coral reef fish larvae. *Limnology and Oceanography* 55(2): 909–919. http://dx.doi.org/10.4319/lo.2010.55.2.0909

Irvine SB, Stevens JD, Laurenson LJB. **2006**. Surface bands on deepwater squalid dorsal-fin spines: an alternative method for ageing *Centroselachus crepidater*. *Canadian Journal of Fisheries and Aquatic Sciences* 63(3): 617–627.

Ishida M. **1994**. Phylogeny of the suborder Scorpaenodei (Pisces: Scorpaeniformes). *Bulletin of the Nansei National Fisheries Research Institute* 27: 1–112. Ishida M, Amaoka K. **1992**. A new species of the fish genus *Idiastion* (Pisces: Scorpaenidae) from the Kyushu-Palau Ridge, western Pacific. *Japanese Journal of Ichthyology* 38(4): 357–360.

Ishihara H. **1987**. Revision of the western North Pacific species of the genus *Raja*. *Japanese Journal of Ichthyology* 34(3): 241–285.

Ishihara H, Homma K, Nakamura R. **2001**. The occurrence of albinism in individuals of the manta ray and Japanese common skate found in the western Pacific. *I.O.P. Diving News* 12(7): 2–5.

Ishihara H, Taniuchi T, Tanaka S, Srivastava MP. 1998. Investigation of the freshwater elasmobranchs in the River Ganges (pp. 41–55). In: Tanaka S (ed) Adaptability and conservation of freshwater elasmobranchs. Report of Research Project, Grant-in-Aid for International Scientific Research (Field Research) in the financial year of 1996 and 1997. 119 pp.

Ishii N, Imamura H. 2008. Phylogeny of the family Congiopodidae (Perciformes: Scorpaenoidea), with a proposal of new classification. *Ichthyological Research* 55(2): 148–161. http://dx.doi.org/10.1007/s10228-007-0014-1

Ishiyama R. **1958**. Studies on the rajid fishes (Rajidae) found in the waters around Japan. *Journal of the Shimonoseki College of Fisheries* 7(2–3): 191–394.

Ito RY, Hawn DR, Collette BB. **1994**. First record of the butterfly kingfish *Gasterochisma melampus* (Scombridae) from the North Pacific Ocean. *Japanese Journal of Ichthyology* 40(4): 482–486.

IUCN (International Union for Conservation of Nature). 2003–2021. IUCN Red List of Threatened Species (by year). http://www.iucnredlist.org

Ivanin NA. 1989. Morphometric characters of the alfoncino, *Beryx splendens*, from submarine ridges in the temperate zone of the Indian Ocean. *Journal of Ichthyology* 29(4): 159–162.

Ivanova NV, Zemlak TS, Hanner RH, Hebert PDN. 2007. Universal primer cocktails for fish DNA barcoding. *Molecular Ecology Notes* 7(4): 544–548.

Ivantsoff W. 1984. Atherinidae. Silversides. In: Fischer W, Bianchi G (eds) FAO species identification sheets for fishery purposes. Western Indian Ocean (Fishing Area 51). Vol 1. Rome: FAO. [unpaginated]

Ivantsoff W, Crowley LELM. 1991. Review of the Australian silverside fishes of the genus *Atherinomorus* (Atherinidae). *Australian Journal of Marine and Freshwater Research* 42(5): 479–505.

Ivantsoff W, Crowley LELM. 1999. Atherinidae. Silversides (or hardyheads) (pp. 2113–2139). In: Carpenter KE, Niem VH (eds) FAO species identification guide for fisheries purposes. The living marine resources of the western central Pacific. Vol. 4. Bony fishes part 2 (pp. 2069–2790). Rome: FAO. [also cited as 2000] Iversen ES, Yoshida HO. **1957**. Notes on the biology of the wahoo in the Line Islands. *Pacific Science* **11**(4): 370–379.

Iwai T, Asano H. 1958. On the luminous cardinal fish, *Apogon ellioti* Day. *Scientific Report, Yokosuka City Museum* (3): 5–13.

Iwamoto T. **1970**. Macrourid fishes of the Gulf of Guinea. The R/V *Pillsbury* deep-sea biological expedition to the Gulf of Guinea, 1964–65. 19. *Studies in Tropical Oceanography* 4(part 2): 316–431.

Iwamoto T. **1973**. Macrourids (Gadiformes: Pisces) collected off Angola by the R/V *Undaunted*, with the description of a new species. *Proceedings of the Biological Society of Washington* 86(31): 373–384.

Iwamoto T. 1999. Bregmacerotidae. Codlets (codlings) (pp. 1997–1998). In: Carpenter K, Niem VH (eds) FAO species identification guide for fishery purposes. The living marine resources of the western central Pacific. Vol. 3. Batoid fishes, chimaeras and bony fishes part 1 (pp. 1397–2068). Rome: FAO.

Iwamoto T, Staiger JC. **1976**. Percophidid fishes of the genus *Chrionema* Gilbert. *Bulletin of Marine Science* 26(4): 488–498.

Iwamoto T, Stein DL. **1974**. A systematic review of the rattail fishes (Macrouridae: Gadiformes) from Oregon and adjacent waters. *Occasional Papers of the California Academy of Sciences* 111: 1–79.

Iwata A, Hosoya S, Niimura Y. **1998**. *Echinogobius hayashii*, a new genus and species of Gobiidae. *Ichthyological Research* 24(2): 113–119.

Iwatsuki Y. **2013**. Review of the *Acanthopagrus latus* complex (Perciformes: Sparidae) with descriptions of three new species from the Indo-West Pacific Ocean. *Journal of Fish Biology* 83(1): 64–95. https://doi.org/10.1111/jfb.12151

Iwatsuki Y, Akazaki M, Yoshino T. **1993**. Validity of a lutjanid fish, *Lutjanus ophuysenii* (Bleeker) with a related species, *L. vitta* (Quoy et Gaimard). *Japanese Journal of Ichthyology* 40(1): 47–59.

Iwatsuki Y, Al-Mamry JM, Heemstra PC. 2016. Validity of a blue stripe snapper, *Lutjanus octolineatus* (Cuvier 1828) and a related species, *L. bengalensis* (Bloch 1790) with a new species (Pisces; Lutjanidae) from the Arabian Sea. *Zootaxa* 4098(3): 511–528. https://doi.org/10.11646/zootaxa.4098.3.5

Iwatsuki Y, Bogorodsky SV, Tanaka F, Mal AO, Ali AH. 2015. Range extension of *Gerres infasciatus* (Perciformes: Gerreidae) from the Red Sea and the Arabian Gulf, with distributional implications for the *G. filamentosus* complex. *Cybium* 39(2): 155–160. https://doi.org/10.26028/ cybium/2015-392-006

Iwatsuki Y, Carpenter KE. **2006**. *Acanthopagrus taiwanensis*, a new sparid fish (Perciformes), with comparisons to *Acanthopagrus berda* (Forsskål, 1775) and other nominal species of *Acanthopagrus*. *Zootaxa* 1202: 1–19.
Iwatsuki Y, Carpenter KE. **2009**. *Acanthopagrus randalli* (Perciformes: Sparidae), a new black seabream from the Persian Gulf. *Zootaxa* 2267: 43–54. http://dx.doi. org/10.11646/zootaxa.2267.1.3

Iwatsuki Y, Djawad MI, Burhanuddin AI, Motomura H, Hidaka K. **2000**. A preliminary list of the epipelagic and inshore fishes of Makassar (= Ujung Panang), South Sulawesi, Indonesia, collected mainly from fish markets between 23–27 January 2000, with notes on fishery catch statistics. *Bulletin of the Faculty of Agriculture, Miyazaki University* 47(1–2): 95–114.

Iwatsuki Y, Heemstra PC. 2001. Gerres phaiya: new species of gerreid fish (Teleostei: Perciformes: Gerreidae) from India, with comments on Gerres poieti and the Gerres erythrourus Complex. Copeia 2001(4): 1043–1049.

Iwatsuki Y, Heemstra PC. 2007. A new gerreid fish species and redescription of *Gerres maldivensis* Regan, 1902 from the Indian Ocean (Perciformes: Gerreidae). *Copeia* 2007(1): 85–92.

Iwatsuki Y, Heemstra PC. **2010**. Taxonomic review of the western Indian Ocean species of the genus *Acanthopagrus* Peters, 1855 (Perciformes: Sparidae), with description of a new species from Oman. *Copeia* 2010(1): 123–136. http://dx.doi.org/10.1643/CI-08-073

Iwatsuki Y, Heemstra PC. 2011. A review of the Acanthopagrus bifasciatus species complex (Pisces: Sparidae) from the Indian Ocean, with redescriptions of A. bifasciatus (Forsskål 1775) and A. catenula (Lacepède 1801). Zootaxa 3025: 38–50. https://doi.org/10.11646/zootaxa.3025.1.2

Iwatsuki Y, Heemstra PC. **2011**. *Polysteganus mascarenensis*, a new sparid fish species from Mascarene Islands, Indian Ocean. *Zootaxa* 3018: 13–20. https://doi.org/10.11646/ zootaxa.3018.1.2

Iwatsuki Y, Heemstra PC. 2015. Redescriptions of *Polysteganus coeruleopunctatus* (Klunzinger 1870) and *P. lineopunctatus* (Boulenger 1903), with two new species from Western Indian Ocean. *Zootaxa* 4059(1): 133–150. https://doi.org/10.11646/zootaxa.4059.1.7

Iwatsuki Y, Heemstra PC. **2018**. Taxonomic review of the genus *Argyrops* (Perciformes; Sparidae) with three new species from the Indo-West Pacific. *Zootaxa* 4438(3): 401–442. http://dx.doi.org/10.11646/zootaxa.4438.3.1

Iwatsuki Y, Jawad LA, Al-Mamry JM. 2012. Johnius (Johnius) majan sp. nov., a sciaenid fish (Pisces: Sciaenidae) from Oman, Indian Ocean. Ichthyological Research 59(2): 151– 155. http://dx.doi.org/10.1007/s10228-011-0265-8

Iwatsuki Y, Jawad LA, Tanaka F, Al-Busaidi HJ, Al-Mamry M, Al-Kharusi LH. **2013**. Omani fishes collected in the vicinity of Mutrah, Gulf of Oman and Madrakah, southern Oman through 3 to 13 October 2010. *Bulletin of the Faculty of Agriculture, Miyazaki University* 59: 29–43. Iwatsuki Y, Kimura S. **1997**. *Gerres methueni* Regan, 1920, a senior synonym of *G. rappi* (Barnard, 1927) (Pisces: Gerreidae). *Ichthyological Research* 44(1): 1–7.

Iwatsuki Y, Kimura S. **1998**. A new species, *Gerres infasciatus*, from the Gulf of Thailand (Perciformes: Gerreidae). *Ichthyological Research* 45(1): 79–84.

Iwatsuki Y, Kimura S, Kishimoto H, Yoshino T. **1996**. Validity of the gerreid fish, *Gerres macracanthus* Bleeker, 1854, with designation of a lectotype, and designation of a neotype for *G. filamentosus* Cuvier, 1829. *Ichthyological Research* 43(4): 417–429.

Iwatsuki Y, Kimura S, Yoshino T. 1999. Redescriptions of Gerres baconensis (Evermann & Seale, 1907), G. equulus Temminck & Schlegel, 1844 and G. oyena (Forsskål, 1775), included in the "G. oyena complex", with notes on other related species (Perciformes: Gerreidae). Ichthyological Research 46(4): 377–395.

Iwatsuki Y, Kimura S, Yoshino T. 2001. Gerres limbatus Cuvier and G. lucidus Cuvier from the Indo-Malay Archipelagos, the latter corresponding to young of the former (Perciformes: Gerreidae). Ichthyological Research 48(3): 307–314.

Iwatsuki Y, Kimura S, Yoshino T. 2001. Redescription of *Gerres* longirostris (Lacepède, 1801) and *G. oblongus* Cuvier in Cuvier and Valenciennes, 1830, included in the *Gerres* longirostris complex (Pisces: Perciformes: Gerreidae). Copeia 2001(4): 945–965.

Iwatsuki Y, Kimura S, Yoshino T. **2002**. A new species: *Gerres microphthalmus* (Perciformes: Gerreidae) from Japan with notes on limited distribution, included in the "*G. filamentosus* complex". *Ichthyological Research* 49(2): 133–139.

Iwatsuki Y, Kimura S, Yoshino T. 2006. A new sparid, Acanthopagrus akazakii, from New Caledonia with notes on nominal species of Acanthopagrus. Ichthyological Research 53(4): 406–414.

Iwatsuki Y, Kimura S, Yoshino T. **2007**. A review of the *Gerres subfasciatus* complex from the Indo-West Pacific, with three new species (Perciformes: Gerreidae). *Ichthyological Research* 54: 168–185.

Iwatsuki Y, Kume M, Yoshino T. 2010. A new species, Acanthopagrus pacificus from the western Pacific (Pisces, Sparidae). Bulletin of the National Museum of Nature and Science (Tokyo) (Series A) 36(4): 115–130.

Iwatsuki Y, Maclaine J. 2013. Validity of Crenidens macracanthus Günther 1874 (Pisces: Sparidae) from Chennai (Madras), India, with taxonomic statuses of the congeners. Ichthyological Research 60(3): 241–248. http://dx.doi.org/10.1007/s10228-013-0342-2

Iwatsuki Y, Pogonoski JJ, Last P. **2012**. Revision of the genus *Parequula* (Pisces: Gerreidae) with a new species from southwestern Australia. *Zootaxa* 3425: 42–54. http://dx.doi. org/10.11646/zootaxa.3425.1.3

Iwatsuki Y, Tanaka F, Allen GR. 2015. Lutjanus xanthopinnis, a new species of snapper (Pisces: Lutjanidae) from the Indo-west Pacific, with a redescription of Lutjanus madras (Valenciennes 1831). Journal of the Ocean Science Foundation 17: 22–42. http://dx.doi.org/10.5281/ zenodo.1051774

## J

Jaafar Z, Murdy EO (eds). **2017**. *Fishes out of water: Biology and ecology of mudskippers*. Boca Raton: CRC Press. xxx + 390 pp. https://doi.org/10.1201/9781315119861

Jaafar Z, Parenti LR. **2016**. Systematics of the mudskipper genus *Oxuderces* Eydoux & Souleyet 1848 (Teleostei: Gobiidae: Oxudercinae) with resurrection from synonymy of *O. nexipinnis* (Cantor 1849). *Zoological Journal of the Linnean Society* 180(1): 195–215. https://doi.org/10.1111/ zoj.12482

Jaafar Z, Randall JE. **2009**. A pictorial review and key to the shrimp gobies of the genus *Amblyeleotris* of the Red Sea, with description of a new species. *Smithiana* Bulletin 10: 23–29.

Jabado RW, Ebert DA. **2015**. *Sharks of the Arabian seas: An identification guide.* Dubai: The International Fund for Animal Welfare. 240 pp.

Jabado RW, Kyne PM, Pollom RA, Ebert DA, Simpfendorfer CA, Ralph GM, Dulvy NK. **2017**. *The conservation status of sharks, rays, and chimaeras in the Arabian Sea and adjacent waters.* Vancouver: Environment Agency, Abu Dhabi and IUCN Species Survival Commission Shark Specialist Group. 236 pp.

Jacob PG, Qasim SZ. **1974**. Mud of a mud bank in Kerala, southwest coast of India. *Indian Journal of Marine Science* 3: 115–119.

Jacobsen IP, Bennett MB. **2007**. Description of a new species of catshark, *Atelomycterus marnkalha* n. sp. (Carcharhiniformes: Scyliorhinidae) from north-east Australia. *Zootaxa* 1520: 19–36.

Jacobsen IP, Bennett MB. **2009**. A taxonomic review of the Australian butterfly ray *Gymnura australis* (Ramsay & Ogilby, 1886) and other members of the family Gymnuridae (Order Rajiformes) from the Indo-West Pacific. *Zootaxa* 2228: 1–28. http://dx.doi.org/10.11646/zootaxa.2228.1.1

Jacquinot H, Guichenot A. **1853**. Reptiles et poissons. In: Dumont d'Urville J, *Voyage au Pôle Sud et dans l'Oceanie sur les corvettes* L'Astrolabe *et* La Zélée. 3(2): 1–56, Pls. 1–7.

Jaeger EC. **1978**. A source-book of biological names and terms (3<sup>rd</sup> edition). Springfield, Illinois: Charles C. Thomas. xxxv + 323 pp.

Jahn AE, Haedrich RL. **1987**. Notes on the pelagic squaloid shark *Isistius brasiliensis*. *Biological Oceanography* 5(4): 297–309.

James GD. **1976**. *Cyttus traversi* Hutton: juvenile form of *C. ventralis* Barnard and Davies (Pisces: Zeidae). *Journal of the Royal Society of New Zealand* 6(4): 49–498.

James GD, Inada T, Nakamura I. **1988**. Revision of the oreosomatid fishes (family Oreosomatidae) from the southern oceans, with a description of a new species. *New Zealand Journal of Zoology* 15: 291–326.

James PSBR. **1962**. Observations on shoals of the Javanese cownose ray *Rhinoptera javanica* Müller & Henle from the Gulf of Mannar, with additional notes on the species. *Journal of the Marine Biological Association of India* 4(2): 217–223.

James PSBR. **1966**. Notes on the biology and fishery of the butterfly ray, *Gymnura poecilura* (Shaw) from the Palk Bay and Gulf of Mannar. *Indian Journal of Fisheries* 13(1&2): 150–157.

James PSBR. **1967**. The ribbon-fishes of the family Trichiuridae of India. *Marine Biological Association of India Memoir* 1: 1–228.

James PSBR. **1969**. A new species of silver-belly, *Leiognathus jonesi* (Family Leiognathidae: Pisces) from the Indian seas. *Journal of the Marine Biological Association of India* 11(1&2): 316–319.

James PSBR. **1970**. Further observations on shoals of the Javanese cownose ray *Rhinoptera javanica* Müller & Henle from the Gulf of Mannar with a note on the teeth structure in the species. *Journal of the Marine Biological Association of India* 12(1&2): 151–157.

James PSBR. **1980**. Some observations on the ray *Himantura marginatus* (Blyth) from the Gulf of Mannar. *Journal of the Marine Biological Association of India* 22(1&2): 161–164.

James PSBR. **1984**. Leiognathidae. Ponyfishes, slipmouths, toothponies. In: Fischer W, Bianchi G (eds) *FAO species identification sheets for fishery purposes. Western Indian Ocean (Fishing Area 51)*. Vol. 2. Rome: FAO. [unpaginated]

James PSBR, Badrudeen M. **1991**. A new species of silverbelly *Leiognathus striatus* (Family Leiognathidae: Pisces) from the Gulf of Mannar, India and redescription of *Leiognathus fasciatus* (Lacépède). *Journal of the Marine Biological Association of India* 32(1&2): 217–226.

Jamieson BGM, Grier HJ. **1993**. Influences of phylogenetic position and fertilization biology on spermatozoal ultrastructure exemplified by exocoetoid and poeciliid fish. *Hydrobiologia* 271(1): 11–25.

Janvier P. **1978**. Les nageoires paires des Ostéostracés et la position systématique des Cephalaspidomorphes. *Annales de Paleontologie (Vertebrata)* 64: 113–142.

Janvier P. **1981**. The phylogeny of the Craniata, with particular reference to the significance of fossil "agnathans." *Journal of Vertebrate Paleontology* 1(2): 121–159.

Janvier P. **1996**. *Early vertebrates*. Oxford, UK: Oxford University Press. 408 pp.

Janvier P. **2002**. *Early vertebrates* (reprint of 1996 Oxford monographs on geology and geophysics 33). New York: Oxford Science Publications. 393 pp.

Janvier P. **2008**. Early jawless vertebrates and cyclostome origins. *Zoological Science* 25(10): 1045–1056. http://dx.doi. org/10.2108/zsj.25.1045

Janvier P. **2010**. microRNAs revive old views about jawless vertebrate divergence and evolution. *Proceedings of the National Academy of Sciences* 107(45): 19137–19138. http://dx.doi.org/10.1073/pnas.1014583107

Janvier P, Pradel A. 2016. Elasmobranchs and their extinct relatives: diversity, relationships and adaptations through time (pp. 1–17). In: Shadwick R, Farrell A, Brauner C (eds) Physiology of elasmobranch fishes: structure and interaction with environment. *Fish Physiology* (Series). Vol. 34A. Academic Press. 422 pp. http://dx.doi. org/10.1016/B978-0-12-801289-5.00001-8

Jardas I. **1996**. *Jadranska ihtiofauna*. Zagreb: Školska knijga. 533 pp.

Jatzow R, Lenz H. **1898**. Fische von Ost Afrika, Madagaskar und Aldabra. *Abhandlungen der Senckenbergischen Naturforschenden Gessellschaft* 21(3): 497–531.

Jawad LA, Al-Badri ME, Fricke R. **2014**. New records of thicklips and grunts from the marine waters of Iraq (Teleostei: Haemulidae). *Journal of the Ocean Science Foundation* 12: 18–24. http://dx.doi.org/10.5281/ zenodo.1049121

Jawad LA, Al-Kharusi L, Al-Mamry J. **2011**. On the occurrence of the Egyptian seahorse *Hippocampus suezensis* Duncker, 1940 in Muscat, Sultanate of Oman. *Acta Adriatica* 52(1): 137–140.

Jawad LA, Al-Mamry J. **2009**. First record of *Antennarius coccineus* from the Gulf of Oman and second record of *Antennarius indicus* from the Arabian Sea coast of Oman. *Marine Biodiversity Records* 2(e163): 1–3. http://dx.doi.org/10.1017/S1755267209990923

Jawad LA, Al-Mamry JM. **2013**. New record of the toothpony, *Gazza minuta* (Osteichthyes: Leiognathidae) from the coasts of Muscat City at the sea of Oman, Sultanate of Oman. *Thalassia Salentina* 35: 3–9.

Jawad LA, Al-Mamry JM, Al-Mamary DS, Al-Rasady EH.
2013. First record of the slender ponyfish, *Equulites elongatus* (Günther, 1874) (Family: Leiognathidae) from the coasts of Muscat city at the Sea of Oman, Sultanate of Oman. *Journal of Applied Ichthyology* 29(2): 456–459. http://dx.doi.org/10.1111/jai.12004

 Jawad LA, Ibrahim M. 2014. Confirmed record of whitebarred rubberlip, *Plectorhinchus playfairi* (Pellegrin, 1914) (Pisces: Haemulidae) from Jubail, Saudi Arabia, Arabian Gulf. *International Journal of Marine Science* 4(No. 21): 194–196. Jayabalan N. **1986**. A new species of silverbelly, *Gazza shettyi* (Pisces: Leiognathidae) from the Bay of Bengal. *Matsya* 11 [for 1985]: 42–45.

Jayaprakash AA, Kurup BM, Sreedhar U, Venu S, Divya I, Manjebrayakath H, Pachu VA, Thampy P, Sudhakar S. **2006**. Distribution, diversity, length-weight relationship and recruitment pattern of deep-sea finfishes and shellfishes in the shelf-break area off southwest Indian EEZ. *Journal of the Marine Biological Association of India* 48(1): 56–67.

Jayaram KC. **1981**. *The freshwater fishes of India, Pakistan, Bangladesh, Burma and Sri Lanka – a handbook*. Calcutta: Zoological Survey of India. 475 pp.

Jayaram KC. 1982. Aid to the identification of the siluroid fishes of India, Burma, Sri Lanka, Pakistan and Bangladesh.
5. Ariidae and Plotosidae. *Records of the Zoological Survey of India, Occasional Paper* 37: 1–41.

Jayaram KC. **1984**. Ariidae (often Tachysuridae and sometimes Bagreidae in the literature). Sea catfishes. In: Fischer W, Bianchi G (eds) *FAO species identification sheets for fishery purposes. Western Indian Ocean (Fishing Area 51)*. Vol. 1. Rome: FAO. [unpaginated]

Jayaram KC. **1999**. *The freshwater fishes of the Indian region*. New Delhi: Narendra Publishing House. 551 pp.

Jayaram KC. **2006**. *Catfishes of India*. New Delhi: Narendera Publishing House. 383 pp.

Jayaram KC. **2010**. *The freshwater fishes of the Indian region* (2<sup>nd</sup> edition). New Delhi: Narendra Publishing House. 616 pp.

Jayaram KC, Dhanze JR. **1979**. Siluroid fishes of India, Burma and Ceylon: 22. A preliminary review of the genera of the family Ariidae (Pisces: Siluroidea). *Matsya* 4 [for 1978]: 42–51.

Jayaram KC, Dhanze JR. **1981**. Siluroid fishes of India, Burma, and Ceylon. 23. The specific status of *Tachysurus malabaricus* (Day) (Ariidae: Siluriformes). *Bulletin of the Zoological Survey of India* 4(1): 121–123.

Jayaram KC, Dhanze JR. **1981**. Siluroid fishes of India, Burma, and Ceylon. 24. The systematic status of *Arius satparanus* Chaudhuri (Ariidae: Siluriformes). *Bulletin of the Zoological Survey of India* 4(3): 395–398.

Jayawardane PAAT, Dayaratne P. **1998**. A preliminary analysis of the flyingfish fishery off Kandakuliya in Sri Lanka. *Ceylon Journal of Science* (Biological Sciences) 26: 18–28.

Jehangeer MI. **1984**. National report for Mauritius. *UNEP Regional Seas Reports and Studies* 50: 77–206.

Jenkins JT. **1910**. Notes on fish from India and Persia, with descriptions of new species. *Records of the Indian Museum* (Calcutta) 5(art.12): 123–140.

Jenkins JT. **1910**. Report on the fishes taken by the Bengal fisheries steamer *Golden Crown* Part IV. Pleuronectidae. *Memoirs of the Indian Museum* 3(1): 23–31.

Jenkins OP. **1901**. Descriptions of fifteen new species of fishes from the Hawaiian Islands. *Bulletin of the U.S. Fish Commission* 19(1899): 387–404.

Jenkins OP. **1903**. Report on collections of fishes made in the Hawaiian Islands, with descriptions of new species. *Bulletin of the U.S. Fish Commission* 22(1902): 417–511.

Jennings S, Marshall S, Cuet P, Naim O. **2000**. Chapter 13. The Seychelles (pp. 383–410). In: McClanahan TR, Sheppard CRC, Obura DO (eds) *Coral reefs of the Indian Ocean: Their ecology and conservation*. Oxford: Oxford University Press. xxiii + 525 pp.

Jenyns L. 1840–1842. Fish. In: Darwin C (ed) The zoology of the voyage of H.M.S. Beagle, under the command of Captain Fitzroy, R.N., during the years 1832 to 1836 (issued in 4 parts). London: Smith, Elder, & Co. xvi + 172 pp.

Jeong CH, Nakabo T. **2008**. *Dipturus wuhanlingi*, a new species of skates (Elasmobranchi; Rajidae) from China. *Ichthyological Research* 55: 183–190.

Jeong CH, Nakabo T, Wu H-L. **2007**. A new species of skate (Chondrichthyes: Rajidae), *Okamejei mengae* sp. nov., from the South China Sea. *Korean Journal of Ichthyology* 19(1): 57–65.

Jerdon TC. **1849**. On the fresh-water fishes of southern India. *Madras Journal of Literature and Science* 15(2): 302–346.

Jerdon TC. **1851**. Ichthyological gleanings in Madras. *Madras Journal of Literature and Science* 17(39): 128–151.

Jewett SL, Lachner EA. **1983**. Seven new species of the Indo-Pacific genus *Eviota* (Pisces: Gobiidae). *Proceedings of the Biological Society of Washington* 96(4): 780–806.

Joanne A. **1847**. Voyage île M. de Castelnau dans l'Amérique du Sud. *L'Illustration, Journal Universel* No. 239, Vol. X: 59–62.

Joannis L de. **1835**. Observations sur les poissons du Nil, et description de plusieurs espèces nouvelles. [Also includes: Tableau des poissons du Nil.] *Magasin de Zoologie* 1835 (5 année): 53 pp. [numbered as Classe IV]

Joglekar A. **1971**. *Aseraggodes steinitzi*, a new sole from the Red Sea. *Journal of the Marine Biological Association of India* 12: 166–170.

Joglekar A. **1976**. On a new species of the genus *Zebrias* Jordan and Snyder, 1900 (Pisces: Soleidae) from Kerala coast (India). *Zoologischer Anzeiger* 197(1–2): 67–70.

Johannes RE. **1978**. Reproductive strategies of coastal marine fishes in the tropics. *Environmental Biology of Fishes* 3: 65–84.

Johannes RE. **1981**. *Words of the lagoon: Fishing and marine lore in the Palau district of Micronesia*. Berkeley: University of California Press. 245 pp.

John CM. **1955**. The grey mullets of Kayamkulam Lake, India, and their fishery. *Copeia* 1955(3): 225–230.

Johnson GD. **1980**. The limits and relationships of the Lutjanidae and associated families. *Bulletin of the Scripps Institution of Oceanography of the University of California* 24: 1–114.

Johnson GD. **1983**. *Niphon spinosus*, a primitive epinepheline serranid: corroborative evidence from the larvae. *Japanese Journal of Ichthyology* 35(1): 7–18.

Johnson GD. **1983**. *Niphon spinosus*: a primitive epinepheline serranid, with comments on the monophyly and interrelationships of the Serranidae. *Copeia* 1983(3): 777–787.

Johnson GD. **1984**. Percoidei: development and relationships. In: Moser HG, Richards WJ, Cohen DM, Fahay MP, Kendall SW Jr, Richardson SL (eds) Ontogeny and systematics of fishes. *American Society of Ichthyologists and Herpetologists Special Publication* 1: 464–498.

Johnson GD. **1986**. Scombroid phylogeny: an alternative hypothesis. *Bulletin of Marine Science* **39**(1): 1–41.

Johnson GD. **1992**. Monophyly of the euteleostean clades: Neoteleostei, Eurypterygii, and Ctenosquamata. *Copeia* 1992(1): 8–25.

Johnson GD. **1993**. Percomorph phylogeny: progress and problems. *Bulletin of Marine Science* 52(1): 3–28.

Johnson GD, Britz R. **2005**. A description of the smallest *Triodon* on record (Teleostei: Tetraodontiformes: Triodontidae). *Ichthyological Research* 52(2): 176–181.

Johnson GD, Brothers EB. **1993**. *Schindleria*: a paedomorphic goby (Teleostei: Gobioidei). *Bulletin of Marine Science* 52(1): 441–471.

Johnson GD, Patterson C. **1993**. Percomorph phylogeny: a survey of acanthomorphs and a new proposal. *Bulletin of Marine Science* 52(1): 554–626.

Johnson JW. **1999**. Annotated checklist of the fishes of Moreton Bay, Queensland, Australia. *Memoirs of the Queensland Museum* 43(2): 709–762.

Johnson JW. **2002**. *Naso mcdadei*, a new species of unicornfish (Perciformes: Acanthuridae), with a review of the *Naso tuberosus* species complex. *Australian Journal of Zoology* 50(3): 293–311.

Johnson JW. **2004**. Two new species and two new records of aploactinid fishes (Pisces: Scorpaeniformes) from Australia. *Records of the Australian Museum* 56(2): 179–188.

Johnson JW, Randall JE, Chenoweth SF. **2001**. *Diagramma melanacrum* new species of haemulid fish from Indonesia, Borneo and the Philippines with a generic review. *Memoirs of the Queensland Museum* 46(2): 657–676.

Johnson JW, Worthington Wilmer J. **2015**. *Plectorhinchus caeruleonothus*, a new species of sweetlips (Perciformes: Haemulidae) from northern Australia and the resurrection of *P. unicolor* (Macleay, 1883), species previously confused with *P. schotaf* (Forsskål, 1775). *Zootaxa* 3985(4): 491–522. https://doi.org/10.11646/zootaxa.3985.4.2 Johnson JY. **1862**. Descriptions of some new genera and species of fishes obtained at Madeira. *Proceedings of the Zoological Society of London* 1862(2): 167–180.

Johnson JY. **1862**. Notes on rare and little known fishes taken at Madeira. *Annals and Magazine of Natural History* (Series 3) 10(57): 161–172.

Johnson JY. **1862**. Notes on rare and little known fishes taken at Madeira. *Annals and Magazine of Natural History* (Series 3) 10(58): 274–287.

Johnson JY. **1863**. Description of five new species of fishes obtained at Madeira. *Proceedings of the Zoological Society of London* 1863(1): 36–46.

Johnson JY. **1863**. Descriptions of three new genera of marine fishes obtained at Madeira. *Proceedings of the Zoological Society of London* 1863: 403–410.

Johnson JY. **1865**. Description of a new genus of trichiuroid fishes obtained at Madeira, with remarks on the genus *Dicrotus*, Günther, and on some allied genera of Trichiuridae. *Proceedings of the Zoological Society of London* 1865(2, art.5): 434–437.

Johnson JY. **1866**. Description of *Trachichthys darwinii*, a new species of berycoid fish from Madeira. *Proceedings of the Zoological Society of London* 1866(2): 311–315.

Johnson JY. **1890**. On some new species of fishes from Madeira. *Proceedings of the Zoological Society of London* 1890(2): 452–459.

Johnson RK, Barnett MA. **1975**. An inverse correlation between meristic characters and food supply in mid-water fishes: evidence and possible explanations. *Fishery Bulletin* 73(2): 284–298.

Johnstone J. **1904**. Report on the marine fishes collected by Professor Herdman, at Ceylon, in 1902. Supplementary Report No. 15: 201–222. [Report to the Government of Ceylon on the Pearl Fisheries of the Gulf of Mannar. Published by the Royal Society, London]

Johnstone RW, Muhando CA, Francis J. **1998**. The status of the coral reefs of Zanzibar: one example of a regional predicament. *Ambio* 27: 700–707.

Jones EC. **1971**. *Isistius brasiliensis*, a squaloid shark, the probable cause of crater wounds on fishes and cetaceans. *Fishery Bulletin* 69(4): 791–798.

Jones RS, Randall RH, Cheng Y-M, Kami HT, Mak S-M. **1972.** A marine biological survey of southern Taiwan with emphasis on corals and fishes. *Institute of Oceanography, National Taiwan University, Special Publication* 1: 1–93.

Jones S. **1963**. Synopsis of biological data on the long corseletted frigate mackerel *Auxis thynnoides* Bleeker, 1855. *FAO Fisheries Biology Synopsis* 71: 782–810.

Jones S. **1963**. Synopsis of biological data on the northern bluefin tuna *Kishinoella tonggol* (Bleeker) 1851 (Indian Ocean). *FAO Fisheries Biology Synopsis* 74: 862–876. Jones S. **1964**. A preliminary survey of the common tuna bait fishes of Minicoy and their distribution in the Laccadive archipelago. In: *Proceedings of the Symposium on Scombroid Fishes*, Marine Biological Association of India (MBAI), 12– 15 January 1962, Mandapam. Part 2: 643–680.

Jones S. **1969**. Catalogue of fishes from the Laccadive Archipelago in the reference collections of the Central Marine Fisheries Research Institute. *Bulletin of the Central Marine Fisheries Research Institute* (India) 8: 1–32.

Jones S, Kumaran M. **1965**. New records of fishes from the seas around India. Part 1. *Journal of the Marine Biological Association of India* 6(2): 285–306.

Jones S, Kumaran M. **1965**. New records of fishes from the seas around India. Part 2. *Journal of the Marine Biological Association of India* 7(1): 108–123.

Jones S, Kumaran M. **1966**. New records of fishes from the seas around India. Part 4. *Journal of the Marine Biological Association of India* 8(1): 163–180.

Jones S, Kumaran M. **1970**. New records of fishes from the seas around India. Part 6. *Journal of the Marine Biological Association of India* 10(2): 321–331.

Jones S, Kumaran M. **1980**. *Fishes of the Laccadive Archipelago*. Trivandrum, Kerala, India: The Nature Conservation and Aquatic Sciences Service. 760 pp.

Jones S, Rosa H Jr. 1965. Synopsis of biological data on Indian mackerel Rastrelliger kanagurta (Cuvier) 1817 and short bodied mackerel Rastrelliger brachysoma (Bleeker) 1851. FAO Fisheries Synopsis No. 29. Rome: FAO. 34 pp.

Jones S, Silas EG. **1961**. Indian tunas—a preliminary review, with a key for their identification. *Indian Journal of Fisheries* 7(2): 369–393.

Jones S, Silas EG. **1963**. Tuna and tuna-like fishes from the Indian seas. *FAO Fisheries Report* 6(3): 1775–1796.

Jones S, Silas EG. **1964**. A systematic review of the scombroid fishes of India. In: *Proceedings of the Symposium on Scombroid Fishes*, Marine Biological Association of India (MBAI), 12–15 January 1962, Mandapam. Part 1: 1–105, Pls. 1–9.

Jones S, Sujansingani KH. **1954**. Fish and fisheries of the Chilka Lake with statistics of fish catches for the years 1948–1950. *Indian Journal of Fisheries* 1(1&2): 256–344.

Jordan DS. 1886. List of fishes collected at Havana, Cuba, in December, 1883, with notes and descriptions. *Proceedings of the United States National Museum* 9(551): 31–55.

Jordan DS. **1898**. Description of a species of fish (*Mitsukurina owstoni*) from Japan, the type of a distinct family of lamnoid sharks. Proceedings of the California Academy of Sciences (Series 3) 1(6): 199–204.

Jordan DS (assisted by Sindo M). **1902.** A review of the pediculate fishes or anglers of Japan. *Proceedings of the United States National Museum* 24(1261): 361–381.

Jordan DS. **1903**. Supplementary note on *Bleekeria mitsukurii*, and on certain Japanese fishes. *Proceedings of the United States National Museum* 26(1328): 693–696.

Jordan DS. **1916**. The nomenclature of American fishes as affected by the opinions of the International Commission on Zoological Nomenclature. *Copeia* 1916(29): 25–28.

Jordan DS. **1917**. Notes on *Glossamia* and related genera of cardinal fishes. *Copeia* 1917(44): 46–47.

Jordan DS. **1919**. New genera of fishes. *Proceedings of the Academy of Natural Sciences of Philadelphia* 70 [for 1918]: 341–344.

Jordan DS. **1919**. On *Elephenor*, a new genus of fishes from Japan. *Annals of the Carnegie Museum* 12(3–4): 329–343.

Jordan DS. **1921**. The fish fauna of the California Tertiary. *Stanford University Publications* (University Series, Biological Sciences) 1(4): 237–300.

Jordan DS. **1923**. A classification of fishes including families and genera as far as known. *Stanford University Publications* (University Series, Biological Sciences) 3(2): 77–243.

Jordan DS, Davis BM. **1891**. A preliminary review of the apodal fishes or eels inhabiting the waters of America and Europe. *United States Commission of Fish and Fisheries, Report of the Commissioner* 16(9): 581–677.

Jordan DS, Dickerson MC. **1908**. On a collection of fishes from Fiji, with notes on certain Hawaiian fishes. *Proceedings of the United States National Museum* 34: 603–617.

Jordan DS, Eigenmann CH. **1890**. A review of the genera and species of Serranidae found in the waters of America and Europe. *Bulletin of the U.S. Fish Commission* 8: 329–441.

Jordan DS, Evermann BW. **1896**. The fishes of North and Middle America: a descriptive catalogue of the species of fish-like vertebrates found in the waters of North America, north of the Isthmus of Panama. Part 1. *Bulletin of the United States National Museum* 47: i–ix + 1–1240.

Jordan DS, Evermann BW. **1898**. The fishes of North and Middle America: a descriptive catalogue of the species of fish-like vertebrates found in the waters of North America, north of the Isthmus of Panama. Part 2. *Bulletin of the United States National Museum* 47: i–xxx + 1241–2183.

Jordan DS, Evermann BW. **1898**. The fishes of North and Middle America: a descriptive catalogue of the species of fish-like vertebrates found in the waters of North America, north of the Isthmus of Panama. Part 3. *Bulletin of the United States National Museum* 47: i–xxiv + 2183a–3136.

Jordan DS, Evermann BW. **1900**. The fishes of North and Middle America: a descriptive catalogue of the species of fish-like vertebrates found in the waters of North America, north of the Isthmus of Panama. Part 4. *Bulletin of the United States National Museum* 47: i–ci + 3137–3313, Pls. 1–392. Jordan DS, Evermann BW. **1902**. Preliminary report on an investigation of the fishes and fisheries of the Hawaiian islands. *Report of the Commissioner of the U.S. Commission of Fish and Fisheries* 27(Appendix 11): 353–499.

Jordan DS, Evermann BW. **1903**. Notes on a collection of fishes from the island of Formosa. *Proceedings of the United States National Museum* 25(1289): 315–368.

Jordan DS, Evermann BW. **1903**. Descriptions of new genera and species of fishes from the Hawaiian Islands. *Bulletin of the U.S. Fish Commission* 22(1902): 161–208.

Jordan DS, Evermann BW. **1903**. Description of a new genus and two new species of fishes from the Hawaiian Islands. *Bulletin of the U.S. Fish Commission* 22(1902): 209–210.

Jordan DS, Evermann BW. **1905**. The aquatic resources of the Hawaiian Islands. Part I. The shore fishes of the Hawaiian Islands, with a general account of the fish fauna. *Bulletin of the U.S. Fish Commission* 23(pt 1) (1903): xxviii + 574 pp.

Jordan DS, Evermann BW. **1927**. New genera and species of North American fishes. *Proceedings of the California Academy of Sciences* (Series 4) 16(15): 501–507.

Jordan DS, Evermann BW, Tanaka S. **1927**. Notes on new or rare fishes from Hawaii. *Proceedings of the California Academy of Sciences* (Series 4) 16(20): 649–680.

Jordan DS, Fordice MW. **1887**. A review of the American species of Belonidae. *Proceedings of the United States National Museum* 9(575): 339–361.

Jordan DS, Fowler HW. **1902**. A review of the trigger-fishes, file-fishes, and trunk-fishes of Japan. *Proceedings of the United States National Museum* 25(1287): 251–286.

Jordan DS, Fowler HW. **1902**. A review of the Chaetodontidae and related families of fishes found in the waters of Japan. *Proceedings of the United States National Museum* 25(1296): 513–563.

Jordan DS, Fowler HW. **1902**. A review of the berycoid fishes of Japan. *Proceedings of the United States National Museum* 26(1306): 1–21.

Jordan DS, Fowler HW. **1903**. A review of the dragonets (Callionymidae) and related fishes of the waters of Japan. *Proceedings of the United States National Museum* 25(1305) [for 1902]: 939–959.

Jordan DS, Fowler HW. **1903**. A review of the elasmobranchiate fishes of Japan. *Proceedings of the United States National Museum* 26(1324): 593–674.

Jordan DS, Gilbert CH. **1880**. Description of seven new species of sebastoid fishes, from the coast of California. *Proceedings of the United States National Museum* 3(150): 287–298.

Jordan DS, Gilbert CH. **1880**. Description of a new scorpaenoid fish (*Sebastichthys maliger*), from the coast of California. *Proceedings of the United States National Museum* 3(157): 322–324.

Jordan DS, Gilbert CH. **1881**. Description of *Sebastichthys mystinus*. *Proceedings of the United States National Museum* 4(192): 70–72.

Jordan DS, Gilbert CH. **1882**. Notes on a collection of fishes made by Lieut. Henry E. Nichols, U. S. N., on the west coast of Mexico, with descriptions of new species. *Proceedings of the United States National Museum* 4(221): 225–233.

Jordan DS, Gilbert CH. **1883**. Synopsis of the fishes of North America. *Bulletin of the United States National Museum* 16: iv + 1018 pp.

Jordan DS, Hubbs CL. **1920**. Studies in ichthyology: a monographic review of the family of Atherinidae or Silversides. *Leland Stanford Junior University Publications* (University Series) 40: 1–87.

Jordan DS, Hubbs CL. **1925**. Record of fishes obtained by David Starr Jordan in Japan, 1922. *Memoirs of the Carnegie Museum* 10(2): 93–346.

Jordan DS, Jordan EK. **1922**. A list of the fishes of Hawaii, with notes and descriptions of new species. *Memoirs of the Carnegie Museum* 10(1): 1–92, Pls. 1–4.

Jordan DS, McGregor RC. **1906**. Description of a new species of threadfin (family Polynemidæ) from Japan. *Proceedings of the United States National Museum* 30(1470): 813–815.

Jordan DS, Meek SE. **1885**. A review of the American species of flying fishes (*Exocoetus*). *Proceedings of the United States National Museum* 8(5): 44–67.

Jordan DS, Richardson RE. **1908**. A review of the flat-heads, gurnards, and other mail-cheeked fishes of the waters of Japan. *Proceedings of the United States National Museum* 33(1581): 629–670.

Jordan DS, Richardson RE. **1908**. Fishes from islands of the Philippine Archipelago. *Bulletin of the Bureau of Fisheries* 27: 233–287.

Jordan DS, Richardson RE. **1909**. Catalogue of the fishes of the Island of Formosa or Taiwan, based on the collections of Dr. Hans Sauter. *Memoirs of the Carnegie Museum* 4(4): 159–204.

Jordan DS, Richardson RE. **1910**. *Check-list of the species of fishes known from the Philippine Archipelago*. Publication No. 1. Manila: Bureau of Science. 78 pp.

Jordan DS, Seale A. **1905**. List of fishes collected at Hong Kong by Captain William Finch, with description of five new species. *Proceedings of the Davenport Academy of Sciences* (Iowa) 10: 1–17.

Jordan DS, Seale A. **1905**. List of fishes collected by Dr. Bashford Dean on the island of Negros, Philippines. *Proceedings of the United States National Museum* 28(1407): 769–803.

Jordan DS, Seale A. **1905**. List of fishes collected in 1882– 83 by Pierre Louis Jouy at Shanghai and Hong Kong, China. *Proceedings of the United States National Museum* 29(1433): 517–529. Jordan DS, Seale A. **1906**. The fishes of Samoa. Description of the species found in the archipelago, with a provisional check-list of the fishes of Oceania. *Bulletin of the Bureau of Fisheries* 25 [for 1905]: 173–455.

Jordan DS, Seale A. **1907**. Fishes of the islands of Luzon and Panay. *Bulletin of the Bureau of Fisheries* 26: 1–48.

Jordan DS, Seale A. **1926**. Review of the Engraulidae, with descriptions of new and rare species. *Bulletin of the Museum of Comparative Zoology* 67(11): 355–418.

Jordan DS, Snyder JO. **1900**. A list of fishes collected in Japan by Keinosuke Otaki, and by the United States steamer *Albatross*, with descriptions of fourteen new species. *Proceedings of the United States National Museum* 23(1213): 335–380, Pls. 9–20.

Jordan DS, Snyder JO. **1901**. A review of the apodal fishes or eels of Japan, with descriptions of nineteen new species. *Proceedings of the United States National Museum* 23(1239): 837–890.

Jordan DS, Snyder JO. **1901**. A review of the cardinal fishes of Japan. *Proceedings of the United States National Museum* 23(1240): 891–913.

Jordan DS, Snyder JO. **1901**. A review of the hypostomide and lophobranchiate fishes of Japan. *Proceedings of the United States National Museum* 24(1241): 1–20.

Jordan DS, Snyder JO. **1902**. A review of the gobioid fishes of Japan, with descriptions of twenty-one new species. *Proceedings of the United States National Museum* 24(1244): 33–132.

Jordan DS, Snyder JO. **1902**. A review of the trachinoid fishes and their supposed allies found in the waters of Japan. *Proceedings of the United States National Museum* 24(1263): 461–497.

Jordan DS, Snyder JO. **1903**. Descriptions of two new species of squaloid sharks from Japan. *Proceedings of the United States National Museum* 25(1279): 79–87.

Jordan DS, Snyder JO. **1904**. Notes on collections of fishes from Oahu Island and Laysan Island, Hawaii, with descriptions of four new species. *Proceedings of the United States National Museum* 27(1377): 939–948.

Jordan DS, Snyder JO. **1906**. A review of the Poeciliidae or killifishes of Japan. *Proceedings of the United States National Museum* 31(1486): 287–290.

Jordan DS, Snyder JO. **1907**. Notes on fishes of Hawaii, with descriptions of new species. *Bulletin of the Bureau of Fisheries* 26(623) [for 1906]: 205–218, Pls. 12–13.

Jordan DS, Snyder JO. **1908**. Description of three new species of carangoid fishes from Formosa. *Memoirs of the Carnegie Museum* 4(2): 37–40.

Jordan DS, Starks EC. **1895**. The fishes of Puget Sound. *Proceedings of the California Academy of Sciences* (Series 2) 5: 785–855. Jordan DS, Starks EC. **1901**. A review of the atherine fishes of Japan. *Proceedings of the United States National Museum* 24(1250): 199–206.

Jordan DS, Starks EC. **1904**. A review of the scorpaenoid fishes of Japan. *Proceedings of the United States National Museum* 27(1351): 91–175.

Jordan DS, Starks EC. **1904**. List of fishes dredged by the steamer *Albatross* off the coast of Japan in the summer of 1900, with descriptions of new species and a review of the Japanese Macrouridae. *Bulletin of the U.S. Fish Commission* 22(1902): 577–630.

Jordan DS, Starks EC. 1908. In: Jordan DS, Dickerson MC.

Jordan DS, Starks EC. **1917**. Notes on a collection of fishes from Ceylon with descriptions of new species. *Annals of the Carnegie Museum* 11: 430–460.

Jordan DS, Swain J. **1884**. A review of the American species of marine Mugilidae. *Proceedings of the United States National Museum* 7(18): 261–275.

Jordan DS, Swain J. **1885**. A review of the American species of *Epinephelus* and related genera. *Proceedings of the United States National Museum* 7(447): 358–410.

Jordan DS, Tanaka S. **1927**. Notes on new and rare fishes of the fauna of Japan. *Annals of the Carnegie Museum* 17(3–4): 385–394.

Jordan DS, Tanaka S, Snyder JO. **1913**. A catalogue of the fishes of Japan. *Journal of the College of Science* (Imperial University, Tokyo) 33(1): 1–497.

Jordan DS, Thompson WF. **1911**. A review of the sciaenoid fishes of Japan. *Proceedings of the United States National Museum* **39**(1787): 241–261.

Jordan DS, Thompson WF. **1913**. Notes on a collection of fishes from the island of Shikoku in Japan, with a description of a new species, *Gnathypops iyonis*. *Proceedings of the United States National Museum* 46(2011): 65–72.

Jordan DS, Thompson WF. **1914**. Record of the fishes obtained in Japan in 1911. *Memoirs of the Carnegie Museum* 6(4): 205–313.

Joshi KK, Zacharia PU, Kanthan P. 2012. Description of a new sand lance species, *Bleekeria murtii* (Perciformes: Ammodyidae) from India. *Indian Journal of Fisheries* 59(2): 101–107. http://eprints.cmfri.org.in/id/eprint/8994

Jubb RA. **1957**. The freshwater eel *Anguilla obscura* Günther, 1871, in southern Africa. *Nature* (London) 180: 1216.

Jubb RA. **1961**. The freshwater eels (*Anguilla* spp.) of southern Africa. An introduction to their identification and biology. *Annals of the Cape Provincial Museums* 1: 15–48.

Jubb RA. **1967**. *Freshwater fishes of southern Africa*. Cape Town and Amsterdam: Balkema. vii + 248 pp.

Juillerat E. **1880**. Note sur un lophobranche du genre *Coelonotus. Bulletin de la Société philomathique de Paris* (Série 7) 4: 176. Jungersen HFE. **1910**. Ichthyotomical contributions. II. The structure of the Aulostomidae, Syngnathidae and Solenostomidae. *Det Kongelige Danske Videnskabernes Selskabs Skrifter, 7 Raekke. Naturvidenskabelig og Matematisk Afdeling* 8: 267–364 + Pls. I–VII.

## Κ

Kaga T. **2013**. Phylogenetic systematics of the family Sillaginidae (Percomorpha: order Perciformes). *Zootaxa* 3642(1): 1–105. http://dx.doi.org/10.11646/zootaxa.3642.1.1

Kaga T, Heemstra E. **2013**. First record of a rare sand whiting, *Sillago caudicula* (Perciformes: Sillaginidae), from Madagascar. *Marine Biology Research* 9(3): 316–320. http://dx.doi.org/10.1080/17451000.2012.742547

Kaga T, Ho H-C. **2012**. Redescription of *Sillago (Parasillago) indica* McKay, Dutt & Sujatha, 1985 (Perciformes: Sillaginidae), with a reassignment to the subgenus *Sillago. Zootaxa* 3513: 61–67. https://doi.org/10.11646/ zootaxa.3513.1.3

Kaga T, Imamura H, Nakaya K. 2010. A new sand whiting, Sillago (Sillago) caudicula, from Oman, the Indian Ocean (Perciformes: Sillaginidae). Ichthyological Research 57(4): 367–372. http://dx.doi.org/10.1007/s10228-010-0169-z

Kähsbauer P. **1950**. Beitrag zur Systematik der Syngnathiden (Pisces). *Annalen des Naturhistorischen Museums in Wien* 57: 263–272.

Kähsbauer P. **1976**. Über einige Syngnathiden des Indo-Pazifischen Raumes und der Antillen. *Annalen des Naturhistorischen Museums in Wien* 80: 281–290.

Kailola PJ. 1987. The fishes of Papua New Guinea. A revised and annotated checklist. Vol. 1. Myxinidae to Synbranchidae.
Research Bulletin 41. Port Moresby: Department of Fisheries and Marine Resources. xxxi + 194.

Kailola PJ. 1999. Ariidae (= Tachysuridae). Sea catfishes (fork-tailed catfishes) (pp. 1827–1879). In: Carpenter KE, Niem VH (eds) FAO species identification guide for fishery purposes. The living marine resources of the western central Pacific. Vol. 3. Batoid fishes, chimaeras and bony fishes part 1 (pp. 1397–2068). Rome: FAO.

Kailola PJ. **2004**. A phylogenetic exploration of the catfish family Ariidae (Otophysi: Siluriformes). *The Beagle, Records of the Museums and Art Galleries of the Northern Territory* 20: 87–166.

Kalish JM, Newman SJ, Johnston J. 2002. Chapter 18. Use of bomb radiocarbon to validate the age estimation method for *Epinephelus octofasciatus, Etelis carbunculus* and *Lethrinus nebulosus* from Western Australia (pp. 281–298). In: Kalish JM (ed) *Use of the bomb radiocarbon chronometer to validate fish age.* Final Report to the Fisheries Research and Development Corporation (FRDC) on Project No. 93-109. Canberra: Australian National University. 384 pp.

Kami HT, Ikehara II. **1976**. Notes on the annual juvenile siganid harvest in Guam. *Micronesica* 12(2): 323–325.

Kamohara T. **1933**. On a new fish from Japan. *Dōbutsugaku* Zasshi [Zoological Magazine, Tokyo] 45(539): 389–393.

Kamohara T. 1934. Supplementary notes on the fishes collected in the vicinity of Kochi (VII). *Dōbutsugaku Zasshi* [*Zoological Magazine, Tokyo*] 46(552): 457–463. [In Japanese]

Kamohara T. **1935**. On a new fish of the Zeidae from Kochi, Japan. *Dōbutsugaku Zasshi* [*Zoological Magazine, Tokyo*] 47(558): 245–247. [In Japanese, English description on p. 247]

Kamohara T. 1936. Supplementary note on the fishes collected in the vicinity of Kôchi-shi (IX). Dōbutsugaku Zasshi [Zoological Magazine Tokyo] 48(6): 306–311. [In Japanese, English summary]

Kamohara T. **1937**. A review of the triacanthodid fishes from Prov. Tosa, Shikoku Japan. *Annotationes Zoologicae Japonenses* 16: 5–8.

Kamohara T. 1937. On some rare and one new species of fishes from Japan. Dobutsugaku Zasshi [Zoological Magazine, Tokyo] 49(5): 186–190. [In Japanese, English summary]

Kamohara T. **1938**. On the offshore bottom-fishes of Prov. Tosa, Shikoku, Japan. Tokyo: Maruzen Kobushiki Kaisha. 86 pp.

Kamohara T. **1941**. Descriptions of one new and two rare fishes from Japan. *Annotationes Zoologicae Japonenses* 20(3): 166–168.

Kamohara T. 1943. Some unrecorded and two new fishes from Prov. Tosa, Japan. Bulletin of the Biogeographical Society of Japan 13(17): 125–137.

Kamohara T. 1952. Revised descriptions of the offshore bottom-fishes of Prov. Tosa, Shikoku, Japan. *Research Reports of the Kôchi University* (Natural Science Series) 3: 1–222. [In Japanese, English summary]

Kamohara T. **1953**. A review of the fishes of the family Chlorophthalmidae found in the waters of Japan. *Japanese Journal of Ichthyology* 3(1): 1–6.

Kamohara T. **1954**. A review of the family Brotulidae found in the waters of Prov. Tosa, Japan. *Reports of the U.S.A. Marine Biological Station* 1(2): 1–14.

Kamohara T. **1954**. Eleven additions to the fish fauna of Prov. Tosa, including one new species of the family Serranidae. *Research Reports of Kochin University* 3(26): 1–6.

Kamohara T. 1962. Notes on six additions to the marine fish fauna of Kochi Prefecture, Japan. *Reports of the U.S.A. Marine Biological Station* 9(2): 1–6.

Kanazawa RH. 1958. A revision of the eels of the genus Conger with descriptions of four new species. Proceedings of the United States National Museum 108(3400): 219–267.

Kannan K, John S, Jhonson JA, Zacharia PU, Joshi KK. 2013. First record of striated fusilier *Caesio striata*  (Teleostei: Caesionidae) from Indian waters. *Marine Biodiversity Records* 6(e106): 1–3. https://doi.org/10.1017/S175526721300081X

Kapoor D, Dayal R, Ponniah AG (eds). **2002**. *Fish biodiversity of India*. Lucknow, India: National Bureau of Fish Genetic Resources. 775 pp.

Kar S, Chakraborty R. 2000. First record of the sunfish *Ranzania laevis* (Pennant) (Pisces: Osteichthyes: Perciformes: Molidae) from the west Bengal coast. *Journal of the Bombay Natural History Society* 97(2): 288–289.

Karmovskaya ES. **1991**. The new species of conger eels (Congridae) from the western Indian Ocean. *Voprosy Ikhtiologii* 31(6): 891–897 [Figure 2a]. [In Russian. English translation in *Journal of Ichthyology* 32(3): 1–8]

Karmovskaya ES. 2006. New species of the genus *Myroconger*, *M. seychellensis* (Myrocongridae, Anguilliformes) from the western Equatorial part of the Indian Ocean. *Voprosy Ikhtiologii* 46(5): 590–593. [In Russian. English translation in *Journal of Ichthyology* 46(8): 563–565]

Karplus I, Szlep R, Tsurnamal M. **1981**. Goby-shrimp partner specificity. I. Distribution in the northern Red Sea and partner specificity. *Journal of Experimental Marine Biology and Ecology* 51(1): 1–19.

Karrer C. 1968. Über Erstnachweise und seltene Arten von Fischen aus dem Südatlantik (argentinisch südbrasilianische Küste) 1. Mitteilungen Zoologische Jahrbücher, Abteilung für Systematik, Ökologie und Geographie der Tiere 95(4): 542–570.

Karrer C. **1973**. Über Fische aus dem Südostatlantik. *Mitteilungen aus dem Zoologischen Museum in Berlin* 49(1): 191–257.

Karrer C. **1975**. Über Fische aus dem Südostatlantik (Teil 2). *Mitteilungen aus dem Zoologischen Museum in Berlin* 51(1): 63–82.

Karrer C. **1976**. Über drei mesopelagische fischarten aus dem Golf von Guinea. *Mitteilungen aus dem Zoologischen Museum in Berlin* 52: 177–182.

Karrer C. **1983**. *Anguilliformes du Canal de Mozambique* (*Pisces, Teleostei*). *Faune Tropicale 23* [for 1982]. Paris: ORSTOM. 116 pp.

Karrer C. 1984. Notes on the synonymies of *Ariomma brevimanum* and *A. luridum* and the presence of the latter in the Atlantic (Teleostei, Perciformes, Ariommatidae). *Cybium* 8(4): 94–95.

Karrer C. 1990. Oreosomatidae (pp. 637–640). In: Quéro J-C, Hureau J-C, Karrer C, Post A, Saldanha L (eds) *Check-list of the fishes of the eastern tropical Atlantic (CLOFETA)*.
Vol. 2. Lisbon: Junta Nacional de Investigação Científica e Tecnológica.

Karrer C, Klausewitz W. **1982**. Tiefenwasser- und Tiefseefische aus dem Roten Meer. II. *Dysomma fuscoventralis* n. sp., ein Tiefsee-Aal aus dem zentralen Roten Meer (Teleostei: Anguilliformes: Synaphobranchidae: Dysomminae). *Senckenbergiana Biologica* 62(4/6): 199–203.

Karuppasamy K, Jawahar P, Kathirvelpandian A, Purushothaman P, Ranjith L, Zacharia PU, Vidhya V. 2020. New distributional record of oblique-barred monocle bream *Scolopsis xenochrous*, Günther, 1872 (Perciformes: Nemipteridae) from Wadge Bank of Southeastern Arabian Sea. *Thalassas: An International Journal of Marine Sciences* 36(2): 517–523. http://dx.doi.org/10.1007/s41208-020-00219-7

Kaschner CJ, Weigmann S, Thiel R. 2015. *Bythaelurus tenuicephalus* n. sp., a new deep-water catshark
(Carcharhiniformes, Scyliorhinidae) from the western Indian Ocean. *Zootaxa* 4013(1): 120–138. https://doi. org/10.11646/zootaxa.4013.1.9

Katayama E, Endo H. 2010. Redescription of a sanddiver, *Trichonotus blochii* (Actinopterygii: Perciformes: Trichonotidae), with confirmation of its validity. *Species Diversity* 15(1): 1–10. http://dx.doi.org/10.12782/ specdiv.15.1

Katayama E, Motomura H, Hiromitsu E. 2012. A new species of *Trichonotus* (Perciformes: Trichonotidae) from Somalia and redescription of *Trichonotus cyclograptus* (Alcock, 1890) with designation of a lectotype. *Zootaxa* 3565: 31–43. http://dx.doi.org/10.11646/zootaxa.3565.1.3

Katayama M. 1934. On the external and internal characters of the bony fishes of the genus *Vegetichthys*, with a description of one new species. *Proceedings of the Imperial Academy*, *Tokyo* 10(7): 435–438.

Katayama M. **1960**. *Fauna Japonica. Serranidae (Pisces)*. Tokyo: Tokyo News Service. viii + 189 pp.

Katayama M. 1975. Serranid fishes of the Okinawa Islands (III). Bulletin of the Faculty of Education Yamaguchi University 25(2): 161–178.

Katayama M. **1978**. The anthiine fish, *Pseudanthias taeniatus*, from Hachijo Island and the coast of Izu, Japan. *Japanese Journal of Ichthyology* 25(3): 216–218.

Katayama M. 1984. Family Serranidae (sea basses)
(pp. 126–138). In: Masuda H, Amaoka K, Araga C, Uyeno T, Yoshino T (eds) *The fishes of the Japanese archipelago*. Tokyo: Tokai University Press. 437 pp.

Katayama M, Amaoka K. **1986**. Two new anthiine fishes from the eastern tropical Atlantic. *Japanese Journal of Ichthyology* 27(3): 213–222.

Katayama M, Takai T. 1954. A new conger-like eel, Muraenesox yamaguchiensis, from the inland Sea of Japan. Japanese Journal of Ichthyology 3(3/4/5): 97–101.

Katayama M, Yamamoto E. 1986. The anthiine fishes, Odontanthias dorsomaculatus sp. nov. and Plectranthias bauchotae Randall, from the Western Indian Ocean. Japanese Journal of Ichthyology 32(4): 387–391. Kaup JJ. **1826**. Beyträge zu Amphibiologie und Ichthiyologie. *Isis (Oken)* 19(1): 87–90.

Kaup JJ. **1853**. Uebersicht der Lophobranchier. *Archiv für Naturgeschichte* **19**(1): 226–234.

Kaup JJ. **1855**. Uebersicht über die species einiger Familien der Sclerodermen. *Archiv für Naturgeschichte* 21(1): 215–233.

Kaup JJ. **1856**. *Catalogue of apodal fish in the collection of the British Museum*. London: British Museum. 163 pp.

Kaup JJ. **1856**. *Catalogue of lophobranchiate fish in the collection of the British Museum*. London: Taylor & Francis. iv + 80 pp.

Kaup JJ. **1856**. Uebersicht der Aale. *Archiv für Naturgeschichte* 22(1): 41–77.

Kaup JJ. **1856**. Einiges über die Unterfamilie Ophidinae. *Archiv für Naturgeschichte* 22(1): 93–100.

Kaup JJ. **1858**. Uebersicht der Familie Gadidae. *Archiv für Naturgeschichte* 24(1): 85–93.

Kaup JJ. 1858. Uebersicht der Plagusinae, der fünften Subfamilie der Pleuronectidae. Archiv für Naturgeschichte 24(1): 105–110.

Kaup JJ. 1859. Einiges über die Acanthopterygiens à joue cuirassée Cuv. Archiv für Naturgeschichte 24(1) [for 1858]: 329–343.

Kaup JJ. **1860**. Ueber die Chaetodontidae. *Archiv für Naturgeschichte* 26(1): 133–156.

Kaup JJ. 1873. Ueber die Familie Triglidae nebst einigen Worten über die Classification. Archiv für Naturgeschichte 39(1): 71–94.

Kawahara R, Miya M, Mabuchi K, Lavoué S, Inoue JG, Satoh TP, Kawaguchi A, Nishida M. **2008**. Interrelationships of the 11 gasterosteiform families (sticklebacks, pipefishes, and their relatives): a new perspective based on whole mitogenome sequences from 75 higher teleosts. *Molecular Phylogenetics and Evolution* 46(1): 224–236. http://dx.doi.org/10.1016/j.ympev.2007.07.009

Kawai T. **2008**. Phylogenetic systematics of the family Peristediidae (Teleostei: Actinopterygii). *Species Diversity* 13(1): 1–34. http://dx.doi.org/10.12782/specdiv.13.1

Kawai T. **2013**. Revision of the peristediid genus *Satyrichthys* (Actinopterygii: Teleostei) with the description of a new species, *S. milleri* sp. nov. *Zootaxa* 3635(4): 419–438. http://doi.org/10.11646/zootaxa.3635.4.5

Kawai T. **2014**. *Satyrichthys kikingeri* Pogoreutz, Vitecek & Ahnelt, 2013, a junior synonym of *Satyrichthys laticeps* (Schlegel, 1852) (Actinopterygii: Teleostei: Peristediidae). *Zootaxa* 3900(1): 135–140.

https://doi.org/10.11646/zootaxa.3900.1.9

Kawai T. **2016**. *Peristedion richardsi* sp. nov. (Actinopterygii: Teleostei: Peristediidae) from Indonesian waters, with synonymy between *Peristedion riversandersoni* Alcock, 1894 and *Peristedion nierstraszi* Weber, 1913. *Zootaxa* 4171(2): 335–346. http://dx.doi.org/10.11646/ zootaxa.4171.2.6

Kawai T, Imamura H, Nakaya K. 2004. Paraheminodus kochiensis Kamohara, 1957 (Teleostei: Peristediidae), a junior synonym of Paraheminodus murrayi (Günther, 1880), with a comparison of Paraheminodus murrayi and Paraheminodus laticephalus (Kamohara, 1952). Ichthyological Research 51(1): 73–76.

Kawai T, Imamura H, Nakaya K. 2004. A new species of armored sea robin, *Paraheminodus kamoharai* (Teleostei: Peristediidae), from the Philippines. *Ichthyological Research* 51(2): 126–130.

Kawai T, Tashiro F. **2008**. First record of armored searobin, *Satyrichthys adeni*, from Suruga Bay, Japan. *Japanese Journal of Ichthyology* 55(1): 43–47.

Kawai T, Tashiro F, Imamura H, Aungtonya C. **2017**. Deep-sea fishes from the Andaman Sea by R/V *Chakratong Tongyai* during 1996–2000. Part 1: Scorpaeniformes. *Phuket Marine Biological Center Research Bulletin* 74: 23–32.

Kearney RE, Lewis AD, Smith BR. 1972. Cruise report Tagula 71-1; survey of skipjack tuna and bait resources in Papua New Guinea waters. *Research Bulletin, Papua New Guinea Department of Agriculture, Stock and Fisheries* 8: 1–145.

Keenleyside MHA (ed). **1991**. *Cichlid fishes: Behaviour, ecology and evolution*. London: Chapman & Hall. xxi + 378 pp.

Keith P, Bosc P, Valade P. **2004**. A new species of *Parioglossus* (Gobioidei, Ptereleotridae) from Seychelles Islands. *Cybium* 28(4): 341–344.

Keith P, Hoareau T, Bosc P. **2005**. The genus *Cotylopus* (Teleostei: Gobioidei) endemic to the rivers of islands of the Indian Ocean with description of a new species from Mayotte (Comoros). *Journal of Natural History* 39(17): 1395–1406.

Keith P, Marquet G, Valade P, Bosc P, Vigneux E. 2006. Atlas des poissons et des Crustacés d'eau douce des Comores, Mascareignes et Seychelles. *Muséum National d'Histoire Naturelle: Patrimoines Naturels* 65: 1–250.

Keith P, Vigneux E, Bosc P. **1999**. Atlas des poissons et des crustacés d'eau douce de la Réunion. Vol. 39. Paris: Patrimoines naturels (M.N.H.N./S.P.N.) 137 pp.

Keivany Y, Nelson JS. 2006. Interrelationships of Gasterosteiformes (Actinopterygii, Percomorpha). *Journal* of Ichthyology 46(Suppl. 1): S84–S96.

Kemp JM. 1998. Zoogeography of the coral reef fishes of the Socotra Archipelago. *Journal of Biogeography* 25(5): 919–933.

Kemp JM. **2000**. Zoogeography of the coral reef fishes of the north-eastern Gulf of Aden, with eight new records of coral reef fishes from Arabia. *Fauna of Arabia* 18: 293–321.

Kemp JM. 2000. Hybridisation of the marine angelfishes Pomacanthus maculosus and P. semicirculatus in the Gulf of Aden. Fauna of Arabia 18: 357–367. Kemp M, Benzoni F. 2000. A preliminary study of coral communities in the northern Gulf of Aden. *Fauna of Arabia* 18: 67–86.

Kendall WC, Goldsborough EL. 1911. XIII. The shore fishes.
In: Reports on the scientific results of the expedition to the tropical Pacific, in charge of Alexander Agassiz, by the U.S. Fish Commission steamer *Albatross*, from August, 1899, to March, 1900. *Memoirs of the Museum of Comparative Zoology* 26(7): 239–344.

Kent P, Hunt JA, Johnstone DW. 1971. *The geology and geophysics of coastal Tanzania*. *Geophysical Paper No. 6*.
London: Institute of Geological Sciences. vi + 101 pp.

Kerstetter DW, Taylor ML. **2008**. Live release of a bigeye sand tiger *Odontaspis noronhai* (Elasmobranchii: Lamniformes) in the western north Atlantic. *Bulletin of Marine Science* 83(3): 465–469.

Kesteven GL, Nakken O, Stromme T (eds). **1981**. The smallpelagic and demersal fish resources of the north-west Arabian Sea – further analysis of the results of the R/V *Dr. Fridtjof Nansen* survey, 1975–1976. Institute of Marine Research, Bergen, Norway. 110 pp.

Key B. 2015. Fish do not feel pain and its implications for understanding phenomenal conciousness. *Biology & Philosophy* 30: 149–165. https://doi.org/10.1007/s10539-014-9469-4

Khalaf KT. **1961**. *The marine and freshwater fishes of Iraq*. Baghdad: Ar-Rabitta Press. 164 pp.

Khalaf MA, Disi AM. **1997**. Fishes of the Gulf of Aqaba. *Publication of the Marine Science Station, Aqaba, Jordan* 8: 252 pp.

Khalaf MA, Kochzius M. **2002**. Changes in trophic community structure of shore fishes at an industrial site in the Gulf of Aqaba, Red Sea. *Marine Ecology Progress Series* 239: 287–299.

Khalaf MA, Krupp F. **2003**. Two new records of fishes from the Red Sea. *Zoology in the Middle East* 30: 55–59.

Khalaf MA, Krupp F. **2008**. A new species of the genus *Symphysanodon* (Perciformes: Symphysanodontidae) from the Gulf of Aqaba, Red Sea. *aqua, International Journal of Ichthyology* 14(2): 85–88.

Khalaf MA, Zajonz U. **2007**. Fourteen additional fish species recorded from below 150 m depth in the Gulf of Aqaba, including *Liopropoma lunulatum* (Pisces: Serranidae), new record for the Red Sea. *Fauna of Arabia* 23: 421–433.

Khan RA. **2003**. Fish faunal resources of Sunderban estuarine system with special reference to the biology of some commercially important species. *Records of the Zoological Survey of India, Occasional Paper* 209: 1–150.

Kiener A. **1961**. Poissons Malgaches. Liste de noms Malgaches de poissons d'eau douce, d'eaux Saumâtres et d'espèces euryhalines. *Bulletin de Madagascar* 179–181: 1–117.

Kiener, A. 1963. Gobioidei (Pisces) nouveaux ou rares de Madagascar. Bulletin du Muséum d'Histoire Naturelle (Série 2) 35(4): 328–333.

Kiener A. 1963. Poissons, pêche et pisciculture à Madagascar. Publication du Centre Technique Forestier Tropical 24: 1–244.

Kiener A, Spillmann CJ. 1969. Contributions à l'étude systématique et écologique des Athérines des côtes françaises. Mémoires du Muséum National d'Histoire Naturelle (Paris) (N.S., Série A, Zoologie) 60(2): 33–74.

Kiener A, Spillmann J. 1973. Atherinidae (pp. 576–578). In: Hureau J-C, Monod T (eds) Check-list of the fishes of the north-eastern Atlantic and of the Mediterranean (CLOFNAM). Vol. I. Paris: UNESCO.

Kikawa S. **1975**. Synopsis on the biology of the shortbill spearfish, *Tetrapterus angustirostris* Tanaka, 1914 in the Indo-Pacific areas. *NOAA Technical Report* NMFS SSRF-675 (Part 3): 39–54.

Kim B-J, Amaoka K. 2001. A new species, *Parupeneus procerigena*, from the Saya de Malha Bank in the western Indian Ocean (Perciformes: Mullidae). *Ichthyological Research* 48(1): 45–50.

Kim B-J, Nakaya K. 2002. Upeneus australiae, a new goatfish (Mullidae: Perciformes) from Australia. Ichthyological Research 49(2): 128–132.

Kim B-J, Nakaya K. 2003. Redescription of a poorly known goatfish, Upeneus doriae (Perciformes: Mullidae) from the Persian Gulf and comparison with U. sulphureus. Korean Journal of Ichthyology 15(2): 109–113.

Kim K-H, Kim YU, Kim Y-S. **1988**. Five species of fish new to Korean waters. *Korean Journal of Fisheries and Aquatic Sciences* 21(2): 105–112.

Kim SY. **2012**. Phylogenetic systematics of the family Pentacerotidae (Actinopterygii: Order Perciformes). *Zootaxa* 3366: 1–111. http://dx.doi.org/10.11646/ zootaxa.3366.1.1

Kimani EN, Mwatha GK, Wakwabi EO, Ntiba JM, Okoth BK.
1996. Fishes of a shallow tropical mangrove estuary, Gazi, Kenya. *Marine and Freshwater Research* 47(7): 857–868.

Kimura K, Imamura H, Kawai T. 2018. Comparative morphology and phylogenetic systematics of the families Cheilodactylidae and Latridae (Perciformes: Cirrhitoidea), and proposal of a new classification. *Zootaxa* 4536(1): 1–72. https://doi.org/10.11646/zootaxa.4536.1.1

Kimura K, Takagishi M, Kawai T, Imamura H, Ho H-C, Tomita T, Tanaka F, Shinohara G. **2017**. Record of a flathead fish, *Rogadius pristiger* (Cuvier, 1829) (Platycephalidae) from Taiwan. *Platax* 14: 46–54.

Kimura S, Golani D, Iwatsuki Y, Tabuchi M, Yoshino T. 2007. Redescriptions of the Indo-Pacific atherinid fishes Atherinomorus forskalii, Atherinomorus lacunosus and Atherinomorus pinguis. Ichthyological Research 54(2): 145–159. Kimura S, Ito T, Perostiwady T, Iwatsuki, Y, Yoshino T, Dunlap PV. 2005. The *Leiognathus splendens* complex (Perciformes: Leiognathidae) with the description of a new species, *Leiognathus kupanensis* Kimura and Peristiwady. *Ichthyological Research* 52(3): 275–291.

Kimura S, Iwatsuki Y, Yoshino T. 2001. Redescriptions of the Indo-West Pacific atherinid fishes, *Atherinomorus* endrachtensis (Quoy and Gaimard, 1825) and A. duodecimalis (Valenciennes in Cuvier and Valenciennes, 1835). Ichthyological Research 48(2): 167–177.

Kimura S, Katahira K, Kuriiwa K. **2013**. The red-fin *Decapterus* group (Perciformes: Carangidae) with the description of a new species, *Decapterus smithvanizi*. *Ichthyological Research* 60(4): 363–379. http://dx.doi.org/10.1007/s10228-013-0364-9

Kimura S, Kimura R, Ikejima K. 2008. Revision of the genus *Nuchequula* with descriptions of three new species (Perciformes: Leiognathidae). *Ichthyological Research* 55(1): 22–42. http://dx.doi.org/10.1007/s10228-007-0011-4

Kimura S, Peristiwady T, Suharti SR. **2003**. Engraulidae. Anchovies. Teri. (pp. 14–15). In: Kimura S, Matsuura K (eds) *Fishes of Bitung, northern tip of Sulawesi, Indonesia*. Tokyo: Ocean Research Institute, University of Tokyo. vi + 244 pp.

Kimura S, Peristiwady T, Suharti SR. 2003. Clupeidae.
Herrings and sardines. Sardin dan Tembang. (pp. 15–17).
In: Kimura S, Matsuura K (eds) *Fishes of Bitung, northern tip of Sulawesi, Indonesia.* Tokyo: Ocean Research Institute, University of Tokyo. vi + 244 pp.

Kimura S, Peristiwady T, Suharti SR. 2003. Scorpaenidae.
Scorpionfishes. Ikan lepu. (pp. 37–41). In: Kimura S,
Matsuura K (eds) *Fishes of Bitung, northern tip of Sulawesi, Indonesia.* Tokyo: Ocean Research Institute, University of Tokyo. vi + 244 pp.

Kimura S, Satapoomin U, Matsuura K (eds). **2009**. *Fishes of Andaman Sea, west coast of southern Thailand*. Tokyo: National Museum of Natural Science. vi + 346 pp.

Kimura S, Yamashita T, Iwatsuki Y. **2000**. A new species, *Gazza rhombea*, from the Indo-West Pacific, with a redescription of *G. achlamys* Jordan & Starks, 1917 (Perciformes: Leiognathidae). *Ichthyological Research* 47(1): 1–12.

King B, Qiao T, Lee MSY, Zhu M, Long JA. 2016. Bayesian morphological clock methods resurrect placoderm monophyly and reveal rapid early evolution in jawed vertebrates. *Systematic Biology* 66(4): 499–516. http://dx.doi. org/10.1093/sysbio/syw107

King D. **1996**. *Reef fishes & corals: East coast of southern Africa*. Cape Town: Struik Publishers. 128 pp.

King D. **1997**. *Reef fishes & corals: East coast of southern Africa* (2<sup>nd</sup> edition). Cape Town: Struik Publishers. 128 pp.

King D, Fraser V. **2001**. *More reef fishes and nudibranchs: East and south coast of southern Africa*. Cape Town: Struik Publishers. 136 pp. King D, Fraser V. **2014**. *The reef guide: East & south coasts of southern Africa*. Cape Town: Struik Nature. 360 pp.

King JE. 1951. Two juvenile pointed-tailed ocean sunfish, *Masturus lanceolatus*, from Hawaiian waters. *Pacific Science* 5(1): 108–109.

King JE, Ikehara II. **1956**. Some unusual fishes from the Central Pacific. *Pacific Science* 10(1): 17–24.

Kishimoto H. 2001. Uranoscopidae. Stargazers (pp. 3519– 3531). In: Carpenter KE, Niem VH (eds) FAO species identification guide for fishery purposes. The living marine resources of the western central Pacific. Vol. 6. Bony fishes part 4 (pp. 3381–4218). Rome: FAO.

Kishimoto H, Katayama M, Kohno H, Tanizawa M. **1983**. Fish fauna of Iriomote-jima, Ryukyu Islands. IV. Serranidae (Anthiinae). *Bulletin of the Institute of Oceanic Research & Development, Tokai University* 5: 33–44.

Kishimoto H, Last PR, Fujii E, Gomon MF. 1988. Revision of a deep-sea stargazer genus *Pleuroscopus*. Japanese Journal of Ichthyology 35(2): 150–158.

Kishinouye K. 1915. A study of the mackerels, cybiids, and tunas. Suisan Gakkai Ho 1(1): 1–24. [In Japanese, English translation in U.S. Fish and Wildlife Service Special Scientific Report – Fisheries 24: 1–14]

Kishinouye K. **1923**. Contributions to the comparative study of the so-called scombroid fishes. *Journal of the College of Agriculture, Imperial University of Tokyo* 8(3): 293–475.

Kittlitz FH. 1834. Beschreibung mehrerer neuer oder wenig gekannter Arten des Geschlechtes Acanthurus, im stillen Ocean, beobachtet und nach dem Leben abgebildet. Museum Senckenbergianum: Abhandlungen aus dem Gebiete der beschreibenden Naturgeschichte, von Mitgliedern der Senckenbergischen Naturforschenden Gesellschaft in Frankfurt am Main 1: 189–196, Pls. 12–13.

Klaus R, Turner JR. **2004**. The marine biotopes of the Socotra Archipelago. *Fauna of Arabia* 20: 45–115.

Klaus R, Turner J, West F. 2002. The marine biotopes of the Socotra Island Group (pp. 11–167). In: Apel M, Hariri K, Krupp F (eds) *Conservation and sustainable* use of biodiversity of Socotra Archipelago. Marine habitat, biodiversity and fisheries surveys and management. Final Report of Phase III. Senckenberg Research Institute, Frankfurt a.M., Germany.

Klausewitz W. **1955**. *Paramicrophis schmidti*, eine neue Seenadel aus Indien (Pisces, Syngnathidae). *Senckenbergiana Biologica* 36(5/6): 325–327.

Klausewitz W. 1959. Fische aus dem Roten Meer. II. Knochenfische der Familie Apogonidae (Pisces: Percomorphi). Senckenbergiana Biologica 40(5/6): 251–262.

Klausewitz W. **1960**. *Eleotriodes pallidus* n. sp. aus dem Indischen Ozean (Pisces, Gobioidea, Eleotridae). *Senckenbergiana Biologica* 41(1/2): 7–9. Klausewitz W. **1960**. Fische aus dem Roten Meer. III. *Tripterygius abeli* n sp (Pisces: Biennioidea, Clinidae). *Senckenbergiana Biologica* 41(1/2): 11–13.

Klausewitz W. **1960**. Fische aus dem Roten Meer. IV. Einige systematische und ökologisch bemerkenswerte Meergrundeln (Pisces, Gobiidae). *Senckenbergiana Biologica* 41(3/4): 149–162.

Klausewitz W. **1960**. Die typen und typoide des Naturmuseums Senckenberg, 23: Pisces, Chondrichthyes, Elasmobranchii. *Senckenbergiana Biologica* 41(5/6): 289–296.

Klausewitz W. **1960**. Fische aus dem Roten Meer. V. Über einige Fische der Gattung *Ecsenius* (Pisces, Salariidae). *Senckenbergiana Biologica* 41(5/6): 297–299.

Klausewitz W. **1961**. Über eine kleine bemerkenswerte Fischsammlung von der Küste von Pakistan. *Senckenbergiana Biologica* 42(5/6): 427–431.

Klausewitz W. **1962**. *Meiacanthus smithi* n. sp. aus dem Indischen Ozean (Pisces, Percomorphi, Blenniidae). *Senckenbergiana Biologica* 43(1): 17–19.

Klausewitz W. **1962**. *Ecsenius lineatus* n. sp. von den Malediven. *Senckenbergiana Biologica* 43(2): 145–157.

Klausewitz W. **1962**. *Gorgasia sillneri*, ein neuer Röhrenaal aus dem Roten Meer (Pisces, Apodes, Heterocongridae). *Senckenbergiana Biologica* 43(6): 433–435.

Klausewitz W. **1963**. *Ecsenius minutus* n. sp. von Malediven. *Senckenbergiana Biologica* 44(5): 357–358.

Klausewitz W. **1964**. Fische aus dem Roten Meere. VI. Taxionomische und ökologische Untersuchungen an einigen Fischarten der Küstenzone. *Senckenbergiana Biologica* 45(2): 123–144.

Klausewitz W. **1964**. Zwei neue Arten von aalartigen Fischen aus dem Indischen Ozean (Pisces, Apodes, Muraenidae, Ophichthidae). *Senckenbergiana Biologica* 45(6): 665–669.

Klausewitz W. **1966**. Fische aus dem Roten Meer. VII. *Siphamia permutata* n. sp. (Pisces, Perciformes, Apogonidae). *Senckenbergiana Biologica* 47(3): 217–222.

Klausewitz W. **1967**. Die physiographische Zonierung der Saumriffe von Sarso. *Meteor Forschungsergebnisse* (Reihe D) 2: 44–68.

Klausewitz W. **1968**. Fische aus dem Roten Meer. VIII. *Biat magnusi* n. sp., eine neue Meergrundel (Pisces, Osteichthyes, Gobiidae). *Senckenbergiana Biologica* 49(1): 13–17.

Klausewitz W. **1968**. Fische aus dem Roten Meer. IX. *Pseudochromis fridmani* n. sp. aus dem Golf von Aqaba (Pisces, Osteichthyes, Pseudochromidae). *Senckenbergiana Biologica* 49(6): 443–450.

Klausewitz W. **1969**. Fische aus dem Roten Meer. XI. *Cryptocentrus sungami* n. sp. (Pisces, Gobiidae). *Senckenbergiana Biologica* 50(1/2): 41–46. Klausewitz W. **1969**. *Nemapterois biocellatus* Fowler, ein Neunachweis für den Indischen Ozean (Pisces, Scleropareiformes, Scorpaenidae). *Senckenbergiana Biologica* 50(5/6): 347–351.

Klausewitz W. **1970**. *Sebastapistes hassi* n. sp. von den Malediven (Pisces: Scleropareiformes: Scorpaenidae). *Senckenbergiana Biologica* 51(1/2): 73–75.

Klausewitz W. **1970**. Wiederfund von *Lotilia graciliosa* (Pisces: Gobiidae). *Senckenbergiana Biologica* 51(3/4): 177–179.

Klausewitz W. **1970**. *Amphiprion allardi* n. sp., ein neuer Anemonfisch von der ostafrikanischen Küste (Pisces: Pomacentnidae). *Senckenbergiana Biologica* 51(3/4): 181–192.

Klausewitz W. 1971. Poeciloconger fasciatus Günther, ein Neunachweis für Madagascar (Pisces: Apodes: Congridae). Senckenbergiana Biologica 52(3/5): 197–200.

Klausewitz W. **1972**. Litoralfische der Malediven. I. Einleitung und Fische der Ordnung Syngnathiformes (Pisces: Teleostei). *Senckenbergiana Biologica* 53(3/4): 199–217.

Klausewitz W. **1972**. The zoogeographical and paleogeographical problem of the Indian Ocean and the Red Sea according to the ichthyofauna of the littoral. *Journal of the Marine Biological Association of India* 14(2): 697–706.

Klausewitz W. **1974**. Litoralfische der Malediven. IV. Die Familie der Drückerfische, Balistidae (Pisces: Tetraodontiformes, Balistoidei). *Senckenbergiana Biologica* 55(1/3): 39–67.

Klausewitz W. **1974**. Fische aus dem Roten Meer. XIII. *Cryptocentrus steinitzi* n. sp., ein neuer "Symbiose-Gobiide" (Pisces: Gobiidae). *Senckenbergiana Biologica* 55(1/3): 69–76.

Klausewitz W. **1974**. Fische aus dem Roten Meer. XIV. *Eilatia latruncularia* n. gen. n. sp. und *Vanderhorstia mertensi* n. sp. vom Golf von Aqaba. *Senckenbergiana Biologica* 55(4/6): 205–212.

Klausewitz W. **1975**. *Cabillus anchialinae*, eine neue Meergrundel von der Sinai-Halbinsel (Pisces: Gobiidae: Gobiinae). *Senckenbergiana Biologica* 56(4/6): 203–207.

Klausewitz W. **1980**. Tiefenwasser- und Tiefseefische aus dem Roten Meer. I. Einleitung und Neunachweis für *Bembrops adenensis* Norman 1939 und *Histiopterus spinifer* Gilchrist 1904 (Pisces: Perciformes: Percophididae, Pentacerotidae). *Senckenbergiana Biologica* 61(1/2): 11–24.

Klausewitz W. 1982. Tiefenwasser- und Tiefseefische aus dem Roten Meer. V. Über die vertikale Verbreitung von *Champsodon omanensis* Regan. Senckenbergiana Maritima 14(1/2): 39–45.

Klausewitz W. 1983. Tiefenwasser-und Tiefseefische aus dem Roten Meer. VII. *Harpadon erythraeus* n. sp. aus der Tiefsee des zentralen Roten Meeres (Pisces: Teleostei: Scopelomorpha: Myctophiformes: Harpadontidae). *Senckenbergiana Biologica* 64(1–3): 34–45. Klausewitz W. **1985**. Fische aus dem Roten Meer. XVII. A new species of the genus *Stalix* from the Gulf of Aqaba, Red Sea (Pisces: Teleostei: Perciformes: Opistognathidae). *Revue française d'Aquariologie Herpétologie* 12(1): 17–22.

Klausewitz W. **2002**. Frankfurt vs Berlin: the Red Sea explorers Wilhelm Hemprich, Christian Ehrenberg and Eduard Rüppell. *Zoology in the Middle East* 27: 7–12.

Klausewitz W, Bauchot M-L. **1967**. Réhabilitation de *Sphryraena forsteri* Cuvier in Cuv. & Val. 1829, et désignation d'un néotype (Pisces, Mugiliformes, Sphryaenidae). *Bulletin du Muséum National d'Histoire Naturelle* (Paris) (Série 2) 39(1): 117–120.

Klausewitz W, Eibl-Eibesfeldt I. **1959**. Neue Röhrenaale von den Maldiven und Nikobaren (Pisces, Apodes, Heterocongridae). *Senckenbergiana Biologica* 40(3/4): 135–153.

Klausewitz W, Fricke HW. **1985**. On the occurrence of *Chaetodon jayakari* Norman in the deep water of the Gulf of Aqaba, Red Sea. *Senckenbergiana Maritima* 17(1/3): 1–13.

Klausewitz W, Frøiland Ø. **1970**. *Scorpaenodes steinitzi* n. sp. von Eilat, Golf von Aqaba (Pisces: Scorpaenidae). *Senckenbergiana Biologica* 51(5/6): 317–321.

Klausewitz W, McCosker JE, Randall JE, Zetzsche H.
1978. Hoplolatilus chlupatyi n. sp., un nouveau poisson marin des Philippines (Pisces, Perciformes, Percoidei, Branchiostegidae). Revue française d'Aquariologie Herpétologie 5(2): 41–48.

Klausewitz W, Nielsen JG. **1965**. On Forsskål's collection of fishes in the Zoological Museum of Copenhagen. *Spolia Zoologica Musei Hauniensis* 22: 1–29.

Klausewitz W, Schneider M. 1986. Tiefenwasser- und Tiefseefische aus dem Roten Meer. XII. Arnoglossus marisrubri n. sp. aus dem Mesobenthos des zentralen Roten Meeres und A. kotthausi nom. nov. vom Epibenthos des nordwestlichen Indischen Ozeans. (Pisces: Pleuronectiformes: Bothidae: Bothinae). Senckenbergiana Maritima 18(3–6): 217–227, Pl. 1. [English summary.]

Klausewitz W, Wongratana T. **1970**. Vergleichende Untersuchungen an *Apolemichthys xanthurus* und *xanthotis* (Pisces: Perciformes: Pomacanthidae). *Senckenbergiana Biologica* 51(5/6): 323–332.

Klausewitz W, Zander CD. **1967**. *Acentrogobius meteori* n. sp. (Pisces, Gobiidae). *Meteor Forschungsergebnisse* (Reihe D) 2: 85–87.

Kleiber D, Blight LK, Caldwell IR, Vincent ACJ. **2011**. The importance of seahorses and pipefishes in the diet of marine animals. *Reviews in Fish Biology and Fisheries* 21(2): 205–223. http://dx.doi.org/10.1007/s11160-010-9167-5

Klein JT. 1776–1777. Neuer Schauplatz der Natur, nach den Richtigsten Beobachtungen und Versuchen, in alphabetischer Ordnung, vorgestellt durch eine Gesellschaft von Gelehrten. Vols. 3 (1766) and 4 (1777). Leipzig: Weidmann. Klein R, Tudhope AW, Chilcott CP, Pätzold J, Abdulkarim Z, Fine M, Fallick AE, Loya Y. **1997**. Evaluating southern Red Sea corals as a proxy record for the Asian monsoon. *Earth and Planetary Science Letters* 148: 381–394.

Klunzinger CB. 1870. Synopsis der fische des Rothen Meeres. I. Theil. Percoiden-Mugiloiden. Verhandlungen der Kaiserliche-Koniglichen zoologisch-botanischen Gesellschaft in Wien 20: 669–834.

Klunzinger CB. 1871. Synopsis der fische des Rothen Meeres.
 II. Theil. Verhandlungen der Kaiserliche-Koniglichen
 Zoologisch-Botanischen Gesellschaft in Wien 21: 441–688.

Klunzinger CB. 1871. Systematische Uebersicht der Fische des Rothen Meers als Anhang und Register zur Synopsis. Verhandlungen der Kaiserliche-Koniglichen Zoologische und Botanische Gesellschaft in Wien 21: 1353–1367. [Indexes to Klunzinger 1870 & 1871 above]

Klunzinger CB. 1879. Die v. Müller'sche Sammlung australischer Fische. Anzeiger der Kaiserlichen Akademie der Wissenschaften, Wien, Mathematisch-Naturwissenschaftliche Classe 16(22): 254–261.

Klunzinger CB. 1884. Die fische de Rothen Meeres. Eine kritische Revision mit Bestimmungstabellen. Teil I. Acanthopteri veri Owen. Stuttgart: E. Schweizerbart'sche Verlagshandlung (E. Koch). ix + 133 pp.

Knapp L, Mincarone MM, Harwell H, Polidoro B, Sanciangco J, Carpenter K. 2011. Conservation status of the world's hagfish species and the loss of phylogenetic diversity and ecosystem function. *Aquatic Conservation: Marine* and Freshwater Ecosystems 21(5): 401–411. http://dx.doi. org/10.1002/aqc.1202

Knapp LW. 1973. Platycephalus beauforti, a new species of flathead (Pisces, Platycephalidae) from the western Pacific. Proceedings of the Biological Society of Washington 86(10): 117–125.

Knapp LW. 1979. Fische des Indischen Ozeans. Ergebnisse der ichthyologischen Untersuchungen während der Expedition des Forschungsschiffes *Meteor* in den Indischen Ozean, Oktober 1964 bis 1965. A. Systematischer Teil, XXII, Scorpaeniformes (Platycephalidae) 4. Fische des Indischen Ozeans. *Meteor Forschungsergebnisse* (Reihe D, Biologie) 29: 48–54.

Knapp LW. 1981. Percophidae. Duckbills. In: Fischer W, Bianchi G, Scott WB (eds) FAO species identification sheets for fishery purposes. Eastern central Atlantic (Fishing Area 34 and part of 47). Vol. 3. Rome: FAO. [unpaginated]

Knapp LW. 1984. Platycephalidae. Spiny flatheads. In: Fischer
W, Bianchi G (eds) FAO species identification sheets for fishery purposes. Western Indian Ocean (Fishing Area 51).
Vol. 3. Rome: FAO. [unpaginated] Knapp LW. **1996**. Review of the genus *Cociella* Whitley (Teleostei: Platycephalidae) with the description of three new species. *Proceedings of the Biological Society of Washington* 109(1): 17–33.

Knapp LW. 1999. Platycephalidae. Flatheads (pp. 2385–2421).
In: Carpenter K, Niem VH (eds) FAO species identification guide for fishery purposes. The living marine resources of the western central Pacific. Vol. 4. Bony fishes Part 2 (pp. 2069–2790). Rome: FAO.

Knapp LW. 2012. Rogadius fehlmanni. A new flathead fish (Scorpaeniformes: Platycephalidae) from Somalia. Proceedings of the Biological Society of Washington 125(1): 61–65. http://dx.doi.org/10.2988/11-20.1

Knapp LW. 2013. Descriptions of four new species of *Thysanophrys* (Scorpaeniformes: Platycephalidae) from the Western Indian Ocean. *Zootaxa* 3608(2): 127–136. https://doi.org/10.11646/zootaxa.3608.2.3

Knapp LW, Heemstra PC. **2011**. *Sorsogona humerosa*, a new flathead fish (Scorpaeniformes: Platycephalidae) from the western Indian Ocean. *Smithiana* Bulletin 13: 75–78.

Knapp LW, Imamura H. 1997. Status of *Platycephalus cantori* Bleeker, 1879 (Teleostei: Platycephalidae). *Proceedings of the Biological Society of Washington* 110(3): 384–387.

Knapp LW, Imamura H. **2014**. *Grammoplites vittatus* (Valenciennes), (Scorpaeniformes, Platycephalidae), removed from synonymy with *Grammoplites scaper* (Linnaeus). *Zootaxa* 3846(3): 447–450. https://doi. org/10.11646/zootaxa.3846.3.8

Knapp LW, Imamura H, Sakashita M. 2000. Onigocia bimaculata, a new species of flathead fish (Scorpaeniformes: Platycephalidae) from the Indo-Pacific. J.L.B. Smith Institute of Ichthyology Special Publication 64: 1–10.

Knapp LW, Randall JE. **2013**. *Sunagocia omanensis*, a new flathead fish (Scorpaeniformes: Platycephalidae) from the Western Indian Ocean, with comments on the distribution of *Sunagocia carbuncula*. *Zootaxa* 3718(1): 97–100. https://doi.org/10.11646/zootaxa.3718.1.9

Knapp LW, Wongratana T. 1987. Sorsogona melanoptera, a flathead fish from the northern Indian Ocean (Teleostei: Platycephaldiae). Proceedings of the Biological Society of Washington 100(2): 381–385.

Kneebone J, Natanson LJ, Andrews AH, Howell WH. **2008**. Using bomb radiocarbon analyses to validate age and growth estimates for the tiger shark, *Galeocerdo cuvier*, in the western North Atlantic. *Marine Biology* 154: 423–434. https://doi.org/10.1007/s00227-008-0934-y

Kner R. 1860. Über einige noch unbeschriebene Fische. Sitzungsberichte der Kaiserlichen Akademie der Wissenschaften. Mathematisch-Naturwissenschaftliche Classe 39(4): 531–547.

- Kner R. 1864. Specielles Verzeichniss der während der Reise der kaiserlichen Fregatte Novara gesammelten Fische. Sitzungsberichte der Kaiserlichen Akademie der Wissenschaften. Mathematisch-Naturwissenschaftliche Classe 49: 481–486.
- Kner R. 1865. Fische. In: Reise der öesterreichischen Fregatte Novara um die Erde in den Jahren 1857–59, unter den Befehlen des Commodore B. von Wüllerstorf-Urbair.
  Vol. 1(1). Wien: Der Kaiserlich-Königlichen Hof- und Staatsdruckerei. pp. 1–109, Pls. 1–5.
- Kner R. 1867. Fische. In: Reise der österreichischen Fregatte Novara um die Erde in den Jahren 1857, 1858, 1859 unter den Befehlen des Commodore B. von Wüllerstorf-Urbair. Vol. 1(3). Wien: Der Kaiserlich-Königlichen Hof- und Staatsdruckerei. pp. 275–433.
- Kner R. 1868. Über neue Fische aus dem Museum der Herren Johann Cäsar Godeffroy & Sohn in Hamburg. Sitzungsberichte der Mathematisch-Naturwissenschaftliche Classe der Kaiserlichen Akademie der Wissenschaften 58(1–2): 26–31.
- Kner R. 1868. IV. Folge neuer Fische aus dem Museum der Herren Johann Cäsar Godeffroy und Sohn in Hamburg. Sitzungsberichte der Mathematisch-Naturwissenschaftliche Classe der Kaiserlichen Akademie der Wissenschaften 58(3–4): 293–356.
- Kner R, Steindachner F. **1867**. Neue Fische aus dem Museum der Herren Johan Cäsar Godeffroy & Sohn in Hamburg. *Sitzungsberichte der Mathematisch-Naturwissenschaftliche Classe der Kaiserlichen Akademie der Wissenschaften* 54(3): 356–395.
- Knudsen SW, Clements KD. 2013. Revision of the family Kyphosidae (Teleostei: Perciformes). Zootaxa 3751(1): 1–101. https://doi.org/10.11646/zootaxa.3751.1.1
- Kobayashi H, Ichikawa T, Suzuki H, Sekimoto M. **1972**. Seasonal migration of the hagfish, *Eptatretus burgeri*. *Japanese Journal of Ichthyology* 19(3): 191–194.
- Kobayashi K, Suzuki K. **1992**. Hermaphroditism and sexual function in *Cirrhitichthys aureus* and the other Japanese hawkfishes (Cirrhitidae: Teleostei). *Japanese Journal of Ichthyology* 38(4): 397–410.
- Kobyliansky SG. 1998. Four new Indo-Pacific species and a new key to species of the genus *Glossanodon* (Argentinidae). *Voprosy Ikhtiologii* 38(6): 725–736. [In Russian. English translation in *Journal of Ichthyology* 38(9): 697–707]
- Kobyliansky SG. 2013. Two new species of green eyes of the genus *Chlorophthalmus* (Chlorophthalmidae, Aulopidae) from the continental slope and submarine rises of the western tropical part of the Indian Ocean. *Journal of Ichthyology* 53(6): 373–379. https://doi.org/10.1134/ S0032945213030077

Kochzius M, Blohm D. **2005**. Genetic population structure of the lionfish *Pterois miles* (Scorpaenidae, Pteroinae) in the Gulf of Aqaba and northern Red Sea. *Gene* 347(2): 295–301.

- Kochzius M, Seidel C, Antoniou A, Botla SK, Campo D, Cariani A, Garcia Vazquez E, Hauschild J, Hervet C, Hjörleifsdottir S, Hreggvidsson G, Kappel K, Landi M, Magoulas A, Marteinsson V, Nölte M, Planes S, Tinti F, Turan C, Venugopal MN, Weber H, Blohm D. 2010. Identifying fishes through DNA barcodes and microarrays. *PLoS ONE* 5(9): e12620. http://dx.doi.org/10.1371/journal. pone.0012620
- Kochzius M, Söller R, Khalaf MA, Blohm D. 2003. Molecular phylogeny of the lionfish genera *Dendrochirus* and *Pterois* (Scorpaenidae, Pteroinae) based on mitochondrial DNA sequences. *Molecular Phylogenetics and Evolution* 28(3): 396–403.
- Koeda K, Teramura A. 2019. Redescription of *Tetragonurus pacificus* (Teleostei: Stromateoidei: Tetragonuridae), based on specimens collected from Taiwan and Tarawa Atoll.
  In: Ho H-C, Koeda K, Hilton EJ (eds) Study on the fish taxonomy and diversity of Taiwan. *Zootaxa* 4702(1): 26–31. https://doi.org/10.11646/zootaxa.4702.1.7
- Koeda K, Yoshino T, Imai H, Tachihara K. 2014. A review of the genus *Pempheris* (Perciformes, Pempheridae) of the Red Sea, with description of a new species. *Zootaxa* 3793(3): 301–330. https://doi.org/10.11646/zootaxa.3793.3.1
- Koelreuter IT. 1766. Piscium rariorum e museo Petropolitano exceptorum descriptiones continuatae. Novi Commentarii Academiae Scientiarum Imperialis Petropolitanae 10 [for 1764]: 329–351, Pl. 8.
- Kohno H. **1984**. Osteology and systematic position of the butterfly mackerel, *Gasterochisma melampus*. *Japanese Journal of Ichthyology* 31(3): 268–286.
- Kok HM, Blaber SJM. **1977**. A new freshwater goby (Teleostei: Gobiidae) from the Pongola floodplain, Zululand, South Africa. *Zoologica Africana* 12(1): 163–168.
- Kondritskaya S. **1970**. The larvae of the swordfish (*Xiphias gladius* (L.)) from Mozambique Channel. *Journal of Ichthyology* 10(6): 853–854.
- Konishi K. **1999**. Developmental and comparative morphology of beryciform larvae (Teleostei: Acanthomorpha), with comments on trachichthyoid relationships. *Bulletin of Seikai National Fisheries Research Institute* 77: 23–92.
- Konovalenko II, Piotrovsky AS. **1988**. First description of a sexually mature amarsipa, *Amarsipus carlsbergi. Journal of Ichthyology* 28(5): 86–89.
- Kossmann R, Räuber H. **1877**. Fische. Wissenschftliche Reise in die Küstengebiete des Rothen Meeres. *Verhandlungen des Naturhistorisch-Medizinischen Vereins zu Heidelberg* 1: 378–420, Pl. 3.

Kossmann R, Räuber H. 1877. Pisces. In: Zoologische Ergebnisse einer im Auftrage der Königlichen Academie der Wissenschaften zu Berlin ausgeführten Reise in Küstengebiete des Rothen Meeres (Erste Hälfte). Leipzig: Wilhelm Engelmann. pp. 3–34, Pls. 1–2.

Kotlyar AN. 1978. A contribution to the taxonomy of the "armless" flounders (Pisces: Bothidae) from southwestern Atlantic. *Voprosy Ikhtiologii* 18(5): 799–813. [In Russian. English translation in *Journal of Ichthyology* 18(5): 708–721]

Kotlyar AN. 1978. Hoplostethus (Hoplostethus) natalensis sp.
N., a new species of the family Trachichthyidae (Berycoidei, Beryciformes) from the south-western part of the Indian Ocean. Zoologicheskii Zhurnal 57(11): 1681–1685.
[In Russian, English summary]

Kotlyar AN. 1979. Paratrachichthys (Aulotrachichthys) sajademalensis, sp. n., a new species of the family Trachichthyidae (Beryciformes) from the Indian Ocean. Voprosy Ikhtiologii 19(4, art.117): 730–732. [In Russian. English translation in Journal of Ichthyology 19(4): 137–140]

Kotlyar AN. **1980**. Systematics and distribution of trachichthyid fishes (Trachichthyidae, Beryciformes) in the Indian Ocean. *Trudy Instituta Okeanologii Imeni P.P. Shirshova* 110: 177–224. [In Russian, English summary]

Kotlyar AN. 1982. First finding of *Parazen pacificus* Kamohara (Zeidae) and *Pentaceros richardsoni* Smith (Pentacerotidae) in the Indian Ocean. *Byulleten Moskovskogo Obshchestva Ispytatelei Prirody Otdel Biologicheskii [Bulletin of the Moscow Society of Naturalists Biological Series]* 87(3): 34–36. [In Russian]

Kotlyar AN. **1984**. Systematics and distribution of fishes of the family Polymixiidae (Polymixioidei, Beryciformes). *Voprosy Ikhtiologii* 24(5): 691–708. [In Russian. English translation in *Journal of Ichthyology* 24(6): 1–20]

Kotlyar AN. 1985. Taxonomy and distribution of Monocentridae (Beryciformes). *Journal of Ichthyology* 25(4): 91–106.

Kotlyar AN. **1986**. On the biology of *Polymixia berndti* Gilbert (Polymixiidae) in the western part of the Indian Ocean. *Journal of Ichthyology* 26(2): 120–127.

Kotlyar AN. 1986. Systematics and distribution of species of the genus *Hoplostethus* Cuvier (Beryciformes, Trachichthyidae). *Trudy Instituta Okeanologii Imeni P.P. Shirshova* 121: 97–140. [In Russian, English summary]

Kotlyar AN. **1987**. Age and growth of alfoncino, *Beryx splendens*. *Journal of Ichthyology* 27(2): 104–111.

Kotlyar AN. **1992**. A new species of the genus *Polymixia* (Polymixiidae, Beryciformes) from the Kyushu- Palau submarine ridge and notes on the other members of the genus. *Journal of Ichthyology* 33(3): 30–49.

Kotlyar AN. **1993**. Beryciform fishes from the western Indian collected in cruise of R/V *Vityaz*. *Trudy Instituta Okeanologii Imeni P.P. Shirshova* 128: 179–198. [In Russian]

Kotlyar AN. **1993**. Discoveries of very large specimens of *Polymixia busakhini* (Polymixiidae) from the eastern shores of Australia. *Journal of Ichthyology* 33(8): 145–147.

Kotlyar AN. **1996**. *Beryciform fishes of the world ocean*. Moscow: VNIRO Publishing. 368 pp. [In Russian]

Kotlyar AN. **2001**. A rare zeid species – *Parazen pacificus*: osteology, systematics, and distribution (Parazenidae, Zeiformes). *Journal of Ichthyology* 41(2): 687–697.

Kotlyar AN. **2002**. New data on beryciform fishes (Beryciformes) from the South China Sea. *Journal of Ichthyology* 42(3): 475–480.

Kottelat M. 2001. Scatophagidae. Scats (pp. 3623–3626). In: Carpenter KE, Niem VH (eds) FAO species identification guide for fishery purposes. The living marine resources of the western central Pacific. Vol. 6. Bony fishes part 4 (pp. 3381–4218). Rome: FAO.

Kottelat M. **2013**. Nomenclature and identity of the tongue soles *Paraplagusia bilineata*, "*Cynoglossus bilineatus*" and *Paraplagusia blochii* (Teleostei: Pleuronectiformes). *Raffles Bulletin of Zoology* 61(2): 763–766.

Kottelat M. **2013**. The fishes of the inland waters of southeast Asia: a catalogue and core bibliography of the fishes known to occur in freshwaters, mangroves and estuaries. *Raffles Bulletin of Zoology* Supplement 27: 1–663.

Kottelat M, Whitten AJ, Kartikasari SN, Wirjoatmodjo S. **1993**. *Freshwater fishes of Western Indonesia and Sulawesi*. Jakarta, Indonesia: Periplus Editions Ltd. 221 pp.

Kotthaus A. **1966**. Fischforschung im Indischen Ozean *Meteor* Expedition 1964–65. *Umschau in Wissenschaft und Technik* jahrgang heft 1966: 118–123.

Kotthaus A. **1967**. Fische des Indischen Ozeans. A. Systematischer Teil I & II. *Meteor Forschungsergebnisse* (Reihe D, Biologie) 1: 1–84.

Kotthaus A. **1968**. Fische des Indischen Ozeans. A. Systematischer Teil III. Ostariophysi und Apodes. *Meteor Forschungsergebnisse* (Reihe D, Biologie) 3: 14–56. www.schweizerbart.de/publications/list/series/meteor

Kotthaus A. 1969. Fische des Indischen Ozeans. Ergebnisse der ichthyologischen Untersuchungen während der Expedition des Forschungsschiffes *Meteor* in den Indischen Ozean, Oktober 1964 bis Mai 1965. A. Systematischer Teil IV: Synentognathi. *Meteor Forschungsergebnisse* (Reihe D, Biologie) 4: 1–30. www.schweizerbart.de/publications/list/ series/meteor

Kotthaus A. **1970**. Fische des Indischen Ozeans. Ergebnisse der ichthyologischen Untersuchungen während der Expedition des Forschungsschiffes *Meteor* in den Indischen Ozean, Oktober 1964 bis Mai 1965. A. Systematiscer Teil, VII Percomorphi (1). *Meteor Forschungsergebnisse* (Reihe D, Biologie) 6: 43–55.

Kotthaus A. 1973. Fische des Indischen Ozeans. Ergebnisse der ichthyologischen Untersuchungen während der Expedition des Forschungsschiffes *Meteor* in der Indischen Ozean, Oktober 1964 bis Mai 1965. A. Systematischer Teil, X Percomorphi (3). *Meteor Forschungsergebnisse* (Reihe D, Biologie) 16: 17–32.

Kotthaus A. 1974. Fische des Indischen Ozeans. Ergebnisse der ichthyologischen Untersuchungen während der Expedition des Forschungsschiffes *Meteor* in den Indischen Ozean, Oktober 1964 bis Mai 1965. A. Systematischer Teil, XI. Percomorphi (4). *Meteor Forschungsergebnisse* (Reihe D, Biologie) 17: 33–54.

Kotthaus A. 1976. Fische de Indischen Ozeans. Ergebnisse der ichthyologischen Untersuchungen während der Expedition des Forschungsschiffes *Meteor* in den Indischen Ozean, Oktober 1964 bis Mai 1965. A. Systematischer Teil, XVII, Percomorphi (7). *Meteor Forschungsergebnisse* (Reihe D, Biologie) 23: 45–61.

Kotthaus A. 1977. Fische des Indischen Ozeans. Ergebnisse der ichthyologischen Untersuchungen während der Expedition des Forschungsschiffes *Meteor* in den indischen Ozean, Oktober 1964 bis Mai 1965. A. Systematischer Teil, XVIII. Percomorphi (8). *Meteor Forschungsergebnisse* (Reihe D, Biologie) 24: 37–53.

Kotthaus A. 1977. Fische des Indischen Ozeans. Ergebnisse der ichthyologischen Untersuchungen während der Expedition des Forschungsschiffes *Meteor* in den Indischen Ozean, Oktober 1964 bis Mai 1965. A. Systematischer Teil, XIX. Percomorphi (9). *Meteor Forschungsergebnisse* (Reihe D, Biologie) 25: 24–44.

Kotthaus A. 1977. Fische des Indischen Ozeans. Ergebnisse der ichthyologischen Untersuchungen während der Expedition des Forschungsschiffes *Meteor* in den Indischen Ozean, Oktober 1964 bis Mai 1965. A. Systematischer Teil, XX. Pleuronectiformes (Heterosomata). *Meteor Forschungsergebnisse* (Reihe D, Biologie) 26: 1–20.

Kotthaus A. 1979. Fische des Indischen Ozeans. Ergebnisse der ichthyologischen Untersuchungen während der Expedition des Forschungsschifes *Meteor* in den Indischen Ozean, Oktober 1964 bis Mai 1965. A. Systematischer Teil, XXI. Diverse Ordungen. *Meteor Forschungsergebnisse* (Reihe D, Biologie) 28: 6–54. www.schweizerbart.de/publications/list/ series/meteor

Koumans FP. **1941**. Gobioid fishes of India. *Memoirs of the Indian Museum* 13(3): 205–329.

Koumans FP. 1944. In: Blegvad H, Løppenthin B.

Koumans FP. **1953**. Gobioidea. In: Weber M, De Beaufort LF (eds) *The fishes of the Indo-Australian Archipelago*. Vol. 10. Leiden: E.J. Brill. xiii + 423 pp. Koumans FP. **1953**. Biological results of the *Snellius* Expedition. XVI. The Pisces and Leptocardii of the *Snellius* expedition. *Temminckia* (Leiden) 9: 177–275.

Kovačić M, Bogorodsky SV. **2013**. Two new species of *Cabillus* (Perciformes: Gobiidae) and the first record of *Cabillus macrophthalmus* from the Western Indian Ocean. *Zootaxa* 3717(2): 179–194. https://doi.org/10.11646/zootaxa.3717.2.4

Kovačić M, Bogorodsky SV. **2014**. A new species of *Hetereleotris* (Perciformes: Gobiidae) from the Red Sea. *Zootaxa* 3764(4): 474–481. http://dx.doi.org/10.11646/ zootaxa.4608.3.5

Kovačić M, Bogorodsky SV, Mal AO. **2014**. A new species of *Hetereleotris* (Perciformes: Gobiidae) from Farasan Island (Red Sea). *Zootaxa* 3846(1): 119–126. https://doi. org/10.11646/zootaxa.3846.1.6

Kovačić M, Bogorodsky SV, Mal AO. **2014**. Two new species of *Coryogalops* (Perciformes: Gobiidae) from the Red Sea. *Zootaxa* 3881(6): 513–531. http://dx.doi.org/10.11646/ zootaxa.3881.6.2

Kovačić M, Bogorodsky SV, Randall JE. **2011**. Redescription of the Red Sea gobiid fish *Ctenogobiops maculosus* (Fourmanoir) and validation of *C. crocineus* Smith. *Zootaxa* 3054: 60–68. https://doi.org/10.11646/zootaxa.3054.1.2

Kovalevskaya NV. **1964**. Materials to embryonal development of flying fishes of the genus *Exocoetus* (Pisces, Exocoetidae). *Trudy Instituta Okeanologii* 73: 204–233.

Kovalevskaya NV. **1965**. Eggs and juveniles of beloniform fishes (Beloniformes, Pisces) from Tonkin Bay. *Trudy Instituta Okeanologii* 53: 124–146.

Kovalevskaya NV. **1972**. Data on reproduction, development and distribution of larvae and juveniles of flying fishes genus *Hirundichthys* (Exocoetidae, Pisces) in the Pacific and Indian Oceans. *Trudy Instituta Okeanologii* 93: 42–69.

Kovalevskaya NV. **1975**. Larvae and juveniles of *Cheilopogon* (*Ptenichthys*) intermedius Parin (Exocoetidae, Beloniformes, Pisces). Voprosy Ikhtiologii 15(5): 747–749.

Kovalevskaya NV. **1977**. Larvae and juveniles of flying fishes of family Exocoetidae – *Cheilopogon cyanopterus* (Val.), *Ch. spilonotopterus* (Bleeker), *Ch. longibarbus* Parin. *Voprosy Ikhtiologii* 17(2) (103): 291–300.

Kovalevskaya NV. **1980**. Reproduction, development and distribution patterns of larvae and juveniles of the oceanic flying fishes in the Pacific and Indian Oceans. *Transactions of the P.P. Shirshov Institute of Oceanology* 97: 212–275. [In Russian]

Kovalevskaya NV. **1982**. Materials on development of flyingfishes (Exocoetidae, *Cypselurus* s.str.). *Trudy Instituta Okeanologii* 118: 107–119.

Koya KPS, Akhilesh KV, Bineesh KK. 2011. A new record of Titan cardinalfish, *Holapogon maximus* (Apogonidae) along the south-west coast of India. *Marine Biodiversity Records* 4(e36): 1–2. https://doi.org/10.1017/S1755267211000340 Krabbenhoft TJ, Munroe TA. 2003. *Symphurus bathyspilus*: A new cynoglossid flatfish (Pleuronectiformes: Cynoglossidae) from deepwaters of the Indo-West Pacific. *Copeia* 2003(4): 810–817.

Krechmer M. **1994**. Marine oddities: batfishes. *Tropical Fish Hobbyist* 42(6#456): 40–45.

Krefft G. 1961. Beryx splendens, ein Erstfund in nordeuropäischen Gewässern nebst Bemerkungen zum Auftreten von B. decadactylus. Archiv für Fischereiwissenschaft 12(1): 24–38.

Krefft G. 1964. Platycephalus indicus (Linnaeus), 1758, ein ueues Faunenelement der ägyptischen Mittelmeerküste. Archiv für Fischereiwissenschaft 14(3): 148–152.

Krefft G. 1968. Neue und erstmalig nachgewiesene Knorpelfische aus dem Archibenthal des Südwestatlantiks, einschließlich einer Diskussion einiger *Etmopterus*-Arten südlicher Meere. Archiv für Fischereiwissenschaft 19(1): 1–2.

Krefft G. **1968**. Knorpelfische (Chondrichthyes) aus dem tropischen Ostatlantik. *Atlantide Report* 10: 33–76.

Krefft G. 1969. Ergebnisse der Forschungsreisen des FFS Walther Herwig nach Südamerika. VI. Fische der Familie Centrolophidae (Perciformes, Stromateoidei). Archiv für Fischereiwissenschaft 20(1): 1–9.

Krefft G. **1974**. Investigations on midwater fish in the Atlantic Ocean. *Berichte der Deutschen Wissenschaftlichen Kommission für Meeresforschung* 23(3): 226–254.

Krefft G. 1976. Ergebnisse der Forschungsreisen des FFS Walther Herwig nach Südamerika. XLI. Fische der Ordnung Beryciformes aus dem Sudwestatlantik. Archiv für Fischereiwissenschaft 26(2–3): 65–86.

Krefft G. **1976**. Distribution patterns of oceanic fishes in the Atlantic Ocean. *Revue des Travaux de l'Institut des Pêches maritimes* 40(3–4): 439–460.

Krefft G. **1980**. Results of the research cruises of FRV *Walther Herwig* to South America. LIII. Sharks from the pelagic trawl catches obtained during Atlantic transects, including some specimens from other cruises. *Archiv für Fischereiwissenschaft* 30(1): 1–16.

Krefft G. 1990. Chimaeridae (pp. 111–113), Rhinochimaeridae (pp. 114–116), Callorynchidae (p. 117). In: Quéro J-C, Hureau J-C, Karrer C, Post A, Saldanha L (eds) *Check-list of the fishes of the eastern tropical Atlantic (CLOFETA)*.
Vol. 1. Lisbon: Junta Nacional de Investigação Científica e Tecnológica.

Krefft G, Tortonese E. 1973. Oxynotidae (pp. 35–36), Squalidae (pp. 37–48). In: Hureau J-C, Monod T (eds) Check-list of the fishes of the north-eastern Atlantic and of the Mediterranean (CLOFNAM). Vol. I. Paris: UNESCO.

Krefft JLG. **1868**. *Deratoptera alfredi* (Prince Alfred's ray). *The Illustrated Sydney News* 5(11 July 1868): 3, 9.

Krishnan S, Mishra SS. **1993**. On a collection of fish from Kakinada–Gopalpur sector of the east coast of India. *Records of the Zoological Survey of India* 93(1–2): 201–240.

Krishnan S, Mishra SS. **1994**. On a collection of fish from middle and south Andaman Group of islands. *Records of the Zoological Survey of India* 94(2–4): 265–306.

Kriwet J. 2003. Dental morphology of the pycnodontid fish †Stemmatodus rhombus (Agassiz 1844) (Neopterygii, †Pycnodontiformes) from the Early Cretaceous with comments on its systematic position. Transactions of the Royal Society of Edinburgh: Earth Sciences 94(2): 145–155.

Kriwet J. **2004**. A new pycnodont fish genus (Neopterygii: Pycnodontiformes) from the Cenomanian (Upper Cretaceous) of Mount Lebanon. *Journal of Vertebrate Paleontology* 24(3): 525–532.

Kriwet J. 2005. A comprehensive study of the skull and dentition of pycnodont fishes. *Zitteliana* (Series A) 45: 135–188.

Kriwet J. 2008. The dentition of the enigmatic pycnodont fish, *Athrodon wittei* (Fricke, 1876) (Neopterygii, Pycnodontiformes; Late Jurassic; NW Germany). *Fossil Record* 11(2): 61–66. http://dx.doi.org/10.1002/ mmng.200800002

Kriwet J, Klug S. **2004**. Late Jurassic selachians (Chondrichthyes, Elasmobranchii) from southern Germany: re-evaluation on taxonomy and diversity. *Zitteliana* A 44: 67–95.

Kriwet J, Schmitz L. **2005**. New insight into the distribution and palaeobiology of the pycnodont fish *Gyrodus*. *Acta Palaeontologica Polonica* 50: 49–56.

Krøyer HN. 1844. Ichthologiske bidrag. 8. Caracanthus typicus. Naturhistorisk Tidsskrift (Kjøbenhavn) (New Series) 1: 264–268.

Krøyer HN. **1845**. Ichthologiske bidrag. *Naturhistorisk Tidsskrift* (Kjøbenhavn) (New Series) 1(3): 213–282.

Krupp F. 1987. Tiefenwasser- und Tiefseefische aus dem Roten Meer. XV. The occurrence of *Cynoglossus acutirostris* Norman 1939 in the Red Sea. *Senckenbergiana Maritima* 19(3/4): 249–259.

Krupp F, Almarri M, Zajonz U, Carpenter K, Almatar S, Zetzsche H. 2000. Twelve new records from the Gulf. *Fauna* of Arabia 18: 323–335.

Krupp F, Apel M, Hariri K. 2002. Conservation and sustainable use of biodiversity of Socotra Archipelago. Marine habitat, biodiversity and fisheries surveys and management. Final overall report including final statement of expenditures. Senckenberg Research Institute, Frankfurt a.M., Germany. 15 pp.

Krupp F, Paulus T. 1991. First record of the coral-reef fish *Pseudanthias fasciatus* (Kamohara, 1954) from the Red Sea (Perciformes: Serranidae). In: W. Büttiker W, Krupp F (eds) *Fauna of Saudi Arabia* 12: 388–392.

Krupp F, Zajonz U, Khalaf MA. 2009. A new species of the deepwater cardinalfish genus *Epigonus* (Perciformes: Epigonidae) from the Gulf of Aqaba, Red Sea. *aqua, International Journal of Ichthyology* 15(4): 223–227.

Kuhl H. 1829. In: Cuvier G. 1829–1830.

Kühlmorgen-Hille G. 1974. Gerreidae. In: Fischer W,
Whitehead PJP (eds) FAO species identification sheets for fishery purposes. Eastern Indian Ocean (Fishing Area 57) and western central Pacific (Fishing Area 71). Vol. 2. Bony fishes [Families Co–L]. Rome: FAO. [unpaginated]

Kuiter RH. **1990**. *Pseudanthias bimaculatus* x *Pseudanthias pleurotaenia*, a hybrid anthiid fish from Indonesia. *Revue française d'Aquariologie Herpétologie* 17(1990) 1: 17–18.

Kuiter RH. 1992. Tropical reef-fishes of the western Pacific Indonesia and adjacent waters. Jakarta, Indonesia: Gramedia Pustaka Utama. xiii + 314 pp.

Kuiter RH. 1993. Coastal fishes of south-eastern Australia.
Bathurst, New South Wales: Crawford House Press; and Honolulu: University of Hawaii Press. xxxi + 437 pp.

Kuiter RH. **1997**. *Guide to sea fishes of Australia*. A comprehensive reference for divers and fishermen. Frenchs Forest, New South Wales: New Holland Publishers. xvii + 434 pp.

Kuiter RH. 1998. Photo guide to fishes of the Maldives. Apollo Bay, Victoria: Atoll Editions. 257 pp. [Also published in 1998 by Windsor Books International and Sea Challengers Press]

Kuiter RH. 1998. Pipefishes of the syngnathid genus Dunckerocampus (Sygnathiformes: Syngnathidae), with a description of a new species from the Indian Ocean. aqua, Journal of Ichthyology and Aquatic Biology 3(2): 81–84.

Kuiter RH. **2000**. *Seahorses, pipefishes and their relatives: a comprehensive guide to Syngnathiformes.* Chorleywood, UK: TMC Publishing. 237 pp.

Kuiter RH. 2001. Revision of the Australian seahorses of the genus *Hippocampus* (Syngnathiformes: Syngnathidae) with descriptions of nine new species. *Records of the Australian Museum* 53: 293–340.

Kuiter RH. **2002**. *Butterflyfishes, bannerfishes and their relatives: a comprehensive guide to Chaetodontidae and Microcanthidae. The Marine Fish Families Series.* Chorleywood, UK: TMC Publishing. 208 pp.

Kuiter RH. **2003**. *Seahorses, pipefishes and their relatives: a comprehensive guide to Syngnathiformes* (2<sup>nd</sup> edition). Chorleywood, UK: TMC Publishing. 237 pp.

Kuiter RH. **2004**. *Basslets, hamlets and their relatives: a comprehensive guide to selected Serranidae and Plesiopidae*. Chorleywood, UK: TMC Publishing. 216 pp.

Kuiter RH. **2004**. Seahorses, seadragons, pipehorses, pipefishes & relatives: pictorial guide to Syngnathiformes. Victoria, Australia: Zoonetics. iv + 231 pp. Kuiter RH. **2004**. *Seahorses, pipefishes, and their relatives: a comprehensive guide to Syngnathiformes* (5<sup>th</sup> revision). Chorleywood, UK: TMC Publishing. 237 pp.

Kuiter RH. **2009**. *Seahorses and their relatives*. Seaford, Australia: Aquatic Photographics. 333 pp.

Kuiter RH, Debelius H. 1994. Southeast Asia tropical fish guide: Indonesia, Philippines, Vietnam, Malaysia, Singapore, Thailand, Andaman Sea. Frankfurt: IKAN-Unterwasserchiv. 321 pp.

Kuiter RH, Debelius H. 1997. Southeast Asia tropical fish guide: Indonesia, Philippines, Vietnam, Malaysia, Singapore, Thailand, Andaman Sea (2<sup>nd</sup> edition). Frankfurt: IKAN-Unterwasserchiv. 321 pp.

Kuiter RH, Debelius H. 1999. Descriptions of a new butterflyfish, *Chaetodon andamanensis*, from the eastern Indian Ocean (Pisces, Perciformes, Chaetodontidae). *Senckenbergiana Biologica* 79(2): 231–235.

Kuiter RH, Debelius H. **2001**. *Surgeonfishes, rabbitfishes and their relatives: A comprehensive guide to Acanthuroidei.* Chorleywood: TMC Publishing. 208 pp.

Kuiter RH, Debelius H. **2006**. *World atlas of marine fishes*. Frankfurt, Germany: Ikan-Unterwasserarchiv. 728 pp.

Kuiter RH, Kozawa T. 1999. Pictorial Guide. Fishes of the Indo-West Pacific: Apogonidae. (2<sup>nd</sup> edition). CDROM, Aquatic Photographics, Seaford, Australia. 130 pp.

Kuiter RH, Randall JE. **1981**. Three look-alike Indo-Pacific labrid fishes, *Halichoeres margaritaceus, H. nebulosus* and *H. miniatus. Revue française d'Aquariologie Herpétologie* 8(1): 13–18.

Kuiter RH, Randall JE. 1995. Four new Indo-Pacific wrasses (Perciformes: Labridae). *Revue française d'Aquariologie Herpétologie* 21(3/4) [for 1994]: 107–118.

Kuiter RH, Tonozuka T. **2001**. *Pictorial guide to Indonesian reef fishes. Part 1. Eels – Snappers, Muraenidae – Lutjanidae.* Zoonetics, Australia. 302 pp.

Kulbicki M, Beets J, Chabanet P, Cure K, Darling E, Floeter SR, Galzin R, Green A, Harmelin-Vivien M, Hixon M, Letourneur Y, Lison de Loma T, McClanahan T, McIlwain J, MouTham G, Myers R, O'Leary JK, Planes S, Vigliola L, Wantiez L. 2012. Distributions of Indo-Pacific lionfishes *Pterois* spp. in their native ranges: implications for the Atlantic invasion. *Marine Ecology Progress Series* 446: 189–205. http://dx.doi.org/10.3354/meps09442

Kulbicki M, Parravicini V, Bellwood DR, Arias-Gonzàlez E, Chabanet P, Floeter SR, Friedlander A, McPherson J, Myers RE, Vigliola L, Mouillot D. 2013. Global biogeography of reef fishes: a hierarchical quantitative delineation of regions. *PLoS ONE* 8(12): e81847. https://doi.org/10.1371/journal. pone.0081847

Kulkarni CV. **1940**. On the systematic position, structural modifications, bionomics and development of a remarkable new family of cyprinodont fishes from the province of

Bombay. *Records of the Indian Museum* (Calcutta) 42(2): 379–423.

- Kulkarni CV. **1948**. The osteology of Indian cyprinodonts. Part I. Comparative study of the head skeleton of *Aplocheilus*, *Oryzias*, and *Horaichthys*. *Proceedings of the National Institute of Sciences of India* 14(2): 65–119.
- Kulongowski C. **2010**. Revision of the ariid catfish genus *Galeichthys* Valenciennes (subfamily Galeichthyinae), with description of a new species from South Africa and designation of a neotype for *G. ater* Castelnau. *Smithiana* Bulletin 12: 19–23.

Kumar S, Tamura K, Nei M. 2004. MEGA3: integrated software for molecular evolutionary genetics analysis and sequence alignment. *Brief Bioinformatics* 5(2): 150–163.

Kumaran M, Randall JE. **1984**. Mullidae. Goatfishes. In:
Fischer W, Bianchi G (eds) FAO species identification sheets for fishery purposes. Western Indian Ocean (Fishing Area 51).
Vol. 3. Rome: FAO. [unpaginated]

Kume M, Yoshino T. 2008. Acanthopagrus chinshira, a new sparid fish (Perciformes: Sparidae) from the East Asia. Bulletin of the National Museum of Nature and Science (Tokyo) (Series A) Supplement No. 2: 47–57.

Kuo S-R, Shao K-T. 1999. Species composition of fish in the coastal zones of the Tsengwen Estuary, with descriptions of five new records from Taiwan. *Zoological Studies* 38(4): 391–404.

Kuraku S, Hoshiyama D, Katoh K, Suga H, Miyata T. 1999. Monophyly of lampreys and hagfishes supported by nuclear DNA-coded genes. *Journal of Molecular Evolution* 49(6): 729–735.

Kuraku S, Kuratani S. **2006**. Time scale for cyclostome evolution inferred with a phylogenetic diagnosis of hagfish and lamprey cDNA sequences. *Zoological Science* 23(12): 1053–1064.

Kurian CV. **1953**. A preliminary survey of the bottom fauna and bottom deposits of the Travancore Coast within the 15-fathom line. *Proceedings of the National Institute of Sciences of India* 19(6): 747–775.

Kuroda N. **1950**. A research on *Bembrops caudimacula* Steindachner. *Japanese Journal of Ichthyology* 1(1): 57–61.

Kuronuma K. **1940**. The heterosomate fishes collected in deep waters of Japan. I. *Bulletin of the Biogeographical Society of Japan* 10(3): 29–61.

Kuronuma K. 1941. Notes on rare fishes taken off the Pacific coast of Japan. Bulletin of the Biogeographical Society of Japan 11(8): 37–67.

Kuronuma K, Abe Y. **1972**. *Fishes of Kuwait*. Kuwait City: Kuwait Institute for Scientific Research. 123 pp.

Kuronuma K, Abe Y. **1986**. *Fishes of the Arabian Gulf*. Kuwait City: Kuwait Institute for Scientific Research. 357 pp.

Kurup BM, see also Madhusoodana Kurup B.

Kurup BM, Samuel CT. **1980**. On the little known fish *Hyporhamphus* (*Hyporhamphus*) *xanthopterus* (Valenciennes) from the Vembanad Lake (Kerala), with a key for identification of halfbeaks (Pisces: Hemiramphidae) of the Vembanad Lake. *Bulletin of the Department of Marine Sciences, University of Cochin* 11(2): 1–9.

Kurup BM, Samuel CT. **1983**. Systematics and distribution of fishes of the family Leiognathidae (Pisces) of the Vembanad Lake, Kerala (S. India). *Records of the Zoological Survey of lndia* 80(3&4): 387–411.

 Kurup BM, Samuel CT. 1985. A re-description of the little known rabbitfish *Siganus lineatus* (Cuvier and Valenciennes) (Pisces: Siganidae) with notes on siganid fishes of the Vembanad Lake. *Fishery Technology* 22: 62–65.

Kuwamura T. **1981**. Life history and population fluctuation in the labrid fish, *Labroides dimidiatus*, near the northern limit of its range. *Publications of the Seto Marine Biological Laboratory* 26: 95–117.

Kuwamura T. **1981**. Diurnal periodicity of spawning activity in free-spawning labrid fishes. *Japanese Journal of Ichthyology* 28: 343–348.

Kuwamura T. **1984**. Social structure of the protogynous fish *Labroides dimidiatus*. *Publications of the Seto Marine Biological Laboratory* **29**(1–3): 117–177.

Kyushin K, Amaoka K, Nakaya K, Ida H. **1977**. *Fishes of Indian Ocean*. Tokyo: Japan Marine Fishery Resource Research Center. 392 pp.

Kyushin K, Amaoka K, Nakaya K, Ida H, Tanino Y, Senta T (eds). 1982. Fishes of the South China Sea. Tokyo: Japan Marine Fishery Resource Research Center. 333 pp.

Labhart P. **1978**. Die Arten der Gattung *Oryzias* Jordan and Snyder, 1907. *Deutsche Killifish Gemeinschaft* 10: 53–58.

Laboute P, Grandperrin R. **2000**. *Poissons de Nouvelle-Calédonie*. Nouméa: Editions Catherine Ledru. 520 pp.

Lacepède BGE. **1798–1803**. *Histoire naturelle des poissons* (5 vols). Paris: Plassan.

Lachner EA. **1951**. Studies of certain apogonid fish from the Indo-Pacific, with descriptions of three new species. *Proceedings of the United States National Museum* 101(3290): 581–610.

Lachner EA. **1953**. Family Apogonidae: cardinal fishes (pp. 412–498). In: Schultz LP, Herald ES, Lachner EA, Welander AD, Woods LP (eds) *Fishes of the Marshall and Marianas Islands. Vol. 1. Families from Asymmetrontidae through Siganidae. Bulletin of the United States National Museum* No. 202: i–xxxii + 1–685, Pls. 1–74.

Lachner EA. **1954**. A revision of the goatfish genus *Upeneus* with descriptions of two new species. *Proceedings of the United States National Museum* 103(3330): 497–532.

Lachner EA. 1955. Populations of the berycoid fish family Polymixiidae. Proceedings of the United States National Museum 105(3356): 189–206.

Lachner EA. **1966**. Family Echeneidae: diskfishes (pp. 74–80). In: Schultz LP, Woods LP, Lachner EA (eds) *Fishes of the Marshall and Marianas Islands. Vol. 3. Families from Kraemeriidae through Antennariidae. Bulletin of the United States National Museum* No. 202: i–vii + 1–176, Pls. 124–148.

Lachner EA. **1973**. Echeneididae (pp. 637–640). In: Hureau J-C, Monod T (eds) *Check-list of the fishes of the northeastern Atlantic and of the Mediterranean (CLOFNAM)*. Vol. I. Paris: UNESCO.

Lachner EA. 1986. Echeneididae (pp. 1329–1334). In: Whitehead PJP, Bauchot M-L, Hureau J-C, Nielsen J, Tortonese E (eds) *Fishes of the north-eastern Atlantic and the Mediterranean*. Vol. III. Paris: UNESCO.

Lachner EA, Karnella SJ. **1978**. Fishes of the genus *Eviota* of the Red Sea with descriptions of three new species (Teleostei: Gobiidae). *Smithsonian Contributions to Zoology* 286: 1–23.

Lachner EA, Karnella SJ. **1980**. Fishes of the Indo-Pacific genus *Eviota* with descriptions of eight new species (Teleostei: Gobiidae). *Smithsonian Contributions to Zoology* 315: 1–127.

Lachner EA, McKinney JF. **1974**. *Barbuligobius boehlkei*, a new Indo-Pacific genus and species of Gobiidae (Pisces), with notes on the genera *Callogobius* and *Pipidonia*. *Copeia* 1974(4): 869–879.

Lachner EA, McKinney JF. **1978**. A revision of the Indo-Pacific fish genus *Gobiopsis* with descriptions of four new species (Pisces: Gobiidae). *Smithsonian Contributions to Zoology* 262: 1–52.

Lachner EA, McKinney JF. **1979**. Two new gobiid fishes of the genus *Gobiopsis* and a redescription of *Feia nympha* Smith. *Smithsonian Contributions to Zoology* 299: 1–18.

Lachner EA, McKinney JF. 1981. A new fish species of the genus Vanderhorstia (Teleostei: Gobiidae) from the Amirante Islands, Indian Ocean. Proceedings of the Biological Society of Washington 93(4): 963–970.

Laissius Y. **1973**. Note sur le voyages de Jean-Jacques Dussumiere (1782–1883). *Annales de la Societe des Sciences Naturelle de al Charente-Maritime* 5(5–9): 387–406.

Lal Mohan RS. **1967**. *Tentaculus waltariensis* Rao and Dutt, 1965, a junior synonym of *Pholioides thomaseni* Nielsen 1960 (Pisces: Haliophidae). *Copeia* 1967(2): 458–459.

Lal Mohan RS. **1968**. On the occurrence of the Blennioid fishes *Blennius semifasciatus* Rüppell (Family: Blenniidae) and *Tripterygion fasciatum* (Weber) (Family: Clinidae) along the Indian coast. *Journal of the Marine Biological Association of India* 10(1): 114–117. Lal Mohan RS. **1969**. On a collection of blennies from Gujarat coast with some new records. *Journal of the Marine Biological Association of India* 10(1): 118–125.

Lal Mohan RS. **1969**. On three new genera of sciaenid fishes (Pisces: Sciaenidae) from India. *Current Science* 38(12): 295–296.

Lal Mohan RS. **1970**. *Callionymus jonesii*, a new callionymid fish (Pisces: Callionymidae) from the east coast of India. *Journal of the Marine Biological Association of India* 10(2) [for 1968]: 357–360.

Lal Mohan RS. **1971**. A new species of sciaenid fish *Johnius mannarensis* from the south-east coast of India. *Journal of the Marine Biological Association of India* 11(1–2): 320–323.

Lal Mohan RS. **1971**. *Helcogramma shinglensis*, a new species of tripterygid fish from Gulf of Mannar with a key to the fishes of the family Tripterygidae of eastern and central Indian Ocean. *Senckenbergiana Biologica* 52(3/5): 219–223.

Lal Mohan RS. **1972**. A synopsis of the Indian genera of the fishes of the family Sciaenidae. *Indian Journal of Fisheries* 16(1–2): 82–98.

Lal Mohan RS. **1972**. *Otolithoides brunneus* (Day) 1873, as a junior synonym of *Otolithoides biauritus* (Cantor) 1850 (Pisces: Sciaenidae), with notes on the identity of *O. brunneus* Dutt and Thankam (1968). *Journal of the Marine Biological Association of India* 14(1): 415–417.

Lal Mohan RS. **1976**. Two new species of sciaenid fishes *Johnius elongatus* and *Johnieops macrorhynus* from India. *Matsya* 1: 19–25.

Lal Mohan RS. **1981**. An illustrated synopsis of the fishes of the family Sciaenidae of India. *Indian Journal of Fisheries* 28(1–2): 1–24.

Lal Mohan RS. **1984**. Sciaenidae. Croakers, drums, meagres, weakfishes. In: Fischer W, Bianchi G (eds) *FAO species identification sheets for fishery purposes. Western Indian Ocean (Fishing Area 51)*. Vol. 4. Rome: FAO. [unpaginated]

Lambers PH. **1992**. On the ichthyofauna of the Solnhofen lithographic limestone (Upper Jurassic, Germany). PhD thesis, Rijksuniversiteit Groningen. 336 pp.

Larson HK. **1985**. A revision of the commensal gobiid genus *Bryaninops* (Pisces), with a description of six new species. *The Beagle, Occasional Papers of the Northern Territory Museum of Arts and Sciences* 2(1): 57–93.

Larson HK. 1986. Phyllogobius, a new generic name for Cottogobius platycephalops Smith 1964 (Pisces: Gobiidae), and a redescription of the species. The Beagle, Occasional Papers of the Northern Territory Museum of Arts and Sciences 3(1): 131–136.

Larson HK. **1990**. A revision of the commensal gobiid fish genera *Pleurosicya* and *Luposicya* (Gobiidae), with descriptions of eight new species of *Pleurosicya* and discussion of related genera. *The Beagle, Records of the Northern Territory Museum of Arts and Sciences* 7(1): 1–53. Larson HK. **2001**. A revision of the gobiid fish genus *Mugilogobius* (Teleostei: Gobioidei), and its systematic placement. *Records of the Western Australian Museum* Supplement No. 62: 1–233.

Larson HK. **2009**. Review of the gobiid fish genera *Eugnathogobius* and *Pseudogobiopsis* (Gobioidei: Gobiidae: Gobionellinae), with descriptions of three new species. *Raffles Bulletin of Zoology* 57(1): 127–181.

Larson HK. **2010**. A review of the gobiid fish genus *Redigobius* (Teleostei: Gobionellinae), with descriptions of two new species. *Ichthyological Exploration of Freshwaters* 21(2): 123–191.

Larson HK, Buckle D. **2012**. A revision of the goby genus *Gnatholepis* Bleeker (Teleostei, Gobiidae, Gobionellinae), with description of a new species. *Zootaxa* 3529: 1–69. http://dx.doi.org/10.11646/zootaxa.3529.1.1

Larson HK, Hoese DF. **1980**. Fishes of the Indian Ocean. Results of the ichthyological investigations during the Indian Ocean expedition of the research vessel *Meteor*, October 1964 to May 1965. Systematical Section, XXIII. Gobiidae. *Meteor Forschungsergebnisse* 32: 33–43. www.schweizerbart.de/publications/list/series/meteor

Larson HK, Murdy EO. **2001**. Eleotridae. Sleepers (gudgeons) (pp. 3574–3577); Gobiidae. Gobies (pp. 3578–3603). In: Carpenter KE, Niem VH (eds) *FAO species identification* guide for fishery purposes. The living marine resources of the western central Pacific. Vol. 6. Bony fishes part 4 (pp. 3381– 4218). Rome: FAO.

Larson HK, Pezold FL. **2016**. The correct name for *Oxyurichthys longicauda* Steindachner and a few other errors. *Zootaxa* 4066(2): 171–172. https://doi.org/10.11646/ zootaxa.4066.2.5

Larson HK, Wright J. **2003**. A new genus for the Indo-Pacific goby species *Gobius baliurus* Valenciennes and (Teleostei, Gobiidae, Gobiinae). *The Beagle, Records of the Museums and Art Galleries of the Northern Territory* 19: 127–135.

Last PR. 2001. Amarsipidae. Amarsipas (pp. 3765–3766); Nomeidae. Driftfishes (cigarfishes) (pp. 3771–3775); Ariommatidae. Ariommas (pp. 3780–3783); Tetragonuridae. Squaretails (pp. 3784–3785); Stromateidae. Butterfishes (silver pomfrets) (pp. 3786–3791). In: Carpenter KE, Niem VH (eds) FAO species identification guide for fishery purposes. The living marine resources of the western central Pacific. Vol. 6. Bony fishes part 4 (pp. 3381–4218). Rome: FAO.

Last PR. 2008. New short-snout members of the skate genus Dipturus (Rajoidei: Rajidae) from Australian seas (pp. 53– 98). In: Last PR, White WT, Pogonoski JJ, Gledhill DC (eds) Descriptions of new Australian skates (Batoidea: Rajoidei). Hobart, Australia: CSIRO Marine and Atmospheric Research. 187 pp. Last PR, Bogorodsky SV, Alpermann TJ. **2016**. *Maculabatis ambigua* sp. nov., a new whipray (Myliobatiformes: Dasyatidae) from the western Indian Ocean. *Zootaxa* 4154(1): 66–78. http://dx.doi.org/10.11646/zootaxa.4154.1.4

Last PR, Compagno LJV. **1999**. Rajidae. Hardnose skates (pp. 1452–1456). In: Carpenter KE, Niem VH (eds) *FAO species identification guide for fishery purposes. The living marine resources of the western central Pacific*. Vol. 3. Batoid fishes, chimaeras and bony fishes part 1 (pp. 1397–2068). Rome: FAO.

Last PR, Compagno LJV, Nakaya K. **2004**. *Rhinobatos nudidorsalis*, a new species of shovelnose ray (Batoidea: Rhinobatidae) from the Mascarene Ridge, central Indian Ocean. *Ichthyological Research* 51(2): 153–158.

Last PR, Gledhill DC. 2008. A new skate of the genus *Dentiraja* (Rajoidei: Rajidae) from southern Australia (pp. 109–118).
In: Last PR, White WT, Pogonoski JJ, Gledhill DC (eds) *Descriptions of new Australian skates (Batoidea: Rajoidei)*.
Hobart, Australia: CSIRO Marine and Atmospheric Research. 187 pp.

Last PR, Gledhill DC. **2008**. Two new skates of the genus *Okamejei* (Rajoidei: Rajidae) from the south-east Indian Ocean (pp. 119–134). In: Last PR, White WT, Pogonoski JJ, Gledhill DC (eds) *Descriptions of new Australian skates (Batoidea: Rajoidei)*. Hobart, Australia: CSIRO Marine and Atmospheric Research. 187 pp.

Last PR, Henderson AC, Naylor GJP. **2016**. *Acroteriobatus omanensis* (Batoidea: Rhinobatidae), a new guitarfish from the Gulf of Oman. *Zootaxa* 4144(2): 276–286. http://dx.doi.org/10.11646/zootaxa.4144.2.9

Last PR, Manjaji-Matsumoto BM. **2008**. *Himantura dalyensis* sp. nov., a new estuarine whipray (Myliobatoidei: Dasyatidae) from northern Australia (pp. 283–291). In: Last PR, White WT, Pogonoski JJ (eds) *Descriptions of new Australian chondrichthyans*. Hobart, Australia: CSIRO Marine and Atmospheric Research. 358 pp.

Last PR, Manjaji-Matsumoto BM, Moore ABM. **2012**. *Himantura randalli* sp. nov., a new whipray (Myliobatoidea: Dasyatidae) from the Persian Gulf. *Zootaxa* 3327: 20–32. http://dx.doi.org/10.11646/zootaxa.3327.1.2

Last PR, Manjaji-Matsumoto BM, Naylor GJP, White WT. 2016. Stingrays. Family Dasyatidae (pp. 522–618). In: Last PR, White WT, Carvalho MR de, Séret B, Stehmann MFW, Naylor GJP (eds) *Rays of the world*. CSIRO Publishing & Cornell University Press. 790 pp.

Last PR, Moteki M. **2001**. Bramidae. Pomfrets (pp. 2824–2836). In: Carpenter KE, Niem VH (eds) *FAO species identification guide for fishery purposes. The living marine resources of the western central Pacific.* Vol. 5. Bony fishes part 3 (pp. 2791–3380). Rome: FAO.

Last PR, Motomura H, White WT. 2008. Cephaloscyllium albipinnum sp. nov., a new swellshark (Carcharhiniformes: Scyliorhinidae) from southeastern Australia (pp. 147–157).
In: Last PR, White WT, Pogonoski JJ (eds) Descriptions of new Australian chondrichthyans. Hobart, Australia: CSIRO Marine and Atmospheric Research. 358 pp.

Last PR, Naylor GJP, Manjaji-Matsumoto BM. **2016**. A revised classification of the family Dasyatidae (Chondrichthyes: Myliobatiformes) based on new morphological and molecular insights. *Zootaxa* 4139(3): 345–368. http://dx.doi.org/10.11646/zootaxa.4139.3.2

Last PR, Séret B, Naylor GJP. **2016**. A new species of guitarfish, *Rhinobatos borneensis* sp. nov. with a redefinition of the family-level classification in the order Rhinopristiformes (Chondrichthyes: Batoidea). *Zootaxa* 4117(4): 451–475. https://doi.org/10.11646/zootaxa.4117.4.1

Last PR, Séret B, Stehmann MFW, Weigmann S. **2016**. Skates. Family Rajidae (pp. 204–363). In: Last PR, White WT, Carvalho MR de, Séret B, Stehmann MFW, Naylor GJP (eds) *Rays of the world*. CSIRO Publishing & Cornell University Press. 790 pp.

Last PR, Séret B, White WT. **2008**. New swellsharks (*Cephaloscyllium*: Scyliorhinidae) from the Indo–Australian region (pp. 129–146). In: Last PR, White WT, Pogonoski JJ (eds) *Descriptions of new Australian chondrichthyans*. Hobart, Australia: CSIRO Marine and Atmospheric Research. 358 pp.

Last PR, Stehmann MFW. **2008**. *Rajella challengeri* sp. nov., a new deepwater skate from southern Australia (pp. 135–144). In: Last PR, White WT, Pogonoski JJ, Gledhill DC (eds) *Descriptions of new Australian skates (Batoidea: Rajoidei)*. Hobart, Australia: CSIRO Marine and Atmospheric Research. 187 pp.

Last PR, Stehmann MFW, Séret B. **2008**. *Leucoraja pristispina* sp. nov., a new deepwater skate from Western Australia (pp. 145–154). In: Last PR, White WT, Pogonoski JJ, Gledhill DC (eds) *Descriptions of new Australian skates* (*Batoidea: Rajoidei*). Hobart, Australia: CSIRO Marine and Atmospheric Research. 187pp.

Last PR, Stevens JD. **1994**. *Sharks and rays of Australia* (1st edition). Hobart, Australia: Australian Fisheries Research & Development Corporation, CSIRO. vii + 513 pp., Pls. 1–84.

Last PR, Stevens JD. 2008. Bythaelurus incanus sp. nov., a new deepwater catshark (Carcharhiniformes: Scyliorhinidae) from northwestern Australia (pp. 123–127). In: Last PR, White WT, Pogonoski JJ (eds) Descriptions of new Australian chondrichthyans. Hobart, Australia: CSIRO Marine and Atmospheric Research. 358 pp.

Last PR, Stevens JD. **2009**. *Sharks and rays of Australia* (2nd edition). Collingwood, Australia: CSIRO Publishing. 644 pp.

Last PR, Weigmann S, Dumale D. **2016**. A new skate genus *Orbiraja* (Rajiformes: Rajidae) from the Indo-West Pacific. *Zootaxa* 4184 (1): 52–62. https://doi.org/10.11646/ zootaxa.4184.1.3

Last PR, White WT. **2008**. Two new saddled swellsharks (*Cephaloscyllium*: Scyliorhinidae) from eastern Australia (pp. 159–170). In: Last PR, White WT, Pogonoski JJ (eds) *Descriptions of new Australian chondrichthyans*. Hobart, Australia: CSIRO Marine and Atmospheric Research. 358 pp.

Last PR, White WT, Caira JN, Dharmadi, Fahmi, Jensen K, Lim APK, Manjaji-Matsumoto BM, Naylor GJP, Pogonoski JJ, Stevens JD, Yearsley GK. 2010. Sharks and rays of Borneo. Australia: CSIRO Marine and Atmospheric Research. 304 pp.

Last PR, White WT, Carvalho MR de, Séret B, Stehmann MFW, Naylor GJP (eds). **2016**. *Rays of the world*. Ithaca: Cornell University Press & Melbourne: CSIRO Publishing. 790 pp.

Last PR, White WT, Naylor GJP. **2016**. Sawfishes. Family Pristidae (pp. 58–64). In: Last PR, White WT, Carvalho MR de, Séret B, Stehmann MFW, Naylor GJP (eds.) *Rays of the world*. CSIRO Publishing & Cornell University Press. 790 pp.

Last PR, White WT, Naylor G. **2016**. Three new stingrays (Myliobatiformes: Dasyatidae) from the Indo-West Pacific. *Zootaxa* 4147(4): 377–402. http://dx.doi.org/10.11646/ zootaxa.4147.4.2

Last PR, White WT, Pogonoski JJ (eds). 2008. *Descriptions* of new Australian chondrichthyans. Research Paper No. 022. Hobart, Australia: CSIRO Marine and Atmospheric Research. 358 pp.

Last PR, White WT, Pogonoski JJ (eds). **2010**. *Descriptions of new sharks and rays from Borneo*. Research Paper No. 032. Hobart, Australia: CSIRO Marine and Atmospheric Research. 165 pp.

Last PR, White WT, Pogonoski JJ, Gledhill DC (eds). 2008.
Descriptions of new Australian skates (Batoidea: Rajoidei).
Research Paper No. 021. Hobart, Australia: CSIRO Marine and Atmospheric Research. 187 pp.

Last PR, White WT, Serét B. **2016**. Taxonomic status of maskrays of the *Neotrygon kuhlii* speces complex (Myliobatoidae: Dasyatidae) with the description of three new species from the Indo-West Pacific. *Zootaxa* 4083(4): 533–561. https://doi.org/10.11646/zootaxa.4083.4.5

Latham JF. **1794**. An essay on the various species of sawfish. *Transactions of the Linnean Society of London* 2(art.25): 273–282.

 Latreille PA. 1804. Tableau méthodique des poissons. In: Nouveaux dictionnaire d'histoire naturelle. 1<sup>re</sup> Edition. Vol. 24. Paris. pp. 71–105. Lau PPF, Li LWH. **2000**. *Identification guide to fishes in the live seafood trade of the Asia-Pacific region*. WWF Hong Kong and Agriculture, Fisheries and Conservation Department. Hong Kong. 137pp.

Lauder GV, Liem KF. **1983**. The evolution and interrelationships of the actinopterygian fishes. *Bulletin of the Museum of Comparative Zoology* 150(3): 95–197.

Lautredou A-C, Motomura H, Gallut C, Ozouf-Costaz C, Cruaud C, Lecointre G, Dettai A. **2013**. New nuclear markers and exploration of the relationships among Serraniformes (Acanthomorpha, Teleostei): the importance of working at multiple scales. *Molecular Phylogenetics and Evolution* 67(1): 140–155. http://dx.doi.org/10.1016/j. ympev.2012.12.020

Lavergne E, Calvès I, Meistertzheim A, Charrier G, Zajonz U, Laroche J. **2014**. Complex genetic structure of a euryhaline marine fish in temporarily open/closed estuaries from the wider Gulf of Aden. *Marine Biology* 161(5): 1113–1126. http://dx.doi.org/10.1007/s00227-014-2404-z

Lavergne E, Zajonz U, Sellin L. 2013. Length-weight relationship and seasonal effects of the Summer Monsoon on condition factor of *Terapon jarbua* (Forsskål 1775) from the wider Gulf of Aden including Socotra Island. *Journal of Applied Ichthyology* (29): 274–277. [online in 2012] https://doi.org/10.1111/j.1439-0426.2012.02018.x

Lavergne E, Zajonz U, Krupp F, Naseeb F, Aideed MS. **2016**. Diversity and composition of estuarine and lagoonal fish assemblages of Socotra Island, Yemen. *Journal of Fish Biology* 88(5): 2004–2026. http://dx.doi.org/10.1111/ jfb.12964

Lavoué S. **2016**. Was Gondwanan breakup the cause of the intercontinental distribution of Osteoglossiformes? A time-calibrated phylogenetic test combining molecular, morphological, and paleontological evidence. *Molecular Phylogenetics and Evolution* 99: 34–43. http://dx.doi. org/10.1016/j.ympev.2016.03.008

Lavoué S, Miya M, Saitoh K, Ishiguro NB, Nishida M. **2007**. Phylogenetic relationships among anchovies, sardines, herrings and their relatives (Clupeiformes), inferred from whole mitogenome sequences. *Molecular Phylogenetics and Evolution* 43: 1096–1105.

Lavoué S, Sullivan JP. **2004**. Simultaneous analysis of five molecular markers provides a well-supported phylogenetic hypothesis for the living bony-tongue fishes (Osteoglossomorpha: Teleostei). *Molecular Phylogenetics and Evolution* 33(1): 171–185.

Lay GT, Bennett ET. 1839. Fishes (pp. 41–75). In: Beechey FW (ed) *The zoology of Captain Beechey's voyage*. London: HG Bohn. 180 pp.

Leach WE. **1814**. *The zoological miscellany; being descriptions of new, or interesting animals*. Vol. 1. London: E. Nodder & Son. 144 pp.

Le Danois Y. **1959**. Étude ostéologique, myologique et systématique des poissons du sous-ordre des orbiculates. *Annales de l'Institut Océanographique* (Monaco) (N.S.) 36(1): 1–273.

Le Danois Y. **1964**. Étude anatomique et systématique des Antennaires, de l'Ordre des Pédiculates. *Mémoires du Muséum National d'Histoire Naturelle* (Paris) (N.S., Série A, Zoologie) 31(1): 1–162.

Le Danois Y. **1970**. Étude sur des poissons pédiculates de la famille des Antennariidae récoltés dans la Mer Rouge et description d'une espèce nouvelle. *Israel Journal of Zoology* 19(2): 83–94.

Le Danois Y. **1975**. Étude osteo-mylogique et révision systématique de la famille des Lophiidae (Pédiculates Haploptérygiens). *Mémoires du Muséum National d'Histoire Naturelle* (Paris) (N.S. Série A, Zoologie) 91: iii + 127 pp.

Le Danois Y. **1979**. Révision systématique de la famille des Chaunacidae (Pisces: Pediculati). *Uo* (Japanese Society of Ichthyologists) 30: 1–76.

Lee M-Y, Chen H-M, Shao K-T. 2009. A new species of deepwater tonguefish genus *Symphurus* (Pleuronectiformes: Cynoglossidae) from Taiwan. *Copeia* 2009(2): 342–347. http://dx.doi.org/10.1643/CI-08-080

Lee M-Y, Munroe TA, Chen H-M. 2009. A new species of tonguefish (Pleuronectiformes: Cynoglossidae) from Taiwanese waters. *Zootaxa* 2203: 49–58. http://dx.doi. org/10.11646/zootaxa.2203.1.4

Lee M-Y, Munroe TA, Kai Y. **2016**. Description of a new cryptic species of tonguefish (Pleuronectiformes: Cynoglossidae: *Symphurus*) from shallow waters off Japan. *Ichthyological Research* 64(1) [2017]: 71–83. http://dx.doi. org/10.1007/s10228-016-0541-8

Lee M-Y, Munroe TA, Shao K-T. **2014**. Description of a new cryptic, shallow-water tonguefish (Pleuronectiformes: Cynoglossidae: *Symphurus*) from the western North Pacific Ocean. *Journal of Fish Biology* 85(3): 563–585. https://doi.org/10.1111/jfb.12440

Lee S-C. **1983**. The family Syngnathidae (Pisces: Syngnathiformes) of Taiwan. *Bulletin of the Institute of Zoology, Academia Sinica* 22(1): 67–82.

Legaspi VA. **1956**. A contribution to the life history of the nomeid fish *Psenes cyanophrys* Cuvier and Valenciennes. *Bulletin of Marine Science of the Gulf and Caribbean* 6(3): 179–199.

Leis JM. **1978**. Systematics and zoogeography of the porcupinefishes (*Diodon*, Diodontidae, Tetraodontiformes), with comments on egg and larval development. *Fishery Bulletin* 76(3): 535–567.

Leis JM. **2006**. Nomenclature and distribution of the species of the porcupinefish family Diodontidae (Pisces, Teleostei). *Memoirs of the Museum of Victoria* 63(1): 77–90.

Leis JM. **2008**. Larval development in the lutjanid subfamily Lutjaninae (Pisces): the genus *Pinjalo*. *Zootaxa* 1760: 37–49. https://doi.org/10.5281/zenodo.181919

Leis JM. **2015**. Is dispersal of larval reef fishes passive? (pp. 223–226). In: Mora C (ed) *Ecology of fishes on coral reefs*. Cambridge: Cambridge University Press. xiv + 374 pp. http://dx.doi.org/10.1017/CBO9781316105412.025

Leis JM, Carson-Ewart BM. **2000**. *The larvae of Indo-Pacific coastal fishes: An identification guide to marine fish larvae*. Leiden: Brill. 850 pp.

Leis JM, Hoese DF, Trnski T. **1993**. Larval development in two genera of the Indo-Pacific gobioid fish family Xenisthmidae: *Allomicrodesmus* and *Xenisthmus*. *Copeia* 1993(1): 186–196.

Leis JM, Moyer JT. **1985**. Development of eggs, larvae and pelagic juveniles of three Indo-Pacific ostraciid fishes (Tetraodontiformes): *Ostracion meleagris, Lactoria fornasini* and *L. diaphana. Japanese Journal of Ichthyology* 32(2): 189–202.

Leis JM, Randall JE. **1982**. *Chilomycterus spilostylus*, a new species of Indo-Pacific burrfish (Pisces, Tetraodontiformes, Diodontidae). *Records of the Australian Museum* 42(3): 363–371.

Leis JM, Rennis DS. **2000**. Cirrhitidae (hawkfishes) (pp. 403– 407); Pinguipedidae (grubfishes, sandperches) (pp. 565–568). In: Leis JM, Carson-Ewart BM (eds) *The larvae of Indo-Pacific coastal fishes: an identification guide to marine fish larvae*. Leiden: Brill. 850 pp.

Leis JM, Siebeck UE, Hay AC, Paris CB, Chateau O, Wantiez L. 2015. In situ orientation of fish larvae can vary among regions. Marine Ecology Progress Series 537: 191–203. http://dx.doi.org/10.3354/meps11446

Leis JM, Trnski T. **1989**. *The larvae of Indo-Pacific shorefishes*. Honolulu: University of Hawaii Press. 371 pp.

Leis JM, Trnski T, Beckley LE. **2002**. Larval development of *Pagellus natalensis*, and what larval morphology indicates about relationships in the perciform fish family Sparidae (Teleostei). *Marine and Freshwater Research* 53(2): 367–376.

Leis JM, Van der Lingen C. **1997**. Larval development and relationships of the perciform Family Dichistiidae (=Coracinidae), the galjoen fishes. *Bulletin of Marine Science* 60(1): 100–116.

Lema T de, Lucena CAS de, Saenger S, Oliveira MFT de.
1979. Primeiro levantamento dos Tetraodontiformes do extremo sul do Brasil, Uruguai e Argentina (Teleostei: Acanthopterygii). *Comunicações do Museu de Ciências da PUCRGS* 20: 1–84.

Lenz H. **1881**. Ein neuer Fisch und zwei neue Myriapoden von Nossi-Bé. *Zoologischer Anzeiger* 4(93): 506–508.

Leslie RW, Grant WS. **1991**. Redescription of the southern African anglerfish *Lophius vomerinus* Valenciennes, 1837 (Lophiiformes: Lophiidae). *Copeia* 1991(3): 787–800. Lesson RP. **1828**. Description du nouveau genre Ichthyophis et de plusieurs espèces inédites ou peu connues de poissons, recueillis dans le voyage autour du monde de la Corvette *La Coquille. Mémoires de la Société d'Histoire Naturelle, Paris* (Série 2) 4: 397–412.

Lesson RP. **1829–1831**. Chapitre X. Poissons. In: Duperrey LI (ed) *Voyage autour du monde, exécuté par ordre du Roi, sur la corvette de La Majesté* La Coquille, *pendant les années 1822*, *1823*, *1824 et 1825.... Zoologie*. Vol. 2 (Part 1). Paris: Arthur Bertrand. pp. 66–238; Atlas, Pls. 1–38.

Lesueur CA. **1814**. Note sur deux poissons non encore décrits du genre *Callionyme* et de l'ordre des Jugulaires. *Bulletin des Sciences, par la Société Philomathique de Paris* (Série 3) 1 [for 1812]: pp. 5–6, Pls. 2.

Lesueur CA. **1818**. Description of several new species of North American fishes. *Journal of the Academy of Natural Sciences of Philadelphia* 1(2): 222–235; 359–368.

Lesueur CA. **1821**. Observations on several genera and species of fish, belonging to the natural family of the Esoces. *Journal of the Academy of Natural Sciences of Philadelphia* 2(1): 124–138.

Lesueur CA. **1822**. Description of a *Squalus*, of a very large size, which was taken on the coast of New-Jersey. *Journal of the Academy of Natural Sciences of Philadelphia* 2(2): 343–352.

Lesueur CA. **1825**. Description of a new fish of the genus *Salmo*. *Journal of the Academy of Natural Sciences of Philadelphia* 5(1): 48–51.

Letourneur Y. **1991**. Modifications du peuplement de poissons du platier récifal de Saint-Pierre (Île de la Réunion, Océan Indien) consécutives au passage du cyclone Firinga. *Cybium* 15(2): 159–170.

Letourneur Y. **1992**. Dynamique des peuplements ichtyologiques des platiers récifaux de l'île de la Réunion. Doctoral thesis, Univesité Aix-Marseille, Marseille. 244 pp.

Letourneur Y. **1998**. Length-weight relationship of some marine fish species inf Réunion Island, Indian Ocean. *Naga*, *The ICLARM Quarterly* 21(4): 37–39.

Letourneur Y. **1998**. Composition, structures et réseaux trophiques des peuplements de poissons de la côte au vent de l'île de La Réunion. *Cybium* 22(3): 267–283.

Letourneur Y, Chabanet P. **1994**. Variations spatiotemporelles de l'ichtyofaune des platiers récifaux à la Réunion. *Cybium* 18: 25–38.

Letourneur Y, Chabanet P, Durville P, Taquet M, Teissier E, Parmentier M, Quéro J-C, Pothin K. **2004**. An updated checklist of the marine fish fauna of Reunion Island, southwestern Indian Ocean. *Cybium* 28(3): 199–216.

Letourneur Y, Harmelin-Vivien M-L, Galzin R. **1993**. Impact of hurricane Firinga on fish community structure on fringing reefs of Reunion Island, S.W. Indian Ocean. *Envionmental Biology of Fishes* 37(2): 109–120. Leu M-Y, Fang L-S, Mok H-K. **1999**. First records of the schindleriid fishes, *Schindleria pietschmanni* and *S. praematura*, from Taiwan. *Acta Zoologica Taiwanica* 10(1): 72. [Abstract]

Leu M-Y, Fang L-S, Mok H-K. **2008**. First record of *Schindleria pietschmanni* (Schindler, 1931) (Actinopterygii: Schindleriidae) from Taiwan. *Platax* 5: 15–21.

Li C, Orti G. **2007**. Molecular phylogeny of Clupeiformes (Actinopterygii) inferred from nuclear and mitochondrial DNA. *Molecular Phylogenetics and Evolution* 44(1): 386–398.

Li G-Q, Wilson MVH. **1996**. Phylogeny of Osteoglossomorpha (pp. 163–174). In: Stiassny MLJ, Parenti LR, Johnson GD (eds) *Interrelationships of fishes*. San Diego: Academic Press. 496 pp.

Lichtenstein MHC. **1823**. Verzeichniss der Doubletten des zoologischen Museums der Königlichen Universität zu Berlin, nebst Beschreibung vieler bisher unbekannter Arten von Säugethieren, Vögeln, Amphibien und Fischen, herausgegeben von Dr H. Lichtenstein. Universität zu Belin, Berlin: T. Trautwin. 118 pp.

Liénard E. **1836**. Sur quelques poissons de l'île Maurice. *L'Institut* 4(167, 175): 240, 305.

Liénard E. 1840. Description d'une nouvelle espèce du genre mole (*Orthagoriscus*, Schn.) découverte à l'île Maurice. *Revue Zoologique par la Société Cuvierienne* (Paris) 3: 291–292.

Liénard F. **1839**. Poissons. In: Bouton L (ed) (*Dixième*) Rapport Annuel sur les travaux de la Société d'Histoire Naturelle de l'île Maurice 10: 31–47.

Liénard F. 1842. Poissons. In: Bouton L (ed) Rapport Annuel sur les travaux de la Société d'Histoire Naturelle de l'Ile Maurice 13 [for 1842–1843]: 57–95.

Lieske E, Myers R. **1994**. *Coral reef fishes: Caribbean, Indian Ocean and Pacific Ocean including the Red Sea*. Collins Pocket Guide. London: HarperCollins. 400 pp.

Lieske E, Myers R. 2002. Coral reef fishes: Caribbean, Indian Ocean and Pacific Ocean including the Red Sea (revised edition). Princeton, New Jersey: Princeton University Press. 400 pp.

Lieske E, Myers R. **2004**. *Collins coral reef guide: Red Sea to Gulf of Aden, South Oman.* London: HarperCollins. 384 pp.

Lin Y-J, Qurban MA, Shen KN, Chao NL. 2019. Delimitation of tigertooth croaker *Otolithes* species (Teleostei: Sciaenidae) from the western Arabian Gulf using integrative approach, with a description of *Otolithes arabicas* sp. nov. *Zoological Studies* 58: 10. https://doi.org/10.6620/ ZS.2019.58-10

Linck HG. **1790**. Versuch einter Eintheilung der Fische nach den Zähnen. *Magazin für das Neueste aus der Physik und Naturgeschichte* (Gotha) 6(3, art.3): 28–38.

Lindberg GU, Krasyukova ZV. **1969**. Fishes of the Sea of Japan and of adjacent areas of the Sea of Okhotsk and the Yellow Sea, Part 3: Teleostomi. XXIX. Perciformes. Percoidei. (XC. Serranidae – CXLIV. Champsodontidae). Zoology Institute and Academy of Science, USSR. 479 pp. [In Russian]

Lindberg GU, Krasyukova ZV. 1975. Fishes of the Sea of Japan and adjacent territories of the Okhotsk and Yellow Sea, Part 4. Teleostomi. XXIX. Perciformes. 2. Blennioidei – 13. Gobioidei. (CXLV. Fam. Anarhichadidae – CLXXV. Fam. Periophthalmidae). Izdateljestvo Nauka, Leningradskoie Otdeleie, Leningrad. 463 pp. [In Russian. English translation in 1989 – see next entry]

Lindberg GU, Krasyukova ZV. **1989**. *Fishes of the Sea of Japan and the adjacent areas of the Sea of Okhotsk and the Yellow Sea, Part 4*. Leningrad: Nauka. 602 pp. [Original in Russian dated 1975]

Lindberg GU, Legeza MI. **1959**. Fishes of the Japanese Sea and the adjacent parts of the Okhotsk and Yellow Seas, Part 1. Amphioxi Petromyzones Myxini Elasmobranchii Holocephali. Leningrad: Nauka. 208 pp. [In Russian]

Lindberg GU, Legeza MI. 1965. Fishes of the Japanese Sea and the adjacent parts of the Okhotsk and Yellow Seas, Part 2. Teleostomi, XII. Acipenseriformes, XXVIII. Polynemiformes. 391 pp. [in Russian, English translation 1969, Israel Prog. Sci. Transl.]

Linnaeus C. **1751**. *Philosophia Botanica*. Stockholm & Amsterdam.

Linnaeus C. **1758**. *Systema naturae per regna tria naturae*, *secundum classes, ordines, genera, species, cum characteribus, differentiis, synonymis, locis* (10<sup>th</sup> edition). Vol. 1. Holmiae: Laurentii Salvii. 824 pp.

Linnaeus C. 1764. Museum S. R. M. Adolphi Friderici Regis Suecorum, Gothorum, Vandalorumque, in quo animalia rariora imprimis et exotica... Aves, Amphibia, Pisces (Pisces: pp. 49–111, Pls. 1–32). Holmiae. Tomus Secundi Prodromus. 133 pp.

Linnaeus C. 1766. Systema naturae per regna tria naturae secundum, classes, ordines, genera, species, cum characteribus, differentiis synonymis, locis (12<sup>th</sup> edition). Vol. 1. Holmiae: Laurentii Salvii. 532 pp.

Linnaeus C. **1771**. *Mantissa Plantarum altera. Regnum Animalia*. Appendix. Holmiae: Laurentii Salvii. 584 pp.

Lipkin Y, Silva PC. **2002**. Marine algae and seagrasses of the Dahlak Archipelago, southern Red Sea. *Nova Hedwigia* 75: 1–90.

Liston JJ. **2008**. A review of the characters of the edentulous pachycormiforms *Leedsichthys*, *Asthenocormus* and *Martillichthys* nov. gen. (pp. 181–198). In: Arratia G, Schultze H-P, Wilson MVH (eds) *Mesozoic Fishes 4 – Homology and phylogeny. Proceedings of the international meeting, Miraflores de la Sierra, 2005*. Munich: Verlag Dr. Friedrich Pfeil. 502 pp. Liu C-H, Shen S-C. **1991**. A revision of the mugilid fishes from Taiwan. *Bulletin of the Institute of Zoology Academia Sinica* (Taipei) 30(4): 273–288.

Liu K-M, Lee M-L, Joung S-J, Chang Y-C. **2009**. Age and growth estimates of the sharptail mola, *Masturus lanceolatus*, in waters of eastern Taiwan. *Fisheries Research* 95(2–3): 154–160. http://dx.doi.org/10.1016/j. fishres.2008.08.013

Liu M, Sadovy de Mitcheson Y. 2009. Gonad development during sexual differentiation in hatchery-produced orangespotted grouper (*Epinephelus coioides*) and humpback grouper (*Cromileptes altivelis*) (Pisces: Serranidae, Epinephelinae). Aquaculture 287: 191–202. http://dx.doi.org/10.1016/j.aquaculture.2008.10.027

Lloris D. **1981**. Peces capturados, en el SO Africano, durante la campana "Benguela I" (Noviembre 1979). *Resultados Expediciones Científicas* 9: 17–28.

Lloris D. **1982**. Peces capturados, en el SO Africano, durante la campaña "Benguela II" (Julio-Septiembre 1980). *Resultados Expediciones Cientificas* 10: 3–15.

Lloris D. **1986**. Ictiofauna demersal y aspectos biogeográphicos de la costa sudoccidental de África (SWA/NAMIBIA). *Monografías de Zoología Marina* 1: 9–432.

Lloris D, Rucabado J. **1991**. *Heteronarce rierai*, a new narkid ray from Mozambique, western Indian Ocean. *Japanese Journal of Ichthyology* 37(4): 327–332.

Lloyd RE. **1906**. Natural history notes from the R.I.M.S. ship *Investigator*, Capt. T.H. Heming, R.N. (retired), commanding. Series 3, No. 14. Notes on the skull of the genus *Aulastomatomorpha*, with descriptions of some new deep-sea fish. *Annals and Magazine of Natural History* (Series 7) 18(106): 306–311.

Lloyd RE. **1907**. Contributions to the fauna of the Arabian Sea, with descriptions of new fishes and crustacea. *Records of the Indian Museum* (Calcutta) 1(1): 1–12.

Lloyd RE. **1908**. *Illustrations of the zoology of the Royal Indian* marine surveying steamer Investigator ... Fishes. Part 9. Calcutta. Pls. 39–43.

Lloyd RE. **1908**. On two species of eagle rays (Myliobatidae), with notes on the skull of the genus *Ceratoptera*. *Records of the Indian Museum* (Calcutta) 2(2): 175–180.

Lloyd RE. **1909**. A description of the deep-sea fish caught by the R.I.M.S. ship *Investigator* since the year 1900, with supposed evidence of mutation in *Malthopsis*. *Memoirs of the Indian Museum* 2(3): 139–180.

Lloyd RE. **1909**. *Illustrations of the zoology of the Royal Indian marine survey ship* Investigator ... Fishes. Part 10. Calcutta. Pls. 44–50.

Lo P-C, Lin S-H, Nor SAH, Chen W-J. **2017**. Molecular exploration of hidden diversity in the Indo-Pacific sciaenid clade. *PLoS ONE* 12(4): e0176623. https://doi.org/10.1371/ journal.pone.0176623 Lobel PS. **1979**. Description of a new Hawaiian gobiid fish of the genus *Trimma*. *Breviora* 456: 1–15.

Locket NA. **1980**. Some advances in coelacanth biology. *Proceedings of the Royal Society of London* B 208(1172): 265–307.

Loewenberg U. **2006**. *Stock Assessment Component. Fisheries Data Frame Survey. 2nd and final Part. Fisheries Monitoring, Control and Surveillance Project* (EUROPEAID/111995/C/ SV/YE). Mission report to the European Commission.

Löfling P. **1758**. Iter Hispanicum, eller resa til Spanska länderna uti Europa och America, förrättad ifrän År 1751 til År 1756 ... utgifven efter dess frånfålle af C. Linnåus. Stockholm: tryck på L. Salvii köstrad. 316 pp.

Loiselle PV, Stiassny MLJ. **2007**. Rehabilitation of the Malagasy endemic *Kuhlia sauvagii* Regan, 1913 (Teleostei: Perciformes), with the designation of a neotype for *Centropomis rupestris* Lacépéde [sic], 1802. *American Museum Novitates* 3561: 1–13, Pl. 1.

Lomolino MV, Riddle BR, Whittaker RJ, Brown JH. **2010**. *Biogeography* (4th edition). Sunderland: Sinauer Associates. 764 pp.

Loneux M. **2006**. The Castelnau's fish collection and watercolour notebooks (pp. 91–94). In: Segers H, Desmet P, Baus E (eds) *Tropical biodiversity: Science, data, conservation. Proceedings of the 3rd GBIF Science Symposium, Brussels, 18–19 April 2005.* 167 pp.

Long DJ, McCosker JE. **1999**. A new species of deep-water skate, *Rajella eisenhardti*, (Chondrichthyes: Rajidae) from the Galápagos Islands. *Proceedings of the Biological Society of Washington* 112(1): 45–51.

Long JA, Choo B, Clement A. 2019. The evolution of fishes through geological time (pp. 3–29). In: Johanson Z, Underwood C, Richter M (eds) *Evolution and development* of fishes. Cambridge, UK: Cambridge University Press. 274 pp. http://dx.doi.org/10.1017/9781316832172.002

Long JA, Mark-Kurik E, Johanson Z, Lee MSY, Young GC, Zhu M, Ahlberg PE, Newman M, Jones R, Den Blaauwen J, Choo B, Trinajstic K. 2015. Copulation in antiarch placoderms and the origin of gnathostome internal fertilization. *Nature* 517: 196–199. http://dx.doi.org/10.1038/nature13825

Longo SJ, Faircloth BC, Meyer A, Westneat MW, Alfaro ME, Wainwright PC. 2017. Phylogenomic analysis of a rapid radiation of misfit fishes (Syngnathiformes) using ultraconserved elements. *Molecular Phylogenetics* and Evolution 113: 33–48. http://dx.doi.org/10.1016/j. ympev.2017.05.002

López-Arbarello A. 2004. The record of Mesozoic fishes from Gondwana (excluding India and Madagascar) (pp. 597–624). In: Arratia G, Tintori A (eds) Mesozoic Fishes 3 – Systematics, paleoenvironments and biodiversity. Proceedings of the international meeting, Serpianao, 2001. Munich: Verlag Dr. Friedrich Pfeil. 649 pp. López-Horgue MA, Poyato-Ariza FJ, Cavin L, Bermúdez-Rochas DD. **2014**. Cenomanian transgression in the Basque-Cantabrian Basin (northern Spain) and associated faunal replacement. *Journal of Iberian Geology* 40(3): 489–506.

http://dx.doi.org/10.5209/rev\_JIGE.2014.v40.n3.42819

Losse GF. **1968**. The elopoid and clupeoid fishes of East African coastal waters. *Journal of the East Africa Natural History Society and National Museum* 27(2): 77–115.

Lotan R. **1970**. Systematic remarks on fishes of the family Salariidae in the Red Sea. *Israel Journal of Zoology* 18(4) [for 1969]: 363–378.

Lourie SA. **2003**. Measuring seahorses. *Project Seahorse Technical Report* No. 4 (version 1.0). Project Seahorse, Fisheries Centre, University of British Columbia. 15 pp.

Lourie SA, Foster S, Cooper E, Vincent A. **2004**. *A guide to the identification of seahorses*. Project Seahorse and TRAFFIC North America. Washington, DC: University of British Columbia and World Wildlife Fund. 120 pp.

Lourie SA, Kuiter RH. **2008**. Three new pygmy seahorse species from Indonesia (Teleostei: Syngnathidae: *Hippocampus*). *Zootaxa* 1963: 54–68. http://dx.doi. org/10.11646/zootaxa.1963.1.4

Lourie SA, Pritchard JC, Casey SP, Truong SK, Hall HJ, Vincent ACJ. **1999**. The taxonomy of Vietnam's exploited seahorses. *Biological Journal of the Linnean Society* 66(2): 231–256.

Louw E, O'Toole MJ. 1977. Larval development of Sardinops ocellata (Pisces: Clupeidae). Annals of the South African Museum 72(7): 125–145.

Lovejoy NR. **2000**. Reinterpreting recapitulation: systematics of needlefishes and their allies (Teleostei: Beloniformes). *Evolution* 54(4): 1349–1362.

Lovejoy NR, Iranpour M, Collette BB. **2004**. Phylogeny and jaw ontogeny of beloniform fishes. *Integrative and Comparative Biology* 44(5): 366–377.

Løvtrup S. **1977**. *The phylogeny of Vertebrata*. New York: John Wiley & Sons. xii + 330 pp.

Lowe RT. **1833**. Description of *Alepisaurus*, a new genus of fishes. *Proceedings of the Zoological Society of London* 1833(1): 104.

Lowe RT. **1834**. A collection of fishes made in that island [Madeira] ... They include the following species ... regarded as hitherto undescribed ... [extracts from a letter presented to Zoological Society, Dec. 24, 1833]. *Proceedings of the Zoological Society of London* 1833(1): 142–144.

Lowe RT. 1836. Piscium Maderensium species quaedam novae, vel minus rite cognitae breviter descriptae. *Transactions of the Cambridge Philosophical Society* 6(1): 195–202.
[Sometimes cited as 1838; cover of part 1 is dated 1836; part 3 and volume cover (pts 1–3) dated 1838.]

Lowe RT. **1838**. A synopsis of the fishes of Madeira; with the principal synonyms, Portuguese names, and characters of the new genera and species. *Transactions of the Zoological Society of London* 2(3, art. 14): 173–200.

Lowe RT. 1839. A supplement to a synopsis of the fishes of Madeira. Proceedings of the Zoological Society of London 1837(7): 76–92. [Also published in 1849 in Transactions of the Zoological Society of London 3(1): 1–20]

Lowe RT. **1841**. Description of certain new species of Madeiran fishes, with additional information relating to those already described. *Proceedings of the Zoological Society of London* 8: 36–39.

Lowe RT. **1843**. Notices of fishes newly observed or discovered in Madeira during the years 1840, 1841, and 1842. *Proceedings of the Zoological Society of London* 11: 81–95.

Lowe RT. **1843–1860**. *A history of the fishes of Madeira, with original figures from nature of all species, by Hon. C.E.C. Norton and M. Young* (5 parts). London: J. Van Voorst.

Lowe RT. 1846. On a new genus of the family Lophidae (les pectorales pediculées, Cuv.) discovered in Madeira. *Proceedings of the Zoological Society of London* 14: 81–84. [Also published in *Transactions of the Zoological Society of London* 1846 3(4, art.15): 339–344]

Lowe RT. **1850**. An account of fishes discovered or observed in Madeira since the year 1842. *Proceedings of the Zoological Society of London* 18: 247–253.

Loya Y. **1972**. Community structure and species diversity of hermatypic corals at Eilat, Red Sea. *Marine Biology* 13: 100–123.

Lozano [Rey] L. 1934. Algunos peces pelágicos o de profundidad procedentes del Mediterráneo occidental. *Boletín de la real Sociedad Española de Historia Natural* 34: 85–91.

Lozano Rey L. **1960**. Peces fisoclistos. Part 3a. *Memorias de la Real Academia de Ciencias de Madrid* 14(3): 1–615.

Lubbock R. **1975**. Fishes of the family Pseudochromidae (Perciformes) in the northwest Indian Ocean and Red Sea. *Journal of Zoology* (London) 176(1): 115–157.

Lubbock R. **1976**. Fishes of the family Pseudochromidae (Perciformes) in the central Indian Ocean. *Journal of Natural History* 10(2): 167–177.

Lubbock R. **1977**. Fishes of the family Pseudochromidae (Perciformes) in the Western Indian Ocean. *Ichthyological Bulletin of the J.L.B. Smith Institute of Ichthyology* 35: 1–21.

Lubbock R. **1980**. The shore fishes of Ascension Island. *Journal* of Fish Biology 17(3): 283–303.

Lubbock R, Allen GR. **1978**. A distinctive new *Anthias* (Teleostei: Serranidae) from the western Pacific. *Records of the Western Australian Museum* 6 (2): 259–268.

Lubbock R, Polunin NVC. **1977**. Notes on the Indo-West Pacific genus *Ctenogobiops* (Teleostei: Gobiidae), with descriptions of three new species. *Revue Suisse de Zoologie* 84(2): 505–514. Lubbock R, Randall JE. **1978**. Fishes of the genus *Liopropoma* (Teleostei: Serranidae) in the Red Sea. *Zoological Journal of the Linnean Society* 64(3): 187–195.

Lucena Rosa IM de. **1993**. Systematic study of the family Creediidae (Perciformes: Trachinoidei). PhD thesis, Department of Zoology, University of Alberta, Edmonton. 188 pp.

Ludt WB, Burridge CP, Chakrabarty P. **2019**. A taxonomic revision of Cheilodactylidae and Latridae (Centrarchiformes: Cirrhitoidei) using morphological and genomic characters. *Zootaxa* 4585(1): 121–141. http://dx.doi.org/10.11646/zootaxa.4585.1.7

Lugendo BR. **2007**. Utilisation by fishes of shallow-water habitats including mangroves and seagrass beds along the Tanzanian coast. PhD thesis, Faculty of Sciences, Radboud University Nijmegen, The Netherlands. 168 pp.

Lugendo BR, Nagelkerken I, Jiddawi N, Mgaya YD, Van Der Velde G. **2007**. Fish community composition of a tropical nonestuarine embayment in Zanzibar, Tanzania. *Fisheries Science* 73: 1213–1223.

Lund R. **1977**. *Echinochimaera meltoni*, new genus and species (Chimaeriformes), from the Mississippian of Montana. *Annals of Carnegie Museum* 46: 195–221.

Lund R. **1982**. *Harpagofututor volsellorhinus* new genus and species (Chondrichthyes, Chondrenchelyiformes) from the Namurian Bear Gulch Limestone, *Chondrochelys problematica* Traquair (Visean), and their sexual dimorphism. *Journal of Paleontology* 56(4): 938–958.

Lund R. **1985**. The morphology of *Falcatus falcatus* (St. John and Worthen), a Mississippian stethacanthid chondrichthyan from the Bear Gulch Limestone of Montana. *Journal of Vertebrate Paleontology* 5(1): 1–19. http://dx.doi.org/10.1080/02724634.1985.10011842

Lund R. 1988. New Mississippian Holocephali (Chondrichthyes) and the evolution of Holocephali.
In: Russell DE, Santoro JP, Sigogneau-Russell D (eds) Teeth revisited: Proceedings of the VIIth International Symposium on Dental Morphology, Paris 20–24 Mai 1986. *Mémoires du Muséum National d'Histoire Naturelle* (Série C, Sciences de la terre) 53: 195–205.

Lundberg B, Lipkin Y. **1979**. Natural food of the herbivorous rabbitfish (*Siganus* spp.) in northern Red Sea. *Botanica Marina* 22(3): 173–182.

Lundberg JG, Friel JP. **2003**. Siluriformes. Catfishes. Version 20 January 2003. http://tolweb.org/ Siluriformes/15065/2003.01.20 [The Tree of Life Web Project]

Lunel G. **1881**. Mélanges ichthyologiques. Liste de quelques espèces de poissons, nouvelles pour la faune de l'île Maurice. *Mémoires de la Société de Physique et d'Histoire Naturelle de Genève* 27(2): 266–303. Luther G. **1966.** On the little known fish *Chirocentrus nudus* Swainson from the Indian Seas, and its comparison with *Chirocentrus dorab* (Forsskål). *Journal of the Marine Biological Association of India* 8(1): 193–201.

Lutjeharms JRE. **1991**. The temperature/salinity relationships of the South West Indian Ocean. *South African Geographer* 18(1/2): 15–31.

Lutjeharms JRE. **1998**. Coastal hydrography (pp. 50–61). In: Lubke R, De Moor I (eds) *Field guide to the eastern and southern Cape coasts*. Rondebosch: University of Cape Town Press. 559 pp.

Lutjeharms JRE. 2006. *The Agulhas Current*. Berlin: Springer-Verlag. 329 pp.

Lutjeharms JRE. **2006**. The coastal oceans of south-eastern Africa (pp. 783–834). In: Robinson AR, Brink KH (eds) *The sea, Volume 14B: The global coastal ocean: Interdisciplinary regional studies and syntheses.* Cambridge, Massachusetts: Harvard University Press. 809 pp. [i–xxiv + 783–1567 pp.]

Lutjeharms JRE. 2011. The southern African oceans (pp. 264–267). In: Zietsman [H]L (ed) Observations on environmental change in South Africa. Section 4.
Stellenbosch: SUN MeDIA (under the SUN PRESS imprint). 302 pp.

Lutjeharms JRE, Roberts HR. **1988**. The Natal Pulse: an extreme transient on the Agulhas Current. *Journal of Geophysical Research* 93(C1): 631–645.

Lütken CF. **1852**. Nogle bemaerkninger om naeseborenes stilling hos de i gruppe med *Ophisurus* staaende slaegter af aalefamilien. *Videnskabelige Meddelelser fra den Naturhistoriske Forening i Kjøbenhavn Aaret* 1851(1–2): 1–21.

Lütken CF. **1880**. Spolia Atlantica. Bidrag til Kundskab om Formforandringer hos Fiske under deres Vaext og Udvikling, saerligt hos nogle af Atlanterhavets Højsøfiske. *Det Kongelige Dansk Videnskabernes Selskabs Skrifter, Naturvidenskabelig og mathematisk afdeling, Kjøbenhavn* (Series 5) 12: 409–613.

Lütken CF. 1883. In: Jordan DS, Gilbert CH.

Lütken CF. **1898**. *The Danish* Ingolf *Expedition: The ichthyological results*. Vol. 2(1). Copenhagen: Bianco Luno (F. Dreyer). 39 pp.

## Μ

Mabuchi K, Fraser TH, Song H, Azuma Y, Nishida M. 2014. Revision of the systematics of the cardinalfishes (Percomorpha: Apogonidae) based on molecular analyses and comparative reevaluation of morphological characters. *Zootaxa* 3846(2): 151–203. http://dx.doi.org/10.11646/ zootaxa.3846.2.1 Mabuchi K, Okuda N, Nishida M. **2006**. Molecular phylogeny and stripe pattern evolution in the cardinalfish genus *Apogon. Molecular Phylogenetics and Evolution* 38: 90–99.

Machida Y. 1994. Descriptions of three new and one resurrected species of the bythitid genus *Dinematichthys* (Ophidiiformes). *Japanese Journal of Ichthyology* 40(4): 451–464.

Machida Y, Yoshino T. 1984. First record of Brosmophyciops pautzkei (Bythitidae, Ophidiiformes) from Japan. Memoirs of the Faculty of Science, Kochi University (Series D) (Biology) 5: 37–41.

Macleay W. **1878**. The fishes of Port Darwin. *Proceedings of the Linnean Society of New South Wales* 2(pt 4): 344–367.

Macleay W. **1881**. Descriptive catalogue of the fishes of Australia. Part 1. *Proceedings of the Linnean Society of New South Wales* 5(pt 3): 302–444.

Macleay W. **1881**. Descriptive catalogue of the fishes of Australia. Part 4. *Proceedings of the Linnean Society of New South Wales* 6(pt 2): 202–387.

Macleay W. **1882**. Contribution to a knowledge of the fishes of New Guinea. No. 1. *Proceedings of the Linnean Society of New South Wales* 7(pt 2): 224–250.

Macleay W. **1883**. Contribution to a knowledge of the fishes of New Guinea. No. 3. *Proceedings of the Linnean* Society of *New South Wales* 7(pt 4): 585–598.

Macleay W. **1883**. Contribution to a knowledge of the fishes of New Guinea. No. 4. *Proceedings of the Linnean Society of New South Wales* 8(pt 2): 252–280.

Macpherson E. **1985**. Daily ration and feeding periodicity of some fishes off the coast of Namibia. *Marine Ecology Progress Series* 26: 253–260.

Macpherson E, Roel BA. **1987**. Trophic relationships in the demersal fish community off Namibia. *South African Journal of Marine Science* 5(1): 585–596.

Madden WD. **1973**. The collection of live fishes from a salvaged vessel. *Copeia* 1973(1): 144–145.

Madhupratap M, Haridas P, Ramaiah N, Achuthankutty CT. **1992**. Zooplankton in the southwest coast of India (pp. 99–112). In: Desai BN (ed) *Oceanography of the Indian Ocean*. Rotterdam: AA Balkema. 770 pp.

Madhupratap M, Kumar SP, Bhattathiri PMA, Dileep Kumar M, Raghukumar S, Nair KKC, Ramaiah N. **1996**. Mechanism of the biological response to winter cooling in the northeastern Arabian Sea. *Nature* 384: 549–552.

Madhupratap M, Nair KNV, Gopalakrishnan TC, Haridas P, Nair KKC, Venugopal P, Gauns M. **2001**. Arabian Sea oceanography and fisheries of the west coast of India. *Current Science* 81: 355–360.

Madhusoodana Kurup B, Manojkumar TG, Radhakrishnan KV. **2005**. *Salarias reticulatus* (Pisces: Blennidae [sic]), a new freshwater blenny from Chalakudy River, Kerala

(South India). *Journal of the Bombay Natural History Society* 102(2): 195–197.

Magnus DBE. **1967**. Zur Ökologie sedimentbewohnender *Alpheus* Garnelen (Decapoda, Natantia) des Roten Meeres. *Helgoländer Wissenschaftliche Meeresuntersuchungen* 15(1–4): 506–522.

Magtoon W, Termvidchakorn A. **2009**. A revised taxonomic account of ricefish *Oryzias* (Beloniformes; Adrianichthyidae), in Thailand, Indonesia and Japan. *The Natural History Journal of Chulalongkorn University* 9(1): 35–68.

Mahadevan S, Nayar KN. 1965. Underwater ecological observations in the Gulf of Mannar off Tuticorin Island.Association between a fish (*Gnathanodon*) and a sea snake.*Ibid* 7(1): 197–199.

Mahdi N. **1962**. *Fishes of Iraq*. Baghdad: Ministry of Education. 82 pp.

Maigret J, Ly B. **1986**. *Les poissons de mer de Mauritanie*. Compiègne, France: Sciences Naturelles. 213 pp.

Maisey JG. **1986**. Heads and tails: a chordate phylogeny. *Cladistics* 2(4): 201–256.

Maisey JG. **1989**. *Hamiltonichthys mapesi*, g. & sp. nov. (Chondrichthyes, Elasmobranchii), from the Upper Pennsylvanian of Kansas. *American Museum Novitates* 2931: 1–42.

Maisey JG. **1991**. *Santana fossils: an illustrated atlas*. Neptune City, New Jersey: TFH Publications. 459 pp.

Maisey JG. **2012**. What is an 'elasmobranch'? The impact of palaeontology in understanding elasmobranch phylogeny and evolution. *Journal of Fish Biology* 80(4): 918–951. http://dx.doi.org/10.1111/j.1095-8649.2012.03245.x

Maisey JG, Denton JSS, Burrow C, Pradel A. **2021**. Architectural and ultrastructural features of tessellated calcified cartilage in modern and extinct chondrichthyan fishes. *Journal of Fish Biology* 98(4): 919–941. https://doi. org/10.1111/jfb.14376

Maisey JG, Janvier P, Pradel A, Denton JSS, Bronson A,
Miller R, Burrow CJ. 2019. *Doliodus* and pucapampellids: contrasting perspectives on stem chondrichthyan morphology (pp. 87–109). In: Johanson Z, Underwood C, Richter M (eds) *Evolution and development of fishes*. Cambridge, UK: Cambridge University Press. 274 pp. http://dx.doi.org/10.1017/9781316832172.006

Maisey JG, Miller R, Pradel A, Denton JSS, Bronson A, Janvier P. **2017**. Pectoral morphology in *Doliodus*: bridging the 'acanthodian'-chondrichthyan divide. *American Museum Novitates* 3875: 1–15. https://doi.org/10.1206/3875.1

Maisey JG, Naylor GJP, Ward DJ. **2004**. Mesozoic elasmobranchs, neoselachian phylogeny and the rise of modern elasmobranch diversity (pp. 17–56). In: Arratia G, Tintori A (eds) *Mesozoic Fishes 3 – Systematics*, *paleoenvironments and biodiversity. Proceedings of the international meeting, Serpianao, 2001.* Munich: Verlag Dr. Friedrich Pfeil. 649 pp.

Mallatt J, Sullivan J. **1998**. 28S and 18S rDNA sequences support the monophyly of lampreys and hagfishes. *Molecular Biology and Evolution* 15(12): 1706–1718.

Malm AW. **1877**. *Göteborgs och Bohusläns fauna*, *ryggradsdjuren*. Göteborg: Göteborgs handelstidnings aktiebolags tryckeri. 674 pp.

Mandrytsa SA. **1990**. On the validity of *Ectreposebastes niger* and distribution of species of the genus *Ectreposebastes* (Pisces: Scorpaenidae). In: *Investigations in vertebrates* [sic] *zoology. Trudy Zoologicheskogo Instituta, Akademii Nauk USSR* 213: 29–34. [In Russian, English summary]

Mandrytsa SA. **1990**. The contraction of the musculus adductor mandibulae of the scorpaeniform fishes. *Proceedings of the Zoological Institute, USSR Academy of Sciences* 222: 75–93.

Mandrytsa SA. **1990**. New fish species of the genus *Minous* Cuvier (Pisces: Synanceiidae) from the Indian Ocean. *Biologiya Morya* 6: 68–69. [In Russian, English summary]

Mandrytsa SA. **1992.** The new species and new records of the fishes from the genera *Phenacoscorpius* and *Plectrogenium* in the Pacific, Atlantic and Indian Oceans. *Voprosy Ikhtiologii* 32(4): 10–17. [In Russian, English translation in *Journal of Ichthyology* 32(8): 100–109]

Mandrytsa SA. 1993. Structure of the seismosensory system of the members of the family Pataecidae (Acanthopterygii).
In: *Investigations on taxonomy and morphology of fishes. Trudy Zoologicheskogo Instituta, Akademii Nauk USSR* 235 [for 1991]: 29–36. [In Russian, English summary]

Mandrytsa SA. 1993. Two new species from the genera *Choridactylus* and *Minous* from the Gulf of Aden (Scorpaeniformes, Synanceiidae). *Voprosy Ikhtiologii* 33(1): 137–141. [In Russian, English translation in *Journal of Ichthyology* 33(5): 136–142]

Mandrytsa SA. 2001. Lateral line system and classification of scorpaenoid fishes (Scorpaeniformes: Scorpaenoidei). PhD dissertation, Perm University. 393 pp. [In Russian, English summary]

Mandrytsa SA. **2002**. New species of the genus *Pteropterus* (Scorpaenidae, Scorpaeniformes) from the Indian Ocean. *Voprosy Ikhtiologii* 42(1): 129–130. [In Russian, English translation in *Journal of Ichthyology* 42(1): 124–125]

Mandrytsa SA. **2002**. New records of rare scorpaenid fishes from the genera *Pteropelor* and *Rhinopias* (Scorpaenidae) in the Indian and western Pacific oceans. *Journal of Ichthyology* 42(7): 544–547.

Mandrytsa SA, Usachev SI. **1990**. A new species of the genus *Ocosia* Jordan and Starks (Scorpaeniformes, Tetrarogidae) from the western part of the Indian Ocean. *Voprosy Ikhtiologii* 30(2): 325–327. [In Russian. English translation in *Journal of Ichthyology* 30(3): 130–134] Manilo LG. **1990**. New species of the genus *Parapercis* (Mugiloididae) from the region of the Maldive Islands. *Voprosy ikhtiologii* 30(6): 1016–1019. [In Russian, English translation in *Journal of Ichthyology* 31(1): 151–155]

Manilo LG, Bogorodsky SV. **2003**. Taxonomic composition, diversity and distribution of coastal fishes of the Arabian Sea. *Journal of Ichthyology* 43(Suppl. 1): S75–S149.

Manjaji BM. **2004**. Taxonomy and phylogenetic systematics of the Indo-Pacific whip-tailed stingray genus *Himantura* Müller and Henle 1837 (Chondrichthyes: Myliobatiformes: Dasyatidae). PhD thesis, University of Tasmania. xxii + 607 pp.

Manjaji-Matsumoto BM, Last PR. **2006**. *Himantura lobistoma*, a new whipray (Rajiformes: Dasyatidae) from Borneo, with comments on the status of *Dasyatis microphthamus*. *Ichthyological Research* 53(3): 290–297.

Manjaji-Matsumoto BM, Last PR. **2008**. *Himantura leoparda* sp. nov., a new whipray (Myliobatoidei, Dasyatidae) from the Indo-Pacific (pp. 293–302). In: Last PR, White WT, Pogonoski JJ (eds) *Descriptions of new Australian chondrichthyans*. Hobart, Australia: CSIRO Marine and Atmospheric Research. 358 pp.

Manjaji-Matsumoto BM, Last PR. 2016. Two new whiprays, Maculabatis arabica sp. nov. and M. bineeshi sp. nov. (Myliobatiformes: Dasyatidae), from the northern Indian Ocean. Zootaxa 4144(3): 335–353. http://dx.doi. org/10.11646/zootaxa.4144.3.3

Manjebrayakath H, Akhilesh KV, Pillai NGK. **2012**. Report of *Apogonichthyoides sialis* (Perciformes: Apogonidae) from the west coast of India. *Marine Biodiversity Records* 5(e15): 1–3. http://dx.doi.org/10.1017/S1755267211001175

Manjusha U, Ambrose TV, Remya R, Paul S, Jayasankar J, Vivekanandan E. **2010**. Seasonal and interannual changes in oceanographic features and their impact on small pelagic catches off Kerala. In: *Book of Abstracts, SAFARI International Symposium on Remote Sensing & Fisheries, 15–17 February 2010, Kochi.* p. 83.

Mann DA, Lu Z, Popper AN. **1997**. Clupeid fishes can detect ultrasound. *Nature* 389: 341.

Marais JFK. **1984**. Feeding ecology of major carnivorous fish from four Eastern Cape estuaries. *South African Journal of Zoology* 19(3): 210–223.

Marceniuk AP, Menezes NA. **2007**. Systematics of the family Ariidae (Ostariophysi, Siluriformes), with a redefinition of the genera. *Zootaxa* 1416: 1–126.

Marceniuk AP, Menezes NA, Britto MR. **2012**. Phylogenetic analysis of the family Ariidae (Ostariophysi: Siluriformes), with a hypothesis on the monophyly and relationships of the genera. *Zoological Journal of the Linnean Society* 165(3): 534–669. https://doi.org/10.1111/j.1096-3642.2012.00822.x Marine Research Section. **1986**. *Catalogue of fishes of the Maldives*. Vol. 1. Malé, Republic of Maldives: Marine Research Section, Ministry of Fisheries. pp. 1–160.

Marine Research Section. **1987**. *Catalogue of fishes of the Maldives*. Vol. 2. Malé, Republic of Maldives: Marine Research Section, Ministry of Fisheries. pp. 161–330.

Marine Research Section. **1988**. *Catalogue of fishes of the Maldives*. Vol 3. Malé, Republic of Maldives: Marine Research Section, Ministry of Fisheries. pp. 331–494.

Marine Research Section. **1992**. *Catalogue of fishes of the Maldives*. Vol. 4. Malé, Republic of Maldives: Marine Research Section, Ministry of Fisheries and Agriculture. pp. 495–650.

Marine Research Section. **1997**. *Fishes of the Maldives*. Malé, Republic of Maldives: Marine Research Section, Ministry of Fisheries and Agriculture. 408 pp.

Marino RP, Dooley JK. **1982**. Phylogenetic relationships of the tilefish family Branchiostegidae (Perciformes) based upon comparative myology. *Journal of Zoology* 196: 151–163.

Markle DF, Olney JE. **1990**. Systematics of the pearlfishes (Pisces: Carapidae). *Bulletin of Marine Science* 47(2): 269–410.

Markle DF, Quéro J-C. 1984. Alepocephalidae (including Bathylaconidae, Bathyprionidae) (pp. 228–258),
Leptochilichthyidae (included in Alepocephalidae in CLOFNAM) (pp. 254–255). In: Whitehead PJP, Bauchot M-L, Hureau J-C, Nielsen J, Tortonese E (eds) *Fishes of the north-eastern Atlantic and the Mediterranean*. Vol. I. Paris: UNESCO.

Marshall AD, Compagno LJV, Bennett MB. **2009**. Redescription of the genus *Manta* with resurrection of *Manta alfredi* (Krefft,1868) (Chondrichthyes; Myliobatoidei; Mobulidae). *Zootaxa* 2301: 1–28. https://doi.org/10.11646/ zootaxa.2301.1.1

Marshall AD, Pierce SJ, Bennett MB. **2008**. Morphological measurements of manta rays (*Manta birostris*) with a description of a foetus from the east coast of southern Africa. *Zootaxa* 1717: 24–30. http://dx.doi.org/10.11646/ zootaxa.1717.1.2

Marshall NB. **1950**. Fishes from the Cocos-Keeling Islands. *Bulletin of the Raffles Museum* 22: 166–205.

Marshall NB. **1952**. The *Manihine* expedition to the Gulf of Aqaba 1948–1949. IX. Fishes. *Bulletin of the British Museum* (*Natural History*) *Zoology* 1(8): 221–252.

Marshall NB. **1954**. *Aspects of deep sea biology*. London: Hutchinson. 380 pp.

Marshall NB. **1955**. Alepisauroid fishes. *Discovery Report* 27: 303–336.

Marshall NB. **1966**. The relationships of the anacanthine fishes, *Macruronus, Lyconus*, and *Steindachneria. Copeia* 1966(2): 275–280.

Marshall NB. **1973**. Family Macrouridae. In: Fishes of the western North Atlantic. *Memoir of the Sears Foundation for Marine Research* 1(part 6): 496–662.

Marshall NB, Bourne DW. **1964**. A photographic survey of benthic fishes in the Red Sea and Gulf of Aden, with observations on their population density, diversity, and habits. *Bulletin of the Museum of Comparative Zoology* 132(2): 223–244.

Marshall NB, Cohen DM. **1973**. Order Anacanthini (Gadiformes): characters and synopsis of families. In: Fishes of the western North Atlantic. *Memoir of the Sears Foundation for Marine Research* 1(part 6): 479–495.

Marshall TC. **1951**. *Ichthyological Notes* 1(1): 1–9. Brisbane, Queensland: Department of Harbours and Marine.

Marshall TC. **1957**. *Ichthyological Notes* 1(3): 1–9. Brisbane, Queensland: Department of Harbours and Marine.

Marshall TC. **1964**. *Fishes of the Great Barrier Reef and coastal waters of Queensland*. Sydney: Angus and Robertson. 556 pp.

Martin TJ, Heemstra PC. **1988**. Identification of *Ambassis* species (Pisces: Perciformes, Ambassidae) from South Africa. *South African Journal of Zoology* 23(1): 7–12.

Martinez-Takeshita N, Purcell CM, Chabot CL, Craig MT, Paterson CN, Hyde JR, Allen LG. **2015**. A tale of three tails: cryptic speciation in a globally distributed marine fish of the genus *Seriola*. *Copeia* 103(2): 357–368. https://doi. org/10.1643/CI-124-224

Martini FH, Flescher D. **2002**. Hagfishes (pp. 9–16). Order Myxiniformes. In: Collette BB, Klein-MacPhee G (eds) *Bigelow and Schroeder's fishes of the Gulf of Maine* (3<sup>rd</sup> edition). Washington, DC: Smithsonian Institution Press. 882 pp.

Mary AA, Balasubramanian T, Selvaraju S, Shiny A. **2017**. Description of a new species of clupeid fish, *Amblygaster indiana* (Clupeiformes: Clupeidae), off Eraviputhenthurai, west coast of India. *Zootaxa* 4247(4): 461–468. https://doi.org/10.11646/zootaxa.4247.4.7

Masuda H. **1984**. *Field guide for sea fishes*. Tokyo: Tokai University Press. 227 pp. [In Japanese]

Masuda H, Amaoka K, Araga C, Uyeno T, Yoshino T (eds). **1984**. *The fishes of the Japanese archipelago*. Tokyo: Tokai University Press. 437 pp.

Masuda H, Amaoka K, Araga C, Uyeno T, Yoshino T (eds). **1988**. *The fishes of the Japanese archipelago* (2<sup>nd</sup> edition). Tokyo: Tokai University Press. 456 pp.

Masuda H, Araga C, Yoshino T. **1975**. *Coastal fishes of southern Japan*. Tokyo: Tokai University Press. 379 pp. [In English & Japanese]

Masuda H, Kobayashi Y. **1994**. *Grand atlas of fish life modes: color variation in Japanese fish*. Tokyo: Tokai University Press. 465 pp. Masuda S. **1942**. Notes on the Japanese fishes of the genus *Epinephelus*. *Annotationes Zoologicae Japonenses* 21(2): 106–123.

Masuda S, Ozawa T. **1979**. Reexamination of the holotypes of *Bregmaceros japonicus* Tanaka and *B. nectabanus* Whitley. *Japanese Journal of Ichthyology* 25(4): 266–268.

Matallanas J. **1977**. Nuevas citas y datos biométricos de *Epigonus constanciae* (Giglioli) 1880 (Perciformes, Apogonidae) del Mediterráneo y de Cabo Blanco (Mauritania). *Vie et Milieu* 27(3-A): 369–376.

Mather FJ III, Clark HL, Mason JM Jr. **1975**. Synopsis of the biology of the white marlin, *Tetrapterus albidus* Poey (1861). *NOAA Technical Report* NMFS SSRF-675 (Part 3): 55–94.

Mathews CP, Samuel M. **1987**. Growth, mortality and assessment for groupers from Kuwait. *Kuwait Bulletin of Marine Science* 9: 173–191.

Mathews CP, Samuel M, Baddar MK. **1986**. Sexual maturation, length and age in some species of Kuwait fish related to their suitability for aquaculture. *Kuwait Bulletin of Marine Science* 8: 243–256.

Matsubara K. **1936**. A new bramid fish found in Japan. *Bulletin* of the Japanese Society of Scientific Fisheries 4(5): 297–300.

Matsubara K. **1936**. A new carcharoid shark found in Japan. *Dobutsugaku Zasshi* (Zoological Magazine Tokyo) 48(7): 380–382.

Matsubara K. **1936**. Biometry of two species of Japanese cardinal-fishes, with special reference to their taxonomy. *Journal of the Imperial Fisheries Institute, Tokyo* 31: 119–130.

Matsubara K. **1939**. Studies on the deep-sea fishes of Japan. XI. On an imperfectly known ribbon-fish, *Eumethichthys fiski* (Günther), with a supplement to the genus *Eumethichthys*. *Bulletin of the Biogeographical Society* 9(11): 193–199.

Matsubara K. **1943**. Studies on the scorpaenoid fishes of Japan. Anatomy, phylogeny and taxonomy (I). *Transactions of the Sigenkagaku Kenkyusho* no. 1: 1–170.

Matsubara K. **1943**. Studies on the scorpaenoid fishes of Japan (II). *Transactions of the Sigenkagaku Kenkyusyo* no. 2: 171–486.

Matsubara K. **1953**. On a new pearlfish, *Carapus owasianus*, with notes on the genus *Jordanicus* Gilbert. *Japanese Journal of Ichthyology* 3(1): 29–32.

Matsubara K. **1955**. *Fish morphology and hierarchy*. Vols. 1–3. Tokyo: Ishizaki-Shoten. 1605 pp. [In Japanese]

Matsubara K, Asano H. **1960**. A new eel of the genus *Kaupichthys. Copeia* 1959(4): 293–297.

Matsubara K, Iwai T. **1952**. Studies on some Japanese fishes of the family Gempylidae. *Pacific Science* 6(3): 193–212.

Matsubara K, Iwai T. **1958**. Anatomy and relationships of the Japanese fishes of the family Gempylidae. *Memoirs of the College of Agriculture Kyoto University* (June): 23–54.

Matsubara K, Ochiai A. **1955**. A revision of the Japanese fishes of the family Platycephalidae (the flatheads). *Memoirs of the College of Agriculture Kyoto University* 68: 1–109.

Matsubara K, Ochiai A. **1963**. Report on the flatfish collected by the Amami Islands Expedition in 1958. *Bulletin of the Misaki Marine Biological Institute* (Kyoto University) 4: 83–105.

Matsui T. **1967**. Review of the mackerel genera *Scomber* and *Rastrelliger* with description of a new species of *Rastrelliger*. *Copeia* 1967(1): 71–83.

Matsumoto T, Bayliff WH. **2008**. A review of the Japanese longline fishery for tunas and billfishes in the eastern Pacific Ocean, 1998–2003. *Bulletin of the Inter-American Tropical Tuna Commission* 24(1): 1–187.

Matsumoto WM. **1960**. Notes on the Hawaiian frigate mackerel of the genus *Auxis*. *Pacific Science* 14(2): 173–177.

Matsumoto WM, Skillman RA, Dizon AE. **1984**. Synopsis of biological data on skipjack tuna, *Katsuwonus pelamis*. *NOAA Technical Report* NMFS Circular 451: 1–92.

Matsunuma M, Jawad LA, Motomura H. **2013**. New records of a scorpionfish, *Parapterois macrura* (Scorpaenidae: Pteroinae), from Oman and Somalia, western Arabian Sea. *Biogeography* 15: 49–54. http://hdl.handle.net/10232/22439

Matsunuma M, Motomura H. **2013**. Newly recognized diagnostic characters of the poorly known lionfish *Pterois brevipectoralis* (Scorpaenidae: Pteroinae), with notes on fresh coloration. *Species Diversity* 18(2): 163–173. https://doi.org/10.12782/sd.18.2.163

Matsunuma M, Motomura H. **2014**. A new species of scorpionfish, *Ebosia saya* (Scorpaenidae: Pteroinae), from the western Indian Ocean and notes on fresh coloration of *Ebosia falcata*. *Ichthyological Research* 62(3): 293–312. https://doi.org/10.1007/s10228-014-0445-4

Matsunuma M, Motomura H. **2014**. *Pterois paucispinula*, a new species of lionfish (Scorpaenidae: Pteroinae) from the western Pacific Ocean. *Ichthyological Research* 62(3): 327–346. https://doi.org/10.1007/s10228-014-0451-6

Matsunuma M, Motomura H. **2015**. A new species of scorpionfish, *Ebosia vespertina* (Scorpaenidae: Pteroinae), from the southwestern Indian Ocean. *Ichthyological Research* 63(1): 110–120. http://dx.doi.org/10.1007/s10228-015-0479-2

Matsunuma M, Motomura H, Bogorodsky SV. **2017**. Review of Indo-Pacific dwarf lionfishes (Scorpaenidae: Pteroinae) in the *Dendrochirus brachypterus* complex, with description of a new species from the western Indian Ocean. *Ichthyological Research* 64(4): 369–414. https://doi.org/10.1007/s10228-017-0583-6

Matsunuma M, Sakurai M, Motomura H. **2013**. Revision of the Indo-West Pacific genus *Brachypterois* (Scorpaenidae: Pteroinae), with description of a new species from

northeastern Australia. *Zootaxa* 3693(4): 401–440. https://doi.org/10.11646/zootaxa.3693.4.1

Matsuura K. **1980**. A revision of Japanese balistoid fishes. I. Family Balistidae. *Bulletin of the National Science Museum* (Tokyo) (Series A) 6(1): 27–69.

Matsuura K. **1981**. *Xenobalistes tumidipectoris*, a new genus and species of triggerfish (Tetraodontiformes, Balistidae) from the Marianas Islands. *Bulletin of the National Science Museum* (Tokyo) (Series A) 7(4): 191–200.

Matsuura K. **1982**. A new triacanthodid fish, *Triacanthodes indicus*, from the Indian Ocean. *Japanese Journal of Ichthyology* 28(4): 385–392.

Matsuura K. 1984. Family Tetraodontidae (puffers) (pp. 362–365). In: Masuda H, Amaoka K, Araga C, Uyeno T, Yoshino T (eds) *The fishes of the Japanese archipelago*. Tokyo: Tokai University Press. 437 pp.

Matsuura K. **1985**. Triacanthodidae to Tetraodontidae (pp. 418–781). In: Okamura O (ed) *Fishes of the Okinawa Trough and the adjacent waters. The intensive research of unexploited fishery resources on continental slopes*. Vol. II. Tokyo: Japan Fisheries Resource Conservation Association.

Matsuura K. 1992. A new sharpnose puffer, Canthigaster punctata (Teleostei: Tetraodontidae) from the Mascaren submarine ridge, Western Indian Ocean. Bulletin of the National Science Museum (Tokyo) (Series A) 18(3): 127–130.

Matsuura K. **1994**. *Arothron caeruleopunctatus*, a new puffer from the Indo-western Pacific. *Japanese Journal of Ichthyology* 41(1): 29–33.

Matsuura K. 2001. Triacanthodidae, Triacanthidae, Balistidae, Monacanthidae, Ostraciidae, Aracanidae, Triodontidae, Tetraodontidae (pp. 3902–3957). In: Carpenter KE, Niem VH (eds) FAO species identification guide for fishery purposes. The living marine resources of the western central Pacific. Vol 6. Bony fishes part 4 (pp. 3381–4218). Rome: FAO.

Matsuura K. 2002. A review of two morphologically similar puffers, *Chelonodon laticeps* and *C. patoca. National Science Museum Monograph* 22: 173–178.

Matsuura K. **2015**. Taxonomy and systematics of tetraodontiform fishes: a review focusing primarily on progress in the period from 1980 to 2014. *Ichthyological Research* 62(1): 72–113. [online in 2014] http://dx.doi. org/10.1007/s10228-014-0444-5

Matsuura K, Golani D, Bogorodsky SV. **2011**. The first record of *Lagocephalus guentheri* Miranda Ribeiro, 1915 from the Red Sea with notes on previous records of *L. lunaris* (Actinopterygii, Tetraodontiformes, Tetraodontidae). *Bulletin of the National Museum of Nature and Science* (Tokyo) (Series A) 37(3): 163–169.

Matsuura K, Nguyen NH. **2008**. First record of the rare sharpnose puffer *Canthigaster inframacula* collected from

the South China Sea off Vietnam. *Memoirs of the National Museum of Nature and Science* 45: 59–63.

Matsuura K, Okuno J. **1991**. Redescription of a rare pufferfish, *Arothron carduus* (Cantor, 1849) (Teleostei: Tetraodontidae). *Bulletin of the National Science Museum* (Tokyo) (Series A) 17(3): 157–164.

Matsuura K, Shibukawa K, Shinohara G, Liu G. **2001**. Fishes collected from the shallow waters of Hainan Island, South China Sea. *National Science Museum Monograph* 21: 101–126.

Matsuura K, Tyler JC. **1997**. Tetraodontiform fishes, mostly from deep waters, of New Caledonia. No. 9. In: Séret B (ed) Résultats des Campagnes MUSORSTOM, Vol. 17. *Mémoires du Muséum National d'Histoire Naturelle* (N.S., Série A, Zoologie) 174: 173–208.

Matsuura K, Yamakawa T. **1982**. Rare boxfishes, *Kentrocapros flavofasciatus* and *K. rosapinto*, with notes on their relationships. *Japanese Journal of Ichthyology* 29(1): 31–42.

Matsuura K, Yoshino T. **1984**. Records of three tetraodontoid fishes from Japan. *Japanese Journal of Ichthyology* 31(3): 331–334.

Matsuura K, Yoshino T. **1994**. Records of three tetraodontoid fishes from Japan. *Japanese Journal of Ichthyology* 31: 331–334.

Matsuura K, Yoshino T. **2004**. A new triggerfish of the genus *Abalistes* (Tetraodontiformes: Balistidae) from the western Pacific. *Records of the Australian Museum* 56(2): 189–194.

Maugé AL [as Maugé LA]. **1967**. Contribution préliminaire à l'inventaire ichtyologique de la région de Tuléar. *Recueil des travaux de la station marine d'Endoume* (Marseille) Supplement 7: 101–132.

Maugé AL [as Maugé LA]. **1967**. Contribution préliminaire à l'inventaire ichthyologique de la région de Tuléar. *Annales de l'Université de Madagascar (Faculté des Sciences)* 5: 215–246.

Maugé AL [as Maugé LA]. **1978**. Statut de deux Syngnathidés signalés par H. Sauvage dans la région de Madagascar en 1891. *Cybium* (Série 3) 2(3): 95–99.

Maugé AL. **1980**. Note sur la presence de *Solegnathus hardwicki* (Gray, 1832) dans les eaux de l'île Maurice (Pisces: Syngnathidae). *Cybium* (Série 3) 4(9): 97–101.

Maugé AL. **1980**. La denture entopteryoidienne chez les subadultes de *Holanthias borbonius* (Valenciennes, 1828) (Pisces: Serranidae, Anthiinae). *Cybium* (Série 3) 4(11): 23–28.

Maugé AL. **1981**. *Syngnathus lumbricoides*, espèce nouvelle de syngnathe de l'Océan Indien occidental (Pisces, Teleostei, Syngnathidae). *Cybium* (Série 3) 5(1): 61–64.

Maugé AL. **1984**. Diagnoses préliminaires d'Eleotridae des eaux douces de Madagascar. *Cybium* 8(4): 98–100.

Maugé AL. **1984**. *Ambassis bleekeri* nom. nov. et réhabilitation d'*Ambassis gymnocephalus* (Lacépède, 1801) (Pisces, Teleostei, Ambassidae). *Bulletin du Muséum National* d'*Histoire Naturelle* (Paris) (Série 4) 6(1): 211–218.

Maugé AL. 1986. Ambassidae (pp. 297–298); Gobiidae (pp. 358–388); Eleotridae (pp. 389–398); Periophthalmidae (pp. 399–401). In: Daget J, Gosse J-P, Thys van den Audenaerde DFE (eds) *Check-list of the freshwater fishes of Africa* (*CLOFFA*). Vol. 2. Brussels: Institut Royal des Sciences Naturelles de Belgique. 521 pp.

Maugé AL, Bardach JE. **1985**. Congrogadinae de Madagascar (Pisces, Pseudochromidae) description d'*Halilmuraenoides isostigma* n.g. et n. sp. *Cybium* 9(4): 375–384.

Maugé AL, Bauchot R. 1976. Une espèce nouvelle de chétodon de l'océan Indien occidental: *Chaetodon guezei* (Pisces: Chaetodontidae). *Bulletin du Muséum National d'Histoire Naturelle* (Paris) (Série 3: Zoologie) 55: 89–101.

Maul GE. **1948**. Monografia dos peixes do Museu Municipal do Funchal. Ordem Isospondyli. *Boletim do Museu Municipal do Funchal* 3(art.5): 5–41.

Maul GE. **1952**. Monografia dos peixes do Museu Municipal do Funchal. Familia Gadidae e Bregmacerotidae. *Boletim do Museu Municipal do Funchal* 6(art.15): 1–51.

Maul GE. **1952**. Additions to previously revised families. *Boletim do Museu Municipal do Funchal* 6(art.16): 51–62.

Maul GE. **1954**. Monografia dos peixes do Museu Municipal do Funchal. Ordem Berycomorphi. Additions to previously revised families. *Boletim do Museu Municipal do Funchal* 7(art.17): 1–41.

Maul GE. **1954**. Additions to previously revised families. *Boletim do Museu Municipal do Funchal* 7(art.18): 41–63.

Maul GE. **1955**. Five species of rare sharks new for Madeira including two new to science. *Notulae Naturae* (Academy of Natural Sciences of Philadelphia) 279: 1–13.

Maul GE. **1986**. Berycidae (pp. 740–742). In: Whitehead PJP, Bauchot M-L, Hureau J-C, Nielsen J, Tortonese E (eds) *Fishes of the north-eastern Atlantic and the Mediterranean*. Vol. II. Paris: UNESCO.

May JL, Maxwell JGH. **1986**. *Trawl fish from temperate waters of Australia*. Tasmania: CSIRO Division of Fisheries Research. 492 pp.

Mayer GF. **1974**. A revision of the cardinalfish genus *Epigonus* (Perciformes, Apogonidae), with descriptions of two new species. *Bulletin of the Museum of Comparative Zoology* 146(3): 147–203.

Mayer GF, Tortonese E. **1977**. *Epigonus trewavasae* Poll., a junior synonym of *Epigonus constanciae* (Gigliolo) (Perciformes, Apogonidae). *Breviora* 443: 1–13.

McAllister DE, Randall JE. **1975**. A new species of centrolophid fish from Easter Island and Rapa Iti Island in the south Pacific. *National Museum of Canada Publications in Biology and Oceanography* 8: 1–7. McArdle AF. **1901**. Natural history notes from the Royal Indian marine survey ship *Investigator*, Commander T.H. Heming, R.N., Commanding. Series 3 No. 5. An account of the trawling operations during the surveying season of 1900–1901. *Annals and Magazine of Natural History* (Series 7) 8(48): 517–526.

McCann C, McKnight DG. **1980**. The marine fauna of New Zealand: macrourid fishes (Pisces: Gadidae). *New Zealand Oceanographic Institute Memoir* 61: 1–91.

McClain CR, Balk MA, Benfield MC, Branch TA, Chen C, Cosgrove J, Dove ADM, Gaskins L, Helm RR, Hochberg FG, Lee FB, Marshall A, McMurray SE, Schanche C, Stone SN, Thaler AD. 2015. Sizing ocean giants: patterns of intraspecific size variation in marine megafauna. *PeerJ* 3: e715. https://doi.org/10.7717/peerj.715

McClanahan TR. **1994**. Kenyan coral reef lagoon fish: effects of fishing, substrate complexity, and sea urchins. *Coral Reefs* 13: 231–241.

McClanahan TR. **1997**. Effects of fishing and reef structure on East Africa coral reefs (pp. 1533–1538). In: Lessios HA, Macintyre IG (eds) *Proceedings of the 8th International Coral Reef Symposium* Vol. 2. Panama: Smithsonian Tropical Research Institute.

McClanahan TR. **2011**. Coral reef fish communities in management systems with unregulated fishing and small fisheries closures compared with lightly fished reefs — Maldives vs. Kenya. *Aquatic Conservation: Marine and Freshwater Ecosystems* 21(2): 186–198. http://dx.doi. org/10.1002/aqc.1172

McClanahan TR, Ateweberhan M, Muhando C, Maina J, Mohammed SM. **2007**. Effects of climate and seawater temperature variation on coral bleaching and mortality. *Ecological Monographs* 77: 503–525.

McClanahan TR, Ateweberhan M, Omukoto J, Pearson L. 2009. Recent seawater temperature histories, status, and predictions for Madagascar's coral reefs. *Marine Ecology Progress Series* 380: 117–128. http://dx.doi.org/10.3354/ meps07879

McClanahan TR, Ateweberhan M, Sebastian CR, Graham NAJ, Wilson SK, Bruggemann H, Guillaume M. **2007**. Western Indian Ocean coral communities, bleaching responses, and susceptibility to extinction. *Marine Ecology Progress Series* 337: 1–13. http://dx.doi.org/10.3354/meps337001

McClanahan TR, Castilla JC, White AT, Defeo O. **2009**. Healing small-scale fisheries by facilitating complex socioecological systems. *Reviews in Fish Biology and Fisheries* 19(1): 33–47. http://dx.doi.org/10.1007/s11160-008-9088-8

McClanahan TR, Graham NAJ, Calnan J, MacNeil MA. 2007. Towards pristine biomass: reef fish recovery in coral reef marine protected areas in Kenya. *Ecological Applications* 17: 1055–1067.
McClanahan TR, Graham NAJ, MacNeil MA, Muthiga NA, Cinner JE, Bruggemann JH, Wilson SK. **2011**. Critical thresholds and tangible targets for ecosystem-based management of coral reef fisheries. *Proceedings of the National Academy of Sciences* 108(41): 17230–17233. http://dx.doi.org/10.1073/pnas.1106861108

McClanahan TR, Obura D. **1998**. *Monitoring, training and assessment of the coral reefs of the Masoala Peninsula*. Report by the Coral Reef Conservation Project, Wildlife Conservation Society. [unpaginated]

McClanahan TR, Shafir SH. **1990**. Causes and consequences of sea urchin abundance and diversity in Kenyan coral reef lagoons. *Oecologia* 83: 362–370.

McClanahan TR, Sheppard CRC, Obura DO (eds). **2000**. *Coral reefs of the Indian Ocean: Their ecology and conservation*. Oxford: Oxford University Press. xxiii + 525 pp.

McClelland J. **1839**. Indian Cyprinidae. *Asiatic Researches* 19(2): 217–471.

McClelland J. **1843**. On East Indian Isinglass, its introduction to, and manufacture for, the European market. *Calcutta Journal of Natural History* 3(10): 157–188.

McClelland J. **1844**. Apodal fishes of Bengal. *Calcutta Journal* of Natural History 5(18): 151–226.

McCosker JE. **1970**. A review of the eel genera *Leptenchelys* and *Muraenichthys*, with the description of a new genus, *Schismorhynchus*, and a new species, *Muraenichthys chilensis*. *Pacific Science* 24(4): 506–516.

McCosker JE. **1975**. The eel genus *Phaenomonas* (Pisces, Ophichthyidae). *Pacific Science* **29**(4): 361–363.

McCosker JE. **1977**. The osteology, classification and relationships of the eel family Ophichthidae. *Proceedings of the California Academy of Sciences* (Series 4) 41(1): 1–123.

McCosker JE. **1977**. Flashlight fishes. *Scientific American* 236(3): 106–114.

McCosker JE. **1978**. Synonomy and distribution of *Calloplesiops* (Pisces: Piesiopidae). *Copeia* 1978(4): 707–710.

McCosker JE. **1979**. Inferred natural history of the living coelacanth (pp. 17–24). In: McCosker JE, Lagios MD (eds) The biology and physiology of the living coelacanth. *Occasional Papers of the California Academy of Sciences* 134: 1–175.

McCosker JE. **1982**. A new genus and two new species of remarkable Pacific worm eels (Ophichthidae, subfamily Myrophinae). *Proceedings of the California Academy of Sciences* (Series 4) 43(5): 59–66.

McCosker JE. **1985**. Two new genera and two new species of deepwater western Atlantic worm eels (Pisces: Ophichthidae). *Proceedings of the California Academy of Sciences* (Series 4) 44(2): 9–15.

McCosker JE. **1986**. A new snake-eel, *Ophichthus bennettai*, (Pisces: Ophichthidae) from off western South Africa. *J.L.B. Smith Institute of Ichthyology Special Publication* **39**: 1–4. McCosker JE. **1998**. A revision of the snake eel genus *Callechelys* (Anguilliformes: Ophichthidae) with the description of two new Indo-Pacific species and a new callechelyin genus. *Proceedings of the California Academy of Sciences* (Series 4) 50(7): 185–214.

McCosker JE. **1998**. Snake-eels of the genus *Xyrias* (Anguilliformes: Ophichthidae). *Cybium* 22(1): 7–13.

McCosker JE. **1999**. Pisces Anguilliformes: deepwater snake eels (Ophichthidae) from the New Caledonia region, southwest Pacific Ocean. *Mémoires du Muséum National d'Histoire Naturelle* (Paris) (Série A, Zoologie) 180: 571–588.

McCosker JE. **2002**. Notes on Hawaiian snake eels (Pisces: Ophichthidae), with comments on *Ophichthus bonaparti*. *Pacific Science* 56(1): 23–34.

McCosker JE. **2004**. A new species of finless snake eel (Anguilliformes: Ophichthidae) from Ascension Island, with comments on *Ichthyapus acutirostris*. *Proceedings of the California Academy of Sciences* (Series 4) 55(7): 169–173.

McCosker JE. **2007**. *Luthulenchelys heemstraorum*, a new genus and species of snake eel (Anguilliformes: Ophichthidae) from KwaZulu-Natal, with comments on *Ophichthus rutidoderma* (Bleeker, 1853) and its synonyms. *Smithiana* Bulletin 7: 3–7.

McCosker JE. **2010**. Deepwater Indo-Pacific species of the snake-eel genus *Ophichthus* (Anguilliformes: Ophichthidae), with the description of nine new species. *Zootaxa* 2505: 1–39. http://dx.doi.org/10.11646/ zootaxa.2505.1.1

McCosker JE. **2011**. Two new Indo-Pacific species of the sand-eel genus *Yirrkala* (Anguilliformes: Ophichthidae). *Smithiana* Bulletin 13: 45–50.

McCosker JE, Baranes A, Golani D. **1993**. Description of the adult of *Leptocephalus echeloides* D'Ancona (1928), a deepwater snake eel, genus *Ophichthus* (Ophichthidae), from the Gulf of Aqaba. *Cybium* 17(2): 165–170.

McCosker JE, Böhlke EB. **1984**. Three new species of western Atlantic snake eels (Pisces: Ophichthidae) of the genus *Ophichthus. Proceedings of the Academy of Natural Sciences of Philadelphia* 136: 24–31.

McCosker JE, Böhlke EB, Böhlke JE. **1989**. Family Ophichthidae. Snake eels and worm eels. In: Fishes of the western North Atlantic. *Memoir of the Sears Foundation for Marine Research* 1(part 9): 254–412.

McCosker JE, Chen W-L, Chen H-M. **2009**. Comments on the snake-eel genus *Xyrias* (Anguilliformes: Ophichthidae) with the description of a new species. *Zootaxa* 2289: 61–67. https://doi.org/10.11646/zootaxa.2289.1.6

McCosker JE, Hatooka K, Ohnishi N, Endo H. **2011**. Redescription and designation of a neotype for *Aphthalmichthys kuro* Kuroda 1947, and its placement in *Callechelys* (Anguilliformes: Ophichthidae). *Ichthyological*  *Research* 58(3): 272–277. http://dx.doi.org/10.1007/s10228-011-0233-3

McCosker JE, Hatooka K, Sasaki K, Moyer JT. **1984**. Japanese moray eels of the genus *Uropterygius*. *Japanese Journal of Ichthyology* 31(3): 261–267.

McCosker JE, Hibino Y. **2015**. A review of the finless snake eels of the genus *Apterichtus* (Anguilliformes: Ophichthidae), with the description of five new species. *Zootaxa* 3941(1): 49–78. https://doi.org/10.11646/zootaxa.3941.1.2

McCosker JE, Lagios MD. **1975**. Les petits Peugeots of Grande Comore. *Pacific Discovery* 28(5): 1–6.

McCosker JE, Randall JE. **1977**. Three new species of Indo-Pacific moray eels (Pisces: Muraenidae). *Proceedings of the California Academy of Sciences* (Series 4) 41(3): 161–168.

McCosker JE, Randall JE. **1982**. Synonymies of Indian Ocean eels, with the description of *Gymnothorax enigmaticus*, a moray previously known as *G. ruppeli*. *Proceedings of the California Academy of Sciences* (Series 4) 43(2): 17–24.

McCosker JE, Randall JE. **2001**. A revision of the snake-eel genus *Brachysomophis* (Anguilliformes: Ophichthidae), with description of two new species and comments on the species of *Mystriophis*. *Indo-Pacific Fishes* 33: 1–32.

McCosker JE, Randall JE. **2002**. *Ophichthys melanochir* Bleeker, 1865, a junior synonym of the highfin snake eel *Ophichthus altipennis* (Kaup, 1856). *Copeia* 2002(3): 798–799.

McCosker JE, Rosenblatt RH. **2010**. The fishes of the Galápagos Archipelago: an update. *Proceedings of the California Academy of Sciences* (Series 4) 61(Suppl. 2): 167–195.

McCosker JE, Rosenblatt RH. **1987**. Notes on the biology, taxonomy, and distribution of flashlight fishes (Beryciformes: Anomalopidae). *Japanese Journal of Ichthyology* 34(2): 157–164.

McCosker JE, Rosenblatt RH. **1993**. A revision of the snake eel genus *Myrichthys* (Anguilliformes: Ophichthidae) with the description of a new eastern Pacific species. *Proceedings of the California Academy of Sciences* (Series 4) 48(8): 153–169.

McCosker JE, Smith DG. **1997**. Two new Indo-Pacific morays of the genus *Uropterygius* (Anguilliformes: Muraenidae). *Bulletin of Marine Science* 60(3): 1005–1014.

McCulloch AR. **1911**. Report of the fishes obtained by the F.I.S. *Endeavour* on the coasts of Queensland, New South Wales, Victoria, South Australia and Tasmania. Part I. *Biological Results Endeavour* 1(1): 1–87.

McCulloch AR. **1914**. Report on some fishes obtained by the F.I.S. *Endeavour* on the coasts of Queensland, New South Wales, Victoria, Tasmania, South and South-Western Australia. Part II. *Biological Results Endeavour* 2(3): 77–165.

McCulloch AR. **1915**. Report on some fishes obtained by the F.I.S. *Endeavour* on the coasts of Queensland, New South Wales, Victoria, Tasmania, South and South-Western Australia. Part III. *Biological Results Endeavour* 3(3): 97–170.

McCulloch AR. **1922**. Check list of the fish and fish-like animals of New South Wales. Part III. *Australian Zoologist* 2(3): 86–130.

McCulloch AR. **1923**. Notes on fishes from Australia and Lord Howe Island. *Records of the Australian Museum* 14(1): 1–17.

McCulloch AR. **1923**. Fishes from Australia and Lord Howe Island, No. 2. *Records of the Australian Museum* 14(2): 113–125.

McCulloch AR. **1929**. A check-list of the fishes recorded from Australia. Part 1. *Memoirs of the Australian Museum* (Sydney Memoir) 5: 1–144.

McCulloch AR, Waite ER. **1918**. Some new and little known fishes from South Australia. *Records of the South Australian Museum* (Adelaide) 1(1): 39–78.

McDowall RM. **1980**. First adults of *Schedophilus maculatus* Günther, 1860 (Stromateoidei, Centrolophidae). *Journal of the Royal Society of New Zealand* 10(2): 145–151.

McDowall RM. **1981**. A sub-dorsal fin pore/canal system in the centrolophid fish *Schedophilus maculatus* (Pisces: Stromateoidei). *Copeia* 1981(2): 492–494.

McDowall RM. **1982**. The centrolophid fishes of New Zealand (Pisces: Stromateoidei). *Journal of the Royal Society of New Zealand* 12(2): 103–142.

McDowall RM. 2001. Centrolophidae. Medusafishes (ruffs, barrelfishes) (pp. 3767–3770). In: Carpenter KE, Niem VH (eds) *FAO species identification guide for fishery purposes. The living marine resources of the western central Pacific.* Vol. 6. Bony fishes part 4 (pp. 3381–4218). Rome: FAO.

McDowall RM, Stewart AL. **1999**. Further specimens of *Agrostichthys parkeri* (Teleostei: Regalecidae), with natural history notes (pp. 165–174). In: Séret B, Sire J-Y (eds) *Proceedings of the 5th Indo-Pacific Fish Conference, Nouméa, New Caledonia, 3–8 November 1997*. Paris: Société Française d'Ichtyologie. 866 pp.

McDowell JR. **2002**. Genetic stock structure of the sailfish, Istiophorus platypterus, based on nuclear and mitochondrial DNA. PhD dissertation, Virginia Institute of Marine Science, College of William and Mary. 229 pp.

McDowell JR, Graves JE. **2008**. Population structure of striped marlin (*Kajikia audax*) in the Pacific Ocean based on analysis of microsatellite and mitochondrial DNA. *Canadian Journal of Fisheries and Aquatic Science* 65(7): 1307–1320. http://dx.doi.org/10.1139/F08-054

McDowell SB. **1973**. Suborder Notacanthoidei, Family Notacanthidae. In: Fishes of the western North Atlantic. *Memoir of the Sears Foundation for Marine Research* 1(part 6): 124–207.

McEachran JD, Aschliman N. **2004**. Phylogeny of Batoidea (pp. 79–113). In: Carrier JC, Musick JA, Heithaus MR (eds) *Biology of sharks and their relatives*. Boca Raton, Florida: CRC Press. 596 pp. McEachran JD, Compagno LJV. **1982**. Interrelationships of and within *Breviraja* based on anatomical structures (Pisces: Rajoidei). *Bulletin of Marine Science* 32(2): 399–425.

McEachran JD, Dunn KA. **1998**. Phylogenetic analysis of skates, a morphologically conservative clade of elasmobranchs (Chondrichthyes: Rajidae). *Copeia* 1998(2): 271–290.

McEachran JD, Fechhelm JD. **1982**. A new species of skate from the western Indian Ocean, with comments on the status of *Raja* (*Okamejei*) (Elasmobranchii: Rajiformes). *Proceedings of the Biological Society of Washington* 95(3): 440–450.

McEachran JD, Fechhelm JD. 2005. Fishes of the Gulf of Mexico. Vol. 2. Scorpaeniformes to Tetraodontiformes. Austin, Texas: University of Texas Press. viii + 1004 pp.

McEachran JD, Séret B, Miyake T. **1989**. Morphological variation within *Raja miraletus* and status of *R. ocellifera* (Chondrichthyes, Rajoidei). *Copeia* 1989(3): 629–641.

McEachran JD, Stehmann M. 1984. A new species of skate, *Neoraja carolinensis*, from off the southeastern United States (Elasmobranchii: Rajoidei). *Proceedings of the Biological Society of Washington* 97(4): 724–735.

McIlwain JL, Hermosa GV, Claereboudt M, Al-Oufi HS, Al-Awi M. **2006**. Spawning and reproductive patterns of six exploited finfish species from the Arabian Sea, Sultanate of Oman. *Journal of Applied Ichthyology* 22(2): 167–176.

McKay RJ. **1970**. Additions to the fish fauna of Western Australia. 5. *Western Australian Fisheries Bulletin* 9(5): 3–24.

McKay RJ. **1971**. Two new genera and five new species of percophidid fishes (Pisces: Percophididae) from Western Australia. *Journal of the Royal Society of Western Australia* 54(2): 40–46.

McKay RJ. **1980**. The fishes of the family Sillaginidae from India with a description of a new species. *Journal of the Marine Biological Association of India* 18(2): 375–385.

McKay RJ. **1984**. Haemulidae. Grunts, sweetlips, rubberlips, hotlips. In: Fischer W, Bianchi G (eds) *FAO species identification sheets for fishery purposes. Western Indian Ocean (Fishing Area 51)*. Vol. 2. Rome: FAO. [unpaginated]

McKay RJ. **1985**. A revision of the fishes of the family Sillaginidae. *Memoirs of the Queensland Museum* 22(1): 1–73.

McKay RJ. **1992**. FAO species catalogue. Vol. 14. Sillaginid fishes of the world (family Sillaginidae): an annotated and illustrated catalogue of the sillago, smelt or Indo-Pacific whiting species known to date. *FAO Fisheries Synopsis No. 125* (Vol. 14). Rome: FAO. 87 pp.

McKay RJ. 1999. Sillaginidae. Sillagos (pp. 2614–2629). In: Carpenter KE, Niem VH (eds) FAO species identification guide for fisheries purposes. The living marine resources of the western central Pacific. Vol. 4. Bony fishes part 2 (pp. 2069– 2790). Rome: FAO. McKay RJ. 2001. Haemulidae. Grunts (also sweetlips, rubberlips, hotlips, and velvetchins) (pp. 2961–2989). In: Carpenter KE, Niem VH (eds) *FAO species identification guide for fishery purposes. The living marine resources of the western central Pacific.* Vol. 5. Bony fishes part 3 (pp. 2791–3380). Vol. 5. Rome: FAO.

McKay RJ, McCarthy J. **1989**. A revision of the sillaginid fishes of the Arabian Gulf with a description of *Sillago arabica* new species. *Memoirs of the Queensland Museum* 272: 551–553.

McKay RJ, Randall JE. **1995**. Two new species of *Pomadasys* (Pisces: Haemulidae) from Oman, with a redescription of *P. punctulatus* (Rüppell). *Memoirs of the Queensland Museum* 38(1): 251–255.

McKenna SA, Allen GR (eds). 2005. A rapid marine biodiversity assessment of the coral reefs of northwest Madagascar. *Bulletin of the Rapid Assessment Program* 31. Washington, DC: Conservation International. 124 pp. [also cited as 2003]

McKenna SA, Allen GR. **2005**. List of the reef fishes of Madagascar (Appendix 4, pp. 102–124). In: McKenna SA, Allen GR (eds) A rapid marine biodiversity assessment of the coral reefs of northwest Madagascar. *Bulletin of the Rapid Assessment Program* 31. Washington, DC: Conservation International. 124 pp.

McKinney JF, Springer VG. **1976**. Four new species of the fish genus *Ecsenius* with notes on other species of the genus (Blenniidae: Salariini). *Smithsonian Contributions to Zoology* 236: 1–27.

McPhie RP, Campana SE. **2009**. Bomb dating and age determination of skates (family Rajidae) off the eastern coast of Canada. *ICES Journal of Marine Science* 66: 546–560. https://doi.org/10.1093/icesjms/fsp002

Mead GW. **1957**. On the bramid fishes of the Gulf of Mexico. *Zoologica* (N.Y.) 42: 51–61.

Mead GW. **1966**. Families Bathysauridae, Bathypteroidae, Ipnopidae, Chlorophthalmidae. In: Fishes of the western North Atlantic. *Memoir of the Sears Foundation for Marine Research* 1(part 5): 103–189.

Mead GW. 1972. Bramidae. Dana Report 81: 1-166.

Mead GW, De Falla JE. **1965**. New oceanic cheilodipterid fishes from the Indian Ocean. *Bulletin of the Museum of Comparative Zoology* 134(7): 261–274.

Mead GW, Maul GE. **1958**. *Taractes asper* and the systematic relationships of the Steinegeriidae and Trachyberycidae. *Bulletin of the Museum of Comparative Zoology* 119(6): 393–417.

Meadows MG, Anthes N, Dangelmayer S, Alwany MA, Gerlach T, Schulte G, Sprenger D, Theobald J, Michiels NK. **2014**. Red fluorescence increases with depth in reef fishes, supporting a visual function, not UV protection. *Proceedings of the Royal Society* B 281(1790): 1–10. https://doi.org/10.1098/rspb.2014.1211 Mecklenburg CW, Mecklenburg TA, Thorsteinson TK. **2002**. *Fishes of Alaska*. Bethesda, Maryland: American Fisheries Society. xxxvii + 1037 pp.

Medio D, Sheppard CRC, Gascoigne J. **2000**. Chapter 8. The Red Sea (pp. 231–255). In: McClanahan TR, Sheppard CRC, Obura DO (eds) *Coral reefs of the Indian Ocean: Their ecology and conservation*. Oxford: Oxford University Press. xxiii + 525 pp.

Mee JKL, Hare SR. **1995**. *Coris nigrotaenia*, a new wrasse (Perciformes: Labridae) from the northwest Indian Ocean. *Journal of South Asian Natural History* 1(2): 247–254.

Meek SE. 1897. List of fishes and reptiles obtained by Field Columbian Museum East African expedition to Somaliland in 1896. *Field Columbian Museum* (Zoölogical Series) 1(8) (Publ. 22): 165–184, Pl. 40.

Meek SE, Hildebrand SF. **1923**. The marine fishes of Panama. Part I. *Field Museum of Natural History* (Zoölogical Series) 15 (Publ. 215): i–xi + 1–330, Pls. 1–24.

Mees GF. **1962**. A preliminary revision of the Belonidae. *Zoologische Verhandelingen* (Leiden) 54: 1–96.

Mees GF. **1964**. Further revisional notes on the Belonidae. *Zoologische Mededelingen* (Leiden) 39: 311–326.

Meisler MR. **1987**. Limits and relationships of serranine seabasses, with revisions of *Serranus* and *Mentiperca* (Pisces: Serranidae). PhD dissertation, University of Southern California, Los Angeles, California.

Meisner AD. **2001**. Phylogenetic systematics of the viviparous halfbeak genera *Dermogenys* and *Nomorhamphus* (Teleostei: Hemiramphidae: Zenarchopterinae). *Zoological Journal of the Linnean Society* 133(2): 199–283.

Meisner AD, Collette BB. 1999. Generic relationships of the internally-fertilized southeast Asian halfbeaks (Hemiramphidae: Zenarchopterinae) (pp. 69–76). In: Séret B, Sire J-Y (eds) Proceedings of the 5<sup>th</sup> Indo-Pacific Fish Conference, Nouméa, New Caledonia, 3–8 November 1997. Paris: Société Française d'Ichtyologie. 866 pp.

Meister HS, Wyanski DM, Loefer JK, Ross SW, Quattrini AM, Sulak KJ. **2005**. Further evidence for the invasion and establishment of *Pterois volitans* (Teleostei: Scorpaenidae) along the Atlantic coast of the United States. *Southeastern Naturalist* 4(2): 193–206.

Mellin C, Ferraris J, Galzin R, Kulbicki M, Ponton D. **2006**. Diversity of coral reef fish assemblages: modelling of the species richness spectra from multi-scale environmental variables in the Tuamotu Archipelago (French Polynesia). *Ecological Modelling* 198: 409–425.

Melnikov YS. **1980**. Feeding peculiarities of *Allocyttus verrucosus*, family Oreosomatidae. *Journal of Ichthyology* 20(3): 99–105.

Mendis AS. **1954**. Fishes of Ceylon: a catalogue, key and bibliography. *Bulletin of the Fisheries Research Station, Ceylon* 2: 1–222.

Menni RC, Cousseau MB. **1981**. *Pentaceros richardsoni* (A. Smith, 1844) en la provincia zoogeográfica Argentina (Osteichthys, Pentacerotidae). *Physis* (Buenos Aires) Sección A 39(97): 77–78.

Menni RC, Miquelarena AM. **1979**. Dimorfismo sexual y status de *Centriscops obliquus maculatus* Pozzi y Bordale. 1936 (Osteichthyes, Macrorhamphosidae). *Acta Zoologica Lilloana* 35: 573–585.

Menon AGK. **1961**. On a collection of fish from the Coromandel coast of India including Pondicherry and Karaikkal areas. *Records of the Indian Museum* (Calcutta) 59(4): 369–404.

Menon AGK. **1963**. Taxonomy of the Indian frog-fishes (Family Batrachoididae). *Labdev Journal of Science and Technology* (Kanpur) 1. [2 pages, not numbered]

Menon AGK. **1977**. A systematic monograph of the tongue soles of the genus *Cynoglossus* Hamilton-Buchanan (Pisces: Cynoglossidae). *Smithsonian Contributions to Zoology* 238: 1–129.

Menon AGK. **1980**. A revision of the fringe-lip tongue soles of the genus *Paraplagusia* Bleeker, 1865 (Family Cynoglossidae). *Matsya* 59 [for 1979]: 11–22.

Menon AGK. **1984**. Soleidae. Soles. In Fischer W, Bianchi G (eds) *FAO species identification sheets for fishery purposes. Western Indian Ocean (Fishing Area 51)*. Vol. 4. Rome: FAO. [unpaginated]

Menon AGK. **1999**. Check list – fresh water fishes of India. *Records of the Zoological Survey of India, Occasional Paper* 175: xxviii + 366 pp.

Menon AGK, Rama-Rao KV. **1967**. Occurrence of flying fish, *Hirundichthys speculiger* (Val.), first in the Indian seas. *Bulletin of the National Institute of Sciences of India* 38: 767–770.

Menon AGK, Rama-Rao KV. **1970**. Type-specimens of fishes described in the R.I.M.S. *Investigator* collections (1884–1926). *Copeia* 1970(2): 377–378.

Menon AGK, Rama-Rao KV. **1975**. A catalogue of typespecimens of fishes described in the biological collections of the R.I.M.S. *Investigator* during 1884–1926. *Matsya* 1: 31–48.

Menon AGK, Yazdani GM. **1968**. Catalogue of type-specimens in the Zoological Survey of India. Part 2. Fishes. *Records of the Zoological Survey of India* 61(1–2): 91–190.

Menzies A. **1791**. Description of a new fish (*Echeneis lineata*) found in the Pacific Ocean. *Transactions of the Linnean Society of London* 1(art.21): 187–188.

MEP (Macalister Elliott & Partners Ltd.). 1999. Republic of Yemen: Fisheries Sector Review. Fourth Fisheries Development Project. MEP-YE-108. Report to the European Commission (ALA/91/22) and to the Ministry of Fish Wealth, Government of the Republic of Yemen. Merrett NR. **1968**. *Lepidocybium flavo-brunneum* (Smith, 1849) (Gempylidae) from the western Indian Ocean. *Journal of Natural History* 2(2): 201–204.

Merrett NR. **1973**. A new shark of the genus *Squalus* (Squalidae: Squaloidea) from the equatorial western Indian Ocean; with notes on *Squalus blainvillei*. *Journal of Zoology* (London) 171(1): 93–110.

Metzelaar J. **1919**. *Report on the fishes, collected by Dr. J. Boeke in the Dutch West Indies 1904–1905, with comparative notes on marine fishes of tropical West Africa*. Gravenhage: F.J. Belinfante. 315 pp.

Meuschen FC. **1781**. In: Gronovius LT (ed) *Zoophylacii Gronoviani: Fasciculus Primus Exhibens, Animalia Quadrepeda, Amphibia atque Pisces*. 2 Parts. Lugduni Batavorum. 236 pp.

Meyer-Rochow VB. **1977**. Comparison between 15 *Carapus mourlani* in a single holothurian and 19 *C. mourlani* from starfish. *Copeia* 1977(3): 582–584.

Meynard CN, Mouillot D, Mouquet N, Douzery EJP. **2012**. A phylogenetic perspective on the evolution of Mediterranean teleost fishes. *PLoS ONE* 7(5): e36443 (10 pp). https://doi. org/10.1371/journal.pone.0036443

Michael SW. **1993**. *Reef sharks and rays of the world: A guide to their identification, behavior, and ecology*. Monterey, California: Sea Challengers. 107 pp.

Michael SW. **2001**. *Reef fishes: A guide to their identification, behavior, and captive care.* Vol. 1. Neptune City, New Jersey: TFH-Microcosm. 624 pp.

Michel C. **1996**. *Poissons de l'île Maurice*. Stanley, Rose-Hill, Mauritius: Editions de l'Océan Indien. 137 pp.

Michelin H. 1849. Essai d'une faune de l'lle Maurice publiée avec les matériaux et les notes laissés par Julien Desjardins sous la direction et par les soins de M.F.E. Guérin Méneville. Zoophytes, Echinodermes et Stellérides. *Magazine de Zoologie* (2), VII, 1845: 1–27, Pls. 7–12.

Michiels NK, Anthes N, Hart NS, Herler J, Meixner AJ, Schleifenbaum F, Schulte G, Siebeck UE, Sprenger D, Wucherer MF. 2008. Red fluorescence in reef fish: a novel signalling mechanism? *BMC Ecology* 8(16). https://doi.org/10.1186/1472-6785-8-16

Mihara E, Amaoka K. 2004. Pleuronectiform fishes from New Caledonian waters. Five species of the samarid genus *Plagiopsetta* and *Samaris* (Samaridae). In: Marshall BA, Richer de Forges B (eds) Tropical deep-sea benthos. Vol. 23. *Mémoires du Muséum National d'Histoire Naturelle* (Paris) (N.S., Série A, Zoologie) 191: 611–635.

Mikhaylin SV. **1976**. Characteristics of the distribution of some members of the families Gempylidae and Trichiuridae in the waters of Southwest Africa. *Journal of Ichthyology* 16(2): 319–323.

Mikhaylin SV. **1983**. On intraspecific variations of the principal meristic characters in *Diplospinus multistriatus* (Gempylidae, Perciformes). *Journal of Ichthyology* 23(3): 13–19.

Millard NAH, Broekhuysen GJ. **1970**. The ecology of South African estuaries part X. St. Lucia: a second report. *Zoologica Africana* 5(2): 277–307.

Miller GC. **1974**. Fische des Indischen Ozeans. Ergebnisse der ichthyologischen Untersuchungen während der Expedition des Forschungsschiffes *Meteor* in den Indischen Ozean, Oktober 1964 bis Mai 1965. A. Systematischer Teil 14. Scorpaeniformes (2) Family Peristediidae. *Meteor Forschungsergebnisse* (Reihe D, Biologie) 18: 61–72.

Miller GC, Richards WJ. 2002. Peristediidae. Armoured searobins (armoured gurnards) (pp. 1278–1285). In: Carpenter KE (ed) FAO species identification guide for fishery purposes. The living marine resources of the western central Atlantic. Vol. 2. Bony fishes part 1 (pp. 601–1374). Rome: FAO.

Miller MJ, Tsukamoto K. **2004**. *An introduction to Leptocephali biology and identification*. Tokyo: Ocean Research Institute, University of Tokyo. 96 pp.

Miller PJ. **1978**. The systematic position and origin of *Gobius ocheticus* Norman, 1927, from the Suez Canal. *Zoological Journal of the Linnean Society* 62(1): 39–58.

Miller PJ. **1986**. Gobiidae (pp. 1019–1085). In: Whitehead PJP, Bauchot M-L, Hureau J-C, Nielsen J, Tortonese E (eds) *Fishes of the North-eastern Atlantic and the Mediterranean*. Vol. III. Paris: UNESCO.

Miller PJ. **1988**. Studies on *Silhouettea* Smith 1959 and an account of *Ebomegobius* Herre 1946 (Pisces: Gobiidae). *Senckenbergiana Biologica* 68(4/6): 241–273.

Miller PJ. 1990. Gobiidae (pp. 925–951). In: Quéro J-C,
Hureau J-C, Karrer K, Post A, Saldanha L (eds) *Check-list* of the fishes of the eastern tropical Atlantic (CLOFETA).
Vol. 2. Lisbon: Junta Nacional de Investigação Científica e Tecnológica.

Miller PJ, Fouda MM. **1986**. Notes on the biology of a Red Sea goby, *Silhouettea aegyptia* (Chabanaud, 1933) (Teleostei: Gobiidae). *Cybium* 10(4): 395–409.

Miller PJ, Wongrat P. **1990**. Eleotridae (pp. 952–957). In: Quéro J-C, Hureau J-C, Karrer C, Post A, Saldanha L (eds) *Check-list of the fishes of the eastern tropical Atlantic* (*CLOFETA*). Vol. 2. Lisbon: Junta Nacional de Investigação Científica e Tecnológica.

Miller TL, Cribb TH. **2007**. Phylogenetic relationships of some common Indo-Pacific snappers (Perciformes: Lutjanidae) based on mitochondrial DNA sequences, with comments on the taxonomic position of the Caesioninae. *Molecular Phylogenetics and Evolution* 44(1): 450–460. Millner R, Walsh SJ, Díaz de Astarloa JM. **2005**. Atlantic flatfish fisheries (pp. 240–271). In: Gibson RN (ed) *Flatfishes: Biology and exploitation*. Oxford: Blackwell Science. xxiv + 391 pp.

Millot J. **1953**. "Notre" Coelacanth. *Revue Madagascar* (17): 18–20.

Millot J, Anthony J. **1958**. *Anatomie de Latimeria chalumnae*. *Tome 1, Squellete, Muscles et Formations de soutien*. Paris. 122 pp.

Mimura K et al. 1963. Synopsis of biological data on yellowfin tuna Neothunnus macropterus Temminck and Schlegel 1842 (Indian Ocean). Species synopsis No. 10. FAO Fisheries Biology Synopsis 53: 319–349. [Prepared by K Mimura and staff of the NRFRL, Japan]

Mimura K *et al.* **1963**. Synopsis on the biology of bigeye tuna *Parathunnus mebachi* Kishinouye 1923 (Indian Ocean). Species synopsis No. 11. *FAO Fisheries Biology Synopsis* 54: 350–379. [Prepared by K Mimura and staff of the NRFRL, Japan]

Mincarone MM. 2017. Redescription of the hagfishes (Myxinidae: *Eptatretus*) from southern Africa. *Marine Biology Research* 13(7): 797–810. https://doi.org/10.1080/17 451000.2017.1287924

Mincarone MM, Bernardes RA, Peppes FV. **2007**. Occurrence of *Pleuroscopus pseudodorsalis* Barnard, 1927 (Uranoscopidae) near Rio Grande Plateau, western South Atlantic. *Journal of Fish Biology* 71(4): 1238–1240.

Mincarone MM, McCosker JE. **2004**. *Eptatretus lakeside* sp. nov., a new species of five-gilled hagfish (Myxinidae) from the Galápagos Islands. *Proceedings of the California Academy of Sciences* (Series 4) 55(6): 162–168.

Mincarone MM, Mwale M, Fernholm B. 2011. First record and further description of the Cape hagfish *Myxine capensis* (Myxinidae) off Mozambique, western Indian Ocean. *Journal of Fish Biology* 79(3): 806–811. http://dx.doi. org/10.1111/j.1095-8649.2011.03063.x

Mincarone MM, Stewart AL. **2006**. A new species of giant seven-gilled hagfish (Myxinidae: *Eptatretus*) from New Zealand. *Copeia* 2006(2): 225–229.

Ministry of Agriculture. **2001**. *Revalidation of potential yield estimates in the Indian EEZ*. Report of Ministry of Agriculture, Government of India. 73 pp.

Miranda Ribeiro A de, see De Miranda Ribeiro A.

Mishra SS, Biswas S, Russell BC, Satpathy KK, Selvanayagam M. **2013**. A new species of the genus *Scolopsis* Cuvier, 1830 (Perciformes: Nemipteridae) from southern India and Sri Lanka. *Zootaxa* 3609(4): 443–449. https://doi.org/10.11646/ zootaxa.3609.4.7

Mishra SS, Krishnan S. **2003**. Marine fishes of Pondichery and Karaikal. *Records of the Zoological Survey of India*, *Occasional Paper* 216: 1–53. Mishra SS, Srinivasan, M. **1999**. On a collection of fish from Cannanore-Mangalore sector of the west coast of India. *Records of the Zoological Survey of India* 97(2): 233–257.

Misra KS. **1952**. An aid to the identification of fishes of India, Burma and Ceylon. I. Elasmobranchii and Holocephali. *Records of the Indian Museum* (Calcutta) 49(1): 89–137.

Misra KS. **1962**. An aid to the identification of the common commercial fishes of India and Pakistan. *Records of the Indian Museum* (Calcutta) 57(1–4) [for 1959]: 1–320.

Misra KS. **1969**. Pisces: Elasmobranchii and Holocephali. In: Roonwal ML (ed) *The fauna of India and adjacent countries* (2<sup>nd</sup> edition, Vol. 1). Faridabad: Zoological Survey of India. 276 pp.

Misra KS. **1976**. Teleostomi: Cypriniformes; Siluri. In: *The fauna of India and the adjacent countries. Pisces* (2<sup>nd</sup> edition, Vol. 3). Calcutta: Zoological Survey of India. xxi + 367 pp.

Mitchill SL. **1814**. *Report, in part, of Samuel L. Mitchill, M. D., Professor of Natural History, on the fishes of New-York.* New York: D. Carlisle. 28 pp.

Mitchill SL. **1815**. The fishes of New-York, described and arranged. *Transactions of the Literary and Philosophical Society of New-York* 1(art.5): 355–492.

Mitchill SL. **1818**. Memoir on ichthyology. The fishes of New York, described and arranged. In a supplement to the Memoir... *American Monthly Magazine and Critical Review* 2(4, art.1): 241–248.

Mito S. **1955**. Breeding habits of a percoid fish, *Plesiops* semeion. Science Bulletin of the Faculty of Agriculture, *Kyushu University* 15: 95–99.

Miya M, Friedman M, Satoh TP, Takeshima H, Sado T, Iwasaki W, Yamanoue Y, Nakatani M, Mabuchi K, Inoue JG, Poulsen JY, Fukunaga T, Sato Y, Nishida M. **2013**. Evolutionary origin of the Scombridae (tunas and mackerels): members of a Paleogene adaptive radiation with 14 other pelagic fish families. *PLoS ONE* 8(9): e73535. https://doi.org/10.1371/journal.pone.0073535

Miya M, Kawaguchi A, Nishida M. **2001**. Mitogenomic exploration of higher teleostean phylogenies: a case study for moderate-scale evolutionary genomics with 38 newly determined complete mitochondrial DNA sequences. *Molecular Biology and Evolution* 18(11): 1993–2009.

Miya M, Nishida M. **2015**. The mitogenomic contributions to molecular phylogenetics and evolution of fishes: a 15-year retrospect. *Ichthyological Research* 62: 29–71. https://doi.org/10.1007/s10228-014-0440-9

Miya M, Saitoh K, Wood R, Nishida M, Mayden RL.
2006. New primers for amplifying and sequencing the mitochondrial ND4/ND5 gene region of the Cypriniformes (Actinopterygii: Ostariophysi). *Ichthyological Research* 53(1): 75–81.

Miya M, Satoh TP, Nishida M. **2005**. The phylogenetic position of toadfishes (order Batrachoidiformes) in the higher rayfinned fish as inferred from partitioned Bayesian analysis of 102 whole mitochondrial genome sequences. *Biological Journal of the Linnean Society* 85(3): 289–306.

Miya M, Takeshima H, Endo H, Ishiguro NB, Inoue JG, Mukai T, Satoh TP, Yamaguchi M, Kawaguchi A, Mabuchi K, Shirai SM, Nishida M. **2003**. Major patterns of higher teleostean phylogenies: a new perspective based on 100 complete mitochondrial DNA sequences. *Molecular Phylogenetics and Evolution* 26(1): 121–138.

Miyazaki E, Sasaki K, Mitani T, Ishida M, Uehara S. 2004.
The occurrence of two species of *Macroramphosus* (Gasterosteiformes: Macroramphosidae) in Japan: morphological and ecological observations on larvae, juveniles, and adults. *Ichthyological Research* 51(3): 256–262.

Miyosi Y. **1939**. Description of three new species of elasmobranoiate fishes collected at Hyuga Nada, Japan. *Bulletin of the Biogeographical Society of Japan* 9(5): 91–97.

Möbius K. **1880**. Beiträge zur Meeresfauna der Insel Mauritius und der Seychellen, bearbeitet von K. Möbius, F. Richters und E. von Martens nach Sammlungen, angelegt auf einer Reise nach Mauritius. Berlin: Gutmann. vi + 352 pp. [In German]

Mochizuki K. **1979**. A new percichthyid fish, *Neoscombrops pacificus*, from Japan, with a redescription of *N. annectens* from South Africa. *Japanese Journal of Ichthyology* 26(3): 247–252.

Mochizuki K, Hayashi M. **1989**. Revision of the leiognathid fishes of the genus *Secutor*, with two new species. *Science Report of the Yokosuka City Museum* 37: 83–95.

Mohamed KH. **1958**. On the occurrence of the eel *Neenchelys buitendijki* Weber & de Beaufort in Indian waters. *Journal of the Bombay Natural History Society* 55(3): 511–517.

Mohr E. **1937**. Revision der Centriscidae (Acanthopterygii, Centrisciformes). *Dana Report* 13: 1–70.

Mohsen S. 2002. A preliminary survey of the status of demersal fish species in the Socotra Archipelago (pp. 399–409). In: Apel M, Hariri K, Krupp F (eds) Conservation and sustainable use of biodiversity of Socotra Archipelago. Marine habitat, biodiversity and fisheries surveys and management. Final Report of Phase III. Senckenberg Research Institute, Frankfurt a.M., Germany.

Mohsin AKM, Ambak MA. **1996**. *Marine fishes and fisheries of Malaysia and neighbouring countries*. Kuala Lumpur: Ampang Press. 744 pp.

Moi RD, Johnson GD. **1997**. Dismantling the Trachinoidei: evidence of a scorpaenoid relationship for the

Champsodontidae. *Ichthyological Research* 44(2): 143–176. Mok H-K. **1978**. Scale-feeding in *Tydemania navigatoris* (Pisces: Triacanthodidae). *Copeia* 1978(2): 338–340. Mok H-K. **1988**. Osteological evidence for the monophyly of Cepolidae and Owstoniidae. *Japanese Journal of Ichthyology* 34(4): 507–508.

Møller PR, Jones WJ. 2007. Eptatretus strickrotti n. sp. (Myxinidae): first hagfish captured from a hydrothermal vent. Biological Bulletin 212(1): 55–66.

Møller PR, Knudsen SW, Schwarzhans W, Nielsen JG. **2016**. A new classification of viviparous brotulas (Bythitidae) – with family status for Dinematichthyidae – based on molecular, morphological and fossil data. *Molecular Phylogenetics and Evolution* 100: 391–408.

http://dx.doi.org/10.1016/j.ympev.2016.04.008

Møller PR, Schwarzhans W. **2006**. Review of the Dinematichthyini (Teleostei, Bythitidae) of the Indo-west Pacific. Part II. *Dermatopsis, Dermatopsoides* and *Dipulus* with description of six new species. *The Beagle, Records of the Museums and Art Galleries of the Northern Territory* 22: 39–76.

Møller PR, Schwarzhans W. **2008**. Review of the Dinematichthyini (Teleostei: Bythitidae) of the Indo-west Pacific. Part IV. *Dinematichthys* and two new genera with descriptions of nine new species. *The Beagle, Records of the Museums and Art Galleries of the Northern Territory* 24: 87–146. http://dx.doi.org/10.5962/p.287440

Molteno CJ. **1948**. *The South African tunas: a preliminary study of the economic potentialities*. Cape Town: South African Fishing Industry Research Institute. 34 pp.

Monkolprasit S, Roberts TR. **1990**. *Himantura chaophraya*, a new giant freshwater stingray from Thailand. *Japanese Journal of Ichthyology* 37(3): 203–208.

Montague G. **1811**. An account of five rare species of British fishes. *Memoirs of the Wernerian Natural History Society* 1: 79–101.

Montilla JR. **1935**. A review of the Philippine Menidae and Gerreidae. *Philippine Journal of Science* 58: 281–295.

Mooi RD. **1990**. Egg surface morphology of pseudochromoids (Perciformes: Percoidei), with comments on its phylogenetic implication. *Copeia* 1990(2): 455–475.

Mooi RD. **1993**. Phylogeny of the Plesiopidae (Pisces: Perciformes) with evidence for the inclusion of the Acanthoclinidae. *Bulletin of Marine Science* 52(1): 284–326.

 Mooi RD. 1995. Revision, phylogeny, and discussion of biology and biogeography of the fish genus *Plesiops* (Perciformes: Plesiopidae). *Royal Ontario Museum Life Science Contributions* 159: 1–107.

Mooi RD. **1999**. Families Plesiopidae and Notograptidae (pp. 2578–2587). In: Carpenter KE, Niem VH (eds) *FAO species identification guide for fisheries purposes. The living marine resources of the western central Pacific.* Vol 4. Bony fishes part 2 (pp. 2069–2790). Rome: FAO. Mooi RD, Gill AC. **2004**. Description of a new species of the fish genus *Acanthoplesiops* Regan (Teleostei: Plesiopidae: Acanthoclininae) from Tonga. *Zootaxa* 432: 1–10.

Mooi RD, Gill AC. **2004**. Notograptidae, sister to *Acanthoplesiops* Regan (Teleostei: Plesiopidae: Acanthoclininae), with comments on biogeography, diet and morphological convergence with Congrogadinae (Teleostei: Pseudochromidae). *Zoological Journal of the Linnean Society* 141: 179–205.

Mooi RD, Randall JE. **2014**. *Pempheris bexillon*, a new species of sweeper (Teleostei: Pempheridae) from the Western Indian Ocean. *Zootaxa* 3780(2): 388–398. https://doi.org/10.11646/zootaxa.3780.2.10

Moore ABM, Almojil D, Harris M, Jabado RW, White WT. 2013. New biological data on the rare, threatened shark *Carcharhinus leiodon* (Carcharhinidae) from the Persian Gulf and Arabian Sea. *Marine and Freshwater Research* 65(4): 327–332. http://dx.doi.org/10.1071/MF13160

Moore ABM, Compagno LJV, Fergusson IK. **2007**. The Persian/Arabian Gulf's sole great white shark *Carcharodon carcharias* (Lamniformes: Lamnidae) record from Kuwait: misidentification of a sandtiger shark *Carcharias taurus* (Lamniformes: Odontaspididae). *Zootaxa* 1591: 67–68.

Moore ABM, White WT, Ward RD, Naylor GJP, Peirce R. 2011. Rediscovery and redescription of the smoothtooth blacktip shark *Carcharhinus leiodon* (Carcharhinidae), from Kuwait, with notes on its possible conservation status. *Marine and Freshwater Research* 62(6): 528–539. http://dx.doi.org/10.1071/MF10159

Moreau E. **1881**. *Histoire naturelle des poissons de la France*. Vol. 2. Paris: G Masson. 572 pp.

Morgan GR. **1983**. A preliminary multi-species assessment of Kuwait's gargoor (fish trap) fishery. *Annual Research Report, Kuwait Institute for Scientific Research* 8: 65–66.

Morgan G[R]. **2004**. Country review: India (West Coast). In: De Young C (ed) *Review of the state of world capture fisheries management: Indian Ocean*. FAO Fisheries Technical Paper 488. Rome: FAO, 2006. 458 pp.

Morgan SK, Panes HZ. **2008**. Threatened fishes of the world: *Hippocamus spinosissimus* Weber 1913 (Syngnathidae). *Environmental Biology of Fishes* 82(1): 21–22. http://dx.doi. org/10.1007/s10641-007-9250-5

Morgans JFC. **1959**. Three confusing species of serranid fish, one described as new, from East Africa. *Annals and Magazine of Natural History* (Series 13) 1(10) [for 1958]: 642–656.

Morgans JFC. **1964**. A preliminary survey of bottom fishing on the North Kenya Banks. London: HMSO. iii + 91 pp.

Morgans JFC. **1966**. East African fishes of the *Epinephelus tauvina* complex, with a description of a new species. *Annals and Magazine of Natural History* (Series 13) 8(89) [for 1965]: 257–271.

Morgans JFC. **1982**. Serranid fishes of Tanzania and Kenya. *Ichthyological Bulletin of the J.L.B. Smith Institute of Ichthyology* 46: 1–44.

Morón J, Bertrand B, Last P. **1998**. A check-list of sharks and rays of western Sri Lanka. *Journal of the Marine Biological Association of India* 40(1&2): 142–157.

Morris JA Jr, Freshwater DW. **2008**. Phenotypic variation of lionfish supraocular tentacles. *Environmental Biology of Fishes* 83(2): 237–241. http://dx.doi.org/10.1007/s10641-007-9326-2

Morris JA Jr, Whitfield PE. **2009**. Biology, ecology, control and management of the invasive Indo-Pacific lionfish: An updated integrated assessment. *NOAA Technical Memorandum* NOS NCCOS 99: 1–65.

Morris M. **2002**. Manual of traditional land use in the Soqotra Archipelago. GEF/YEM/96/G32. Royal Botanic Garden, Edinburgh. 540 pp. [unpublished but authoritative; widely used manuscript]

Morris RE. **1974**. A new labrid fish from the North Kenya Banks. *Copeia* 1974(3): 632–634.

Morrow JE. **1954**. Fishes from East Africa with new records and descriptions of two new species. *Annals and Magazine of Natural History* (Series 12) 7(83): 797–820.

Morrow JE. **1964**. Marlins, sailfish and spearfish of the Indian Ocean. In: *Proceedings of the Symposium on Scombroid Fishes*, Marine Biological Association of India (MBAI), 12–15 January 1962, Mandapam. Part 1: 429–440.

Morrow JE, Harbo SJ. **1969**. A revision of the sailfish genus *Istiophorus. Copeia* 1969(1): 34–44.

Moser HG, Richards WJ, Cohen DM, Fahay MP, Kendall SW Jr, Richardson SL (eds). **1984**. Ontogeny and systematics of fishes. *American Society of Ichthyologists and Herpetologists Special Publication* 1: x + 760 pp.

Moteki M, Fujita K, Last P. **1995**. *Brama pauciradiata*, a new bramid fish from the seas off tropical Australia and the central Pacific Ocean. *Japanese Journal of Ichthyology* 41(4): 421–427.

Motomura H. **2002**. Revision of the Indo-Pacific threadfin genus *Polydactylus* (Perciformes: Polynemidae) with a key to the species. *Bulletin of the National Science Museum* (Tokyo) (Series A) 28(3): 171–194.

Motomura H. **2004**. Morphological comparison of a poorly known scorpionfish, *Parapterois macrura*, with a related species, *P. heterura* (Scorpaenidae: Pteroinae). *Zoological Studies* 43(1): 1–7.

Motomura H. **2009**. *Sebastapistes taeniophrys* (Fowler 1943): a valid scorpionfish (Scorpaenidae) from the Philippines. *Ichthyological Research* 56(1): 62–68. http://dx.doi. org/10.1007/s10228-008-0084-8

Motomura H, Arbsuwan S, Musikasinthorn P. **2010**. *Thysanichthys evides*, a senior synonym of *Sebastella littoralis*, and a valid species of *Scorpaenodes*  (Actinopterygii: Scorpaenidae). *Species Diversity* 15(2): 71–81. http://dx.doi.org/10.12782/specdiv.15.71

- Motomura H, Béarez P, Causse R. **2011**. Review of Indo-Pacific specimens of the subfamily Scorpaeninae (Scorpaenidae), deposited in the Muséum national d'Histoire naturelle, Paris, with description of a new species of *Neomerinthe*. *Cybium* 35(1): 55–73. https://doi.org/10.26028/cybium/2011-351-006
- Motomura H, Burhanuddin AI, Iwatsuki Y. **2000**. Distributional implications of a poorly known polynemid fish, *Polydactylus sexfilis* (Pisces: Perciformes), in Japan. *Bulletin of the Faculty of Agriculture, Miyazaki University* 47(1/2): 115–120.
- Motomura H, Burhanuddin AI, Kimura S, Iwatsuki Y. **2001**. Fresh color notes for *Priacanthus prolixus* Starnes, 1988 from the west coast of India (Perciformes: Priacanthidae). *Biogeography* 3: 77–81.
- Motomura H, Causse R, Béarez P, Mishra SS. **2015**. Redescription of the Indo-West Pacific scorpionfish (Scorpaenidae), *Neomerinthe erostris* (Alcock 1896), a senior synonym of *Scorpaena gibbifrons* Fowler 1938, *N. rotunda* Chen 1981, and *N. bathyperimensis* Zajonz & Klausewitz 2002. *Zootaxa* 4021(4): 529–540. https://doi. org/10.11646/zootaxa.4021.4.3
- Motomura H, Causse R, Struthers CD. **2012**. *Phenacoscorpius longilineatus*, a new species of deepwater scorpionfish from the southwestern Pacific Ocean and the first records of *Phenacoscorpius adenensis* from the Pacific Ocean (Teleostei: Scorpaenidae). *Species Diversity* 17(2): 151–160. http://dx.doi.org/10.12782/sd.17.2.151
- Motomura H, Causse R, Struthers CD. **2016**. Redescription of the Indo-Pacific scorpionfish *Scorpaenodes guamensis* (Quoy & Gaimard 1824) (Scorpaenidae), a senior synonym of seven nominal species. *Zootaxa* 4067(3): 345–360. https://doi.org/10.11646/zootaxa.4067.3.4
- Motomura H, Iwatsuki Y. **2001**. A new genus, *Leptomelanosoma*, for the polynemid fish previously known as *Polydactylus indicus* (Shaw, 1804) and a redescription of the species. *Ichthyological Research* 48(1): 13–21.
- Motomura H, Iwatsuki Y. **2001**. Review of *Polydactylus* species (Perciformes: Polynemidae) characterized by a large black anterior lateral line spot, with descriptions of two new species. *Ichthyological Research* 48(4): 337–354.
- Motomura H, Iwatsuki Y, Kimura S. 2001. Redescription of *Polydactylus sexfilis* (Valenciennes *in* Cuvier and Valenciennes, 1831), a senior synonym of *P. kuru* (Bleeker, 1853) with designation of a lectotype (Perciformes: Polynemidae). *Ichthyological Research* 48(1): 83–89.
- Motomura H, Iwatsuki Y, Kimura S, Yoshino T. **2002**. Revision of the Indo-West Pacific polynemid fish genus *Eleutheronema* (Teleostei: Perciformes). *Ichthyological Research* 49(1): 47–61.

- Motomura H, Iwatsuki Y, Yoshino T. **2001**. A new species, *Polydactylus siamensis*, from Thailand and redescription of *P. plebeius* (Broussonet, 1782) with designation of a neotype (Perciformes: Polynemidae). *Ichthyological Research* 48(2): 117–126.
- Motomura H, Johnson JW. **2006**. Validity of the poorly known scorpionfish, *Rhinopias eschmeyeri*, with redescriptions of *R. frondosa* and *R. aphanes* (Scorpaeniformes: Scorpaenidae). *Copeia* 2006(3): 500–515.
- Motomura H, Kanade Y. **2015**. Review of the scorpionfish genus *Pteroidichthys* (Scorpaenidae), with descriptions of two new species. *Zootaxa* 4057(4): 490–510. https://doi.org/10.11646/zootaxa.4057.4.2
- Motomura H, Kullander SO, Yoshino T, Iwatsuki Y. **2002**. Review of seven-spined *Polynemus* species (Perciformes: Polynemidae) with designation of a neotype for *Polynemus paradiseus* Linnaeus, 1758. *Ichthyological Research* 49(4): 307–317.
- Motomura H, Last PR, Yearsley GK. **2007**. Two new species of the scorpionfish genus *Trachyscorpia* (Sebastidae: Sebastolobinae) from the southern Indo-West Pacific, with comments on the distribution of *T. eschmeyeri. Zootaxa* 1466: 19–34.
- Motomura H, Sakurai Y, Senou H, Ho H-C. **2009**. Morphological comparisons of the Indo-West Pacific scorpionfish, *Parascorpaena aurita*, with the closely related species, *P. picta*, with first records of *P. aurita* from East Asia (Scorpaeniformes: Scorpaenidae). *Zootaxa* 2191: 41–57. https://doi.org/10.11646/zootaxa.2191.1.2
- Motomura H, Sakurai Y, Shinohara G. **2009**. First records of a scorpionfish, *Scorpaenodes albaiensis*, from East Asia, with a synopsis of *S. minor* (Actinopterygii: Scorpaeniformes: Scorpaenidae). *Species Diversity* 14(2): 75–87. http://dx.doi.org/10.12782/specdiv.14.75
- Motomura H, Senou H. **2002**. Record of *Polydactylus sexfilis* (Perciformes: Polynemidae) from Hachijo-jima, Izu Islands, Japan with comments on morphological changes with growth and speciation of related species. *Bulletin of the Kanagawa Prefectural Museum* (*Natural Science*) 31: 27–31.
- Motomura H, Shinohara G. **2005**. Assessment of taxonomic characters of *Scorpaenopsis obtusa* and *S. gibbosa* (Scorpaenidae), with first records of *S. obtusa* from Japan and Australia and comments on the synonymy of *S. gibbosa*. *Cybium* 29(3): 295–301.
- Motomura H, Yamashita M, Itou M, Haraguchi Y, Iwatsuki Y. **2012**. First records of the two-tone goatfish, *Upeneus guttatus*, from Japan, and comparisons with *U. japonicus* (Perciformes: Mullidae). *Species Diversity* 17: 7–14. http://dx.doi.org/10.12782/sd.17.1.007
- Moyer JT, Sano M. **1987**. Feeding habits of two sympatric ostraciid fishes at Miyake-Jima, Japan. *Japanese Journal of Ichthyology* 34(1): 108–112.

Mukerji DD. **1935**. Notes on some rare and interesting fishes from the Andaman Islands, with descriptions of two new freshwater gobies. *Records of the Indian Museum* (Calcutta) 37(3): 259–277.

Müller J. 1836. Vergleichende Anatomie des Myxinoiden, der Cyclostomen mit durchbohrtem Gaumen. Erster Theil.
Osteologie und Myologie. Abhandlungen der Königlichen Akademie der Wissenschaften zu Berlin 1834: 65–340.

Müller J. 1838. Vergleichende Neurologie der Myxinoiden. Fortsetzung (3). Abhandlungen der Königlichen Akademie der Wissenschaften zu Berlin 1838: 171–251.

Müller J. 1841. Vergleichende Anatomie der Myxinoiden. Dritte Fortsetzung. Über das Gefässystem. Abhandlungen der Königlichen Akademie der Wissenschaften zu Berlin 1839: 175–304.

Müller J. **1842**. Über die Schwimmblase der Fische, mit Bezug auf einige neue Fischgattungen. Bericht über die zur Bekanntmachung geeigneten Verhandlungen der Königlich Preussischen Akademie der Wissenschaften zu Berlin 1842: 202–210.

Müller J. **1843**. Beiträge zur Kenntniss der natürlichen Familien der Fische. Archiv für Naturgeschichte 9: 292–330.

Müller J, Henle FGJ. **1837**. Über die Gattungen der Haifische und Rochen nach einer ... mit Hrn. Henle unternommenen gemeinschaftlichen Arbeit über die Naturgeschichte der Knorpelfische. Bericht über die zur Bekanntmachung geeigneten Verhandlungen der Königlichen Preussischen Akademie der Wissenschaften zu Berlin 1837: 111–118.

Müller J, Henle FGJ. **1837**. Ueber die Gattungen der Plagiostomen. *Archiv für Naturgeschichte* 3: 394–401, 434.

Müller J, Henle FGJ. 1838. On the generic characters of cartilaginous fishes, with descriptions of new genera. *Magazine of Natural History* (N.S.) 2(art.7): 33–37, 88–91.

Müller J, Henle FGJ. **1838**. Poissons cartilagineux. *L'Institut* 6: 63–65.

Müller J, Henle FGJ. **1838–1841**. *Systematische Beschreibung der Pagiostomen*. Berlin: Veit & Comp. xxii + 200 pp., Pls. 60.

Müller J, Troschel FH. 1848. Reisen in Britisch-Guiana in den Jahren 1840–1844 (Im Auftrag St. Mäjestat des Königs von Preussen ausgeführt von Richard Schomburgk) [title variant: Versuch einer Fauna und Flora von Britisch-Guiana] Vol. 3 (Fische): pp. 618–644.

Müller J, Troschel FH. **1849**. Horae Ichthyologicae. Beschreibung und Abbildung neuer Fische (Berlin) 3: 1–27.

Mundy BC. **1990**. Development of larvae and juveniles of the alfonsins, *Beryx splendens* and *B. decadactylus* (Berycidae, Beryciformes). *Bulletin of Marine Science* 46(2): 257–273.

Muñoz-Chápuli R, Ramos F. **1989**. Morphological comparison of *Squalus blainvillei* and *S. megalops* in the eastern Atlantic, with notes on the genus. *Japanese Journal of Ichthyology* 36(1): 6–21. Muñoz-Chápuli R, Ramos F. **1989**. Review of the *Centrophorus* sharks (Elasmobranchii, Squalidae) of the eastern Atlantic. *Cybium* 13(1): 65–81.

Munro ISR. **1938**. Handbook of Australian Fishes. *Fisheries Newsletter* 17(2): 71–75.

Munro ISR. **1948**. *Sparidentex hasta* (Valenciennes), a new name for *Chrysophrys cuvieri* Day. *Copeia* 1948(4): 275–280.

Munro ISR. **1950**. Revision of *Bregmaceros* with descriptions of larval stages from Australasia. *Proceedings of the Royal Society of Queensland* 61(5): 37–53.

Munro ISR. **1955**. *The marine and fresh water fishes of Ceylon*. Canberra: Department of External Affairs. 351 pp.

Munro ISR. **1956**. *Handbook of Australian fishes* (Nos 1–42). Sydney: Publicity Press. 172 pp. [Reprinted from Fisheries Newsletter]

Munro ISR. **1958**. The fishes of the New Guinea region: a check-list of the fishes of New Guinea incorporating records of species collected by the fisheries survey vessel *Fairwind* during the years 1948 to 1950. *Papua New Guinea Agricultural Journal* 10(4): 97–369. [Reprinted in the *Territory of Papua New Guinea Fisheries Bulletin* No. 1 (1958) with same pagination]

Munro ISR. **1967**. *The fishes of New Guinea*. Port Moresby, New Guinea: Department of Agriculture, Stock and Fisheries. xxxvii + 651 pp.

Munroe TA. **1990**. Eastern Atlantic tonguefishes (*Symphurus*: Cynoglossidae, Pleuronectiformes), with descriptions of two new species. *Bulletin of Marine Science* 47(2): 464–515.

Munroe TA. **1992**. Interdigitation pattern of dorsal-fin pterygiophores and neural spines, an important diagnostic character for symphurine tonguefishes (*Symphurus*: Cynoglossidae: Pleuronectiformes). *Bulletin of Marine Science* 50(3): 357–403.

Munroe TA. **1998**. Systematics and ecology of tonguefishes of the genus *Symphurus* (Cynoglossidae: Pleuronectiformes) from the western Atlantic Ocean. *Fishery Bulletin* 96(1): 1–182.

Munroe TA. **2001**. Cynoglossidae. Tonguesoles (pp. 3890– 3901). In: Carpenter KE, Niem VH (eds) *FAO species identification guide for fishery purposes. The living marine resources of the western central Pacific*. Vol. 6. Bony fishes part 4 (pp. 3381–4218). Rome: FAO.

Munroe TA. **2005**. Systematic diversity of the Pleuronectiformes (pp. 10–41); Distributions and biogeography (pp. 42–67); Tropical flatfish fisheries (pp. 292–318). In: Gibson RN (ed) *Flatfishes: Biology and exploitation*. Oxford: Blackwell Science. xxiv + 391 pp.

Munroe TA. **2006**. New western Indian Ocean tonguefish (Pleuronectiformes: Cynoglossidae, *Symphurus*). *Copeia* 2006(2): 230–234. Munroe TA. **2015**. Systematic diversity of the Pleuronectiformes (pp. 13–51); Distributions and biogeography (pp. 52–82); Tropical flatfish fisheries (pp. 418–460). In: Gibson RN, Nash RDM, Geffen AJ, van der Veer HW (eds) *Flatfishes: Biology and exploitation* (2<sup>nd</sup> edition). John Wiley & Sons, Ltd. 542 pp. https://doi. org/10.1002/9781118501153

Munroe TA. **2017**. Rediscovery of the holotype of the tongue sole *Cynoglossus dollfusi* (Chabanaud, 1931), and impacts of this discovery on Red Sea records for *C. sealarki* Regan, 1908, *C. lingua* Hamilton, 1822, and *C. zanzibarensis* Norman, 1939, and on the taxonomic status of *C. cleopatridis* Chabanaud, 1949 (Pisces: Pleuronectiformes: Cynoglossidae). *Proceedings of the Biological Society of Washington* 130: 5–33. https://doi.org/10.2988/0006-324X-130.1.yy

Munroe TA, Amaoka K. **1998**. *Symphurus hondoensis* Hubbs, 1915, a valid species of western Pacific tonguefish (Pleuronectiformes: Cynoglossidae). *Ichthyological Research* 45(4): 385–391.

Munroe TA, Desoutter M. **2001**. On the authorship, identity and taxonomic position of *Pleuronectes commersonnii* Lacepède, 1802 (Pleuronectiformes, Soleidae). *Cybium* 25(3): 273–277.

Munroe TA, Kong X-Y. **2016**. Resolving uncertainties regarding the nomenclature and status of the tongue soles *Paraplagusia dollfusi* Chabanaud, 1931 and "*Cynoglossus* (*Trulla*) *dollfusi* (Chabanaud, 1937)" (Teleostei: Pleuronectiformes: Cynoglossidae). Proceedings of the Biological Society of Washington 129: 10–23. http://dx.doi.org/10.2988/0006-324X-129.Q1.10

Munroe TA, Krupp F, Schneider M. 1995. Family Cynoglossidae (Lenfuas, lenguetas) (pp. 1039–1059). In: Fischer W, Krupp F, Schneider W, Sommer C, Carpenter KE, Niem VH (eds) *Guía FAO para la identificación de especies para los fines de la pesca. Pacífico centro-oriental*. Vol. 2. Rome: FAO.

Munroe TA, Marsh BN. **1997**. Taxonomic status of three nominal species of Indo-Pacific symphurine tonguefishes (*Symphurus*: Cynoglossidae: Pleuronectiformes). *Ichthyological Research* 44(2): 189–200. https://doi. org/10.1007/BF02678697

Munroe TA, McCosker JE. **2001**. Redescription of *Symphurus diabolicus* Mahadeva and Munroe, a poorly-known, deepsea tonguefish (Pleuronectiformes: Cynoglossidae) from the Galápagos Archipelago. *Revista de Biologica Tropical* 49(Suppl. 1): 187–198.

Munroe TA, Nizinski MS. 2002. Clupeidae. Herrings (shads, menhadens) (pp. 804–830). In: Carpenter KE (ed) FAO species identification guide for fishery purposes. The living marine resources of the western central Atlantic. Vol. 2. Bony fishes part 1 (pp. 601–1374). Rome: FAO. Munroe TA, Wongratana T, Nizinski MS. 1999.

Pristigasteridae. Ilishas, pellonas (pp. 1754–1770); Clupeidae. Herrings (also, sardines, shads, sprats, pilchards, and menhadens) (pp. 1775–1821). In: Carpenter KE, Niem VH (eds) *FAO species identification guide for fishery purposes. The living marine resources of the western central Pacific.* Vol. 3. Batoid fishes, chimaeras and bony fishes part 1 (pp. 1397–2068). Rome: FAO.

Muraleedharan PM, Kumar SP. **2014**. Arabian Sea upwelling – a comparison between coastal and open ocean regimes. *Current Science* 71: 842–846.

Murase A. **2007**. A new species of the blenniid fish, *Laiphognathus longispinis* (Perciformes: Blenniidae), from southern Japan and Taiwan. *Ichthyological Research* 54(3): 287–296.

Murdy EO. **1985**. A review of the gobiid fish genera *Exyrias* and *Macrodontogobius*, with description of a new species of *Exyrias*. *Indo-Pacific Fishes* 10: 1–14.

Murdy EO. **1989**. A taxonomic revision and cladistic analysis of the oxudercine gobies (Gobiidae: Oxudercinae). *Records of the Australian Museum*, Supplement 11: 1–93.

Murdy EO. **2006**. A revision of the gobiid fish genus *Trypauchen* (Gobiidae: Amblyopinae). *Zootaxa* 1343: 55–68.

Murdy EO. **2008**. *Paratrypauchen*, a new genus for *Trypauchen microcephalus* Bleeker, 1860, (Perciformes: Gobiidae: Amblyopinae) with a redescription of *Ctenotrypauchen chinensis* Steindachner, 1867, and a key to '*Trypauchen*' group genera. *aqua, International Journal of Ichthyology* 14(3): 115–128.

Murdy EO. **2011**. Systematics of Amblyopinae (pp. 99–106). In: Patzner RA, Van Tassell JL, Kovačić M, Kapoor BG (eds) *The biology of gobies*. Enfield, New Hampshire: CRC Press, Science Publishers. 722 pp. http://dx.doi.org/10.1201/ b11397-10

Murdy EO, Hoese DF. **1985**. Revision of the gobiid fish genus *Istigobius. Indo-Pacific Fishes* 4: 1–41.

Murdy EO, Randall JE. **2002**. *Taenioides kentalleni*, a new species of eel goby from Saudi Arabia (Gobiidae: Amblyopinae). *Zootaxa* 93: 1–6.

Murdy EO, Shibukawa K. **2001**. A revision of the gobiid fish genus *Odontamblyopus* (Gobiidae: Amblyopinae). *Ichthyological Research* 48(1): 31–43.

Murray BW, Wang JY, Yang S-C, Stevens JD, Fisk A, Svavarsson J. **2008**. Mitochondrial cytochrome b variation in sleeper sharks (Squaliformes: Somniosidae). *Marine Biology* 153(6): 1015–1022. http://dx.doi.org/10.1007/s00227-007-0871-1

Murray JA. **1887**. New species of fish from Kurrachee and the Persian Gulf. *Journal of the Bombay Natural History Society* 2(1): 47–49.

Murty VSR. **1982**. On the fishes of the family Platycephalidae of the seas around India. *Journal of the Marine Biological Association of India* 17(3): 679–694.

Murugan A, Dhanya S, Rajagopal S, Balasubramanian T. **2008**. Seahorses and pipefishes of the Tamil Nadu coast. *Current Science* 95(2): 253–260.

Musick JA. 2000. Latimeria chalumnae. The IUCN Red List of Threatened Species 2000: e.T11375A3274618. https://doi. org/10.2305/IUCN.UK.2000.RLTS.T11375A3274618.en

Muthiah C, Neelakantan B. **1991**. On the occurrence of *Saurida isarankurai* Shindo and Yamada 1972 from the west coast of India. *Journal of the Bombay Natural History Society* 88: 461–463.

Myagkov NA. **1986**. Oxynotidae. In: Gubanov EP, Kondyurin VV, Myagkov NA (eds) *Sharks of the world ocean: Identification handbook*. Moscow. 272 pp. [In Russian]

Myagkov NA, Kondyurin VV. **1986**. Dogfishes, *Squalus* (Squalidae), of the Atlantic Ocean and comparative notes on the species of this genus from other regions. *Journal of Ichthyology* 26(6): 1–18.

Myers GS. **1934**. Three new deep-water fishes from the West Indies. *Smithsonian Miscellaneous Collection* 91(9): 1–12.

Myers GS. **1936**. A new polynemid fish collected in the Sadong River, Sarawak, by D. William T. Hornaday, with notes on the genera of Polynemidae. *Journal of the Washington Academy of Sciences* 26(9): 376–382.

Myers GS, Wade CB. **1941**. Four new genera and ten new species of eels from the Pacific coast of tropical America. *Allan Hancock Pacific Expedition 1932–40: Reports* 9(4): 65–111.

Myers RF. **1989**. *Micronesian reef fishes: A practical guide to the coral reef fishes of the tropical central and western Pacific.* Guam: Coral Graphics. 298 pp.

Myers RF. **1991**. *Micronesian reef fishes: A practical guide to the coral reef fishes of the tropical central and western Pacific* (2<sup>nd</sup> edition). Guam: Coral Graphics. 298 pp.

Myers RF. **1999**. *Micronesian reef fishes: A comprehensive guide to the coral reef fishes of Micronesia* (3<sup>rd</sup> revised and expanded edition). Guam: Coral Graphics. 330 pp.

Myers RF, Shepard JW. **1980**. New records of fishes from Guam, with notes on the ichthyofauna of the southern Marianas. *Micronesica* 16(2): 305–347.

Myrberg AA, Montgomery WL, Fishelson L. **1988**. The reproductive behavior of *Acanthurus nigrofuscus* (Forskal) and other surgeonfishes (Fam. Acanthuridae) off Eilat, Israel (Gulf of Aqaba, Red Sea). *Ethology* 79: 31–61.

Mytilineou C, Politou C-Y, Papaconstantinou C, Kavadas S, D'Onghia G, Sion L. **2005**. Deep-water fish fauna in the Eastern Ionian Sea. *Belgian Journal of Zoology* 135(2): 2 29–233.

### Ν

Nagabhushanam AK, Rao GC. **1972**. An ecological survey of the marine fauna of Minicoy Atoll (Laccadive Archipelago, Arabian Sea). *Mitteilungen aus dem Zoologischen Museum in Berlin* 48(2): 265–324.

Naik SK, Uikey DE, Russell BC, Shanbhag AB. **2002**. Two new records of *Parascolopsis* (Pisces: Nemipteridae) from the west coast of India, with a redescription of *Parascolopsis boesemani* (Rao and Rao). *The Beagle, Records of the Museums and Art Galleries of the Northern Territory* 18: 73–76.

Naim O, Cuet P, Manger V. **2000**. Chapter 12. The Mascarene Islands (pp. 353–381). In: McClanahan TR, Sheppard CRC, Obura DO (eds) *Coral reefs of the Indian Ocean: Their ecology and conservation*. Oxford: Oxford University Press. xxiii + 525 pp.

Nair NB, Nair JR, Kumar KK, Azis PKA, Dharmaraj K, Arunachalam M. 1983. Occurrence of Hyporhamphus xanthopterus (Val.) in the lakes of southern Kerala. Mahasagar – Bulletin of the National Institute of Oceanography 16(4): 479–482.

Nair RJ, Praveen P, Kumar SD, Kuriakose S. 2012. First record of the dwarf monocle bream, *Parascolopsis baranesi* (Family: Nemipteridae) from Indian waters. *Indian Journal of Geo Marine Sciences* 41(5): 395–397.

Nair RV, Appukuttan KK. **1973**. Observations on the food of deep sea sharks *Halaeurus hispidus* (Alcock), *Eridacnis radcliffei* Smith and *Iago omanensis* Compagno and Springer. *Indian Journal of Fisheries* 20(2): 575–583.

Nair RV, Appukuttan KK. **1974**. Observations on the developmental stages of the smooth dogfish, *Eridacnis radcliffei* Smith from Gulf of Mannar. *Indian Journal of Fisheries* 21(1): 141–151.

Nair RV, Appukuttan KK, Rajapandian ME. **1974**. On the systematics and identity of four pelagic sharks of the family Carcharhinidae from Indian region. *Indian Journal of Fisheries* 21(1): 220–232.

Nair RV, Lal Mohan RS. **1971**. On the occurrence of the spiny shark *Echinorhinus brucus* (Bonnaterre) from the east coast of India with a note on its distribution. *Indian Journal of Animal Sciences* 41(10): 1011–1014.

Nair RV, Lal Mohan RS. **1973**. On a new deep sea skate, *Rhinobatos variegatus*, with notes on the deep sea sharks *Halaelurus hispidus*, *Eridacnis radcliffei* and *Eugaleus omanensis* from the Gulf of Mannar. *Senckenbergiana Biologica* 54(1/3): 71–80.

Nakabo T. **1979**. A new and two rare species of the genus *Callionymus* (Callionymidae) from the western Indian Ocean. *Japanese Journal of Ichthyology* 26(3): 231–237.

Nakabo T. **1982**. Revision of genera of the dragonets (Pisces: Callionymidae). *Publications of the Seto Marine Biological Laboratory* 27(1/3): 77–131. Nakabo T. **1982**. Revision of the family Draconettidae. *Japanese Journal of Ichthyology* 28(4): 355–367.

Nakabo T (ed). **1993**. *Fishes of Japan with pictorial keys to the species*. Tokyo: Tokyo University Press. 1474 pp. [In Japanese]

Nakabo T (ed). **2000**. *Fishes of Japan with pictorial keys to the species* (2<sup>nd</sup> edition). Tokyo: Tokyo University Press. 1474 pp. [In Japanese]

Nakabo T. **2001**. *Lagocephalus suezensis*. In: Nakabo *et al*.

Nakabo T (ed). **2002**. *Fishes of Japan with pictorial keys to the species* (English edition). Tokyo: Tokyo University Press. 1749 pp.

Nakabo T, Machido K, Yamaoka K, Nishida K (eds). **2001**. *Fishes of the Kuroshio Current, Japan*. Osaka: Nikkei Osaka PR. 278 pp.

Nakabo T, Senou H, Masuda H. **1993**. *Scorpaenopsis iop*, a new species of Scorpaenidae from southern Japan. *Japanese Journal of Ichthyology* 40(1): 29–33.

Nakamura H. **1935**. On the two species of the thresher shark from Formosan waters. *Memoirs Faculty Science Taihoku Imperial University Formosa* 14(1): 1–6.

Nakamura I. **1975**. Synopsis of the biology of the black marlin, *Makaira indica* (Cuvier), 1831. *NOAA Technical Report* NMFS SSRF-675 (Part 3): 17–27.

Nakamura I. **1984**. Gempylidae. Snake mackerels, barracoutas, escolars and oilfishes. In: Fischer W, Bianchi G (eds) *FAO species identification sheets for fishery purposes. Western Indian Ocean (Fishing Area 51).* Vol. 2. Rome: FAO. [unpaginated]

Nakamura I. **1984**. Istiophoridae. Billfishes (spearfishes, marlins and sailfishes). In: Fischer W, Bianchi G (eds) *FAO species identification sheets for fishery purposes. Western Indian Ocean (Fishing Area 51)*. Vol. 2. Rome: FAO. [unpaginated]

Nakamura I. 1984. Trichiuridae. Cutlassfishes, hairtailfishes, frostfishes, scabbardfishes. In: Fischer W, Bianchi G (eds) FAO species identification sheets for fishery purposes. Western Indian Ocean (Fishing Area 51). Vol. 4. Rome: FAO. [unpaginated]

Nakamura I. **1984**. Xiphiidae. Swordfishes. In: Fischer W, Bianchi G (eds) *FAO species identification sheets for fishery purposes. Western Indian Ocean (Fishing Area 51).* Vol. 5. Rome: FAO. [unpaginated]

Nakamura I. **1985**. FAO species catalogue. Vol. 5. Billfishes of the world. An annotated and illustrated catalogue of marlins, spearfishes and swordfishes known to date. *FAO Fisheries Synopsis No. 125* (Vol. 5). Rome: FAO. 65 pp.

Nakamura I. **1986**. *Important fishes trawled off Patagonia*. Tokyo: Japan Marine Fishery Resource Center. 369 pp. Nakamura I. **1990**. Gempylidae (pp. 402–403); Scombridae (pp. 404–405). In: Gon O, Heemstra PC (eds) *Fishes of the Southern Ocean*. Grahamstown: J.L.B. Smith Institute of Ichthyology. 462 pp.

Nakamura I, Parin NV. **1993**. FAO species catalogue. Vol. 15. Snake mackerels and cutlassfishes of the world (families Gempylidae and Trichiuridae). An annotated and illustrated catalogue of the snake mackerels, snoeks, escolars, gemfishes, sackfishes, domine, oilfish, cutlassfishes, scabbardfishes, hairtails, and frostfishes known to date. *FAO Fisheries Synopsis No. 125* (Vol. 15). Rome: FAO. 136 pp.

Nakamura I, Parin NV. 2001. Gempylidae. Snake mackerels (pp. 3698–3708); Trichiuridae. Cutlassfishes (pp. 3709–3720).
In: Carpenter KC, Niem VH (eds) FAO species identification guide for fishery purposes. The living marine resources of the western central Pacific. Vol. 6. Bony fishes part 4 (pp. 3381– 4218). Rome: FAO.

Nakaya K. **1971**. Descriptive notes on a porbeagle, *Lamna nasus*, from Argentine waters, compared with the North Pacific salmon shark, *Lamna ditropis*. *Bulletin of the Faculty of Fisheries, Hokkaido University* 21(4): 269–279.

Nakaya K. **1988**. Morphology and taxonomy of *Apristurus longicephalus* (Lamniformes, Scyliorhinidae). *Japanese Journal of Ichthyology* 34(4): 431–442.

Nakaya K, Sato K, Iglésias SP. 2008. Occurrence of *Apristurus* melanoasper from the South Pacific, Indian and South Atlantic Oceans (Carcharhiniformes: Scyliorhinidae) (pp. 61–74). In: Last PR, White WT, Pogonoski JJ (eds) Descriptions of new Australian chondrichthyans. Hobart, Australia: CSIRO Marine and Atmospheric Research. 358 pp.

Nakazono A, Nakatani H, Tsukahara H. **1985**. Reproductive ecology of the Japanese reef fish *Parapercis snyderi*. In: Harmelin-Vivien M, Salvat B (eds) *Proceedings of the 5th International Coral Reef Congress* (Tahiti) Vol. 5: 355–360.

Nalbant T. **1979**. Studies on the reef fishes of Tanzania. II. *Neosynchiropus bacescui* gen. n., sp. n., an interesting dragonet fish from Makatumbe coral reefs (Pisces, Perciformes, Callionymoidei). *Travaux du Muséum d'Histoire Naturelle "Grigore Antipa"* 20(1): 349–352.

Nalbant T, Mayer R. **1975**. Studies on the reef fishes of Tanzania. I. New and Interesting species. *Travaux du Muséum d'Histoire Naturelle "Grigore Antipa*" 16: 235–242.

Naqvi SWA, Narvekar PV, Desa E. 2006. Coastal biogeochemical processes in the north Indian Ocean (pp. 723–783). In: Robinson AR, Brink KH (eds) *The sea*, *Vol. 14A: The global coastal ocean: Interdisciplinary regional studies and syntheses*. Cambridge, Massachusetts: Harvard University Press. 840 pp. Narahara A, Bergman HL, Laurent P, Maina JN, Walsh PJ, Wood CM. **1996**. Respiratory physiology of the Lake Magadi tilapia (*Oreochromis alcalicus grahami*), a fish adapted to a hot, alkaline, and frequently hypoxic environment. *Physiological Zoology* 69(5): 1114–1136.

Nardo GD. **1827**. Prodromus observationum et disquisitionum Adriaticae ichthyologiae. *Giornale di fisica, chimica e storia naturale, medicina, ed arti* (Series 2) 10: 22–40.

Nardo GD. **1827**. Prodromus observationum et disquisitionum ichthyologiae Adriaticae. *Isis* (*Oken*) 20(no. 6): 473–489.

Nardo GD. **1840**. Considerazioni sulla famiglia dei pesci *Mola*, e sui caratteri che li distinguono. *Annali delle Scienze del Regno Lombardo-Veneto* 10: 105–112.

Nardo GD. **1840/1841**. Nota sopra uno smisurato individuo della specie *Mola aspera* Nardo, stato preso nelle vicinanze di Venezia. *Atti del Reale Istituto Veneto di Scienze, lettere ed arti* 1: 130.

Natanson LJ, Kohler NE. **1996**. A preliminary estimate of age and growth of the dusky shark *Carcharhinus obscurus* from the south-west Indian Ocean, with comparisons to the Western North Atlantic population. *South African Journal of Marine Science* 17(1): 217–224.

Natanson LJ, Kohler NE, Ardizzone D, Cailliet GM, Wintner SP, Mollet HF. **2006**. Validated age and growth estimates for the shortfin mako, *Isurus oxyrinchus*, in the North Atlantic Ocean. *Environmental Biology of Fishes* 77: 367–383.

Natanson LJ, Winttner SP, Johansson F, Piercy A, Campbell P, De Maddalena A, Gulak SJB, Human B, Fulgosi FC, Ebert DA, Hemida F, Mollen FH, Vanni S, Burgess GH, Compagno LJV, Wedderburn-Maxwell A. **2008**. Ontogenetic vertebral growth patterns in the basking shark *Cetorhinus maximus*. *Marine Ecology Progress Series* 361: 267–278. http://dx.doi.org/10.3354/meps07399

Navarro F de P, Lozano F, Novaz JM, Otero E, Sainz Padro J. 1943. La pesca de arrastre en los fondos del Cabo blanco y del banco Arguín (Africa Sahariana): resultados científicos de dos campañas realizadas por el Instituto Español de Oceanografía en barcos de P.Y.S.B.E. *Trabajos (Instituto Español de Oceanografía)* 18: 1–225.

Naylor GJP, Caira JN, Jensen K, Rosana KAM, Straube N, Lakner C. **2012**. Elasmobranch phylogeny: a mitochondrial estimate based on 595 species (pp. 31–56). In: Carrier JC, Musick JA, Heithaus MR (eds) *Biology of sharks and their relatives* (2<sup>nd</sup> edition). Boca Raton, Florida: CRC Press. 672 pp.

Naylor GJP, Caira JN, Jensen K, Rosana KAM, White WT, Last PR. **2012**. A DNA sequence-based approach to the identification of shark and ray species and its implications for global elasmobranch diversity and parasitology. *Bulletin of the American Museum of Natural History* 367: 1–262. http://dx.doi.org/10.1206/754.1 Naylor GJP, Ryburn JA, Fedrigo O, Lopez A. **2005**. Phylogenetic relationships among the major lineages of modern elasmobranchs (pp. 1–25). In: Hamlett WC (ed) *Reproductive biology and phylogeny of Chondrichthyes: Sharks, batoids and chimaeras. Vol. 3 of series: Reproductive Biology and Phylogeny.* CRC Press, Taylor & Francis Group. 572 pp.

Naylor GJP, Yang L, Corrigan S, Carvalho MR de. **2016**. Phylogeny and classification of rays (pp. 10–15). In: Last PR, White WT, Carvalho MR de, Séret B, Stehmann MFW, Naylor GJP (eds) *Rays of the world*. CSIRO Publishing & Cornell University Press. 790 pp.

Nayudu MR. **1923**. A note on the eggs and early embryonic development of *Cypsilurus*. *Madras Fisheries Bulletin* 15(4): 109–112.

Near TJ. **2009**. Conflict and resolution between phylogenies inferred from molecular and phenotypic data sets for hagfish, lampreys, and gnathostomes. *Journal of Experimental Zoology* (Part B: Molecular and Developmental Evolution) 312(7): 749–761. http://dx.doi. org/10.1002/jez.b.21293

Near TJ, Dornburg A, Eytan RI, Keck BP, Smith WL, Kuhn KL, Moore JA, Price SA, Burbrink FT, Friedman M, Wainwright PC. 2013. Phylogeny and tempo of diversification in the superradiation of spiny-rayed fishes. *Proceedings of the National Academy of Sciences of the United States of America* 110(31): 12738–12743. http://dx.doi.org/10.1073/ pnas.1304661110

Near TJ, Eytan RI, Dornburg A, Kuhn KL, Moore JA, Davis MP, Wainright PC, Friedman M, Smith WL. 2012.
Resolution of ray-finned fish phylogeny and timing of diversification. *Proceedings of the National Academy of Sciences* 109(34): 13698–13703. https://doi.org/10.1073/ pnas.1206625109

Neira FJ, Miskiewicz GA, Trnski T (eds). **1998**. *Larvae of temperate Australian fishes. Laboratory guide for larval fish identification*. Nedlands: University of Western Australia Press. 474pp.

Nekrasov VV. **1966.** A new subspecies of *Trachurus (Trachurus mediterraneus indicus* Necrassov subsp. n.) in the Indian Ocean. *Zoologicheskii Zhurnal* 45(1): 141–144. [In Russian, English summary]

Nekrasov VV. **1970**. Mackerels (Carangidae family) of the eastern coasts of Africa. *Trudy Azovsko-Černomorskoj Naučno-Promyslovoj Ékspedicii* 29: 89–138. [In Russian]

Nekrasov VV. **1978**. Systematic position of horse mackerel of the genus *Trachurus* from the western part of the Indian Ocean. *Journal of Ichthyology* 18(1): 15–19.

Nellen W. **1973**. Fischlarven des Indischen Ozeans, Ergebnisse der Fischbrutuntersuchungen wahrend der ersten Expedition des Forschungsschiffs *Meteor* in den Indischen Ozean und den Persischen Golf, Oktober 1964 bis April 1965. *Meteor Forschungsergebnisse* (Reihe D, Biologie) 14: 1–66. Nelson DR, McKibben JN, Strong WR Jr, Lowe CG, Sisneros JA, Schroeder DM, Lavenberg RJ. **1997**. An acoustic tracking of a megamouth shark, *Megachasma pelagios*: a crepuscular vertical migrator. *Environmental Biology of Fishes* 49(4): 389–399.

Nelson GJ. **1969**. Gill arches and the phylogeny of fishes, with notes on the classification of vertebrates. *Bulletin of the American Museum of Natural History* 141(article 4): 477–552.

Nelson GJ. **1969**. Infraorbital bones and their bearing on the phylogeny and geography of osteoglossomorph fishes. *American Museum Novitates* 2394: 1–37.

Nelson GJ. **1969**. Origin and diversification of teleostean fishes. Annals of the New York Academy of Sciences 167: 18–30.

Nelson GJ. 1973. Relationships of clupeomorphs, with remarks on the structure of the lower jaw in fishes (pp. 333–349). In: Greenwood PH, Miles RS, Patterson C (eds) *Interrelationships* of fishes. London: Academic Press. 536 pp.

Nelson GJ, McCarthy LJ. **1995**. Two new species of gizzard shads of the genus *Nematalosa* (Teleostei, Clupeidae, Dorosomatinae) from Persian/Arabian Gulf. *Japanese Journal of Ichthyology* 41(4): 379–383.

Nelson GJ, Rothman MN. **1973**. The species of gizzard shads (Dorosomatinae) with particular reference to the Indo-Pacific region. *Bulletin of the American Museum of Natural History* 150(article 2): 131–206.

Nelson JS. **1978**. *Bembrops morelandi*, a new percophidid fish from New Zealand, with notes on other members of the genus. *Records of the National Museum of New Zealand* 1(14): 237–241.

Nelson JS. 1978. Limnichthys polyactis, a new species of blennioid fish from New Zealand, with notes on the taxonomy and distribution of other Creediidae (including Limnichthyidae). New Zealand Journal of Zoology 5(2): 351–364.

Nelson JS. **1979**. Some osteological differences between the blennioid fishes *Limnichthys polyactis* and *L. rendahli*, with comments on other species of Creediidae. *New Zealand Journal of Zoology* 6(2): 273–277.

Nelson JS. **1982**. *Pteropsaron heemstrai* and *Osopsaron natalensis* (Perciformes: Percophidae), new fish from South Africa, with comments on *Squamicreedia obtusa* from Australia and on the classification of the subfamily Hemerocoetinae. *J.L.B. Smith Institute of Ichthyology Special Publication* 25: 1–11.

Nelson JS. **1984**. *Fishes of the world* (2<sup>nd</sup> edition). New York: John Wiley & Sons. xv + 523 pp.

Nelson JS. **1985**. On the interrelationships of the genera of Creediidae (Perciformes: Trachinoidei). *Japanese Journal of Ichthyology* 32(3): 283–293. Nelson JS. **1986**. Some characters of Trichonotidae, with emphasis to those distinguishing it from Creediidae (Perciformes: Trachinoidei). *Japanese Journal of Ichthyology* 33(1): 1–6.

Nelson JS. **1994**. *Fishes of the world* (3<sup>rd</sup> edition). New York: John Wiley & Sons, Inc. 600 pp.

Nelson JS. **2006**. *Fishes of the world* (4<sup>th</sup> edition). Hoboken, New Jersey: John Wiley & Sons, Inc. 624 pp.

Nelson JS, Grande TC, Wilson MVH. **2016**. *Fishes of the world* (5<sup>th</sup> edition). Hoboken, New Jersey: John Wiley & Sons. 752 pp.

Nelson JS, Schultze H-P, Wilson MVH (eds). **2010**. *Origin and phylogenetic interrelationships of teleosts*. Munich: Verlag Dr. Friedrich Pfeil. 482 pp.

Nemeth D. **1994**. Systematics and distribution of fishes of the family Champsodontidae (Teleostei: Perciformes), with descriptions of three new species. *Copeia* 1994(2): 347–371.

Newman HH. **1907**. Spawning behavior and sexual dimorphism in *Fundulus heteroclitus* and allied fish. *Biological Bulletin* 12(5): 314–348.

Ng HH, Sparks JS. **2002**. *Plotosus fisadoha*, a new species of marine catfish (Teleostei: Siluriformes: Plotosidae) from Madagascar. *Proceedings of the Biological Society of Washington* 115(3): 564–569.

Ng HH, Sparks JS. **2003**. The ariid catfishes (Teleostei: Siluriformes: Ariidae) of Madagascar, with the description of two new species. *Occasional Papers of the Museum of Zoology, University of Michigan* 735: 1–21.

Ngusaru A. **1997**. Geological history. In: Richmond MD (ed) *A guide to the seashores of eastern Africa and the western Indian Ocean islands*. Department of Research Cooperation. Stockholm: SIDA/SAREC. 464 pp.

Nichols JT. **1920**. A contribution to the ichthyology of Bermuda. *Proceedings of the Biological Society of Washington* 33: 59–64.

Nichols JT. **1923**. A new wrasse and two new cichlids from northeast Africa. *American Museum Novitates* 65: 1–4.

Nichols JT, Breder CM. **1928**. An annotated list of the Synenthognathi with remarks of their development and relationships. *Zoologica* (N.Y.) 8(7): 423–448.

Nichols JT, Breder CM. **1932**. A new "two-winged" flying-fish from Mauritius. *American Museum Novitates* 561: 1–2.

Nichols JT, Breder CM. **1935**. New Pacific flying fishes collected by Templeton Crocker. *American Museum Novitates* 821: 1–4.

Nichols JT, Firth FE. **1939**. Rare fishes off the Atlantic coast, including a new grammicolepid. *Proceedings of the Biological Society of Washington* 52: 85–88.

Nichols OC, Tscherter UT. **2011**. Feeding of sea lampreys *Petromyzon marinus* on minke whales *Balaenoptera acutorostrata* in the St Lawrence Estuary, Canada. *Journal of Fish Biology* 78(1): 338–343. http://dx.doi.org/10.1111/ j.1095-8649.2010.02842.x Nichols P. 2001. Conservation and sustainable use of biodiversity of Socotra Archipelago. Part I: Fisheries overview and management options (90 pp.). Part II: Data collection system for monitoring and assessment (40 pp.). Senckenberg Research Institute, Frankfurt a.M., Germany. 130 pp.

Niebuhr C. **1775**. *Descriptiones animalium avium*, *amphibiorum, piscium, insectorum, vermium; quae in itinere orientali observavit Petrus Forskål*. Post mortem auctoris edidit Carsten Niebuhr. Hauniae. 20 + xxxiv + 164 pp.

Nielsen JG. **1960**. On some fishes from Karachi and Bombay with description of a new genus and species of Haliophidae. *Videnskabelige Meddelelser fra Dansk Naturhistorisk Forening* (Kjøbenhavn) 123: 249–256.

Nielsen JG. **1961**. Heterosomata (Pisces). *Galathea Report* 4: 219–226.

Nielsen JG. **1961**. Psettodoidea and Pleuronectoidea (Pisces: Heterosomata). *Atlantide Report* 6: 101–127.

Nielsen JG. **1963**. Soleoidea (Pisces: Heterosomata). *Atlantide Report* 7: 7–35.

Nielsen JG. **1964**. Heterosomata (Pisces) collected by Dr. Th. Mortensen off South Africa. *Videnskabelige Meddelelser fra Dansk Naturhistorisk Forening* (Kjøbenhavn) 127: 127–134.

Nielsen JG. **1971**. Ergebnisse der Forschungsreisen des FFS *Walther Herwig* nach Südamerika. 16. Redescription of the genus *Selachophidium* (Pisces: Brotulidae) with two new species. *Archiv für Fischereiwissenschaft* 22(1): 17–33.

Nielsen JG. **1974**. *Fish types in the Zoological Museum of Copenhagen*. Copenhagen: Zoological Museum, University of Copenhagen. 115 pp.

Nielsen JG. 1984. Bothidae. Lefteye flounders. In: Fischer W, Bianchi G. (eds) FAO species identification sheets for fishery purposes. Western Indian Ocean (Fishing Area 51). Vol. 1. Rome: FAO. [unpaginated]

Nielsen JG. 1995. A review of the species of the genus *Neobythites* (Pisces: Ophidiidae) from the western Indian Ocean, with descriptions of seven new species. *Ichthyological Bulletin of the J.L.B. Smith Institute of Ichthyology* 62: 1–19.

Nielsen JG. 1997. Deepwater ophidiiform fishes from off New Caledonia with six new species. In: Séret B (ed) Résultats des Campagnes MUSORSTOM, Vol. 17. Mémoires du Muséum National d'Histoire Naturelle (Paris) (N.S, Série A, Zoologie) 174: 51–82.

Nielsen JG. **2002**. Revision of the Indo-Pacific species of *Neobythites* (Teleostei, Ophidiidae), with 15 new species. *Galathea Report* 19: 5–104.

Nielsen JG, Cohen DM. 1999. Family Bythitidae (pp. 94–135).
In: Nielsen JG, Cohen DM, Markle DF, Robins CR (eds)
FAO Species Catalogue. Vol. 18. Ophidiiform fishes of the world (Order Ophidiiformes). An annotated and illustrated catalogue of pearlfishes, cusk-eels, brotulas and other

ophidiiform fishes known to date. *FAO Fisheries Synopsis No. 125* (Vol. 18). Rome: FAO. 178 pp.

Nielsen JG, Cohen DM, Markle DF, Robins CR (eds). 1999. FAO species catalogue. Vol. 18. Ophidiiform fishes of the world (Order Ophidiiformes). An annotated and illustrated catalogue of pearlfishes, cusk-eels, brotulas and other ophidiiform fishes known to date. FAO Fisheries Synopsis No. 125 (Vol. 18). Rome: FAO. 178 pp.

Nielsen JG, Machida Y. **1988**. Revision of the Indo-Pacific fish genus *Glyptophidium* (Ophidiiformes, Ophidiidae). *Japanese Journal of Ichthyology* 35(3): 289–319.

Nielsen JG, Quéro J-C. **1991**. Quelques Ophidiiformes de l'île de la Réunion: description d'une espèce nouvelle. *Cybium* 15(3): 193–198.

Nielsen JG, Smith DG. **1978**. The eel family Nemichthyidae (Pisces: Anguilliformes). *Dana Report* 88: 1–71.

Nielsen JG, Schwarzhans W, Uiblein F. 2014. Review of the Indo-West Pacific ophidiid genera *Sirembo* and *Spottobrotula* (Ophidiiformes, Ophidiidae), with description of three new species. *Marine Biology Research* 11(2) [2015]: 113–134. http://dx.doi.org/10.1080/17451000 .2014.904885

Nielsen JG, Uiblein F. **1993**. Tiefenwasser- und Tiefseefische aus dem Roten Meer. XVI. A new species of *Neobythites* from the NW Indian Ocean and the Red Sea (Pisces: Ophidiiformes: Ophidiidae). *Senckenbergiana Maritima* 23(4/6): 109–113.

Nielsen JG, Uiblein F. **2014**. The West Pacific *Neobythites bimarginatus* (Ophidiidae) recorded from off Madagascar. *Cybium* 38(4): 309–310. https://doi.org/10.26028/ cybium/2014-384-010

Ninni E. **1934**. I *Callionymus* dei mari d'Europa. Con un'Aggiunta di quelli esotici esistenti nei Musei d'Italia ed una nuova specie di *Callionymus* del Mar Rosso. *Notas y resúmenes, Instituto Español de Oceanografía* (Madrid) (Serie 2) 85: 1–59.

Nishikawa Y, Nakamura I. **1978**. Postlarvae and juveniles of the gempylid fish *Neoepinnula orientalis* (Gilchrist and von Bonde) from the north Arabian Sea. *Bulletin of the Far Seas Fisheries Research Laboratory* 16: 75–91.

Nolf D. **1985**. *Otolithi piscium*. In: Schultze HP (ed) *Handbook of paleoichthyology*. Vol. 10. Stuttgart and New York: Gustav Fischer Verlag. 145 pp.

Norman JR. **1922**. Three new fishes from Zululand and Natal, collected by Mr. H.W. Bell Marley; with additions to the fish fauna of Natal. *Annals and Magazine of Natural History* (Series 9) 9(52): 318–322.

Norman JR. **1926**. A synopsis of the rays of the family Rhinobatidae, with a revision of the genus *Rhinobatus*. *Proceedings of the Zoological Society of London* 1926(4): 941–982. Norman JR. **1927**. Zoological results of the Cambridge Expedition to the Suez Canal, 1924. Report on the fishes. *Transactions of the Zoological Society of London* 22(part 3, no.12): 375–390.

Norman JR. **1927**. The flatfishes (Heterosomata) of India, with a list of the specimens in the Indian Museum. Part I. *Records of the Indian Museum* (Calcutta) 29(1): 7–48. Pls. 2–7.

Norman JR. **1928**. The flatfishes (Heterosomata) of India, with a list of the specimens in the Indian Museum. Part II. *Records of the Indian Museum* (Calcutta) 30(2): 173–215.

Norman JR. **1929**. Note on the fishes of the Suez Canal. *Proceedings of the Zoological Society of London* 1930: 615–616.

Norman JR. **1930**. Oceanic fishes and flatfishes collected in 1925–27. *Discovery Reports* 2: 261–369.

Norman JR. **1931**. Four new fishes from the Gold Coast. *Annals and Magazine of Natural History* (Series 10) 7(40): 352–359.

Norman JR. 1931. Notes on flatfishes (Heterosomata). I. Notes of flatfishes of the family Bothidae in the British Museum, with descriptions of three new species. *Annals and Magazine of Natural History* (Series 10) 8(47): 507–510.

Norman JR. **1934**. *A systematic monograph of the flatfishes (Heterosomata). Vol. 1. Psettodidae, Bothidae, Pleuronectidae.* London: British Museum, Natural History. viii + 459 pp.

Norman JR. **1935**. Coast Fishes. Part 1. The south Atlantic. *Discovery Reports* 12: 1–58.

Norman JR. **1935**. Notes on the fishes of the family Scorpaenidae recorded from the British coasts, with a description of a new species. *Proceedings of the Zoological Society of London* 1935(3): 611–614.

Norman JR. **1935**. The European and South African sea breams of the genus *Spondyliosoma* and related genera. *Annals of the South African Museum* 32(1:2): 5–22.

Norman JR. **1937**. Fishes. British Australian New Zealand Antarctic Research Expedition Reports (Series B) 1(2): 51–88.

Norman JR. **1937**. Coast fishes. Part 2. The Patagonian region. *Discovery Reports* 16: 1–150.

Norman JR. 1939. Fishes. The John Murray Expedition 1933– 34. Scientific Reports, John Murray Expedition 7(1): 1–116.

Norman JR. **1944**. Notes on the blennioid fishes. I. A provisional synopsis of the genera of the family Blenniidae. *Annals and Magazine of Natural History* (Series 11) 10(72) [for 1943]: 793–812.

Norman JR. **1957**. *A draft synopsis of the orders, families and genera of recent fish and fish-like vertebrates*. London: British Museum (Natural History). 649 pp.

Normark BB, McCune AR, Harrison RG. **1991**. Phylogenetic relationships of neopterygian fishes, inferred from mitochondrial DNA sequences. *Molecular Biology and Evolution* 8(6): 819–334.

Notarbartolo di Sciara G. 1987. A revisionary study of the genus *Mobula* Rafinesque, 1810 (Chondrichthyes: Mobulidae) with the description of a new species. *Zoological Journal of the Linnean Society* 91(1): 1–91.

Notarbartolo di Sciara G. **1988**. Natural history of the rays of the genus *Mobula* in the Gulf of California. *Fisheries Bulletin* 86(1): 45–66.

Notarbartolo di Sciara G, Adnet S. Bennett M, Broadhurst MK, Fernando D, Jabado RW, Laglbauer BJL, Stevens G. 2019.
Taxonomic status, biological notes, and conservation of the longhorned pygmy devil ray *Mobula eregoodoo* (Cantor, 1849). *Aquatic Conservation: Marine and Freshwater Ecosystems* 29(10): 1–19. https://doi.org/10.1002/aqc.3230

Notarbartolo di Sciara G, Fernando D, Adnet S, Cappetta H, Jabado RW. **2017**. Devil rays (Chondrichthyes: *Mobula*) of the Arabian Seas, with a redescription of *Mobula kuhlii* (Valenciennes in Müller and Henle, 1841). *Aquatic Conservation: Marine and Freshwater Ecosystems* 27: 197–218. https://doi.org/10.1002/aqc.2635

Nouguier J, Refait D. **1990**. *Poissons de l'Océan Indien: les Iles Maldives*. Paris: Réalisations Editoriales Pédagogiques. 304 pp.

Novikov NP. **2002**. On the ecology of the African chimaera *Hydrolagus africanus* (Gilchrist) of the Madagascar and Mozambique Seamounts. *Journal of Ichthyology* 42(3): 271–274.

Nozaki M, Ichikawa T, Tsuneki K, Kobayashi H. **2000**. Seasonal development of gonads of the hagfish, *Eptatretus burgeri*, correlated with their seasonal migration. *Zoological Science* 17(2): 225–232.

Ntiba MJ, Jaccarini V. **1988**. Age and growth parameters of *Siganus sutor* in Kenyan marine inshore water, derived from numbers of otolith mircobands and fish lengths. *Journal of Fish Biology* 33(3): 465–470.

Nunes JLS, Piorski NM. **2009**. A dorsal fold in *Gymnura micrura* (Bloch and Scheneider, 1801) (Chondrichthyes: Gymnuridae). *Brazilian Archives of Biology and Technology* 52(2): 479–482.

http://dx.doi.org/10.1590/S1516-89132009000200027

Nursall JR. 1996. The phylogeny of pycnodont fishes. (pp. 125–152). In: Arratia G, Viohl G (eds) Mesozoic Fishes – Systematics and paleoecology. Proceedings of the 1<sup>st</sup> International Meeting on Mesozoic Fishes, Eichstätt, 1993. Munich: Verlag Dr. Friedrich Pfeil. 576 pp.

Nursall JR. 1999. The pycnodontiform bauplan: the morphology of a successful taxon (pp. 189–214). In: Arratia G, Schultze H-P (eds) *Mesozoic Fishes 2 – Systematics and fossil record. Proceedings of the international meeting, Buckow, 1997.* Munich: Verlag Dr. Friedrich Pfeil. 604 pp.

Nursall JR. **2010**. The case for pycnodont fishes as the fossil sister-group of teleosts (pp. 37–60). In: Nelson JS, Schultze H-P, Wilson MVH (eds) *Origin and phylogenetic* 

*interrelationships of teleosts*. Munich: Verlag Dr. Friedrich Pfeil. 482 pp.

- Nybelin O. **1966**. On certain Triassic and Liassic representatives of the family Pholidophoridae s. str. *Bulletin of the British Museum (Natural History) Geology* 11(8): 351–432.
- Nybelin O. **1971**. Versuch einer taxonomischen Revision der *Anaethalion*-Arten des Weissjura Deutschlands. *Acta Regiae Societatis Scientiarum et Litterarum Gothoburgensis* (Zoologica) 2: 1–53.
- Nyström E. **1887**. Redogörelse for den Japanska Fisksamlingen i Upsala Universitets Zoologiska Museum. Bihang till Kongl. *Svenska vetenskaps-akademiens handlingar* (Stockholm) 13(pt 4, no. 4): 1–54.
- Nyunja J, Ntiba M, Onyari J, Mavut K, Soetaert K, Bouillon S. 2009. Carbon sources supporting a diverse fish community in a tropical coastal ecosystem (Gazi Bay, Kenya). *Estuarine, Coastal and Shelf Science* 83(3): 333–341. http://dx.doi. org/10.1016/j.ecss.2009.01.009

# 0

- Obura D[O]. **2003**. Pemba, Mozambique. Mozambique Coral Reef Monitoring Program. NICOA-CDS/WWF/CORDIO East Africa. Coral species survey, Pemba, Mozambique. [unpaginated]
- Obura D[O], Church J. **2004**. Coral reef monitoring in Kiunga Marine Reserve, Kenya. 1998–2003. CORDIO SAREC Marine Science Program, Stockholm University. [unpaginated]
- Obura D[O], Machano H. **2004**. Coral species survey, Mafia Island Marine Park. CORDIO SAREC Marine Science Program, Stockholm University. 16 pp.
- Obura DO, Mbije N, Church J, Ngowo R, King A, Nur M.
  2006. Ecological status of coral reefs of the Mnazi Bay Ruvuma Estuary Marine Park, Tanzania (pp. 1077–1086). Proceedings of 10<sup>th</sup> International Coral Reef Symposium, Okinawa, Japan, June 27 to July 2, 2004. 1997 pp.
- Obura DO, Muthiga NA, Watson M. **2000**. Chapter 7. Kenya (pp. 199–229). In: McClanahan TR, Sheppard CRC, Obura DO (eds) *Coral reefs of the Indian Ocean: their ecology and conservation*. Oxford: Oxford University Press. xxiii + 525 pp.
- Obura D[O], Uku JN, Wawiye OP, Mwachireya S, Mdodo R.
  2000. Kenya, reef status and ecology (pp. 25–34). In: Souter D, Obura D, Lindén O (eds) *Coral reef degradation in the Indian Ocean status report*. CORDIO SAREC Marine Science Program, Stockholm University. 208 pp.
- Ochiai A. **1963**. *Fauna Japonica*. *Soleinae (Pisces)*. Tokyo: Biogeographical Society of Japan. 114 pp.
- Ochiai A, Okada K. **1966**. On the two allied red gurnards referable to *Chelidonichthys* from the Pacific Ocean. *Bulletin of the Misaki Marine Biological Institute* (Kyoto University) 9: 1–6.

- Oelschlager HA. **1974**. Ergebnisse der Forschungsreisen des FFS *Walther Herwig* nach Südamerika. XXXI. Das Jugendstadium van *Lampris guttatus* (Brünnich, 1788) (Osteichthyes, Allotriognathi), ein Beitrag zur Kenntnis seiner Entwicklung. *Archiv für Fischereiwissenschaft* 25(Beih. 1): 3–19.
- Ogilby JD. **1888**. Description of a new genus and species of deep-sea fish from Lord Howe Island. *Proceedings of the Linnean Society of New South Wales* (Series 2) 3(3): 1313.
- Ogilby JD. **1889**. The reptiles and fishes of Lord Howe Island. *Memoirs of the Australian Museum* 2(art.3): 49–74.
- Ogilby JD. **1890**. Description of a new *Tetrodon* from New South Wales. *Records of the Australian Museum* 1(4): 81–82.
- Ogilby JD. **1895**. On two new genera and species of fishes from Australia. *Proceedings of the Linnean Society of New South Wales* (Series 2) 10(2): 320–324.
- Ogilby JD. **1897**. Some new genera and species of fishes. *Proceedings of the Linnean Society of New South Wales* 22(2): 245–257.
- Ogilby JD. **1897**. On a *Trachypterus* from New South Wales. *Proceedings of the Linnean Society of New South Wales* 22(3): 646–659.
- Ogilby JD. **1898**. New genera and species of fishes. *Proceedings* of the Linnean Society of New South Wales 23(1): 32–41.
- Ogilby JD. **1899**. Additions to the fauna of Lord Howe Island. *Proceedings of the Linnean Society of New South Wales* 23(4): 730–745.
- Ogilby JD. **1903**. Studies in the ichthyology of Queensland. *Proceedings of the Royal Society of Queensland* 18: 7–27.
- Ogilby JD. **1908**. New or little known fishes in the Queensland Museum. *Annals of the Queensland Museum* 9(pt 1): 1–41.
- Ogilby JD. **1908**. On new genera and species of fishes. *Proceedings of the Royal Society of Queensland* 21: 1–26.
- Ogilby JD. **1910**. On new or insufficiently described fishes. *Proceedings of the Royal Society of Queensland* 23: 1–55.
- Ogilby JD. **1910**. On some new fishes from the Queensland coast. *Endeavour* Series 1: 85–139.
- Ogilby JD. **1912**. On some Queensland fishes. *Memoirs of the Queensland Museum* 1: 26–65.
- Ogilby JD. **1913**. On six new or rare Queensland fishes. *Memoirs of the Queensland Museum* 2: 81–89.
- Ogilby JD. **1915**. Edible fishes of Queensland. Part 3. Carangidae. *Memoirs of the Queensland Museum* 3: 99–116.
- Ohnishi N, Yanagisawa Y, Kohda M. **1997**. Sneaking by harem masters of the sandperch, *Parapercis snyderi*. *Environmental Biology of Fishes* 50(2): 217–223.
- Ohta I, Tachihara K. **2004**. Larval development and food habitats of the marbled parrotfish, *Leptoscarus vaigiensis*, associated with drifting algae. *Ichthyological Research* 51(1): 63–69.
- Ojeda RFR. **1978**. *Apterygopectus avilesi* nuevo género y nueva especie de lenguado para aguas australes chilenas (Pisces: Pleuronectiformes). *Noticiaro Mensual* (Museo Nacional de Histoire Natural, Santiago, Chile) 23(267): 3–10.

Okada Y, Suzuki K. **1952**. A new blennioid fish from the Sea of Japan. *Report of the Faculty of Fisheries, Prefectural University of Mie* 1(2): 75–77.

Okada Y, Suzuki K. **1956**. On the similarity of the osteological characters found between Owstoniidae and Cepolidae. *Report of the Faculty of Fisheries, Prefectural University of Mie* 2(2): 185–194.

Okada Y, Suzuki K. **1958**. Notes on the young of rare fish *Taractes steindachneri* (Döderlein). *Report of the Faculty of Fisheries, Prefectural University of Mie* 2(2): 195–198.

Okamoto M. **2012**. Two new species of the genus *Epigonus* (Perciformes: Epigonidae) from the South Pacific, with a definition of the *Epigonus constanciae* group. *Ichthyological Research* 59(3): 242–254. http://dx.doi.org/10.1007/s10228-012-0284-0

Okamoto M, Bartsch P, Motomura H. **2012**. *Epigonus merleni*, a junior synonym of *Epigonus macrops* (Actinopterygii: Perciformes: Epigonidae). *Species Diversity* 17(2): 123–126. http://dx.doi.org/10.12782/sd.17.2.123

Okamoto M, Golani D. **2017**. Three new species of the genus *Acropoma* (Perciformes: Acropomatidae) from the Indian Ocean. *Ichthyological Research* 65(1): 101–114. http://dx.doi. org/10.1007/s10228-017-0595-2

Okamoto M, Motomura H. **2011**. *Epigonus carbonarius*, a new species of deepwater cardinalfish (Perciformes: Epigonidae) from the Marquesas Islands, with a redefinition of the *Epigonus oligolepis* group. *Ichthyological Research* 58(2): 155–160. http://dx.doi.org/10.1007/s10228-011-0205-7

Okamoto M, Motomura H. **2013**. Two new species of deepwater cardinalfish from the Indo-Pacific, with a definition of the *Epigonus pandionis* group (Perciformes: Epigonidae). *Ichthyological Research* 60(4): 301–311. http://dx.doi.org/10.1007/s10228-013-0352-0

Okamura O. **1970**. *Fauna Japonica. Macrourina (Pisces)*. Tokyo: Academic Press of Japan. 216 pp.

Okamura O. **1970**. Studies on the macrouroid fishes of Japan: morphology, ecology and phylogeny. *Reports of the USA Marine Biological Station* 17(1–2): 1–179.

Okamura O. 1984. Parapercis somaliensis Schultz (p 291). In: Masuda H, Amaoka K, Araga C, Uyeno T, Yoshino T (eds) The fishes of the Japanese archipelago. Tokyo: Tokai University Press. 437 pp.

Okamura O (ed). **1985**. *Fishes of the Okinawa Trough and the adjacent waters. The intensive research of unexploited fishery resources on continental slopes.* Vol. II. Tokyo: Japan Fisheries Resource Conservation Association. 364 pp (pp. 418–781). [In: Japanese & English]

Okamura O (ed). **1986**. *Fishes of the East China Sea and the Yellow Sea*. Nagasaki: Sekai Regional Fisheries Research Laboratory. 501 pp.

Okamura O, Amaoka K. **1997**. *Sea fishes of Japan*. Tokyo: Yama-Kei Publishers. 784 pp. Okamura O, Amaoka K, Mitani F (eds). **1982**. *Fishes of the Kyushu-Palau Ridge and Tosa Bay. The intensive research of unexploited fishery resources on continental slopes.* Tokyo: Japan Fisheries Resource Conservation Association. 435 pp.

Okamura O, Kitajima T (eds). **1984**. *Fishes of the Okinawa Trough and the adjacent waters. The intensive research of unexploited fishery resources on continental slopes*. Vol. I. Tokyo: Japan Fisheries Resource Conservation Association. 414 pp. [In Japanese & English]

Oken L. **1816**. *Lehrbuch der Naturgeschicte*. Vol. 3. Zoologie. Leipzig. 1270 pp.

Oken L. 1817. Cuviers und Okens Zoologien neben einander gestellt. Isis, oder Encyclo. Zeitung von Oken 8(148): 1779– 1782 [for 1179–1182].

Okiyama M. **1972**. Morphology and identification of the young ipnopid, "*Macristiella*" from the tropical western Pacific. *Japanese Journal of Ichthyology* 19(3): 145–153.

Olfers JFM von. **1831**. *Die Gattung Torpedo in ihren naturhistorischen und antiquarischen Bezeichnungen erläutert.* Berlin: K. Akademie der Wissenschaft. 35 pp.

Olivar M-P, Fortuño JM. **1991**. Guide to ichthyoplankton of the southeast Atlantic (Benguela Current region). *Scientia Marina* 55(1): 1–383.

Olney JE, Johnson GD, Baldwin CC. 1993. Phylogeny of lampridiform fishes. *Bulletin of Marine Science* 52(1): 137–169.

Olney JE, Markle DF. **1979**. Description and occurrence of vexillifer larvae of *Echiodon* (Pisces: Carapidae) in the western North Atlantic and notes on other carapid vexillifers. *Bulletin of Marine Science* 29(3): 365–379.

Opic P, Conand F, Bourret P. **1994**. *Poissons commerciaux du Sud-Ouest de l'océan Indien*. Paris: Orstom Editions. 91 pp.

Oren OH. **1957**. Changes in temperature of the eastern Mediterranean Sea in relation to the catch of the Israel trawl fishery during the years 1954–55 and 1955–56. *Bulletin de l'Institut Océanographique* (Monaco) 1102: 1–15.

Ormond RFG. **1980**. Aggressive mimicry and other interspecific feeding associations among Red Sea coral reef predators. *Journal of Zoology* (London) 191: 247–262.

Orr JW, Fritzsche RA. **1993**. Revision of the ghost pipefishes, family Solenostomidae (Teleostei: Syngnathoidei). *Copeia* 1993(1): 168–182.

Orr JW, Fritzsche RA, Randall JE. **2002**. *Solenostomus halimeda*, a new species of ghost pipefish (Teleostei: Gasterosteiformes) from the Indo-Pacific, with a revised key to the known species of the family Solenostomidae. *aqua*, *Journal of Ichthyology and Aquatic Biology* 5(3): 99–108.

Orrell TM, Carpenter KE. **2004**. A phylogeny of the fish family Sparidae (porgies) inferred from mitochondrial sequence data. *Molecular Phylogenetics and Evolution* 32(2): 425–434. Orrell TM, Carpenter KE, Musick JA, Graves JE. **2002**. A phylogenetic and biogeographic analysis of the Sparidae (Perciformes: Percoidei) based on cytochrome *b* sequences. *Copeia* 2002(3): 618–631.

Orrell TM, Collette BB, Johnson GD. **2006**. Molecular data support separate scombroid and xiphioid clades. *Bulletin of Marine Science* 79(3): 505–519.

Orts S. **1993**. *Field guide to Seychelles commercial fishes: Lutjanidae, Lethrinidae, Serranidae: their range in the southwestern Indian Ocean*. Victoria, Seychelles: Seychelles Fishing Authority. 171 pp.

Osbeck P. **1765**. *Reise nach Ostindien und China*. Rostock, 552 pp. [Translated from the Swedish by JG Georgi]

Osorio B. **1894**. D'algunas especies a juntar ao catalogo dos peixes de Portugal de Capello. *Jornal do Sciências Mathemáticas, Physicas e Naturaes, Lisboa* (Series 2) 3(11): 186–188.

Osorio B. **1917**. Nota sôbre algumas espécies de peixes que vivem no Atlântico ocidental. *Arquivo da Universidade de Lisboa* 4: 103–131.

Otero O. **2004**. Anatomy, systematics and phylogeny of both Recent and fossil latid fishes (Teleostei, Perciformes, Latidae). *Zoological Journal of the Linnean Society* 141: 81–133.

O'Toole MJ. **1976**. Incidental collections of small and juvenile fishes from egg and larval surveys off South West Africa (1972–1974). *Fisheries Bulletin* (South Africa) 8: 23–33.

O'Toole MJ. **1977**. Development and distributional ecology of the larvae of the West Coast sole, *Austroglossus microlepis*. *Fisheries Bulletin* (South Africa) 9: 32–45.

Oyugi D. **2005**. Preliminary investigations on the ichthyodiversity of Kilifi Creek, Kenya. *Western Indian Ocean Journal of Marine Science* 4(1): 11–20.

Ozawa T, Fukui A. **1986**. Studies on the development and distribution of the bothid larvae in the Western North Pacific (pp. 321–423). In: Ozawa T (ed) *Studies on the oceanic ichthyoplankton in the Western North Pacific*. Fukuoka: Kyushu University Press. 430 pp.

### Ρ

Paepke H-J, Fricke R. 1992. Kritischer Katalog der Typen der Fischsammlung des Zoologischen Museums Berlin.
Teil 4: Scorpaeniformes. *Mitteilungen aus dem Zoologischen Museum in Berlin* 68(2): 267–293.

Paepke H-J, Schmidt K. 1988. Kritischer Katalog der Typen der Fischsammlung des Zoologischen Museums Berlin.
Teil 2: Agnatha, Chondrichthyes. *Mitteilungen aus dem Zoologischen Museum in Berlin* 64(1): 155–189.

Pakhorukov NP. **2001**. Distribution and behavior of the bottom and demersal fishes of the Rio Grande Plateau (the Atlantic Ocean). *Journal of Ichthyology* 41(4): 300–307.

Palko BJ, Beardsley GL, Richards WJ. **1981**. Synopsis of the biology of the swordfish, *Xiphias gladius* Linnaeus. *NOAA Technical Report* NMFS Circular 441: 1–21.

Palko BJ, Beardsley GL, Richards WJ. **1982**. Synopsis of the biological data on dolphin-fishes, *Coryphaena hippurus* Linnaeus and *Coryphaena equiselis* Linnaeus. *NOAA Technical Report* NMFS Circular 443: 1–28.

Pallas PS. **1767**. [Description of *Sciaena jaculatrix*. Appended to a letter from Dr. John Albert Schlosser.] *Philosophical Transactions of the Royal Society of London* 56(1766): 186–188.

Pallas PS. 1767–1780. Spicilegia Zoologica quibus novae imprimis et obscurae animalium species iconibus, descriptionibus atque commentariis illustrantur. Berolini: Gottl. August. Lange. [Pisces: Vol. 1 (1769; fasc. vii), pp. 1–42, Pls. 1–6; Vol. 1 (1770, fasc. viii): 1–56, Pls. 1–5.]

Pallas PS. 1814. Zoographia Rosso-Asiatica, sistens omnium animalium in extenso Imperio Rossico et adjacentibus maribus observatorum recensionem, domicilia, mores et descriptiones, anatomen atque icones plurimorum. Vol. 3. Petropoli. 428 pp.

Palmer G. **1950**. Additions to the fish fauna of Christmas Island, Indian Ocean. *Bulletin of the Raffles Museum* 23: 200–205.

Palmer G. 1961. The dealfishes (Trachipteridae) of the Mediterranean and north-east Atlantic. Bulletin of the British Museum (Natural History) Zoology 7(7): 335–351.

Palmer G. **1970**. New records, and one new species, of teleost fishes from the Gilbert Islands. *Bulletin of the British Museum (Natural History) Zoology* 19(6): 213–234.

Palmer G. 1973. Lamprididae (p. 328). In: Hureau J-C, Monod T (eds) Check-list of the fishes of the north-eastern Atlantic and of the Mediterranean (CLOFNAM). Vol. I. Paris: UNESCO.

Palmer G, Oelschläger HA. 1976. Use of the name *Lampris* gutatus (Brunnich, 1788) in preference to *Lampris regius* (Bonnaterre, 1788) for the Opah. *Copeia* 1976(2): 366–367.

Palsson WA, Pietsch TW. 1989. Revision of the acanthopterygian fish family Pegasidae (Order Gasterosteiformes). *Indo-Pacific Fishes* 18: 1–38, Pl. 1.

Pandy AK, Sandhu GS. 1992. Encyclopaedia of fishes and fisheries of India. Vol. 6. Identification of bony fishes. New Delhi: Anmol Publication. 385 pp.

Pappe CWL. 1853. Synopsis of the edible fishes of the Cape of Good Hope. Cape Town: Van de Sandt de Villiers & Tier. 34 pp.

Pappenheim P. 1914. II. Die Tiefseefische. In: Die Fische der deutschen S\u00fcdpolar-Expedition 1901–1903. Vol. 15 (Zoologie v. 7): 161–200.

Pardo BG, Machordom A, Foresti F, Porto-Foresti F, Azevedo MF, Banon R, Sanchez L, Martinez P. **2005**. Phylogenetic analysis of flatfish (Order Pleuronectiformes) based on mitochondrial 16s rDNA sequences. *Scientia Marina* 69(4): 531–43.

Parenti LR. 1987. Phylogenetic aspects of tooth and jaw structure of the medaka, *Oryzias latipes*, and other beloniform fishes. *Journal of Zoology* 211(3): 561–572.

Parenti LR. **1993**. Relationships of atherinomorph fishes (Teleostei). *Bulletin of Marine Science* 52(1): 170–196.

Parenti LR. 1999. Order Beloniformes. Adrianichthyidae.
Ricefishes (pp. 2149–2150). In: Carpenter KE, Niem VH (eds) FAO species identification guide for fishery purposes. The living marine resources of the western central Pacific.
Vol. 4. Bony fishes part 2 (pp. 2069–2790). Rome: FAO.

Parenti LR. 2005. The phylogeny of atherinomorphs: evolution of a novel fish reproductive system (pp. 13–30). In: Uribe MC, Grier HJ (eds) *Viviparous fishes*. Mexico: New Life Publications. 603 pp.

Parenti LR. **2008**. A phylogenetic analysis and taxonomic revision of ricefishes, *Oryzias* and relatives (Beloniformes, Adrianichthyidae). *Zoological Journal of the Linnean Society* 154(3): 494–610. http://dx.doi.org/10.1111/j.1096-3642.2008.00417.x

Parenti LR, Grier HJ. **2004**. Evolution and phylogeny of gonad morphology in bony fishes. *Integrative and Comparative Biology* 44(5): 333–348.

Parenti P. **2003**. On the status of species classified in the genus *Perca* by Johann Julius Walbaum (1792). *Zoological Studies* 42(4): 491–505.

Parenti P, Randall JE. **1998**. First record of the labrid fish *Pteragogus flagellifer* (Valenciennes, 1839) from the Red Sea. *Fauna of Arabia* 17: 473–475.

Parenti P, Randall JE. **2000**. An annotated checklist of the species of the Labroid fish families Labridae and Scaridae. *Ichthyological Bulletin of the J.L.B. Smith Institute of Ichthyology* 68: 1–97.

Parenti P, Randall JE. **2011**. Checklist of the species of the families Labridae and Scaridae: an update. *Smithiana* Bulletin 13: 29–44.

Parenti P, Randall JE. **2018**. A checklist of wrasses (Labridae) and parrotfishes (Scaridae) of the world: 2017 update. *Journal of the Ocean Science Foundation* 30: 11–27. https://doi.org/10.5281/zenodo.1247552

Parin NV. **1958**. A new species of flying fish from the western Pacific Ocean – *Cypselurus vitiazi* Parin, sp. n. (Pisces Exocoetidae). *Zoologicheskii Zhurnal* 37(9): 1412–1416. [In Russian, English summary]

Parin NV. **1960**. Distribution of flying fishes (family Exocoetidae) in the western and central parts of the Pacific Ocean. *Trudy Instituta Okeanologii* 41: 153–162. [In Russian, English summary]

Parin NV. **1960**. The flying fishes (Exocoetidae) of the Northwest Pacific. *Trudy Instituta Okeanologii* 31: 205–285. [In Russian, English summary] Parin NV. **1961**. A contribution to the knowledge of the flyingfish fauna (family Exocoetidae) of the Pacific and Indian oceans. *Trudy Instituta Okeanologii* 43: 40–91. [In Russian, English summary]

Parin NV. **1961**. Principles of classification of the flying fishes (families Oxyporhamphidae and Exocoetidae). *Trudy Instituta Okeanologii* 43: 92–183. [In Russian, English summary]

Parin NV. **1961**. *Oxyporhamphus meristocystis* (Pisces, Oxyporhamphidae) – a new species of flying halfbeak from the waters of the Indo-Malayan Archipelago. *Voprosy Ikhtiologii* 1(3[20]): 391–394. [In Russian, English summary]

Parin NV. 1964. Taxonomic status, geographic variation, and distribution of the oceanic halfbeak *Euleptorhamphus viridis* (van Hasselt) (Hemirhamphidae, Pisces). *Trudy Instituta Okeanologii* 73: 185–203. [In Russian, English summary]

Parin NV. **1967**. Review of the marine belonids [needlefishes] of the western Pacific and Indian oceans. *Trudy Instituta Okeanologii* 84: 3–83. [In Russian, English summary]

Parin NV. **1970**. *Ichthyofauna of the epipelagic zone*. Jerusalem: Israel Program for Scientific Translations. 206 pp. [Translated from the 1968 Russian]

Parin NV. **1982**. New species of the genus *Draconetta* and a key to the species of the family Draconettidae (Osteichthyes). *Zoologicheskii Zhurnal* 61(4): 554–563. [In Russian, English summary]

Parin NV. **1983**. *Aphanopus mikhailini* sp. n. and *A. intermedius* sp. n. (Trichiuridae, Perciformes) two new scabbardfishes from the temperate waters of the southern hemisphere and the tropical Atlantic. *Journal of Ichthyology* 23(3): 1–12.

Parin NV. 1984. Exocoetidae. Flyingfishes. In: Fischer W,
Bianchi G (eds) FAO species identification sheets for fishery purposes. Western Indian Ocean (Fishing Area 51).
Vol. 2. Rome: FAO. [unpaginated]

Parin NV. **1984**. Oceanic ichthyologeography: an attempt to review the distribution and origin of pelagic and bottom fishes outside continental shelves and neritic zones. *Archiv für Fischereiwissenschaft* 35(1): 5–41.

Parin NV. **1987**. Species of spiny dogfish of the genus *Squalus* from southeastern Pacific seamounts. *Voprosy Ikhtiologii* 27(4): 531–538. [In Russian, English translation in *Journal of Ichthyology* 27(5): 43–50]

Parin NV. **1989**. A review of the genus *Rexea* (Gempylidae) with descriptions of three new species. *Voprosy Ikhtiologii* 29(1): 3–23. [In Russian, English translation in *Journal of Ichthyology* 29(2): 86–105]

Parin NV. **1990**. Percophid fishes (Percophidae) from the Sala y Gomez Ridge (Southeast Pacific). *Voprosy Ikhtiologii* 30(1): 3–12. [In Russian, English translation in *Journal of Ichthyology* 30(1): 68–79] Parin NV. **1991**. Three new species of the bentho-pelagic fish genus *Plagiogeneion* from the southern Pacific and Indian oceans (Teleostei: Emmelichthyidae). *Proceedings of the Biological Society of Washington* 104(3): 459–467.

Parin NV. **1996**. On the species composition of flying fishes (Exocoetidae) in the west-central part of the tropical Pacific. *Voprosy Ikhtiologii* 36(3): 300–307. [In Russian, English translation in *Journal of Ichthyology* 36(5): 357–364]

Parin NV. 1999. Exocoetidae. Flyingfishes (pp. 2162–2179). In: Carpenter KE, Niem VH (eds) FAO species identification guide for fishery purposes. The living marine resources of the western central Pacific. Vol. 4. Bony fishes part 2 (pp. 2069– 2790). Rome: FAO.

Parin NV. 2003. An annotated catalogue of fish-like vertebrates and fishes of the seas of Russia and adjacent countries: Part 3. Orders Perciformes (excluding suborders Gobioidei, Zoarcoidei and Stichaeoidei) and Tetraodontiformes. *Voprosy Ikhtiologii* 43(Suppl. 1): S1–S40. [Original text in English]

Parin NV, Abramov AA. **1986**. Materials for a revision of the genus *Epigonus* Rafinesque (Perciformes, Epigonidae): species from the submarine ridges of the southeastern Pacific and preliminary review of the "*Epigonus robustus* species-group". *Trudy Instituta Okeanologii* 121: 173–194. [In Russian, English summary]

Parin NV, Abramov AA. 1986. Two new species of benthopelagic fishes of the genus *Epigonus* (Apogonidae) from the western tropical part of the Indian Ocean. *Byulleten Moskovskogo Obshchestva Ispytatelei Prirody Otdel Biologicheskii* [Bulletin of the Moscow Society of Naturalists Biological Series] 91(3): 53–57. [In Russian]

Parin NV, Becker VE. 1970. Materials for a revision of the trichiuroid fishes of the genus *Benthodesmus*, with the description of four new species and one new subspecies. *Proceedings of the Biological Society of Washington* 83(33): 351–364.

Parin NV, Becker VE. 1972. Materials on taxonomy and distribution of some trichiuroid fishes (Pisces: Trichiuroidae, Scombrolabracidae, Gempylidae and Trichiuridae). *Trudy Instituta Okeanologii* 93: 110–204. [In Russian, English summary]

Parin NV, Bekker VE. **1973**. Gempylidae (pp. 457–460). In: Hureau J-C, Monod T (eds) *Check-list of the fishes of the north-eastern Atlantic and of the Mediterranean (CLOFNAM)*. Vol. I. Paris: UNESCO. [author also as Becker]

Parin NV, Belyanina TN. 1998. Age and geographical variability and distribution of *Cheilopogon furcatus* (Exocoetidae, Beloniformes), with descriptions of two new subspecies. *Voprosy Ikhtiologii* 38(5): 581–597. [In Russian, English translation in *Journal of Ichthyology* 38(8): 557–573]

Parin NV, Belyanina TN. **2002**. A review of flying fishes of the subgenus *Danichthys* (genus *Hirundichthys*, Exocoetidae). *Journal of Ichthyology* 42(Suppl. 1): S23–S44.

Parin NV, Belyanina TN. 2007. On the distribution of *Glossanodon mildredae* (Argentinidae) in the Indian Ocean. *Voprosy Ikhtiologii* 47(3): 425–426. [In Russian, English translation in *Journal of Ichthyology* 47(5): 408–409.]

Parin NV, Besednov LN. **1965**. Flying fishes (Oxyporhamphidae and Exocoetidae) in the Gulf of Tonkin. *Trudy Instituta Okeanologii* 80: 104–117.

Parin NV, Bogorodsky SV. 2011. Distribution and morphology of the flying fish *Cypselurus hexazona* placed in a separate subgenus *Zonocypselurus* subgen. nov. *Voprosy Ikhtiologii* 51(5): 683–686. [In Russian, English translation in *Journal* of *Ichthyology* 51(8): 658–661] http://dx.doi.org/10.1134/ S0032945211050110

Parin NV, Borodulina OD. 1986. Preliminary review of the bathypelagic fish genus *Antigonia* Lowe (Zeiformes, Caproidae). *Trudy Instituta Okeanologii* 121: 141–172. [In Russian, English summary]

Parin NV, Borodulina OD. **2005**. *Antigonia* (Caproidae) from the western Indian Ocean: 1. Species with nine spiny rays in the dorsal fin. *Journal of Ichthyology* 45(6): 417–428.

Parin NV, Borodulina OD. **2006**. Antigonias (*Antigonia*, Caproidae) of the western Indian Ocean: 2. Species with eight spiny rays in the dorsal fin. *Journal of Ichthyology* 46(3): 203–211.

Parin NV, Collette BB, Scherbachev YN. **1980**. Preliminary review of the marine halfbeaks (Hemiramphidae, Beloniformes) of the tropical Indo-West Pacific. *Trudy Instituta Okeanologii* 97: 7–173. [In Russian, English summary]

Parin NV, Fedoryako BI. 1981. On age variation, ecology and distribution of *Nomeus gronovii* Gmelin (Nomeidae, Stromateoidei) (pp. 94–98). In: Parin NV (ed) *Fishes of the open ocean*. Moscow: PP Shirshov Institute of Oceanology. 119 pp. [In Russian]

Parin NV, Kukuyev YI. **1983**. Reestablishment of the validity of *Lampris immaculata* Gilchrist and the geographical distribution of Lampridae. *Journal of Ichthyology* 23(1): 1–12.

Parin NV, Lakshminarayana D. **1993**. Flying fishes (Exocoetidae) from the coastal waters of southeastern India. *Voprosy Ikhtiologii* 33(1): 53–60.

Parin NV, Mikhailin SV. 1982. Tentoriceps cristatus (Klunzinger) (Trichiuridae) in the Indian Ocean (pp. 48–54). In: Parin NV (ed) Poorly known fishes of the open ocean. Moscow: Instituta Okeanologii. 140 pp. [In Russian]

Parin NV, Nesis KN, Sagaidachnyi AY, Shcherbachev YN.
1993. Fauna of Walters Seamount (southwestern part of the Indian Ocean). *Trudy Instituta Okeanologii* 128: 199–216.
[In Russian, English summary]

Parin NV, Permitin YY. **1969**. Materials on the pelagic fish fauna of the Atlantic. A new genus of stromateoid fishes, *Pseudocichthys* (Pisces: Centrolophidae). *Voprosy Ikhtiologii* 9(6): 981–987. [In Russian, English translation in *Problems in Ichthyology* 9(6): 789–794]

Parin NV, Piotrovsky AS. **2004**. Stromateoid fishes (suborder Stromateoidei) of the Indian Ocean (species composition, distribution, biology, and fisheries). *Journal of Ichthyology* 44(Suppl. 1): S33–S62.

Parin NV, Pokhil'skaya GN. 1968. Age variability and the distribution of a rare oceanic fish *Eumecichthys fiski* (Pisces, Lophotidae). *Voprosy Ikhtiologii* 8(6): 1015–1021. [In Russian, English translation in *Problems in Ichthyology* 8(6): 808–812]

Parin NV, Shakhovskoy IB. **2000**. A review of the flying fish genus *Exocoetus* (Exocoetidae) with descriptions of two new species from the southern Pacific Ocean. *Journal of Ichthyology* 40(Suppl. 1): S31–S63.

Parin NV, Shcherbachev YN. 1972. A new species of halfbeak – *Rhynchorhamphus arabicus* Parin et Shcherbachev (Beloniformes, Hemiramphidae) from south Yemen. *Voprosy Ikhtiologii* 12(3): 569–571. [In Russian, English translation in *Journal of Ichthyology* 12(3): 523–526]

Parin NV, Shcherbachev YN, Pakhorukov NP. **1995**. Bottom and near-bottom fishes of the Rio Grande Submarine Rise (southwestern part of the Atlantic Ocean). *Voprosy Ikhtiologii* 35(6): 740–747. [In Russian, English translation in *Journal of Ichthyology* 41: 300–307]

Parin NV, Sundaram S. **2011**. On the living colouration of flying fish *Cheilopogon abei* (Exocoetidae). *Problems of Fisheries* 12(2-46): 411–412. [In Russian]

Park M. **1797**. Descriptions of eight new fishes from Sumatra. *Transactions of the Linnean Society of London* 3(art.9): 33–38.

Parmentier É. **2004**. *Encheliophis chardewalli*: a new species of Carapidae (Ophidiiformes) from French Polynesia, with a redescription of *Encheliophis vermicularis*. *Copeia* 2004(1): 62–67.

Parmentier É, Bailly N. **2002**. New record of *Carapus dubius* (Carapidae) off Madagascar?. *Cybium* 26(1): 79–80.

Parmentier É, Castillo G, Chardon M, Vandewalle P. 2000.
Phylogenetic analysis of the pearlfish tribe Carapini (Pisces: Carapidae). *Acta Zoologica* (Stockholm) 81: 293–306.

Parmentier É, Lanterbecq D, Todesco M, Eeckhaut I. 2010. Unique morphologies of *Encheliophis vermiops* (Carapidae) with revised diagnosis of the genus. *Ichthyological Research* 57: 85–90. http://dx.doi.org/10.1007/s10228-009-0138-6

Parmentier M. **1988**. *Dendrochirus biocellatus* (Fowler, 1938), type de genre monotypique *Nemapterois* Fowler, 1938, considéré comme synonyme récent de *Dendrochirus* Swainson, 1839; spécimen à 3 ocelles photographié de nuit à Saint-Gilles (La Réunion). *Revue française d'Aquariologie Herpétologie* 15(4): 1. Parr AE. **1930**. Teleostean shore and shallow-water fishes from the Bahamas and Turks Island. Scientific results of the third oceanographic expedition of the *Pawnee* 1927. *Bulletin of the Bingham Oceanographic Collection* (Yale University) 3(art.4): 1–148.

Parr AE. **1933**. Deepsea Berycomorphi and Percomorphi from the waters around the Bahama and Bermuda Islands (scientific results of the third oceanographic expedition of the *Pawnee* 1927). *Bulletin of the Bingham Oceanographic Collection* (Yale University) 3(art.6): 1–51.

Parr AE. **1951**. Preliminary revision of the Alepocephalidae, with the introduction of a new family, Searsidae. *American Museum Novitates* 1531: 1–21.

Parrish BB, Sharmann DP. **1956**. Some remarks on methods used in herring "Racial" investigations, with special reference to otolith studies. *Rapports et procès-verbaux des Réunions* 143(2): 66–80.

Parrish RH, Serra R, Grant WS. **1989**. The monotypic sardine, *Sardina* and *Sardinops*: their taxonomy, distribution, stock structure, and zoogeography. *Canadian Journal of Fisheries and Aquatic Sciences* 46(11): 2019–2036.

Passerotti MS, Andrews AH, Carlson JK, Wintner SP, Goldman KJ, Natanson LJ. **2014**. Maximum age and missing time in the vertebrae of sand tiger shark (*Carcharias taurus*): validated lifespan from bomb radiocarbon dating in the western North Atlantic and southwestern Indian Oceans. *Marine and Freshwater Research* 65(8): 674–687. https://doi.org/10.1071/MF13214

Pati S. **1978**. Systematic comparison of the stromateid fishes of the genus *Pampus* Bonaparte (family Stromateidae). *Journal of the Marine Biological Association of India* 20(1–2): 50–64.

Patterson C. **1965**. The phylogeny of the chimaeroids. *Philosophical Transactions of the Royal Society of London* (Series B, Biological Sciences) 249(757): 101–219.

Patterson C. **1968**. The caudal skeleton in Lower Liassic pholidophorid fishes. *Bulletin of the British Museum* (*Natural History*) *Geology* 16(5): 201–239.

Patterson C. 1973. Interrelationships of holosteans (pp. 233–305). In: Greenwood PH, Miles RS, Patterson C (eds) *Interrelationships of fishes*. London: Academic Press. 536 pp.

Patterson C. **1975**. The braincase of pholidophorid and leptolepid fishes, with a review of the actinopterygian braincase. *Philosophical Transactions, Royal Society of London* (Series B) 269(899): 275–579.

Patterson C. **1975**. The distribution of Mesozoic freshwater fishes. *Mémoires du Muséum National d'Histoire Naturelle* (Paris) (N.S., Série A, Zoologie) 88: 156–174.

Patterson C. **1977**. The contribution of paleontology to teleostean phylogeny (pp. 579–643). In: Hecht MK, Goody PC, Hecht BM (eds) *Major patterns in vertebrate evolution*. New York: Plenum Press. 917 pp. Patterson C. 1994. Bony fishes (pp. 57–84). In: Prothero DR, Schoch RM (eds) *Major features of vertebrate evolution*. *Vol. 7. Short courses in paleontology*. Knoxville, Tennessee: Paleontological Society, University of Tennessee. 270 pp.

Patterson C, Johnson GD. **1995**. The intermuscular bones and ligaments of teleostean fishes. *Smithsonian Contributions to Zoology* 559: 1–83.

Patterson C, Rosen DE. 1977. Review of ichthyodectiform and other Mesozoic teleost fishes and the theory and practice of classifying fossils. *Bulletin of the American Museum of Natural History* 158(art.2): 83–172.

Pauca M. **1930**. Fische aus der Walfischbay Sudwestafrika. Annalen des Naturhistorischen Museums in Wien 44: 33–37.

Paulin CD. **1982**. Scorpionfishes of New Zealand (Pisces: Scorpaenidae). *New Zealand Journal of Zoology* 9: 437–450.

Paulin CD. **1995**. Description of a new genus and two new species of bythitid fishes, and a redescription of *Bidenichthys consobrinus* (Hutton) from New Zealand. *Journal of Natural History* 29(1): 249–258.

Paulin CD, Stewart AL, Roberts CD, McMillan PJ. 1989. New Zealand fish. A complete guide. Wellington: National Museum of New Zealand. Miscellaneous Series No. 19: xiv + 279 pp.

Paulus T. 1992. Syngnathus safina n. sp. and first record of S. macrophtalmus [sic] Duncker 1915 from the Gulf of Aqaba, Red Sea (Pisces: Osteichthyes: Syngnathidae). Senckenbergiana Biologica 72(1/3): 27–33.

Paulus T. 1999. Syngnathidae. Pipefishes and seahorses (pp. 2264–2276). In: Carpenter KE, Niem VH (eds) FAO species identification guide for fishery purposes. The living marine resources of the western central Pacific. Vol. 4. Bony fishes part 2 (pp. 2069–2790). Rome: FAO.

Pavan-Kumar A, Kumar R, Pitale P, Shen K-N, Borsa P. 2018. Neotrygon indica sp. nov., the Indian-Ocean blue spotted maskray (Myliobatoidei, Dasyatidae). Comptes Rendus Biologies 341(2): 120–130. http://dx.doi.org/10.1016/j. crvi.2018.01.004

Pavlov YP. **1989**. Some data on the morphometrics and distribution of species of the genus *Taractes* (Bramidae) in the Pacific Ocean. *Journal of Ichthyology* 29(6): 164–166.

Pavlov YP. **1991**. Information on morphometrics and ecology of pomfrets of the genus *Brama* in the southeastern Pacific Ocean. *Journal of Ichthyology* 31(2): 120–124.

Paxton JR, Hoese DF, Allen GR, Hanley JE. **1989**. *Zoological catalogue of Australia*. Vol. 7. Pisces. Petromyzontidae to Carangidae. Canberra: Australian Government Publishing Service. xii + 665 pp.

Pears RJ. **2005**. Comparative demography and assemblage structure of serranid fishes: implications for conservation and fisheries management. PhD thesis, James Cook University, Townsville. Pearson J. **1912**. Biological survey of Trincomalee harbour. *Spolia Zeylanica* 8: 30–40.

Peden AE, Ostermann W. **1980**. Three fish species previously unknown from marine waters off British Columbia. *Syesis* 13: 215–217.

Pellegrin J. 1898. Contribution à l'étude ichtyologique des îles Mariannes, d'après les envois de M. Marche. Bulletin du Muséum National d'Histoire Naturelle (Paris) 4(5): 228–229.

Pellegrin J. 1906. Sur un Salarias de la Baie de Tadjourah. Bulletin du Muséum National d'Histoire Naturelle (Paris) (Série 1) 12(2): 93–94.

Pellegrin J. 1914. Sur une collection de poissons de Madagascar. Bulletin de la Société Zoologique de France 39: 221–234.

Pellegrin J. **1919**. Sur les *Eleotris* des eaux douces de Madagascar. Description d'une espèce nouvelle. *Bulletin de la Société Zoologique de France* 44: 266–271.

Pellegrin J. 1932. Poissons de Madagascar recueillis par M. Waterlot. Description d'une variété nouvelle. *Bulletin de la Société Zoologique de France* 57: 225–228.

Pellegrin J. **1932**. Poissons de Madagascar recueillis par M. Decary. Description d'une variété nouvelle. *Bulletin de la Société Zoologique de France* 57: 291–297.

Pellegrin J. 1932. Poissons nouveaux de Madagascar recueillis par M. Catala. Bulletin de la Société Zoologique de France 57: 424–427.

Pellegrin J. 1933. Les poissons des eaux douces de Madagascar et des iles voisines (Comores, Seychelles, Mascareignes). Mémoires de l'Académie Malagache 14: 1–222.

Pellegrin J. **1935**. Poissons de Madagascar recueillis par M. Catala description d'un *Sicydium* nouveau. *Bulletin de la Société Zoologique de France* 60: 69–73.

Pellegrin J. **1935**. Poisson marin nouveau de la Réunion de la famille des Serranides. *Bulletin du Muséum National d'Histoire Naturelle* (Paris) (Série 2) 7(1): 51–53.

Pellegrin J, Chevey P. 1940. Poissons nouveaux ou rares de Cochinchine. Description de deux espèces et de deux variétés. *Bulletin de la Société Zoologique de France* 65: 153–158.

Peng Z, Diogo R, He S. 2009. Teleost fishes (Teleostei) (pp. 335–338). In: Hedges SB, Kumar S (eds) *The timetree of life*. New York: Oxford University Press. 551 pp.

Peng Z, He S, Wang J, Wang W, Diogo R. **2006**. Mitochondrial molecular clocks and the origin of the major Otocephalan clades (Pisces: Teleostei): a new insight. *Gene* 370: 113–124.

Pennant T. 1776. British zoology (4th edition). Vol. 3: Class III. Reptiles. Class IV. Fish. Benjamin White: London. 425 pp., Pls. 1–73.

Penrith MJ. **1964**. A marked extension of the known range of *Tetrapturus angustirostris* in the Indian Ocean. *Copeia* 1964(1): 231–232.

Penrith MJ. 1965. A new species of flatfish, Mancopsetta milfordi, from South Africa, with notes on the genus Mancopsetta. Annals of the South African Museum 48(7):181–188.

Penrith MJ. **1967**. The fishes of Tristan da Cunha, Gough Island and the Vema Seamount. *Annals of the South African Museum* 48(22): 523–548.

Penrith MJ. **1969**. New records of deep-water fishes from South West Africa. *Cimbebasia* (Series A) 1(3): 59–75.

Penrith MJ. **1972**. Earliest description and name for the whale shark. *Copeia* 1972(2): 362.

Penrith MJ. **1972**. Sex reversal in the sparid fish *Chrysoblephus laticeps*. *Koedoe* 15: 135–139.

Penrith MJ. **1972**. The behaviour of reef-dwelling sparid fishes. *Zoologica Africana* 7(1): 43–48.

Penrith MJ. **1976**. Distribution of shallow water marine fishes around southern Africa. *Cimbebasia* (Series A) 4(7): 137–154.

Penrith MJ. 1976. First confirmed record of *Lithognathus lithognathus* sympatric with *L. aureti* (Pisces: Sparidae). *Madoqua* 9(3): 53–54.

Penrith MJ. **1978**. An annotated check-list of the inshore fishes of southern Angola. *Cimbebasia* (Series A) 4(11): 179–190.

Penrith MJ. **1980**. Three additions to the marine fish fauna of southern Africa. *Madoqua* 12(1): 59–62.

Penrith MJ. **1982**. Southernmost records of *Pomadasys jubelini*. *Madoqua* 13(1): 91–93.

Penrith MJ. **1982**. Notes on marine fishes collected in the vicinity of Bosluisbaai. *Madoqua* 13(2): 159–168.

Penrith MJ, Cram DL. 1974. The Cape of Good Hope: a hidden barrier for billfishes. NOAA Technical Report NMFS SSRF-675 (Part 2): 175–187.

Penrith MJ, Penrith M-L. **1969**. A new species of *Lithognathus* (Pisces: Sparidae) from the northern coast of South West Africa. *Cimbebasia* (Series A) 1(5): 99–111.

Penrith MJ, Penrith M-L. **1971**. The status of *Batrichthys apiatus* (Cuvier & Valenciennes) (Pisces: Batrachoididae) with notes on four western southern African species of batrachoid fishes. *Cimbebasia* (Series A) 2(3): 46–52.

Penrith MJ, Penrith M-L. **1972**. The Blenniidae of western southern Africa. *Cimbebasia* (Series A) 2(5): 65–90.

Penrith MJ, Penrith M-L. **1972**. Redescription of *Pandaka silvana* (Barnard) (Pisces: Gobiidae). *Annals of the South African Museum* 60(2): 105–108.

Penrith MJ, Penrith M-L. **1983**. Range extensions of blennoid fishes on the southern African west coast. *South African Journal of Zoology* 18(2): 144–147.

Penrith MJ, Talbot FH. **1973**. An additional billfish (Pisces: Istiophoridae) recorded from South Africa. *South African Journal of Science* 69(4): 121–122. Penrith M-L. 1965. Studies on the South African Clinidae.
1. Description of a new species of *Pavoclinus*, and redescription of *Gynutoclinus rotundifrons* (Barnard). *Annals of the South African Museum* 48(10): 211–217.

Penrith M-L. **1965**. Note on an extension of the known ranges of distribution of some species of Clinidae (Pisces). *South African Journal of Science* 61(12): 425–426.

Penrith M-L. **1967**. Studies on the South African Clinidae. II. Two new species of *Clinus* from the Western Cape. *Annals of the South African Museum* 50(4): 43–59.

Penrith M-L. **1969**. *Apletodon pellegrini* (Chabanaud) and other clingfishes (Pisces: Gobiesocidae) from South West Africa. *Annals of the South African Museum* 55(2): 123–134.

Penrith M-L. **1969**. Pisces (pp. 202–236). In: Day JH *A guide to marine life on South African shores*. Cape Town: AA Balkema. 300 pp.

Penrith M-L. **1969**. The systematics of the fishes of the family Clinidae in South Africa. *Annals of the South African Museum* 55(1): 1–121.

Penrith M-L. 1970. The distribution of the fishes of the family Clinidae in Southern Africa. Annals of the South African Museum 55(3): 135–150.

Penrith M-L, Penrith MJ. **1967**. A new species of mullet of the genus *Ellochelon* (Pisces: Mugilidae) from St. Lucia estuary Zululand. *Durban Museum Novitiates* 8(8): 69–75.

Penrith M-L, Penrith MJ. **1970**. Note on the differentiation of two sympatrically occurring clingfishes in the Western Cape. *South African Journal of Science* 66(12): 392–393.

Pequeño G. **1989**. Peces de Chile. Lista sistemática revisada y comentada. *Revista de Biología Marina y Oceanografía* 24(2): 1–132.

Pequeño GR, Lamilla J, Crovetto AE. 1991. Captura de Somniosus cf. pacificus Bigelow & Schroeder, 1944, frente a Valdivia, Chile, con notas sobre su contenido gástrico (Chondrichthyes, Squalidae). Estudios oceanológicos 10: 117–122.

Pereira MAM. **2000**. *Preliminary checklist of reef-associated fishes of Mozambique*. Maputo, Mozambique: (CDS-MICOA). 21 pp.

Péron F. **1807**. *Voyage de découvertes aux terres Australes, exécuté par ordre de sa majesté l'Empereur et Roi, sur les Corvettes le* Géographe, *le* Naturaliste *et la Goelette le* Casuarina, *pendant les années 1800, 1801, 1803 et 1804* (2 vols). Paris: Arthus Bertrand.

Perry G. **1810–1811**. Ichthyology [unpaginated]. In: *Arcana*, or, *The Museum of Natural History: containing the most recent discovered objects embellished with coloured plates*, *and corresponding descriptions: with extracts relating to animals, and remarks of celebrated travellers; combining a general survey of nature*. London: James Stratford. 384 pp.; Pls. 1–84. Peters WCH. **1844**. Über einige neue Fische und Amphibien aus Angola und Mozambique. *Monatsberichte der Königlichen Preussischen Akademie der Wissenschaften zu Berlin* 1844: 32–37.

Peters WCH. **1852**. Diagnosen von neuen Flussfischen aus Mozambique. *Monatsberichte der Königlichen Preussischen Akademie Wissenschaften zu Berlin* 1852: 257–276, 681–685.

Peters WCH. 1855. Uebersicht der in Mossambique beobachteten Seefische. Bericht über die zur Bekanntmachung geeigneten Verhandlungen der Königlichen Preussischen Akademie der Wissenschaften zu Berlin 1855: 428–466.

Peters WCH. **1855**. Übersicht der in Mossambique beobachten Fische. *Archiv für Naturgeschichte* 21(2–3): 234–282.

Peters WCH. **1864**. Uber einige neue Saugethiere ... Amphibien ... und Fische. *Monatsberichte der Königlichen Preussischen Akademie der Wissenschaften zu Berlin* 1864: 381–399.

Peters WCH. **1865**. Uber einige Bloch'sche Arten der Fisch-Gattung Serranus. Monatsberichte der Königlichen Preussischen Akademie der Wissenschaften zu Berlin 1865: 97–111.

Peters WCH. **1866**. Mittheilung über Fische (*Protopterus, Auliscops, Labrax, Labracoglossa, Nematocentris, Serranus, Scorpis, Opisthognathus, Scombresox, Acharnes, Anguilla, Gymnomuraena, Chilorhinus, Ophichthys, Helmichthys). Monatsberichte der Königlichen Preussischen Akademie der Wissenschaften zu Berlin* 1866: 509–526.

Peters WCH. **1867**. Mitteilung über Fische. *Monatsberichte der Königlichen Preussischen Akademie der Wissenschaften zu Berlin* 1866: 509–526.

Peters WCH. 1868. Naturwissenschaftliche Reise nach Mossambique auf befehl Seiner Majestät des Königs Friedrich Wilhelm IV in den Jahren 1842 bis 1848 ausgeführt. Zoologie. IV. Flussfische. Berlin: G. Reimer, viii + 116 pp.

Peters WCH. **1868**. Über eine neue Untergattung der Flederthiere, so wie über neue Gattungen und Arten von Fischen. *Monatsberichte der Königlichen Preussischen Akademie der Wissenschaft zu Berlin* 1868: 145–148.

Peters WCH. **1869**. Über die von Hrn. Dr. F. Jagor in dem ostindischen Archipel gesammelten und dem Königlichen Zoologischen Museum übergebenen Fische. *Monatsberichte der Königlichen Preussischen Akademie der Wissenschaft zu Berlin* 1868: 254–281.

Peters WCH. **1869**. Ueber neue oder weniger bekannte Fische des Berliner Zoologischen Museums. *Monatsberichte der Königlichen Preussischen Akademie der Wissenschaft zu Berlin* 1869: 703–711.

Peters WCH. **1876**. Übersicht der von Hrn. Prof. Dr. K. Möbius in Mauritius und bei den Seychellen gesammelten Fische. *Monatsberichte der Königlichen Preussischen Akademie Wissenschaften zu Berlin* 1876: 435–447. Peters WCH. 1877. Ubersicht der während der von 1874 bis 1876 unter der Commando des Hrn. Capitän z.
S. Freiherrn von Schleinitz ausgeführten Reise S.M.S. *Gazelle* gesammelten und von der Kaiserlichen Admiralität der Königlichen Akademie der Wissenschaften überstanden Fische. *Monatsberichte der Königlichen Preussischen Akademie Wissenschaften zu Berlin* 1876: 831–854.

Peters WCH. **1883**. Liste des poissons collectionnés à Maurice et aux Seychelles par le Professeur-Docteur K. Möbius et classifiés par M.W. Peters. *Transactions of the Royal Society of Arts and Sciences of Mauritius* 11: 49–58.

Pethiyagoda R. **1991**. *Freshwater fishes of Sri Lanka*. Colombo: Wildlife Heritage Trust of Sri Lanka. xiii + 362 pp.

Pethiyagoda R. **1994**. *Rabaulichthys stigmaticus*: first record from Sri Lanka (Pisces: Serranidae: Anthiinae). *Journal of South Asian Natural History* 1(1): 15–16.

Petit G. **1931**. Une espèce nouvelle du genre *Foa* présentant un cas d'incubation bucco-branchiale. *Bulletin du Muséum National d'Histoire Naturelle* (Série 2) 3(1): 91–95.

Petit G. 1934. Un Fierasfer nouveau de Madagascar. Bulletin du Muséum National d'Histoire Naturelle (Paris) (Série 2) 6(4): 393–397.

Petit G. **1936**. Un *Gobius* nouveau de Madagascar. *Bulletin du Muséum National d'Histoire Naturelle* (Paris) (Série 2) 8(5): 388–393.

Pezold F. **1998**. Three new species of *Oxyurichthys* (Teleostei: Gobiidae) from the Indian and Pacific Oceans. *Copeia* 1998(3): 687–695.

Pezold FL, Larson HK. **2015**. A revision of the fish genus *Oxyurichthys* (Gobioidei: Gobiidae) with descriptions of four new species. *Zootaxa* 3988(1): 1–95. https://doi. org/10.11646/zootaxa.3988.1.1

Pfaff JR. **1933**. Report on the fishes collected by Mr. Harry Madsen during Professor O. Olufsen's Expedition to French Sudan in the years 1927–28. *Videnskabelige Meddelelser fra Dansk Naturhistorisk Forening, Kjøbenhavn* 94: 273–315, Pl. 6.

Pfaff JR. **1942**. Papers from Dr. Th. Mortensen's Pacific expedition 1914–16. LXXI. On a new genus and species of the family Gobiesocidae from the Indian Ocean, with observations on sexual dimorphism in the Gobiesocidae, and on the connection of certain gobiesocids with echinids. *Videnskabelige Meddelelser fra Dansk Naturhistorisk Forening, Kjøbenhavn* 105: 413–422.

Pfaff JR. **1953**. Kystomradadernes fisk. *Galatheas Jordomsejling* (København): 139–152.

Pfeffer GJ. **1893**. Ostafrikanische Fische gesammelt von Herrn Dr. F. Stuhlmann in Jahre 1888 und 1889. *Jahrbuch der Hamburgischen Wissenschaftlichen Anstalten* 10: 131–177.

Pfeffer GJ. **1894**. Die Thierwelt Ost-Afrikas und der Nachbargebiete. Lief. v. Die Fische Ost-Afrikas. *Fische Ost-Afrikas* 3: i–xviii + 72 pp. Philippi RA. **1857**. Ueber einige Chilenische Vögel und Fische. *Archiv Naturgeschichte* 23(1): 262–272.

Philippi RA. 1887. Historia natural. Sobre los tiburones y algunos otros peces de Chile. [Plus:] Apendice sobre el pejeespada, peje-aguja, peje-perro i vieja negra. Anales de la Universidad de Chile Sec. 1 71: 535–574, Pls. 1–8.

Philippi RA. **1892**. Algunos peces de Chile. *Anales del Museo Nacional de Chile* (Section 1, Zoologia) 3: 1–16. [In Spanish]

Phillipps WJ. **1924**. *Agrostichthys*, a new genus of ribbon fishes. *Proceedings of the Zoological Society of London* 1924(2): 539–540.

Phillipps WJ. **1932**. Notes on new fishes from New Zealand. *New Zealand Journal of Science and Technology* 13(4): 226–234.

Picaglia L. 1895. Pesci del Mar Rosso pescati nella campagna idrographica della Regia Nave *Scilla* nel 1891–92, Collezione aggiunta delle specie del Mar Rosso e del Golfo di Aden. *Atti della Società dei Naturalisti e Matematici, Modena* 3(13): 22–40.

Pichon M. 1972. The coral reefs of Madagascar (pp. 367–410).
In: Battistini R, Richard-Vindard G (eds) *Biogeography and ecology in Madagascar*. The Hague: Junk. 765 pp.

Pichon M, Benzoni F, Chaineau C, Dutrieux E. 2010. Field guide to the hard corals of the southern coast of Yemen. Paris: Muséum National d'Histoire Naturelle | Mèze: Biotope. 256 pp.

Pierce SJ, White WT, Marshall AD. 2008. New record of the smalleye stingray, *Dasyatis microps* (Myliobatiformes: Dasyatidae), from the western Indian Ocean. *Zootaxa* 1734: 65–68. http://dx.doi.org/10.11646/zootaxa.1734.1.5

Pietsch TW. **1978**. Evolutionary relationships of the sea moths (Teleostei: Pegasidae) with a classification of gasterosteiform families. *Copeia* 1978(3): 517–529.

Pietsch TW. **1984**. The genera of frogfishes (family Antennariidae). *Copeia* 1984(1): 27–44.

Pietsch TW. **1989**. Phylogenetic relationships of trachinoid fishes of the family Uranoscopidae. *Copeia* 1989(2): 253–303.

Pietsch TW (ed). 1995. Historical portrait of the progress of ichthyology, from its origins to our own time. Edited and annotated by TW Pietsch, translated from the French by AJ Simpson. Baltimore, Maryland: Johns Hopkins University Press. xxiv + 366 pp.

Pietsch TW, Anderson WD Jr (eds). 1997. Collection building in ichthyology and herpetology. American Society of Ichthyologists and Herpetologists Special Publication 2. Lawrence, Kansas: Allen Press. xiii + 593 pp.

Pietsch TW, Grobecker DB. 1987. Frogfishes of the world: Systematics, zoogeography, and behavioral ecology. Stanford: Stanford University Press. xxiv + 420 pp. Pietsch TW, Zabetian CP. **1990**. Osteology and interrelationships of the sand lances (Teleostei: Ammodytidae). *Copeia* 1990(1): 78–100.

Pietschmann V. **1912**. Eine neue *Mugil*-art aus dem Schatt el Arab. *Anzeiger der Akademie der Wissenschaften in Wien* 49(17): 268–270.

Pietschmann V. **1913**. Fische des Wiesbadener Museums. *Jahrbücher des Nassauischen Vereins für Naturkunde* (Wiesbaden) 66: 170–201.

Pietschmann V. **1934**. Drei neue Fische aus den hawaiischen Küstengewässern. *Anzeiger der Akademie der Wissenschaften in Wien* 71: 99–100.

Pietschmann V. **1938**. Hawaiian shore fishes. *Bulletin of the Bernice Pauahi Bishop Museum* 156: 3–55.

Pillay SR. 1962. A revision of Indian Mugilidae. Parts I–II. Journal of the Bombay Natural History Society 59(1): 254–270.

Pillay TVR. **1951**. Structure and development of the scales of five species of grey mullets of Bengal. *Proceedings of the National Institute of Sciences of India* 17(6): 413–424.

Pillay TVR. 1953. Mugil poecilus Day, same as Mugil troscheli Bleeker. Journal of the Bombay Natural History Society 51(1&2): 378–383.

Pilosof D, Fishelson L. **1981**. *Mysteries of the Red Sea*. Ramat-Gan: Massada Ltd. 167 pp. [In Hebrew]

Pinault M, Daydé A, Fricke R. **2014**. First observation of the slow dragonet *Callionymus aagilis* Fricke, 1999 in its natural environment. *Western Indian Ocean Journal of Marine Science* 12(1): 87.

Pinault M, Loiseau N, Chabanet P, Durville P, Magalon H, Quod JP, Galzin R. 2013. Marine fish communities in shallow volcanic habitats. *Journal of Fish Biology* 82(6): 1821–1847.

Pinchot G. **1930**. *To the South Seas: the cruise of the schooner Mary Pinchot to the Galapagos, the Marquesas, and the Tuamotu islands, and Tahiti*. New York City: Blue Ribbon Books, inc. 500 pp.

Pinchuk VI. 1969. A finding of new specimens of the rare shark *Heterodontus ramalheira* (Heterodontiformes, Heterodontidae). *Zoologicheskii Zhurnal* 48(2): 295–297.

Pinheiro HT, Bernardi G, Rocha LA. 2016. Pempheris gasparinii, a new species of sweeper fish from Trindade Island, southwestern Atlantic (Teleostei, Pempheridae). ZooKeys 561: 105–115. http://dx.doi.org/10.3897/ zookeys.561.7263

Piotrovsky AS. 1987. Two new species of stromateoid fishes (Centrolophidae and Ariommidae) from the western tropical Indian Ocean. *Voprosy Ikhtiologii* 27(3): 506–509.
[In Russian. Author spelling also seen as Piotrovskij and Piontrovskiy. English translation in *Journal of Ichthyology* 27(5): 153–157 with author as Piontrovskiy] Piotrovsky AS. 1994. Butterfishes (Stromateoidei) of southern Africa and adjacent regions (distribution, biology, fisheries). Proceedings of the Society for Scientific Research, Institute of Marine Fisheries and Oceanography 40: 69–77. [In Russian]

Piotrovsky AS, Pruťko VG. **1980**. The occurrence of the goblin shark, *Scapanorhynchus owstoni* (Chondrichthyes, Scapanorhynchidae) in the Indian Ocean. *Journal of Ichthyology* 20(1): 124–125.

Playfair RL. 1867 [also 1866]. In: Playfair RL, Günther A.

Playfair RL. **1867**. On the fishes of Cachar. *Proceedings of the Zoological Society of London* 1867(1): 14–17, Pl. 3.

Playfair RL. **1868**. The fishes of Seychelles. *Proceedings of the Zoological Society of London* 1867(3): 846–872. [also cited as 1867]

Playfair RL. **1868**. On a collection of fishes from Madagascar. *Proceedings of the Zoological Society of London* 1868(1): 9–12.

Playfair RL. **1869**. Further contributions to the ichthyology of Zanzibar. *Proceedings of the Zoological Society of London* 1869: 239–241.

Playfair RL, Günther A. 1867. The fishes of Zanzibar, with a list of the fishes of the whole east coast of Africa. London: John Van Voorst. xiv, 153 pp., Pls. 21. [also cited as 1866; reprint 1971, Newton K Gregg, Kentfield, California]

Plessis Y, Fourmanoir P. **1966**. Pleurosycia taisnei, nouvelle espèce de Gobiidae de Nouvelle-Calédonie. Bulletin du Muséum National d'Histoire Naturelle (Paris) (Série 2) 37(5): 764–765.

Plumstead EE, Prinsloo JF, Schoonbee HJ. 1991. A survey of the fish fauna of Transkei estuaries. Part 4: The Mntafufu and Mzamba River estuaries. *South African Journal of Zoology* 26(4): 153–163.

Poey F. **1858–1861**. *Memorias sobre la historia natural de la Isla de Cuba, acompañadas de sumarios Latinos y extractos en Francés*. Vol. 2. La Habana: Imprenta de la Viuda de Barcina. 442 pp.

Poey F. **1863**. Descriptions des poissons nouvelles ou peu connues. *Proceedings of the Academy of Natural Sciences of Philadelphia* 15: 180–188, 227.

Poey F. **1868**. Synopsis piscium cubensium. Catalogo Razonado de los peces de la isla de Cuba. *Repertorio Fisico-Natural de la Isla de Cuba* 2: 279–484.

Poey F. **1873**. *Grammicolepis brachiusculus*, tipe de una nueva familia en la clase de los peces. *Anales de la Sociedad Española de Historia Natural, Madrid* 2: 403–406.

Pogoreutz C, Vitecek S, Ahnelt H. **2013**. A new species of *Satyrichthys* (Teleostei: Peristediidae) from the Maldives Archipelago (Indian Ocean). *Zootaxa* 3694(2): 153–160. https://doi.org/10.11646/zootaxa.3694.2.4

Polanco FA, Acero PA, Betancur-R R. **2016**. No longer a circumtropical species: revision of the lizardfishes in the *Trachinocephalus myops* species complex, with description

of a new species from the Marquesas Islands. *Journal of Fish Biology* 89(2): 1302–1323. https://doi.org/10.1111/jfb.13038

Poll M. **1949**. Resultats scientifiques des croisières du Navire-École Belge *Mercator*. IV. *Mémoires, Institut Royal des Sciences Naturelles de Belgique* (Série 2) 33: 173–269.

Poll M. 1951–1959. Poissons. In: *Expédition océanographic belge dans les eaux côtières africaines de l'Atlantique sud (1948–1949)* (5 vols). Bulletin de l'Institut royal des sciences naturelles de Belgique, Bruxelles. [unpaginated]

Poll M. 1971. Revision systématique des daurades du genre Dentex de la côte africaine tropicale occidentale et de la Méditerranée. Mémoire de la Classe des Sciences de l'Académie Royale de Belgique 40(1): 1–51.

Poll M, Gosse J-P. **1995**. Genera des poissons d'eau douce de l'Afrique. *Mémoire de la Classe des Sciences de l'Académie Royale de Belgique* 9: 1–324.

Polunin NVC, Lubbock R. **1977**. Prawn-associated gobies (Teleostei: Gobiidae) from the Seychelles, western Indian Ocean: systematics and ecology. *Journal of Zoology* (London) 183(1): 63–101.

Polunin NVC, Lubbock R. **1979**. Five new prawn-associated gobies (Teleostei: Gobiidae) of the genus *Amblyeleotris*. *Bulletin of the British Museum (Natural History) Zoology* 36(4): 239–249.

Pondella DJ II, Craig MT, Franck JPC. **2003**. The phylogeny of *Paralabrax* (Perciformes: Serranidae) and allied taxa inferred from partial 16S and 12S mitochondrial ribosomal DNA sequences. *Molecular Phylogenetics and Evolution* 29(1): 176–184.

Ponomareva L. **1968**. Euphausiids of the Red Sea collected in summer 1966 by RV *Academician S. Vavilov. Marine Biology* 1: 263–265.

Popper D, Gunderman N. **1975**. Some ecological and behavioural aspects of siganid populations in the Red Sea and Mediterranean coasts of Israel in relation to their suitability for aquaculture. *Aquaculture* 6(2): 127–141.

Popper D, Pitt R, Zohar Y. **1979**. Experiments on the propagation of Red Sea siganids and some notes on their reproduction in nature. *Aquaculture* 16(2): 177–181.

Popta CML. **1922**. Vierte und letzte Fortsetzung der Beschreibung von neuen Fischarten der Sunda-Expedition. *Zoologische Mededelingen (Leiden)* 7(2): 27–39.

Poss SG. **1994**. Scorpaenidae (pp. 477–494). In: Gomon MF, Glover JCM, Kuiter RH (eds) *The fishes of Australia's south coast*. Adelaide: State Print. 992 pp.

Poss SG. 1999. Scorpaenidae. Scorpionfishes (also, lionfishes, rockfishes, stingfishes, stonefishes, and waspfishes) (pp. 2291–2352). In: Carpenter KE, Niem VH (eds) FAO species identification guide for fishery purposes. The living marine resources of the western central Pacific. Vol. 4. Bony fishes part 2 (pp. 2069–2790). Rome: FAO. Poss SG, Collette BB. **1990**. *Scorpaenodes immaculatus*, a new species of scorpionfish (Osteichthyes: Scorpaenidae) from Walters Shoals, Madagascar Ridge. *Proceedings of the Biological Society of Washington* 103(3): 543–549.

Poss SG, Duhamel G. **1991**. *Neomerinthe bauchotae*, a new scorpionfish (Scorpaenidae) from Saint Paul and Amsterdam Islands (southern Indian Ocean), with comments on the limits of the genus. *Cybium* 15(2): 93–102.

Poss SG, Eschmeyer WN. **1975**. The Indo-West Pacific scorpionfish genus *Ocosia* Jordan and Starks (Scorpaenidae, Tetraroginae), with description of three new species. *Matsya* 1: 1–18.

Poss SG, Eschmeyer WN. **1978**. Two new Australian velvetfishes, genus *Paraploactis* (Scorpaeniformes: Aploactinidae), with a revision of the genus and comments on the genera and species of the Aploactinidae. *Proceedings of the California Academy of Sciences* (Series 4) 41(18): 401–426.

Poss SG, McCosker JE, Baldwin CC. **2010**. A new species of *Scorpaenodes* (Pisces: Scorpaenidae) from the Galápagos and Cocos islands with discussions of the limits of *Socrpaenodes* and *Thysanichthys. Proceedings of the California Academy of Sciences* (Series 4) 61(2): 235–267.

Poss SG, Mee JKL. **1995**. A new species of *Choridactylus* (Pisces: Scorpaenoidei) from southern Oman. *Japanese Journal of Ichthyology* 42(1): 1–6.

Poss SG, Rama Rao KV. **1984**. Scorpaenidae. Scorpionfishes, rockfishes, rosefishes, stingfishes, stonefishes, turkeyfishes and waspfishes. In: Fischer W, Bianchi G (eds) *FAO species identification sheets for fishery purposes. Western Indian Ocean (Fishing Area 51)*. Vol. 4. Rome: FAO. [unpaginated]

Poss SG, Springer VG. **1983**. *Eschmeyer nexus*, a new genus and species of scorpaenid fish from Fiji. *Proceedings of the Biological Society of Washington* 96(2): 309–316.

Post A, Quéro J-C. **1991**. Distribution et taxonomie des *Howella* (Perciformes, Percichthyidae) de l'Atlantique. *Cybium* 15(2): 111–128. [In French]

Postel E. **1962**. *Myripristis (Holotrachys) guezei* poisson téléosteen nouveau de l'île de la Réunion. *Bulletin du Muséum National d'Histoire Naturelle* (Paris) (Série 2) 34(2): 158–162.

Postel E, Du Buit M-H. **1964**. Liste des poissons observés à la criée de Concarneau en fin Julliet-début Août 1964: tailles maximales enregistrées. *Bulletin de la Société Scientifique de Bretagne* 39(1–2): 113–117.

Postel E, Fourmanoir P, Guézé P. **1963**. Serranides de la Réunion. *Mémoires de l'Institut français d'Afrique Noire* 68: 339–384.

Postel E, Roux C. **1964**. *Scorpaena folgori*, poisson téléostéen nouveau des Iles du Cap Vert. *Bulletin du Muséum National d'Histoire Naturelle* (Paris) (Série 2) 36(2): 165–171. Potter IC, Gill HS. **2003**. Adaptive radiation of lampreys. *Journal of Great Lakes Research* 29(Suppl. 1): 95–112.

Potthoff T, Kelley S. **1982**. Development of the vertebral column, fins and fin supports, branchiostegal rays, and squamation in the swordfish, *Xiphias gladius*. *Fishery Bulletin* 80: 161–186.

Potthoff T, Richards WJ, Ueyanagi S.**1980**. Development of *Scombrolabrax heterolepis* (Pisces, Scombrolabraciae) and comments on familial relationships. *Bulletin of Marine Science* 30(2): 329–357.

Poulton WB. **1898**. Natural selection the cause of mimetic resemblance and common warning colours. *Journal of the Linnean Society of London* (Zoology) 26(172): 558–612.

Pouyaud L, Wirjoatmodjo S, Rachmatika I, Tjakrawidjaja AH, Hadiaty RK, Hadie W. **1999**. Une nouvelle espèce de coelacanthe. Preuves génétiques et morphologiques. *Comptes rendus de l'Académie des Sciences. Série 3. Sciences de la vie* 322: 261–267. [In French, abridged version in English]

Poyato-Ariza FJ, Wenz S. **2002**. New insight into pycnodontiform fishes. *Geodiversitas* 24(1): 139–248.

Pradervand P. **2000**. National marine linefish system: Spatial and temporal distribution of ragged tooth shark *Carcharias taurus* in shore anglers' catches from Umtumvuna River to Cape Point. *Oceanographic Research Institute, Data Report* 2000(7): 4.

Pratap HB. **1985**. Occurrence of *Solenostoma cynopterum* [sic] Bleeker 1854 from Mbudya Island, Dar-Es-Salaam. *Indian Journal of Fisheries* 32(1): 133–134.

Priest MA, DiBattista JD, McIlwain JL, Taylor BM, Hussey NE, Berumen ML. 2015. A bridge too far: dispersal barriers and cryptic speciation in an Arabian Peninsula grouper (*Cephalopholis hemistiktos*). Journal of Biogeography 43(4) [2016]: 820–832. http://dx.doi.org/10.1111/jbi.12681

Prochazka K, Griffiths CL. **1991**. *Cancelloxus longior*, a new species of xenopoclinin fish (Perciformes: Clinidae) from South Africa. J.L.B. Smith Institute of Ichthyology Special Publication 51: 1–6.

Prochazka K, Griffiths CL. **1992**. The intertidal fish fauna of the west coast of South Africa – species, community and biogeographic patterns. *South African Journal of Zoology* 27(3): 115–120. https://doi.org/10.1080/02541858.1992.11 448271

Prokofiev AM. **2007**. Osteology and some other morphological characters of *Howella sherborni*, with a discussion of the systematic position of the genus (Perciformes, Percoidei). *Journal of Ichthyology* 47(6): 413–426.

Prokofiev AM. **2007**. The osteology of *Bathysphyraenops simplex* and the diagnosis of the Howellidae (Perciformes: Percoidei) family. *Journal of Ichthyology* 47(8): 566–578.

Prokofiev AM. **2007**. Osteology of *Florenciella lugubris* (Percoidei: Epigonidae). *Journal of Ichthyology* 47(9): 715–725.

Prokofiev AM. 2008. New species and new records of lizardfishes of the genus *Synodus* (Teleostei, Synodontidae) from the western Indian Ocean. *Zoologicheskii Zhurnal* 87(4): 424–435. [In Russian]

Prokofiev AM. 2010. On a new finding of the rare species Owstonia pectinifer (Perciformes: Cepolidae). Journal of Ichthyology 50(5): 408–412. http://dx.doi.org/10.1134/S0032945210050073

Psomadakis PN, Osmany HB, Moazzam M. 2015. FAO species identification guide for fishery purposes. Field identification guide to the living marine resources of Pakistan. Rome: FAO. x + 386 pp., Pls. 1–42.

Purkis SJ. **2011**. The most temperature-adapted corals have an Achilles' Heel. *Marine Pollution Bulletin* 62(2): 246–250. http://dx.doi.org/10.1016/j.marpolbul.2010.11.005

Putnam FW. **1874**. Notes on the genus *Bdellostoma*. *Proceedings* of the Boston Society of Natural History 16: 156–160.

Pylaie M de la, *see* De la Pylaie M.

Pyle RL. **1990**. *Centropyge debelius*, a new species of angelfish (Teleostei: Pomacanthidae) from Mauritius and Réunion. *Revue française d'Aquariologie Herpétologie* 17(2): 47–52.

Pyle RL. 2005. Recent discoveries of new fishes inhabiting deep Pacific coral reefs, with biogeographic implications. Paper read at the 7th Indo-Pacific Fish Conference, 16–20 May, 2005. [collaborators listed as Randall JE, Earle JL, Greene BD, Greenfield DW] https://doi.org/10.5281/zenodo.16942

### Q

Quast JC. **1965**. Osteological characteristics and affinities of the hexagrammid fishes, with a synopsis. *Proceedings of the California Academy of Sciences* (Series 4) 31(21): 563–600.

Quensel C. **1806**. Försök att närmare bestämma och naturligare uppställa svenska arterna af flunderslägtet. *Kungliga Svenska Vetenskapsakademiens handlingar* 27: 44–56, 203–233.

Quéro J-C. **1970**. Les poissons de la famille des Searside capture dans l'Atlantique Nord-est Campagnes du *President-Thedore Tissier* et de la *Thalassa. Revue des Travaux de l'Institut des Pêches Maritimes* 34: 261–276.

Quéro J-C. **1974**. *Hoplostethus cadenati* sp. nov. Pisces, Beryciformes, Trachichthyidae poisson nouveau de l'Atlantique est. *Revue des Travaux de l'Institut des Pêches Maritimes* 38(1): 103–109.

Quéro J-C. **1979**. Remarques sur le *Grammicolepis brachiusculus* (Pisces: Zeiformes, Grammicolepidae) observé au port de la Rochelle. *Annales de la Société des Sciences Naturelles de la Charente-Maritime* 6(6): 573–576. Quéro J-C. **1986**. Caproidae (pp. 777–779). In: Whitehead PJP, Bauchot M-L, Hureau J-C, Nielsen J, Tortonese E (eds) *Fishes of the north-eastern Atlantic and the Mediterranean*. Vol. II. Paris: UNESCO.

Quéro J-C. **1997**. Soleidae et cynoglossidae (Pleuronectiformes) de l'île de la Réunion (Océan Indien): description d'une nouvelle espèce. *Cybium* 21(3): 319–329.

Quéro J-C, Desoutter M. **1990**. Soleidae de Madagascar. *Cybium* 14(2): 105–112.

Quéro J-C, Golani D. **1990**. Description d'*Engyprosopon hureaui* n. sp. (Pleuronectiformes, Bothidae) du golfe d'Akaba. *Cybium* 14(1): 37–42.

Quéro J-C, Hensley DA, Maugé AL. **1988**. Pleuronectidae de l'île de la Réunion et de Madagascar. I. *Poecilopsetta*. *Cybium* 12(4): 321–330.

Quéro J-C, Hensley DA, Maugé AL. **1989**. Pleuronectidae de l'île de la Réunion et de Madagascar. II. Genres *Samaris* et *Samariscus*. *Cybium* 13(2): 105–114.

Quéro J-C, Hureau J-C, Karrer K, Post A, Saldanha L (eds). **1990**. *Check-list of the fishes of the eastern tropical Atlantic (CLOFETA)* (3 vols). Lisbon: Junta Nacional de Investigação Científica e Tecnológica. xxxii + 1492 pp. [Vol. 1, i–xxxii + 1–519 pp.; Vol. 2, 520–1080 pp.; Vol. 3, 1081–1492 pp.]

Quéro J-C, Maugé A. **1989**. Cynoglossidae de l'île de la Réunion et de Madagascar. *Cybium* 13(4): 391–394.

Quéro J-C, Ozouf-Costaz C. **1991**. *Ostracoberyx paxtoni*, nouvelle espèce des côtes est de l'Australie. Remarques sur les modifications morphologiques des *Ostracoberyx* au cours de leur croissance (Perciformes, Ostracoberycidae). *Cybium* 15(1): 43–54.

Quéro J-C, Randall JE. **1990**. Mugiloididae (p. 892). In: Quéro J-C, Hureau J-C, Karrer C, Post A, Saldanha L (eds) *Checklist of the fishes of the eastern tropical Atlantic (CLOFETA)*. Vol. 2. Lisbon: Junta Nacional de Investigação Científica e Tecnológica.

Quéro J-C, Saldanha L. **1995**. Poissons anguilliformes de l'île de la Réunion (Océan Indien): description d'une nouvelle espèce. *Cybium* 19(1): 61–88.

Quéro J-C, Spitz J, Vayne J-J. **2009**. *Symphysanodon pitondelafournaisei*: une nouvelle espèce de Symphysanodontidae (Perciformes) de l'île de La Réunion (France, océan Indien). *Cybium* 33(1): 73–77.

Quéro J-C, Spitz J, Vayne J-J. **2010**. *Chromis durvillei*: une nouvelle espèce de Pomacentridae de l'île de la Réunion (France, océan Indien) et premier signalement pour l'île de *Chromis axillaris*. *Cybium* 33(4) [for 2009]: 321–326. https://doi.org/10.26028/cybium/2009-334-004

Quéro J-C, Spitz J, Vayne J-J. **2011**. Une éruption volcanique débusque *Neocentropogon profundus* (Tetrarogidae, Scorpaenoidei) à l'île de la Réunion (Océan Indien). *Cybium* 35(2): 99–103. https://doi.org/10.26028/ cybium/2011-352-003

Quod J-P, Naim O, Abdourazi F. **2000**. The Comoros Archipelago (pp. 243–267). In: Sheppard CRC (ed) *Seas at the Millennium: An environmental evaluation. Vol. 2.* Oxford: Pergamon Press. 920 pp.

- Quoy JRC, Gaimard JP. 1824–1825. Chapter IX. Description des Poissons (pp. 192–401). In: de Freycinet L Voyage autour du Monde...exécuté sur les corvettes de L. M. L'Uranie et La Physicienne, pendant les années 1817, 1818, 1819 et 1820.
  Paris: Pillet Aîné, pp. 1–328 in 1824; pp. 329–616 in 1825.
- Quoy JRC, Gaimard JP. **1830**. Atlas. In: *Dictionnaire Classique d'Histoire Naturelle* 17: 114, fig. 2.
- Quoy JRC, Gaimard JP. 1834. Poissons: In: Voyage de découvertes de l'Astrolabe exécuté par ordre du Roi, pendant les années 1826–1827–1828–1829, sous le commandement de M J Dumont d'Urville. Vol. 3. Paris: J. Tastu. pp. 647–720.
- Qureshi MR. **1972**. Sharks, skates and rays of the Arabian Sea. *Pakistan Journal of Scientific and Industrial Research* 15(4–5): 294–311.

# R

Radcliffe L. 1911. Notes on some fishes of the genus Amia, family of Cheilodipteridae, with descriptions of four new species from the Philippine Islands. [Scientific results of the Philippine cruise of the Fisheries steamer Albatross, 1907–1910. No. 12.] Proceedings of the United States National Museum 41(1853): 245–261, Pls. 20–25.

Radcliffe L. 1912. Descriptions of fifteen new fishes of the family Cheilodipteridae, from the Philippine Islands and contiguous waters. [Scientific results of the Philippine cruise of the Fisheries steamer *Albatross*, 1907–1910. No. 13.] *Proceedings of the United States National Museum* 41(1868): 431–446, Pls. 34–38.

Radcliffe L. **1912**. New pediculate fishes from the Philippine Islands and contiguous waters. *Proceedings of the United States National Museum* 42(1896): 199–214.

Radcliffe L. 1913. Descriptions of seven new genera and thirtyone new species of fishes of the families Brotulidae and Carapidae from the Philippine Islands and the Dutch East Indies. *Proceedings of the United States National Museum* 44(1948): 135–176, Pls. 7–17.

Radcliffe L, Smith HM. **1913**. Descriptions of a new family, two genera, and twenty-nine new species of anacanthine fishes from the Philippine Islands and contiguous waters. *Proceedings of the United States National Museum* 43(1924): 105–140.

Radhakrishnan N. **1967**. Notes on the maturity and spawning of *Opisthopterus tardoore* (Cuvier). *Indian Journal of Fisheries* 10A(1): 102–106.

Radhakrishnan N. **1968**. Some aspects of the biology of the long finned herring, *Opisthopterus tardoore* (Cuvier) from the inshore regions at Karwar. *Indian Journal of Fisheries* 10(1): 634–641.

Rafinesque CS. **1810**. *Caratteri di alcuni nuovi generi e nuovi specie di animali e piante della Sicilia, con varie osservazioni sopra i medisimi*. Palermo: Sanfilippo. 105 pp. [Reprint, 1967, Asher-Amsterdam]

Rafinesque CS. **1810**. *Indice d'ittiologia siciliana; ossia, catalogo metodico dei nomi latini, italiani, e siciliani dei pesci, che si rinvengono in Sicilia disposti secondo un metodo naturale e seguito da un appendice che contiene la descrizione de alcuni nuovi pesci siciliani*. Messina: G. del Nobolo. 70 pp. [Reprint, 1967, Asher-Amsterdam]

Rafinesque CS. **1815**. *Analyse de la nature, ou tableau de l'univers et des corps organisés*. Palerme. 224 pp.

Rahimian H, Pehpuri A. **2006**. Intertidal fishes of Qeshm Island, the Persian Gulf. I. Gobiidae (Pisces: Perciformes). *Journal of Science, University of Tehran* 33: 69–76.

Rahman AKA. **1976**. The fresh water pipe fishes of Bangladesh, with the description of a new species. *Bangladesh Journal of Zoology* 4(2): 43–50. [Page numbers erroneously printed as 43–50, correctly 105–112]

Rahman AKA. 1989. Freshwater fishes of Bangladesh. Zoological Society of Bangladesh. Department of Zoology, University of Dhaka. 364 pp.

Raj BS. **1941**. Notes on the common Madras sea-horse, *Hippocampus kuda multiannularis*, a new sub-species. *Proceedings of the Indian Science Congress* 27(3): 156.

Raj BS. **1954**. The problem of the apparent discontinuous distribution of *Harpadon nehereus* (Ham). *Proceedings of the Indian Academy of Sciences* (Section B) 40(2): 58–68.

Ramaiyan V, Rao TVS. 1970. A scleropareid fish Acanthosphex leurynnis (Pisces: Bathyaploactidae) from inshore waters off Porto Novo, South India. Copeia 1970(1): 192–193.

Ramamirtham CP, Rao DS. **1974**. On upwelling along the west coast of India. *Journal of the Marine Biological Biological Association of India* 15(1): 306–317.

Ramanathan N, Natarajan R. 1980. The flat fishes of Porto Novo (India) (Pisces, Pleuronectiformes). *Bulletin Zoologisch Museum* (Universiteit van Amsterdam) 7(10): 89–116.

Rama-Rao KV. **1967**. A new sole *Zebrias cochinensis* from India. *Journal of the Zoological Society of India* 19(1–2): 99–100.

Rama-Rao KV. **1970.** A systematic review of the family Scorpaenidae (Pisces) of the Indian seas and adjacent waters. PhD dissertation, University of Gorakhpur.

Rama-Rao KV, Badrudeen M. **1973**. *Inimicus sinense* (Valenciennes) (Synanceiidae: Pisces) a new record from India and Ceylon. *Journal of the Marine Biological Biological Association of India* 15(1): 418–421.

Ramaswami LS. **1946**. A comparative account of the skull of *Gambusia*, *Oryzias*, *Aplocheilus* and *Xiphophorus* (Cyprinodontes: Teleostomi). *Spolia Zeylanica* 24: 181–192. Ramesh R, Nammalwar P, Gowri VS. 2008. Database on coastal information of Tamilnadu. Report submitted to Environmental Information System (ENVIS) Centre, Department of Environment, Government of Tamilnadu, Chennai. Institute for Ocean Management, Anna University, Chennai. 132 pp.

Ramsay EP. **1881**. On a new species of *Regalaecus*, from Port Jackson. *Regalaecus jacksonensis*, sp. nov. *Proceedings of the Linnean Society of New South Wales* 5(4): 631–633.

Ramsay EP, Ogilby JD. **1886**. A contribution to the knowledge of the fish-fauna of New Guinea. *Proceedings of the Linnean Society of New South Wales* (Series 2) 1(1): 8–20.

Ramsay EP, Ogilby JD. **1886**. Descriptions of new or rare Australian fishes. *Proceedings of the Linnean Society of New South Wales* 10(4): 575–579.

Ramsay EP, Ogilby JD. **1887**. Descriptions of new Australian fishes. *Proceedings of the Linnean Society of New South Wales* (Series 2) 2(2): 181–184.

Ramsay EP, Ogilby JD. **1887**. Notes on the genera of Australian fishes. *Proceedings of the Linnean Society of New South Wales* (Series 2) 2(3): 561–564.

Randall HA, Allen GR. 1977. A revision of the damselfish genus *Dascyllus* (Pomacentridae) with the description of a new species. *Records of the Australian Museum* 31(9): 349–385.

Randall JE. 1955. A revision of the surgeon fish genus *Ctenochaetus*, family Acanthuridae, with descriptions of five new species. *Zoologica* (N.Y.) 40(pt 4, no. 15): 149–166, Pls. 1–2.

Randall JE. **1956**. A revision of the surgeon fish genus *Acanthurus. Pacific Science* 10(2): 159–235.

Randall JE. **1958**. A review of the labrid fish genus *Labroides*, with descriptions of two new species and notes on ecology. *Pacific Science* 12(4): 327–347.

Randall JE. 1963. Review of the hawkfishes (family Cirrhitidae). Proceedings of the United States National Museum 114(3472): 389–451.

Randall JE. **1964**. A revision of the filefish genera *Amanses* and *Cantherhines*. *Copeia* 1964(2): 331–361.

Randall JE. **1964**. Notes on the groupers of Tahiti, with description of a new serranid fish genus. *Pacific Science* 18(3): 281–296.

Randall JE. **1972**. A revision of the labrid fish genus *Anampses*. *Micronesica* 8(1–2): 151–189.

Randall JE. **1973**. Size of the great white shark (*Carcharodon*). *Science* **181**(4095): 169–170.

Randall JE. **1975**. A revision of the Indo-Pacific angelfish genus *Genicanthus*, with descriptions of three new species. *Bulletin of Marine Science* 25(3): 393–421.

Randall JE. **1977**. Contribution to the biology of the whitetip reef shark (*Triaenodon obesus*). *Pacific Science* 31(2): 143–164.

Randall JE. **1978**. A revision of the Indo-Pacific labrid fish genus *Macropharyngodon*, with descriptions of five new species. *Bulletin of Marine Science* 28(4): 742–770.

Randall JE. **1979**. A review of the serranid fish genus *Anthias* of the Hawaiian Islands, with descriptions of two new species. *Contributions in Science* (Los Angeles) 302: 1–13.

Randall JE. **1980**. Notes on the Indian Ocean hawkfishes *Cirrhitichthys bleekeri* and *C. guichenoti* (Pisces: Cirrhitidae). *Matsya* 6: 1–8.

Randall JE. 1980. Revision of the fish genus *Plectranthias* (Serranidae: Anthiinae) with descriptions of 13 new species. *Micronesica* 16(1): 101–187.

Randall JE. 1981. Luzonichthys earlei — a new species of anthiine fish from the Hawaiian Islands. Freshwater and Marine Aquarium 4(9): 13–18.

Randall JE. **1981**. A review of the Indo-Pacific sand tilefish genus *Hoplolatilus* (Perciformes: Malacanthidae). *Freshwater and Marine Aquarium* 4(12): 39–46.

Randall JE. **1981**. Two new species and six new records of labrid fishes from the Red Sea. *Senckenbergiana Maritima* 13(1/3): 79–109.

Randall JE. **1982**. A review of the labrid fish genus *Hologymnosus. Revue française d'Aquariologie Herpétologie* 9(1): 13–20.

Randall JE. **1983**. *Red Sea reef fishes*. London: Immel Publishing. 192 pp.

Randall JE. **1984**. Two new Indo-Pacific mugiloidid fishes of the genus *Parapercis*. *Freshwater and Marine Aquarium* 7(12): 41–49.

Randall JE. **1985**. *Underwater guide to Hawaiian reef fishes*. Newton Square, Pennsylvania: Harrowood Books. 70 pp.

Randall JE. 1987. A preliminary synopsis of the groupers (Perciformes: Serranidae: Epinephelinae) of the Indo-Pacific region (pp. 89–188). In: Polovina JJ, Ralston S (eds) *Tropical snappers and groupers: Biology and fisheries management*. Boulder, Colorado: Westview Press. 659 pp.

Randall JE. **1988**. Three new damselfishes of the genus *Chromis* (Perciformes: Pomacentridae) from the Indian Ocean. *Revue française d'Aquariologie Herpétologie* 15(2): 49–56.

Randall JE. **1988**. Three new Indo-Pacific damselfishes of the genus *Chromis* (Pomacentridae). *Memoirs of the Museum of Victoria* 49(1): 73–81.

Randall JE. **1992**. *Diver's guide to fishes of Maldives*. London: Immel Publishing. 193 pp.

Randall JE. 1993. Acanthurus tristis, a valid Indian Ocean surgeonfish (Perciformes: Acanthuridae). J.L.B. Smith Institute of Ichthyology Special Publication 54: 1–8.

Randall JE. **1994**. Twenty-two new records of fishes from the Red Sea. *Fauna of Saudi Arabia* 14: 259–275.

Randall JE. **1994**. A new genus and six new gobiid fishes (Perciformes: Gobiidae) from Arabian waters. *Fauna of Saudi Arabia* 14: 317–340. Randall JE. **1994**. *Ilisha compressa*, a new species of clupeid fish from the Persian Gulf. *Raffles Bulletin of Zoology* 42(4): 893–899.

 Randall JE. 1994. Two new damselfishes (Perciformes: Pomacentridae) from Arabian waters. *Revue française* d'Aquariologie Herpétologie 21(1–2): 39–48.

Randall JE. 1995. A review of the wrasses of the genus *Cirrhilabrus* (Perciformes: Labridae) from the western Indian Ocean. *Revue française d'Aquariologie Herpétologie* 22(1–2): 19–26.

Randall JE. **1995**. A review of the triplefin fishes (Perciformes: Blennioidei: Triptrygiidae) of Oman, with descriptions of two new species of *Enneapterygius*. *Revue française d'Aquariologie Herpétologie* 22(1–2): 27–34.

Randall JE. **1995**. *Coastal fishes of Oman*. Bathurst, Australia: Crawford House Publishing. 439 pp.

Randall JE. **1996**. *Shore fishes of Hawai'i*. Vida, Oregon: Natural World Press. 216 pp.

Randall JE. **1996**. Two new anthiine fishes of the genus *Plectranthias* (Perciformes: Serranidae), with a key to the species. *Micronesica* 29(2): 113–131.

Randall JE. **1997**. *Coastal fishes of Oman* (avec des photographies additionnelles de John P. Hoover). Honolulu: University of Hawaii Press. 439 pp.

Randall JE. **1998**. Revision of the Indo-Pacific squirrelfishes (Beryciformes: Holocentridae: Holocentrinae) of the genus *Sargocentron*, with descriptions of four new species. *Indo-Pacific Fishes* 27: 1–105, Pls. 1–11.

Randall JE. **1999**. Revision of the Indo-Pacific labrid fishes of the genus *Pseudocheilinus*, with descriptions of three new species. *Indo-Pacific Fishes* 28: 1–34, Pls. 1–2.

Randall JE. 2000. Revision of the Indo-Pacific labrid fishes of the genus *Stethojulis*, with descriptions of two new species. *Indo-Pacific Fishes* 31: 1–42, Pls. 1–6.

Randall JE. **2001**. Five new Indo-Pacific gobiid fishes of the genus *Coryphopterus*. *Zoological Studies* 40(3): 206–225.

Randall JE. 2001. Hawkfish *Amblycirrhitus indicus* Fowler, 1938: a junior synonym of *Amblycirrhitus pinos* (Mowbray, 1927), the result of locality error. *Copeia* 2001(3): 870–871.

Randall JE. 2001. Mullidae. Goatfishes (surmullets) (pp. 3175–3200). In: Carpenter KE, Niem VH (eds) FAO species identification guide for fishery purposes. The living marine resources of the western central Pacific. Vol. 5. Bony fishes part 3 (pp. 2791–3380). Rome: FAO.

Randall JE. 2001. Pinguipedidae (= Parapercidae, Mugiloididae). Sandperches (pp. 3501–3510). In: Carpenter KE, Niem VH (eds) FAO species identification guide for fishery purposes. The living marine resources of the western central Pacific. Vol. 6. Bony fishes part 4 (pp. 3381–4218). Rome: FAO. Randall JE. **2001**. Revision of the generic classification of the hawkfishes (Cirrhitidae), with descriptions of three new genera. *Zootaxa* 12: 1–12.

Randall JE. **2002**. *Surgeonfishes of the world*. Honolulu: Mutual Publishing and Bishop Museum Press. 123 pp.

Randall JE. **2004**. Revision of the goatfish genus *Parupeneus* (Perciformes: Mullidae), with descriptions of two new species. *Indo-Pacific Fishes* 36: 1–64.

Randall JE. **2005**. A review of mimicry in marine fishes. *Zoological Studies* 44(3): 299–328.

Randall JE. **2005**. *Reef and shore fishes of the South Pacific. New Caledonia to Tahiti and the Pitcairn Islands.* Honolulu: University of Hawai'i Press. xii + 707 pp.

Randall JE. **2005**. Review of clupeotoxism, an often fatal illness from the consumption of clupeoid fishes. *Pacific Science* 59(1): 73–77.

Randall JE. **2007**. *Reef and shore fishes of the Hawaiian Islands*. Honolulu: Sea Grant Program, University of Hawai'i. xiv + 546 pp.

Randall JE. **2007**. *Vanderhorstia opercularis*, a new shrimp goby from the northern Red Sea. *Electronic Journal of Ichthyology* 3(1): 18–25.

Randall JE. 2008. Six new sandperches of the genus *Parapercis* from the western Pacific, with description of a neotype for *P. maculata* (Bloch and Schneider). *Raffles Bulletin of Zoology* Supplement 19: 159–178.

Randall JE. **2009**. Five new Indo-Pacific lizardfishes of the genus *Synodus* (Aulopiformes: Synodontidae). *Zoological Studies* 48(3): 402–417.

Randall JE. **2011**. Two new serranid fishes of the genus *Pseudanthias* from the western Indian Ocean. *Smithiana* Bulletin 13: 79–87.

Randall JE. **2013**. Seven new species of labrid fishes (*Coris*, *Iniistius*, *Macropharyngodon*, *Novaculops*, and *Pteragogus*) from the Western Indian Ocean. *Journal of the Ocean Science Foundation* 7: 1–43. https://doi.org/10.5281/ zenodo.1041964

Randall JE, Aida K, Hibiya T, Matsuura N, Kamiya H, Hashimoto Y. 1971. Grammistin, the skin toxin of soapfishes and its significance in the classification of the Grammistidae. *Publications of the Seto Marine Biological Laboratory* 19(2–3): 157–190.

Randall JE, Allen GR. **1973**. A revision of the gobiid fish genus *Nemateleotris*, with descriptions of two new species. *Quarterly Journal of the Taiwan Museum* 26(3/4): 347–367.

Randall JE, Allen GR. **1982**. *Chromis pelloura*. A new species of damselfish from the northern Red Sea. *Freshwater and Marine Aquarium* 5(11): 15–19.

Randall JE, Allen GR. 1987. Four new serranid fishes of the genus *Epinephelus* (Perciformes: Epinephelinae) from Western Australia. *Records of the Western Australian Museum* 13(3): 387–411.

Randall JE, Allen GR. 2004. Gomphosus varius x Thalassoma lunare, a hybrid labrid fish from Australia. aqua, Journal of Ichthyology and Aquatic Biology 8(3): 135–139.

Randall JE, Allen GR. 2005. Neopomacentrus sororius, a new species of damselfish from the Indian Ocean, with description of a neotype for its sister species, N. azysron (Bleeker). aqua, Journal of Ichthyology and Aquatic Biology 10(2): 73–80.

Randall JE, Allen GR, Anderson WD Jr. 1987. Revision of the Indo-Pacific lutjanid genus *Pinjalo*, with description of a new species. *Indo-Pacific Fishes* 14: 1–17.

Randall JE, Allen GR, Smith-Vaniz WF. 1978. Illustrated identification guide to commercial fishes: Bahrain, Iran, Iraq, Kuwait, Oman, Qatar, Saudi Arabia, United Arab Emirates. Rome: FAO. v + 221 pp.

Randall JE, Allen GR, Steene RC. **1990**. *Fishes of the Great Barrier Reef and Coral Sea*. Honolulu: University of Hawaii Press; Bathurst, Australia: Crawford House Press. xx + 507 pp.

Randall JE, Allen GR, Steene RC. **1997**. *Fishes of the Great Barrier Reef and Coral Sea* (2<sup>nd</sup> edition). Honolulu: University of Hawaii Press; Bathurst, Australia: Crawford House Press. xx + 557 pp.

Randall JE, Anderson RC. 1993. Annotated checklist of the epipelagic and shore fishes of the Maldive Islands. *Ichthyological Bulletin of the J.L.B. Smith Institute of Ichthyology* 59: 1–48.

Randall JE, Anderson RC. 1997. Chlorurus rhakoura, a new species of parrotfish (Perciformes: Labroidei: Scaridae) from Sri Lanka. Journal of South Asian Natural History 2(2): 155–164.

Randall JE, Arnold RJ. **2012**. *Uranoscopus rosette*, a new species of stargazer (Uranoscopidae: Trachinoidei) from the Red Sea. *aqua, International Journal of Ichthyology* 18(4): 209–218.

Randall JE, Baldwin CC. 1997. Revision of the serranid fishes of the subtribe Pseudogrammina, with descriptions of five new species. *Indo-Pacific Fishes* 26: 1–56, Pl. 1.

Randall JE, Bauchot ML. 1993. Case 2842. Naucrates
Rafinesque, 1810 and Xyrichtys Cuvier, 1814 (Osteichthyes, Perciformes): Proposed conservation. Bulletin of Zoological Nomenclature 50(4): 277–281.

Randall JE, Bauchot M-L, Ben-Tuvia A, Heemstra PC. 1985. Cephalopholis argus Schneider, 1801 and Cephalopholis sexmaculata (Rüppell, 1830) (Osteichthyes, Serranidae): proposed conservation by suppression of Bodianus guttatus Bloch, 1790, Anthias argus Bloch, 1792 and Serranus zanana Valenciennes 1828. Z.N. (S.) 2470. Bulletin of Zoological Nomenclature 42(4): 374–378.

Randall JE, Ben-Tuvia A. 1983. A review of the groupers (Pisces: Serranidae, Epinephelinae) of the Red Sea, with description of a new species of *Cephalopholis*. *Bulletin of Marine Science* 33(2): 373–426. Randall JE, Bineesh KK. **2014**. Review of the fishes of the genus *Pempheris* (Perciformes: Pempheridae) of India, with description of a new species and a neotype for *P. mangula* Cuvier. *Journal of the Ocean Science Foundation* 10: 20–40.

Randall JE, Bogorodsky SV. **2016**. Preliminary review of the pempherid fish genus *Parapriacanthus* of the western Indian Ocean, with descriptions of five new species. *Journal of the Ocean Science Foundation* 20: 1–24. http://doi.org/10.5281/zenodo.47480

Randall JE, Bogorodsky SV, Alpermann TJ, Satapoomin U, Mooi RD, Mal AO. **2014**. *Pempheris flavicycla*, a new pempherid fish from the Indian Ocean, previously identified as *P. vanicolensis* Cuvier. *Journal of the Ocean Science Foundation* 9 [for 2013]: 1–23.

Randall JE, Bogorodsky SV, Krupp F, Rose JM, Fricke R. 2013. *Epinephelus geoffroyi* (Klunzinger, 1870) (Pisces, Serranidae), a valid species of grouper endemic to the Red Sea and Gulf of Aden. *Zootaxa* 3641(5): 524–532. http://dx.doi.org/10.11646/zootaxa.3641.5.2

Randall JE, Bogorodsky SV, Mal AO. **2013**. Four new soles (Pleuronectiformes: Soleidae) of the genus *Aseraggodes* from the western Indian Ocean. *Journal of the Ocean Science Foundation* 8: 1–17. http://dx.doi.org/10.5281/ zenodo.1041968

Randall JE, Böhlke JE. **1981**. The status of the cardinalfishes *Apogon evermanni* and *A. anisolepis* (Perciformes: Apogonidae) with description of a related new species from the Red Sea. *Proceedings of the Academy of Natural Sciences of Philadelphia* 133: 129–140.

Randall JE, Brock VE. **1960**. Observations on the ecology of epinepheline and lutjanid fishes of the Society Islands, with emphasis on food habits. *Transactions of the American Fisheries Society* 89: 9–16.

Randall JE, Bruce RW. **1983**. The parrotfishes of the subfamily Scarinae of the western Indian Ocean with descriptions of three new species. *Ichthyological Bulletin of the J.L.B. Smith Institute of Ichthyology* **47**: 1–39.

Randall JE, Choat JH. **1980**. Two new parrotfishes of the genus *Scarus* from the central and South Pacific, with further examples of sexual dichromatism. *Zoological Journal of the Linnean Society* 70(4): 383–419, Pls. 1–8.

Randall JE, Clements KD. 2001. Second revision of the surgeonfish genus *Ctenochaetus* (Perciformes: Acanthuridae), with descriptions of two new species. *Indo-Pacific Fishes* 32: 1–33, Pls. 1–6.

Randall JE, Compagno LJV. **1995**. A review of the guitarfishes of the genus *Rhinobatos* (Rajiformes: Rhinobatidae) from Oman, with description of a new species. *Raffles Bulletin of Zoology* 43(2): 289–298.

Randall JE, Condé B. **1989**. *Frontilabrus caeruleus*, nouveaux genre et espèce de Labridé des Maldives. *Revue française d'Aquariologie Herpétologie* 15(3) [for 1988]: 89–92.

Randall JE, Connell AD. 2013. Nemateleotris exquisita, a new microdesmid fish from the Indian Ocean (Perciformes: Microdesmidae). Journal of the Ocean Science Foundation 8: 18–29.

Randall JE, Connell AD, Victor BC. **2015**. Review of the labrid fishes of the Indo-Pacific genus *Pseudocoris* with a description of two new species. *Journal of the Ocean Science Foundation* 16: 1–55.

Randall JE, De Bruin GHP. **1988**. The butterflyfish *Prognathodes guyotensis* from the Maldive Islands, a first record for the Indian Ocean. *Cybium* 12(2): 145–149.

Randall JE, DiBattista JD. 2013. A new species of damselfish (Pomacentridae) from the Indian Ocean. *aqua*, *International Journal of Ichthyology* 19(1): 1–16.

Randall JE, DiSalvo LH. 1997. *Rhinopias cea*, a new species of scorpionfish from Easter Island. *Bulletin of Marine Science* 60(3): 1035–1039.

Randall JE, Dooley JK. 1974. Revision of the Indo-Pacific branchiostegid fish genus *Hoplolatilus*, with descriptions of two new species. *Copeia* 1974(2): 457–471.

Randall JE, Dor M. 1981. Description of a new genus and species of labrid fish from the Red Sea. *Israel Journal of Zoology* 29(4) [for 1980]: 153–162.

Randall JE, Downing N, McCarthy LJ, Stanaland BE, Tarr AB.
1994. Fifty-one new records of fishes from the Arabian Gulf. *Fauna of Saudi Arabia* 14: 220–258.

Randall JE, Earle JL. 1994. Doryrhamphus aurolineatus, a new pipefish (Syngnathidae) from Masirah Island, Oman. Fauna of Saudi Arabia 14: 282–286.

Randall JE, Earle JL. 2002. Review of Hawaiian razorfishes of the genus *Iniistius* (Perciformes: Labridae). *Pacific Science* 56(4): 389–402, Pls. 1–3.

Randall JE, Earle JL. **2004**. *Novaculoides*, a new genus for the Indo-Pacific labrid fish *Novaculichthys macrolepidotus*. *aqua, Journal of Ichthyology and Aquatic Biology* 8(1): 37–43.

Randall JE, Earle JL. 2008. Two new Indo-Pacific sand lances of the genus Ammodytoides (Perciformes: Ammodytidae). Pacific Science 62(4): 603–612. https://doi.org/10.2984/1534-6188(2008)62[603:TNISLO]2.0.CO;2

Randall JE, Edwards AJ. **1984**. A new labrid fish of the genus *Thalassoma* from the Pitcairn Group, with a review of related Indo-Pacific species. *Journal of Aquariculture and Aquatic Sciences* 4(2): 13–32.

Randall JE, Emery AR. 1971. On the resemblance of the young of the fishes *Platax pinnatus* and *Plectorhynchus chaetodontoides* to flatworms and nudibranchs. *Zoologica* (N.Y.) 56(1): 115–119.

Randall JE, Emery AR. 1983. A new labrid fish of the genus Cirrhilabrus from the Chagos Archipelago, Indian Ocean. Journal of Aquariculture and Aquatic Sciences 3(2): 21–24. Randall JE, Eschmeyer WN. **2002**. Revision of the Indo-Pacific scorpionfish genus *Scorpaenopsis*, with descriptions of eight new species. *Indo-Pacific Fishes* 34 [for 2001]: 1–79.

Randall JE, Ferraris CJ Jr. 1981. A revision of the Indo-Pacific labrid fish genus *Leptojulis* with descriptions of two new species. *Revue française d'Aquariologie Herpétologie* 8(3): 89–96.

Randall JE, Fraser TH. **1999**. Clarification of the western Pacific cardinalfish species *Apogon trimaculatus* and *A. rhodopterus*, with decription of a similar new species. *Raffles Bulletin of Zoology* 47(2): 617–633.

Randall JE, Fraser TH, Lachner EA. **1990**. On the validity of the Indo-Pacific cardinalfishes *Apogon aureus* (Lacepède) and *A. fleurieu* (Lacepède), with description of a related new species from the Red Sea. *Proceedings of the Biological Society of Washington* 103(1): 39–62.

Randall JE, Golani D. 1995. Review of the moray eels (Anguilliformes: Muraenidae) of the Red Sea. *Bulletin of Marine Science* 56(3): 849–880.

Randall JE, Gon O. **2006**. Review of the soles of the genus *Aseraggodes* of the western Indian Ocean, with descriptions of three new species. *Israel Journal of Zoology* 51(3): 165–190.

Randall JE, Goren M. **1993**. A review of the gobioid fishes of the Maldives. *Ichthyological Bulletin of the J.L.B. Smith Institute of Ichthyology* 58: 1–37.

Randall JE, Greenfield DW. **1996**. Revision of the Indo-Pacific holocentrid fishes of the genus *Myripristis*, with descriptions of three new species. *Indo-Pacific Fishes* 25: 1–61, Pls. 1–9.

Randall JE, Greenfield DW. 2001. A preliminary review of the Indo-Pacific gobiid fishes of the genus *Gnatholepis*. *Ichthyological Bulletin of the J.L.B. Smith Institute of Ichthyology* 69: 1–17.

Randall JE, Guézé P. **1980**. The goatfish *Mulloidichthys mimicus* n. sp. (Pisces: Mullidae) from Oceania, a mimic of the snapper *Lutjanus kasmira* (Pisces: Lutjanidae). *Bulletin du Muséum National d'Histoire Naturelle* (Paris) (Série 4: Section A: Zoologie, Biologie et Écologie Animale) 2(2): 603–609.

Randall JE, Guézé P. 1981. The holocentrid fishes of the genus Myripristis of the Red Sea, with clarification of the Murdjan and Hexagonus complexes. Contributions in Science (Los Angeles) 334: 1–16.

Randall JE, Guézé P. **1984**. *Parupeneus margaritatus*, a new species of goatfish (Mullidae) from the Persian Gulf and Gulf of Oman. *Cybium* 8(4): 9–17.

Randall JE, Harmelin-Vivien ML. 1977. A review of the labrid fishes of the genus *Paracheilinus* with description of two new species from the western Indian Ocean. *Bulletin du Muséum National d'Histoire Naturelle* (Série 3: Zoologie) 436 (Zoologie 306): 329–342. Randall JE, Heemstra E. **2009**. Three new goatfishes of the genus *Parupeneus* from the western Indian Ocean, with resurrection of *P. seychellensis. Smithiana* Bulletin 10: 37–50.

Randall JE, Heemstra PC. **1978**. Reclassification of the Japanese cirrhitid fishes, *Serranocirrhitus latus* and *Isobuna japonica* to the Anthiinae. *Japanese Journal of Ichthyology* 25(3): 165–172.

Randall JE, Heemstra PC. **1985**. A review of the squirrelfishes of the subfamily Holocentrinae from the western Indian Ocean and Red Sea. *Ichthyological Bulletin of the J.L.B. Smith Institute of Ichthyology* 49: 1–27.

Randall JE, Heemstra PC. 1986. Epinephelus truncatus Katayama, a junior synonym of the Indo-West Pacific serranid fish Epinephelus retouti Bleeker. Japanese Journal of Ichthyology 33(1): 51–56.

Randall JE, Heemstra PC. **1991**. Revision of Indo-Pacific groupers (Perciformes: Serranidae: Epinephelinae), with descriptions of five new species. *Indo-Pacific Fishes* 20: 1–332.

Randall JE, Heemstra PC. **2006**. Review of the Indo-Pacific fishes of the genus *Odontanthias* (Serranidae: Anthiinae), with descriptions of two new species and a related genus. *Indo-Pacific Fishes* 38: 1–32.

Randall JE, Heemstra PC. 2008. Ammodytoides xanthops, a new species of sandlance (Perciformes: Ammodytidae) from Mozambique. Smithiana Bulletin 9: 21–25.

Randall JE, Heemstra PC. 2008. Meganthias filiferus, a new species of anthine fish (Perciformes: Serranidae), from the Andaman Sea off southwestern Thailand. Phuket Marine Biological Center Research Bulletin 68: 5–9. [also cited as 2007]

Randall JE, Hoese DF. 1985. Revision of the Indo-Pacific dartfishes, genus *Ptereleotris* (Perciformes: Gobioidei). *Indo-Pacific Fishes* 7: 1–36.

Randall JE, Hoese DF. **1986**. Revision of the groupers of the Indo-Pacific genus *Plectropomus* (Perciformes: Serranidae). *Indo-Pacific Fishes* 13: 1–31.

Randall JE, Holcom RR. **2001**. *Antennatus linearis*, a new Indo-Pacific species of frogfish (Lophiiformes: Antennariidae). *Pacific Science* 55(2): 137–144.

Randall JE, Hoover JP. **1993**. *Pseudanthias marcia*, a new serranid fish from Oman. *Revue française d'Aquariologie Herpétologie* 20(2): 47–52.

Randall JE, Hoover JP. **1995**. *Scarus zufar*, a new species of parrotfish from southern Oman, with comments on endemism of the area. *Copeia* 1995(3): 683–688.

Randall JE, Hutomo M. 1988. Redescription of the Indo-Pacific serranid fish *Pseudanthias bimaculatus* (Smith). *Copeia* 1988(3): 669–673.

Randall JE, Ida H. **2014**. Three new species of sand lances (Perciformes: Ammodytidae) from the southwest Indian Ocean. *Journal of the Ocean Science Foundation* 12: 1–11. Randall JE, Ida H, Earle JL. **1994**. *Ammodytoides pylei*, a new species of sand lance (Ammodytidae) from the Hawaiian Islands. *Pacific Science* 48(1): 80–89.

Randall JE, Ida H, Kato K, Pyle RL, Earle JL. **1997**. *Annotated checklist of the inshore fishes of the Ogasawara Islands*. Tokyo: National Science Museum. 74 pp.

Randall JE, Johnson JW. **2000**. *Perca lineata* and *P. vittata* established as valid species of *Plectorhinchus* (Perciformes: Haemulidae). *Memoirs of the Queensland Museum* 45(2): 477–482.

Randall JE, Johnson JW. **2007**. Revision of the soleid fish genus *Pardachirus. Indo-Pacific Fishes* **39**: 1–22, Pls. 1–4.

Randall JE, Kay JC. **1974**. *Stethojulis axillaris*, a junior synonym of the Hawaiian labrid fish *Stethojulis balteata*, with a key to the species of the genus. *Pacific Science* 28(2): 101–107.

Randall JE, Khalaf MA. 2003. Redescription of the labrid fish Oxycheilinus orientalis (Günther), a senior synonym of O. rhodochrous (Günther), and the first record from the Red Sea. Zoological Studies 42(1): 135–139.

Randall JE, King DR. 2009. Parupeneus fraserorum, a new species of goatfish (Perciformes: Mullidae) from South Africa and Madagascar. Smithiana Bulletin 10: 31–35.

Randall JE, King DR. **2010**. *Halichoeres zulu*, a new labrid fish from South Africa. *Smithiana* Bulletin 11: 17–23.

Randall JE, Klausewitz W. **1973**. A review of the triggerfish genus *Melichthys*, with description of a new species from the Indian Ocean. *Senckenbergiana Biologica* 54(1/3): 57–69.

Randall JE, Klausewitz W. 1977. Centropyge flavipectoralis, a new angelfish from Sri Lanka (Ceylon) (Pisces: Teleostei: Pomacanthidae). Senckenbergiana Biologica 57(4/6) [for 1976]: 235–240.

Randall JE, Klausewitz W. **1986**. Tiefenwasser- und Tiefseefische aus dem Roten Meer. XIV. New records of the serranid fish *Epinephelus radiatus* (Day) from the Red Sea and Gulf of Oman (Pisces: Perciformes: Serranidae). *Senckenbergiana Maritima* 18(3/6): 229–237.

Randall JE, Kotthaus A. 1977. Suezichthys tripunctatus, a new deep-dwelling Indo-Pacific labrid fish. Meteor Forschungsergebnisse (Reihe D, Biologie) 24: 33–36.

Randall JE, Kuiter RH. **1989**. The juvenile Indo-Pacific grouper *Anyperodon leucogrammicus*, a mimic of the wrasse *Halichoeres purpurescens* and allied species, with a review of the recent literature on mimicry of fishes. *Revue française d'Aquariologie Herpétologie* 16(2): 51–56.

Randall JE, Kuiter RH. **2007**. *Wetmorella tanakai*, a new wrasse (Perciformes: Labridae) from Indonesia and the Philippines. *aqua, International Journal of Ichthyology* 13(1): 1–6.

Randall JE, Kulbicki M. **1998**. Two new cardinalfishes (Perciformes: Apogonidae) of the *Apogon cyanosoma* complex from the western Pacific, with notes on the status of *A. wassinki* Bleeker. *Revue française d'Aquariologie Herpétologie* 25(1/2): 31–39.
Randall JE, Kulbicki M. 2006. A review of the goatfishes of the genus *Upeneus* (Perciformes: Mullidae) from New Caledonia and the Chesterfield Bank, with a new species and four new records. *Zoological Studies* 45(3): 298–307.

Randall JE, Lachner EA. 1986. The status of the Indo-West Pacific cardinalfishes Apogon aroubiensis and A. nigrofasciatus. Proceedings of the Biological Society of Washington 99(1): 110–120.

Randall JE, Lachner EA, Fraser TH. **1985**. A revision of the Indo-Pacific apogonid fish genus *Pseudamia*, with descriptions of three new species. *Indo-Pacific Fishes* 6: 1–23.

Randall JE, Lim KKP (eds). 2000. A checklist of the fishes of the South China Sea. *Raffles Bulletin of Zoology* Supplement 8: 569–667.

Randall JE, Lourie SA. **2009**. *Hippocampus tyro*, a new seahorse (Gasterosteiformes: Syngnathidae) from the Seychelles. *Smithiana* Bulletin 10: 19–21.

Randall JE, Lubbock R. 1981. A revision of the serranid fishes of the subgenus *Mirolabrichthys* (Anthiinae: *Anthias*), with description of five new species. *Contributions in Science* (Los Angeles) 333: 1–27.

Randall JE, Lubbock R. 1981. Labrid fishes of the genus *Paracheilinus*, with descriptions of three new species from the Philippines. *Japanese Journal of Ichthyology* 28(1): 19–30, Pls. 1–2.

Randall JE, Lubbock R. **1982**. A new Indo-Pacific dartfish of the genus *Ptereleotris* (Perciformes: Gobiidae). *Revue française d'Aquariologie Herpétologie* 9(2): 41–46.

Randall JE, Matsuura K, Zama A. **1978**. A revision of the triggerfish genus *Xanthichthys*, with description of a new species. *Bulletin of Marine Science* 28(4): 688–706.

Randall JE, Maugé LA [Maugé AL]. 1978. Holacanthus guezei, a new angelfish from Reunion. Bulletin du Muséum National d'Histoire Naturelle (Série 3) 514 (Zoologie 353): 297–303.

Randall JE, McCarthy LJ. 1988. A new damselfish of the genus *Chromis* (Perciformes: Pomacentridae) from the Persian Gulf and Gulf of Oman. *Revue française d'Aquariologie Herpétologie* 14(4): 133–136.

Randall JE, McCarthy LJ. **1989**. *Solea stanalandi*, a new sole from the Persian Gulf. *Japanese Journal of Ichthyology* 36(2): 196–199.

Randall JE, McCosker JE. **1975**. The eels of Easter Island with a description of a new moray. *Contributions in Science* (Los Angeles) 264: 1–32.

Randall JE, McCosker JE. **1992**. Revision of the fish genus *Luzonichthys* (Perciformes: Serranidae: Anthiinae), with descriptions of two new species. *Indo-Pacific Fishes* 21: 1–21.

Randall JE, McCosker JE. **1993**. Social mimicry in fishes. *Revue française d'Aquariologie Herpétologie* 20(1): 5–8. Randall JE, McCosker JE. 2002. Parapercis lata, a new species of sandperch (Perciformes: Pinguipedidae) from the central Pacific. Proceedings of the California Academy of Sciences (Series 4) 72: 1–19.

Randall JE, Mee JKL. **1994**. *Pardachirus balius*, a new sole (Pleuronectiformes: Soleidae) from Oman. *Fauna of Saudi Arabia* 14: 341–347.

Randall JE, Miroz A. **2001**. *Thalassoma lunare* x *Thalassoma rueppellii*, a hybrid labrid fish from the Red Sea. *aqua*, *Journal of Ichthyology and Aquatic Biology* 4(4): 131–134.

Randall JE, Munroe TA. **2008**. *Soleichthys dori*, a new sole (Pleuronectiformes: Soleidae) from the Red Sea. *Electronic Journal of Ichthyology, Bulletin of the European Ichthyology Society* 2: 76–84.

Randall JE, Nelson GJ. 1979. Scarus japanensis, S. quoyi and S. iserti – valid names for parrotfishes presently known as S. capistratoides, S. blochii and S. croicensis. Copeia 1979(2): 206–212.

Randall JE, Ormond FG. **1978**. On the Red Sea parrotfishes of Forsskål, *Scarus psittacus* and *S. ferrugineus*. *Zoological Journal of the Linnean Society* 63(3): 239–248.

Randall JE, Parenti P. **1999**. Rejection of nine old labrid fish names in order to conserve well-established taxa. *Revue française d'Aquariologie Herpétologie* 26(1/2): 29–32.

Randall JE, Parenti P. **2014**. Parrotfishes are not wrasses. *Reef Encounter* 29(1): 16–18.

Randall JE, Poss SG. **2002**. Redescription of the Indo-Pacific scorpionfish and reallocation to the genus *Sebastapistes*. *Pacific Science* 56(1): 57–64.

Randall JE, Pyle RL. **2001**. Four new serranid fishes of the anthiine genus *Pseudanthias* from the South Pacific. *Raffles Bulletin of Zoology* 49(1): 19–34.

Randall JE, Pyle RM. **1989**. A new species of anthine fish of the genus *Rabaulichthys* (Perciformes: Serranidaie [sic]) from the Maldive Islands. *J.L.B. Smith Institute of Ichthyology Special Publication* 47: 1–7.

Randall JE, Randall HA. **1960**. Examples of mimicry and protective resemblance in tropical marine fishes. *Bulletin of Marine Science of the Gulf and Caribbean* 10(4): 444–480.

Randall JE, Randall HA. **1981**. A revision of the labrid fish genus *Pseudojuloides* with descriptions of five new species. *Pacific Science* **35**(1): 51–74.

Randall JE, Randall HA. **2001**. Review of the fishes of the genus *Kuhlia* (Perciformes: Kuhliidae) of the central Pacific. *Pacific Science* 55(3): 227–256.

Randall JE, Senou H. **2001**. Review of the Indo-Pacific gobiid fish genus *Lubricogobius*, with description of a new species and a new genus for *L. pumilus*. *Ichthyological Research* 48(1): 3–12.

Randall JE, Schultz JK. 2008. Cirrhitops mascarenensis, a new species of hawkfish from the Mascarene Islands, southwestern Indian Ocean. Smithiana Bulletin 9: 15–20. Randall JE, Shen S-C. 1978. A review of the labrid fishes of the genus *Cirrhilabrus* from Taiwan, with description of a new species. *Bulletin of the Institute of Zoology Academia Sinica* (Taipei) 17(1): 13–24.

Randall JE, Shimizu T. **1994**. *Plectranthias pelicieri*, a new anthiine fish (Perciformes: Serranidae) from Mauritius, with notes on *P. gardineri*. *Japanese Journal of Ichthyology* 41(2): 109–115.

Randall JE, Shimizu T, Yamakawa T. **1982**. A revision of the Holocentrid fish genus *Ostichthys*, with description of four new species and a related new genus. *Japanese Journal of Ichthyology* 29(1): 1–25.

Randall JE, Smith CL. 1988. Two new species and a new genus of cardinalfishes (Perciformes: Apogonidae) from Rapa, South Pacific Ocean. *American Museum Novitates* 2926: 1–9.

Randall JE, Smith MM. **1982.** A review of the labrid fishes of the genus *Halichoeres* of the western Indian Ocean, with descriptions of six new species. *Ichthyological Bulletin of the J.L.B. Smith Institute of Ichthyology* 45: 1–26.

Randall JE, Smith MM, Aida K. 1980. Notes on the classification and distribution of the Indo-Pacific soapfish, *Belanoperca chabanaudi* (Perciformes: Grammistidae). *J.L.B. Smith Institute of Ichthyology Special Publication* 21: 1–8.

Randall JE, Spreinat A. 2004. The subadult of labrid fish Novaculoides macrolepidotus, a mimic of waspfishes of the genus Ablabys. aqua, Journal of Ichthyology and Aquatic Biology 8(2): 45–48.

Randall JE, Springer VG. **1973**. The monotypic Indo-Pacific labrid fish genera *Labrichthys* and *Diproctacanthus* with description of a new related genus, *Larabicus*. *Proceedings of the Biological Society of Washington* 86(23): 279–297.

Randall JE, Springer VG. **1975**. *Labroides pectoralis*, a new species of labrid fish from the tropical Western Pacific. *Uo* (Japanese Society of Ichthyologists) 25: 4–11, 22.

Randall JE, Stanaland BE. **1989**. A new dottyback of the genus *Pseudochromis* (Teleostei; Perciformes; Pseudochromidae) from the northwestern Indian Ocean. *Revue française d'Aquariologie Herpétologie* 15(4): 105–110.

Randall JE, Steene RC. 1983. *Rhinecanthus lunula*, a new species of triggerfish from the South Pacific. *Freshwater and Marine Aquarium* 6(7): 45–51.

Randall JE, Stroud GJ. **1985**. On the validity of the mugiloidid fish *Parapercis robinsoni* Fowler. *Japanese Journal of Ichthyology* 32(1): 93–99.

Randall JE, Tarr AB. **1994**. *Trichonotus arabicus* (Perciformes: Trichonotidae), a new species of sand diver from the Arabian Gulf and Oman. *Fauna of Saudi Arabia* 14: 309–316.

Randall JE, Taylor LR Jr. **1988**. Review of the Indo-Pacific fishes of the serranid genus *Liopropoma*, with descriptions of seven new species. *Indo-Pacific Fishes* 16: 1–47.

Randall JE, Van Egmond J. 1994. Marine fishes from the Seychelles: 108 new records. In: van der Land J (ed) Results of the 'Oceanic Reefs' Expedition to the Seychelles (1992– 1993), Vol. 1. Zoologische Verhandelingen Leiden 297: 43–83.

Randall JE, Victor BC. **2013**. *Bodianus atrolumbus* (Valenciennes 1839), a valid species of labrid fish from the southwest Indian Ocean. *Journal of the Ocean Science Foundation* 8: 44–61. http://dx.doi.org/10.5281/ zenodo.1041974

Randall JE, Victor BC. **2014**. Four new fishes of the genus *Pempheris* (Perciformes: Pempheridae) from the western Indian Ocean. *Journal of the Ocean Science Foundation* 12: 61–83. http://dx.doi.org/10.5281/zenodo.1049113

Randall JE, Victor BC. **2015**. Descriptions of thirty-four new species of the fish genus *Pempheris* (Perciformes: Pempheridae), with a key to the species of the western Indian Ocean. *Journal of the Ocean Science Foundation* 18: 1–77. http://doi.org/10.5281/zenodo.1045550

Randall JE, Victor BC, Alpermann TJ, Bogorodsky SV, Mal AO, Satapoomin U, Bineesh KK. **2014**. Rebuttal to Koeda *et al.* (2014) on the Red Sea fishes of the perciform genus *Pempheris. Zootaxa* 3887(3): 377–392. http://dx.doi. org/10.11646/zootaxa.3887.3.5

Randall JE, Whitehead PJP. **1985**. *Epinephelus cyanopodus* (Richardson), a senior synonym of *E. hoedtii* (Bleeker), and comparison with the related *E. flavocaeruleus* (Lacepède). *Cybium* 9(1): 29–39.

Randall JE, Williams JT, Rocha LA. **2008**. The Indo-Pacific tetraodontid fish *Canthigaster coronata*, a complex of three species. *Smithiana* Bulletin 9: 3–13.

Randall JE, Williams JT, Smith DG, Kulbicki M, Mou Tham G, Labrosse P, Kronen M, Clua E, Mann BS. **2003**. Checklist of the shore and epipelagic fishes of Tonga. *Atoll Research Bulletin* 502: ii + 1–35.

Randall JE, Wrobel L. **1988**. A new species of soldierfish of the genus Ostichthys and records of O. archiepiscopus and O. sandix from Tahiti. Japanese Journal of Ichthyology 35(3): 243–246.

Randall JE, Yamakawa T. **2006**. *Parapercis phenax* from Japan and *P. banoni* from the southeast Atlantic, new species of pinguipedid fishes previously identified as *P. roseoviridis*. *Zoological Studies* 45(1): 1–10.

Ranzani C. **1839**. Dispositio familiae Molarum in genera et in species. *Novi Commentarii Academiae Scientiarum Instituti Bononiensis* 3: 63–82, Pls. 6.

Ranzani C. **1841**. De nonnullis novis speciebus Piscium, Opusculum tertium. Nuovi annali delle scienze naturali e rendiconto dei lavori dell'Accademia della Scienze dell'Instituto di Bologna con appendice agraria. Bologna Anno III, Tomo V: 60–66.

Ranzani C. **1842**. De novis speciebus piscium. Dissertatio secunda. *Novi Commentarii Academiae Scientiarum Instituti Bononiensis* 5: 3–22.

Ranzani C. 1842. De novis speciebus piscium. Dissertatio IV. Novi Commentarii Academiae Scientiarum Instituti Bononiensis 5: 339–365.

Rao BVS. 1974. On the little known electric ray *Narke* (*Bengalichthys*) *impennis* from India. *Copeia* 1974(3): 791–792.

Rao DM, Rao KS. 1981. A revision of the genus Scolopsis Cuvier (Pisces: Nemipteridae) with descriptions of two new species from Indian waters. Proceedings of the Koniklijke Nederlandse Akademie van Wetenschappen (Series C) 84(1): 131–141.

Rao DV, Devi K, Ranjan PT. 2000. An account of ichthyofauna of Andaman and Nicobar Islands, Bay of Bengal. Occasional Papers of the Zoological Survey of India 178: 1–434.

Rao VV. 1966. Platycephalus bengalensis sp. nov. from Bay of Bengal. Annals and Magazine of Natural History (Series 13) 9(97–99): 123–127.

Rao VV. 1970. Sexual dimorphism in coloration among band fishes of the family Cepolidae. *Journal of the Bombay Natural History Society* 66(2) [for 1969]: 388–390.

Rao VV, Dutt S. **1966**. A new fish *Tentaculus waltairiensis* gen. et sp. nov. (family: Haliophidae) from Indian waters. *Annals* and Magazine of Natural History (Series 13) 8(92) [for 1965]: 455–459.

Rapson WS, Schwartz HM, van Rensburg NJ. 1945. South African fish products. Part XIII. The jacopever, *Sebastichthys capensis* (Gmelin) and the sancord, *Helicolenus maculatus* (C. and V.). *Journal of the Society of Chemical Industry*, London (Series B) 64: 47–50.

Rass TS. **1971**. Fish (pp. 15–18). In: *The life of animals*. Vol. 4. Moscow: Prosveschenie. 575 pp. [In Russian]

Ratnasingham S, Hebert PDN. **2007**. BOLD: The Barcode of Life Data system (www.barcodinglife.org). *Molecular Ecology Notes* 7(3): 355–364.

Rauther M. **1924**. Cyclostomi. In: *Bronn's Klassen und* Ordnungen des Tier-Reichs 6(1): 583–701.

Raven, HC. 1939. Notes on the anatomy of *Ranzania truncate*. *American Museum Novitates* 1038: 1–7.

Raven HC, Pflueger A. 1939. On the anatomy and evolution of the locomotor apparatus of the nipple-tailed sunfish (*Masturus lanceolatus*). Bulletin of the American Museum of Natural History 76(4): 143–150.

Ray C. **1969**. Marine parks and conservation in Kenya. Report to Director of Parks on behalf of the African Wildlife Leadership Foundation. [online at http://www.reefbase.org/ resource\_center/publication/pub\_4657.aspx]

Reader SE, Leis JM. 1996. Larval development in the lutjanid subfamily Caesioninae (Pisces): the genera *Caesio*, *Dipterygonotus*, *Gymnocaesio* and *Pterocaesio*. *Bulletin of Marine Science* 59: 310–369. Reeves C, De Wit M. **2000**. Making ends meet in Gondwana: retracing the transforms of the Indian Ocean and reconnecting continental shear zones. *Terra Nova* 12: 272–280.

Regan CT. 1902. On the fishes from the Maldive Islands. I.
Dredged. II. Freshwater. (pp. 272–281). In: Gardiner JS
(ed) The fauna and geography of the Maldive and Laccadive archipelagoes: being the account of the work carried on and of the collections made by an expedition during the years 1899 and 1900. Vol. 1(3). Cambridge: Cambridge University Press. 346 pp.

Regan CT. **1903**. Descriptions de poissons nouveaux faisant partie de la collection du Musée d'Histoire Naturelle de Genève. *Revue Suisse de Zoologie* 11(2): 413–418.

Regan CT. **1904**. Descriptions of three new marine fishes from South Africa. *Annals and Magazine of Natural History* (Series 7) 14(80): 128–130.

Regan CT. **1905**. On a collection of fishes from the Inland Sea of Japan made by Mr. R. Gordon Smith. *Annals and Magazine of Natural History* (Series 7) 15(85): 17–26.

Regan CT. **1905**. On fishes from the Persian Gulf, the Sea of Oman, and Karachi, collected by Mr. F.W. Townsend. *Journal of the Bombay Natural History Society* 16: 318–333.

Regan CT. **1906**. Descriptions of some new sharks in the British Museum Collection. *Annals and Magazine of Natural History* (Series 7) 18(108, art.70): 435–440.

Regan CT. **1906**. Descriptions of new or little-known fishes from the coast of Natal. *Annals of the Natal Museum* 1(1): 1–6.

Regan CT. **1907**. Descriptions of the teleostean fish *Velifer hypselopterus*, and of a new species of the genus *Velifer*. *Proceedings of the Zoological Society of London* 1907: 633–634.

Regan CT. **1908**. A collection of fishes from the coasts of Natal, Zululand, and Cape Colony. *Annals of the Natal Museum* 1(3): 241–255.

Regan CT. **1908**. A synopsis of the sharks of the family Scyliorhinidae. *Annals and Magazine of Natural History* (Series 8) 1(6): 453–465.

Regan CT. **1908**. A synopsis of the sharks of the family Squalidae. *Annals and Magazine of Natural History* (Series 8) 2(7): 39–57.

Regan CT. 1908. Report on the marine fishes collected by Mr. J. Stanley Gardiner in the Indian Ocean. *Transactions of the Linnean Society of London* (2<sup>nd</sup> series) Zoology 12(3, art.14): 217–255.

Regan CT. **1909**. A collection of fishes made by Dr. C.W. Andrews, F.R.S., at Christmas Island. *Proceedings of the Zoological Society of London* 1909(2): 403–406.

Regan CT. **1909**. The classification of teleostean fishes. *Annals and Magazine of Natural History* (Series 8) 3(13): 75–86.

Regan CT. **1911**. The anatomy and classification of the teleostean fishes of the order Iniomi. *Annals and Magazine of Natural History* (Series 8) 7(37): 120–133.

Regan CT. **1912**. A synopsis of the myxinoids of the genus *Heptatretus* or *Bdellostoma*. *Annals and Magazine of Natural History* (Series 8) 9(53): 534–536.

Regan CT. **1912**. The classification of the blennioid fishes. *Annals and Magazine of Natural History* (Series 8) 10(57): 265–280.

Regan CT. 1912. New fishes from Aldabra and Assumption, collected by Mr. J.C.F. Fryer. *Transactions of the Linnean Society of London* (2<sup>nd</sup> series) Zoology 15(2, no. 18): 301–302.

Regan CT. **1913**. A collection of fishes made by Professor Francisco Fuentes at Easter Island. *Proceedings of the General Meetings for Scientific Business of the Zoological Society of London* 1913(3): 368–374.

Regan CT. **1913**. The Antarctic fishes of the Scottish National Antarctic Expedition. *Transactions of the Royal Society of Edinburgh* 49(2): 229–292.

Regan CT. 1913. The osteology and classification of the teleostean fishes of the order Scleroparei. *Annals and Magazine of Natural History* (Series 8) 11(62): 169–184.

Regan CT. **1913**. A revision of the myxinoids of the genus *Myxine*. *Annals and Magazine of Natural History* (Series 8) 11(64): 395–398.

Regan CT. **1913**. Note on *Myxine capensis*. *Annals and Magazine of Natural History* (Series 8) 12(68): 229.

Regan CT. **1914**. Diagnoses of new marine fishes collected by the British Antarctic (*Terra Nova*) Expedition. *Annals and Magazine of Natural History* (Series 8) 13(73): 11–17.

Regan CT. **1914**. Fishes. British Antarctic (*Terra Nova*) expedition, 1910. *Natural History Report (Zoology)* 1(1): 1–54.

Regan CT. **1915**. A collection of fishes from Lagos. *Annals and Magazine of Natural History* (Series 8) 15(85): 124–130.

Regan CT. 1916. Larval and post-larval fishes. British Antarctic (*Terra Nova*) expedition, 1910. *Natural History Report* (*Zoology*) 1(4): 125–156.

Regan CT. **1916**. Fishes from Natal, collected by Mr. Romer Robinson. *Annals of the Durban Museum* 1(3): 167–170.

Regan CT. **1917**. Additions to the fish fauna of Natal. *Annals of the Durban Museum* 1(5): 458–459.

Regan CT. **1917**. A revision of the clupeid fishes of the genera *Sardinella, Harengula, &c. Annals and Magazine of Natural History* (Series 8) 19(113): 377–395.

Regan CT. **1918**. Further additions to the fish fauna of Natal. *Annals of the Durban Museum* 2(2): 76–77.

Regan CT. 1919. Fishes from Durban, Natal, collected by Messrs. H.W. Bell Marley and Romer Robinson. Annals of the Durban Museum 2(4): 197–204.

Regan CT. **1920**. A revision of the flat-fishes (Heterosomata) of Natal. *Annals of the Durban Museum* 2(5): 205–222.

Regan CT. **1920**. Freshwater fishes from Madagascar. *Annals* and Magazine of Natural History (Series 9) 5(29): 419–424.

Regan CT. **1921**. New fishes from deep water off the coast of Natal. *Annals and Magazine of Natural History* (Series 9) 7(41): 412–420.

Regan CT. **1921**. Three new fishes from South Africa, collected by Mr. H.W. Bell Marley. *Annals of the Durban Museum* 3(1): 1–2.

Reid A, Macfadyen G, Grinyer T, Al-Sakaf H. **2002**. Socotra Archipelago Master Plan 2001–2010. YEM/B7-3000/ IB/97/0787. Phase II: Final Report. Project 5: Fisheries Feasibility Study. [unpaginated]

Reid ED. **1943**. Review of the genera of blennioid fishes related to *Ophioblennius*. *Journal of the Washington Academy of Sciences* 33(12): 373–384.

Reinhardt JCH. 1825. Ichthyologiske bidrag. In: Orsted HC (ed) Oversigt over det Kongelige Danske Videnskabernes Selskabs Forhandlinger og dets Medlemmers Arbeider (Kjøbenhavn) (1824–25). pp. 1–35.

Reinhardt JCH. **1837**. Icthyologiske bidrag til den Grönlandske fauna. Indledning, indeholdende tillaeg og forandringer i den fabriciske fortegnelse paa Grönlandske hvirveldyr. *Det Kongelige Danske videnskabernes selskabs naturvidenskabelige og mathematiske afhandlinger* 7: 83– 196. [dated 1838]

Reinthal PN, Stiassny MLJ. **1997**. Revision of the Madagascan genus *Ptychochromoides* (Teleostei: Cichlidae), with description of a new species. *Ichthyological Exploration of Freshwaters* 7(4): 353–368.

Renaud CB. **1997**. Conservation status of northern hemisphere lampreys (Petromyzontidae). *Journal of Applied Ichthyology* 13(3): 143–148.

Renaud CB. **2011**. Lampreys of the world: an annotated and illustrated catalogue of lamprey species known to date. *FAO Species Catalogue for Fishery Purposes* No. 5. Rome: FAO. 109 pp.

Renaud CB, Economidis PS. **2010**. *Eudontomyzon graecus*, a new nonparasitic lamprey from Greece (Petromyzontiformes: Petromyzontidae). *Zootaxa* 2477: 37–48. http://dx.doi.org/10.11646/zootaxa.2477.1.3

Rendahl H. 1921. Results of Dr E. Mjöberg's Swedish Scientific Expedition to Australia, 1910–1913. XXVIII.
Fische. *Kungliga Svenska Vetenskapsakademiens handlingar* 61(9): 1–24.

Rendahl H. **1922**. A contribution to the ichthyology of northwest Australia. *Meddelelser Fra Det Zoologiske Museum, Kristiania* 5: 163–197.

Rendahl H. **1933**. Zur Osteologie und Myologie des Schultergürtels und der Brustflosse einiger Scleroparei. *Arkiv för Zoologi* 26A(12): 1–50. Rendahl H. **1933**. Studien über die Scleroparei. I. Zur Kenntnis der kranialen Anatomie der Agoniden. *Arkiv för Zoologi* 26A(13): 1–106.

Rennis DS, Hoese DF. **1985**. A review of the genus *Parioglossus*, with descriptions of six new species (Pisces, Gobioidei). *Records of the Australian Museum* 36(4): 169–201.

Restrepo V, Prince ED, Scott GP, Uozumi Y. **2003**. ICCAT stock assessments of Atlantic billfish. *Marine and Freshwater Research* 54(4): 361–367.

Retzius AJ. **1799**. *Lampris*, en ny fiskslaegt. *Kongliga Vetenskaps Academiens Handlingar* (Stockholm) 20: 91–100.

Rey LL, see Lozano Rey L.

Rhasis Erazi RA. **1943**. *Leiognathus mediterraneus* nov sp. *Compte Rendu Annuel / Société Turque Sciences Physiques et Naturelles* 10(1942–43): 49–53.

Richards WJ. **1968**. Eastern Atlantic Triglidae (Pisces, Scorpaeniformes). *Atlantide Report* 10: 77–114.

Richards WJ. **1969**. Elopoid leptocephali from Angolan waters. *Copeia* 1969(3): 515–518.

Richards WJ. **1984**. Triglidae. Gurnards and searobins. In: Fischer W, Bianchi G (eds) *FAO species identification sheets for fishery purposes. Western Indian Ocean (Fishing Area 51).* Vol. 5. Rome: FAO. [unpaginated]

Richards WJ. **1992**. Comments on the genus *Lepidotrigla* (Pisces: Triglidae) with descriptions of two new species from the Indian and Pacific Oceans. *Bulletin of Marine Science* 51(1): 45–65.

Richards WJ. 1999. Triglidae. Gurnards, sea robins (also, armoured gurnards, armoured sea robins) (pp. 2359–2382).
In: Carpenter KE, Niem VH (eds) *FAO species identification guide for fishery purposes. The living marine resources of the western central Pacific.* Vol. 4. Bony fishes part 2 (pp. 2069–2790). Rome: FAO.

Richards WJ. 2000. Family Peristediidae (armoured gurnards) (p. 607). In: Randall JE, Lim KKP (eds) A checklist of the fishes of the South China Sea. *The Raffles Bulletin of Zoology* 2000 Supplement 8: 569–667.

Richards WJ. 2005. Xiphiidae: swordfish (pp. 2241–2243).
In: Richards WJ (ed) Early stages of Atlantic fishes. An identification guide for the western central North Atlantic.
Boca Raton, Florida: CRC Press. 1312 pp.

Richards WJ, Luthy SA. 2005. Istiophoridae: billfishes
(pp. 2231–2240). In: Richards WJ (ed) Early stages of
Atlantic fishes. An identification guide for the western central
North Atlantic. Boca Raton, Florida: CRC Press. 1312 pp.

Richards WJ, Miller PJ. **1981**. Peristediidae. Armoured searobins. In: Fischer W, Bianchi G, Scott WB (eds) *FAO species identification sheets for fishery purposes. Eastern central Atlantic (Fishing Area 34 and part of 47)*. Vol. 3. Rome: FAO. [unpaginated] Richards WJ, Saksena VP. **1977**. Systematics of the gurnards, genus *Lepidotrigla* (Pisces, Triglidae), from the Indian Ocean. *Bulletin of Marine Science* 27(2): 208–222.

Richards WJ, Saksena VP. **1980**. *Trigla (Triglaporus) africana* Smith, a synonym of *Chelidonichthys lastoviza* (Bonnaterre) (Pisces: Triglidae). *Copeia* 1980(1): 156–157.

Richards WJ, Yato T. **2014**. Revision of the subgenus *Parapterygotrigla* (Pisces, Triglidae, *Pterygotrigla*). *Zootaxa* 3768(1): 23–42. https://doi.org/10.11646/zootaxa.3768.1.2

Richards WJ, Yato T, Last PR. **2003**. Revision of the gurnard fish subgenus *Otohime* (Triglidae: *Pterygotrigla*). *Smithiana*, *Publications in Aquatic Biodiversity* Bulletin 2: 1–18.

Richardson AJ, Maharaj G, Compagno LJV, Leslie RW, Ebert DA, Gibbons MJ. 2000. Abundance, distribution, morphometrics, reproduction and diet of the Izak catshark. *Journal of Fish Biology* 56(3): 552–576.

Richardson J. **1840**. On some new species of fishes from Australia. *Proceedings of the Zoological Society of London* 8: 25–30.

Richardson J. **1842**. Contributions to the ichthyology of Australia. *Annals and Magazine of Natural History* (Series 1) 9(55): 15–36.

Richardson J. **1842**. Contributions to the ichthyology of Australia. *Annals and Magazine of Natural History* (Series 1) 9(57): 207–218.

Richardson J. **1842**. Contributions to the ichthyology of Australia. *Annals and Magazine of Natural History* (Series 1) 9(59): 384–393.

Richardson J. **1843**. Report on the present state of the ichthyology of New Zealand. *Report of the British Association for the Advancement of Science* 12: 12–30.

Richardson J. **1844–1845**. Ichthyology. In: Hinds RB (ed) *The zoology of the voyage of H.M.S.* Sulphur, *under the command of Captain Sir Edward Belcher R.N., C.B., F.R.G.S., etc., during the years 1836–42*. London: Smith, Elder & Co. pp. 51–150, Pls. 35–64 [Part 1(1844): 51–70, Pls. 35–44; Part 2(1845): 71–98, Pls. 45–54; Part 3(1845): 99–150, Pls. 55–64]

Richardson J. 1844–1848. Ichthyology of the voyage of H.M.S. *Erebus & Terror* under the command of Captain Sir James Clark Ross, R.N., F.R.S. In: Richardson J, Gray JE (eds) *The zoology of the voyage of H. M. S.* Erebus & Terror, *under the command of Captain Sir J. C. Ross, R. N., F. R. S., during the years 1839 to 1843.* Vol. 2(2). London: Edward Newman. viii + 139 pp., Pls. 1–60. [1844: 1–16; 1845: 17–52; 1846: 53–74; 1848: i–viii + 75–139 pp.]

Richardson J. 1845. Generic characters of *Gasterochisma* melampus, a fish which inhabits Port Nicholson, New Zealand. Annals and Magazine of Natural History (Series 1) 15(99): 346.

Richardson J. **1846**. Descriptions of six fish (pp. 484–497). In: Stokes JL (ed) *Discoveries in Australia; with an account of the coasts and rivers explored and surveyed during the voyage*  of H.M.S. Beagle, in the years 1837-38-39-40-41-42-43. Also a narrative ... islands of the Arafura Sea. Vol. 1. London: T. & W. Boone. 521 pp.

Richardson J. **1846**. Report on the ichthyology of the seas of China and Japan. *Report of the British Association for the Advancement of Science 15<sup>th</sup> meeting* [1845]: 187–320.

Richardson J. 1848. Fishes. In: Adams A (ed) The zoology of the voyage of H.M.S. Samarang under the command of Captain Sir Edward Belcher, C.B., F.R.A.S., F.G.S., during the years 1843–46. London: Reeve and Benham. 28 pp.

Richardson J. **1849**. Description of Australian fish. *Proceedings of the Zoological Society of London* 3(art.5): 133–185.

Richardson J. 1856. Ichthyology (pp. 204–331). In: The Encyclopaedia Britannica, or Dictionary of Arts, Sciences, and General Literature (8<sup>th</sup> edition). Vol 12. Edinburgh: A & C Black.

Richardson MK, Admiraal J, Wright GM. **2011**. Developmental anatomy of lampreys. *Biological Reviews* 85(1): 1–33. http:// dx.doi.org/10.1111/j.1469-185X.2009.00092.x

Richter C, Abu-Hilal A. 2006. Seas of the Arabian region (pp. 1373–1412). In: Robinson AR, Brink K (eds) *The sea*, *Vol. 14B: The global coastal ocean: Interdisciplinary regional studies and syntheses.* Cambridge, Massachusetts: Harvard University Press. 809 pp. (i–xxiv + 783–1567).

Riegl B, Schleyer MH, Cook PJ, Branch GM. **1995**. Structure of Africa's southernmost coral communities. *Bulletin of Marine Science* 56: 648–663.

Rijnsdorp AD, Witthames PR. **2005**. Ecology of reproduction (pp. 68–93). In: Gibson RN (ed) *Flatfishes: Biology and exploitation*. Oxford: Blackwell Science. xxiv + 391 pp.

Rilov G, Benayahu Y. 2000. Fish assemblage on natural vs. vertical artificial reefs: the rehabilitation perspective. *Marine Biology* 36: 931–942.

Rimbu N, Lohmann G, Felis T, Pätzold J. **2003**. Shift in ENSO teleconnections recorded by a northern Red Sea coral. *Journal of Climate* 16: 1414–1422.

Rincón G, Stehmann MFW, Vooren CM. 2001. Results of the research cruises of FRV *Walther Herwig* to South America. LXXIV. *Benthobatis kreffti* n. sp. (Chondrichthyes, Torpediniformes, Narcinidae), a new deep-water electric ray from off South Brazil and the third species of the genus. *Archive of Fishery and Marine Research* 49(1): 45–60.

Risk MJ, Sluka R. 2000. Chapter 11. The Maldives: a nation of atolls (pp. 325–352). In: McClanahan TR, Sheppard CRC, Obura DO (eds) *Coral reefs of the Indian Ocean: Their ecology and conservation*. Oxford: Oxford University Press. xxiii + 525 pp.

Risso A. **1810**. *Ichthyologie de Nice, ou histoire naturelle des poissons du departement des Alpes Maritimes*. Paris: F. Schoell. xxxvi + 388 pp.

Risso A. **1827**. *Histoire naturelle des principales productions de l'Europe méridionale, et particulièrement de celles des* 

environs de Nice et des Alpes maritimes. Vol. 3. Paris & Strasbourg: FG Levrault. xvi + 480 pp., Pls. 1–16.

Ritchie W. **1918**. *The history of the South African College 1829–1918*. T. Maskew Miller. **918** pp.

Riva-Rossi C, Barrasso DA, Quiroga AP, Baigún C, Basso NG.
2020. Revalidation of the Argentinian pouched lamprey *Geotria macrostoma* (Burmeister, 1868) with molecular and morphological evidence. *PLoS ONE* 15(5): e0233792 (26 pp). http://dx.doi.org/10.1371/journal.pone.0233792

Rivas LR. **1964**. Western Atlantic serranid fishes (groupers) of the genus *Epinephelus*. *Quarterly Journal of the Florida Academy of Sciences* 27(1): 17–30.

Rivas LR. **1975**. Synopsis of biological data on blue marlin, *Makaira nigricans* Lacepède, 1802. *NOAA Technical Report* NMFS SSRF-675 (Part 3): 1–16.

Roberts C. **1989**. Reproductive mode in the percomorph fish genus *Polyprion* Oken. *Journal of Fish Biology* 34: 1–9.

Roberts CD. **1993**. Comparative morphology of spined scales and their phylogenetic significance in the Teleostei. *Bulletin of Marine Science* 52(1): 60–113.

Roberts CD, Stewart AL, Struthers CD (eds). 2015. *The Fishes* of New Zealand. 4 volumes. Wellington: Te Papa Press.

Roberts CM, Shepherd ARD, Ormond RFG. **1992**. Large-scale variation in assemblage structure of Red Sea butterflyfishes and angelfishes. *Journal of Biogeography* **19**: 239–250.

Roberts TR. **1978**. An ichthyological survey of the Fly River in Papua New Guinea with descriptions of new species. *Smithsonian Contributions to Zoology* 281: 1–72.

Roberts TR. **1998**. Systematic observations on tropical Asian medakas or ricefishes of the genus *Oryzias*, with descriptions of four new species. *Ichthyological Research* 45(3): 213–224.

Roberts TR. **2012**. *Systematics, biology, and distribution of the species of the oceanic oarfish genus* Regalecus *(Teleostei, Lampridiformes, Regalecidae).* Paris: Publications Scientifiques du Museum. 268 pp.

Robertson DR, Reinboth R, Bruce RW. **1982**. Gonochorism, protogynous sex-change and spawning in three Sparisomatinine parrotfishes from the Western Indian Ocean. *Bulletin of Marine Science* **32**(4): 868–879.

Robillard M, Séret B. **2006**. Cultural importance and decline of sawfish (Pristidae) populations in West Africa. *Cybium* 30(4): 23–30.

Robins CH. **1971**. The comparative morphology of the synaphobranchid eels of the Straits of Florida. *Proceedings of the Academy of Natural Sciences of Philadelphia* 123(7): 153–204.

Robins CH, Robins CR. **1970**. The eel family Dysommidae (including the Dysomminidae and Nettodaridae), its osteology and composition, including a new genus and species. *Proceedings of the Academy of Natural Sciences of Philadelphia* 122(6): 293–335. Robins CH, Robins CR. 1975. New genera and species of dysommine and synaphobranchine eels (Synaphobranchidae) with an analysis of the Dysomminae. *Proceedings of the Academy of Natural Sciences of Philadelphia* 127(18): 249–280.

Robins CR. **1975**. Synopsis of biological data on the longbill spearfish, *Tetrapturus pfluegeri* Robins and de Sylva. *NOAA Technical Report* NMFS SSRF-675 (Part 3): 28–38.

Robins CR, Bailey RM, Bond CE, Brooker JR, Lachner EA, Lea RN, Scott WB. **1991**. Common and scientific names of fishes from the United States and Canada. Fifth edition. *American Fisheries Society, Special Publication* 20: 1–183.

Robins CR, Böhlke JE. **1970**. The first Atlantic species of the ammodytid fish genus *Embolichthys*. *Notulae Naturae* (Academy of Natural Sciences of Philadelphia) 430: 1–11.

Robins CR, de Sylva DP. **1963**. A new western Atlantic spearfish, *Tetrapturus pfluegeri*, with a redescription of the Mediterranean spearfish *Tetrapturus belone*. *Bulletin of Marine Science of the Gulf and Caribbean* 13(1): 84–122.

Robins CR. Nielsen JG. 1970. The R/V *Pillsbury* Deep-Sea Biological Expedition to the Gulf of Guinea, 1964–65 – 16 – *Snyderidia bothrops* a new tropical, amphi-Atlantic species (Pisces, Carapidae). *Studies in Tropical Oceanography* 4(2): 285–293.

Robinson GA. **1976**. Sex reversal in the dageraad *Chrysoblephus cristiceps* (Pisces, Sparidae). *Koedoe* 19: 43–48.

Robison BH. 1975. Observations on living juvenile specimens of the slender mola, *Ranzania laevis* (Pisces, Molidae). *Pacific Science* 29(1): 27–29. https://doi.org/10125/907

Rocha-Olivares A, Rosenblatt RH, Vetter RD. **1999**. Cryptic species of rockfishes (*Sebastes*: Scorpaenidae) in the southern hemisphere inferred from mitochondrial lineages. *Journal of Heredity* 90: 404–411.

Rocha-Olivares A, Rosenblatt RH, Vetter RD. **1999**. Molecular evolution, systematics, and zoogeography of the rockfish subgenus *Sebastomus* (*Sebastes*, Scorpaenidae) based on mitochondrial cytochrome *b* and control region sequences. *Molecular Phylogenetics and Evolution* 11(3): 441–458.

Rocliffe S, Peabody S, Samoilys M, Hawkins J. **2014**. Towards a network of Locally Managed Marine Areas (LMMAs) in the Western Indian Ocean. *PLoS ONE* 9(7): e103000. https://doi.org/10.1371/journal.pone.0103000

Rodriguez M-J, Motta H, Whittington MW, Schleyer M. 2000. Chapter 4. Coral reefs of Mozambique (pp. 107–130). In: McClanahan TR, Sheppard CRC, Obura DO (eds) Coral reefs of the Indian Ocean: Their ecology and conservation. Oxford: Oxford University Press. xxiii + 525 pp.

Roel BA, Melo YC. 1990. Reproductive biology of the round herring *Etrumeus whiteheadi*. South African Journal of Marine Science 9(1): 177–187. Rofen RR. **1958**. The marine fishes of Rennell Island, British Solomon Islands (pp. 149–218). In: Wolff T, Bradley JD (eds) *The natural history of Rennell Island, British Solomon Islands. Vol. 1. Vertebrates.* Copenhagen: Danish Science Press. 228 pp.

Rofen RR. **1963**. Diagnoses of new genera and species of alepisauroid fishes of the family Paralepididae. *Aquatica* 2: 1–7.

Rofen RR. **1966**. Family Paralepididae. Barracudinas. In: Fishes of the western North Atlantic. *Memoir of the Sears Foundation for Marine Research* 1(part 5): 205–461.

Romanov EV, Samorov VV. **1994**. On discoveries of the crocodile shark, *Pseudocarcharias kamoharai* (Pseudocarchariidae), in the equatorial Indian Ocean. *Journal of Ichthyology* 34(4): 155–157.

Rosa IL, Rosa RS. **1987**. *Pinguipes* Cuvier and Valenciennes and Pinguipedidae Günther, the valid names for the fish taxa usually known as Mugiloides and Mugiloididae. *Copeia* 1987(4): 1048–1051.

Röse AF. **1793**. *Petri Artedi Angermannia – Sueci synonymia nominum piscium fere omnium;... Ichthyologiae pars IV* (Editio II). Grypeswaldiae. i–ii + 1–140.

Rose JD, Arlinghaus R, Cooke SJ, Diggles BK, Sawynok W, Wynne CDL. **2014**. Can fish really feel pain? *Fish and Fisheries* 15(1): 97–113. https://doi.org/10.1111/faf.12010 [first published online in 2012]

Rosen BR. 1971. The distribution of reef coral genera in the Indian Ocean (pp. 263–299). In: Stoddart DR, Yonge CM (eds) Regional variation in Indian Ocean coral reefs. Proceedings of a Symposium, organized jointly by the Royal Society of London and the Zoological Society of London. London: Academic Press. 584 pp.

Rosen DE. **1964**. The relationships and taxonomic position of the halfbeaks, killifishes, silversides and their relatives. *Bulletin of the American Museum of Natural History* 127(5): 217–268.

Rosen DE. **1973**. Interrelationships of higher euteleostean fishes (pp. 397–513). In: Greenwood PH, Miles RS, Patterson C (eds) *Interrelationships of fishes*. London: Academic Press. 536 pp.

Rosen DE. **1984**. Zeiforms as primitive plectognath fishes. *American Museum Novitates* 2782: 1–45.

Rosen DE, Bailey RM. **1963**. The poeciliid fishes (Cyprinodontiformes), their structure, zoogeography, and systematics. *Bulletin of the American Museum of Natural History* 126(art.1): 1–176.

Rosen DE, Forey PL, Gardiner BG, Patterson C. 1981. Lungfishes, tetrapods, palaeontology and plesiomorphy. Bulletin of the American Museum of Natural History 167(art.4): 163–275. Rosen DE, Parenti LR. **1981**. Relationships of *Oryzias*, and the groups of atherinomorph fishes. *American Museum Novitates* 2719: 1–25.

Rosen DE, Patterson C. **1969**. The structure and relationships of the paracanthopterygian fishes. *Bulletin of the American Museum of Natural History* 141(art.3): 359–474.

Rosenblatt RH, Butler JL. **1977**. The ribbonfish genus *Desmodema*, with the description of a new species (Pisces: Trachipteridae). *Fishery Bulletin* 75(4): 843–855.

Rosenblatt RH, Johnson GD. 1976. Anatomical considerations of pectoral swimming in the opah, *Lampris guttatus*. *Copeia* 1976(2): 367–370.

Rossignol M, Blache J. 1961. Sur le statut spécifique de deux poissons pélagiques du Golfe de Guinée. Anchoviella guineensis nov. sp. (Clupeiformes, Engraulidae) Atherina lopeziana nov. sp. (Mugiliformes, Atherinidae). Bulletin du Muséum National d'Histoire Naturelle (Paris) (Série 2) 33(3): 285–293.

Rossignol M, Blache J. 1961. Sur un poisson Stromatiidae nouveau du Golfe de Guinée, *Psenes benardi* nov sp. *Bulletin du Muséum National d'Histoire Naturelle* (Paris) (Série 2) 33(4): 384–386.

Roule L. 1916. Notice préliminaire sur quelques espèces nouvelles ou rares des poissons provenant des croisières de S. A. S. le Prince de Monaco. Bulletin de l'Institut Océanographique (Monaco) 320: 1–32.

Roule L. **1921**. Sur un nouveau poisson abyssal (*Scombrolabrax heterolepis*, nov. gen. nov. sp.) pêché dans les eaux de l'île Madère. Comptes rendus hebdomadaires des séances de l'Académie des Sciences 172(24): 1534–1536.

Roule L. 1922. Description de *Scombrolabrax heterolepis* nov. gen. nov. sp., poisson abyssal nouveau de l'île Madère. *Bulletin de l'Institut Océanographique* (Monaco) 408: 1–8.

Roule L. 1923. Un cas probable de mutation chez les poissons. Comptes rendus des séances de la Société de Biologie et de ses filiales, Centre National de la Recherche (Paris Scientifique) 89: 1027–1028.

Roule L. **1924**. Descriptions d'une forme nouvelle d'un poisson appartenant à la famille des Bérycidés, *Actinoberys jugeati* nov. gen. nov. sp. = ? mutation de *Beryx decadactylus* C.V.; suivie d'une revision de cette famille. *Bulletin du Muséum National d'Histoire Naturelle* (Paris) (Série 1) 30(1): 69–74.

Roule L, Angel F. **1933**. Poissons provenant des campagnes du Prince Albert I de Monaco. *Résultats des campagnes scientifiques accomplies sur son yacht par Albert 1<sup>er</sup> Monaco* Fasc. 86: 1–115.

Roux C. 1957. Poissons marins (pp. 139–253). In: Collignon J, Rossignol M, Roux C (eds) Mollusques, crustacés, poissons marins des côtes d'A.E.F. en collection au centre d'Océanographie de l'Institut d'Études Centrafricaines de Pointe-Noire. Paris: ORSTOM. 369 pp. Roux C. **1971**. Révision des poissons marins de la famille des Batrachoididae de la côte occidentale Africaine. *Bulletin du Muséum National d'Histoire Naturelle* (Paris) (Série 2) 4(4): 626–643.

Roux C. 1990. Uranoscopidae (pp. 897–898). In: Quéro J-C, Hureau J-C, Karrer C, Post A, Saldanha L (eds) *Check-list of the fishes of the eastern tropical Atlantic (CLOFETA)*.
Vol. 2. Lisbon: Junta Nacional de Investigação Científica e Tecnológica.

Roux C, Collette BB. **1981**. Batrachoididae. Toadfishes. In: Fischer W, Bianchi G, Scott WB (eds) *FAO species identification sheets for fishery purposes. Eastern central Atlantic (Fishing Area 34 and part of 47)*. Vol. 1. Rome: FAO. [unpaginated]

Roux C, Whitley GP. 1972. Perulibatrachus, nouveau nom de genre de poissons téléostéens de la famille des Batrachoididae, en remplacement de Parabatrachus Roux 1970. Bulletin du Muséum National d'Histoire Naturelle (Paris) (Série 3) 6 [for 1971]: 349–350.

Roux-Estève R. **1956**. Resultats scientifiques des campagnes de la *Calypso*. X. Poissons. *Annales de l'Institut Océanographique* (Monaco) (N.S.) 32: 61–115.

Roux-Estève R, Fourmanoir P. **1955**. VII. Poissons capturés par la mission de la *Calypso* en Mer Rouge. *Annales de l'Institut Océanographique* (Monaco) (N.S.) 30: 195–203.

Roxas HA. **1934**. A review of Phillippine Mugilidae. *Philippine Journal of Science* 54(3): 393–431.

Rüppell WPES. **1828–1830**. *Atlas zu der Reise im nördlichen Afrika von Eduard Rüppell. Fische des Rothen Meeres*. Part 3. Frankfurt-am-Main: Heinrich Ludwig Brönner. pp. 95–141.

Rüppell WPES. **1835–1838**. *Neue Wirbelthiere zu der Fauna von Abyssinien gehörig. Fische des Rothen Meeres.* Frankfurtam-Main: Siegmund Schmerber. 148 pp.

Rüppell WPES. **1835**. Neuer Nachtrag von Beschreibungen und Abbildungen neuer Fische, im Nil entdeckt. *Museum Senckenbergianum: Abhandlungen aus dem Gebiete der beschreibenden Naturgeschichte, von Mitgliedern der Senckenbergischen Naturforschenden Gesellschaft in Frankfurt am Main* 2: 1–28.

Rüppell WPES. **1852**. Verzeichniss der in dem Museum der Senckenbergischen naturforschenden Gesellschaft aufgestellten Sammlungen. Vierte Abtheilung: Fische und deren Skelette. Frankfurt am Main. i–iv + 1–40.

Russ GR, Alcala AC. **1998**. Natural fishing experiments in marine reserves 1983–1993: roles of life history and fishing intensity in family responses. *Coral Reefs* 17(4): 399–416.

Russell BC. **1983**. *Nelabrichthys*, a new genus of labrid fish (Perciformes: Labridae) from the southern Indian and Atlantic oceans. *J.L.B. Smith Institute of Ichthyology Special Publication* 27: 1–7. Russell BC. **1985**. Revision of the Indo-Pacific labrid fish genus *Suezichthys*, with descriptions of four new species. *Indo-Pacific Fishes* 2: 1–21.

Russell BC. **1986**. Review of the western Indian Ocean species of *Nemipterus* Swainson 1839, with description of a new species. *Senckenbergiana Biologica* 67(1/3): 19–35.

Russell BC. **1990**. FAO species catalogue. Vol. 12. Nemipterid fishes of the world (threadfin breams, whiptail breams, monocle breams, dwarf monocle breams, and coral breams) Family Nemipteridae. An annotated and illustrated catalogue of nemipterid species known to date. *FAO Fisheries Synopsis No. 125* (Vol. 12). Rome: FAO. 149 pp.

Russell BC. **1996**. *Parascolopsis capitinis*, a new species of nemipterid fish from Sri Lanka. *Journal of South Asian Natural History* 2(1): 63–66.

Russell BC. 2001. Family Nemipteridae. Threadfin breams (also whiptail breams, monocle breams, dwarf monocle breams, and coral breams) (pp. 3051–3089). In: Carpenter KE, Niem VH (eds) FAO species identification guide for fishery purposes. The living marine resources of the western central Pacific. Vol. 5. Bony fishes part 3 (pp. 2791–3380). Rome: FAO.

Russell BC. **2011**. *Saurida golanii*, a new deep water lizardfish (Pisces: Synodontidae) from the Gulf of Aqaba, northern Red Sea. *Zootaxa* 3098: 21–25. http://dx.doi.org/10.11646/ zootaxa.3098.1.2

Russell BC. **2015**. A new species of *Saurida* (Pisces: Synodontidae) from the Mascarene Plateau, western Indian Ocean. *Zootaxa* 3947(3): 440–446. http://dx.doi. org/10.11646/zootaxa.3947.3.10

Russell BC, Allen GR, Lubbock HR. 1976. New cases of mimicry in marine fishes. *Journal of Zoology* (London) 180: 407–423.

Russell BC, Cressey RF. **1979**. Three new species of Indo-West Pacific lizardfish (Synodontidae). *Proceedings of the Biological Society of Washington* 92(1): 166–175.

Russell BC, Golani D. **1993**. A review of the fish genus *Parascolopsis* (Nemipteridae) of the Western Indian Ocean, with description of a new species from the Northern Red Sea. *Israel Journal of Zoology* 39(4): 337–347.

Russell BC, Golani D, Tikochinski Y. **2015**. *Saurida lessepsianus* a new species of lizardfish (Pisces: Synodontidae) from the Red Sea and Mediterranean Sea, with a key to *Saurida* species in the Red Sea. *Zootaxa* 3956(4): 559–568. http://dx.doi.org/10.11646/zootaxa.3956.4.7

Russell BC, Tweddle D. **2013**. A new species of *Nemipterus* (Pisces: Nemipteridae) from the western Indian Ocean. *Zootaxa* 3630(1): 191–197. https://doi.org/10.11646/ zootaxa.4895.4.7

Russell BC, Westneat MW. **2013**. A new species of *Suezichthys* (Teleostei: Perciformes: Labridae) from the south-eastern Pacific, with a redefinition of the genus and a key to species.

*Zootaxa* 3640(1): 88–94. http://dx.doi.org/10.11646/ zootaxa.3640.1.7

Russell P. **1803**. *Descriptions and figures of two hundred fishes; collected at Vizagapatam on the coast of Coromandel* (2 volumes in one). London: W. Bulmer & Co. [unpaginated]

Rutter CM. **1897**. A collection of fishes obtained in Swatore, China, by Miss Adele M Fielde. *Proceedings of the Academy of Natural Sciences of Philadelphia* 49: 56–90.

## S

Sabaj MH. **2016**. Standard symbolic codes for institutional resource collections in herpetology and ichthyology: an Online Reference. Version 6.5 (16 August 2016). American Society of Ichthyologists and Herpetologists, Washington, DC. [accessible online at http://www.asih.org]

Sabatés A, Olivar MP. **1990**. Early development and spawning of *Sebastes capensis* in the Southeast Atlantic. *Marine Biology* 107(3): 389–395.

Sadovy Y, Cornish AS. **2000**. *Reef fishes of Hong Kong*. Hong Kong University Press. xi + 321 pp.

Sadovy Y, Donaldson TJ. **1995**. Sexual pattern of *Neocirrhites armatus* (Cirrhitidae) with notes on other hawkish species. *Environmental Biology of Fishes* 42(2): 143–150.

Sadovy Y, Kulbicki M, Labrosse P, Letourneur Y, Lokani P, Donaldson TJ. 2003. The humphead wrasse, *Cheilinus undulatus*: synopsis of a threatened and poorly known giant coral reef fish. *Reviews in Fish Biology and Fisheries* 13: 327–364.

Sadovy de Mitcheson Y, Liu M. **2008**. Functional hermaphroditism in teleosts. *Fish and Fisheries* 9: 1–43. https://doi.org/10.1111/j.1467-2979.2007.00266.x

Saeed B, Ivantsoff W, Aarn A. **2006**. Descriptive anatomy of *Iso rhothophilus* (Ogilby), with a phylogenetic analysis of *Iso* and a redefinition of Isonidae (Atheriniformes). *aqua, Journal of Ichthyology and Aquatic Biology* 11(1): 25–43.

Saeed S. 2000. Preliminary survey on the status of shark stocks in Socotra (123–128). In: Hariri K, Krupp F (eds) Conservation and sustainable use of biodiversity of Socotra Archipelago. Marine habitat, biodiversity and fisheries surveys and management. Report of Phase II. 2 Vols. Senckenberg Research Institute, Frankfurt a.M., Germany.

Saemundsson B. 1922. Zoologiske Meddelelser fra Island. XIV. Fiske, ny for Island, of supplerende Oplysninger om andre, tidligere kendte. Videnskabelige Meddelelser fra Dansk Naturhistorisk Forening (Kjøbenhavn) 74: 159–201, Pls. 3–5.

Sætersdal G, Bianchi G, Stromme T. 1999. The Dr. Fridtjof Nansen Programme 1975–1993. Investigations of fishery resources in developing countries. History of the programme and review of results. FAO Fisheries Technical Paper No. 391. Rome: FAO. 434 pp. Sainsbury KJ, Kailola PJ, Leyland GG. **1984**. *Continental shelf fishes of northern and north-western Australia: an illustrated guide*. Canberra, Australia: Clouston & Hall and Peter Pownwall Fisheries Information Service. 375 pp.

Sakai K, Nakabo T. **1995**. Taxonomic review of the Indo-Pacific kyphosid fish, *Kyphosus vaigiensis* (Quoy and Gaimard). *Japanese Journal of Ichthyology* 42(1): 61–70.

Sakai K, Nakabo T. **2004**. Two new species of *Kyphosus* (Kyphosidae) and a taxonomic review of *Kyphosus bigibbus* Lacepède from the Indo-Pacific. *Ichthyological Research* 51(1): 20–32.

Sakai K, Nakabo T. 2006. Taxonomic reviews of two Indo-Pacific sea chubs, *Kyphosus cinerascens* (Forsskål, 1775) and *Kyphosus sydneyanus* (Günther, 1886). *Ichthyological Research* 53(4): 337–356.

Sakai K, Nakabo T. 2014. Taxonomic review of *Kyphosus* (Pisces: Kyphosidae) in the Atlantic and eastern Pacific oceans. *Ichthyological Research* 61(3): 265–292. http:// dx.doi.org/10.1007/s10228-014-0395-x

Sakaizumi M. 1985. Electrophoretic comparison of proteins in five species of *Oryzias* (Pisces: Oryziatidae). *Copeia* 1985(2): 520–522.

Sakamoto K. **1984**. Interrelationships of the family Pleuronectidae (Pisces: Pleuronectiformes). *Memoirs of the Faculty of Fisheries Hokkaido University* 31(1–2): 95–215.

Salazar-Hermoso F, Ochoa-López E, Villavicencio-Garayzar CJ. **1999**. Stranding records of the oarfish in and around Bahia de La Paz, Mexico. *California Fish and Game* 85(2): 70–74.

Saldanha L, Quéro J-C. **1994**. *Channomuraena bauchotae* (Anguilliformes: Muraenidae), nouvelle espèce de l'Ile de la Réunion, Océan Indien. *Cybium* 18(3): 307–313.

Salm RV. **1983**. Coral reefs of the Western Indian Ocean: a threatened heritage. *Ambio* 12: 349–354.

Salm RV, Mee JKL. **1989**. *Chaetodon dialeucos* sp. nov. A new species of shallow water butterflyfish from the northwest Indian Ocean. *Freshwater and Marine Aquarium* 12(3): 8, 9, 11, 131.

Samuel CT. **1961**. On the ambulatory mechanism in *Choridactylus multibarbis* Richardson. *Bulletin of the Central Research Institute* (University of Kerala) 8: 79–84.

Samuel CT. **1963**. Bottom fishes collected by R.V. *Conch* off the Kerala coast. *Bulletin of the Department of Marine Biology Oceanography* (University of Kerala) 1: 97–121.

Sanches JG. **1963**. Contribuição para o estudo dos peixes teleósteos da ilha de Inhaca (Moçambique). *Memórias da Junta de Investigações do ultramar, Lisboa* (Series 2) 44: 1–207.

Sanders M, Morgan G. **1989**. Review of the fisheries resources of the Red Sea and Gulf of Aden. *FAO Fisheries Technical Paper* No. 304. Rome: FAO. 138 pp. Sandknop EM, Watson W. 1996. Howellidae: pelagic basslets (pp. 1072–1079). In: Moser HG (ed) *The early stages of fishes in the California Current Region. California Cooperative Oceanic Fisheries Investigations (CalCOFI) Atlas No. 33.* La Jolla, California: Scripps Institution of Oceanography. 1517 pp.

Sansom IJ, Andreev PS. 2019. The Ordovician enigma: fish, first appearances and phylogenetic controversies (pp. 59–70). In: Johanson Z, Underwood C, Richter M (eds) *Evolution and development of fishes*. Cambridge, UK: Cambridge University Press. 274 pp. http://dx.doi. org/10.1017/9781316832172.004

Santini F. **2006**. A new species of Triacanthodidae (Tetraodontiformes, Acanthomorpha) from the central Pacific. *Cybium* 30(3): 195–198.

Santini F, Soresen L, Marcroft T, Dornburg A, Alfaro ME. 2013. A multilocus molecular phylogeny of boxfishes (Aracanidae, Ostraciidae; Tetraodontiformes). *Molecular Phylogenetics and Evolution* 66(1): 153–160. http://dx.doi. org/10.1016/j.ympev.2012.09.022

Santini F, Tyler JC. 2003. A phylogeny of the families of fossil and extant tetraodontiform fishes (Acanthomorpha, Tetraodontiformes), Upper Cretaceous to Recent. *Zoological Journal of the Linnean Society* 139(4): 565–617.

Saramma D. 1963. Bottom fishes collected by the research vessel *Conch*, off the Kerala Coast during 1958–'63: Heterosomata. *Bulletin of the Department of Marine Biology and Oceanography, University of Kerala* 1: 57–80.

Sarang JD, Katkar BN, Deshmukh VD. 2011. Occurrence of dusky sweeper *Pempheris adusta* Bleeker, 1877 in Ratnagiri waters, Maharashtra. *Marine Fisheries Information Service* 208: 22–24.

Saravanan R, Vijayanand P, Arumugam M, Rajagopal S, Thangavel B. **2014**. First record of Titan cardinal fish, *Holapogon maximus* (Apogonidae) along the south east coast of Tamilnadu, India. *Indian Journal of Geo-Marine Sciences* 43(2): 229–232. http://nopr.niscair.res.in/ handle/123456789/27259

Sardou J. **1974**. Contribution à la connaissance de la faune ichthyologique Malgache: découverte de poissons de la famille des Schindleriidae dans le canal de Mozambique, à Nosy-Bé et étude d'une collection de *Schindleria*. *Cahiers ORSTOM* (Série Océanographie) 12(1): 3–15.

Sasaki D, Kimura S. **2011**. First records of a silverside (Atheriniformes: Atherinidae), *Hypoatherina temminckii*, from Japan. *Japanese Journal of Ichthyology* 58(1): 87–91. https://doi.org/10.11369/jji.58.87

Sasaki D, Kimura S. **2012**. Descriptions of two new silversides, *Hypoatherina golanii* and *Hypoatherina lunata*, from the Indo-West Pacific (Atheriniformes: Atherinidae). *Ichthyological Research* 60(2) [2013]: 103–111. http://dx.doi. org/10.1007/s10228-012-0318-7 Sasaki D, Kimura S. **2014**. Taxonomic review of the genus *Hypoatherina* Schultz 1948 (Atheriniformes: Atherinidae). *Ichthyological Research* 61(3): 207–241. http://dx.doi. org/10.1007/s10228-014-0391-1

Sasaki K. **1994**. *Argyrosomus beccus*, a new sciaenid from South Africa. *Japanese Journal of Ichthyology* 41(1): 35–38.

Sasaki K. **1995**. A review of the Indo-West Pacific sciaenid genus *Panna* (Teleostei, Perciformes). *Japanese Journal of Ichthyology* 42(1): 27–37.

Sasaki K. **1995**. Two new species of *Atrobucca* (Sciaenidae) from the Bay of Bengal. *Japanese Journal of Ichthyology* 42(3–4): 269–275.

Sasaki K. 1996. Sciaenid fishes of the Indian Ocean (Teleostei, Perciformes). *Memoirs of the Faculty of Science, Kochi* University (Series D) (Biology) 16/17: 83–95.

Sasaki K. **1997**. Resurrection of two East African species of *Johnius* (Perciformes, Sciaenidae). *Ichthyological Research* 44(3): 311–315.

Sasaki K. 2001. Family Sciaenidae. Croakers (drums) (pp. 3117–3174). In: Carpenter KE, Niem VH (eds) FAO species identification guide for fishery purposes. The living marine resources of the western central Pacific. Vol. 5. Bony fishes part 3 (pp. 2791–3380). Rome: FAO.

Sasaki K, Kailola PJ. **1988**. Three new Indo-Australian species of the sciaenid genus *Atrobucca*, with a reevaluation of generic limit. *Japanese Journal of Ichthyology* 35(3): 261–277.

Sato T. **1978**. A synopsis of the sparoid fish genus *Lethrinus*, with the description of a new species. *Bulletin of the University Museum, University of Tokyo* 15: i–v + 1–70, Pls. 1–12.

Sato T. 1984. Lethrinidae (including part of Pentapodidae of authors). Emperors, pig-face breams, large-eye breams. In: Fischer W, Bianchi G (eds) FAO species identification sheets for fishery purposes. Western Indian Ocean (Fishing Area 51). Vol. 2. Rome: FAO. [unpaginated]

Sato T, Gomon MF, Nakabo T. 2010. Two new Australian species of the *Paraulopus nigripinnis* complex (Aulopiformes: Paraulopidae). *Ichthyological Research* 57(3): 254–262.

http://dx.doi.org/10.1007/s10228-010-0162-6

Sato T, Nakabo T. **2002**. Paraulopidae and *Paraulopus*, a new family and genus of aulopiform fishes with revised relationships within the order. *Ichthyological Research* 49(1): 25–46.

Sato T, Nakabo T. **2003**. A revision of the *Paraulopus oblongus* group (Aulopiformes: Paraulopidae) with description of a new species. *Ichthyological Research* 50(2): 164–177.

Saunders DC. **1960**. A survey of the blood parasites in the fish of the Red Sea. *Transactions of the American Microscopical Society* 79(3): 239–252.

Sauvage H-E. **1873**. Notice sur quelques poissons d'espèces nouvelles ou peu connues provenant des mers de l'Inde et de la Chine. *Nouvelles Archives du Muséum d'Histoire Naturell, Paris* 9(pt 2): 49–62.

Sauvage H-E. **1875**. Sur la faune ichthyologique de l'île Saint-Paul. *Comptes rendus hebdomadaires des séances de l'Académie des Sciences* 81: 987–988.

Sauvage H-E. 1878. Description de poissons nouveaux ou imparfaitement connus de la collection du Muséum d'Histoire Naturelle: famille des scorpenidées, des platycéphalidées et des triglidées. Nouvelle Archives du Muséum National d'Histoire Naturelle, Paris (Série 2) 1: 109–158.

Sauvage H-E. **1879**. Mémoire sur la faune ichthyologique de l'île Saint-Paul. *Archives de Zoologie Expérimentale et Générale* 8: 1–46.

Sauvage H-E. 1879. Description de quelques poissons d'espèces nouvelles de la collection du Muséum d'Histoire Naturelle. *Bulletin de la Société Philomathique de Paris* (Série 7) 3: 204–209.

Sauvage H-E. **1880**. Description des Gobioïdes nouveaux ou peu connus de la collection du Muséum d'Histoire Naturelle. *Bulletin de la Société Philomathique de Paris* (Série 7) 4: 40–58.

Sauvage H-E. **1881**. Recherches sur la faune ichthyologique de l'Asie et description d'espèces nouvelles de l'Indo-Chine. *Nouvelles Archives du Muséum d'Histoire Naturelle, Paris* (Série 2) 4: 123–194, Pls 5–8.

Sauvage H-E. **1882**. Description de quelques poissons de la collection du Muséum d'histoire naturelle. *Bulletin de la Société Philomathique de Paris* (Série 7) 6: 168–176.

Sauvage H-E. **1883**. Descriptions de quelques poissons de la collection du Muséum d'Histoire Naturelle. *Bulletin de la Société Philomathique de Paris* (Série 7) 7: 156–161.

Sauvage H-E. 1891. *Histoire physique, naturelle et politique de Madagascar. Vol. XVI. Histoire naturelle des poisons* (2 volumes; text and atlas). Paris: Alfred Grandidier. 543 pp.

Sawai E, Yamanoue Y, Nyegaard M, Sakai Y. **2018**. Redescription of the bump-head sunfish *Mola alexandrini* (Ranzani 1839), senior synonym of *Mola ramsayi* (Giglioli 1883), with designation of a neotype for *Mola mola* (Linnaeus 1758) (Tetraodontiformes: Molidae). *Ichthyological Research* 65(1): 142–160. http://dx.doi. org/10.1007/s10228-017-0603-6

Sazonov YI, Ivanov AN. **1980**. Slickheads (Alepocephalidae and Leptochilichthyidae) from the thalassobathyal zone of the Indian Ocean. *Trudy Institute of Oceanography* (USSR) 110: 7–104. [In Russian, English summary]

Schaaf-Da Silva JA, Ebert DA. **2008**. A revision of the western North Pacific swellsharks, genus *Cephaloscyllium* Gill 1862 (Chondrichthyes: Carcharhiniformes: Scyliorhinidae), including descriptions of two new species. *Zootaxa* 1872: 1–28. http://dx.doi.org/10.11646/zootaxa.1872.1.1

- Schaaf-Da Silva JA, Ebert DA. 2008. A re-description of the eastern Pacific swellshark, *Cephaloscyllium ventriosum* (Garman 1880) (Chondrichthyes: Carcharhiniformes: Scyliorhinidae), with comments on the status of *C. uter* (Jordan & Gilbert 1896). *Zootaxa* 1872: 59–68. https://doi.org/10.11646/zootaxa.1872.1.5
- Schils T. 2002. Macroalgal assemblages of the Socotra Archipelago (pp. 383–389). In: Apel M, Hariri K, Krupp F (eds) Conservation and sustainable use of biodiversity of Socotra Archipelago. Marine habitat, biodiversity and fisheries surveys and management. Final Report of Phase III. Senckenberg Research Institute, Frankfurt a.M., Germany.
- Schils T. **2006**. The tripartite biogeographical index: a new tool for quantifying spatio-temporal differences in distribution patterns. *Journal of Biogeography* 33(4): 560–572.
- Schils T, Coppejans E. 2003. Phytogeography of upwelling areas in the Arabian Sea. *Journal of Biogeography* 30(9): 1339–1356.
- Schils T, Coppejans E. 2003. Spatial variation in subtidal plant communities around the Socotra Archipelago and their biogeographic affinities within the Indian Ocean. *Marine Ecology Progress Series* 251: 103–114.
- Schindler O. 1930. Ein neuer Hemirhamphus aus dem pazifischen Ozean. Anzeiger der Akademie der Wissenschaften in Wien 67(9): 79–80.
- Schindler O. 1931. Ein neuer Hemirhamphus aus dem Pazifischen Ozean. Anzeiger der Akademie der Wissenschaften in Wien, Mathematisch-Naturwissenschaftliche Klasse 68(1): 2–3.
- Schindler O. 1932. Sexually mature larval Hemirhamphidae from the Hawaiian Islands. *Bulletin of the Bernice Pauahi Bishop Museum* 97: 1–28, Pls. 1–10.
- Schinz HR, Cuvier G. **1822**. *Das Thierreich eingetheilt nach dem Bau der Thiere als Grundlage ihrer Naturgeschichte und der vergleichenden Anatomie von dem Herrn Ritter von Cuvier*. Vol. 2. Stuttgart and Tübingen: J.S. Cotta.
- Schlegel H. **1852**. Beschrijving eener nieuwe soort van visschen *Peristedion laticeps*. *Bijdragen tot de Dierkunde* 1: 43–44.
- Schlegel H, Müller S. 1840–1844. Overzigt den uit de Sunda en Moluksche zeeën bekende visschen, van de geslachten Amphiprion, Premnas, Pomacentrus, Glyphisodon, Dascyllus en Heliases. Verhandelingen over de natuurlijke geschiedenis der Nederlandsche overzeesche bezittingen / door de Leden der Natuurkundige Commissie in Indië en andere schrijvers; uitgegeven ... door C. J. Temminck Temminck. Zoologie 2: 17–26.
- Schleyer MH. 2000. Chapter 3. South African coral communities (pp. 83–106). In: McClanahan TR, Sheppard CRC, Obura DO (eds) Coral reefs of the Indian Ocean: Their ecology and conservation. Oxford: Oxford University Press. xxiii + 525 pp.

Schmeltz JDE. **1879**. Museum Godeffroy. Catalog VII. Wirbelthiere (Animalia vertebrata) und Nachträge zu Catalog V & VI aus den übrigen Thierklassen. Hamburg (L. Friederichsen & Co.). viii + 99 pp.

Schmid H, Randall JE. **1997**. First record of the tripletail, *Lobotes surinamensis* (Pisces: Lobotidae) from the Red Sea. *Fauna of Saudi Arabia* 16: 353–355.

Schmidt J. **1921**. Contributions to the knowledge of the young of the sunfishes *Mola* and *Ranzania*. *Meddeleser Fra Kommissionen For Havundersogelser* 4(6): 1–13.

- Schmidt J. **1931**. Oceanographic expedition of the *Dana*, 1928–30. *Nature* 127: 444–446.
- Schmidt PJ. 1931. Fishes of Japan, collected in 1901. Transactions of the Pacific Committee Academy of Sciences of the USSR. 2: 1–176.
- Schnakenbeck W. **1931**. Über einige Meeresfische aus Südwestafrika. *Mitteilungen aus dem Zoologischen Museum in Hamburg* 44: 23–45.

Schneider JG. 1801. In: Bloch ME, Schneider JG.

Schneider JG, Forster JR. 1801. In: Bloch ME, Schneider JG.

Schneider M, Janke M. 2013. First record of gorgeous swallowtail *Meganthias natalensis*, an anthiine fish (Acanthopterygii: Serranidae: Anthiinae) from Kenyan waters. *Marine Biodiversity Records* 6(e22): 1–4. http://dx.doi.org/10.1017/S1755267212001327

Schneider MF, Buramuge VA, Aliasse L, Serfontein F.
2005. Checklist of vertebrates of Mozambique. Maputo, Moçambique: Universidade Eduardo Mondlane. 227 pp.

- Scholte P, De Geest P. 2010. The climate of Socotra Island (Yemen): a first-time assessment of the timing of the monsoon wind reversal and its influence on precipitation and vegetation patterns. *Journal of Arid Environments* 74(11): 1507–1515. http://dx.doi.org/10.1016/j. jarideny.2010.05.017
- Schott FA, Fischer J. 2000. Winter monsoon circulation of the northern Arabian Sea and Somali Current. *Journal of Geophysical Research* 105(C3): 6359–6376.
- Schott F, Swallow JC, Fieux M. **1990**. The Somali current at the equator: annual cycle of currents and transports in the upper 1000 m and connection to neighbouring latitudes. *Deep Sea Research, Part A: Oceanographic Research Papers* 37(12): 1825–1848.

Schrank F von P. 1798. Fauna Boica. Durchgedachte Geschichte der in Baiern einheimischen und zahmen Thiere. Nurnberg. [unpaginated] https://doi.org/10.5962/bhl.title.44923

- Schrey WC. **1978**. Fast schon eine "Rarität" Die Gattung *Oryzias. Die Aquarien- und Terrarien-Zeitschrift* 10: 335–338.
- Schuhmacher H, Krupp F, Randall JE. **1989**. *Pseudanthias heemstrai*, a new species of anthiine fish (Perciformes: Serranidae) from the Gulf of Aqaba, Red Sea. *Fauna of Saudi Arabia* 10: 338–346.

Schultz ET. **1986**. *Pterois volitans* and *Pterois miles*: two valid species. *Copeia* 1986(3): 686–690.

Schultz LP. **1940**. Two new genera and three new species of cheilodipterid fishes, with notes on other genera of the family. *Proceedings of the United States National Museum* 88(3085): 403–423.

Schultz LP. 1943. Fishes of the Phoenix and Samoan islands collected in 1939 during the expedition of the U.S.S. *Bushnell. Bulletin of the United States National Museum* 180: i–x + 1–316.

Schultz LP. **1946**. A revision of the genera of mullets, fishes of the family Mugilidae, with descriptions of three new genera. *Proceedings of the United States National Museum* 96(3204): 377–395.

Schultz LP. 1948. A. revision of six subfamilies of atherine fishes, with descriptions of new genera and species. *Proceedings of the United States National Museum* 98(3220): 1–48.

Schultz LP. **1950**. Correction for "A revision of six subfamilies of atherine fishes, with descriptions of new genera and species." *Copeia* 1950(2): 150.

Schultz LP. **1950**. Three new species of fishes of the genus *Cirrhitus* (family Cirrhitidae) from the Indo-Pacific. *Proceedings of the United States National Museum* 100(3270): 547–552.

Schultz LP. 1953. Families Synodontidae: lizardfishes (pp. 30–42); Enchelidae: worm eels (pp. 60–83); Muraenidae: moray eels (pp. 98–159); Atherinidae: silversides (pp. 287–310); Mugilidae: mullets (pp. 310–322); Serranidae: groupers, sea basses (pp. 328–388); Pseudochromidae (pp. 388–411); Lutjanidae: snappers (pp. 521–556). In: Schultz LP, Herald ES, Lachner EA, Welander AD, Woods LP (eds) *Fishes of the Marshall and Marianas Islands. Vol. 1. Families from Asymmetrontidae through Siganidae. Bulletin of the United States National Museum* 202: i–xxxii + 1–685, Pls. 1–74.

Schultz LP. 1953. Review of the Indo-Pacific anemone fishes, genus Amphiprion, with descriptions of two new species. Proceedings of the United States National Museum 103(3323): 187–201.

Schultz LP. **1956**. A new pinecone fish, *Monocentris reedi*, from Chile, a new family record for the eastern Pacific. *Proceedings of the United States National Museum* 106(3365): 237–239.

Schultz LP. 1957. The frogfishes of the family Antennariidae. Proceedings of the United States National Museum 107(3383): 47–105.

Schultz LP. **1958**. Review of the parrotfishes, family Scaridae. *Bulletin of the United States National Museum* 214: i–v + 1–143.

Schultz LP. **1960**. Families Labridae (pp. 121–238); Trichonotidae (pp. 273–281); Clinidae (pp. 281–300); Brolutidae (pp. 382–389); Callionymidae (pp. 397–410). In: Schultz LP, Chapman WM, Lachner EA, Woods LP (eds) Fishes of the Marshall and Marianas Islands. Vol. 2. Families from Mullidae through Stromateidae. Bulletin of the United States National Museum No. 202: i–ix + 1–438, Pls. 75–123.

Schultz LP. **1964**. Three new species of frogfishes from the Indian and Pacific oceans, with notes on other species (family Antennariidae). *Proceedings of the United States National Museum* 116(3500): 171–182.

Schultz LP. 1966. Family Platycephalidae: flatheads (pp. 45–51). In: Schultz LP, Woods LP, Lachner EA (eds) *Fishes of the Marshall and Marianas Islands. Vol. 3. Families from Kraemeriidae through Antennariidae. Bulletin of the United States National Museum* No. 202: i–vii + 1–176, Pls. 124–148.

Schultz LP. 1968. Four new fishes of the genus *Parapercis* with notes on species from the Indo-Pacific area (family Mugiloididae). *Proceedings of the United States National Museum* 124(3636): 1–16.

Schultz LP. **1969**. The taxonomic status of the controversial genera and species of parrotfishes with a descriptive list (family Scaridae). *Smithsonian Contributions to Zoology* 17: 1–49.

Schultz LP, Chapman WM. 1960. Subfamily Salariinae (pp. 302–372). In: Schultz LP, Chapman WM, Lachner EA, Woods LP (eds) *Fishes of the Marshall and Marianas Islands. Vol. 2. Families from Mullidae through Stromateidae. Bulletin of the United States National Museum* 202: i–ix + 1–438, Pls. 75–123.

Schultz LP, Chapman WM, Lachner EA, Woods LP (eds). 1960. Fishes of the Marshall and Marianas islands. Vol. 2. Families from Mullidae through Stromateidae. Bulletin of the United States National Museum No. 202: i–ix + 1–438, Pls. 75–123.

Schultz LP, Herald ES, Lachner EA, Welander AD, Woods LP (eds). 1953. Fishes of the Marshall and Marianas Islands.
Vol. 1. Families from Asymmetrontidae through Siganidae.
Bulletin of the United States National Museum No. 202: i-xxxii + 1–685, Pls. 1–74.

Schultz LP, Marshall NB. **1954**. A review of the labrid fish genus *Wetmorella* with descriptions of new forms from the tropical Indo-Pacific. *Proceedings of the United States National Museum* 103(3327): 439–447.

Schultz LP, Woods LP. **1948**. A new name for *Synchiropus altivelis* Regan, with a key to the genera of the fish family Callionymidae. *Journal of the Washington Academy of Sciences* 38(12): 419–420.

Schultz LP, Woods LP. **1949**. Keys to the genera of echelid eels and the species of *Muraenichthys* of the Pacific, with two new species. *Journal of the Washington Academy of Sciences* 39(5): 169–174. Schultz LP, Woods LP, Lachner EA (eds). 1966. Fishes of the Marshall and Marianas Islands. Vol. 3. Families from Kraemeriidae through Antennariidae. Bulletin of the United States National Museum No. 202: i–vii + 1–176, Pls. 124–148.

Schultze HP, Arratia G. 2013. The caudal skeleton of basal teleosts, its conventions, and some of its major evolutionary novelties in a temporal dimension (pp. 187–246). In: Arratia G, Schultze H-P, Wilson MVH (eds) *Mesozoic Fishes 4 – Homology and phylogeny. Proceedings of the international meeting, Miraflores de la Sierra, 2005*. Munich: Verlag Dr. Friedrich Pfeil. 502 pp.

Schultze L. 1907. Die Fischerei an der Westküste Süd-Afrikas: Bericht über Untersuchungen an der Deutsch-S-W-Afrikanischen Küste und am Kap der Guten Hoffnung; der Kolonialabteilung des Auswärtigen amtes, erstattet. Berlin: O. Salle. 57 pp.

Schumann EH. **1987**. The coastal ocean off the east coast of South Africa. *Transactions of the Royal Society of South Africa* **46**: 215–229.

Schwartz FJ. 1990. Mass migratory congregation and movements of several species of cownose rays, genus *Rhinoptera*: a world-wide review. *Journal of the Elisha Mitchell Scientific Society* 106(1): 10–13.

Schwarzhans W. **1994**. Sexual and ontogenetic dimorphism in otoliths of the family Ophidiidae. *Cybium* 18(1): 71–98.

Schwarzhans W, Møller PR. **2007**. Review of the Dinematichthyini (Teleostei: Bythitidae) of the Indo-West Pacific. Part III. *Beaglichthys, Brosmolus, Monothrix* and eight new genera with description of 20 new species. *The Beagle, Records of the Museums and Art Galleries of the Northern Territory* 23: 29–110.

Schwarzhans W, Møller PR. **2011**. New Dinematichthyini (Teleostei: Bythitidae) from the Indo-west Pacific, with the description of a new genus and five new species. *The Beagle, Records of the Museums and Art Galleries of the Northern Territory* 27: 161–177.

Schwarzhans W, Møller PR, Nielsen JG. 2005. Review of the Dinematichthyini (Teleostei: Bythitidae) of the Indo-West Pacific. Part I. *Diancistrus* and two new genera with 26 new species. *The Beagle, Records of the Museums and Art Galleries of the Northern Territory* 21: 73–163.

Schwarzhans W, Nielsen JG. 2011. Revision of the genus Microbrotula (Teleostei: Bythitidae), with description of two new species and a related new genus. The Beagle, Records of the Museums and Art Galleries of the Northern Territory 27: 147–160.

Schwarzhans W, Prokofiev AM. 2017. Reappraisal of Synagrops Günther, 1887 with rehabilitation and revision of Parascombrops Alcock, 1889 including description of seven new species and two new genera (Perciformes: Acropomatidae). Zootaxa 4260(1): 1–74. http://dx.doi. org/10.11646/zootaxa.4260.1.1 Schwimmer DR, Stewart JD, Williams GD. **1994**. Giant fossil coelacanths of the Late Cretaceous in the eastern United States. *Geology* 22(6): 503–506.

Scoles DR, Collette BB, Graves JE. **1998**. Global phylogeography of mackerels of the genus *Scomber*. *Fishery Bulletin* 96(4): 823–842.

Scoles DR, Graves JE. 1993. Genetic analysis of the population structure of yellowfin tuna, *Thunnus albacares*, from the Pacific Ocean. *Fishery Bulletin* 91(4): 690–698.

Scopoli JA. 1777. Introductio ad historiam naturalem, sistens genera lapidum, plantarum et animalium hactenus detecta, caracteribus essentialibus donata, in tribus divisa, subinde ad leges naturae. Prague. x + 506 pp.

Scott EOG. **1934**. Observations on some Tasmanian fishes, with descriptions of new species. *Papers and Proceedings of the Royal Society of Tasmania* [for 1933]: 31–53.

Scott TD, Glover CJM, Southcott RV. **1974**. *The marine and freshwater fishes of South Australia*. South Australia Government. 392 pp.

Scott WB, Scott MG. **1988**. Atlantic fishes of Canada. *Canadian Bulletin of Fisheries and Aquatic Sciences* 219: xxx +730 pp.

Seale A. **1901**. Report of a mission to Guam. Part II – Fishes. Occasional Papers of the Bernice Pauahi Bishop Museum of Polynesian Ethnology and Natural History 1(3): 61–128.

Seale A. **1901**. New Hawaiian fishes. Occasional Papers of the Bernice Pauahi Bishop Museum of Polynesian Ethnology and Natural History 1(4): 1–15.

Seale A. **1906**. Fishes of the South Pacific. Occasional Papers of the Bernice Pauahi Bishop Museum of Polynesian Ethnology and Natural History 4(1): 1–89.

Seale A. **1910**. New species of Philippine fishes. *Philippine Journal of Science* (Section A) 4(6): 491–543.

Seale A. **1917**. New species of apodal fishes. *Bulletin of the Museum of Comparative Zoology* 61(4): 79–94.

Seale A. **1935**. The Templeton Crocker expedition to western Polynesian and Melanesian islands. *Proceedings of the California Academy of Sciences* (Series 4) 21(27): 337–378.

Seba A. **1758**. *Locupletissimi rerum naturalium thesauri accurate descriptio et iconibus artificiosissimus expressio per universam physices historiam*. Vol. 3. Amsterdam: H.K. Arksteus. [unpaginated]

Seegers L. **1997**. *Killifishes of the world: Old world killis I: Ricefishes, lampeyes,* Aphyosemion (Aqualog Reference Book). Mörfelden-Walldorf: Verlag A.C.S. 160 pp.

Seki MP, Mundy BC. **1991**. Some notes on the early life stages of the Pacific pomfret, *Brama japonica*, and other Bramidae from the central North Pacific Ocean. *Japanese Journal of Ichthyology* 38(1): 63–68.

Sen TK. 1986. Description of a new species, *Pisodonophis assamensis*, a new eel from lower Assam with a key to the Indian Ophichthidae (family: Ophichthidae / genus: *Pisodonophis*). *Bulletin of the Zoological Survey of India* 7(2–3) [for 1985]: 241–244.

Seno M. 2000. Ecsenius minutus. I.O.P. Diving News 11(11): 4, fig. 2.

Senou H. **1997**. Redescription of a mullet, *Chelon melinopterus* (Perciformes: Mugilidae). *Bulletin of the Kanagawa Prefectural Museum, Natural Science* 26: 51–55.

Senou H, Mishiku A, Ito M, Motomura H. 2013. First records of a rare unicornfish, *Naso mcdadei* (Perciformes: Acanthuridae), from Japan, with notes on biogeographical implications for the species. *Bulletin of the Kanagawa Prefectural Museum, Natural Science* 2013(42): 91–96.

Senou H, Randall JE, Okiyama M. 1995. Chelon persicus, a new species of mullet (Perciformes: Mugilidae) from the Persian Gulf. Bulletin of the Kanagawa Prefectural Museum, Natural Science 25: 71–78.

Senou H, Yunokawa K. 1995. New record of an anthiine fish, *Plectranthias pelicieri* (Perciformes: Serranidae) from the western Pacific. *I.O.P. Diving News* 6(9): 4–6. [In Japanese, English abstract]

Senta T. **1973**. A new sparoid fish, *Gymnocranius elongatus*, from the southern South China Sea. *Japanese Journal of Ichthyology* 20(3): 135–144.

Senta T. **1975**. Redescription of trichiurid fish *Tentoriceps cristatus* and its occurrence in the South China Sea and the Straits of Malacca. *Japanese Journal of Ichthyology* 21(4): 175–182.

Senta T, Tan S-M. **1975**. On *Pristipomoides multidens* and *P. typus* (family Lutjanidae). *Japanese Journal of Ichthyology* 22(2): 68–76.

Séret B. 1986. Deep water skates of Madagascar. Part I. Anacanthobatidae (Pisces, Chondrichthyes, Batoidea), second record of the skate *Anacanthobatis ori* (Wallace, 1967) from off Madagascar. *Cybium* 10(4): 307–326.

Séret B. 1987. Halaelurus clevai, sp. n., a new species of catshark (Scyliorhinidae) from off Madagascar, with remarks on the taxonomic status of the genera Halaelurus Gill and Galeus Rafinesque. J.L.B. Smith Institute of Ichthyology Special Publication 44: 1–28.

Séret B. 1989. Deep water skates of Madagascar. Part 2. Rajidae. Gurgesiella (Fenestraja) maceachrani sp. n. Cybium 13(1): 55–64.

Séret B. 1989. Deep water skates of Madagascar. Part 3. Rajidae (Pisces, Chondrichthyes, Batoidea). *Raja (Dipturus) crosnieri* sp. n. *Cybium* 13(2): 115–130.

Séret B. 2003. Chondrichthyes: 1. Carcharhinidae. 2. Pristidae.
3. Dasyatidae. (Vol. 1: 74–96). In: Paugy D, Lévêque C, Teugels GG (eds) The fresh and brackish water fishes of West Africa. [Faune des poissons d'eau douce et saumâtres de l'Afrique de l'Ouest.] Vols. 1 & 2. Paris: Muséum National d'Histoire Naturelle. 1272 pp.

Séret B. 2016. Rhinobatidae. Guitarfishes (pp. 1357–1364). In: Carpenter KE, De Angelis N (eds) FAO species identification guide for fishery purposes. The living marine resources of *the eastern central Atlantic*. Vol. 2. Bivalves, gastropods, hagfishes, sharks, batoid fishes and chimaeras. FAO: Rome. pp. 665–1509.

Séret B, Last PR. 2007. Four new species of deep-water catsharks of the genus *Parmaturus* (Carcharhiniformes: Scyliorhinidae) from New Caledonia, Indonesia and Australia. *Zootaxa* 1657: 23–39.

Séret B, McEachran JD. 1986. Catalogue critique des types de poissons du Muséum national d'Histoire Naturelle. (Suite). Poissons batoïdes (Chrondricththyes, Elasmobranchii, Batoidea). Bulletin du Muséum National d'Histoire Naturelle (Paris) (Série 4, Section A: Zoologie, Biologie et Écologie Animales) 8(4) suppl.: 3–50.

Séret B, Sire J-Y (eds). 1999. Proceedings of the 5<sup>th</sup> Indo-Pacific Fish Conference, Nouméa, New Caledonia, 3–8 November 1997. Paris: Société Française d'Ichtyologie. 866 pp.

Serventy DL. **1948**. *Allothunnus fallai* a new genus and species of tuna from New Zealand. *Records of the Canterbury Museum* 5(3): 131–135.

Serventy DL. 1956. The southern bluefin tuna, *Thunnus thynnus maccoyii* (Castelnau) in Australian waters. *Australian Journal of Marine and Freshwater Research* 7(1): 1–43.

Serventy DL. **1956**. Additional observations on the biology of the northern bluefin tuna, *Kishinoella tonggol* (Bleeker) in Australia. *Australian Journal of Marine and Freshwater Research* 7(1): 44–64.

Seshagiri Rao BV. **1974**. On the little known electric ray *Narke* (*Bengalichthys*) *impennis* from India. *Copeia* 1974(3): 791–792.

Seshagiri Rao BV. **1975**. A new species of clupeid fish, *Ilisha sirishai* from Visakhapatnam, India. *Hydrobiologia* 47(3–4): 463–468.

Setiamarga DHE, Miya M, Yamanoue Y, Mabuchi K, Satoh TP, Inoue JG, Nishida M. **2008**. Interrelationships of Atherinomorpha (medakas, flyingfishes, killifishes, silversides, and their relatives): the first evidence based on whole mitogenome sequences. *Molecular Phylogenetics and Evolution* 49(2): 598–605. http://dx.doi.org/10.1016/j. ympev.2008.08.008

Setna SB, Sarangdhar PN. **1946**. Selachian fauna of the Bombay waters. (A classificatory representation with a key for their identification.) *Proceedings of the National Institute of Sciences of India* 12(5): 243–259.

Setna SB, Sarangdhar PN. **1949**. Studies on the development of some Bombay elasmobranchs. *Records of the Indian Museum* (Calcutta) 47(2): 203–216.

SFSA (The Sea Fishes of Southern Africa), *see* Smith JLB. 1949, 1950, 1953, 1961, 1965.

Shakhovskoy IB, Parin NV. **2010**. A comparative description and distribution of the flying fishes — *Cypselurus poecilopterus*, *C. simus*, and *C. callopterus*, sorted out into the species group of spotwing species of the subgenus *Poecilocypselurus. Journal of Ichthyology* 50(8): 559–579. http://dx.doi.org/10.1134/S0032945210080011

- Shakhovskoy IB, Parin NV. 2013. A review of flying fishes of the subgenus *Hirundichthys* (genus *Hirundichthys*, Exocoetidae). Part 1. Oceanic species: *H. speculiger*, *H. indicus* sp. nova. *Journal of Ichthyology* 53(2): 117–145. https://doi.org/10.1134/S003294521301013X
- Shakhovskoy IB, Parin NV. 2013. A review of flying fishes of the subgenus *Hirundichthys* (genus *Hirundichthys*, Exocoetidae). Part 2. Nerito-oceanic species: *H. oxycephalus*, *H. affinis. Journal of Ichthyology* 53(8): 509–540. http://dx.doi.org/10.1134/S0032945213050093 [Also in Russian in *Voprosy Ikhtiologii* 53(5): 507–539]
- Shakhovskoy IB, Parin NV. 2019. A review of the flying fish genus Cypselurus (Beloniformes: Exocoetidae). Part 1. Revision of the subgenus Zonocypselurus Parin et Bogorodsky, 2011 with descriptions of one new subgenus, four new species and two new subspecies and reinstatement of one species as valid. *Zootaxa* 4589(1): 1–71. http://dx.doi. org/10.11646/zootaxa.4589.1.1
- Shallenberger RJ, Madden WD. **1973**. Luring behavior in the scorpionfish, *Iracundus signifer*. *Behaviour* 47(1/2): 33–47.
- Shao K-T, Chen J-P. **1987**. Fishes of the Family Platycephalidae (Teleostei: Platycephaloidae) of Taiwan with descriptions of two new species. *Bulletin of the Institute of Zoology Academia Sinica (Taipei)* 26(1): 77–94.
- Shao K-T, Ho H-C, Lin P-L, Lee P-F, Lee M-Y, Tsai C-Y, Liao Y-C, Lin Y-C. 2008. A checklist of the fishes of southern Taiwan, Northern South China Sea. *The Raffles Bulletin of Zoology* Supplement 19: 233–271.
- Shaw G. **1791**. Description of the *Stylephorus chordatus*, a new fish. *Transactions of the Linnean Society of London* 1(art.7): 90–92, Pl. 6.
- Shaw G. **1803**. *General zoology or systematic natural history*. Vols. IV(1) and IV(2): Pisces. London: G. Kearsley. i–v + 1–186, Pls. 1–25; i–xiii + 187–632, Pls. 26–92.
- Shaw G. **1804**. *General zoology or systematic natural history*. Vols. V(1) and V(2): Pisces. London: G. Kearsley. i–v + 1–250, pls. 93–132; i–vi + 251–463, Pls. 132–182.
- Shaw G, Nodder FP. **1789–1813**. *The naturalist's miscellany, or coloured figures of natural objects, drawn and described from nature* (24 vols). J. Cooper, London. [unpaginated]
- Shcherbachev YN. 1973. The biology and distribution of the dolphins (Pisces, Coryphaenidae). *Voprosy Ikhtiologii* 13: 219–230. [In Russian, English translation in *Journal of Ichthyology* 13: 182–191]
- Shcherbachev YN. **1978**. Long-nosed Chimaeras (Rhinochimaeridae, Chimaeriformes) from the waters of South Africa. *Trudy Instituta Okeanologii* 3: 7–9. [In Russian]

- Shcherbachev YN. 1980. Preliminary review of deep-sea ophidiids (Ophidiidae, Ophidiiformes) of the Indian Ocean. *Trudy Instituta Okeanologii Imeni P.P. Shirshova* 110: 105–176. [In Russian, English summary]
- Shcherbachev YN. 1981. Preliminary review of the Indian Ocean Chlorophthalmidae (Myctophiformes) (pp. 47–67).In: Parin NV (ed) *Fishes of the open ocean*. Moscow: PP Shirshov Institute of Oceanology. 119 pp. [In Russian]
- Shcherbachev YN. **1987**. Preliminary list of thalassobathyal fishes of the tropical and subtropical waters of the Indian Ocean. *Journal of Ichthyology* 27(2): 37–46.

Shcherbachev YN, Pakhorukov NP. 1982. On the distribution of *Trachyscorpia capensis* (Gilchrist et von Bonde)
Pisces, Scorpaenidae. *Bulletin Moscow Society for Nature Experimentation* 87(4): 68–69.

Shcherbachev YN, Parin NV, Pakhorukov NP, Piotrovsky AS. **1986**. Mesobenthic and mesobenthopelagic fishes from submarine rises in the western Indian Ocean. *Trudy Instituta Okeanologii* 121: 195–214. [In Russian]

- Shelmerdine RL, Cliff G. 2006. Sharks caught in the protective gill nets off KwaZulu-Natal, South Africa. 12. The African angel shark Squatina africana (Regan). African Journal of Marine Science 28(3–4): 581–588.
- Shen S-C. **1969**. Additions to the study on the flatfishes in the adjacent waters of Hong Kong. *Report of the Institute of Fishery Biology of Ministry of Economic Affairs and National Taiwan Unversity, Taipei* 2(3): 19–27.
- Shen S-C. **1971**. A new genus of clinid fishes form the Indo-West-Pacific, with a redescription of *Clinus nematopterus*. *Copeia* 1971(4): 697–707.
- Shen S-C. **1982**. Study of the pleuronectid fishes (family Pleuronectidae) from Taiwan. *Quarterly Journal of the Taiwan Museum* 35: 197–213.
- Shen S-C. **1983**. Study on the bothid fishes (Family Bothidae) from Taiwan. *Journal of Taiwan Museum* 36(1): 1–42.
- Shen S-C. 1993. Mugilidae (pp. 437–441). In: Shen S-C [chief editor] et al. Fishes of Taiwan. Taipei: Department of Zoology, National Taiwan University. 960 pp. [In Chinese]
- Shen S-C. **1997**. A review of the genus *Scolopsis* of nemipterid fishes, with descriptions of three new records from Taiwan. *Zoological Studies* 36(4): 345–352.
- Shen S-C. **1998**. *Ariosoma* Swainson, 1838, a senior synonym of *Poeciloconger* Günther, 1871, with redescriptions of *A. fasicatus. Acta Zoologica Taiwanica* 9(2): 105–110.
- Shen S-C [chief editor], Lee SC, Shao KT, Mok HC, Chen CH, Chen CC, Tzeng CS. **1993**. *Fishes of Taiwan*. Taipei: Department of Zoology, National Taiwan University. xx + 960 pp., Pls. 310. [In Chinese]
- Shen S-C, Lim PC. 1974. Study on the plectognath fishes.B. The family Canthigasteridae. *Bulletin of the Institute of Zoology, Academia Sinica* 13(1): 15–34.

Shen S-C, Lim PC, Ting WH. 1975. Ecological and morphological study on fish-fauna from the waters around Taiwan and its adjacent islands. 8. Study on the plectognath fishes. C. The family Tetraodontidae. Acta Oceanographioa Taiwanica 5: 152–178.

Shen S-C, YangT-H, Lin JJ. **1986**. A review of the blenniid fishes in the waters around Taiwan & its adjacent islands. *Taiwan Museum Special Publications Series* 5: 1–74.

Shepard JW, Meyer KA. 1978. A new species of the labrid fish genus Macropharyngodon from southern Japan. Japanese Journal of Ichthyology 25(3): 159–164.

Sheppard CRC. **1981**. The groove and spur structures of Chagos atolls and their coral zonation. *Estuarine Coastal and Shelf Science* **12**: 549–560.

Sheppard CRC. 2000. Chapter 1. Coral reefs of the Western Indian Ocean: an overview (pp. 3–38). In: McClanahan TR, Sheppard CRC, Obura DO (eds) Coral reefs of the Indian Ocean: Their ecology and conservation. Oxford: Oxford University Press. xxiii + 525 pp.

Sheppard CRC. 2000. Chapter 15. The Chagos Archipelago (pp. 445–470). In: McClanahan TR, Sheppard CRC, Obura DO (eds) Coral reefs of the Indian Ocean: Their ecology and conservation. Oxford: Oxford University Press. xxiii + 525 pp.

Sheppard CRC. **2000**. The Red Sea (pp. 35–45). In: Sheppard CRC (ed) Seas at the Millennium: An environmental evaluation. Vol. 2. Oxford: Pergamon Press. 920 pp.

Sheppard CRC. 2000. The Chagos Archipelago, central Indian Ocean (pp. 221–232). In: Sheppard CRC (ed) Seas at the Millennium: An environmental evaluation. Vol. 2. Oxford: Pergamon Press. 920 pp.

Sheppard CRC, Ateweberhan M, Bowen BW, Carr P, Chen CA, Clubbe C, Craig MT *et al.* 2012. Reefs and islands of the Chagos Archipelago, Indian Ocean: why it is the world's largest no-take marine protected area. *Aquatic Conservation* of Marine and Freshwater Ecosystems 22(2): 232–261. http://dx.doi.org/10.1002/aqc.1248

Sheppard C[RC], Loughland R. **2002**. Coral mortality and recovery in response to increasing temperature in the southern Arabian Gulf. *Aquatic Ecosystem Health and Management* 5(4): 395–402. http://dx.doi. org/10.1080/14634980290002020

Sheppard CRC, Price A, Roberts CM. **1992**. *Marine ecology* of the Arabian region: Patterns and processes in the extreme tropical environments. London: Academic Press. 359 pp.

Sheppard CRC; Salm RV. 1988. Reef and coral communities of Oman, with the description of a new coral species (Order Scleractinia, genus Acanthastrea). Journal of Natural History 22: 263–279.

Sheppard CRC, Sheppard ALS. **1991**. Corals and coral communities of Arabia. *Fauna of Arabia* 12: 3–170.

Sheppard CRC, Wilson SC, Salm RV, Dixon D. 2000. Chapter 9. Reefs and coral communities of the Arabian Gulf and Arabian Sea (pp. 257–293). In: McClanahan TR, Sheppard CRC, Obura, DO (eds) Coral reefs of the Indian Ocean: Their ecology and conservation. Oxford: Oxford University Press. xxiii + 525 pp.

Shibukawa K, Allen G. **2007**. Review of the cheek-spine goby genus *Gladiogobius* (Actinopterygii, Perciformes, Gobiidae), with descriptions of two new species from the Indo-West Pacific. *Bulletin of the National Museum of Nature and Science* (Tokyo) (Series A) 33(4): 193–206.

Shibukawa K, Aonuma Y. 2007. Three new species of the deepdwelling goby genus *Obliquogobius* (Perciformes: Gobiidae: Gobiinae) from Japan, with comments on the limits of the genus. *Bulletin of the National Museum of Nature and Science* (Tokyo) (Series A) Supplement No.1: 137–152.

Shibukawa K, Ida H. **2013**. *Ammodytoides kanazawai*, a new species of sand lance (Perciformes: Ammodytidae) from the Ogasawara Islands, Japan. *Bulletin of the National Museum of Nature and Science* (Tokyo) (Series A) Supplement No. 7: 25–30.

Shibukawa K, Suzuki T. 2007. Two new species of the cheekspine goby genus Asterropteryx (Perciformes: Gobiidae: Gobiinae) from the western Pacific. Bulletin of the National Museum of Nature and Science (Tokyo) (Series A) Supplement No. 1: 109–121.

Shibukawa K, Yoshino T, Allen GR. 2010. Ancistrogobius, a new cheek-spine goby genus from the West Pacific and Red Sea, with descriptions of four new species (Perciformes: Gobiidae: Gobiinae). Bulletin of the National Museum of Nature and Science (Tokyo) (Series A) Supplement No. 4: 67–87.

Shiino SM. **1976**. List of common names of fishes of the world: those prevailing among English-speaking nations. *Science Report of Shima Marineland* 4: 1–262.

Shimada K, Yoshino T. **1984**. A new trichonotid fish from the Yaeyama Islands, Okinawa Prefecture, Japan. *Japanese Journal of Ichthyology* 31(1): 15–19.

Shimizu T, Yamakawa T. **1979**. Review of the squirrelfishes (subfamily Holocentrinae: order Beryciformes) of Japan, with a description of a new species. *Japanese Journal of Ichthyology* 26(2): 109–147.

Shindo S, Yamada U. **1972**. Descriptions of three new species of the lizardfish genus *Saurida*, with a key to its Indo-Pacific species. *Uo* (Japanese Society of Ichthyologists) 11&12: 1–13; 1–14.

Shinohara G, Imamura H. 2005. Anatomical description and phylogenetic classification of the orbicular velvetfishes (Scorpaenoidea: *Caracanthus*). *Ichthyological Research* 52(1): 64–76. Shinohara G, Imamura H. **2007**. Revisiting recent phylogenetic studies of "Scorpaeniformes". *Ichthyological Research* 54(1): 92–99.

Shipp RL. **1974**. The pufferfishes (Tetraodontidae) of the Atlantic Ocean. *Publications of the Gulf Coast Research Laboratory Museum* 41: 1–162.

Shirai S. **1992**. Phylogenetic relationships of the angel sharks, with comments on elasmobranch phylogeny (Chondrichthyes, Squaliformes). *Copeia* 1992(2): 505–518.

Shirai S. **1992**. Squalean phylogeny: A new framework of *"squaloid" sharks and related taxa*. Sapporo: Hokkaido University Press. 151 pp.

Shirai S. **1996**. Phylogenetic interrelationships of neoselachians (Chondrichthyes: Euselachii) (pp. 9–34). In: Stiassny MLJ, Parenti LR, Johnson GD (eds) *Interrelationships of fishes*. San Diego: Academic Press. 496 pp.

Shirai S, Tachikawa H. 1993. Taxonomic resolution of the *Etmopterus pusillus* species group (Elasmobranchii, Etmopteridae) with a description of *E. bigelowi*, n. sp. *Copeia* 1993(2): 483–495.

Shirai Y, Kitazawa H. **1998**. Peculiar feeding behavior of *Asterorhombus intermedius* in an aquarium. *Japanese Journal of Ichthyology* 45(1): 47–50. [In Japanese]

Shomura RS, Williams F (eds). 1974. Proceedings of the International Billfish Symposium, Kailua-Kona, Hawaii, 9–12 August 1972. Part 2. Review and contributed papers. NOAA Technical Report NMFS SSRF-675: 344 pp.

Shomura RS, Williams F (eds). 1975. Proceedings of the International Billfish Symposium, Kailua-Kona, Hawaii, 9–12 August 1972. Part 3. Species synopses. NOAA Technical Report NMFS SSRF-675: 159 pp.

Short G, Claassens L, Smith R, De Brauwer M, Hamilton H, Stat M, Harasti D. **2020**. *Hippocampus nalu*, a new species of pygmy seahorse from South Africa, and the first record of a pygmy seahorse from the Indian Ocean (Teleostei, Syngnathidae). *ZooKeys* 934: 141–156. http://dx.doi. org/10.3897/zookeys.934.50924

Short G, Smith R, Motomura H, Harasti D, Hamilton H. **2018**. *Hippocampus japapigu*, a new species of pygmy seahorse from Japan, with a redescription of *H. pontohi* (Teleostei, Syngnathidae). *ZooKeys* 779: 27–49. http://dx.doi. org/10.3897/zookeys.779.24799

Shpigel M. **1997**. *Fishes of the Red Sea*. Ra'anana, Israel: Red Sea Magazine. 159 pp.

Shpigel M, Fishelson L. 1989. Food habits and prey selection of three species of groupers from the genus *Cephalopholis* (Serranidae: Teleostei). *Environmental Biology of Fishes* 24(1): 67–73.

Shpigel M, Fishelson L. 1991. Experimental removal of piscivorous groupers of the genus *Cephalopholis* (Serranidae) from coral habitats in the Gulf of Aqaba (Red Sea). *Environmental Biology of Fishes* 31(2): 131–138.

Shpigel M, Fishelson L. 1991. Territoriality and associated behaviour in three species of the genus *Cephalopholis* (Pisces: Serranidae) in the Gulf of Aqaba, Red Sea. *Journal* of Fish Biology 38(6): 887–896.

Shu D-G, Luo H-L, Conway Morris S, Zhang X-L, Hu S-X, Chen L, Han J, Zhu M, Li Y, Chen L-Z. **1999**. Lower Cambrian vertebrates from south China. *Nature* 402: 42–46. https://doi.org/10.1038/46965

Shu D-G, Conway Morris S, Han J, Zhang Z-F, Yasui K, Janvier P, Chen L, Zhang X-L, Liu J-N, Li Y, Liu HQ.
2003. Head and backbone of the Early Cambrian vertebrate *Haikouichthys. Nature* 421: 526–529. https://doi. org/10.1038/nature01264

Siddall M, Rohling EJ, Almogi-Labin A, Hemleben C, Meischner D, Schmelzer I, Smeed DA. 2003. Sea-level fluctuations during the last glacial cycle. *Nature* 423: 853–858.

Siddiqui PJ, Amir SA, Masroor R. 2014. The sparid fishes of Pakistan, with new distribution records. *Zootaxa* 3857(1):71–100. https://doi.org/10.11646/zootaxa.3857.1.3

Siebeck UE, Parker AN, Sprenger D, Mäthger LM, Wallis G. 2010. A species of reef fish that uses ultraviolet patterns for covert face recognition. *Current Biology* 20(5) [for 2009]: 407–410. https://doi.org/10.1016/j.cub.2009.12.047

Siegel JA. **1978**. Revision of the dalatiid shark genus *Squaliolus*: anatomy, systematics, ecology. *Copeia* 1978(4): 602–614.

Siegel JA, Pietsch TW, Robison BH, Abe T. **1977**. *Squaliolus sarmenti* and *S. alii*, synonyms of the dwarf deepsea shark, *Squaliolus laticaudus*. *Copeia* 1977(4): 788–791.

Silas EG. **1959**. On the natural distribution of the Indian cyprinodont fish *Horaichthys setnai* Kulkarni. *Journal of the Marine Biological Association of India* 1(2): 256.

Silas EG. 1963. Synopsis of biological data on double-lined mackerel *Grammatorcynus bicarinatus* (Quoy and Gaimard) (Indo-Pacific). *FAO Fisheries Biology Synopsis* 72 (Species Synopsis 29): 811–833.

Silas EG. 1963. Synopsis of biological data on oriental bonito Sarda orientalis (Temminck and Schlegel) 1842 (Indian Ocean). FAO Fisheries Biology Synopsis 73 (Species Synopsis 30): 834–861.

Silas EG. **1963**. Synopsis of biological data on dogtooth tuna *Gymnosarda unicolor* (Ruppell) 1838 (Indo-Pacific). *FAO Fisheries Biology Synopsis* 75 (Species Synopsis 32): 877–899.

Silas EG. 1964. Aspects of the taxonomy and biology of the oriental bonito *Sarda orientalis* (Temminck and Schlegel).
In: *Proceedings of the Symposium on Scombroid Fishes*, Marine Biological Association of India (MBAI), 12–15 January 1962, Mandapam. Part 1: 283–308.

Silas EG. **1964**. *Cybium croockewitii* Bleeker (1850) and *C. koreanum* Kishinouye (1915) considered synonyms of *Scomberomorus guttatus* (Bloch and Schneider) with a redescription and annotated bibliography of *S. guttatus*. In: *Proceedings of the Symposium on Scombroid Fishes*, Marine

Biological Association of India (MBAI), 12–15 January 1962, Mandapam. Part 1: 309–342.

Silas EG, Rajagopalan M. **1974**. Studies on demersal fishes of the deep neritic waters and the continental slope 2 on *Trichiurus auriga* Klunzinger, with notes on its biology. *Journal of the Marine Biological Association of India* 16(1): 253–274.

Silas EG, Selvaraj GSD. 1972. Descriptions of the adult and embryo of the bramble shark *Echinorhinus brucus* (Bonnaterre) obtained from the continental slope of India. *Journal of the Marine Biological Association of India* 14(1): 395–401.

Sirajudheen TK, Bijukumar A. **2011**. First record of reticulated moray, *Muraena retifera* (Anguilliformes: Muraenidae) from the Indian coast. *Marine Biodiversity Records* 4(e90): 1–3. http://dx.doi.org/10.1017/S1755267211000984

Sivasubramaniam K, Ibrahim MA. **1982**. *Common fishes of Qatar. Scientifc Atlas of Qatar Doha, No. 1.* Doha, Qatar: Doha Modern Printing Press. 176 pp.

Sivertsen E. **1945**. Fishes of Tristan da Cunha, with remarks on age and growth based on scale readings. *Results of the Norwegian Scientific Expedition to Tristan da Cunha 1937–1938* 12: 1–44.

Skaramuca B, Kožul V, Tutman P. **2001**. Biological requirements for increasing aquaculture production in the southern Adriatic Sea. *Naše More* 48(5–6): 244–247.

Skelton PH. 1993. A complete guide to the freshwater fishes of southern Africa. Halfway House: Southern Book Publishers. xiii + 388 pp.

Skelton PH. 2001. A complete guide to the freshwater fishes of southern Africa (2<sup>nd</sup> edition). Cape Town: Struik Publishers. xvi + 395 pp.

Skelton PH, Jubb RA, Bruton MN. **1980**. Additions to the checklist and recent changes to the scientific names of south ern African freshwater fishes. *Journal of the Limnological Society of Southern Africa* 6(2): 109–112.

Sloane H. 1707. A voyage to the islands Madera, Barbados, Nieves, S. Christophers and Jamaica, with the natural history of the herbs and trees, four-footed beasts, fishes, birds, insects, reptiles, &c. of the last of those islands. 2 Volumes. Printed by B. M. (i.e. R. Bentley and M. Magnes) for the author; London. 763 pp. + 274 plates.

Sluka RD. **2013**. Coastal marine fish biodiversity along the west coast of India. *Journal of Threatened Taxa* 5(1): 3574–3579. http://dx.doi.org/10.11609/JoTT.o3187.118

Smale MJ. 2002. Occurrence of *Carcharias taurus* in nursery areas of the Eastern and Western Cape, South Africa. *Marine and Freshwater Research* 53(2): 551–556.

Smale MJ, Buxton CD. 1998. Subtidal and intertidal fishes (pp. 135–155). In: Lubke R, De Moor I (eds) *Field guide to the eastern and southern Cape coasts*. Rondebosch: University of Cape Town Press. 559 pp. Smale MJ, Cliff G. 1998. Cephalopods in the diets of four shark species (*Galeocerdo cuvier*, *Sphyrna lewini*, *S. zygaena* and *S. mokarran*) from KwaZulu-Natal, South Africa. South African Journal of Marine Science 20(1): 241–253.

Smale MJ, Compagno LJV, Human BA. **2002**. First megamouth shark from the western Indian Ocean and South Africa. *South African Journal of Science* 98(7/8): 349–350.

Smale MJ, Heemstra PC. **1997**. First record of albinism in the great white shark, *Carcharodon carcharias* (Linnaeus, 1758). *South African Journal of Science* **93**(5): 243–245.

Smale MJ, Sauer WHH, Hanlon RT. **1995**. Attempted ambush predation on spawning squids *Loligo vulgaris reynuadii* by benthic pyjama sharks, *Poroderma africanum*, off South Africa. *Journal of the Marine Biological Association of the UK* 75(3): 739–742.

Smale MJ, Watson G, Hecht T. **1995**. Otolith atlas of southern African marine fishes. *Ichthyological Monograph of the J.L.B. Smith Institute of Ichthyology* 1: i–iv + 253 pp.

Smith A. **1828**. Descriptions of new, or imperfectly known objects of the animal kingdom, found in the south of Africa. *South African Commercial Advertiser* 3(145): 2.

Smith A. **1829**. Contributions to the natural history of South Africa, &c. *Zoological Journal* 4(16): 433–444.

Smith A. **1837**. On the necessity for a revision of the groups included in the Linnean genus *Squalus*. *Proceedings of the Zoological Society of London* 5: 85–86.

Smith A. 1838–1849. Illustrations of the zoology of South Africa, consisting chiefly of figures and descriptions of the objects of natural history collected during an expedition into the interior of South Africa in the years 1834, 1835 and 1836; fitted out by "The Cape of Good Hope Association for Exploring Central Africa": together with a summary of African zoology, and an inquiry into the geographical ranges of species in that quarter of the globe. 5 Volumes [Vol. 4: Pisces]. London: Smith Elder & Co. [unpaginated]

Smith CL. 1971. A revision of the American groupers: Epinephelus and allied genera. Bulletin of the American Museum of Natural History 146(art.2): 67–242.

Smith CL, Bailey RM. **1961**. Evolution of the dorsal-fin supports of percoid fishes. *Papers of the Michigan Academy of Science, Arts and Letters* **46**: 345–363.

Smith CL, Tyler JC. **1969**. Observations on the commensal relationship of the western Atlantic pearlfish, *Carapus bermudensis*, and holothurians. *Copeia* 1969(1): 206–208.

Smith CR, Baco AR. **2003**. Ecology of whale falls at the deepsea floor. *Oceanography and Marine Biology: An Annual Review* 41: 311–354.

Smith DG. **1970**. Notacanthiform leptocephali in the western North Atlantic. *Copeia* 1970(1): 1–9.

Smith DG. **1989**. Family Chlopsidae. False morays. In: Fishes of the western North Atlantic. *Memoir of the Sears Foundation for Marine Research* 1(part 9): 72–97. Smith DG. **2002**. *Enchelycore nycturanus*, a new moray eel from South Africa (Teleostei: Anguilliformes: Muraenidae). *Zootaxa* 104: 1–6.

Smith DG. 2012. A checklist of the moray eels of the world (Teleostei: Anguilliformes: Muraenidae). Zootaxa 3474: 1–64. http://dx.doi.org/10.11646/zootaxa.3474.1.1

Smith DG, Bogorodsky SV, Mal AO, Alpermann TJ. **2019**. Review of the moray eels (Anguilliformes: Muraenidae) of the Red Sea, with description of a new species. *Zootaxa* 4704(1): 1–87. http://dx.doi/10.11646/zootaxa.4704.1.1

Smith DG, Böhlke EB. 1997. A review of the Indo-Pacific banded morays of the *Gymnothorax reticularis* group, with descriptions of three new species (Pisces, Anguilliformes, Muraenidae). Proceedings of the Academy of Natural Sciences of Philadelphia 148: 177–188.

Smith DG, Böhlke JE. 1983. Neenchelys retropinna: a new worm eel (Pisces: Ophichthidae) from the Indian Ocean. Proceedings of the Academy of Natural Sciences of Philadelphia 135: 80–84.

Smith DG, Brokovich E, Einbinder S. 2008. *Gymnothorax* baranesi, a new moray eel (Anguilliformes: Muraenidae) from the Red Sea. Zootaxa 1678: 63–68. http://dx.doi.org/10.11646/zootaxa.1678.1.4

Smith DG, Johnson GD. 2007. A new species of *Pteropsaron* (Teleostei: Trichonotidae: Hemerocoetinae) from the western Pacific, with notes on related species. *Copeia* 2007(2): 364–377.

Smith DG, Karmovskaya ES. 2003. A new genus and two new species of congrid eels (Teleostei: Anguilliformes: Congridae) from the Indo-West Pacific, with a redescription and osteology of *Chiloconger dentatus*. *Zootaxa* 343: 1–19.

Smith HM. 1912. The squaloid sharks of the Philippine Archipelago, with descriptions of new genera and species. *Proceedings of the United States National Museum* 41(1877): 677–685, Pls. 50–54.

Smith HM. **1913**. Description of a new carcharioid shark from the Sulu archipelago. *Proceedings of the United States National Museum* 45(2003): 599–601.

Smith HM. 1932. Contributions to the ichthyology of Siam.I. Descriptions of a new genus and three new species of Siamese gobies. *Journal of the Siam Society*, Natural History Supplement 8(4): 255–262.

Smith HM. **1934**. Contributions to the ichthyology of Siam. IX–XIX. *Journal of the Siam Society*, Natural History Supplement 9(3): 287–325, Pls. 10–14.

Smith HM. **1938**. Status of the oriental fish genera *Aplocheilus* and *Panchax*. *Proceedings of the Biological Society of Washington* 51: 165–166.

Smith HM. **1945**. The freshwater fishes of Siam, or Thailand. *Bulletin of the United States National Museum* 188: 1–622. Smith HM, Pope TEB. **1906**. List of fishes collected in Japan in 1903, with descriptions of new genera and species. *Proceedings of the United States National Museum* 31(1489): 459–499.

Smith HM, Radcliffe L. 1911. Descriptions of three new fishes of the family Chaetodontidæ from the Philippine Islands.
[Scientific results of the Philippine cruise of the Fisheries steamer Albatross, 1907–1910, No. 9] Proceedings of the United States National Museum 40(1822): 319–326.

Smith HM, Radcliffe L. 1912. In: Smith HM.

Smith HM, Radcliffe L. 1912. In: Radcliffe L.

Smith HM, Radcliffe L. 1912. Description of a new family of pediculate fishes from Celebes. *Proceedings of the United States National Museum* 42(1917): 579–581, Pl. 72.
Smith HM, Radcliffe L. 1913. In: Radcliffe L.

Smith JLB. **1931**. New and little known fish from the south and east coasts of Africa. *Records of the Albany Museum* 4(1): 145–160.

Smith JLB. **1933**. The South African species of the genus *Hemirhamphus* Cuv. *Transactions of the Royal Society of South Africa* 21(2): 129–150.

Smith JLB. 1934. The growth changes of *Pteroplatea natalensis*, G. & T. *Transactions of the Royal Society of South Africa* 22(1): 83–87.

Smith JLB. **1934**. Marine fishes of seven genera new to South Africa. *Transactions of the Royal Society of South Africa* 22(1): 89–100.

Smith JLB. **1934**. The Triglidae of South Africa. *Transactions of the Royal Society of South Africa* 22(4): 321–336.

Smith JLB. **1935**. New and little known fishes from South Africa. *Records of the Albany Museum Grahamstown* 4(2): 169–235.

Smith JLB. **1935**. The South African species of the family Aluteridae. *Records of the Albany Museum Grahamstown* 4(2): 358–364.

Smith JLB. **1935**. The fishes of the family Mugilidae in South Africa. *Annals of the South African Museum* 30(5): 587–644.

Smith JLB. **1935**. The "Galjoen" fishes of South Africa. *Transactions of the Royal Society of South Africa* 23(3): 265–276.

Smith JLB. **1936**. The genus *Tripterodon* Playfair. *Transactions of the Royal Society of South Africa* 23(4): 303–310.

Smith JLB. **1936**. Two interesting new fishes from South Africa. *Transactions of the Royal Society of South Africa* 24(1): 1–6.

Smith JLB. **1936**. New gobioid and cyprinid fishes from South Africa. *Transactions of the Royal Society of South Africa* 24(1): 47–55.

Smith JLB. **1937**. New records of South African fishes. *Annals of the Natal Museum* 8(2): 167–197.

Smith JLB. 1938. The South African fishes of the families Sparidae and Denticidae. *Transactions of the Royal Society of South Africa* 26(3): 225–305. Smith JLB. **1938**. A new gobioid fish from South Africa. *Transactions of the Royal Society of South Africa* 26(4): 319–320.

Smith JLB. **1939**. A surviving fish of the order Actinistia. *Transactions of the Royal Society of South Africa* 27(1): 47–50.

Smith JLB. **1939**. New records and descriptions of marine fishes from Portugese East Africa. *Transactions of the Royal Society of South Africa* 27(3): 215–222.

Smith JLB. **1939**. A living coelacanthid fish from South Africa. *Transactions of the Royal Society of South Africa* 28(1): 1–106.

Smith JLB. **1939**. A living fish of Mesozoic type. *Nature* (London) 143: 455–456.

Smith JLB. **1939**. The living coelacanthid fish from South Africa. *Nature* (London) 143: 748–750.

Smith JLB. 1939. A living fossil. The Cape Naturalist 1(6): 187–194. [Reprinted in Annual report of the Smithsonian Institution [for 1940]: 321–328]

Smith JLB. 1940. Sparid fishes from Portuguese East Africa, with a note on the genus *Gymnocranius* Klunzinger. *Transactions of the Royal Society of South Africa* 28(2): 175–182.

Smith JLB. **1941**. The genus *Gymnocranius* Klunzinger, with notes on certain rare fishes from Portuguese East Africa. *Transactions of the Royal Society of South Africa* 28(5): 441–452.

Smith JLB. **1942**. The genus *Austrosparus* Smith. *Transactions* of the Royal Society of South Africa 29(4): 279–283.

Smith JLB. **1943**. Interesting early juvenile stadia of certain well-known South African fishes. *Transactions of the Royal Society of South Africa* 30(1): 49–57.

Smith JLB. **1943**. Interesting new fishes of three genera new to South Africa, with a note on *Mobula diabolus* (Shaw). *Transactions of the Royal Society of South Africa* 30(1): 67–77.

Smith JLB. **1946**. The fishes of the family Clinidae in South Africa. *Annals and Magazine of Natural History* (Series 11) 12(92): 535–546.

Smith JLB. 1947. New species and new records of fishes from South Africa. Annals and Magazine of Natural History (Series 11) 13(108): 793–821.

Smith JLB. **1948**. Brief revisions and new records of South African marine fishes. *Annals and Magazine of Natural History* (Series 11) 14(113): 335–346.

Smith JLB. **1948**. New clinid fishes from the south western Cape, with notes on other fishes. *Annals and Magazine of Natural History* (Series 11) 14(118): 732–736.

Smith JLB. 1948. A generic revision of the mugilid fishes of South Africa. Annals and Magazine of Natural History (Series 11) 14(120): 833–843. Smith JLB. **1949**. Forty-two fishes new to South Africa with notes on others. *Annals and Magazine of Natural History* (Series 12) 2(14): 97–111.

Smith JLB. **1949**. A new aracanid fish from South Africa. *Annals and Magazine of Natural History* (Series 12) 2(17): 354–359.

Smith JLB. **1949**. Interesting fishes of three genera new to South Africa. *Annals and Magazine of Natural History* (Series 12) 2(17): 367–374.

Smith JLB. **1949**. The stromateid fishes of South Africa. *Annals and Magazine of Natural History* (Series 12) 2(23): 839–851.

Smith JLB. **1949**. *The sea fishes of southern Africa*. South Africa: Central News Agency, Ltd. 550 pp. [Followed by revised editions in 1950, 1953, 1961, 1965 and 1977 (under the title *Smith's Sea Fishes*), and succeeded by Smith & Heemstra's *Smiths' Sea Fishes*]

Smith JLB. **1949**. Taxonomy and ecology of the fishes of southern Africa. *African Regional Scientific Conference, Johannesburg, Communication* No. C(j) 2: 1–3.

Smith JLB. **1950**. Two noteworthy non-marine fishes from South Africa. *Annals and Magazine of Natural History* (Series 12) 3(32): 705–710.

Smith JLB. **1950**. *Pomadasys operculare* Playfair, and the South African seas. *Annals and Magazine of Natural History* (Series 12) 3(33): 778–785.

Smith JLB. 1950. A new dogfish from South Africa with notes on other chondrichthyan fishes. *Annals and Magazine of Natural History* (Series 12) 3(34): 878–887.

Smith JLB. 1950. The sea fishes of southern Africa (2<sup>nd</sup> impression). South Africa: Central News Agency Ltd. 376 pp.

Smith JLB. **1951**. A case of poisoning by the stonefish, *Synanceja verrucosa*. *Copeia* 1951(3): 207–210.

Smith JLB. **1951**. Thirteen noteworthy additions to the southeast African marine fauna. *Annals and Magazine of Natural History* (Series 12) 4(37): 49–66.

Smith JLB. **1951**. A new galeorhinid shark from South Africa, with notes on other species. *Annals and Magazine of Natural History* (Series 12) 4(37): 86–93.

Smith JLB. **1951**. The fishes of the family Veliferidae from south east Africa. *Annals and Magazine of Natural History* (Series 12) 4(41): 497–510.

Smith JLB. 1951. The genus Paragobioides Kendall & Goldsborough, 1911. Annals and Magazine of Natural History (Series 12) 4(41): 518–527.

Smith JLB. **1951**. The fishes of the family Cirrhitidae of the western Indian Ocean. *Annals and Magazine of Natural History* (Series 12) 4(43): 625–652.

Smith JLB. 1951. A juvenile of the man-eater, Carcharodon carcharias Linn. Annals and Magazine of Natural History (Series 12) 4(44): 729–736. Smith JLB. 1951. Trigger action in *Quinquarius capensis* Cuvier (1829), with a description of the adult form. *Annals and Magazine of Natural History* (Series 12) 4(45): 873–882.

Smith JLB. 1951. Sexual dimorphism in the genus Naso Lacepede, 1802, with a description of a new species and new records. Annals and Magazine of Natural History (Series 12) 4(47): 1126–1132.

Smith JLB. **1952**. The fishes of the family Haliophidae. *Annals and Magazine of Natural History* (Series 12) 5(49): 85–101.

Smith JLB. 1952. Plesiopid fishes from South and East Africa. Annals and Magazine of Natural History (Series 12) 5(50): 139–151.

Smith JLB. **1952**. A new hound shark from South Africa, and new records. *Annals and Magazine of Natural History* (Series 12) 5(51): 223–226.

Smith JLB. **1952**. The fishes of the family Batrachoididae from South Africa and East Africa. *Annals and Magazine of Natural History* (Series 12) 5(52): 313–339.

Smith JLB. **1952**. Preliminary notes on fishes of the family Plectorhynchidae from South and East Africa, with descriptions of two new species. *Annals and Magazine of Natural History* (Series 12) 5(55): 711–716.

Smith JLB. **1952**. Two chondrichthyan fishes new to South Africa. *Annals and Magazine of Natural History* (Series 12) 5(56): 760–765.

Smith JLB. **1952**. *Carcharinus zambezensis* Peters, 1852, with notes on other chondrichthyan fishes. *Annals and Magazine of Natural History* (Series 12) 5(57): 857–863.

Smith JLB. **1952**. Tropical fishes recently found in South Africa. *Annals and Magazine of Natural History* (Series 12) 5(59): 1020–1025.

Smith JLB. **1952**. Discovery of *Tetragonurus* Risso, 1810, in South African waters. *Nature* (London) 170: 496.

Smith JLB. **1953**. A shark new to the African coast. *South African Angler* 7(11): 8.

Smith JLB. **1953**. The genus *Tetragonurus* Risso, 1810. *Annals and Magazine of Natural History* (Series12) 6(61): 53–66.

Smith JLB. 1953. An interesting new plectorhynchid fish from East Africa. *Annals and Magazine of Natural History* (Series 12) 6(62): 158–160.

Smith JLB. **1953**. The fishes of the family Pseudogrammidae from East Africa. *Annals and Magazine of Natural History* (Series 12) 6(68): 548–560.

Smith JLB. 1953. The giant "Cushionhead Parrotfish" of Kenya, *Callyodon muricatus* (Valenciennes), and its growth stadia. *Annals and Magazine of Natural History* (Series 12) 6(68): 620–622.

Smith JLB. 1953. Fishes taken in the Moçambique Channel by Mussolini P. Fajardo. *Memórias do Museu Dr. Álvaro de Castro* 2: 5–21.

Smith JLB. **1953**. Problems of the coelacanth. *South African Journal of Science* 49(9): 279–281.

Smith JLB. **1953**. *The sea fishes of southern Africa* (3<sup>rd</sup> edition; revised enlarged edition). Cape Town: Central News Agency. 564 pp.

Smith JLB. 1953. The second coelacanth. Copeia 1953(1): 72.

Smith JLB. **1953**. The second coelacanth. *Nature* (London) 171: 99–101.

Smith JLB. **1953**. The shark, *Isurus oxyrinchus*, in South African waters. *Nature* (London) 171: 977.

Smith JLB. **1954**. Two interesting new anthiid fishes from East Africa. *Annals and Magazine of Natural History* (Series 12) 7(73): 1–6.

Smith JLB. **1954**. Pseudoplesiopsine fishes from South and East Africa. *Annals and Magazine of Natural History* (Series 12) 7(75): 195–208.

Smith JLB. 1954. The Anisochromidae, a new family of fishes from East Africa. *Annals and Magazine of Natural History* (Series 12) 7(76): 298–302.

Smith JLB. **1954**. Fishes new to Africa obtained by deep-line fishing in Kenya waters, with a revision of the East African species of the genus *Pristipomoides* Bleeker, 1852. *Annals and Magazine of Natural History* (Series 12) 7(79): 481–492.

Smith JLB. **1954**. Apogonid fishes of the subfamily Pseudamiinae from south-east Africa. *Annals and Magazine of Natural History* (Series 12) 7(82): 775–795.

Smith JLB. **1954**. Aberrant serraniform fishes from East Africa. *Annals and Magazine of Natural History* (Series 12) 7(83): 861–872.

Smith JLB. **1955**. Four rare serraniform fishes from East Africa. *Annals and Magazine of Natural History* (Series 12) 7(84): 925–933.

Smith JLB. **1955**. Siphamine fishes from South and East Africa. *Annals and Magazine of Natural History* (Series 12) 8(85): 61–66.

Smith JLB. **1955**. An interesting new gobiiform fish from South Africa. *Annals and Magazine of Natural History* (Series 12) 8(86): 106–110.

Smith JLB. **1955**. An especially colourful new pseudochromid fish. *Annals and Magazine of Natural History* (Series 12) 8(86): 145–148.

Smith JLB. **1955**. The fishes of Aldabra. Part 1. *Annals and Magazine of Natural History* (Series 12) 8(88): 304–312.

Smith JLB. 1955. The fishes of the family Anthiidae of the western Indian Ocean. Annals and Magazine of Natural History (Series 12) 8(89): 337–350.

Smith JLB. **1955**. The fishes of the family Pomacanthidae in the western Indian Ocean. *Annals and Magazine of Natural History* (Series 12) 8(89): 377–384.

Smith JLB. **1955**. The fishes of the family Carapidae in the western Indian Ocean. *Annals and Magazine of Natural History* (Series 12) 8(90): 401–416.

Smith JLB. **1955**. The genus *Pyramodon* Smith & Radcliffe, 1913. *Annals and Magazine of Natural History* (Series 12) 8(91): 545–550.

Smith JLB. **1955**. East African unicorn fishes from Mozambique. *South African Journal of Science* 51(6): 169–174.

Smith JLB. 1955. New species and new records of fishes from Mozambique. Part 1. *Memórias do Museu Dr. Álvaro de Castro* 3: 3–27.

Smith JLB. **1956**. Self-inflation in a gobioid fish. *Nature* (London) 177: 714.

Smith JLB. **1956**. The striped marlin (*Makaira audax* Phillipi) in South Africa. *Nature* (London) 177: 758.

Smith JLB. **1956**. A marlin in Angola, with a note on *Makaria hershelii* (Gray), 1838. *Nature* (London) 177: 1246–1247.

Smith JLB. **1956**. The fishes of Aldabra. Part II. *Annals and Magazine of Natural History* (Series 12) 8(93): 689–697.

Smith JLB. **1956**. The fishes of Aldabra. Part III. *Annals and Magazine of Natural History* (Series 12) 8(96): 886–896.

Smith JLB. 1956. The fishes of Aldabra. Part IV. Annals and Magazine of Natural History (Series 12) 8(96): 928–937.

Smith JLB. **1956**. An extraordinary fish from South Africa. *Annals and Magazine of Natural History* (Series 12) 9(97): 54–57.

Smith JLB. **1956**. Two new plectorhynchid fishes from Ceylon with a note on *Sciaena foetela* Forskal, 1775. *Annals and Magazine of Natural History* (Series 12) 9(98): 97–101.

Smith JLB. **1956**. A new dealfish from South Africa. *Annals and Magazine of Natural History* (Series 12) 9(102): 449–452.

Smith JLB. 1956. An interesting new gobioid fish from Madagascar, with a note on *Cryptocentrus oni* Tomiyama, 1936. *Annals and Magazine of Natural History* (Series 12) 9(104): 553–556.

Smith JLB. **1956**. The parrot fishes of the family Callyodontidae of the western Indian Ocean. *Ichthyological Bulletin* (Rhodes University) 1: 1–23.

Smith JLB. 1956. Swordfish, marlin and sailfish in South and East Africa. *Ichthyological Bulletin* (Rhodes University) 2: 23–34.

Smith JLB. **1956**. The fishes of the family Sphyraenidae in the western Indian Ocean. *Ichthyological Bulletin* (Rhodes University) 3: 37–46.

Smith JLB. **1956**. Pugnacity of marlins and swordfish. *Nature* (London) 178: 1065.

Smith JLB. **1956**. The genus *Luzonichthys* Herre. *Copeia* 1956(4): 251.

Smith JLB. **1957**. A new shark from South Africa. *South African Journal of Science* 53(10): 261–264.

Smith JLB. 1957. A remarkable new unicorn fish from East Africa. Annals and Magazine of Natural History (Series 12) 9(105) [for 1956]: 686–688. Smith JLB. 1957. The fishes of Aldabra. Part V. Annals and Magazine of Natural History (Series 12) 9(106) [1956]: 721–729.

Smith JLB. 1957. The fishes of Aldabra. Part VI. Annals and Magazine of Natural History (Series 12) 9(107) [1956]: 817–829.

Smith JLB. **1957**. The fishes of Aldabra. Part VII. *Annals and Magazine of Natural History* (Series 12) 9(108): 888–892.

Smith JLB. **1957**. Deep-line fishing in northern Mozambique, with the description of a new pentapodid fish. *Annals and Magazine of Natural History* (Series 12) 10(110): 121–124.

Smith JLB. **1957**. Fishes of Aldabra. Part VIII. *Annals and Magazine of Natural History* (Series 12) 10(113): 395–400.

Smith JLB. **1957**. Dasyatid rays new to South Africa. *Annals and Magazine of Natural History* (Series 12) 10(114): 429–431.

Smith JLB. 1957. The rare shark *Hemipristis elongatus* (Klunzinger), 1871, from Zanzibar and Mozambique. *Annals and Magazine of Natural History* (Series 12) 10(115): 555–560.

Smith JLB. 1957. A new shark from Zanzibar, with notes on Galeorhinus Blainville. Annals and Magazine of Natural History (Series 12) 10(116): 585–592.

Smith JLB. **1957**. A preliminary survey of the Scylliogaleid dogfishes of South Africa. *South African Journal of Science* 53(14): 353–359.

Smith JLB. **1957**. Four interesting new fishes from South Africa. *South African Journal of Science* 53(8): 219–222.

Smith JLB. 1957. The fishes of the family Scorpaenidae in the western Indian Ocean. Part 1. The subfamily Scorpaeninae. *Ichthyological Bulletin* (Rhodes University) 4: 49–72.

Smith JLB. 1957. The fishes of the family Scorpaenidae in the western Indian Ocean, Part II. The subfamilies Pteroinae, Apistinae, Setarchinae and Sebastinae. *Ichthyological Bulletin* (Rhodes University) 5: 75–88.

Smith JLB. **1957**. Sharks of the genus *Isurus* Rafinesque, 1810. *Ichthyological Bulletin* (Rhodes University) 6: 91–96.

Smith JLB. 1957. List of the fishes of the family Labridae in the western Indian Ocean with new records and five new species. *Ichthyological Bulletin* (Rhodes University) 7: 99–114.

Smith JLB. 1957. The labrid fishes of the subgenus Julis Cuvier, 1814 (in Coris Lacepede, 1802) from South and East Africa. Ichthyological Bulletin (Rhodes University) 8: 117–120.

Smith JLB. **1957**. Two rapid fatalities from stonefish stabs. *Copeia* 1957(3): 249.

Smith JLB. **1958**. Dasyatid rays new to South Africa. *Annals and Magazine of Natural History* (Series 12) 10(114) [for 1957]: 429–431.

Smith JLB. **1958**. The fishes of Aldabra. Part IX. *Annals and Magazine of Natural History* (Series 12) 10(119): 833–842.

Smith JLB. **1958**. The fishes of Aldabra. Part X. *Annals and Magazine of Natural History* (Series 13) 1(1): 57–63.

Smith JLB. **1958**. Tetraodont fishes from South and East Africa. *Annals and Magazine of Natural History* (Series 13) 1(2): 156–160.

Smith JLB. 1958. The genus *Limnichthys* Waite, 1904 in African seas. *Annals and Magazine of Natural History* (Series 13) 1(4): 247–249.

Smith JLB. 1958. The gunnellichthid fishes with description of two new species from East Africa and of *Gunnellichthys* (*Clarkichthys*) bilineatus (Clark), 1936. Ichthyological Bulletin (Rhodes University) 9: 123–129.

Smith JLB. **1958**. Sharks of the genus *Pterolamiops* Springer, 1951 with notes on isurid sharks. *Ichthyological Bulletin* (Rhodes University) 10: 131–134.

Smith JLB. 1958. The fishes of the family Eleotridae in the western Indian Ocean. *Ichthyological Bulletin* (Rhodes University) 11: 137–163.

Smith JLB. 1958. Fishes of the families Tetrarogidae, Caracanthidae and Synanciidae from the western Indian Ocean with further notes on scorpaenid fishes. *Ichthyological Bulletin* (Rhodes University) 12: 167–181.

Smith JLB. **1958**. New and rare fishes from South Africa. *South African Journal of Science* 54(5): 123–129.

Smith JLB. **1958**. Shark attacks in South Africa. *South African Journal of Science* 54(6): 150–152.

Smith JLB. **1958**. Rare fishes from South Africa. *South African Journal of Science* 54(12): 319–323.

Smith JLB. 1958. The marine fishes of Inhaca (pp. 113–116).
In: MacNae W, Kalk M (eds) *A natural history of Inhaca Island, Mocambique*. Johannesburg: Witwatersrand University Press. 163 pp.

Smith JLB. **1958**. The mystery killer, the new shark *Carcharhinus Vanrooyeni*. *Veld & Vlei* 3(9): 12–14, 28.

Smith JLB. 1959. Gobioid fishes of the families Gobiidae, Periophthalmidae, Trypauchenidae, Taenioididae, and Kraemeriidae of the western Indian Ocean. *Ichthyological Bulletin* (Rhodes University) 13: 185–225. Pls. 9–13.

Smith JLB. 1959. Fishes of the families Blenniidae and Salariidae of the western Indian Ocean. *Ichthyological Bulletin* (Rhodes University) 14: 229–252.

Smith JLB. **1959**. Serioline fishes (yellowtails : amberjacks) from the western Indian Ocean. *Ichthyological Bulletin* (Rhodes University) 15: 255–261.

Smith JLB. **1959**. The identity of *Scarus gibbus* Ruppell, 1828 and of other parrotfishes of the family Callyodontidae from the Red Sea and the western Indian Ocean. *Ichthyological Bulletin* (Rhodes University) 16: 265–282.

Smith JLB. 1959. Fishes of the family Lethrinidae from the western Indian Ocean. *Ichthyological Bulletin* (Rhodes University) 17: 285–295. Smith JLB. **1960**. A new grammicolepid fish from South Africa. *Annals and Magazine of Natural History* (Series 13) 3(28): 231–235.

Smith JLB. **1960**. Fishes of the family Gobiidae in South Africa. *Ichthyological Bulletin* (Rhodes University) 18: 299–314.

Smith JLB. **1960**. Coral fishes of the family Pomacentridae from the western Indian Ocean and the Red Sea. *Ichthyological Bulletin* (Rhodes University) 19: 317–347.

Smith JLB. **1960**. Two interesting fishes from South Africa. *South African Journal of Science* 56(4): 91–92.

Smith JLB. **1961**. *Oreosoma atlanticum* Cuvier, 1829, in South Africa, with notes on other zeid fishes. *Annals and Magazine of Natural History* (Series 13) 3(33) [for 1960]: 565–569.

Smith JLB. **1961**. A new clinid fish from South Africa. *Annals and Magazine of Natural History* (Series 13) 3(35): 689–691.

Smith JLB. **1961**. A rare lutianid fish from Kenya. *Annals and Magazine of Natural History* (Series 13) 3(36): 753–755.

Smith JLB. **1961**. A new stromateid fish from South Africa, and another new to that area. *South African Journal of Science* 57(6): 158–160.

Smith JLB. **1961**. Fishes of the family Xenopoclinidae. *Ichthyological Bulletin* (Rhodes University) 20: 351–356.

Smith JLB. 1961. Fishes of the family Anthiidae from the western Indian Ocean and the Red Sea. *Ichthyological Bulletin* (Rhodes University) 21: 359–369.

Smith JLB. 1961. Fishes of the family Apogonidae of the western Indian Ocean and the Red Sea. *Ichthyological Bulletin* (Rhodes University) 22: 373–418.

Smith JLB. **1961**. *The sea fishes of southern Africa* (4<sup>th</sup> edition). Cape Town: Central News Agency. 580 pp.

Smith JLB. 1962. Fishes from the Cape described by Lichtenstein in 1823. South African Journal of Science 58(2): 39–40.

Smith JLB. **1962**. The sparid genus *Lithognathus*, Swainson, 1839 with description of an interesting new species. *South African Journal of Science* 58(4): 109–114.

Smith JLB. 1962. The moray eels of the western Indian Ocean and the Red Sea. *Ichthyological Bulletin* (Rhodes University) 23: 421–444.

Smith JLB. **1962**. Sand-dwelling eels of the western Indian Ocean and the Red Sea. *Ichthyological Bulletin* (Rhodes University) 24: 447–466.

Smith JLB. 1962. Fishes of the family Gaterinidae of the western Indian Ocean and the Red Sea, with a resumé of all known Indo-Pacific species. *Ichthyological Bulletin* (Rhodes University) 25: 469–502.

Smith JLB. 1962. The rare "furred-tongue" Uraspis uraspis (Günther) from South Africa, and other new records from there. Ichthyological Bulletin (Rhodes University) 26: 505–511. Smith JLB. **1962**. Nomenclatorial change for a long-known South African fish. *Annals of the South African Museum* 46(10): 257–260.

Smith JLB. **1963**. Fishes of the family Syngnathidae from the Red Sea and the western Indian Ocean. *Ichthyological Bulletin* (Rhodes University) 27: 515–543.

Smith JLB. 1963. Fishes of the families Draconettidae and Callionymidae from the Red Sea and the western Indian Ocean. *Ichthyological Bulletin* (Rhodes University) 28: 547–564.

Smith JLB. 1963. The identity of *Hapalogenys pictus* Tortonese, 1935. Annals and Magazine of Natural History (Series 13) 5(58) [1962]: 637–638.

Smith JLB. **1963**. New species and new records of fishes from the western Indian Ocean. *Annals and Magazine of Natural History* (Series 13) 6(61): 33–37.

Smith JLB. **1963**. Shark attacks in the South African seas. In: Gilbert PW (ed) *Sharks and survival*. Boston: D.C. Heath & Co. 578 pp.

Smith JLB. 1964. A new gobiid fish of unusual type from Mozambique. Annals and Magazine of Natural History (Series 13) 7(75): 173–176.

Smith JLB. 1964. Fishes collected by Dr. Th. Mortensen off the coast of South Africa in 1929, with an account of the genus *Cruriraja* Bigelow & Schroeder, 1954 in South Africa. Videnskabelige Meddelelser fra Dansk Naturhistorisk Forening, Kjøbenhavn 126: 283–300.

Smith JLB. **1964**. Fishes of the family Pentacerotidae. *Ichthyological Bulletin* (Rhodes University) 29: 567–578.

Smith JLB. 1964. Scombroid fishes of South Africa. In: Proceedings of the Symposium on Scombroid Fishes, Marine Biological Association of India (MBAI), 12–15 January 1962, Mandapam. Part 1: 165–183.

Smith JLB. 1964. The clingfishes of the western Indian Ocean and the Red Sea. *Ichthyological Bulletin* (Rhodes University) 30: 581–596.

Smith JLB. 1964. The rediscovery in South Africa of a rare form of labrid fish from Mauritius and its sexual dichromatic relation to *Lepidaplois axillaris* (Bennett 1831). *Annals and Magazine of Natural History* (Series 13) 7(81): 177–183.

Smith JLB. **1965**. *Acanthurus bahianus* Castelnau, 1855, in the southeast Atlantic Ocean. *Copeia* 1965(1): 110–111.

Smith JLB. **1965**. A rare anthiid fish from Cook Island, Pacific, with a résumé of related species. *Annals and Magazine of Natural History* (Series 13) 7(81): 533–537.

Smith JLB. **1965**. The discovery in Mozambique of the little known eel *Ophichthys tenuis* Günther, 1870, a redescription of the type of *Caecula pterygera* Vahl, 1794, notes on other species and on generic relationships. *Annals and Magazine of Natural History* (Series 13) 7(84) [for 1964]: 711–723.

Smith JLB. **1965**. Fishes of the family Atherinidae of the Red Sea and the western Indian Ocean with a new freshwater genus and species from Madagascar. *Ichthyological Bulletin* (Rhodes University) 31: 601–632.

Smith JLB. **1965**. New records and descriptions of fishes from Southwest Africa. *Department of Ichthyology, Rhodes University, Occasional Paper* 3: 13–23.

Smith JLB. **1965**. New records and new species of fishes from South Africa, chiefly from Natal. *Department of Ichthyology, Rhodes University, Occasional Paper* 4: 27–42.

Smith JLB. **1965**. *Kaupichthys diodontus* Schultz, in the western Indian Ocean. A problem in systematics. *Department of Ichthyology, Rhodes University, Occasional Paper* 5: 45–54.

Smith JLB. **1965**. *The sea fishes of southern Africa* (5<sup>th</sup> edition). South Africa: Central News Agency. 580 pp.

Smith JLB. 1966. An interesting new eel of the family Xenocongridae from Cook Island, Pacific. Annals and Magazine of Natural History (Series 13) 8(89): 297–301.

Smith JLB. **1966**. An interesting new callionymid fish from Madagascar and the first record of a clingfish from there. *Annals and Magazine of Natural History* (Series 13) 8(90): 321–324.

Smith JLB. **1966**. The rare northeastern Atlantic fish *Centrolophus brittannicus*, in South Africa. *Annals and Magazine of Natural History* (Series 13) 8(92): 505–509.

Smith JLB. **1966**. A new clingfish from southern Mozambique. Annals and Magazine of Natural History (Series 13) 8(95) [for 1965]: 641–644.

Smith JLB. **1966**. A new stromateid fish from South Africa with illustration of the unique rare *Centrolophus huttoni* Waite, 1910. *Annals and Magazine of Natural History* (Series 13) 9(97–99): 1–3.

Smith JLB. 1966. A rare acanthurid fish from Mauritius in South Africa. *Annals and Magazine of Natural History* (Series 13) 9(97–99): 5–8.

Smith JLB. **1966**. Certain rare fishes from South Africa with other notes. *Department of Ichthyology, Rhodes University, Occasional Paper* 7: 65–80.

Smith JLB. **1966**. Fishes of the sub-family Nasinae with a synopsis of the Prionurinae. *Ichthyological Bulletin* (Rhodes University) 32: 635–682.

Smith JLB. **1966**. Interesting fishes from South Africa. Department of Ichthyology, Rhodes University, Occasional Paper 8: 83–94.

Smith JLB. 1966. The rare big-eye *Pristigenys niphonia* (C & V) in South Africa. *Department of Ichthyology, Rhodes University, Occasional Paper* 9: 97–102.

Smith JLB. **1967**. The lizard shark *Chlamydoselachus anguineus* Garman in South Africa. *Department of Ichthyology, Rhodes University, Occasional Paper* 10: 105–114.

Smith JLB. 1967. A new squalid shark from South Africa with notes on the rare *Atractophorus armatus* Gilchrist. *Department of Ichthyology, Rhodes University, Occasional Paper* 11: 117–136.

Smith JLB. **1967**. Studies in carangid fishes No. 1. Naked thoracic areas. *Department of Ichthyology, Rhodes University, Occasional Paper* 12: 139–141.

Smith JLB. 1967. Studies in carangid fishes No. 2. The identity of Scomber malabaricus Bloch-Schneider, 1801. Department of Ichthyology, Rhodes University, Occasional Paper 13: 143–153.

Smith JLB. 1967. Studies in carangid fishes No. 3. The genus Trachinotus Lacepede, in the western Indian Ocean. Department of Ichthyology, Rhodes University, Occasional Paper 14: 157–166.

Smith JLB. **1967**. Pugnacity of the whale shark, *Rhincodon*. *Copeia* 1967(1): 237.

Smith JLB. **1967**. Two flatfishes new to South and East Africa. *Journal of Natural History* 1(4): 457–464.

Smith JLB. **1967**. A new liparine fish from the Red Sea. *Journal of Natural History* 2(1) [for 1968]: 105–109.

Smith JLB. **1968**. A new labrid fish from deep water of Mozambique. *Copeia* 1968(2): 343–345.

Smith JLB. 1968. New and interesting fishes from deepish water off Durban, Natal and southern Mozambique. *Investigational Report. Oceanographic Research Institute* (Durban) 19: 1–30.

Smith JLB. **1968**. Studies in carangid fishes No. 4. The identity of *Scomber sansun* Forsskal, 1775. *Department of Ichthyology, Rhodes University, Occasional Paper* 15: 173–184.

Smith JLB. 1969. Fishes of Inhaca recorded by Professor J.L.B.
Smith (pp. 131–136). In: Macnae W, Kalk M (eds) *A natural history of Inhaca Island, Moçambique* (revised edition).
Johannesburg: Witwatersrand University Press. 163 pp.

Smith JLB. 1970. Studies in carangid fishes No. 5. The genus Chorinemus Cuvier, 1831 in the western Indian Ocean. J.L.B. Smith Institute of Ichthyology, Rhodes University, Occasional Paper 17: 217–228.

Smith JLB. 1971. Marine fishes (pp. 231–255). In: Potgieter DJ, Du Plessis PC, Skaife SH (eds) Animal life in southern Africa. A comprehensive compilation of articles on zoology from Protozoa to Man. Cape Town, South Africa: Nasou Limited. 469 pp.

Smith JLB. **1977**. *Smith's sea fishes* (4<sup>th</sup> impression). Sandton: Valiant Publishers. 580 pp.

Smith JLB, Smith MM. 1963. The fishes of Seychelles.Grahamstown: Department of Ichthyology, Rhodes University. 215 pp.

Smith JLB, Smith MM. **1966**. *Fishes of the Tsitsikama Coastal National Park*. National Parks Board of Trustees of the Republic of South Africa. 161 pp. Smith JLB, Smith MM. 1969. The fishes of Seychelles (2<sup>nd</sup> edition). Grahamstown: J.L.B. Smith Institute of Ichthyology. 223 pp.

Smith JLB, Smith MM. 1986. Family No. 183: Sparidae.In: Smith MM, Heemstra PC (eds) *Smiths' sea fishes*.Johannesburg: Macmillan South Africa. 1047 pp.

Smith MM. **1969**. *Echidna tritor* (Vaillant & Sauvage, 1875), the large adult of *Echidna polyzona* (Richardson, 1845), and other interesting fishes collected by Dr R.A.C. Jensen in southern Mozambique waters. *Memórias do Instituto de Investigação Científica de Moçambique* (Série A) 9(1967–1968): 293–308.

Smith MM. **1969**. Early paintings of Cape fishes by Hendrik Claudius in the Africana Museum. *Africana Notes and News* 18(8): 325–327.

Smith MM. 1972. Studies in carangid fishes No. 6. Key to the western Indian Ocean species of the genus *Carangoides* Bleeker, 1851, with a description of *Carangoides nitidus* Smith. J.L.B. Smith Institute of Ichthyology, Rhodes University, Occasional Paper 18: 229–239.

Smith MM. **1973**. Identity of *Caranx armatus* (Pisces: Carangidae). *Copeia* 1973(2): 352–355.

Smith MM. **1975**. Common and scientific names of the fishes from southern Africa. Part 1: Marine fishes. *J.L.B. Smith Institute of Ichthyology Special Publication* 14: 1–178.

Smith MM. 1977. A new species of Argyrosomus (Pisces: Sciaenidae) from Natal, South Africa. Journal of Zoology (London) 181(4): 561–566.

Smith MM. **1977**. A note on *Anisochromis kenyae* Smith, 1954. *Ichthyological Bulletin of the J.L.B. Smith Institute of Ichthyology* 35: 22–23.

Smith MM. **1977**. *Sea and shore dangers. Their recognition, avoidance and treatment*. Grahamstown: J.L.B. Smith Institute of Ichthyology. 66 pp.

Smith MM. 1978. A new Polysteganus (Pisces, Sparidae) from Mauritius. Proceedings of the Biological Society of Washington 91(3): 563–568.

Smith MM. 1979. *Rhabdosargus thorpei*, a new sparid fish from South Africa, with a key to the species of *Rhabdosargus*. *Copeia* 1979(4): 702–709.

Smith MM. 1**979**. *Balistoides niger* vs *Balistoides conspicillum*. *Tropical Fish Hobbyist* 27(5#275): 44–47.

Smith MM. 1980. Marine fishes of Maputaland (pp. 164–187). In: Bruton MN, Cooper KH (eds) *Studies on the ecology* of Maputaland. Grahamstown: Rhodes University, and Durban: Natal Branch of the Wildlife Society of Southern Africa. 560 pp.

Smith MM. **1980**. A review of the South African cheilodactylidae fishes (Pisces: Perciformes), with descriptions of two new species. *Ichthyological Bulletin of the J.L.B. Smith Institute of Ichthyology* 42: 1–14.

Smith MM, Heemstra PC (eds). **1986**. *Smiths' sea fishes*. Johannesburg: Macmillan South Africa. 1047 pp.

Smith MM, Heemstra PC (eds). **1988**. *Smiths' sea fishes*. Johannesburg: Southern Book Publishers. 1047 pp.

Smith MM, Heemstra PC (eds). 1995. Smiths' sea fishes (3<sup>rd</sup> impression). Johannesburg: Southern Book Publishers. 1047 pp.

Smith MM, Heemstra PC (eds). **2003**. *Smiths' sea fishes*. Cape Town: Struik Publishers. 1047 pp.

Smith R. **1886**. On *Tetraodon setosus*, a new species allied to *Tetraodon meleagris* Lacép. *Bulletin of the California Academy of Sciences* 2: 155.

Smith WD, Bizzarro JJ, Richards VP, Nielsen J, Márquez-Farías F, Shivji MS. **2009**. Morphometric convergence and molecular divergence: the taxonomic status and evolutionary history of *Gymnura crebripunctata* and *Gymnura marmorata* in the eastern Pacific Ocean. *Journal* of Fish Biology 75(4): 761–783. https://doi.org/10.1111/ j.1095-8649.2009.02300.x

Smith WL, Craig MT. 2007. Casting the percomorph net widely: the importance of broad taxonomic sampling in the search for the placement of serranid and percid fishes. *Copeia* 2007(1): 35–55. https://doi.org/10.1643/0045-8511(2007)7[35:CTPNWT]2.0.CO;2

Smith WL, Smith KR, Wheeler WC. **2009**. Mitochondrial intergenic spacer in fairy basslets (Serranidae: Anthiinae) and the simultaneous analysis of nucleotide and genomic data. *American Museum Novitates* 3652: 1–10. http://dx.doi. org/10.1206/518.1

Smith WL, Webb JF, Blum SD. 2003. The evolution of the laterophysic connection with a revised phylogeny and taxonomy of butterflyfishes (Teleostei: Chaetodontidae). *Cladistics* 19(4): 287–306.

Smith WL, Wheeler WC. **2004**. Polyphyly of the mailcheeked fishes (Teleostei: Scorpaeniformes): evidence from mitochondrial and nuclear sequence data. *Molecular Phylogenetics and Evolution* 32(2): 627–646.

Smith WL, Wheeler WC. **2006**. Venom evolution widespread in fishes: a phylogenetic road map for the bioprospecting of piscine venoms. *Journal of Heredity* 97(3): 206–217.

Smith-Vaniz WF. **1969**. A new species of *Meiacanthus* (Pisces: Blenniidae: Nemophini) from the Red Sea, with a review of the Indian Ocean species. *Proceedings of the Biological Society of Washington* 82: 349–354.

Smith-Vaniz WF. 1976. The saber-toothed blennies, tribe Nemophini (Pisces: Blenniidae). *Monographs of the Academy of Natural Sciences of Philadelphia* Monograph 19: 1–196. Smith-Vaniz WF. 1983. Opistognathus margaretae, a new species of jawfish (Perciformes: Opistognathidae) from the Indian Ocean, with notes on O. nigromarginatus Rüppell and O. muscatensis Boulenger. J.L.B. Smith Institute of Ichthyology Special Publication 30: 1–10.

Smith-Vaniz WF. 1984. Carangidae: relationships. In: Moser HG, Richards WJ, Cohen DM, Fahay MP, Kendall SW Jr, Richardson SL (eds) Ontogeny and systematics of fishes. *American Society of Ichthyologists and Herpetologists Special Publication* 1: 522–530.

Smith-Vaniz WF. 1986. Family No. 210: Carangidae; Family No. 225: Opistognathidae; Family No. 226: Cepolidae.
In: Smith MM, Heemstra PC (eds) *Smiths' sea fishes*.
Johannesburg: Macmillan South Africa. 1047 pp.

Smith-Vaniz WF. 1987. The saber-toothed blennies, tribe Nemophini (Pisces: Blenniidae): an update. Proceedings of the Academy of Natural Sciences of Philadelphia 139: 1–52.

Smith-Vaniz WF. **1989**. Revision of the jawfish genus *Stalix* (Pisces: Opistognathidae), with descriptions of four new species. *Proceedings of the Academy of Natural Sciences of Philadelphia* 141: 375–407.

Smith-Vaniz WF. 1999. Opistognathidae. Jawfishes (pp. 2588–2589); Carangidae. Jacks and scads (also trevallies, queenfishes, runners, amberjacks, pilotfishes, pampanos, etc.) (pp. 2659–2790). In: Carpenter KE, Niem VH (eds) FAO species identification guide for fisheries purposes. The living marine resources of the western central Pacific. Vol. 4. Bony fishes part 2 (pp. 2069–2790). Rome: FAO.

Smith-Vaniz WF. 2001. Family Cepolidae. Bandfishes (pp. 3331–3332). In: Carpenter KE, Niem VH (eds) FAO species identification guide for fishery purposes. The living marine resources of the western central Pacific. Vol 5. Bony fishes part 3 (pp. 2791–3380). Rome: FAO.

Smith-Vaniz WF. **2009**. Three new species of Indo-Pacific jawfishes (*Opistognathus*: Opistognathidae), with the posterior end of the upper jaw produced as a thin flexible lamina. *aqua, International Journal of Ichthyology* 15(2): 69–108.

Smith-Vaniz WF. **2010**. New species of Indo-Pacific jawfishes (*Opistognathus*: Opistognathidae) from the western Indian Ocean and Red Sea. *Smithiana* Bulletin 12: 39–54.

Smith-Vaniz WF, Bineesh KK, Akhilesh KV. 2012. *Opistognathus pardus*, a new species of jawfish (Teleostei: Opistognathidae) from the western Indian Ocean. Zootaxa 3523: 20–24. http://dx.doi.org/10.11646/zootaxa.3523.1.2

Smith-Vaniz WF, Johnson GD. 1990. Two new species of Acanthoclininae (Pisces: Plesiopidae) with a synopsis and phylogeny of the subfamily. *Proceedings of the Academy of Natural Sciences of Philadelphia* 142: 211–260. Smith-Vaniz WF, Johnson GD. 2016. Hidden diversity in deepwater bandfishes: review of *Owstonia* with descriptions of twenty-one new species (Teleostei: Cepolidae: Owstoniinae). *Zootaxa* 4187(1): 1–103. http://dx.doi. org/10.11646/zootaxa.4187.1.1

Smith-Vaniz WF, Johnson GD, Randall JE. 1988. Redescription of *Gracila albomarginata* (Fowler and Bean) and *Cephalopholis polleni* (Bleeker) with comments on the generic limits of selected Indo-Pacific groupers (Pisces: Serranidae: Epinephelinae). *Proceedings of the Academy of Natural Sciences of Philadelphia* 140(2): 1–23.

Smith-Vaniz WF, Randall JE. 1994. Scomber dentex Bloch & Schneider, 1801 (currently Caranx or Pseudocaranx dentex) and Caranx lugubris Poey, [1860] (Osteichthyes, Perciformes): proposed conservation of the specific names. Bulletin of Zoological Nomenclature 51(4): 323–329.

Smith-Vaniz WF, Rose JM. **2012**. *Adelotremus leptus*, a new genus and species of sabertooth blenny from the Red Sea (Teleostei: Blenniidae: Nemophini). *Zootaxa* 3249: 39–46. http://dx.doi.org/10.11646/zootaxa.3249.1.4

Smith-Vaniz WF, Satapoomin U, Allen GR. 2001. Meiacanthus urostigma, a new fangblenny from the northeastern Indian Ocean, with discussion and examples of mimicry in species of Meiacanthus (Teleostei: Blenniidae: Nemophini). aqua, Journal of Ichthyology and Aquatic Biology 5(1): 25–43.

Smith-Vaniz WF, Springer VG. **1971**. Synopsis of the Tribe Salariini, with description of five new genera and three new species (Pisces: Blenniidae). *Smithsonian Contributions to Zoology* 73: 1–72.

Smith-Vaniz WF, Staiger JC. 1973. Comparative revision of Scomberoides, Oligoplites, Parona, and Hypacanthus with comments on the phylogenetic position of Campogramma (Pisces: Carangidae). Proceedings of the California Academy of Sciences (Series 4) 39(13): 185–256.

Smitt FA. **1892**. *Skandinaviens Fiskar. Vol. 1*. P.A. Norstedt & Soners, Stockholm. 566 pp.

Smitt FA. 1900. Preliminary notes on the arrangement of the genus *Gobius*, with an enumeration of its European species. *Öfversigt af Kongliga Vetenskaps-Akademiens förhandlingar* 56(6) [for 1899]: 543–555.

Sneath PHA, Sokal RR. **1973**. *Numerical taxonomy: The principles and practice of numerical classification*. San Francisco: W.H. Freeman and Co. xvi + 573 pp.

Sneddon LU. **2003**. Trigeminal somatosensory innervation of the head of the rainbow trout with particular reference to nociception. *Brain Research* 972: 44–52.

Snodgrass RE, Heller E. 1905. Papers from the Hopkins-Stanford Galapagos Expedition, 1898–1899. XVII.
Shore fishes of the Revillagigedo, Clipperton, Cocos and Galapagos Islands. *Proceedings of the Washington Academy* of Sciences 6: 333–427. Snyder JO. **1904**. A catalogue of the shore fishes collected by the steamer *Albatross* about the Hawaiian Islands in 1902. *Bulletin of the U.S. Fish Commission* 22 [for 1902]: 513–538.

Snyder JO. **1908**. Descriptions of eighteen new species and two new genera of fishes from Japan and the Riu Kiu islands. *Proceedings of the United States National Museum* 35(1635): 93–111.

Snyder JO. **1909**. Descriptions of new genera and species of fishes from Japan and the Riu Kiu Islands. *Proceedings of the United States National Museum* 36(1688): 597–610.

Snyder JO. **1911**. Descriptions of new genera and species of fishes from Japan and the Riu Kiu Islands. *Proceedings of the United States National Museum* 40(1836): 525–549.

Soh CL. **1976**. Some aspects of the biology of *Siganus canaliculatus* (Park) 1797. PhD thesis, National University of Singapore. 293 pp.

Sommer C, Schneider W, Poutiers J-M. **1996**. *FAO species identification field guide for fishery purposes. The living marine resources of Somalia.* Rome: FAO. 376 pp.

Song HY, Mabuchi K, Satoh TP, Moore JA, Yamanoue Y, Miya M, Nishida M. 2014. Mitogenomic circumscription of a novel percomorph fish clade mainly comprising "Syngnathoidei" (Teleostei). *Gene* 542(2): 146–155. http://dx.doi.org/10.1016/j.gene.2014.03.040

Song YS, Kim J-K, Kang J-H, Kim SY. **2017**. Two new species of the genus *Atractoscion*, and resurrection of the species *Atractoscion atelodus* (Günther 1867) (Perciformes: Sciaenidae). *Zootaxa* 4306(2): 223–237. http://dx.doi.org/10.11646/zootaxa.4306.2.3

Sorbini L. **1981**. The cretaceous fishes of Nardò. 1. Order Gasterosteiformes (Pisces). *Bollettino del Museo Civico di Storia Naturale di Verona* 8: 1–27.

South JF. **1845**. *Thunnus* (pp. 620–622). In: Smedley E, Rose HJ, Rose HJ (eds) *Encyclopedia Metropolitana; or, Universal Dictionary of Knowledge*. Vol 25. London.

South African Museum. **1920**. *Report of the Trustees of the South African Museum for the Years Ended 31st December 1918, 1919*. Cape Town: Cape Times Ltd., Government Printers. [unpaginated]

Spalding MD, Fox HE, Halpern BS, McManus MA, Molnar J, Allen GR, Davidson N, Jorge ZA, Lombana AL, Lourie SA, Martin KD, McManus E, Recchia CA, Robertson J. 2007. Marine ecoregions of the world: a bioregionalization of coastal and shelf areas. *Bioscience* 57(7): 573–583.

Sparks JS. **2004**. A new and presumably extinct species of *Ptychochromoides* (Teleostei: Perciformes: Cichlidae) from central Madagascar. *Zootaxa* 524: 1–15.

Sparks JS. **2006**. A new species of ponyfish (Teleostei: Leiognathidae: *Photoplagios*) from Madagascar, with a phylogeny for *Photoplagios* and comments on the status of *Equula lineolata* Valenciennes. *American Museum Novitates* 3526: 1–20. Sparks JS, Baldwin ZH. **2012**. A new species of *Parapercis* (Teleostei: Pinguipedidae) from Madagascar. *Zootaxa* 3204: 31–39. http://dx.doi.org/10.11646/zootaxa.3204.1.3

Sparks JS, Chakrabarty P. **2007**. A new species of ponyfish (Teleostei: Leiognathidae: *Photoplagios*) from the Philippines. *Copeia* 2007(3): 622–629.

Sparks JS, Chakrabarty P. **2015**. Description of a new genus of ponyfishes (Teleostei: Leiognathidae), with a review of the current generic-level composition of the family, *Zootaxa* 3947(2): 181–190. http://dx.doi.org/10.11646/ zootaxa.3947.2.2

Sparks JS, Chakrabarty P. 2019. Description of a new species of ponyfish (Teleostei, Leiognathidae, Equulitini, *Photolateralis*) from the Gulf of Oman. *American Museum Novitates* 3929: 1–16. http://dx.doi.org/10.1206/3929.1

Sparks JS, Dunlap PV. 2004. A clade of non-sexually dimorphic ponyfishes (Teleostei: Perciformes: Leiognathidae): phylogeny, taxonomy, and description of a new species. *American Museum Novitates* 3459: 1–21.

Sparks JS, Dunlap PV, Smith WL. 2005. Evolution and diversification of a sexually dimorphic luminescent system in ponyfishes (Teleostei: Leiognathidae), including diagnoses for two new genera. *Cladistics* 21(4): 305–327.

Sparks JS, Nelson DW. **2004**. Review of the Malagasy sicydiine gobies (Teleostei: Gobiidae), with description of a new species and comments on the taxonomic status of *Gobius lagocephalus* Pallas, 1770. *American Museum Novitates* 3440: 1–20.

Sparks JS, Smith WL. 2004. Phylogeny and biogeography of the Malagasy and Australasian rainbowfishes (Teleostei: Melanotaenioidei): Gondwanan vicariance and evolution in freshwater. *Molecular Phylogenetics and Evolution* 33(3): 719–734.

Sprague J, Clements D, Conlin T, Edwards P, Frazer K, Schaper K, Segerdell E, Song P, Sprunger B, Westerfield M. **2003**. The Zebrafish Information Network (ZFIN): the zebrafish model organism database. *Nucleic Acids Research* 31(1): 241–243.

Springer S. 1950. A revision of north American sharks allied to the genus *Carcharhinus*. *American Museum Novitates* 1451: 1–13.

Springer S. **1965**. A review of western Atlantic cat sharks, Scyliorhinidae, with descriptions of a new genus and five new species. *Fishery Bulletin* 65(3): 581–624.

Springer S. **1968**. *Triakis fehlmanni*, a new shark from the coast of Somalia. *Proceedings of the Biological Society of Washington* 81: 613–624.

Springer S. **1979**. A revision of the catsharks, family Scyliorhinidae. *NOAA Technical Report* NMFS Circular 422: 1–147. Springer S, D'Aubrey JD. **1972**. Two new scyliorhinid sharks from the east coast of Africa with notes on related species. *Investigational Report. Oceanographic Research Institute* (Durban) 29: 1–19.

Springer S, Waller RA. **1969**. *Hexanchus vitulus*, a new sixgill shark from the Bahamas. *Bulletin of Marine Science* 19(1): 159–174.

Springer VG. **1963**. Two species of Indo-West Pacific blenniid fishes erroneously described from the western Atlantic Ocean. *Copeia* 1963(2): 452–454.

Springer VG. **1964**. A revision of the Carcharhinid shark genera *Scoliodon*, Loxodon, and Rhizoprionodon. *Proceedings of the United States National Museum* 115(3493): 559–632.

Springer VG. **1967**. Revision of the circumtropical shorefish genus *Entomacrodus* (Blenniidae: Salariinae). *Proceedings of the United States National Museum* 122(3582): 1–150.

Springer VG. **1968**. The Indo-Pacific blenniid fish genus *Stanulus*, with description of a new species from the Great Barrier Reef (Blenniidae; Blenniinae; Salariini). *Proceedings of the Biological Society of Washington* 81: 111–122.

Springer VG. 1971. Revision of the fish genus Ecsenius (Blenniidae, Blenniinae, Salariini). Smithsonian Contributions to Zoology 72: 1–74.

Springer VG. 1972. Synopsis of the tribe Omobranchini with descriptions of three new genera and two new species (Pisces: Blenniidae). *Smithsonian Contributions to Zoology* 130: 1–31.

Springer VG. **1976**. *Cirrisalarias bunares*, new genus and species of blenniid fish from the Indian Ocean. *Proceedings of the Biological Society of Washington* 89(13): 199–203.

Springer VG. **1981**. Notes on blenniid fishes of the tribe Omobranchini, with descriptions of two new species. *Proceedings of the Biological Society of Washington* 94(3): 699–707.

Springer VG. **1982**. Pacific Plate biogeography, with special reference to shorefishes. *Smithsonian Contributions to Zoology* 367: 1–182.

Springer VG. **1985**. *Oman ypsilon*, a new genus and species of blenniid fish from the Indian Ocean. Proceedings of the Biological Society of Washington 98(1): 90–97.

Springer VG. **1986**. Family No. 235: Blenniidae. In: Smith MM, Heemstra PC (eds) *Smiths' sea fishes*. Johannesburg: Macmillan South Africa. 1047 pp.

Springer VG. **1988**. The Indo-Pacific blenniid fish genus *Ecsenius*. *Smithsonian Contributions to Zoology* **4**65: 1–134.

Springer VG. 1988. Rotuma lewisi, new genus and species of fish from the southwest Pacific (Gobioidei, Xenisthmidae). Proceedings of the Biological Society of Washington 101(3): 530–539.

Springer VG. **1989**. *Ecsenius*: the world's most interesting genus of marine fishes. Part 1. *Tropical Fish Hobbyist* 37(8#398): 38–52.

Springer VG. **1989**. *Ecsenius*: the world's most interesting genus of marine fishes. Part 2. *Tropical Fish Hobbyist* 37(9#399): 50–61.

- Springer VG. 2001. Family Blenniidae. Blennies (combtooth and sabertooth blennies) (pp. 3538–3546). In: Carpenter KE, Niem VH (eds) FAO species identification guide for fishery purposes. The living marine resources of the western central Pacific. Vol. 6. Bony fishes part 4 (pp. 3381–4218). Rome: FAO.
- Springer VG, Allen GR. **2004**. *Ecsenius caeruliventris* and *E. shirleyae*, two new species of blenniid fishes from Indonesia, and new distribution records for other species of *Ecsenius*. *Zootaxa* 791: 1–12.

Springer VG, Bath H, Randall JE. 1998. Remarks on the species of the Indian Ocean fish genus *Alloblennius* Smith-Vaniz & Springer 1971, (Blenniidae). *aqua, Journal of Ichthyology and Aquatic Biology* 3(1): 19–24.

- Springer VG, Bauchot M-L. **1994**. Identification of the taxa Xenocephalidae, *Xenocephalus*, and *X. armatus* (Osteichthyes: Uranoscopidae). *Proceedings of the Biological Society of Washington* 107(1): 79–89.
- Springer VG, Fraser TH. **1976**. Synonymy of the fish families Cheilobranchidae (=Alabetidae) and Gobiesocidae, with descriptions of two new species of *Alabes*. *Smithsonian Contributions to Zoology* 234: 1–23.
- Springer VG, Fricke R. **2000**. Description of two new blenniid fish species: *Entomacrodus lemuria* from the western Indian Ocean and *E. williamsi* from the western Pacific Ocean. *Proceedings of the Biological Society of Washington* 113(2): 386–396.
- Springer VG, Gomon MF. **1975**. Revision of the blenniid fish genus *Omobranchus* with descriptions of three new species and notes on other species of the tribe Omobranchini. *Smithsonian Contributions to Zoology* 177: 1–135.
- Springer VG, Johnson GD. **2004**. Study of the dorsal gill-arch musculature of teleostome fishes, with special reference to the Actinopterygii. *Bulletin of the Biological Society of Washington* 11: 1–260.

Springer VG, Randall JE. **1974**. Two new species of the labrid fish genus *Cirrhilabrus* from the Red Sea. *Israel Journal of Zoology* 23(1): 45–54.

Springer VG, Randall JE. **1999**. *Ecsenius polystictus*, new species of blenniid fish from Mentawai Islands, Indonesia, with notes on other species of *Ecsenius*. *Revue française* d'Aquariologie Herpétologie 26(1–2): 39–48.

Springer VG, Smith CL, Fraser TH. **1977**. *Anisochromis straussi*, new species of protogynous hermaphroditic fish, and synonymy of Anisochromidae, Pseudoplesiopidae, and Pseudochromidae. *Smithsonian Contributions to Zoology* 252: 1–15.

Springer VG, Smith-Vaniz WF. **1968**. Systematics and distribution of the monotypic Indo-Pacific blenniid fish

genus Atrosalarias. Proceedings of the United States National Museum 124(3643): 1–12.

- Springer VG, Smith-Vaniz WF. **1972**. Mimetic relationships involving fishes of the family Blenniidae. *Smithsonian Contributions to Zoology* 112: 1–36.
- Springer VG, Spreitzer AE. **1978**. Five new species and a new genus of Indian Ocean blenniid fishes, tribe Salariini, with a key to the genera of the tribe. *Smithsonian Contributions to Zoology* 268: 1–20. https://doi.org/10.5479/si.00810282.268

Springer VG, Williams JT. 1994. The Indo-Pacific blenniid fish genus Istiblennius reappraised: a revision of Istiblennius, Blenniella, and Paralticus, new genus. Smithsonian Contributions to Zoology 565: 1–193.

Srinivasan KN, Sivaraja V, Huys I, Sasaki T, Cheng B, Kumar TKS, Sato K, Tytgat J, Yu C, San BCC, Ranganathan S, Bowie HJ, Kini RM, Gopalakrishnakone P. 2002. kappa-Hefutoxin1, a novel toxin from the scorpion *Heterometrus fulvipes* with uniquestructure and function. Importance of the functional diad in potassium channel selectivity. *Journal of Biological Chemistry* 277(33): 30040–30047.

Srinivasarengan S. 1981. Occurrence of a large shoal of Javanese cownose ray, *Rhinoptera javanica* Muller and Henle in the Bay of Bengal off Madras. *Indian Journal of Fisheries* 26(1&2): 239.

Srivastava GJ. 1967. A new species of freshwater fish of the genus *Hemiramphus* Cuv. from Gorakhpur, Uttar Pradesh, India. *Journal of the Bombay Natural History Society* 64(1): 93–94.

Srivastava GJ. **1968**. *Fishes of eastern Uttar Pradesh*. Varanasi, India: Vishwavidyalaya Prakashan. 163 pp.

- SSF (Smiths' Sea Fishes), *see* Smith JLB 1977; Smith MM, Heemstra PC (eds) 1986, 1988, 1995, 2003.
- Staiger JC. **1986**. Atlantic flyingfishes of the genus *Cypselurus*, with descriptions of the juveniles. *Bulletin of Marine Science* 15(3): 672–725.

Stahl BJ. 1999. Chondrichthyes III: Holocephali. In: Schultze H-P (ed) Handbook of paleoichthyology. Vol. 4. Munich: Verlag Dr. Friedrich Pfeil. 164 pp.

Starck WA II. 1969. Ecsenius (Anthiiblennius) midas, a new subgenus and species of mimic blenny from the western Indian Ocean. Notulae Naturae (Academy of Natural Sciences of Philadelphia) 419: 1–9.

Starks EC. **1911**. A possible line of descent of the gobioid fishes. *Science* 33(854): 747–748.

Starnes WC. 1988. Revision, phylogeny and biogeographic comments on the circumtropical marine percoid fish family Priacanthidae. *Bulletin of Marine Science* 43(2): 117–203.

Starnes WC. 1999. Priacanthidae. Bigeyes (pp. 2590–2601). In: Carpenter KE, Niem VH (eds) FAO species identification guide for fisheries purposes. The living marine resources of the western central Pacific. Vol. 4. Bony fishes part 2 (pp. 2069–2790). Rome: FAO. Stauch A. 1965. Caractères morphologiques et biogéographiques de deux *Heteromycteris a*fricains (Pisces, Teleostei, Heterosomata, Soleidae). *Cahiers ORSTOM* (Série Océanographie) 3(2): 5–10.

Stauch A. 1965. Sur la répartition géographique d'Arnoglossus imperialis (Raf. 1810) et description d'une espèce nouvelle, Arnoglossus blachei (Pisces: Teleostei, Heterosomata, Bothidae). Bulletin du Muséum National d'Histoire Naturelle (Paris) (Série 2) 37(2): 252–260.

Stauch A. 1967. Description d'une nouvelle espèce de Bothinae: Arnoglossus entomorhynchus n. sp. (Pisces: Teleostei, Heterosomata). Bulletin du Muséum National d'Histoire Naturelle (Paris) (Série 2) 39(4): 660–664.

Stauch A, Blache J. 1964. Contribution à la connaissance de genre *Ateleopus* Schlegel 1846 (Pisces, Teleostei, Ateleopoidei, Ateleopidae) dans l'Atlantique Oriental. *Cahiers ORSTOM* (Série Océanographie) 2(2): 47–54.

Stauch A, Blanc M. 1964. Dagetichthys lakdoensis n. g., n. sp., téléostéen pleuronectiforme du bassin de la Haute-Bénoué. Bulletin du Muséum National d'Histoire Naturelle (Série 2) 36(2): 172–177.

Stauch A, Cadenat J. 1965. Revision du genre *Psettodes* Bennett 1831 (Pisces, Teleostei, Heterosomata). Office de la recherche scientifique et technique Outre-Mer, Paris 3(4): 19–30.

Stead DG. 1907. Note on a small collection of fishes from Suwarow Island. Sydney: New South Wales Board of Fisheries. 9 pp.

Stebbins RC, Kalk M. **1961**. Observations on the natural history of the mud-skipper, *Periophthalmus sobrinus*. *Copeia* 1961(1): 18–27.

Steene RC. **1978**. *Butterfly and angelfishes of the world*. Vol. 1. New York: John Wiley & Sons. 144 pp.

Stehmann MFW. 1970. Vergleichend morphologische und anatomische Untersuchungen zur Neuordnung der Systematik der nordostatlantischen Rajidae (Chondrichthyes, Batoidei). Archiv für Fischereiwissenschaft 21(2): 73–163.

Stehmann MFW. 1976. Revision der Rajoiden-Arten des nördlichen Indischen Ozean und Indopazifik (Elasmobranchii, Batoidea, Rajiformes). *Beaufortia* 24(315): 133–175.

Stehmann MFW. 1986. Notes on the systematics of the rajid genus *Bathyraja* and its distribution in the world oceans (pp. 261–268). In: Uyeno T, Arai R, Taniuchi T, Matsuura K (eds) *Indo-Pacific fish biology: Proceedings of the Second International Conference on Indo-Pacific Fishes*. Ichthyological Society of Japan. 985 pp.

Stehmann MFW. 1990. Rhynchobatidae (p. 22), Rhinobatidae (pp. 23–27), Platyrhinidae (p. 28), Rajidae (pp. 29–50),
Pristidae (pp. 51–54). In: Quéro J-C, Hureau J-C, Karrer K, Post A, Saldanha L (eds) *Check-list of the fishes of the eastern tropical Atlantic (CLOFETA)*. Vol. 1. Lisbon: Junta Nacional de Investigação Científica e Tecnológica.

Stehmann MFW. **1995**. First and new records of skates (Chondrichthyes, Rajiformes, Rajidae) from the West African continental slope (Morocco to South Africa), with descriptions of two new species. *Archive of Fishery and Marine Research* 43(1): 1–119.

Stehmann MFW, Bürkel DL. 1984. Chimaeridae (pp. 212–215); Rhinochimaeridae (pp. 216–218).
In: Whitehead PJP, Bauchot M-L, Hureau J-C, Nielsen J, Tortonese E (eds) *Fishes of the north-eastern Atlantic and the Mediterranean*. Vol. I. Paris: UNESCO.

Stehmann MFW, Lenz W. 1973. Ergebnisse der Forschungsreisen des FFS Walther Herwig nach Südamerika. XXVI. Systematik und Verbreitung der Artengruppe — Seriolella punctata (Schneider, 1801), S. porosa Guichenot, 1848, S. dobula (Günther, 1869) sowie taxonomische Bemerkungen zu Hyperoglyphe Günther, 1859 und Schedophilus Cocco, 1839 (Osteichthyes, Stromateoidei, Centrolophidae). Archiv für Fischereiwissenschaft 23(3): 179–201.

Stein DL. 1979. The genus *Psednos* a junior synonym of *Paraliparis*, with a redescription of *Paraliparis micrurus* (Barnard) (Scorpaeniformes: Liparidae). *Matsya* 4: 5–10.

Steindachner F. 1861. Beiträge zur Kenntniss der Gobioiden. Sitzungsberichte der Mathematisch-Naturwissenschaftlichen Classe der Kaiserlichen Akademie der Wissenschaften 42(23): 283–292.

Steindachner F. 1861. Ichthyologische Mittheilungen (II). Verhandlungen der Kaiserlich-Königlichen zoologischbotanischen Gesellschaft in Wien 11: 133–144. Pl. 4.

Steindachner F. 1861. Ichthyologische Mittheilungen (III). Verhandlungen der Kaiserlich-Königlichen zoologischbotanischen Gesellschaft in Wien 11: 175–182. Pl. 5.

Steindachner F. 1862. Ichthyologische Mittheilungen (IV). Verhandlungen der KaiserlichKöniglichen zoologischbotanischen Gesellschaft in Wien 12: 497–504. Pl. 14.

Steindachner F. 1863. Ichthyologische Mittheilungen (V). Verhandlungen der KaiserlichKöniglichen zoologischbotanischen Gesellschaft in Wien 13: 1111–1114, Pls. 23–24.

Steindachner F. 1864. Ichthyologische Notizen. Anzeiger der Kaiserlichen Akademie der Wissenschaften, Mathematisch-Naturwissenschaftlichen Classe 1(5): 37–38.

Steindachner F. 1864. Serranus (Cerna) ongus Bloch, Günther (?), an Serranus angustifrons, n. sp. (pp. 230–231).
Ichthyologische Mittheilungen (VII). Verhandlungen der Kaiserlich-Königlichen zoologisch-botanischen Gesellschaft in Wien 14: 223–232. Pls. 7–8.

Steindachner F. 1866. Fortsetzung der ichthyologischen Notizen. Anzeiger der Kaiserlichen Akademie der Wissenschaften, Mathematisch-Naturwissenschaftlichen Classe 3(3): 19–20.

Steindachner F. **1866**. Ichthyologische Mittheilungen (IX). Verhandlungen der Kaiserlich-Königlichen zoologischbotanischen Gesellschaft in Wien 16: 761–796. Steindachner F. 1866. Üeber eine neue Mustelus-Art von Port Natal. Sitzungsberichte der Mathematisch-Naturwissenschaftlichen Classe der Kaiserlichen Akademie der Wissenschaften 53(1): 482–483.

Steindachner F. 1866. Zur Fischfauna von Port Jackson in Australien. Sitzungsberichte der Kaiserlichen Akademie der Wissenschaften, Mathematisch-Naturwissenschaftliche Classe 53: 424–481.

Steindachner F. 1867. Ichthyologische Notizen, vierte Folge. Anzeiger der Kaiserlichen Akademie der Wissenschaften, Mathematisch-Naturwissenschaftlichen Classe 4(8): 63–64.

Steindachner F. 1867. Ichthyologische Notizen (IV). Sitzungsberichte der Mathematisch-Naturwissenschaftlichen Classe der Kaiserlichen Akademie der Wissenschaften 55(1): 517–534.

Steindachner F. 1867. Ichthyologische Notizen (V). Über eine neue Scopelus- und Monacanthus-Art aus China (pp. 711–713). Sitzungsberichte der Mathematisch-Naturwissenschaftlichen Classe der Kaiserlichen Akademie der Wissenschaften 55: 701–717.

Steindachner F. 1867. Ichthyologische Notizen (V). Folge. Anzeiger der Kaiserlichen Akademie der Wissenschaften in Wien 4(14): 119–120.

Steindachner F. 1867. Über einige neue und seltene Meeresfische aus China. Sitzungsberichte der Mathematisch-Naturwissenschaftlichen Classe der Kaiserlichen Akademie der Wissenschaften 55(4–5): 585–592.

Steindachner F. 1868. Ichthyologische Notizen (VII). Sitzungsberichte der Kaiserlichen Akademie der Wissenschaften. Mathematisch-Naturwissenschaftliche Classe 57(1): 965–1008.

Steindachner F. **1869**. Eine Abhandlung über neue oder seltene Fische des Wiener-Museums, welche zum grössten Theile aus Mazatlan und China stammen. *Anzeiger der Akademie der Wissenschaften in Wien* 6(16): 125–126.

Steindachner F. 1869. Ichthyologische Notizen (VIII). Sitzungsberichte der Kaiserlichen Akademie der Wissenschaften. Mathematisch-Naturwissenschaftliche Classe 60(1): 120–139.

Steindachner F. 1870. Ichthyologische Notizen (X). Sitzungsberichte der Kaiserlichen Akademie der Wissenschaften. Mathematisch-Naturwissenschaftliche Classe 61(1): 623–642.

Steindachner F. 1870. Zur Fischfauna des Senegal (2 Abtheilung). Sitzungsberichte der Kaiserlichen Akademie der Wissenschaften. Mathematisch-Naturwissenschaftliche Classe 60(1): 945–995.

Steindachner F. 1876. Ichthyologische Beiträge (V). Sitzungsberichte der Kaiserlichen Akademie der Wissenschaften. Mathematisch-Naturwissenschaftliche Classe 74(1 Abtheilung): 49–240. Steindachner F. 1878. Ichthyologische Beiträge (VI). Sitzungsberichte der Kaiserlichen Akademie der Wissenschaften. Mathematisch-Naturwissenschaftliche Classe 77(1): 379–392.

Steindachner F. 1879. Ichthyologische Beiträge (VIII). Sitzungsberichte der Kaiserlichen Akademie der Wissenschaften. Mathematisch-Naturwissenschaften Classe 80(1–2): 119–191.

Steindachner F. 1880. Ichthyologische Beiträge (IX). 1.
Über eine Sammlung von Flussfischen von Tohizona auf Madagascar. II. Über zwei neue Agonus-Arten aus Californien. III. Über einige Fischarten aus dem nördlichen Japan, gesammelt vom Professor Dybowski. Sitzungsberichte der Kaiserlichen Akademie der Wissenschaften. Mathematisch-Naturwissenschaftliche Classe 82(1): 238–266.

Steindachner F. 1881. Ichthyologische Beiträge (X). Sitzungsberichte der Kaiserlichen Akademie der Wissenschaften. Mathematisch-Naturwissenschaftliche Classe 83(1 Abtheilung): 179–219.

Steindachner F. 1882. Beiträge zur Kenntniss der fische Afrika's. 1. Beitrag zur Kenntniss der Meeresfische Senegambiens. Denkschriften der Mathematisch-Naturwissenschaftlichen Classe der Kaiserlichen Akademie der Wissenschaften 44: 19–54.

Steindachner F. 1882. Ichthyologische Beiträge (XII). Sitzungsberichte der Kaiserlichen Akademie der Wissenschaften. Mathematisch-Naturwissenschaftliche Classe 86(1): 61–82.

Steindachner F. 1887. Ichthyologische Beiträge (XIV). Anzeiger der Kaiserlichen Akademie der Wissenschaften. Mathematisch-Naturwissenschaftliche Classe 24(19): 230–231.

Steindachner F. 1891. Ichthyologische Beiträge (XV). I. Über einige seltene und neue Fischarten aus dem canarischen Archipel. II. Über einige Characinen-Arten aus Südamerika. III. Pomacentrus grandidieri, n. sp. Sitzungsberichte der Kaiserlichen Akademie der Wissenschaften. Mathematisch-Naturwissenschaftliche Classe 100: 343–374.

Steindachner F. 1891. Über einige neue und seltene Fische von dem canarischen Archipel, aus den Flüssen Südamerika's und von Madagascar unter dem Titel: 'Ichthyologische Beiträge' (XV). Anzeiger der Akademie der Wissenschaften in Wien 28(18): 171–174.

Steindachner F. **1892**. Über einige neue und seltene Fischarten aus der ichthyologischen Sammlung des Kaiserlich-Königlichen Naturhistorischen Hofmuseums. *Anzeiger der Akademie der Wissenschaften in Wien* 29: 130–134.

Steindachner F. 1892. Über einige neue und seltene Fischarten aus der ichthyologischen sammlung des Kaiserlich-Königlichen Naturhistorischen Hofmuseums. Denkschriften der Kaiserlichen Akademie der Wissenschaften in Wien, Mathematisch-Naturwissenschaftliche Classe 59(1): 357–384. Steindachner F. 1893. Ichthyologische Beiträge (XVI). Sitzungsberichte der Kaiserlichen Akademie der Wissenschaften. Mathematisch-Naturwissenschaftliche Classe 102(1): 215–243.

Steindachner F. **1894**. Die Fische Liberia's. *Notes from the Leyden Museum* 16(1–2): 1–96.

Steindachner F. 1895. Vorläufige Mittheilung über einige neue Fischarten aus der ichthyologischen Sammlung des k. k. naturhistorischen Hofmuseums in Wien. Anzeiger der Kaiserlichen Akademie der Wissenschaften, Wien, Mathematisch-Naturwissenschaftliche Classe 32(18): 180–183.

Steindachner F. **1898**. Über einige neue Fischarten aus dem rothen Meere. *Anzeiger der Akademie der Wissenschaften in Wien* 35(19): 198–200.

Steindachner F. 1898. Über einige neue Fischarten aus dem rothen Meere, gesammelt während der I. und II. Österreichischen Expedition nach dem rothen Meere in dem Jahren 1895–1896 und 1897–1898. Sitzungsberichte der Kaiserlichen Akademie der Wissenschaften. Mathematisch-Naturwissenschaftliche Classe 107: 780–788.

Steindachner F. 1899. Berichtet über eine von Herrn Prof. O. Simony während der südarabischen Expedition in Sokotra entdeckte neue Sespina-Art, die zugleich einer besonderen Subgattung (*Hakaria*) angehört, und charakteristik dieselbe. Anzeiger der Akademie der Wissenschaften in Wien 36: 161–162.

Steindachner F. 1900. Fische aus dem Stillen Ocean. Ergebnisse einer Reise nach dem Pacific (Schauinsland, 1896–1897). Anzeiger der Akademie der Wissenschaften in Wien 37(16): 174–178.

Steindachner F. 1902. Über zwei neue Fischarten aus dem Rothen Meere. Anzeiger der Akademie der Wissenschaften in Wien 39(25): 336–338.

Steindachner F. 1902. Wissenschaftliche Ergebnisse der suedarabischen Expedition in den Jahren 1898 bis 1899.
Fische von Suedarabien und Socotra. Anzeiger der Kaiserlichen Akademie der Wissenschaften in Wien 39: 316–318.

Steindachner F. **1903**. Fische. *Abhandlungen herausgegeben von der Senckenbergischen Naturforschenden Gesellschaft* 25: 413–464.

Steindachner F. 1903. Fische aus Südarabien und Sokotra. Denkschriften der Kaiserlichen Akademie der Wissenschaften in Wien, Mathematisch-Naturwissenschaftliche Classe 71: 123–168.

Steindachner F. 1906. Zur Fischfauna der Samoa-Inseln. Sitzungsberichte der Kaiserlichen Akademie der Wissenschaften. Mathematisch-Naturwissenschaften Classe 115(1): 1369–1425. Steindachner F. **1907**. [Pomadasys stridens] Denkschriften der Kaiserlichen Akademie der Wissenschaften in Wien, Mathematisch-Naturwissenschaftliche Classe 71(1): p. 130.

Steindachner F, Döderlein L. 1883. Beiträge zur Kentniss der Fische Japan's. (I). Denkshriften der Kaiserlichen Akademie der Wissenchaften in Wien, Mathematisch-Naturwissenschaftlichen Classe 47: 211–242.

Steindachner F, Döderlein L. 1883. Beiträge zur Kenntniss der Fische Japan's. (II). Anzeiger der Kaiserlichen Akademie der Wissenschaften in Wien, Mathematisch-Naturwissenschaftliche Classe 20(15): 123–125.

Steindachner F, Döderlein L. **1884**. Beiträge zur Kentniss der fische Japan's. (III). *Denkshriften der Kaiserlichen Akademie der Wissenchaften in Wien, Mathematisch-Naturwissenschaftlichen Classe* 49: 171–212.

Steindachner F, Döderlein L. 1887. Beiträge zur Kentniss der Fische Japan's. (IV). Denkshriften der Kaiserlichen Akademie der Wissenchaften in Wien, Mathematisch-Naturwissenschaftlichen Classe 53: 257–296.

Steinitz H, Ben-Tuvia A. **1955**. Fishes from Eylath (Gulf of Aqaba), Red Sea. Second report. *Bulletin of the Sea Fisheries Research Station, Haifa* 11: 1–15.

Steinke D, Zemlak TS, Hebert PDN. **2009**. Barcoding Nemo: DNA-based identifications for the ornamental fish trade. *PLoS ONE* 4(7): e6300 (5 pp).

https://doi.org/10.1371/journal.pone.0006300

Stephens HR. 2011. Taxonomic revision of the flatfish genera *Zebrias* Jordan and Snyder, 1900 and *Pseudaesopia* Chabanaud, 1934 with Notes on *Aesopia* Kaup, 1858 (Pleuronectiformes: Soleidae). MSc thesis, University of Ottawa. 295 pp.

Stephenson AB. 1977. Metavelifer multiradiatus (Regan, 1907) (Pisces: Veliferidae) a new record from New Zealand waters. Records of the Auckland Institute and Museum 14: 143–144.

Stergiou KI. **1988**. Feeding habits of the Lessepsian migrant *Siganus luridus* in the eastern Mediterranean, its new environment. *Journal of Fish Biology* 33(4): 531–543.

Stern N, Goren M. 2013. First record of the moray eel *Gymnothorax reticularis*, Bloch, 1795 in the Mediterranean Sea, with a note on its taxonomy and distribution. *Zootaxa* 3641(2): 197–200. http://dx.doi.org/10.11646/ zootaxa.3641.2.8

Stevens JD. 1984. Life-history and ecology of sharks at Aldabra Atoll, Indian Ocean. *Proceedings of the Royal Society of London* B 222(1226): 79–106.

Stewart JD. 1999. A new genus of Saurodontidae (Teleostei: Ichthyodectiformes) from Upper Cretaceous rocks of the western interior of North America (pp. 335–360).
In: Arratia G, Schultze H-P (eds) *Mesozoic Fishes 2 – Systematics and fossil record. Proceedings of the international meeting, Buckow, 1997.* Munich: Verlag Dr. Friedrich Pfeil. 604 pp. Steyskal GC. **1980**. The grammar of family-group names as exemplified by those of fishes. *Proceedings of the Biological Society of Washington* 93(1): 168–177.

Stiassny MLJ. **1990**. Notes on the anatomy and relationships of the bedotiid fishes of Madagascar, with a taxonomic review of the genus *Rheocles* (Atherinomorpha: Bedotiidae). *American Museum Novitates* 2979: 1–33.

Stiassny MLJ. **1993**. What are grey mullets? *Bulletin of Marine Science* 52(1): 197–219.

Stiassny MLJ, Harrison IJ. 2000. Notes on a small collection of fishes from the Parc National de Marojejy, northeastern Madagascar, with a description of a new species of the endemic genus *Bedotia* (Atherinomorpha: Bedotiidae).
In: Goodman SM (ed) *A floral and faunal inventory of the Parc National de Marojejy, Madagascar: with reference to elevational variation. Fieldiana Zoology* (New Series) 97: 143–156.

Stiassny MLJ, Moore JA. **1992**. A review of the pelvic girdle of acanthomorph fishes, with comments on hypotheses of acantomorph intrarelationships. *Zoological Journal of the Linnean Society* 104(3): 209–242.

Stiassny MLJ, Parenti LR, Johnson GD (eds). 1996. Interrelationships of fishes. San Diego: Academic Press. xiii + 496 pp.

Stiassny MLJ, Raminosoa N. 1994. The fishes of the inland waters of Madagascar (pp. 133–148). In: Teugels GG, Guégan J-F, Albaret J-J (eds) Biological diversity of African fresh- and brackish water fishes. Geographical overviews presented at the PARADI Symposium, Senegal, 15–20 November 1993. *Musée Royal de l'Afrique Centrale, Annales Zoologie* 275. 177 pp.

Stinton FC. **1967**. The otoliths of the teleostean fish *Antigona capros* and their taxonomic significance. *Bocagiana* 13: 1–7.

Stobbs RE. 1980. Feeding habits of the giant clingfish Chorisochismus dentex (Pisces: Gobiesocidae). South African Journal of Zoology 15(3): 146–149.

Stock DW, Whitt GS. **1992**. Evidence from 18S ribosomal RNA sequences that lampreys and hagfishes form a natural group. *Science* 257(5071): 787–789.

Stone NR, Shimada K. 2019. Skeletal anatomy of the bigeye sand tiger shark, *Odontaspis noronhai* (Lamniformes: Odontaspididae), and its implications for lamniform phylogeny, taxonomy, and conservation biology. *Copeia* 107(4): 632–652. http://dx.doi.org/10.1643/CG-18-160

Storer DH. 1853. A history of the fishes of Massachusetts. Memoirs of the American Academy of Arts and Sciences (New Series) 5(1, art.8): 122–168, Pls. 9–16.

Storey MH. 1939. Contributions toward a revision of the ophichthyid eels. I. The genera *Callechelys* and *Bascanichthys*, with descriptions of new species and notes on *Myrichthys. Stanford Ichthyological Bulletin* 1(3): 61–84. Strahan R. 1975. *Eptatretus longipinnis*, n. sp., a new hagfish (family Eptatretidae) from south Australia, with a key to the 5–7 gilled Eptatretidae. *Australian Zoologist* 18(3): 137–148.

Stransky C, MacLellan SE. **2005**. Species separation and zoogeography of redfish and rockfish (genus *Sebastes*) by otolith shape analysis. *Canadian Journal of Fisheries and Aquatic Sciences* 62(1): 2265–2276.

Strasburg DW. **1960**. A new Hawaiian engraulid fish. *Pacific Science* 14(4): 395–399.

Strasburg DW. **1964**. Further notes on the identification and biology of echeneid fishes. *Pacific Science* 18(1): 51–57.

Strasburg DW. 1967. Observations on the biology of the lousefish, *Phtheirichthys lineatus* (Menzies). *Pacific Science* 21(2): 260–265.

Strasburg DW, Schultz LP. **1953**. The blenniid fish genera *Cirripectus* and *Exallias* with descriptions of two new species from the tropical Pacific. *Journal of the Washington Academy of Sciences* **43**(4): 128–135.

Straube N, Leslie RW, Clerkin PJ, Ebert DA, Rochel E, Corrigan SL, Chenhong L, Naylor GJP. 2015. On the occurrence of the southern lanternshark, *Etmopteus* granulosus, off South Africa, with comments on the validity of *E. compagnoi*. Deep Sea Research Part II: Topical Studies in Oceanography 115: 11–17. https://doi.org/10.1016/j. dsr2.2014.04.004

Straube N, Li C, Claes JM, Corrigan S, Naylor GJP. 2015. Molecular phylogeny of Squaliformes and first occurrence of bioluminescence in sharks. *BMC Evolutionary Biology* 15(art.162): 1–10. https://doi.org/10.1186/s12862-015-0446-6

Streelman JT, Alfaro M, Westneat MW, Bellwood DR, Karl SA. 2002. Evolutionary history of the parrotfishes: biogeography, ecomorphology, and comparative diversity. *Evolution* 56(5): 961–971.

Streets TH. 1877. Ichthyology (pp. 43–102). In: Contributions to the natural history of the Hawaiian and Fanning Islands and Lower California, made in connection with the United States North Pacific Surveying Expedition, 1873–75. Bulletin of the United States National Museum 1(7): 1–172.

Strömman PH. **1896**. *Leptocephalids in the University Zoological Museum at Upsala*. Upsala: Almqvist & Wiksell. 53 pp.

Stroud GJ. 1984. The taxonomy and biology of fishes of the genus *Parapercis* (Teleostei: Mugiloididae) in Great Barrier Reef waters. PhD dissertation, Department of Marine Biology, James Cook University, North Queensland.

Subba Rao DV, Al-Yamani, F. 2000. The Arabian Gulf (pp. 1–16). In: Sheppard CRC (ed) Seas at the Millennium: An environmental evaluation. Vol. 2. Oxford: Pergamon Press. 920 pp.

Suda Y, Tachikawa H, Baba O. **1986**. Adult form of the stromateoid fish, *Nomeus gronovii*, from the north Pacific. *Japanese Journal of Ichthyology* 33(3): 319–322.

Suda Y, Tominaga Y. **1983**. The percoid genus *Sphyraenops*, from the Pacific Ocean, with discussion on *Scombrosphyraena*. *Japanese Journal of Ichthyology* 30(3): 291–296.

Sugiyama K, Iwata A, Senou H, Yoshino T. 2000. First record of a goby, *Bathygobius niger* (Perciforms; Gobiidae) from western Pacific, and availability and validity of the scientific name. *I.O.P. Diving News* 11(9): 2–5. [In Japanese]

Sujatha K. **1995**. Finfish constituents of trawl by-catch off Visakhapatnam. *Fishery Technology* 32(1): 56–60.

Sujatha K. **2002**. Batoid fishes off Visakhapatnam, north east coast of India. *Journal of the Marine Biological Association of India* 44(1–2): 155–162.

Sulak KJ. 1984. Chlorophthalmidae (pp. 412–420). In: Whitehead PJP, Bauchot M-L, Hureau J-C, Nielsen J, Tortonese E (eds) *Fishes of the north-eastern Atlantic and the Mediterranean*. Vol. I. Paris: UNESCO.

Sultana M, Raman K, Abdul Kadir PM, Srinivasagam S, Rao GRM. **1980**. Observations on the biology of *Hemiramphus* gaimardi (Cuvier and Valenciennes) of Pulicat Lake. Journal of the Marine Biological Association of India 22(1&2): 89–109.

Sulya LL, Box BE, Gunter G. **1960**. Distribution of some blood constituents in fishes from the Gulf of Mexico. *American Journal of Physiology* **199**(6): 1177–1180.

Sumich JL. **1992**. *An introduction to the biology of marine life*. Dubuque, Iowa: Wm. C. Brown. 449 pp.

Sumith JA, Munkittrick KR, Athukorale N. **2011**. Fish assemblage structure of two contrasting stream catchments of the Mahaweli River Basin in Sri Lanka: hallmarks of human exploitation and implications for conservation. *Open Conservation Biology Journal* 5: 25–44. http://dx.doi. org/10.2174/1874839201105010025

Suresh GK, Thomas PA. **2007**. Three new records of apogonids (Order – Perciformes, Suborder – Percoidei) from the Indian seas. *Journal of the Marine Biological Association of India* 49(1): 105–108.

Suzuki T, Bogorodsky SY, Randall JE. **2012**. Gobiid fishes of the genus *Bryaninops* from the Red Sea, with description of two new species and two new records. *Zootaxa* 3170: 1–17. http://dx.doi.org/10.11646/zootaxa.3170.1.1

Suzuki T, Nakabo T. **1996**. Revision of the genus *Acanthaphritis* (Percophidae) with the description of a new species. *Ichthyological Research* **43**(4): 441–454.

Suzuki T, Senou H. **1999**. *Atrosalarias hosokawai*, a new species of blenny (Perciformes: Blenniidae) from the western Pacific. *Ichthyological Research* 46(3): 259–265.

Svetovidov AN. 1948. Gadiformes (Treskoobraznye). In: Pavlovskii EN, Shtakel'berg AA (eds) Fauna of the U.S.S.R.
Fishes. Zoological Institute of the Academy of Sciences of the U.S.S.R. 9(4): 221 pp. Svetovidov AN. **1949**. Fishes of Iran from the material collected by Academician F.H. Pavlovsky. *Trudy Instituta Okeanologii* 8: 859–869.

Svetovidov AN. **1986**. Review of the three-bearded rocklings of the genus *Gadropsarus* Rafinesque, 1810 (Gadidae) with description of a new species. *Journal of Ichthyology* 26(1): 114–135.

Swainson W. 1838. The natural history of fishes, amphibians and reptiles, or monocardian animals. Vol. 1. London: Longman, Orme, Brown, Green, & Longmans, and John Taylor. 368 pp.

Swainson W. 1839. On the natural history and classification of fishes, amphibians and reptiles Vol. 2. London: Longman, Orme, Brown, Green, & Longmans, and John Taylor. 452 pp.

## Т

Tabeta O, Ishida K. **1975**. Occurrence of the stromateoid fish *Ariomma brevimanus* in southern Japan. *Japanese Journal of Ichthyology* 22(3): 175–178.

Takata Y, Sasaki K. **2005**. Branchial structures in the Gasterosteiformes, with special reference to myology and phylogenetic implications. *Ichthyological Research* 52(1): 33–49.

Talbot FH. **1957**. The fishes of the genus *Lutianus* of the East African coast. *Annals and Magazine of Natural History* (Series 12) 10(112): 241–258.

Talbot FH. **1959**. On *Plectropomus maculatus* (Bloch) and *P. marmoratus* (n. sp.) from East Africa (Pisces: Serranidae). *Annals and Magazine of Natural History* (Series 13) 1(11) [for 1958]: 748–752.

Talbot FH. **1960**. Additions to the South African Museum collection of marine fishes. *Annals of the South African Museum* 45(2): 257–259.

Talbot FH. **1964**. The South African tunas. In: *Proceedings of the Symposium on Scombroid Fishes*, Marine Biological Association of India (MBAI), 12–15 January 1962, Mandapam. Part 1: 187–209.

Talbot FH. **1969**. The branchiostegid fish *Hoplolatilus fronticinctus* (Günther) from the Bay of Bengal. *Journal of the Marine Biological Association of India* 11(1&2): 309–310.

Talbot FH, Penrith MJ. **1961**. *Thunnus obesus* Lowe in South Africa. *South African Journal of Science* 57(9): 240.

Talbot FH, Penrith MJ. **1962**. Tunnies and marlins of South Africa. *Nature* (London) 193: 558–559.

Talbot FH, Penrith MJ. **1963**. Synopsis of biological data on species of the genus *Thunnus* (Sensu lato) (South Africa). *FAO Fisheries Biology Synopsis* 62: 608–646.

Talbot FH, Penrith MJ. **1968**. The tunas of the genus *Thunnus* in South African waters. Part I. Introduction, systematics, distribution and migrations. *Annals of the South African Museum* 52(1): 1–41. Talbot FH, Penrith M-L. **1965**. *Ctenogobius cloatus* Smith, 1960 a synonym of *Ctenogobius saldanha* (Barnard, 1927). *Annals of the South African Museum* 48(8): 189–193.

Talbot MMJ-F, Baird D. **1985**. Feeding of the estuarine round herring *Gilchristella aestuarius* (G & T) (Stolephoridae). *Journal of Experimental Marine Biology and Ecology* 87(3): 199–214.

Talwar PK. **1964**. Studies on the food and feeding relationships of the halfbeak fishes (Hemiramphidae) from the Gulf of Mannar and Palk Bay. *Indian Journal of Fisheries* 9(1A): 1–9.

Talwar PK. **1964**. A contribution to the biology of the halfbeak, *Hyporhamphus georgii* (Cuv. and Val.) (Hemirhamphidae). *Indian Journal of Fisheries*. 9(1A): 168–196.

Talwar PK. **1968**. Taxonomic status of *Sphyraena acutipinnis* Day, 1876. (Sphyraenidae, Pisces). *Journal of Natural History* 2(2): 197–200.

Talwar PK. **1968**. Studies on the biology of *Hemirhamphus marginatus* (Forskal) (Hemirhamphidae – Pisces). *Journal of the Marine Biological Association of India* 9(1): 61–69.

Talwar PK. 1971. Taxonomic position of *Corvina albida* Cuvier, 1830 [Pisces: Sciaenidae]. *Proceedings of the Zoological Society, Calcutta* 23(2) [for 1970]: 191–193.

Talwar PK. **1973**. A new bathypelagic fish, *Sphenanthias whiteheadi* (Pisces: Owstoniidae), from India. *Proceedings of the Zoological Society, Calcutta* 25(2) [for 1972]: 87–91.

Talwar PK. **1973**. A note on the systematics of the bathypelagic fish, *Mulichthys squamiceps* Lloyd. *Indian Journal of Animal Sciences* **43**(8): 799–801.

Talwar PK. **1973**. On some interesting bathypelagic fishes from the continental slope off the south west coast of India. *Proceedings of the Zoological Society, Calcutta* 26: 1–14.

Talwar PK. **1974**. On a new bathypelagic shark, *Scyliorhinus* (*Halaelurus*) *silasi* (Fam. Scyliorhinidae) from the Arabian Sea. *Journal of the Marine Biological Association of India* 14(2): 779–783.

Talwar PK. **1975**. On *Chrionema chryseres* Gilbert, a rare bathypelagic fish in the Indian Ocean. *Journal of the Bombay Natural History Society* 70(2): 390–392.

Talwar PK. **1976**. A contribution to the systematics of *Arius tenuispinis* Day, 1877 (Pisces: Ariidae). *Records of the Zoological Survey of India* 69: 291–294.

Talwar PK. **1976**. On a new bathypelagic fish, *Holanthias perumali* from the Arabian Sea, with a record of *Holanthias rhodopeplus* (Günther) from Indian Seas. *Journal of Natural History* 10(4): 361–365.

Talwar PK. **1977**. The rare deep-water scorpion-fish *Snyderina guentheri* in Indian seas. *Copeia* 1977(3): 580–581.

Talwar PK. **1980**. A new species of *Atrobucca* (Pisces: Sciaenidae) from the Arabian Sea. *Bulletin of the Zoological Survey of India* 3(1–2): 23–26. Talwar PK. **1981**.The electric rays of the genus *Heteronarce* Regan (Rajiformes: Torpedinidae), with the description of a new species. *Bulletin of the Zoological Survey of India* 3(3): 147–151.

Talwar PK. 1986. On a new bathypelagic fish, *Paracolopsis jonesi* (Pisces: Nemipteridae) from the Arabian Sea. In: James PSBR (ed) *Recent Advances in Marine Biology. Dr. S. Jones' 70<sup>th</sup> birthday commemoration volume*. New Delhi: Today's and Tomorrow's Publishers. i–xxx + 591 pp.

Talwar PK. **1995**. *Fauna of India and the adjacent countries*. *Pisces. Perciformes: Sciaenidae*. Kolkata, India: Zoological Survey of India. 144 pp.

Talwar PK, Jhingran AG. 1991. Inland fishes of India and adjacent countries (2 Volumes). New Delhi, Bombay, Calcutta: Oxford & IBH Publishing Co. 1158 pp.

Talwar PK, Jhingran AG. 1992. Inland fishes of India and adjacent countries. Vol. 2. Rotterdam: A.A. Balkema. pp. 543–1158. [reprint of Talwar & Jhingran 1991, see above]

Talwar PK, Kacker RK. **1984**. *Commercial sea fishes of India*. Calcutta: Zoological Survey of India. 997 pp.

Talwar PK, Mukherjee P. **1977**. A note on a new bathypelagic eel, *Ariosoma gnanadossi*, from the Bay of Bengal. *Indian Journal of Animal Sciences* 47(7): 432–434.

Talwar PK, Sen TK. **1971**. On some fishes of the Madras coast with description of a new species of the family Clinidae. *Records of the Zoological Survey of India* 65(1–4): 243–251.

Talwar PK, Whitehead PJP. **1971**. The clupeoid fishes described by Francis Day. *Bulletin of the British Museum (Natural History) Zoology* 22(2): 59–85.

Tan HH, Ng HH. **2000**. The catfishes (Teleostei: Siluriformes) of central Sumatra. *Journal of Natural History* 34(2): 267–303.

Tan SM, Yong LP, Senta T, Kuang HK. **1982**. *A colour guide to the fishes of the South China Sea and the Andaman Sea*. Marine Fisheries Research Department, SEAFDEC, Singapore. 54 pp.

Tanaka F, Iwatsuki Y. **2013**. *Rhabdosargus niger* (Perciformes: Sparidae), a new sparid species from Indonesia, with taxonomic status of the nominal species synonymized under *Rhabdosargus sarba*. *Ichthyological Research* 60(4): 343–352. http://dx.doi.org/10.1007/s10228-013-0360-0

Tanaka S. **1905**. On two new species of *Chimaera*. *Journal of the College of Science, Imperial University, Tokyo* 20(art.11): 1–14.

Tanaka S. **1908**. Notes on some Japanese fishes, with descriptions of fourteen new species. *Journal of the College of Science, Imperial University, Tokyo* 23(7): 1–54.

Tanaka S. **1911**. *Figures and descriptions of the fishes of Japan, including Riukiu Islands, Bonin Islands, Formosa, Kurile Islands, Korea and southern Sakhalin*. Vols. 1–4. Tokyo Printing Company. pp. 1–70, Pls. 1–20. [In Japanese and English]
Tanaka S. 1912. Figures and descriptions of the fishes of Japan, including the Riukiu Islands, Bonin Islands, Formosa, Kurile Islands, Korea, and southern Sakhalin. Vols. 5–10. Imperial University, Tokyo. pp. 71–186, Pls. 21–50. [In Japanese and English]

Tanaka S. **1913**. *Figures and descriptions of the fishes of Japan, including Riukiu Islands, Bonin Islands, Formosa, Kurile Islands, Korea and southern Sakhalin.* Vols. 11–14. Tokyo Printing Company. pp. 187–246, Pls. 51–70. [In Japanese and English]

Tanaka S. **1914**. *Figures and descriptions of the fishes of Japan, including Riukiu Islands, Bonin Islands, Formosa, Kurile Islands, Korea and southern Sakhalin.* Vols. 15–18. Tokyo Printing Company. pp. 247–318, Pls. 71–105. [In Japanese and English]

Tanaka S. 1915. Figures and descriptions of the fishes of Japan, including Riukiu Islands, Bonin Islands, Formosa, Kurile Islands, Korea and southern Sakhalin. Vols. 19–21: 319–382. Tokyo Printing Company. [In Japanese and English]

Tanaka S. 1915. Ten new species of Japanese fishes. Dobutsugaku Zasshi (Zoological Magazine Tokyo) 27(325): 565–568. [In Japanese]

Tanaka S. **1916**. Three new species of Japanese fishes. *Dobutsugaku Zasshi* (Zoological Magazine Tokyo) 28(330): 141–144. [In Japanese]

Tanaka S. 1916. Figures and descriptions of the fishes of Japan, including Riukiu Islands, Bonin Islands, Formosa, Kurile Islands, Korea and southern Sakhalin. Vols. 22–24: 383–440.
Tokyo Printing Company. [In Japanese and English]

Tanaka S. **1917**. Eleven new species of fishes of Japan. *Dobutsugaku Zasshi* (Zoological Magazine Tokyo) 29(339): 7–12. [In Japanese]

 Tanaka S. 1917. Figures and descriptions of the fishes of Japan, including Riukiu Islands, Bonin Islands, Formosa, Kurile Islands, Korea and southern Sakhalin. Vols. 25–26: 441–474. Tokyo Printing Company. [In Japanese and English]

Tanaka S. **1917**. Six new species of Japanese fishes. *Dobutsugaku Zasshi* (Zoological Magazine Tokyo) 29(340): 37–40. [In Japanese]

Tanaka S. 1917. Six new species of Japanese fishes. Dobutsugaku Zasshi (Zoological Magazine Tokyo) 29(345): 198–201. [In Japanese]

Tanaka S. 1917. Three new species of Japanese fishes.*Dobutsugaku Zasshi* (Zoological Magazine Tokyo) 29: 268–269. [In Japanese]

 Tanaka S. 1918. Figures and descriptions of the fishes of Japan, including Riukiu Islands, Bonin Islands, Formosa, Kurile Islands, Korea and southern Sakhalin. Vols. 27–30: 475–557.
 Tokyo Printing Company. [In Japanese and English]

Tanaka S. 1918. Twelve new species of fishes of Japan. Dobutsugaku Zasshi (Zoological Magazine Tokyo) 30(356): 223–227. [In Japanese] Tanaka Y, Ohyama T. **1991**. Reproductive behavior, egg and larval development of the hawkfishes, *Oxycirrhites typus* and *Cirrhitops hubbardi*, in the aquarium. *Bulletin of the Institute of Oceanic Research and Development, Tokai University* 11/12: 41–57.

Tanaka Y, Suzuki K. **1991**. Spawning, eggs and larvae of the hawkfish, *Cirrhitichthys aureus*, in an aquarium. *Japanese Journal of Ichthyology* 38(3): 283–288.

Taniuchi T, Yanagisawa F. **1987**. Albinism and lack of second dorsal fin in an adult tawny nurse shark, *Nebrius concolor*, from Japan. *Japanese Journal of Ichthyology* 34(3): 393–395.

Taquet M, Diringer A. 2007. Poissons de l'océan Indien et de la mer Rouge. Versailles, France: Éditions Quae. 527 pp. [In French]

Taquet M, Diringer A. 2012. Poissons de l'océan Indien et de la mer Rouge. Versailles, France. Éditions Quae. 679 pp. [In French]

Taquet M, Diringer A. 2013. Fishes of the Indian Ocean and Red Sea. Second edition. Versailles, France. Éditions Quae. 527 p.

Taschenberg O. **1883**. Beitrage zur Fauna der Insel Sokotra, vorzuglich nach dem von Dr. Emil Riebeck aus Halle a S. gesammelten Materiale zusammengestellt. *Zeitschrift für Naturwissenschaften* 56(2): 157–185.

Taverne L. **1973**. Les affinites systématique des téléosteens fossiles de l'ordre des Ichthyodectiformes. *Bulletin de l'Institut Royal des Sciences Naturelles de Belgique, Sciences de la Terre* 49(6): 1–11.

Taverne L. **1974**. Deuxième note sur la position systématique des téléostéens fossils de l'ordre des Ichthyodectiformes. *Bulletin de la Société Belge de Géologie* 83(1): 49–53.

Taverne L. **1975**. Considérations sur la position systématique des genres fossiles *Leptolepis* et *Allothrissops* au sein des téléostéens primitifs et sur l'origine et le polyphylétisme des poissons téléostéens. *Bulletin de la Classe des Sciences, Académie Royale de Belgique* (5<sup>ième</sup> Série) 61: 336–371.

Taverne L. 1979. Ostéologie, phylogénèse et systématique des téléostéens fossiles et actuels du super-ordre des ostéoglossomorphes. Troisiéme partie. Evolution des structures ostéologiques et conclusions générales relatives à la phylogénèse et à la systématique du super ordre. Addendum. Mémoires de l'Académie Royale des Sciences, des Lettres et des Beaux-Arts de Belgique (Classe des Sciences) (2<sup>ième</sup> série) 43(3): 1–268.

Taverne L. **1989**. *Crossognathus* Pictet, 1858 du Crétacé inférieur de l'Europe et systématique, paléozoogéographie et biologie des Crossognathiformes nov. ord. (Téléostéens) du Crétacé et du Tertiaire. *Palaeontographica* (Abteilung A Band A207 Lieferung 1–3) 207: 79–105.

Taylor EC. **1983**. Rabbitfishes for your marine tank. *Tropical Fish Hobbyist* 31(6#324): 8–14.

Taylor LR, Compagno LJV, Struhsaker PJ. **1983**. Megamouth – a new species, genus, and family of lamnoid shark (*Megachasma pelagios*, family Megachasmidae) from the Hawaiian Islands. *Proceedings of the California Academy of Sciences* (Series 4) 43(8): 87–110.

Taylor WR. **1964**. Fishes of Arnhem Land. *Records of the American-Australian scientific expedition to Arnhem Land* 4: 45–307.

Taylor WR. 1986. Ariidae (pp. 153–159). In: Daget J, Gosse
J-P, Thys van den Audenaerde DFE (eds) *Check-list of the freshwater fishes of Africa (CLOFFA)*. Vol. 2. Brussels: Institut Royal des Sciences Naturelles de Belgique. 521 pp.

Tea Y-K, Gill AC. 2016. Synchiropus sycorax, a new species of dragonet from the Philippines (Teleostei: Callionymidae). Zootaxa 4173(1): 85–93. http://dx.doi.org/10.11646/ zootaxa.4173.1.8

Tea Y-K, Pinheiro HT, Shepherd B, Rocha LA. **2019**. *Cirrhilabrus wakanda*, a new species of fairy wrasse from mesophotic ecosystems of Zanzibar, Tanzania, Africa (Teleostei, Labridae). *ZooKeys* 863: 85–96. https://doi. org/10.3897/zookeys.863.35580

Temminck CJ, Schlegel H. **1843–1850**. Pisces. In: *Fauna Japonica, sive descriptio animalium, quae in itinere per Japoniam suscepto annis 1823–30 collegit, notis observationibus et adumbrationibus illustravit Ph. Fr. de Siebold.* Batavia: Lugduni Batavorum. 324 pp.

Teng H-T. **1959**. Studies on the elasmobranch fishes from Formosa. Part 6. A new species of deep sea shark (*Centrophorus niaukang*) from Formosa. *Reports of the Laboratory of Fishery Biology of the Taiwan Fisheries Research Institute* 9: 1–6.

Teng H-T. 1962. Classification and distribution of the Chondrichthyes of Taiwan. Maizuru: Ogawa Press. 304 pp. [In Japanese]

Tennent JE. **1861**. *Sketches of the natural history of Ceylon, with narratives and anecdotes illustrative of the habits and instincts of the mammalia, birds, reptiles, fishes, insects, &c. including a monograph of the elephant and a description of the modes of capturing and training it.* London: Longman, Green, Longman, and Roberts. xxiv + 500 pp.

Terashima H, Mosaheb JI, Paupiah CN, Chineah V. 2001. *Field guide to coastal fishes of Mauritius*. Petite Riviere, Mauritius: Albion Fisheries Research Centre, Ministry of Fisheries. vii + 191 pp.

Tesch FW. **1977**. *The eel. Biology and management of anguillid eels*. London: Chapman & Hall. 434 pp.

Tesch FW. **2003**. *The eel* (5<sup>th</sup> edition). Oxford: Wiley-Blackwell. 416 pp.

Tesfamichael D, Rossing P, Saeed H. **2012**. Reconstruction of Yemen's catches in the Gulf of Aden from 1950–2010. Appendix B. In: Tesfamichael D, Pauly D (eds) Catch reconstruction for the Red Sea large marine ecosystem by countries (1950–2010). Fisheries Centre Research Reports (University of British Columbia) 20(1): 135–152.

Tessier E, Chabanet P, Pothin K, Soria M, Lasserre G. **2005**. Visual censuses of tropical fish aggregations on artificial reefs: slate versus video recording techniques. *Journal of Experimental Marine Biology and Ecology* **315**(1): 17–30.

Teugels GG, Janssens LJM, Bogaert J, Dumalin M. **1985**. Sur une collection de poissons de rivière des Comores. *Cybium* 9(1): 41–56.

Thies D, Leidner A. **2011**. Sharks and guitarfishes (Elasmobranchii) from the Late Jurassic of Europe. *Palaeodiversity* 4: 63–184.

Thiollière VJ de l'I. 1857. Essai sur le faune de l'île de Woodlark ou Moiou. Ichthyologie. Annales de la Société d'Agriculture, Histoire Naturelle et Arts Utiles de Lyon (Série 2) 8(1856): 417–504.

Thomas PA. **1969**. The goat-fishes (family Mullidae) of the Indian seas. *Memoirs of the Marine Biological Association of India* 3: 1–174.

Thominot A. **1878**. Sur un *Eleotris* d'espèce nouvelle. *Bulletin de la Société Philomathique de Paris* (Série 7) 2: 256.

Thominot A. **1886**. Sur quelques poissons nouveaux appartenant à la collection du Muséum d'Histoire Naturelle. *Bulletin de la Société Philomathique de Paris* (Série 7) 10(4): 161–171.

Thompson BA, Suttkus BA. **2002**. A revision of Indo-Pacific *Bembrops*, family Percophidae (suborder Trachinoidei). *Marine and Freshwater Research* 53(2): 283–295.

Thompson W. **1837**. Notes relating to the natural history of Ireland, with a description of a new genus of fishes *(Echiodon)*. *Proceedings of the Zoological Society of London* Part 5(54): 52–63.

Thompson W. **1840**. On a new genus of fishes from India. *Magazine of Natural History* 4(6): 184–187.

Thompson WW. **1914**. I. Catalogue of fishes of the Cape Province. *Marine Biological Report* 2: 132–167.

Thompson WW. **1916**. II. Catalogue of fishes of the Cape Province. *Marine Biological Report* 3: 69–135.

Thompson WW. **1918**. III. Catalogue of fishes of the Cape Province. *Marine Biological Report* 4: 75–177.

Thomson DA, Findley LT, Kerstitch AN. **1979**. *Reef fishes of the Sea of Cortez: The rocky-shore fishes of the Gulf of California*. Tucson: University of Arizona. 302 pp.

Thomson JM. **1954**. The Mugilidae of Australia and adjacent seas. *Australian Journal of Marine and Freshwater Research* 5(1): 70–131.

Thomson JM. **1974**. Mugilidae. In: Fischer W, Whitehead PJP (eds) FAO species identification sheets for fishery purposes. Eastern Indian Ocean (Fishing Area 57) and western central Pacific (Fishing Area 71). Vol. 3. Bony fishes [Families M– Sci]. Rome: FAO. [unpaginated] Thomson JM. **1997**. The Mugilidae of the world. *Memoirs of the Queensland Museum* 41(3): 457–562.

Thomson JM, Luther G. 1984. Mugilidae. Mullets. In: Fischer W, Bianchi G (eds) FAO species identification sheets for fishery purposes. Western Indian Ocean (Fishing Area 51).Vol. 3. Rome: FAO. [unpaginated]

Thorson G. **1971**. *Life in the sea*. New York, Toronto: World University Library, McGraw-Hill Book Company. 257 pp. [Translated from Danish by MC Meilgaard and A Laurie]

Thresher RE. **1984**. *Reproduction in reef fishes*. Neptune City, New Jersey: TFH Publications. 399 pp.

Thunberg CP. **1787–1821**. *Museum naturalium Academiae Upsaliensis. ... Praeside*. CP Thunberg, etc. 33 pts. Upsaliae.

Thunberg CP. **1791**. Tvänne utländska fiskar, beskrifne. *Kongliga Vetenskaps Akademiens nya Handlingar* (Stockholm) 12(7–9): 190–192.

Thunberg CP. **1792**. Åtskillige förut okánde Fiskar af abborslágtet. *Kongliga Vetenskaps Akademiens nya Handlingar* (Stockholm) 13: 141–143.

Thunberg CP. **1793**. Beskrifning på nya fiskar utaf abborrslägtet ifrån Japan. *Kongliga Vetenskaps Academiens Handlingar* (Stockholm) 14: 55–56, Pl. 1.

Thunberg CP. **1793**. Beskrifning på nya fisk-arter utaf abborslägtet ifrån Japan. *Kongliga Vetenskaps Academiens Handlingar* (Stockholm) 14: 198–200, Pl. 7.

Thunberg CP. **1793**. Sidsta fortsåttningen af beskrifningen på nya fiskarter utaf abbor-slägtet ifrån Japan. *Kongliga Vetenskaps Academiens Handlingar* (Stockholm) 14: 296–298, Pl. 9.

Thys TM, Whitney J, Hearn A, Weng KC, Peñaherrera C, Jawad LA, Alfaro-Shigueto J, Mangel JC, Karl SA. **2013**. First record of the southern ocean sunfish, *Mola ramsayi*, in the Galápagos Marine Reserve. *Marine Biodiversity Records* 6(e70): 1–4. https://doi.org/10.1017/S1755267213000377

Tibbetts IR, Collette BB, Isaac R, Kreiter P. **2007**. Functional and phylogenetic implications of the vesicular swimbladder of *Hemiramphus* and *Oxyporhamphus convexus* (Beloniformes: Teleostei). *Copeia* 2007(4): 808–817.

Tiberi C, Ebinger C, Ballu V, Stuart G, Oluma B. **2005**. Inverse models of gravity data from the Red Sea-Aden-East African rifts triple junction zone. *Geophysical Journal International* 163: 775–787.

Tickell SR. **1865**. Description of a supposed new genus of the Gadidae, Arakan. *Journal of the Asiatic Society of Bengal* 34(2): 32–33.

Tighe KA. **1992**. *Boehlkenchelys longidentata*, a new genus and species of chlopsid eel (Teleostei: Anguilliformes) from the Indo-West Pacific region. *Proceedings of the Biological Society of Washington* 105(1): 19–22.

Tilesius WG von, see Tilesius von Tilenau WG.

Tilesius von Tilenau WG. **1812**. Abbildungen und Beschreibungen einiger Fische aus Japan und einiger

Mollusken aus Brasilien, welche bey Gelegenheit der ersten Russischen Kaiserlichen Erdumseglung lebendig beobachtet wurden. *Denkschriften der Bayerischen Akademie der Wissenschaften.* 38 pp.

Tilesius von Tilenau WG. **1814**. Atlas zur Reise um die Welt, unternommen auf Befehl Seiner Kaiserlichen Majestät Alexander des Ersten auf den Schiffen Nadeshda und Neva unter dem Commando des Captains von Krusenstern. Atlas. St. Petersburg, Pls. 1–106.

Tilesius von Tilenau WG. **1820**. De piscium Australium novo genere icone illustrato. *Mémoires de l'Académie Impériale des Sciences de St. Petersbourg* 7: 301–310.

Tillier J-B. **1902**. Le Canal de Suez et sa faune ichthyologique. *Mémoires de la Société zoologique de France* 15: 279–320.

Tilney RL, Hecht T. **1990**. The food and feeding habits of two co-occurring marine catfish, *Galeichthys feliceps* and *G. ater* (Osteichthyes: Ariidae) along the south-east coast of South Africa. *Journal of Zoology* 221(2): 171–193.

Tinker SW. **1944**. *Hawaiian fishes: a handbook of the fishes found among the islands of the central Pacific Ocean.* Honolulu: Tongg Publishing. 404 pp.

Tinker SW. **1978**. *Fishes of Hawaii: a handbook of the marine fishes of Hawaii and the central Pacific Ocean.* Honolulu: Hawaiian Service. 532 pp.

Tinley KL. 1971. Determinants of coastal conservation dynamic and diversity of the environment as exemplified by the Mozambican coast (pp. 125–153). In: *Nature conservation as a form of land use*. SARRCUS Proceedings, Gorongosa National Park, Mozambique, 13–17 September 1971. Pretoria: Southern African Regional Commission for the Conservation and Utilization of the Soil. 153 pp.

 Tobias WJ. 1976. Ecology of Siganus argenteus (Pisces: Siganidae) in relation to its mariculture potential on Guam. University of Guam Marine Laboratory Technical Report 29: 58–93.

Tomczak M, Godfrey JS. **2003** *Regional oceanography: An introduction* (2<sup>nd</sup> edition). Delhi: Daya Publishing House. 390 pp.

Tominaga Y. **1963**. A revision of the fishes of the family Pempheridae of Japan. *Journal of the Faculty of Science, University of Tokyo, Section IV Zoology* 10(pt 1): 269–290.

Tominaga Y. **1968**. Internal morphology, mutual relationships and systematic position of the fishes belonging to the family Pempheridae. *Japanese Journal of Ichthyology* 15(2): 43–95.

Tominaga Y, Sakamoto K, Matsuura K. **1996**. Posterior extension of the swimbladder in percoid fishes, with a literature survey of other teleosts. *Bulletin of the University Museum, University of Tokyo* 36: 1–73.

Tomiyama I, Abe T. **1953–1958**. *Figures and descriptions of the fishes of Japan (a continuation of Dr. Shigeho Tanaka's work)*. Vols. 49–59. Tokyo: Kazama Shobo. pp. 961–1247.

Topp RW, Girardin DL. **1971**. An adult louvar, *Luvarus imperialis* (Pisces, Luvaridae) from the Gulf of Mexico. *Copeia* 1971(1): 181–182.

Torchio M. **1973**. Soleidae (pp. 628–634). In: Hureau J-C, Monod T (eds). *Check-list of the fishes of the north-eastern Atlantic and of the Mediterranean (CLOFNAM)*. Vol. I. Paris: UNESCO.

Torii A, Harold AS, Ozawa T. 2003. Redescription of type specimens of three *Bregmaceros* species (Gadiformes: Bregmacerotidae): *B. bathymaster, B. rarisquamosus*, and *B. cayorum. Memoirs of the Faculty of Fisheries, Kagoshima University* 52: 23–32.

Torii A, Harold AS, Ozawa T, Iwatsuki Y. **2003**. Redescription of *Bregmaceros mcclellandi* Thompson, 1840 (Gadiformes: Bregmacerotidae). *Ichthyological Research* 50(2): 129–139.

Tortonese E. **1936**. Un nuovo Percoide dell'Oceano Indiano (*Hapalogenys pictus*, n. sp.). *Bollettino dei Musei Zoologia ed Anatomia Comparata della R. Università di Torino* (Series 3) 45(67): 1–4.

Tortonese E. **1937**. Pesci del mar Rosso. *Bollettino dei Musei di Zoologia ed Anatomia Comparata della R. Università di Torino* (Series 3) 45(1935–36): 153–218.

Tortonese E. **1941**. Pesci marini della Somalia Italiana raccolti dal Marchese Negrotto Cambiaso. *Atti della Reale Academia Ligure di Scienze e Lettres* 1(1): 1–12.

Tortonese E. **1948**. Ricerche zoologiche nel Canale de Suez e dintorni. II. Pesci. *Archivio Zoologico Italiano* 33: 275–292.

Tortonese E. **1952**. Un Percoide marino e batifilo nuova per l'ittiofauna italiana (*Epigonus denticulatus* Dieuz.). *Bollettino di Pesca, Piscicoltura e Idrobiologia* (n.s.) 7(1): 72–74.

Tortonese E. **1953**. Su alcuni pesci Indo-Pacifici immigrati nel Mediterraneo orientale. *Bolletino di Zoologia* 20(4–6): 73–81. https://doi.org/10.1080/11250005309436878

 Tortonese E. 1956. Spedizione subacquea italiana nel Mar Rosso. Ricerche zoologiche. IV. Plagiostomi. VI. Plettognati.

Rivista di Biologia Coloniale 14: 5–21, 73–86.

Tortonese E. **1968**. Fishes from Eilat (Red Sea). *Bulletin of the Sea Fisheries Research Station, Haifa* 51: 6–30.

Tortonese E. **1970**. *Osteichthyes (Pesci ossei), Parte Prima*. Fauna d'Italia, 10. Bologna: Calderini. xiii + 565 pp.

Tortonese E. **1976**. Gobioid fishes from the Gulf of Aden. *Monitore Zoologica Italiano (N.S.) Supplemento* 7(4): 187–193.

Tortonese E. **1980**. Poissons observés près de la Côte Arabe de la Mer Rouge (Arabie Saoudite). *Cybium* (Série 3) 4(9): 61–68.

Tortonese E. **1983**. List of fishes observed near Jeddah (Saudi Arabia). *Journal of the Faculty of Marine Science* 3: 105–110.

Tortonese E. 1990. Molidae (pp. 1077–1079). In: Quéro J-C, Hureau J-C, Karrer K, Post A, Saldanha L (eds) *Check-list* of the fishes of the eastern tropical Atlantic (CLOFETA).
Vol. 2. Lisbon: Junta Nacional de Investigação Científica e Tecnológica. Tortonese E, Casanova Queirolo L. **1970**. Contributo allo studio dell'ittiofauna del Mar Ligure orientale. *Annali del Museo civico di storia naturale "Giacomo Doria"* 78: 21–46.

Tretjakoff D. **1926**. Die Wirbelsäule des Neunauges. *Anatomische Anzeiger* 61: 387–396.

Trewavas E. **1964**. The sciaenid fishes with a single mental barbel. *Copeia* 1964(1): 107–117.

Trewavas E. **1971**. The syntypes of the sciaenid *Corvina albida* Cuvier and the status of *Dendrophysa hooghliensis* Sinha and Rao and *Nibea coibor* (nec. Hamilton) of Chu, Lo & Wu. *Journal of Fish Biology* 3(4): 453–461.

Trewavas E. 1973. Sciaenidae (pp. 396–401); Mugilidae (pp. 567–574). In: Hureau J-C, Monod T (eds) Checklist of the fishes of the north-eastern Atlantic and of the Mediterranean (CLOFNAM). Vol. I. Paris: UNESCO.

Trewavas E. 1974. Sciaenidae, Otolithes cuvieri, Sciaen Otol 1. In: Fischer W, Whitehead PJP (eds) FAO species identification sheets for fishery purposes. Eastern Indian Ocean (Fishing Area 57) and western central Pacific (Fishing Area 71). Vol. 3. Bony fishes [Families M–Sci]. Rome: FAO. [unpaginated]

Trewavas E. **1977**. The sciaenid fishes (croakers or drums) of the Indo-West-Pacific. *Transactions of the Zoological Society of London* 33(4): 253–541, Pls. 1–14.

Trewavas E, Ingham SE. **1972**. A key to the species of Mugilidae (pisces) in the northeastern Atlantic, with explanatory notes. *Journal of Zoology* 167(1): 15–29.

Trinajstic K, Boisvert C, Long J, Maksimenko A, Johanson Z. 2015. Pelvic and reproductive structures in placoderms (stem gnathostomes). *Biological Reviews of the Cambridge Philosophical Society* 90(2): 467–501. http://dx.doi.org/10.1111/brv.12118

Troschel FH. **1840**. Ueber einige Bloch'sche Fisch-Arten. *Archiv für Naturgeschichte* 6(pt 1): 267–281.

Trott LB. **1970**. Contributions to the biology of carapid fishes (Paracanthopterygii: Gadiformes). *University of California Publications in Zoology* 89: 1–60.

Trott LB. **1981**. A general review of the pearlfishes (Pisces, Carapidae). *Bulletin of Marine Science* **31**(3): 623–629.

Trott LB, Trott EE. **1972**. Pearlfishes (Carapidae: Gadiformes) collected from Puerto Galera, Mindoro, Philippines. *Copeia* 1972(4): 839–843.

Trunov IA. **1971**. Occurrence of *Lamprogrammus exutus* Nyb. et Poll (fam. Brotulidae) at the coasts of South West Africa. *Voprosy Ikhtiologii* 11(4): 733–734. [in Russian]

Trunov IA. **1972**. Some fish species rare and new to the southeastern Atlantic. *Voprosy Ikhtiologii* 12(3): 439–444. [In Russian]

Trunov IA. **1975**. Notes on the suborder Stromateoidei in the southeastern Atlantic. *Voprosy Ikhtiologii* 15(3): 390–401.

Trunov IA. **1976**. New species and species recorded for the first time in the pelagic area of the tropical Atlantic of the families Serranidae, Emmelichthyidae and Ariommidae. *Journal of Ichthyology* 16(2): 229–238.

Trunov IA. **1982**. Species of the family Regalecidae (Lampridiformes) from the southeastern Atlantic Ocean. *Journal of Ichthyology* 22(1): 1–6.

Trunov IA. 2006. Ichthyofauna of seamounts around the island of Ascension and St. Helena Island (Atlantic Ocean). *Voprosy Ikhtiologii* 46(4): 471–477. [In Russian, English translation in *Journal of Ichthyology* 46(7): 493–499.]

Trunov IA, Kukuev EI. **1982**. In: Trunov IA, Zeiformes of the Thalassobathyal of the southeastern Atlantic. *Bulletin of the Moscow Society of Naturalists Biological Series* 87(2): 41–53. [In Russian]

Trunov IA, Malevanyy. **1974**. On the ecology of searobins (Triglidae) off Namibia. *Journal of Ichthyology* 14(3): 367–373.

Tsuda RT, Bryan PG. **1973**. Food preference of juvenile *Siganus rostratus* and *S. spinus* in Guam. *Copeia* 1973(3): 604–606.

Tsuda RT, Bryan PG, Fitzgerald WJ, Tobias WJ. 1976. Juvenileadult rearing of *Siganus* (Pisces: Siganidae) in Guam. University of Guam Marine Laboratory Technical Report 29: 19–26.

Tsuneki K, Ouji M, Saito H. **1983**. Seasonal migration and gonadal changes in the hagfish *Eptatretus burgeri*. *Japanese Journal of Ichthyology* 29(4): 429–440.

Tucker DW. 1953. The fishes of the genus Benthodesmus (family Trichiuridae). Proceedings of the Zoological Society of London 123(1): 171–195.

Tucker DW. **1954**. Report on the fishes collected by S.Y. *Rosaura* in the north and central Atlantic, 1937–38. *Bulletin of the British Museum (Natural History) Zoology* 2(6): 163–214.

Tucker DW. **1956**. Studies on the trichiuroid fishes. 3. A preliminary revision of the family Trichiuridae. *Bulletin of the British Museum (Natural History) Zoology* 4(3): 163–214.

Turan C, Gunduz I, Gurlek M, Yaglioglu D, Erguen D. 2009. Systematics of Scorpaeniformes species in the Mediterranean Sea inferred from mitochondrial 16S rDNA sequence and morphological data. *Folia Biologica* (Kraków) 57(3–4): 219– 226. http://dx.doi.org/10.3409/fb57\_1-2.219-226

Turner J, Jago C, Daby D, Klaus R. 2000. The Mascarene region (pp. 253–268). In: Sheppard CRC (ed) Seas at the Millennium: An environmental evaluation. Vol. 2. Oxford: Pergamon Press. 920 pp.

Turner W. **1867**. On a remarkable mode of gestation in an undescribed species of *Arius (A. boakeii). Journal of Anatomy and Physiology* 1(1): 78–82. Turon JM, Rucabado J, Lloris D, Macpherson E. 1986. Datos pess de las expediciones realizadas en aguas de Namibia durante los Anos 1981 a 1984. In: Macpherson E (ed) Resultados de las Expediciones Oceanográfico-Pesqueras "Benguela III" (1981) a "Benguela VII" (1984) y "Valdivia I" realizadas en el Atlántico Sudoriental (Namibia). Vol. 17. Barcelona: Datos Informativos, Instituto de Ciencias del Mar. 345 pp.

Tutman PJ, Freyhof J, Dulčić J, Glamuzina B, Geiger MF. 2017. Lampetra soljani, a new brook lamprey from the southern Adriatic Sea basin (Petromyzontiformes: Petromyzontidae). Zootaxa 4273 (no. 4): 531–548. http://dx.doi.org/10.11646/ zootaxa.4273.4.4

Tweddle D, Anderson ME. **2008**. A collection of marine fishes from Angola, with notes on new distribution records. *Smithiana* Bulletin 8: 3–24.

Tyler EHM, Speight MR, Henderson P, Manica A. **2009**. Evidence for a depth refuge effect in artisanal coral reef fisheries. *Biological Conservation* 142(3): 652–667. http://dx.doi.org/10.1016/j.biocon.2008.11.017

Tyler JC. **1965**. The trunkfish genus *Acanthostracion* (Ostraciontidae, Plectognath) in the Western Atlantic: two species rather than one. *Proceedings of the Academy of Natural Sciences of Philadelphia* 117(1): 1–18.

Tyler JC. **1966**. A new species of serranoid fish of the family Anthiidae from the Indian Ocean. *Notulae Naturae* (Academy of Natural Sciences of Philadelphia) 389: 1–6.

Tyler JC. **1966**. A new species of damselfish (Pomacentridae) from the western Indian Ocean, *Pristotis judithae*. *Notulae Naturae* (Academy of Natural Sciences of Philadelphia) 393: 1–6.

Tyler JC. 1966. Bathyphylax omen, a new species of triacanthodid plectognath fish from the Indian Ocean. Notulae Naturae (Academy of Natural Sciences of Philadelphia) 395: 1–5.

Tyler JC. **1967**. A diagnosis of the two transversely barred Indo-Pacific pufferfishes of the genus *Canthigaster* (*valentini* and *coronatus*). *Proceedings of the Academy of Natural Sciences of Philadelphia* 119(2): 53–73.

Tyler JC. **1967**. A redescription of *Triodon macropterus* Lesson, a phyletically important plectognath fish. *Proceedings of the Koninklijke Nederlandse Academie van Wetenschappen* (Series C, Biological and Medical Sciences) 70(1): 84–96.

Tyler JC. **1968**. A monograph on plectognath fishes of the superfamily Triacanthoidea. *Monographs of the Academy of Natural Sciences of Philadelphia* 16: 1–364.

Tyler JC. **1970**. A redescription of the inquiline carapid fish *Onuxodon parvibrachium*, with a discussion of the skull structure and the host. *Bulletin of Marine Science* 20(1): 148–164.

Tyler JC. **1970**. New records of triacanthoid plectognath fishes. *Notulae Naturae* (Academy of Natural Sciences of Philadelphia) 435: 1–7.

Tyler JC. **1971**. Habitat preferences of the fishes that dwell in shrub corals on the Great Barrier Reef. *Proceedings of the Academy of Natural Sciences Philadelphia* 123: 1–26.

Tyler JC. **1980**. Osteology, phylogeny and higher classification of the fishes of the order Plectognathi (Tetraodontiformes). *NOAA Technical Report* NMFS Circular 434: 1–422.

Tyler JC. **1983**. Records of fishes of the family Triacanthodidae (Tetradontiformes) from the western Indian Ocean off East Africa. *J.L.B. Smith Institute of Ichthyology Special Publication* 31: 1–13.

Tyler JC. **1997**. New species of *Paratriacanthodes* spikefish (Triacanthodidae: Tetraodontiformes) from the South China Sea. *Proceedings of the Biological Society of Washington* 110(2): 310–313.

Tyler JC, Johnson GD, Nakamura I, Collette BB. **1989**. Morphology of *Luvarus imperialis* (Luvaridae), with a phylogenetic analysis of the Acanthuroidei (Pisces). *Smithsonian Contributions to Zoology* 485: i–vi + 1–78.

Tyler JC, O'Toole B, Winterbottom R. **2003**. Phylogeny of the genera and families of zeiform fishes, with comments on their relationships with tetraodontiforms and caproids. *Smithsonian Contributions to Zoology* 618: i–iv + 1–110.

Tyler JC, Paxton JR. **1979**. New genus and species of pufferfish (Tetraodontidae) from Norfolk Island, southwest Pacific. *Bulletin of Marine Science* 29(2): 202–215.

Tyler JC, Sorbini C. **1999**. Phylogeny of the fossil and recent genera of fishes of the family Scatophagidae (Squamipinnes). *Bollettino del Museo Civico di Storia Naturale di Verona* 23: 353–393.

# U

Uchida RN. **1981**. Synopsis of biological data on frigate tuna, *Auxis thazard*, and bullet tuna, *A. rochei. NOAA Technical Report* NMFS Circular 436: 1–63.

Ueyanagi S, Wares PG. **1975**. Synopsis of biological data on striped marlin *Tetrapterus audax* (Phillippi), 1887. *NOAA Technical Report* NMFS SSRF-675 (Part 3): 132–159.

Uiblein F. 1991. Ontogenetic shifts in resource use and shoaling tendency related to body size in Red Sea goatfish (*Parupeneus forsskali*, Mullidae). *Marine Ecology* 12(2): 153–161.

Uiblein F. **1995**. Morphological variability between populations of *Neobythites stefanovi* (Pisces: Ophidiidae) from the deep Red Sea and the Gulf of Aden. *Marine Ecology Progress Series* 124: 23–29.

Uiblein F. **2007**. Goatfishes (Mullidae) as indicators in tropical and temperate coastal habitat monitoring and management. *Marine Biology Research* 3(5): 275–288.

Uiblein F. **2011**. Taxonomic review of Western Indian Ocean goatfishes of the genus *Mulloidichthys* (Family Mullidae), with description of a new species and remarks on colour and body form variation in Indo-West Pacific species. *Smithiana* Bulletin 13: 51–73.

Uiblein F. **2021**. Taxonomic review of the "*posteli-species* group" of goatfishes (genus *Parupeneus*, Mullidae), with description of a new species from the northern Red Sea. *Cybium* 45(1): 63–77. https://doi.org/10.26028/cybium/2021-451-008

Uiblein F, Causse R. **2013**. A new deep-water goatfish of the genus *Upeneus* (Mullidae) from Vanuatu, South Pacific. *Zootaxa* 3666(3): 337–344. http://dx.doi.org/10.11646/ zootaxa.3666.3.4

Uiblein F, Gledhill D. **2015**. A new goatfish of the genus *Upeneus* (Mullidae) from Australia and Vanuatu, with interand intraspecific comparisons. *Marine Biology Research* 11: 475–491. [online in 2014] https://doi.org/10.1080/17451 000.2014.958088

Uiblein F, Gledhill DC, Pavlov DA, Hoang TA, Shaheen S. 2019. Three new goatfishes of the genus *Upeneus* (Mullidae) from the Indo-Pacific, with a redescription of colour patterns in *U. margarethae. Zootaxa* 4683(2): 151–196. https://doi.org/10.11646/zootaxa.4683.2.1

Uiblein F, Gledhill DC, Peristiwady T. **2017**. Two new goatfishes of the genus *Upeneus* (Mullidae) from Australia and Indonesia. *Zootaxa* 4318(2): 295–311. http://dx.doi. org/10.11646/zootaxa.4318.2.4

Uiblein F, Gouws G. **2014**. A new goatfish species of the genus *Upeneus* (Mullidae) based on molecular and morphological screening and subsequent taxonomic analysis. *Marine Biology Research* 10(7): 655–681. https://doi.org/10.1080/17 451000.2013.850515

Uiblein F, Gouws G. **2015**. Distinction and relatedness – taxonomic and genetic studies reveal a new species group of goatfishes (*Upeneus*; Mullidae). *Marine Biology Research* 11(10): 1021–1042. http://dx.doi.org/10.1080/17451000.20 15.1064963

Uiblein F, Gouws G, Gledhill DC, Stone K. 2016. Just off the beach: intrageneric distinctiveness of the bandtail goatfish *Upeneus taeniopterus* (Mullidae) based on a comprehensive alpha taxonomy and barcoding approach. *Marine Biology Research* 12(7): 675–694. http://dx.doi.org/10.1080/1745100 0.2016.1190458

Uiblein F, Gouws G, Johnson JW, Lobel PS, Shaheen S. **2020**. Junior synonymy of *Mulloides armatus* and intraspecific comparisons of the yellowstripe goatfish *Mulloidichthys flavolineatus* (Mullidae) using a comprehensive alphataxonomy approach. *Cybium* 44(2): 137–155. https://doi. org/10.26028/cybium/2020-442-005

Uiblein F, Gouws G, Lisher M, Maluene BS. **2020**. *Upeneus floros*, a new goatfish from South Africa and Mozambique,

with updated taxonomic accounts for *U. guttatus* and *U. pori* and a key to Western Indian Ocean *Upeneus* species (Mullidae). *Zootaxa* 4834(4): 523–555. https://doi. org/10.11646/zootaxa.4834.4.3

- Uiblein F, Heemstra PC. 2010. A taxonomic review of the western Indian Ocean goatfishes of the genus *Upeneus* (Family Mullidae), with descriptions of four new species. *Smithiana* Bulletin 11: 35–71.
- Uiblein F, Heemstra PC. 2011. A new goatfish, Upeneus seychellensis sp. nov. (Mullidae), from the Seychelles Bank, with remarks on Upeneus guttatus and a key to Western Indian Ocean Upeneus species. Marine Biology Research 7(7): 637–550. https://doi.org/10.1080/17451000.2010.547202
- Uiblein F, Heemstra PC. **2011**. Description of a new goatfish species, *Upeneus randalli* n. sp. (Mullidae), from the Persian Gulf, with remarks and identification keys for the genus *Upeneus. Scientia Marina* 75(3): 585–594. http://dx.doi.org/10.3989/scimar.2011.75n3585
- Uiblein F, Hoang TA, Alama U, Causse R, Chacate OE, Fahmi, Garibay S, Matiku P. **2018**. A new species and new records of goatfishes of the genus *Parupeneus* (Mullidae) from the Indian Ocean, with updated occurrence information for *P. jansenii* in the Western Pacific. *Cybium* 42(3): 229–256. https://doi.org/10.26028/10.26028/cybium/2018-423-002
- Uiblein F, Hoang TA, Gledhill D. 2017. Redescription and new records of Jansen's goatfish, *P. jansenii* (Mullidae), from the Western Pacific and Eastern Indian Ocean. *Zootaxa* 4344(3): 541–559. http://dx.doi.org/10.11646/ zootaxa.4344.3.6
- Uiblein F, Lisher M. 2013. A new goatfish of the genus Upeneus (Mullidae) from Angoche, northern Mozambique. Zootaxa 3717(1): 85–95. https://doi.org/10.11646/zootaxa.3717.1.7
- Uiblein F, McGrouther M. **2012**. A new deep-water goatfish of the genus *Upeneus* (Mullidae) from northern Australia and the Philippines, with a taxonomic account of *U. subvittatus* and remarks on *U. mascareinsis. Zootaxa* 3550: 61–70. http://dx.doi.org/10.11646/zootaxa.3550.1.4
- Uiblein F, Nielsen JG. **2018**. Review of the *steatiticus*-species group of the cuskeel genus *Neobythites* (Ophidiidae) from the Indo-Pacific, with description of two new species. *Zootaxa* 4387(1): 157–173. http://dx.doi.org/10.11646/ zootaxa.4387.1.7
- Uiblein F, Nielsen, JG. **2019**. Redescription of the ocellusbearing cuskeel *Neobythites kenyaensis* (Ophidiidae), with new Southeast African records and remarks on intraspecific morphological and colour variation. *Cybium* 43(1): 109– 116. https://doi.org/10.26028/cybium/2019-431-011
- Uiblein F, White W. **2015**. A new goatfish of the genus *Upeneus* (Mullidae) from Lombok, Indonesia and first verified record of *U. asymmetricus* for the Indian Ocean. *Zootaxa* 3980: 51–66. https://doi.org/10.11646/zootaxa.3980.1.3

- UNEP/Nairobi Convention Secretariat. **2009**. *Transboundary diagnostic analysis of land-based sources and activities in the Western Indian Ocean*. United Nations Environment Programme (UNEP)/Nairobi Convention Secretariat and Western Indian Ocean Marine Sciences Association (WIOMSA). 355 pp. [http://www.unep.org/ nairobiconvention]
- Ünlü E, Balci K, Meriç N. **2000**. Aspects of the biology of *Liza abu* (Mugilidae) in the Tigris River (Turkey). *Cybium* 241(1): 27–43.
- Uwa H. 1986. Karyotype evolution and geographical distribution in the ricefish, genus *Oryzias* (Oryziidae) (pp. 867–876). In: Uyeno T, Arai R, Taniuchi T, Matsuura K (eds) *Indo-Pacific fish biology: Proceedings of the Second International Conference on Indo-Pacific Fishes*. Tokyo: Ichthyological Society of Japan. 985 pp.
- Uwa H, Iwamatsu T, Saxena OP. **1983**. Karyotype and cellular DNA content of the Indian ricefish, *Oryzias melastigma*. *Proceedings of the Japan Academy* (Series B) 59(3): 43–47.
- Uwa H, Parenti LR. **1988**. Morphometric and meristic variation in ricefishes, genus *Oryzias*: a comparison with cytogenetic data. *Japanese Journal of Ichthyology* 35(2): 159–166.
- Uyeno T, Arai R, Taniuchi T, Matsuura K (eds). **1986**. *Indo-Pacific fish biology: Proceedings of the Second International Conference on Indo-Pacific Fishes*. Tokyo: Ichthyological Society of Japan. xii + 985 pp.
- Uyeno T, Matsuura K, Fujii E (eds). **1983**. *Fishes trawled off Suriname and French Guiana*. Japan Marine Fishery Resource Research Center. 519 pp.

# V

- Vachon J, Chapleau F, Desoutter-Meniger M. 2008. Révision taxinomique du genre Solea et réhabilitation du genre Barnardichthys (Soleidae; Pleuronectiformes). Cybium 32(1): 9–26. https://doi.org/10.26028/cybium/2008-321-002
- Vachon J, Desoutter M, Chapleau F. 2005. Solea bleekeri Boulenger, 1898, a junior synonym of *Pegusa nasuta* (Pallas,1814), with the recognition and redescription of *Solea turbynei* Gilchrist, 1904 (Pleuronectiformes: Soleidae). *Cybium* 29(4): 315–319.
- Vahl M. 1794. Beskrivelse af en nye fiske-slaegt, *Caecula*. *Skrivter af Naturhistorie-Selskabet Kiøbenhavn* 3(2): 149–156. [In Danish, English translation in Smith JLB. 1965. *Annals and Magazine of Natural History* (Series 13) 7(84): 711–723.]
- Vahl M. **1797**. Beskrivelse tvende nye arter af Bredflab-slaegten *Lophius. Skrivter af Naturhhistorie-Selskabet Kiøbenhavn* 4: 212–216.
- Vaillant LL. **1873**. Sur le pretendu *Serranus phaeton*. *Bulletin de la Société Philomatique de Paris* (Série 10) 6: 94.

Vaillant LL. **1877**. Sur une espèce nouvelle du genre Cheilodipterus. *Bulletin de la Société Philomathique de Paris* (6<sup>th</sup> Série) 12: 27–30.

Vaillant LL. 1888. Poissons. In: Bouvier E-L, Gravier C, Milne-Edwards A, Perrier E (eds) *Expeditions scientifiques du* Travailleur *et du* Talisman *pendant les années 1880, 1881, 1882, 1883*. Paris: G. Masson. 406 pp.

Vaillant LL, Bocourt F. 1874–1915. Études sur les poissons. In: Hamy ET (ed) *Mission scientifique au Mexique et dans l'Amérique centrale. Recherches zoologiques*. Quatrième partie. Imprimerie Nationale, Paris. Part 4: 1–265, Pls. 20.

Vaillant LL, Sauvage HE. 1875. Note sur quelques espèces nouvelles de poissons des îles Sandwich. *Revue et Magasin de Zoologie* (Série 3) 3: 278–287.

Valenciennes A. **1832**. Descriptions de plusieurs espèces nouvelles de poissons du genre *Apogon*. *Nouvelles Annales du Muséum d'Histoire Naturelle, Paris* 1: 51–59.

Valenciennes A. 1837–1844. Ichthyologie des îles Canaries, ou histoire naturelle des poissons rapportés par Webb & Berthelot. In: Webb PB, Berthelot S (eds) *Histoire naturelle des îles Canaries*. Paris, 1835–1850. Vol. 2(pt 2): 1–109, Pls. 26.

Valenciennes A. 1841. In: Müller J, Henle FGJ. 1838–1841.

Valenciennes A. **1855**. Ichthyologie (pp. 297–351). In: du Petit-Thouars A (ed) *Voyage autour du monde sur la frégate* La Vénus *pendant les années 1836–1839: mammifères, oiseaux, reptiles, et poissons*. Paris: Gide et J. Baudry. 351 pp.

Valenciennes A. 1862. Description de quelques espèces nouvelles de poissons envoyées de Bourbon par M. Morel, directeur du Muséum d'Histoire naturelle de cette île. *Comptes Rendus Hebdomadaires des séances de l'Académie des Sciences* 54: 1165–1170.

Van Damme K, Banfield L. 2011. Past and present human impacts on the biodiversity of Socotra Island (Yemen): implications for future conservation. *Zoology in the Middle East* Supplementum 3: 31–88. https://doi.org/10.1080/09397 140.2011.10648899

Vandelli D. **1797**. Florae, et faunae Lusitanicae specimen. *Memorias de agriculture: premiados pela Academia Real das Sciencias de Lisboa* 1: 37–79.

Van der Elst R. 1976. Game fish of the east coast of southern Africa. I. The biology of the elf, *Pomatomus saltatrix* (Linnaeus), in the coastal waters of Natal. *Investigational Report. Oceanographic Research Institute* (Durban) 44: 1–59.

Van der Elst R. **1981**. *A guide to the common sea fishes of southern Africa* (1<sup>st</sup> edition). Cape Town: Struik. 367 pp.

Van der Elst R. 1985. A guide to the common sea fishes of southern Africa (1<sup>st</sup> edition; softcover). Cape Town: Struik. 367 pp.

Van der Elst R. **1988**. *A guide to the common sea fishes of southern Africa* (2<sup>nd</sup> edition). Cape Town: Struik. 398 pp.

Van der Elst R. **1993**. A guide to the common sea fishes of southern Africa (3<sup>rd</sup> edition). Cape Town: Struik. 398 pp.

Van der Elst R, Collette BB. **1984**. Game fishes of the east coast of southern Africa. 2. Biology and systematics of the queen mackerel *Scomberomorus plurilineatus*. *Investigational Report*. *Oceanographic Research Institute* (Durban) 55: 1–12.

Van der Elst RP, Everett BI (eds). **2015**. Offshore fisheries of the Southwest Indian Ocean: their status and the impact on vulnerable species. *Oceanographic Research Institute, Special Publication* 10: 448 pp.

Van der Elst RP, King D. 1990. Everyone's guide to sea fishes of southern Africa. Johannesburg: Central News Agency. 112 pp.

Van der Elst RP, Vermeulen R. **1986**. *Sharks and stingrays*. Cape Town: Struik Publishers. 64 pp.

Van der Elst RP, Wallace JH. 1976. Identification of the juvenile mullet of the east coast of South Africa. *Journal of Fish Biology* 9(4): 371–374.

Van der Laan R, Eschmeyer WN, Fricke R. **2014**. Family-group names of recent fishes. *Zootaxa* 3882(1): 1–230. https://doi. org/10.11646/zootaxa.3882.1.1

Van der Lingen CD. **2002**. Diet of sardine *Sardinops sagax* in the southern Benguela upwelling ecosystem. *South African Journal of Marine Science* 24: 301–316.

Van der Straten KM, Collette BB, Leung LK-P, Johnston SD. 2006. Sperm morphology of the black marlin (*Makaira indica*) differs from scombroid sperm. *Bulletin of Marine Science* 79(3): 839–845.

Van Hasselt JC. 1823. Uittreksel uit een' brief van Dr. J.C. van Hasselt, aan den Heer C.J. Temminck, geschreven uit Tjecande, Residentie Bantam, den 28<sup>sten</sup> December 1822. *Algemeene Konst- en Letter-bode voor het Jaar II* 35: 130–133.

Van Hasselt JC. 1824. Extrait d'une seconde lettre sur les poissons de Java, écrite par M. van Hasselt à M. C.J. Temminck, datée de Tjecande, résidence de Bantam, 29 Décembre 1822. *Bulletin des Sciences Naturelles et de Géologie* (Férussac), Paris 2: 374–377.

Van Oijen MJP. **2001**. *Squalus edwardsii* (currently *Haploblepharus edwardsii*; Chondrichthyes, Carcharhiniformes): proposed attribution to Schinz (1822) and conservation of *edwardsii* as the correct original spelling. *Bulletin of Zoological Nomenclature* 58(4): 294–296.

Van Rampelbergh M, Fleitmann D, Verheyden S, Cheng H, Edwards L, De Geest P, De Vleeschouwer D, Burns SJ, Matter A, Claeys P, Keppens E. 2013. Mid- to late-Holocene Indian Ocean Monsoon variability recorded in four speleothems from Socotra Island, Yemen. *Quaternary Science Reviews* 65: 129–142. http://dx.doi.org/10.1016/j. quascirev.2013.01.016

Vari RP. 1978. The terapon perches (Percoidei, Teraponidae). A cladistic analysis and taxonomic revision. *Bulletin of the American Museum of Natural History* 159(art.5): 179–340. Vari RP. **1992**. Redescription of *Mesopristes elongatus* (Guichenot, 1866), an endemic Malagasy fish species (Pisces, Terapontidae). *American Museum Novitates* 3039: 1–7.

Vari RP. 2001. Family Terapontidae (= Therapontidae, Theraponidae, Teraponidae). Terapon-perches (terapongrunters) (pp. 3305–3316). In: Carpenter KE, Niem VH (eds) FAO species identification guide for fishery purposes. The living marine resources of the western central Pacific. Vol. 5. Bony fishes part 3 (pp. 2791–3380). Rome: FAO.

Vasil'eva ED. **2007**. *Fish of the Black Sea. Key to marine, brackish-water, euryhaline, and anadromous species with color illustrations, collected by S.V. Bogorodsky.* Moscow: VNIRO Publishing. 237 pp.

Vasiliu GD. **1931**. Beschreibung der Ceylon-Fische aus der Sammlung Plate. *Jenaische Zeitschrift für Naturwissenschaft* 65(2): 319–360, Pls. 7–10.

Vassilev MV, Pehlivanov LZ. **2005**. Checklist of Bulgarian freshwater fishes. *Acta Zoologica Bulgarica* 57(2): 161–190.

Veldhuis M, Kraay G, van Bleijswijk J, Baars M. 1997.
Seasonal and spatial variability in phytoplankton biomass, productivity and growth in the northwestern Indian
Ocean: the southwest and northeast monsoon, 1992–1993.
Deep Sea Research Part I: Oceanographic Research Papers 44(4): 425–449.

Venkataraman K. **2003**. *Checklist of Marine Fauna of Tamil Nadu*. Chennai: Marine Biological Station, Zoological Survey of India, Department of Environment, Government of Tamil-Nadu. 88 pp.

Venkataraman K, Srinivasan M, Satyanarayana Ch, Prabakar D. 2002. Faunal diversity of Gulf of Mannar Biosphere Reserve. *Zoological Survey of India* (Conservation Area Series) 15: 1–77.

Venu S, Madhusoodana Kurup B. 2003. Distribution and biology of the deep-sea fish *Psenopsis cyanea* (Alcock) inhabiting continental slope of the west coast of India. *Journal of the Marine Biological Association of India* 44(1&2) [for 2002]: 176–186.

Veron JEN, Turak E. 2005. Zooxanthellate scleractinia of Madagascar (pp. 23–25). In: McKenna SA, Allen GR (eds) A rapid marine biodiversity assessment of the coral reefs of northwest Madagascar. *Bulletin of the Rapid Assessment Program* 31. Washington, DC: Conservation International. 124 pp.

Viana ST de F, Carvalho MR de. **2016**. Redescription of *Squalus acutipinnis* Regan 1908, a valid species of spiny dogfish from southern Africa (Chondrichthyes: Squaliformes: Squalidae). *Copeia* 104(2): 539–553. https://doi.org/10.1643/CI-14-217

Victor BC. **2015**. How many coral reef fish species are there? Cryptic diversity and the new molecular taxonomy (pp. 76–87). In: Mora C (ed) *Ecology of fishes on coral reefs*. Cambridge, UK: Cambridge University Press. 388 pp. http://dx.doi.org/10.1017/CBO9781316105412.010

Victor BC. 2016. Halichoeres gurrobyi, a new labrid fish (Teleostei: Labridae) from Mauritius in the southwestern Indian Ocean, with a review of the *H. zeylonicus* species complex. Journal of the Ocean Science Foundation. 22: 10–27. https://doi.org/10.5281/zenodo.57088

Victor BC. **2016**. Two new species in the spike-fin fairy-wrasse species complex (Teleostei: Labridae: *Cirrhilabrus*) from the Indian Ocean. *Journal of the Ocean Science Foundation* 23: 21–50. https://doi.org/10.5281/zenodo.163217

Victor BC, Edward JMB. **2016**. *Pseudojuloides labyrinthus*, a new labrid fish (Teleostei: Labridae) from the western Indian Ocean. *Journal of the Ocean Science Foundation* 21: 58–70. https://doi.org/10.5281/zenodo.55594

Victor BC, Randall JE. 2014. Pseudojuloides edwardi, n. sp. (Perciformes: Labridae): an example of evolution of maledisplay phenotype outpacing divergence in mitochondrial genotype. Journal of the Ocean Science Foundation 11: 1–12.

Vijay Anand PE, Pillai NGK. **2007**. Coral reef fish abundance and diversity of seagrass beds in Kavaratti atoll, Lakshadweep, India. *Indian Journal of Fisheries* 54(1): 11–20.

Vijayaraghavan P. **1973**. Studies on fish eggs and larvae from Indian waters. I. Development of egg and larvae of *Hirundichthys (Hirundichthys) coromandelensis* (Hornell). *Indian Journal of Fisheries* 20(1): 108–137.

Vilasri V. 2013. Comparative anatomy and phylogenetic systematics of the family Uranoscopidae (Actinopterygii: Perciformes). *Memoirs of the Faculty of Fisheries Hokkaido* University 55(1/2): 1–106. http://hdl.handle.net/2115/53996

Viljoen A, Cyrus DP. **2002**. Temporal variation in the recruitment of larval fish into the Mhlathuze Estuary in northern KwaZulu-Natal, South Africa. *Marine and Freshwater Research* 53(2): 439–445.

Villanueva R. **1993**. Diet and mandibular growth of *Octopus magnificus* (Cephalopoda). *South African Journal of Marine Science* 13(1): 121–126.

Vincent A, Foster SJ, Koldewey HJ. **2011**. Conservation and management of seahorses and other Syngnathidae. *Journal of Fish Biology* 78(6): 1681–1724. http://dx.doi.org/10.1111/ j.1095-8649.2011.03003.x

Vine NG. **1998**. Aspects of the biology of the doublesash butterflyfish, *Chaetodon marleyi* (Pisces: Chaetodontidae). Masters thesis, Rhodes University, South Africa. 108 pp.

Vineesh N, Mohitha C, Bineesh KK, Kumar KG, Gopalakrishnan A, Basheer VS. 2014. DNA barcoding and taxonomic notes on the genus *Macolor* (Perciformes: Lutjanidae) from Indian waters. *Journal of the Ocean Science Foundation* 13: 16–26. Vivekanandan E. 2011. Marine Fisheries Policy Brief 3: Climate change and Indian marine fisheries. CMFRI Special Publication 105: 1–97. http://eprints.cmfri.org.in/id/ eprint/8440

Vivekanandan E, Krishnakumar PK. **2010**. Spatial and temporal differences in the coastal fisheries along the east coast of India. *Indian Journal of Marine Sciences* 39(3): 380–387. http://nopr.niscair.res.in/handle/123456789/10674

Voigt M, Weber D. **2011**. *Field guide for sharks of the genus* Carcharhinus. Munich: Verlag Dr. Friedrich Pfeil. 151 pp.

Volz W. **1903**. Neue Fische aus Sumatra. *Zoologischer Anzeiger* 26(703): 553–559.

Von Bonde C. 1922. The Heterosomata (flat fishes) collected by the S.S. Pickle. Union of South Africa Fisheries and Marine Biological Survey Vol. 2(Special Report 1): 1–29.

Von Bonde C. 1923. Shallow-water fishes procured by the S.S. Pickle. Union of South Africa Fisheries and Marine Biological Survey Vol. 3(Special Report 1): 1–40.

Von Bonde C. **1925**. A collection of Heterosomata from Portuguese East Africa. *Transactions of the Royal Society of South Africa* 12(1): 285–294.

Von Bonde C. **1929**. Union of South Africa Fisheries and Marine Biological Survey Report 7: 4–62.

Von Bonde C. **1934**. A collection of marine fishes from Zanzibar. *Annals of the Natal Museum* 7(3): 435–458.

Von Bonde C, Swart DB. **1923**. The Platostomia (skates and rays) collected by the S.S. *Pickle. Union of South Africa Fisheries and Marine Biological Survey* Report 3(art.5): 1–22.

Von Holten HS. **1802**. *Trichiurus gladius*, en ny fisk fra Portugal. *Skrivter af Naturhistorie-Selskabet Kjøbenhavn* 5: 19–26.

Von Krusenstern AJ. **1810–1812**. *Reise urn die Welt, in den Jahren 1803, 1804, 1805 und 1806*, etc. St. Petersburg. (Atlas only).

Von Tschudi JJ. 1845. Ichthyologie. In: Untersuchungen über die Fauna Peruana. St. Gallen: Scheitlin & Zollikofer. xxx + 35 pp.

Voronina EP. 2009. New species of righteye flounder Samariscus leopardus sp. nov. (Samaridae, Pleuronectiformes) from the Saya de Malha Bank (Indian Ocean). Voprosy Ikhtiologii 49(5): 593–603. [In Russian. English translation in Journal of Ichthyology 49(8): 575–584]

Voronina EP, Volkova GA. 2007. Catalog of specimens in the collection of the Zoological Institute, Russian Academy of Sciences. Osteichthyes, Scorpaeniformes, Scorpaenoidei, Congiopodoidei, Platycephaloidei, Anoplopomatoidei, Hexagrammoidei, Scorpaenoidei. St. Petersburg: Zoological Institute, Russian Academy of Sciences. 189 pp. [In Russian]

Vossoughi GH, Vosoughi AR. **1999**. Study of batoid fishes in northern part of Hormoz Strait, with emphasis on some species new to the Persian Gulf and Sea of Oman. *Indian Journal of Fisheries* 46(3): 301–306. Vrijenhoek RC. 1994. Unisexual fish: model systems for studying ecology and evolution. *Annual Review of Ecology* and Systematics 25: 71–96. https://doi.org/10.1146/annurev. es.25.110194.00

Vrijenhoek RC. 1999. Parthenogenesis and natural clones (pp. 695–702). In: Knobil E, Neill JD (eds) *Encyclopedia of Reproduction* Vol. 3. San Diego: Academic Press. xxxvi + 1072 pp.

# W

Wade CB. **1946**. Two new genera and five new species of apodal fishes from the eastern Pacific. *Allan Hancock Pacific Expedition 1932–40, Los Angeles* 9(7): 181–213, Pls. 24–28.

Waite ER. 1894. New or rare fishes from Maroubra, N. S. W. Proceedings of the Linnean Society of New South Wales (Series 2) 9(2): 215–227, Pl. 17.

Waite ER. 1899. Scientific results of the trawling expedition of H.M.C.S. *Thetis* off the coast of New South Wales, in February and March, 1898. *Memoirs of the Australian Museum*, Sydney 4(pt 1): 2–132.

Waite ER. **1904**. Additions to the fish fauna of Lord Howe Island. No. 4. *Records of the Australian Museum* 5(3): 135–186.

Waite ER. **1905**. Notes on fishes from Western Australia. No. 3. *Records of the Australian Museum* 6(2): 55–82, Pls. 8–17.

Waite ER. **1910**. Notes on New Zealand fishes. *Transactions of the New Zealand Institute* 42: 384–391.

Waite ER. **1911**. Additions to the fish fauna of New Zealand: No. II. *Transactions and Proceedings of the New Zealand Institute* 43(2) [for 1910]: 49–51.

Waite ER. 1911. Scientific results of the New Zealand government trawling expedition, 1907. Pisces - part 2. *Records of the Canterbury Museum* 1(3): 157–272, Pls. 24–57.

Waite ER, Hale HM. **1921**. Review of the lophobranchiate fishes (pipe-fishes and sea-horses) of South Australia. *Records of the South Australian Museum (Adelaide)* 1(4): 293–324.

Wajih S, Naqvi A, Narvekar PV, Desa E. 2006. Coastal biogeochemical processes in the North Indian Ocean (pp. 723–783). In: Robinson AR, Brink KH (eds) *The sea*, *Vol. 14A: The global coastal ocean: Interdisciplinary regional studies and syntheses*. Cambridge, Massachusetts: Harvard University Press. 840 pp.

Wakiya Y. **1924**. The carangoid fishes of Japan. *Annals of the Carnegie Museum* 15(2–3): 139–244.

Walbaum JJ. 1792. Petri Artedi sueci genera piscium. In quibus systema totum ichthyologiae proponitur cum classibus, ordinibus, generum characteribus, specierum differentiis, observationibus plurimis. Redactis speciebus 242 ad genera 52. Ichthyologiæ pars III. Grypeswaldiae: Impensis Ant. Ferdin. Röse. 723 pp. Wallace EM, Tringali MD. 2010. Identification of a novel member in the family Albulidae (bonefishes). *Journal of Fish Biology* 76(8): 1972–1983. http://dx.doi.org/10.1111/ j.1095-8649.2010.02639.x

Wallace EM, Tringali MD. **2016**. Fishery composition and evidence of population structure and hybridization in the Atlantic bonefish species complex (*Albula* spp.). *Marine Biology* 163(art.142): 1–15. http://dx.doi.org/10.1007/ s00227-016-2915-x

Wallace JH. 1967. The batoid fishes of the east coast of southern Africa. Part I: sawfishes and guitarfishes. *Investigational Report. Oceanographic Research Institute* (Durban) 15: 1–32.

Wallace JH. 1967. The batoid fishes of the east coast of southern Africa. Part II: manta, eagle, duckbill, cownose, butterfly and sting rays. *Investigational Report. Oceanographic Research Institute* (Durban) 16: 1–56.

Wallace JH. 1967. The batoid fishes of the east coast of southern Africa. Part III: skates and electric rays. *Investigational Report. Oceanographic Research Institute* (Durban) 17: 1–62.

Wallace JH. **1969**. Final report on the skate and ray project. Bulletin of the South African Assosication for Marine Biological Research 7: 24–26.

Wallace JH. 1975. The estuarine fishes of the east coast of South Africa. I. Species composition and length distribution in the estuarine and marine environments. II. Seasonal abundance and migrations. *Investigational Report*. *Oceanographic Research Institute* (Durban) 40: 1–72.

Wallace JH. 1975. The estuarine fishes of the east coast of South Africa. III. Reproduction. *Investigational Report*. *Oceanographic Research Institute* (Durban) 41: 1–51.

Wallace JH, Kok HM, Buxton CD, Bennett B. 1984. Inshore small-mesh trawling survey of the Cape south coast. Part 1. Introduction, methods, stations and catches. *South African Journal of Zoology* 19(3): 154–164.

Wallace JH, Van der Elst RP. 1975. The estuarine fishes of the east coast of South Africa. IV. Occurrence of juveniles in estuaries. V. Ecology, estuarine dependence and status. *Investigational Report. Oceanographic Research Institute* (Durban) 42: 1–63.

Wallace-Fincham BP. **1987**. The food and feeding of *Etrumeus whiteheadi* Wongratana 1983, off the Cape Province of South Africa. Masters thesis, University of Cape Town, South Africa. 117 pp.

Walmsley-Hart SA, Sauer WHH, Buxton CD. 1999. The biology of the skates *Raja wallacei* and *R. pullopunctata* (Batoidea: Rajidae) on the Agulhas Bank, South Africa. *South African Journal of Marine Science* 21(1): 165–179. Walsh M, Randall JE. 2004. Thalassoma jansenii x T. quinquevittatum and T. nigrofasciatum x T. quinquevittatum, hybrid labrid fishes from Indonesia and the Coral Sea. aqua, Journal of Ichthyology and Aquatic Biology 9(2): 69–74.

Walter JP, Ebert DA. 1991. Preliminary estimates of age of the bronze whaler *Carcharhinus brachyurus* (Chondrichthyes: Carcharhinidae) from southern Africa, with a review of some life history parameters. *South African Journal of Marine Science* 10(1): 37–44.

Walters V. **1960**. Synopsis of the lampridiform suborder Veliferoidei. *Copeia* 1960(3): 245–247.

Walters V. **1963**. On two hitherto overlooked teleost families: Guentheridae (Ateleopodiformes) and Radiicephalidae (Lampridiformes). *Copeia* 1963(2): 455–457.

Walters V. **1963**. Order Giganturoidei. In: Fishes of the western North Atlantic. *Memoir of the Sears Foundation for Marine Research* 1(part 4): 566–577.

Walters V, Fierstine HL. **1964**. Measurements of Swimming Speeds of Yellowfin Tuna and Wahoo. *Nature* 202: 208–209.

Walters V, Fitch JE. **1960**. The families and genera of the lampridiform (Allotriognath) suborder Trachipteroidei. *California Fish and Game* 46(4): 441–452.

Waples RS. **1981**. A biochemical and morphological review of the lizardfish genus *Saurida* in Hawaii, with the description of a new species. *Pacific Science* **35**(3): 217–235.

Waples RS, Randall JE. **1989**. A revision of the Hawaiian lizardfishes of the genus *Synodus*, with descriptions of four new species. *Pacific Science* 42(3–4): 178–213. [also cited as 1988]

Warashina I, Hisada K. 1972. Geographical distribution and body length composition of two tuna-like fishes, *Gasterochisma melampus* Richardson and Allothunnus fallai Serventy, taken by Japanese tuna longline fishery. Bulletin of the National Research Institute of Far Seas Fisheries 6: 51–75.

Ward RD, Hanner R, Hebert PDN. **2009**. The campaign to DNA barcode all fishes, FISH-BOL. *Journal of Fish Biology* 74(2): 329–356. http://dx.doi.org/10.1111/j.1095-8649.2008.02080.x

Ward RD, Zemlak TS, Innes BH, Last PR, Hebert PDN. **2005**. DNA barcoding Australia's fish species. *Philosophical Transactions of the Royal Society B (Biological Sciences)* 360(1462): 1847–1857.

Washington BB, Eschmeyer WN, Howe KM. 1984.
Scorpaeniformes: relationships. In: Moser HG, Richards
WJ, Cohen DM, Fahay MP, Kendall SW Jr, Richardson SL
(eds) Ontogeny and systematics of fishes. *American Society* of Ichthyologists and Herpetologists Special Publication 1: 438–447.

Wass RC. **1984**. An annotated checklist of the fishes of Samoa. NOAA Technical Report NMFS SSRF-781: 1–43. Wassink H. 1988. Eine gelungene Zucht des Mirakelfisches, Calloplesiops altivelis (Steindachner, 1903). Die Aquarienund Terrarien-Zeitschrift 41(3): 126–130.

Wassink H. 1989. Calloplesiops altivelis (Steindachner, 1903) reproduction réussie. Aquarama 105: 42–47. [adapted by Jean-Jacques Eckert from Wassink, H. 1988; see above]

Watson G, Smale MJ. 1998. Reproductive biology of shortnose spiny dogfish, *Squalus megalops*, from the Agulhas Bank, South Africa. *Marine and Freshwater Research* 49(7): 695–703.

Watson RE. **1991**. A provisional review of the genus *Stenogobius* with descriptions of a new subgenus and thirteen new species. (Pisces: Teleostei: Gobiidae). *Records of the Western Australian Museum* 15(3): 571–654.

Watson RE. **1995**. Review of the freshwater goby genus *Cotylopus* (Teleostei: Gobiidae: Sicydiinae). *Ichthyological exploration of Freshwaters* 6(1): 61–70.

Watson RE, Marquet G, Pöllabauer C. **2000**. New Caledonia fish species of the genus *Sicyopterus* (Teleostei: Gobioidei: Sicydiinae). *aqua, Journal of Ichthyology and Aquatic Biology* 4(1): 5–34.

Weber M. 1895. Fische von Ambon, Java, Thursday Island, dem Burnett-Fluss und von der Süd-Küste von Neu-Guinea.
In: Zoologische Forschungsreisen in Australien und dem Malayischen Archipel: mit Unterstützung des Herrn Dr. Paul von Ritter ausgeführt in den Jahren 1891–1893 von Richard Semon. Bd. 5: 257–276.

Weber M. **1897**. Beitrage zur Kenntniss der Fauna von Süd-Afrika. Zoologische Jahrbücher, Abteilung für Systematik, Geographie und Biologie der Tiere (Jena) 10: 135–199.

Weber M. **1902**. *Photoblepharon* n gen. *Siboga* Expedition, Introd. p. 108.

Weber M. **1909**. Diagnosen neuer Fische der *Siboga*-Expedition. *Notes from the Leyden Museum* 31(4): 143–169.

Weber M. 1911. Die Fische der Aru- und Kei-Inseln. Ein Beitrag zur Zoogeographie dieser Inseln. Abhandlungen der Senckenbergischen Naturforschenden Gesellschaft 34(1): 1–49.

Weber M. **1913**. *Die Fische der* Siboga-*Expedition*. Leiden: E.J. Brill. 710 pp.

Weber M, De Beaufort LF. 1915. Die Insel Nias bei Sumatra (pp. 268–276). In: Kleiweg de Zwaan JP (ed) Kraniologische Untersuchungen Niassischer Schädel Springer. 325 pp.

Weber M, De Beaufort LF. 1916. The fishes of the Indo-Australian Archipelago. III. Ostariophysi: II Cyprinoidea, Apodes, Synbranchi. Vol. 3. Leiden: E.J. Brill. xv + 455 pp.

Weber M, De Beaufort LF. **1922**. The fishes of the Indo-Australian Archipelago. IV. Heteromi, Solenichthyes, Synentognathi, Percesoces, Labyrinthici, Microcyprini. Vol. 4. Leiden: E.J. Brill. xiii + 410 pp. Weber M, De Beaufort LF. **1929**. The fishes of the Indo-Australian Archipelago. V. Anacanthini, Allotriognathi, Heterostomata, Berycomorphi, Percomorphi: families Kuhliidae, Apogonidae, Plesiopidae, Pseudoplesiopidae, Priacanthidae, Centropomidae. Vol. 5. Leiden: E.J. Brill. xiv + 458 pp.

Weber M, De Beaufort LF. 1931. *The fishes of the Indo-Australian Archipelago. VI. Perciformes (continued)*.Vol. 6. Leiden: E.J. Brill. vii + 448 pp.

Weed WH III. **1961**. A new species of *Aseraggodes* (Soleidae) from Ceylon. *Copeia* 1961(3): 292–295.

Wegner NC, Snodgrass OE, Dewar H, Hyde JR. 2015. Wholebody endothermy in a mesopelagic fish, the opah, *Lampris* guttatus. Science 348(6236): 786–789. http://dx.doi. org/10.1126/science.aaa8902

Weigmann S. **2016**. Annotated checklist of the living sharks, batoids and chimaeras (Chondrichthyes) of the world, with a focus on biogeographical diversity. *Journal of Fish Biology* 88(3): 837–1037. http://dx.doi.org/10.1111/jfb.12874

Weigmann S, Ebert DA, Clerkin PJ, Stehmann MFW, Naylor GJP. **2016**. *Bythaelurus bachi* n. sp., a new deep-water catshark (Carcharhiniformes, Scyliorhinidae) from the southwestern Indian Ocean, with a review of *Bythaelurus* species and a key to their identification. *Zootaxa* 4208(5): 401–432. https://doi.org/10.11646/zootaxa.4208.5

Weigmann S, Ebert DA, Séret B. 2021. Resolution of the Acroteriobatus leucospilus species complex, with a redescription of A. leucospilus (Norman, 1926) and descriptions of two new western Indian Ocean species of Acroteriobatus (Rhinopristiformes, Rhinobatidae). Marine Biodiversity 51(art.58): 1–30. https://doi.org/10.1007/ s12526-021-01208-6

Weigmann S, Gon O, Leeney RH, Barrowclift E, Berggren P, Jiddawi N, Temple AJ. 2020. Revision of the sixgill sawsharks, genus *Pliotrema* (Chondrichthyes, Pristiophoriformes), with descriptions of two new species and a redescription of *P. warreni* Regan. *PLoS ONE* 15(3): e0228791 (56 pp). http://dx.doi.org/10.1371/journal. pone.0228791

Weigmann S, Kaschner CJ. 2017. Bythaelurus vivaldii, a new deep-water catshark (Carcharhiniformes, Scyliorhinidae) from the northwestern Indian Ocean off Somalia. Zootaxa 4263(1): 97–119. https://doi.org/10.11646/zootaxa.4263.1.4

Weigmann S, Kaschner CJ, Thiel R. 2018. A new microendemic species of the deep-water catshark genus *Bythaelurus* (Carcharhiniformes, Pentanchidae) from the northwestern Indian Ocean, with investigations of its feeding ecology, generic review and identification key. *PLoS ONE* 13(12): e0207887 (51 pp). https://doi.org/10.1371/ journal.pone.0207887

Weigmann S, Séret B, Stehmann M. **2021**. *Notoraja hesperindica* sp. nov., a new colorful deep-sea softnose skate

(Elasmobranchii, Rajiformes, Arhynchobatidae) and first generic record from the western Indian Ocean. *Marine Biodiversity* 51(art.35): 1–19. https://doi.org/10.1007/ s12526-021-01162-3

Weigmann S, Stehmann MFW. 2016. Sinobatis brevicauda n. sp., a new deep-water legskate (Rajiformes, Anacanthobatidae) and first generic record from the western Indian Ocean. Zootaxa 4137(4): 478–500. http://dx.doi.org/10.11646/zootaxa.4137.4.2

Weigmann S, Stehmann MFW, Thiel R. **2013**. *Planonasus parini* n. g. and n. sp., a new genus and species of false cat sharks (Carchariniformes, Pseudotriakidae) from the deep northwestern Indian Ocean off Socotra Islands. *Zootaxa* 3609(2): 163–181. http://dx.doi.org/10.11646/ zootaxa.3609.2.3

Weigmann S, Stehmann MFW, Thiel R. **2014**. Complementary redescription of *Anacanthobatis ori* (Wallace, 1967) and its assignment to *Indobatis* n. g. (Elasmobranchii, Anacanthobatidae), with comments on other legskates. *Zootaxa* 3779(2): 101–132. https://doi.org/10.11646/ zootaxa.3779.2.1

Weinberg S. **1999**. *A fish caught in time: the search for the coelacanth*. London: Fourth Estate. 239 pp.

Weiss G, Hubold G, Bainy ACD. **1987**. Larval development of the zeiform fishes *Antigonia capros* Lowe, 1843 and *Zenopsis conchifer* (Lowe, 1852) from the south west Atlantic. *Cybium* 11(1): 79–91.

Weitkamp DE, Sullivan RD. **1939**. Fishes. The John Murray Expedition 1933–34. *Scientific Reports, John Murray Expedition* 25 Nov. 7: 1–116.

Welander AD, Schultz LP. **1951**. *Chromis atripectoralis*, a new damselfish from the tropical Pacific, closely related to *C. caeruleus*, family Pomacentridae. *Journal of the Washington Academy of Sciences* **41**(3): 107–110.

Wellington GM. **1978**. Undersea wonders of the Galapagos. *National Geographic Magazine* 154(3): 363–381.

Werner TB, Allen GR (eds). **1998**. *A rapid biodiversity assessment of the coral reefs of Milne Bay Province, Papua New Guinea*. RAP Working Paper No. 11. Conservation International: Washington DC, USA. [unpaginated]

Westneat MW. 1993. Phylogenetic relationships of the tribe Cheilinini (Labridae: Perciformes). Bulletin of Marine Science 52: 351–394.

Westneat MW, Alfaro ME. **2005**. Phylogenetic relationships and evolutionary history of the reef fish family Labridae. *Molecular Phylogenetics and Evolution* 36: 370–390.

Westneat MW, Wainwright PC. **1989**. Feeding mechanism of *Epibulus insidiator* (Labridae; Teleostei): evolution of a novel functional system. *Journal of Morphology* 202(2): 129–150. Wetzel J, Wourms JP. **1995**. Adaptations for reproduction and development in the skin-brooding ghost pipefishes, *Solenostomus. Environmental Biology of Fishes* 44(4): 363–384.

Wheeler AC. **1955**. A preliminary revision of the fishes of the genus *Aulostomus*. *Annals and Magazine of Natural History* (Series 12) 8(92): 613–623.

Wheeler AC. **1958**. The Gronovius fish collection; a catalogue and historical account. *Bulletin of the British Museum* (*Natural History*) *Historical Series* 1(5): 185–249.

Wheeler AC. 1973. Uranoscopidae (p. 451). In: Hureau J-C, Monod T (eds) Check-list of the fishes of the north-eastern Atlantic and of the Mediterranean (CLOFNAM).
Vol. I. Paris: UNESCO.

Wheeler AC. **1980**. The sources of Linnaeus's knowledge of fishes. *Svenska Linnésällskapets Årsskrift* 1: 156–211.

Wheeler JFG. **1961**. The genus *Lethrinus* in the western Indian Ocean. *Colonial Office, Fishery Publications* (London) 15: 1–51, Pls. 1–3.

Wheeler JFG, Ommanney FD. **1953**. Report on the Mauritius-Seychelles fisheries survey 1948–1949. *Colonial Office Fishery Publications* 1(3): 1–145.

White EI, Moy-Thomas JA. **1941**. Notes on the nomenclature of fossil fishes. Part III. Homonyms M–Z. *Annals and Magazine of Natural History* (Series 11) 7(40, art.25): 395–400.

White J. **1790**. *Journal of a voyage to New South Wales with sixty-five plates of non descript animals, birds, lizards, serpents, curious cones of trees and other natural productions.* Piccadilly, London: J. Debrett. i–xvi, 1–299, Pls. 1–65.

White MG, North AW. **1980**. *Pseudoicichthys australis* (Pisces: Centrolophidae): an addition to the marine fauna of South Georgia and confirmation of Antarctic distribution. *British Antarctic Survey Bulletin* 50(October): 113–115.

White WT. **2012**. A redescription of *Carcharhinus dussumieri* and *C. sealei*, with resurrection of *C. coatesi* and *C. tjutjot* as valid species (Chondrichthyes: Carcharhinidae). *Zootaxa* 3241: 1–34. http://dx.doi.org/10.11646/zootaxa.3241.1.1

White WT. **2014**. A revised generic arrangement for the eagle ray family Myliobatidae, with definitions for the valid genera. *Zootaxa* 3860(2): 149–166. https://doi.org/10.11646/ zootaxa.3860.2.3

White WT, Corrigan S, Yang L, Henderson AC, Bazinet AL, Swofford DL, Naylor GJP. **2017**. Phylogeny of the manta and devilrays (Chondrichthyes: Mobulidae), with an updated taxonomic arrangement for the family. *Zoological Journal of the Linnean Society* 20: 1–26. https://doi.org/10.1093/ zoolinnean/zlx018

White WT, Dharmadi. 2007. Species and size compositions and reproductive biology of rays (Chondrichthyes, Batoidea) caught in target and non-target fisheries in eastern Indonesia. *Journal of Fish Biology* 70(6): 1809–1837. White WT, Ebert DA. **2008**. *Cephaloscyllium hiscosellum* sp. nov., a new swellshark (Carcharhiniformes: Scyliorhinidae) from northwestern Australia (pp. 171–178). In: Last PR, White WT, Pogonoski JJ (eds) *Descriptions of new Australian chondrichthyans*. Hobart, Australia: CSIRO Marine and Atmospheric Research. 358 pp.

White WT, Ebert DA, Naylor GJP, Ho H-C, Clerkin P, Veríssimo A, Cotton CF. 2013. Revision of the genus *Centrophorus* (Squaliformes: Centrophoridae):
Part 1—Redescription of *Centrophorus granulosus* (Bloch & Schneider), a senior synonym of *C. acus* Garman and *C. niaukang* Teng. *Zootaxa* 3752(1): 35–72. https://doi. org/10.11646/zootaxa.3752.1.5

White WT, Giles J, Dharmadi, Potter IC. 2006. Data on the bycatch fishery and reproductive biology of mobulid rays (Myliobatiformes) in Indonesia. *Fisheries Research* 82(1–3): 65–73.

White WT, Last PR. **2013**. Notes on shark and ray types at the South China Sea Fisheries Research Institute (SCSFRI) in Guangzhou, China. *Zootaxa* 3752(1): 228–248. http://dx.doi.org/10.11646/zootaxa.3752.1.14

White WT, Last PR. **2016**. Eagle rays. Family Myliobatidae (pp. 706–725); Pelagic eagle rays. Family Aetobatidae (pp. 726–731). In: Last PR, White WT, Carvalho MR de, Seret B, Stehmann MFW, Naylor GJP (eds) *Rays of the world*. CSIRO Publishing & Cornell University Press. 790 pp.

White WT, Last PR, Dharmadi. **2005**. Description of a new species of catshark, *Atelomycterus baliensis* (Carcharhiniformes: Scyliorhinidae) from eastern Indonesia. *Cybium* 29(1): 33–40.

White WT, Last PR, Naylor GJP, Harris M. 2010. Resurrection and redescription of the Borneo broadfin shark *Lamiopsis tephrodes* (Fowler, 1905) (Carcharhiniformes: Carcharhinidae) (pp. 45–59). In: Last PR, White WT, Pogonoski JJ (eds) *Descriptions of new sharks and rays from Borneo*. Hobart, Australia: CSIRO Marine and Atmospheric Research. 165 pp.

White WT, Last PR, Stevens JD. **2007**. *Halaelurus maculosus* n. sp. and *H. sellus* n. sp., two new species of catshark (Carcharhiniformes: Scyliorhinidae) from the Indo-West Pacific. *Zootaxa* 1639: 1–21.

White WT, Last PR, Stevens JD, Yearsley GK, Fahmi, Dharmadi. 2006. Economically important sharks and rays of Indonesia. ACIAR Monograph Series No. 124. Canberra: Australian Centre for International Agricultural Research. 329 pp.

White WT, Naylor GJP. **2016**. Resurrection of the family Aetobatidae (Myliobatiformes) for the pelagic eagle rays, genus *Aetobatus*. *Zootaxa* 4139(3): 435–438. http://dx.doi.org/10.11646/zootaxa.4139.3.10

White WT, Vaz DFB, Ho H-C, Ebert DA, Carvalho MR de, Corrigan S, Rochel E, Carvalho M de, Tanaka S, Naylor GJP. **2014**. Redescription of *Symnodon ichiharai* Yano and Tanaka 1984 (Squaliformes: Somniosidae) from the western North Pacific, with comments on the definition of somniosid genera. *Ichthyological Research* 62(2) [2015]: 213–229. https://doi.org/10.1007/s10228-014-0430-y

White WT, Weigmann S. **2014**. *Carcharhinus humani* sp. nov., a new whaler shark (Carcharhiniformes: Carcharhinidae) from the western Indian Ocean. *Zootaxa* 3821(1): 71–87. http://doi.org/10.11646/zootaxa.3821.1.5

Whitehead PJP. 1962. A review of the Indo-Pacific gizzard shad genera *Nematalosa*, *Clupanodon* and *Konosirus* (Pisces: Dorosomatidae). *Bulletin of the British Museum (Natural History) Zoology* 9(2): 89–102.

Whitehead PJP. **1963**. A revision of the recent round herrings (Pisces: Dussumieriidae). *Bulletin of the British Museum* (*Natural History*) Zoology 10(6): 305–380.

Whitehead PJP. 1964. Xiphias platypterus Shaw & Nodder, 1792 (Pisces): application to validate this nornen oblitum for the Indian Ocean sailfish (genus Istiophorus) Z.N.(S.) 1657. Bulletin of Zoological Nomenclature 21(6): 444–446.

Whitehead PJP. **1965**. A preliminary revision of the Indo-Pacific Alosinae (Pisces: Clupeidae). *Bulletin of the British Museum (Natural History) Zoology* 12(4): 115–156.

Whitehead PJP. **1965**. A review of the elopoid and clupeoid fishes of the Red Sea and adjacent regions. *Bulletin of the British Museum (Natural History) Zoology* 12(7): 225–281.

Whitehead PJP. **1967**. Indian anchovies collected by the Anton Bruun and Te Vega, 1963–64. *Journal of the Marine Biological Association of India* 9(1): 13–37.

Whitehead PJP. **1973**. A synopsis of the clupeoid fishes of India. *Journal of the Marine Biological Association of India* 14(1): 160–256.

Whitehead PJP. 1981. Clupeidae. In: Fischer W, Bianchi G, Scott WB (eds) FAO species identification sheets for fishery purposes. Eastern central Atlantic (Fishing Area 34 and part of 47). Vol. 1. Rome: FAO. [unpaginated]

Whitehead PJP. **1985**. FAO species catalogue. Vol. 7. Clupeoid fishes of the world (suborder Clupeoidei). An annotated and illustrated catalogue of the herrings, sardines, pilchards, sprats, anchovies and wolf-herrings. Part 1: Chirocentridae, Clupeidae and Pristigasteridae. *FAO Fisheries Synopsis No. 125* (Vol. 7, part 1, pp. 1–303). Rome: FAO.

Whitehead PJP, Bauchot M-L, Hureau J-C, Nielsen J, Tortonese E (eds). 1984–1986. Fishes of the north-eastern Atlantic and the Mediterranean (3 vols). Paris: UNESCO. Vol. I (1984): 1–510; Vol. II (1986): 517–1007; Vol. III (1986): 1015–1473.

Whitehead PJP, Ivantsoff W. **1983**. *Atherina lacunosa* and the fishes described by J.R. Forster. *Japanese Journal of Ichthyology* 29(4): 355–364.

Whitehead PJP, Nelson GJ, Wongratana T. **1988**. FAO species catalogue. Vol. 7. Clupeoid fishes of the world (suborder Clupeoidei). An annotated and illustrated catalogue of the

herrings, sardines, pilchards, sprats, shads, anchovies and wolf-herrings. Part 2. Engraulidae. *FAO Fisheries Synopsis No.* 125 (Vol. 7, part 2, pp. 305–579). Rome: FAO.

Whitehead PJP, Talwar PK. **1976**. Francis Day (1829–1889) and his collections of Indian Fishes. *Bulletin of the British Museum (Natural History) Historical Series* 5(1): 1–189.

Whitehead PJP, Wongratana T. 1984. Clupeidae. Herrings, shads, sardinellas, sprats, sardines. In: Fischer W, Bianchi G (eds) FAO species identification sheets for fishery purposes. Western Indian Ocean (Fishing Area 51). Vol. 1. Rome: FAO. [unpaginated]

Whitehead PJP, Wongratana T. 1984. Engraulidae. Anchovies. In: Fischer W, Bianchi G (eds) FAO species identification sheets for fishery purposes. Western Indian Ocean (Fishing Area 51). Vol. 2. Rome: FAO. [unpaginated]

Whitehead PJP, Wongratana T. **1986**. Family No. 54: Clupeidae. In: Smith MM, Heemstra PC (eds) Smiths' sea fishes. Johannesburg: Macmillan South Africa. 1047 pp.

Whitehouse RH. **1923**. A statistical study of young fishes from Silavatturai Lagoon, Tuticorin. *Madras Fisheries Bulletin* 17(3): 49–103.

Whitehouse RH. **1927**. The grey mullets of Tuticorin. *Madras Fisheries Bulletin* 15(3): 71–98.

Whitfield AK. **1994**. An estuary-association classification for the fishes of southern Africa. *South African Journal of Science* 90: 411–417.

Whitfield AK. **1995**. Threatened fishes of the world: *Syngnathus watermeyeri* Smith, 1963 (Syngnathidae). *Environmental Biology of Fishes* 44: 152.

Whitfield AK. **1998**. Biology and ecology of fishes in southern African estuaries. *Ichthyological Monograph of the J.L.B. Smith Institute of Ichthyology* 2: 1–223.

Whitfield PE, Gardner T, Vives SP, Gilligan MR, Courtenay WR, Ray GC, Hare JA. 2002. Biological invasion of the Indo-Pacific lionfish (*Pterois volitans*) along the Atlantic coast of North America. *Marine Ecology Progress Series* 235: 289–297.

Whitley GP. **1929**. Fishes from Ongtong Java, Melanesia. *Proceedings of the Linnean Society of New South Wales* 54(2): 91–95.

Whitley GP. **1929**. Studies in ichthyology. No. 3. *Records of the Australian Museum* 17(3): 101–143.

Whitley GP. **1930**. Additions to the check-list of the fishes of New South Wales, No. 3. *Australian Zoologist* 6(2): 117–123.

Whitley GP. **1930**. Five new generic names for Australian fishes. *Australian Zoologist* 6(3): 250–251.

Whitley GP. **1930**. Ichthyological miscellanea. *Memoirs of the Queensland Museum* 10(1): 8–31.

Whitley GP. **1931**. New names for Australian fishes. *Australian Zoologist* 6(4): 310–334.

Whitley GP. **1932**. Fishes. *Great Barrier Reef Expedition* 1928–29: Scientific Reports 4(9): 267–316.

Whitley GP. **1932**. Studies in ichthyology. No. 6. *Records of the Australian Museum* 18(6): 321–348.

Whitley GP. **1933**. Studies in ichthyology. No. 7. *Records of the Australian Museum* 19(1): 60–112.

Whitley GP. **1934**. Notes on some Australian sharks. *Memoirs* of the Queensland Museum 10(4): 180–200.

Whitley GP. **1935**. Fishes from Princess Charlotte Bay, North Queensland. *Records of the South Australian Museum* (*Adelaide*) 5(3): 345–363.

Whitley GP. **1935**. Studies in ichthyology. No. 9. *Records of the Australian Museum* 19(4): 215–250.

Whitley GP. **1936**. The Australian devil ray, *Daemomanta alfredi* (Krefft), with remarks on the superfamily Mobuloidea (order Batoidei). *The Australian Zoologist* 8(3): 164–188.

Whitley GP. **1937**. The Middleton and Elizabeth Reefs, South Pacific Ocean. *The Australian Zoologist* 8(4): 199–231.

Whitley GP. **1938**. Ray's bream and its allies in Australia. *The Australian Zoologist* 9(2): 191–194.

Whitley GP. **1939**. Taxonomic notes on sharks and rays. *The Australian Zoologist* 9(3): 227–262.

Whitley GP. **1940**. *The fishes of Australia. Part I. The sharks, rays, devil-fish, and other primitive fishes of Australia and New Zealand.* Australian Zoological Handbook. Sydney: Royal Zoological Society of New South Wales. 280 pp.

Whitley GP. **1940**. The *Nomenclator Zoologicus* and some new fish names. *Australian Naturalist* 10(7): 241–243.

Whitley GP. **1941**. Ichthyological notes and illustrations. *Australian Zoologist* 10(1): 1–50, Pls. 1–2.

Whitley GP. **1943**. Ichthyological notes and illustrations (Part 2). *Australian Zoologist* 10(2): 167–187.

Whitley GP. **1945**. New sharks and fishes from Western Australia. Part 2. *Australian Zoologist* 11(1): 1–42, Pl. 1.

Whitley GP. **1948**. New sharks and fishes from Western Australia. Part 4. *Australian Zoologist* 11(3): 259–276.

Whitley GP. **1948**. Studies in ichthyology. No. 13. *Records of the Australian Museum* 22(1): 70–94.

Whitley GP. **1950**. Clingfishes. *Australian Museum Magazine* 10(4): 124–128.

Whitley GP. **1951**. Studies in ichthyology. No. 15. *Records of the Australian Museum* 22(4): 389–408.

Whitley GP. **1954**. More new fish names and records. *Australian Zoologist* 12(1): 57–62.

Whitley GP. **1955**. Taxonomic notes on fishes. *Proceedings of the Royal Zoological Society of New South Wales* 1953–54: 44–57.

Whitley GP. **1959**. Ichthyological snippets. *Australian Zoologist* 12(4): 310–323.

Whitley GP. **1961**. A new scorpion fish from Queensland. *North Queensland Naturalist* 29(127): 9–10.

Whitley GP. **1964**. Fishes from the Coral Sea and the Swain Reefs. *Records of the Australian Museum* 26(5): 145–195.

Whitley GP. **1965**. Illustrations and records of fishes. *Australian Zoologist* 13(2): 103–120.

Whitley GP. **1970**. Ichthyological quiddities. *Australian Zoologist* 15(3): 242–247.

Whitley GP, Colefax AN. **1938**. Fishes from Nauru, Gilbert Islands, Oceania. *Proceedings of the Linnean Society of New South Wales* 63(parts 3–4, Nos. 277–278): 282–304.

Wickler W. 1968 Mimikry: Nachahmung und Täuschung in der Natur. München: Kindler Verlag GmbH. 256 pp.
[English translation: Mimicry in plants and animals. London: Wiedenfeld and Nicolson. 255 pp.]

Widder EA. **1998**. A predatory use of counterillumination by the squaloid shark, *Isistius brasiliensis*. *Environmental Biology of Fishes* 53(3): 267–273.

Wilderbuer T, Leaman B, Zhang CI, Fargo J, Paul L. 2005. Pacific flatfish fisheries (pp. 272–291). In: Gibson RN (ed) *Flatfishes: Biology and exploitation*. Oxford: Blackwell Science. xxiv + 391 pp.

Wiley EO. **1979**. Ventral gill arch muscles and the interrelationships of gnathostomes, with a new classification of the Vertebrata. *Zoological Journal of the Linnean Society* 67(2): 149–179.

Wiley EO, Johnson GD. 2010. A teleost classification based on monophyletic groups (pp. 123–182). In: Nelson J, Schultze H-P, Wilson MVH (eds) Origin and phylogenetic interrelationships of teleosts. Munich: Verlag Dr. Friedrich Pfeil. 482 pp.

Wiley ML, Collette BB. 1970. Breeding tubercles and contact organs in fishes: their occurrence, structure and significance. *Bulletin of the American Museum of Natural History* 143(art.3): 143–216.

Wilkinson CD (ed). 2008. Status of coral reefs of the world: 2008. Townsville, Australia: Global Coral Reef Monitoring Network and Reef and Rainforest Research Center. 296 pp.

Willey A. **1910**. The occurrence of *Solenostoma* off the coast of Ceylon. *Spolia Zeylanica* 6(23): 102–107, Pl. 1.

Williams F. 1958. Fishes of the family Carangidae in British East African waters. *Annals and Magazine of Natural History* (Series 13) 1(6): 369–430, Pls. 6–16.

Williams F. 1959. The barracudas (genus Sphyraena) in British East African waters. Annals and Magazine of Natural History (Series 13) 2(14): 92–128.

Williams F. 1960. On Scomberomorus lineolatus (C.V.) 1831, from British East African waters (Pisces: Scombridae). Annals and Magazine of Natural History (Series 13) 3(27): 183–192.

Williams F. 1961. On Uraspis wakiyai sp. nov. (Pisces, Carangidae), from the western Indian Ocean, with a review of the species of Uraspis Bleeker, 1855, S.S. Annals and Magazine of Natural History (Series 13) 4(38) 65–87. Williams F. 1963. Synopsis of biological data on little tuna Euthynnus affinis (Cantor) 1859 (Indian Ocean). Species Synopsis No. 5, FAO Fisheries Biology Synopsis 48: 167–179.

Williams F. **1964**. The scombroid fishes of East Africa. *Symposium Series, Marine Biological Association of India* 1(1): 107–164.

Williams F. **1965**. Further notes on the biology of East African pelagic fishes of the families Carangidae and Sphyraenidae. *East African Journal of Agriculture and Forestry* 31: 141–168.

Williams F. **1966**. Food of longline-caught yellowfin tuna from east African waters. *East African Agricultural and Forestry Journal* 31: 375–382.

Williams F, Heemstra PC, Shameem A. 1980. Notes on the Indo-Pacific carangid fishes of the genus *Carangoides* Bleeker. 2. The *Carangoides armatus* group. *Bulletin of Marine Science* 30(1): 13–20.

Williams F, Venkataramani VK. 1978. Notes on Indo-Pacific carangid fishes of the genus *Carangoides* Bleeker. 1. The *Carangoides malabaricus* group. *Bulletin of Marine Science* 28(3): 501–511.

Williams JT. **1983**. Synopsis of the pearlfish subfamily Pyromodontinae (Pisces: Carapidae). *Bulletin of Marine Science* 33(4): 846–854.

Williams JT. **1984**. Studies on *Echiodon* (Pisces: Carapidae), with description of two new Indo-Pacific species. *Copeia* 1984(2): 410–422.

Williams JT. **1984**. Synopsis and phylogenetic analysis of the pearlfish subfamily Carapinae (Pisces: Carapidae). *Bulletin of Marine Science* 34(3): 386–397.

Williams JT. **1988**. Revision and phylogenetic relationships of the blenniid fish genus *Cirripectes*. *Indo-Pacific Fishes* 17: 1–78.

Williams JT. 2010. A new species of blenny, *Cirripectes heemstraorum*, from Cape Vidal, South Africa (Family Blenniidae). *Smithiana* Bulletin 12: 3–7.

Williams JT, Bogorodsky SV. **2010**. *Entomacrodus solus*, a new species of blenny (Perciformes: Blenniidae) from the Red Sea. *Zootaxa* 2475: 64–68. http://dx.doi.org/10.11646/zootaxa.2475.1.5

Williams JT, Delrieu-Trottin E, Planes S. 2012. A new species of Indo-Pacific fish, *Canthigaster criobe*, with comments on other *Canthigaster* (Tetraodontiformes: Tetraodontidae) at the Gambier Archipelago. *Zootaxa* 3523: 80–88. http://dx.doi.org/10.11646/zootaxa.3523.1.9

Williams JT, Howe JC. **2003**. Seven new species of the triplefin fish genus *Helcogramma* (Tripterygiidae) from the Indo-Pacific. *aqua, Journal of Ichthyology and Aquatic Biology* 7(4): 151–176.

Williams JT, McCormick CJ. **1990**. Two new species of the triplefin fish genus *Helcogramma* (Tripterygiidae) from the western Pacific. *Copeia* 1990(4): 1020–1030.

Williams VR, Clarke TA. **1983**. Reproduction, growth and other aspects of the biology of the gold spot herring, *Herklotsichthys quadrimaculatus* (Clupeidae), a recent introduction to Hawaii. *Fishery Bulletin* 81(3): 587–597.

Wilson MVH, Bruner JC. 2004. Mesozoic fish assemblages of North America (pp. 575–595). In: Arratia G, Tintori A (eds) Mesozoic Fishes 3 – Systematics, paleoenvironments and biodiversity. Proceedings of the international meeting, Serpianao, 2001. Munich: Verlag Dr. Friedrich Pfeil. 649 pp.

Wilson MVH, Murray AM. **2008**. Osteoglossomorpha: phylogeny, biogeography, and fossil record and the significance of key African and Chinese taxa. *Geological Society of London Special Publication* 295(1): 185–219. http://dx.doi.org/10.1144/SP295.12

Wilson PC, Beckett JS. **1970**. Atlantic Ocean distribution of the pelagic stingray, *Dasyatis violacea*. *Copeia* 1970(4): 696–707.

Winstanley RH. **1978**. Food of the trevalla *Hyperoglyphe porosa* (Richardson) off southeastern Australia. *New Zealand Journal of Marine and Freshwater Research* 12(1): 77–79.

Winterbottom R. **1974**. Rediscovery of certain type specimens of fishes from the collections of the government marine survey made by J.D.F. Gilchrist and the S.S. *Pickle. J.L.B. Smith Institute of Ichthyology Special Publication* 12: 1–10.

Winterbottom R. **1976**. Additions to, and range extensions of, the South African marine ichthyofauna. *Zoologica Africana* 11(1): 59–73.

Winterbottom R. 1976. Notes on South African gobies possessing free upper pectoral fin rays (Pisces: Gobiidae). *J.L.B. Smith Institute of Ichthyology Special Publication* 16: 1–11.

Winterbottom R. **1976**. On *Clinus nematopterus* Günther with notes on other South African clinid fishes. *South African Journal of Science* 72(6): 178–180.

Winterbottom R. **1978**. Range extensions and additions to the South African marine ichthyofauna, with the description of a new species of congrogadid from Kwazulu. *Zoologica Africana* 13(1): 41–56.

Winterbottom R. **1979**. A new genus and species of the family Congrogadidae (Pisces: Perciformes) from the western Indian Ocean. *Bulletin of Marine Science* 29(3): 298–302.

Winterbottom R. **1980**. A new genus and three new species of the family Congrogadidae (Pisces, Perciformes) from Natal, South Africa. *Annals of the South African Museum* 83(1): 1–12.

Winterbottom R. **1980**. Two new species of the Congrogadidae (Pisces: Perciformes) from the Indo-West Pacific. *Copeia* 1980(3): 396–402.

Winterbottom R. **1982**. A revision of the congrogadid fish genus *Halidesmus* (Pisces: Perciformes), with the description of a new species from Kenya and a list of the species included in the family. *Canadian Journal of Zoology* 60(5): 754–763. Winterbottom R. **1984**. A review of the gobiid fish genus *Trimma* from the Chagos Archipelago, central Indian Ocean, with the description of seven new species. *Canadian Journal of Zoology* 62(4): 695–715.

Winterbottom R. **1985**. Revision of the congrogadid *Haliophis* (Pisces: Perciformes), with the description of a new species from Indonesia, and comments on the endemic fish fauna of the northern Red Sea. *Canadian Journal of Zoology* 63(2): 209–217.

Winterbottom R. 1985. Two new gobiid fish species (in *Priolepis* and *Trimma*) from the Chagos Archipelago, central Indian Ocean. *Canadian Journal of Zoology* 63(4): 748–754.

Winterbottom R. 1986. Revision and vicariance biogeography of the subfamily Congrogadinae (Pisces: Perciformes: Pseudochromidae). *Indo-Pacific Fishes* 9: 1–34.
[Cover date Oct. 1985]

Winterbottom R. **1992**. Evolution of *Naso thynnoides* and the status of *N. minor* (Acanthuridae; Actinopterygii). *Japanese Journal of Ichthyology* 38(4): 375–378.

Winterbottom R. **1993**. Search for the gobioid sister group (Actinopterygii: Percomorpha). *Bulletin of Marine Science* 52(1): 395–414.

Winterbottom R. **1993**. Myological evidence for the phylogeny of recent genera of surgeonfishes (Percomorpha, Acanthuridae), with comments on the Acanthuroidei. *Copeia* 1993: 21–39.

Winterbottom R. **1995**. Red Sea gobiid fishes of the genus *Trimma*, with the description of two new species. *Revue française d'Aquariologie Herpétologie* 22(3–4): 93–98.

Winterbottom R. **1996**. A new species of the congrogadin genus *Rusichthys* (Perciformes; Pseudochromidae), with notes on its osteology. *Canadian Journal of Zoology* 74(3): 581–584.

Winterbottom R. **2000**. Four new species of *Trimma* (Gobiidae), from the Indian and western Pacific oceans. *aqua, Journal of Ichthyology and Aquatic Biology* 4(2): 57–66.

Winterbottom R. **2001**. Two new gobiid fish species in *Trimma* and *Trimmatom* (Teleostei: Gobiidae) from the Indian and Western Pacific Oceans. *aqua, Journal of Ichthyology and Aquatic Biology* 5(4): 19–24.

Winterbottom R. **2002**. A redescription of *Cryptocentrus crocatus* Wongratana, a redefinition of *Myersina* Herre (Acanthopterygii; Gobiidae), a key to the species, and comments on relationships. *Ichthyological Research* 49(1): 69–75.

Winterbottom R. **2003**. A new species of the gobiid fish *Trimma* from the western Pacific and northern Indian Ocean coral reefs, with a description of its osteology. *Zootaxa* 218: 1–24.

Winterbottom R, Alofs KA, Marseu A. **2011**. Life span, growth and mortality in the western Pacific goby *Trimma benjamini*, and comparisons with *T. nasa. Environmental Biology of Fishes* 91(3): 295–301. https://doi.org/10.1007/ s10641-011-9782-6

Winterbottom R, Anderson RC. **1997**. A revised checklist of the epipelagic and shore fishes of the Chagos Archipelago, central Indian Ocean. *Ichthyological Bulletin of the J.L.B. Smith Institute of Ichthyology* 66: 1–28.

Winterbottom R, Anderson RC. **1999**. Fishes of the Chagos Archipelago. *Linnean Society of London, Occasional Publication* 2: 101–117.

Winterbottom R, Burridge M. **1992**. Revision of *Egglestonichthys* and of *Priolepis* species possessing a transverse pattern of cheek papillae (Teleostei; Gobiidae), with a discussion of relationships. *Canadian Journal of Zoology* 70(10): 1934–1946.

Winterbottom R, Burridge M. **1993**. Revision of the species of *Priolepis* possessing a reduced transverse cheek papillae pattern and no predorsal scales (Teleostei; Gobiidae). *Canadian Journal of Zoology* 71(3): 494–514.

Winterbottom R, Burridge M. **1993**. Revision of the Indo-Pacific *Priolepis* species possessing a reduced transverse pattern of cheek papillae, and predorsal scales (Teleostei; Gobiidae). *Canadian Journal of Zoology* 71(10): 2056–2076.

Winterbottom R, Burridge-Smith M. **1987**. The occurrence of the Japanese wrasse, *Stethojulis maculata* (Labridae), in Fiji. *Japanese Journal of Ichthyology* 33(4): 410–413.

Winterbottom R, Emery AR. **1981**. A new genus and two new species of gobiid fishes (Perciformes) from the Chagos Archipelago, central Indian Ocean. *Environmental Biology of Fishes* 6(2): 139–149.

Winterbottom R, Emery AR. **1986**. Review of the gobioid fishes of the Chagos Archipelago, central Indian Ocean. *Royal Ontario Museum Life Science Contributions* 142: i–v + 1–82.

Winterbottom R, Emery AR, Holm E. **1989**. An annotated checklist of the fishes of the Chagos Archipelago, central Indian Ocean. *Royal Ontario Museum Life Science Contributions* 145: 1–226.

Winterbottom R, Harold AS. **2005**. *Gobiodon prolixus*, a new species of gobiid fish (Teleostei: Perciformes: Gobiidae) from the Indo-west Pacific. *Proceedings of the Biological Society of Washington* 118(3): 582–589.

Winterbottom R, Hoese DF. 1988. A new genus and four new species of fishes from the Indo-West Pacific (Pisces; Perciformes; Gobiidae), with comments on relationships. *Royal Ontario Museum Life Sciences Occasional Paper* 37: 1–17.

Winterbottom R, Hoese DF. **2015**. A revision of the Australian species of *Trimma* (Actinopterygii, Gobiidae), with descriptions of six new species and redescriptions of

twenty-three valid species. *Zootaxa* 3934(1): 1–102. http://dx.doi.org/10.11646/zootaxa.3934.1.1

Winterbottom R, Randall JE. 1994. Two new species of congrogadins (Teleostei; Pseudochromidae), with range extensions for four other species. *Canadian Journal of Zoology* 72: 750–756.

Winterbottom R, Southcott L. 2007. Two new species of the genus *Trimma* (Percomorpha: Gobioidei) from western Thailand. *aqua*, *International Journal of Ichthyology* 13(2): 69–76.

Winterbottom R, Southcott L. **2008**. Short lifespan and high mortality in the western Pacific coral reef goby *Trimma nasa*. *Marine Ecology Progress Series* 366: 203–208. http://dx.doi.org/10.3354/meps07517

Wintner SP. **2000**. Preliminary study of vertebral growth rings in the whale shark, *Rhincodon typus*, from the east coast of South Africa. *Environmental Biology of Fishes* 59(4): 441–451.

Wintner SP, Cliff G. **1996**. Age and growth determination of the blacktip shark, *Carcharhinus limbatus*, from the east coast of South Africa. *Fishery Bulletin* 94(1): 135–144.

Wintner SP, Dudley SFJ, Kistnasamy N, Everett B. **2002**. Age and growth estimates for the Zambezi shark, *Carcharhinus leucas*, from the east coast of South Africa. *Marine and Freshwater Research* 53(2): 557–566.

Wirtz P, Ferreira CEL, Floeter SR, Fricke R, Gasparini JL, Iwamoto T, Rocha L, Sampaio CLA, Schliewen UK. 2007. Coastal fishes of São Tomé and Principe islands, Gulf of Guinea (eastern Atlantic Ocean) – an update. *Zootaxa* 1523: 1–48.

Witzell WN. **1978**. *Apolectus niger* (family Apolectidae): synonymy and systematics. *Matsya* 3 [for 1977]: 72–82.

Wong MYL, Munday PL, Jones GP. **2005**. Habitat patch size, facultative monogamy and sex change in a coral-dwelling fish, *Caracanthus unipinna*. *Environmental Biology of Fishes* 74(2): 141–150.

Wongratana T. **1977**. *Sillago intermedius*, a new species of sand whiting from the Gulf of Thailand (Pisces: Sillaginidae). *Natural History Bulletin of the Siam Society* 26(3–4): 257–262, Pls. 9–10.

Wongratana T. **1980**. Systematics of clupeoid fishes of the Indo-Pacific region (2 vols). PhD thesis, University of London.

Wongratana T. **1983**. Diagnoses of 24 new species and proposal of a new name for a species of Indo-Pacific Clupeoid fishes. *Japanese Journal of Ichthyology* 29(4): 385–407.

Wongratana T. **1987**. Two new species of anchovies of the genus *Stolephorus* (Engraulidae), with a key to species of *Engraulis, Encrasicholina*, and *Stolephorus. American Museum Novitates* 2876: 1–8.

Wongratana T, Munroe TA, Nizinski MS. **1999**. Order Clupeiformes. Engraulidae. Anchovies (pp. 1698–1753). In: Carpenter KE, Niem VH (eds) *FAO species identification guide for fishery purposes. The living marine resources of the western central Pacific.* Vol. 3. Batoid fishes, chimaeras and bony fishes part 1 (pp. 1397–2068). Rome: FAO.

- Woodland DJ. 1984. Gerreidae. Silver-biddies, mojarras. In:
  Fischer W, Bianchi G (eds) FAO species identification sheets for fishery purposes. Western Indian Ocean (Fishing Area 51).
  Vol. 2. Rome: FAO. [unpaginated]
- Woodland DJ. 1984. Siganidae. Spinefoots, rabbitfishes. In:
  Fischer W, Bianchi G (eds) FAO species identification sheets for fishery purposes. Western Indian Ocean (Fishing Area 51).
  Vol. 4. Rome: FAO. [unpaginated]

Woodland DJ. **1990**. Revision of the fish family Siganidae with descriptions of two new species and comments on distribution and biology. *Indo-Pacific Fishes* 19: 1–136.

- Woodland DJ. 2001. Gerreidae. Mojarras (silverbiddies) (pp. 2946–2960). In: Carpenter KE, Niem VH (eds) FAO species identification guide for fishery purposes. The living marine resources of the western central Pacific. Vol. 5. Bony fishes part 3 (pp. 2791–3380). Rome: FAO.
- Woodland DJ. 2001. Siganidae. Rabbitfishes (spinefoots) (pp. 3627–3650). In: Carpenter KE, Niem VH (eds) FAO species identification guide for fishery purposes. The living marine resources of the western central Pacific. Vol. 6. Bony fishes part 4 (pp. 3381–4218). Rome: FAO.
- Woodland DJ, Anderson RC. **2014**. Description of a new species of rabbitfish (Perciformes: Siganidae) from southern India, Sri Lanka and the Maldives. *Zootaxa* 3811(1): 129–136. http://dx.doi.org/10.11646/zootaxa.3811.1.8
- Woodland DJ, Randall JE. **1979**. *Siganus puelloides*, a new species of rabbitfish from the Indian Ocean. *Copeia* 1979(3): 390–393.
- Wood-Mason J, Alcock A. 1891. On the uterine villiform papillae of *Pteroplataea micrura*, and their relation to the embryo, being natural history notes from H.M. Indian marine survey steamer *Investigator*, Commander R.F. Hoskyn, R.N., Commanding. No. 22. *Proceedings of the Royal Society of London* 49: 4 unnumbered pp [Plates], 359–367

Woods LP. **1966**. Family Pleuronectidae (pp. 66–68). In: Schultz LP, Woods LP, Lachner EA (eds) *Fishes of the Marshall and Marianas Islands. Vol. 3. Families from Kraemeriidae through Antennariidae. Bulletin of the United States National Museum* No. 202: i–vii + 1–176, Pls. 124–148.

Woods LP, Schultz LP. 1953. Family Exocoetidae: flyingfishes (pp. 175–190). In: Schultz LP, Herald ES, Lachner EA, Welander AD, Woods LP (eds) Fishes of the Marshall and Marianas Islands. Vol. 1. Families from Asymmetrontidae through Siganidae. Bulletin of the United States National Museum No. 202: i–xxxii + 1–685, Pls. 1–74. Woods LP, Sonoda PM. 1973. Order Berycomorphi (Beryciformes). In: Fishes of the western North Atlantic. *Memoir of the Sears Foundation for Marine Research* 1(part 6): 263–396.

Woodward AS. 1895. Catalogue of the fossil fishes in the British Museum (Natural History). Part 3. Containing the Actinopterygian Teleostomi of the orders Chondrostei (concluded), Protospondyli, Aetheospondyli, and Isospondyli (in part). London: British Museum of Natural History. 544 pp.

Woodward AS. **1942**. The beginning of the teleostean fishes. *Annals and Magazine of Natural History* (Series 11) 9(60): 909–912.

Worthington J. **1905**. Contributions to our knowledge of the myxinoids. *The American Naturalist* 39(465): 625–663.

Wourms JP, Bayne O. **1973**. Development of the viviparous brotulid fish, *Dinematichthys iluocoeteoides*. *Copeia* 1973(1): 32–40.

Wright J. **2000**. A fish far from home? *The Fish-Watcher* (Newsletter of the East Coast Fish-watch Project) 3: 1–2.

Wu H-L, Shao K-T, Lai C-F, Chong D-H, Lin P-L. **1999**. *Latin-Chinese dictionary of fish names by classification system*. Taiwan: The Sueichan Press. 602 pp. [In Chinese]

Wucherer MF, Michiels NK. **2012**. A fluorescent chromatophore changes the level of fluorescence in a reef fish. *PLoS ONE* 7(6): e37913. https://doi.org/10.1371/ journal.pone.0037913

Wuitner È. 1936. Collection de Poissons de Mer, pêchés vers 1870 à l'Île de la Réunion (Mer des Indes) offerte par le baron Vidal de Léry. Annales de l'Association des Naturalistes Levallois-Perret 22 (for 1935): 59–76.

Wyrtki K. **1973**. Physical oceanography of the Indian Ocean (pp. 18–36). In: Zeitzschel B, Gerlach SA (eds) *The biology of the Indian Ocean*. Berlin: Springer. 555 pp.

# Y

Yabe M. 1985. Comparative osteology and myology of the superfamily Cottoidea and its phylogenetic classification. *Memoirs of the Faculty of Fisheries, Hokkaido University* 32(1): 1–130.

Yamada U. **2002**. Peristediidae (pp. 610–613, 1523). In: Nakabo T (ed) *Fishes of Japan with pictorial keys to the species* (English edition): Parts I and II. Tokyo: Tokai University Press. 1749 pp.

Yamada U, Yagishita N. 2013. Peristediidae (pp. 727–731, 1951–1952). In: Nakabo T (ed) *Fishes of Japan with pictorial keys to the species* (3<sup>rd</sup> edition). Hadano: Tokai University Press. 2428 pp. Yamakawa T. 1984. Family Trachichthyidae (slimeheads) (p. 109). In: Masuda H, Amaoka K, Araga C, Uyeno T, Yoshino T (eds) *The fishes of the Japanese archipelago*. Tokyo: Tokai University Press. 437 pp.

Yamakawa T, Machida Y, Gushima K. 1995. First record of the coral catshark, *Atelomycterus marmoratus*, from Kuchierabu Island, southern Japan. *Japanese Journal of Ichthyology* 42(2): 193–195.

Yamamoto M. 1982. Comparative morphology of the peripheral olfactory organ in teleosts. In: Hara TJ (ed) Chemoreception in fishes. *Developments in Aquaculture and Fisheries Science* 8: 39–59.

Yamane S, Okamura O. 1966. A review of the Pacific species of the berycoid fish belonging to the genus *Polymixia*. *Bulletin of the Misaki Marine Biological Institute* (Kyoto University) 9: 13–20.

Yamanoue Y, Matsuura K. 2003. Redescription of Neoscombrops cynodon (Regan, 1921), a senior synonym of Neoscombrops annectens Gilchrist, 1922 (Perciformes: Acropomatidae). Ichthyological Research 50(3): 288–292.

Yamanoue Y, Miya M, Matsuura K, Katoh M, Sakai H, Nishida M. 2008. A new perspective on phylogeny and evolution of tetraodontiform fishes (Pisces: Acanthopterygii) based on whole mitochondrial genome sequences: basal ecological diversification? *BMC Evolutionary Biology* 8(art. 212). http://dx.doi.org/10.1186/1471-2148-8-212

Yamanoue Y, Miya M, Matsuura K, Miyazawa S, Tsukamoto N, Doi H, Takahashi H, Mabuchi K, Nishida M, Sakai H. 2009. Explosive speciation of *Takifugu*: another use of fugu as a model system for evolutionary biology. *Molecular Biology* and Evolution 26: 623–629. http://dx.doi.org/10.1093/ molbev/msn283

Yamanoue Y, Miya M, Matsuura K, Yagishita N, Mabuchi K, Sakai H, Katoh M, Nishida M. 2007. Phylogenetic position of tetraodontiform fishes within the higher teleosts: Bayesian inferences based on 44 whole mitochondrial genome sequences. *Molecular Phylogenetics* and Evolution 45(1): 89–101. http://dx.doi.org/10.1016/j. ympev.2007.03.008

Yamashita T, Kimura S. **2001**. A new species, *Gazza squamiventralis*, from the east coast of Africa (Perciformes: Leiognathidae). *Ichthyological Research* 48(2): 161–166.

Yamashita T, Kimura S, Iwatsuki Y. **1998**. Validity of the leiognathid fish, *Gazza dentex* (Valenciennes in Cuvier and Valenciennes, 1835), with designation of a lectotype, and redescription of *G. minuta* (Bloch, 1795). *Ichthyological Research* 45(3): 271–280.

Yamashita Y, Golani D, Motomura H. **2011**. A new species of *Upeneus* (Perciformes: Mullidae) from southern Japan. *Zootaxa* 3107(1): 47–58. http://dx.doi.org/10.11646/ zootaxa.3107.1.3 Yano K, Ahmed A, Gambang AC, Idris AH, Solahuddin AR, Aznan Z. 2005. Sharks and rays of Malaysia and Brunei, Darussalam. Marine Fishery Resources Development and Managment Department, Southeast Asian Fishery Development Center. Kuala Terengganu, Malaysia. 557 pp.

Yano K, Matsuura K. 2002. A review of the genus Oxynotus (Squaliformes, Oxynotidae). Bulletin of the National Science Museum (Tokyo) (Series A) 28(2): 109–117.

Yano K, Miya M, Aizawa M, Noichi T. **2007**. Some aspects of the biology of the goblin shark, *Mitsukurina owstoni*, collected from the Tokyo Submarine Canyon and adjacent waters, Japan. *Ichthyological Research* 54(4): 388–398.

Yano K, Stevens JD, Compagno LJV. **2004**. A review of the systematics of the sleeper shark genus *Somniosus* with redescriptions of *Somniosus (Somniosus) antarcticus* and *Somniosus (Rhinoscymnus) longus* (Squaliformes: Somniosidae). *Ichthyological Research* 51(4): 360–373.

Yano K, Stevens JD, Compagno LJV. **2007**. Distribution, reproduction and feeding of the Greenland shark *Somniosus (Somniosus) microcephalus*, with notes on two other sleeper sharks, *Somniosus (Somniosus) pacificus* and *Somniosus (Somniosus) antarcticus. Journal of Fish Biology* 70(2): 374–390.

Yatou T. 1985. Scorpaenidae (pp. 562–575). In: Okamura O (ed) Fishes of the Okinawa Trough and the adjacent waters. The intensive research of unexploited fishery resources on continental slopes. Vol. II. Tokyo: Japan Fisheries Resource Conservation Association. [In Japanese and English]

Yatou T. 1985. Satyrichthys magnus Yatou, sp. nov.
(pp. 591–593). In: Okamura O (ed) Fishes of the Okinawa Trough and the adjacent waters. The intensive research of unexploited fishery resources on continental slopes. Vol. II.
Tokyo: Japan Fisheries Resource Conservation Association.

Yatou T, Okamura O. 1985. Satyrichthys isokawae Yatou et Okamura, sp. nov. (pp. 587–589). In: Okamura O (ed) Fishes of the Okinawa Trough and the adjacent waters. The intensive research of unexploited fishery resources on continental slopes. Vol. II. Tokyo: Japan Fisheries Resource Conservation Association.

Yogo Y, Nakazono A, Tsukahara H. **1980**. Ecological studies on the spawning behavior of the parrotfish, *Scarus sordidus* Forsskål. *Scientific Bulletin of the Faculty of Agriculture, Kyushu University* 34: 105–114.

Yokota L, Carvalho MR de. **2017**. Taxonomic and morphological revision of butterfly rays of the *Gymnura micrura* (Bloch & Schneider 1801) species complex, with the description of two new species (Myliobatiformes: Gymnuridae). *Zootaxa* 4332: 1–74. https://doi.org/10.11646/zootaxa.4332.1.1

- Yokota L, Lessa RP. **200**7. Reproductive biology of three ray species: *Gymnura micrura* (Bloch & Schneider, 1801), *Dasyatis guttata* (Bloch & Schneider, 1801) and *Dasyatis marianae* Gomes, Rosa & Gadig, 2000, caught by artisanal fisheries in northeastern Brazil. *Cahiers de Biologie Marine* 48(3): 249–257. http://dx.doi.org/10.21411/ CBM.A.7B3707B0
- Yoshida HO. **1979**. Synopsis of biological data on tunas of the genus *Euthynnus*. *NOAA Technical Report* NMFS Circular 429: 1–59.
- Yoshida HO. **1980**. Synopsis of biological data on bonitos of the genus *Sarda*. *NOAA Technical Report* NMFS Circular 432: 1–50.
- Yoshida HO, Otsu T. 1963. Synopsis of biological data on albacore *Thunnus germo* (Lacépède), 1800 (Pacific and Indian oceans). Species Synopsis No. 9. FAO Fisheries Biology Synopsis 52: 274–318.
- Yoshida T, Motomura H. **2016**. A new cardinalfish, *Verulux solmaculata* (Perciformes: Apogonidae), from Papua New Guinea and Australia. *Ichthyological Research* 64(1): 64–70. http://dx.doi.org/10.1007/s10228-016-0539-2
- Yoshigou H, Sasaki K, Yoshino T. 2004. The twin-spot butterfly ray, *Gymnura bimaculata* (Norman, 1925) (Elasmobranchii: Myliobatiformes; Gymnuridae): reconfirmation of the synonymy of the species *G. japonica* (Temminck and Schlegel, 1850). *Bulletin of the Hoshizaki Green Foundation* 7: 155–167.
- Yoshino T. **1984**. Parapercis schauinslandi (Steindachner) (p. 292). In: Masuda H, Amaoka K, Araga C, Uyeno T, Yoshino T (eds) The fishes of the Japanese archipelago. Tokyo: Tokai University Press. xxii + 437 pp.
- Yoshino T, Kishimoto H. 2008. Plotosus japonicus, a new eeltail catfish (Siluriformes: Plotosidae) from Japan. Bulletin of the National Museum of Natural Science (Tokyo) (Series A) Supplement No. 2: 1–11.
- Yoshino T, Kon T, Miura A. 1999. Morphological differences between *Beryx splendens* Lowe and *B. mollis* Abe (Teleostei: Beryciformes: Berycidae). *Bulletin Faculty Science*, *University of the Ryukyus* 67: 77–86.
- Yoshino T, Kon T, Okabe S. **1999**. Review of the genus *Limnichthys* (Perciformes: Creediidae) from Japan, with description of a new species. *Ichthyological Research* 46(1): 73–83.
- Yoshino T, Kotlyar AN. **2001**. World distribution of the Baloon Alfonsin, *Beryx mollis* (Pisces: Beryciformes: Berycidae). *Bulletin Faculty Science, University of the Ryukyus* 72: 119–123.
- Yusuf M, Kassem FNE. **2000**. Establishment of fishermen's cooperative societies (pp. 105–112). In: Hariri K, Krupp F (eds) *Conservation and sustainable use of biodiversity of Socotra Archipelago. Marine habitat, biodiversity and fisheries surveys and management.* Report of Phase II. 2

Vols. Senckenberg Research Institute, Frankfurt a.M., Germany.

- Ζ
- Zajonz U. **2006**. *Plectranthias klausewitzi* n. sp. (Teleostei, Perciformes, Serranidae), a new anthiine fish from the deep waters of the southern Red Sea. *aqua, International Journal of Ichthyology* 12(1): 19–26.
- Zajonz U, Akester S. 2005. The Pilot CZM Areas of Bir Ali Burum and Sharma – Jethmun. Guidelines to (Participatory Livelihood-centred) Fisheries Management (Delivery C02). Report to the Environment Protection Authority, Yemen and the World Bank-GEF (Grant No. TF 023492). MacAlister Elliott & Partners Ltd, U.K.; Scientific Council for Systems and Applied Science, Sana'a. 56 pp. + I Annex.
- Zajonz U, Deodatus F, Al-Harrani G. **2010**. Phase II Report (Progress Report Assessment & Analysis, Part 1). *Yemen Strategic Environmental Assessment of Coastal Zone Management*. Report to the World Bank and the Environment Protection Authority, Yemen. xi + 138 pp. + 7 Annexes.
- Zajonz U, Khalaf MA. **2002**. Inshore fishes of the Socotra Archipelago: diversity and community structure (pp. 237–296). In: Apel M, Hariri KI, Krupp F (eds) *Conservation and sustainable use of biodiversity of Socotra Archipelago. Marine habitat, biodiversity and fisheries surveys and management*. Final Report of Phase III. Senckenberg Research Institute, Frankfurt a.M., Germany.
- Zajonz U, Khalaf MA, Krupp F. **2000**. Coastal fish assemblages of the Socotra Archipelago (pp.127–170). In: Apel M, Hariri KI (eds) *Conservation and sustainable use of biodiversity of Socotra Archipelago: Marine Habitat, Biodiversity and Fisheries Surveys and Management*. Progress Report of Phase III. Frankfurt: Senckenberg Research Institute and Natural History Museum.
- Zajonz U, Klausewitz W. **2002**. *Neomerinthe bathyperimensis* sp. nov. from deep waters of the southern Red Sea. *Journal of Fish Biology* 61(6): 1481–1488.
- Zajonz U, Lavergne E, Bogorodsky S, Krupp F. **In press**. Biogeography of the coastal fishes of the Socotra Archipelago: challenging current ecoregional concepts. *PLoS ONE*.
- Zajonz U, Lavergne E, Bogorodsky SV, Saeed FN, Aideed MS, Krupp F. **2019**. Coastal fish diversity of the Socotra Archipelago, Yemen. *Zootaxa* 4636(1): 1–108 pp. https://doi.org/10.11646/zootaxa.4636.1.1
- Zajonz U, Lavergne E, Klaus R, Krupp F, Aideed MS, Naseeb FS. **2016**. The coastal fishes and fisheries of the Socotra Archipelago, Yemen. *Marine Pollution Bulletin* 105(2): 660–675. https://doi.org/10.1016/j. marpolbul.2015.11.025

Zajonz U, Saeed F. 2002. Inshore fish monitoring programme for the Socotra Archipelago (pp. 297–337). In: Apel M, Hariri K, Krupp F (eds) Conservation and sustainable use of biodiversity of Socotra Archipelago. Marine habitat, biodiversity and fisheries surveys and management. Final Report of Phase III. Senckenberg Research Institute, Frankfurt a.M., Germany.

Zama A, Yasuda F. **1979**. Annotated list of fishes from the Ogasawara Islands: Supplement 1, with zoogeographical notes on the fish fauna. *Journal of the Tokyo University of Fisheries* 65(2): 139–163.

Zambelli G. **1986**. Note sui Pholidophoriformes. VI. Pholidophorinae subfamiglia nuova del Triassico superiore. *Rivista Museo Civico Scienze Naturali "E. Caffi"* 10: 1–32.

Zehren SJ. **1979**. The comparative osteology and phylogeny of the Beryciformes (Pisces: Teleostei). *Evolutionary Monographs* 1: 1–389.

Zehren SJ. **1987**. Osteology and evolutionary relationships of the boarfish genus *Antigonia* (Teleostei: Caproidae). *Copeia* 1987(3): 564–592.

Zhu M, Yu X, Ahlberg PE, Choo B, Lu J, Qiao T, Qu Q, Zhao W, Jia L, Blom H, Zhu Y. **2013**. A Silurian placoderm with osteichthyan-like marginal jaw bones. *Nature* 502: 188–193. http://dx.doi.org/10.1038/nature12617

Zintzen V, Roberts CD, Anderson MJ, Stewart AL, Struthers CD, Harvey ES. **2011**. Hagfish predatory behaviour and slime defence mechanism. *Scientific Reports* 1(art. 131): 1–6. http://dx.doi.org/10.1038/srep00131

Zogaris S, Vidalis A, Fricke R. **2014**. First record of the Oman blenny *Oman ypsilon* Springer, 1985 (Teleostei: Blenniidae) from Kuwait and the Persian/Arabian Gulf. *Cahiers de Biologie Marine* 56(1) [2015]: 77–80. http://dx.doi. org/10.21411/CBM.A.72068B8E Zsilavecz G. **2001**. *Pavoclinus caerulopunctatus*, a new species of clinid fish (Perciformes: Clinidae) from South Africa. *J.L.B. Smith Institute of Ichthyology Special Publication* 66: 1–8.

Zsilavecz G. **2005**. *Coastal fishes of the Cape Peninsula and False Bay. A divers' identification guide*. Cape Town: Southern Underwater Research Group Press. 108 pp.

Zuffa M, Van Grevelynghe G, De Maddalena A, Storai T. 2002. Records of the white shark, *Carcharodon carcharias* (Linnaeus, 1758), from the western Indian Ocean. *South African Journal of Science* 98(7–8): 347–349.

Zugmayer E. **1911**. Diagnoses des poissons nouveaux provenant des campagnes du Yacht *Princesse Alice* (1901 à 1910). *Bulletin de l'Institut Océanographique* (Monaco) 193: 1–14.

Zugmayer E. **1911**. Poissons provenant des campagnes du yacht *Princesse Alice*. *Résultats des campagnes scientifiques accomplies sur son yacht par Albert 1er Monaco* Fasc. 35: 1–171.

Zugmayer E. **1913**. Diagnoses de stomiatidés nouveaux provenant des campagnes du yacht *Hirondelle II* (1911 et 1912). *Bulletin de l'Institut Océanographique* (Monaco) 253: 1–7.

Zugmayer E. **1913**. Die Fische von Balutschistan. *Abhandlungen der Akademie der Wissenchaften der Munchen* 26(6): 1–35.

Zuiew B. **1793**. Biga Mvraenarvm, novae species descriptae. *Nova Acta Academiae Scientiarum Imperialis Petropolitanae* 7 [for 1789]: 296–301, Pls. 1–7. COASTAL FISHES OF THE WESTERN INDIAN OCEAN

# SYSTEMATIC ACCOUNTS

# **MYXINIFORMES AND PETROMYZONTIFORMES:** LIVING REPRESENTATIVES OF JAWLESS FISHES, AND THEIR RELATIONSHIPS WITH OTHER VERTEBRATES

## Fabio Di Dario and Michael Maia Mincarone

The living representatives of jawless fishes, the hagfishes (Myxiniformes) and lampreys (Petromyzontiformes), comprise two distinct lineages of primitive-looking yet highly specialised craniates. These two groups were first combined in the Cyclostomes (later recognised as Cyclostomi or Cyclostomata) by Duméril (1806), based on a combination of some unusual morphological characters: a cartilaginous skeleton, absence of the opercle and branchial membranes, absence of paired fins, and the presence of a round mouth, at the tip of a cylindrical, naked and slimy body. With the advance of the cladistic paradigm in the 1970s, the shared morphological attributes of hagfishes and lampreys were best interpreted as symplesiomorphies (e.g., Løvtrup 1977). In addition, comparative morphological studies also revealed that lampreys might be phylogenetically closer to the Gnathostomata, the monophyletic group that comprises all jawed vertebrates. Lampreys and gnathostomes share, for instance, the presence of arcualia (singular arcualium). In lampreys the arcualia are rudimentary, bilaterally-paired cartilaginous structures on the dorsolateral side of the notochord. Arcualia are also known as neurapophyses, interdorsals or basidorsals, and in the gnathostomes they develop into the neural arches (Richardson et al. 2011). Whereas a single lamprey might have up to 130 arcualia (Tretjakoff 1926), those structures are completely absent in hagfishes and more basal clades of the Chordata, leading to the conclusion that the name Vertebrata should be restricted to the clade composed by the Petromyzontiformes and Gnathostomata. In that view (e.g., Janvier 1981, 2002), hagfishes are not vertebrates but members of a more encompassing clade — the Craniata. Lampreys and gnathostomes alone also share other key morphological and physiological features, such as a lateral line, a complex adenohypophysis, extrinsic eye muscles, true lymphocytes, and the nervous-system control of the heart (Janvier 2002).

In the 18th century and during most of the 19th, hagfishes and lampreys were considered 'degenerate representatives' of the 'Ostracodermi,' a non-monophyletic assemblage of bony armoured, jawless Palaeozoic craniates (Janvier 2008). Cyclostomes and ostracoderms were first combined in the 'Agnatha' by Cope (1889), who viewed this group as ancestral to the Gnathostomata. If hagfishes and lampreys were degenerate ostracoderms, then the absence of bones, enamel and dentine in those animals would be secondary, since those mineralised tissues are present in the skeleton of most ostracoderms. During the 1970s and 1980s, however, the Myxiniformes were gradually moved to the base of the Craniata, implying that their ancestors might never have had mineralised tissues after all, whereas the Petromyzontiformes were still perceived as an offshoot of the 'Ostracodermi' (e.g., Janvier 1978).

The debate as to whether or not the Cyclostomata comprises a monophyletic lineage is one of the most interesting topics in comparative biology, because it ultimately relates to a deeper understanding of vertebrate origins. The hypothesis of Cyclostomata monophyly was revived at the beginning of the 'molecular revolution' in systematics in the 1990s (Stock & Whitt 1992; Mallatt & Sullivan 1998; Kuraku et al. 1999). Since then, almost all phylogenetic reconstructions based on molecular data recovered the Cyclostomata as a monophyletic assemblage (e.g., Delarbre et al. 2002; Kuraku & Kuratani 2006; Heimberg et al. 2010). As succinctly stated by Janvier (2008: 1046), "These results leave morphologists and physiologists greatly perplexed," given the amount of morphological, physiological and biochemical synapomorphies that support a closer relationship between the Petromyzontiformes and Gnathostomata (recently summarised by Near 2009). If lampreys are actually related to hagfishes, then one would expect that the Cyclostomata might be the sister group of the remaining vertebrates, including fossil ostracoderms, since the Myxiniformes have been recognised as a basal clade in the Craniata for almost 50 years. However, even that view is now under dispute: it is possible that the Cyclostomata, if monophyletic, evolved from an ancestor morphologically more similar to ostracoderms than to gnathostomes (Heimberg et al. 2010; Janvier 2010), implying that living lampreys and hagfishes would indeed be "offshoots" of the 'Ostracodermi' after all.

Summing up, relationships among the three major living groups of craniates – the Myxiniformes, Petromyzontiformes and Gnathostomata – are still under debate. Difficulties in accessing the phylogenetic position of hagfishes and lampreys are in part related to their highly specialised feeding mechanisms and biology.

The 87 valid species of living hagfishes have an entirely marine distribution, inhabiting the cold or deep parts of oceans of both hemispheres. Lengths typically range from ~25–100 cm. The recently described *Eptatretus goliath* from New Zealand is the largest species recorded so far, reaching at least 127 cm TL and ~6.2 kg (Mincarone & Stewart 2006).

Hagfishes usually live in burrows in soft mud-bottom habitats, in depths ranging from the surface to at least 2 743 m, but a species of Eptatretus was photographed below 5 000 m in the tropical Pacific (Sumich 1992). In cold waters at higher latitudes some species live in shallower depths. Recent studies also indicate that hagfishes might live in association with rocky and deep coral reef habitats (Mincarone & McCosker 2004; Fernholm & Quattrini 2008) or hydrothermal vent sites (Møller & Jones 2007). Hagfishes have a nocturnal pattern of activity both in aquaria and in the wild (Worthington 1905; Fernholm 1974). Seasonal migration in response to habitat shifts or reproductive season is apparently unusual, and has been reported only for Eptatretus burgeri (Kobayashi et al. 1972; Fernholm 1974; Tsuneki et al. 1983; Nozaki et al. 2000). The reproductive biology of hagfishes is practically unknown, but might be inferred on the basis of collected specimens and from observations made on animals held in aquaria. Hagfishes have a single gonad, which is differentiated anteriorly as an ovary and posteriorly as a testicle in immature individuals. Adult individuals are functionally males or females, however. Fertilisation is probably external. Females produce ~20-30 relatively large eggs (15-40 mm long), laid in batches. Eggs are approximately sausage-shaped, heavily yolked, and covered by a horny shell. There is little information on the development of hagfishes, but it is probably direct, without a larval stage, with young emerging at ~45 mm TL (Gorbman 1983, 1997; Helfman et al. 2009).

Information on other aspects of the biology of hagfishes is also scarce, simply because most species are deepsea dwellers, and thus both hard to observe and to collect. Although hagfishes are jawless vertebrates, two eversible keratinised tooth plates located in the lower mouth chamber are used in conjunction with a single, curved, horny tooth at the roof of the mouth to grasp prey or perforate the tegument of dead or dying animals (Martini & Flescher 2002; Helfman et al. 2009; Zintzen et al. 2011). Hagfishes are well known for their scavenging habits, which are particularly relevant during the onset of so-called whale fall communities (Smith & Baco 2003). They might penetrate the carcass of large whales and other animals through natural orifices on their bodies, consuming their prey from the inside out. They also use a very elaborate mechanism for perforating the tegument, which probably evolved in order to compensate for the absence of paired fins, which provide a counterforce or anchor point for the biting action in aquatic gnathostomes. This mechanism is succinctly described as follows: the eversible tooth plates are used to firmly grasp the skin of the prey; a knot, produced by twisting the posterior portion of the hagfish, moves forward along its body and reaches the head region; that knot is then pressed against the tegument of the prey as a means of levering off a piece of flesh (Helfman et al. 2009). A species

of *Neomyxine* from New Zealand was observed in active predation using this knotting mechanism to remove live red bandfish (*Cepola hastii*) from their burrows (Zintzen *et al.* 2011). Hagfishes also prey on a large variety of invertebrates, and the force generated during their bite actually matches or exceeds that of several gnathostomes. Surprisingly, the Myxiniformes might be regarded as the oldest living lineage of chordate predators (Zintzen *et al.* 2011). It has also been recently discovered that hagfishes are able to absorb dissolved organic matter through the skin and gills. This ability probably originated as an adaptation associated with their scavenging lifestyle, allowing hagfishes to maximise sporadic opportunities for acquiring nutrients (Glover *et al.* 2011).

The eyes of hagfishes are small and subcutaneous, and also devoid of musculature and true lenses. Two pairs of sensory tentacles or barbels encircle the single nasopharyngeal duct at the tip of the head, and a third pair of barbels is located near the mouth (Janvier 2002; Martini & Flescher 2002); when searching for food, hagfishes keep their barbels in constant contact with the substrate (Zintzen et al. 2011). The inner ear is also relatively simplified and contains a single vertical semicircular canal. The number of branchial openings ranges from 5-14 in Eptatretus, to a single common opening in Myxine. An opening for the pharyngocutaneous duct on the left side of the body, behind the gill apertures, connects the pharynx to the exterior (Janvier 2002). A series of 57-207 ventrolateral pairs of slime glands can produce copious quantities of mucus, used mostly in defence against predation but also during hunting and probably reproduction (Gorbman 1997; Helfman et al. 2009; Zintzen et al. 2011).

Lampreys do not occur in the Western Indian Ocean or on the African continent, and for that reason only a brief summary of the group is given. The most distinctive morphological feature of an adult lamprey is the oral disc, which is a highly muscular structure. The mouth, at the centre of the oral disc, is encircled by a series of relatively well-developed keratinised teeth. The primary function of the oral disc is to provide a suction mechanism for the lamprey, which attaches itself to the skin of larger fish and, occasionally, cetaceans (Nichols & Tscherter 2011). A piston-like tongue, also covered with horny teeth, is used in conjunction with the teeth of the oral disc to abrade the skin of the prey, and blood, other body fluids and bits of skin, are ingested during the process. Adult lampreys are sometimes considered parasites, but since they harm other organisms only during feeding, they might be best regarded as highly specialised predators. Adults of about half of the known species never feed, and the Caspian lamprey Caspiomyzon wagneri feeds exclusively as a scavenger (Renaud 1997; Potter & Gill 2003).

Three families and ~48 species of lampreys are known (Renaud & Economidis 2010; Renaud 2011; Tutman *et al.*  2017; Riva-Rossi *et al.* 2020). Petromyzontidae is the most diverse family, with 8 genera and 43 species occurring in the Northern Hemisphere (Renaud 2011; Nelson *et al.* 2016; Tutman *et al.* 2017). Species of Geotriidae and Mordaciidae occur exclusively in the southern portions of South America and Oceania (Nelson *et al.* 2016). Two species of pouched lampreys (*Geotria australis* and *G. macrostoma*) are recognised in the Geotriidae, whereas 1 genus (*Mordacia*) with 3 species is recognised in the Mordaciidae (Renaud 2011; Riva-Rossi *et al.* 2020). Interestingly, the extinct but exquisitely preserved and fascinating *Priscomyzon riniensis* is known from the Late Devonian (~360 MYA) in a marine or estuarine environment of southern Africa (Gess *et al.* 2006).

# FAMILY **MYXINIDAE**

#### Hagfishes

Michael Maia Mincarone

Body eel-like, subcylindrical and slender, slightly deeper than wide at prebranchial, branchial and trunk regions; tail strongly compressed, paddle-shaped. Mouth jawless, with 2 sets of laterally everting keratinous cusps (or 'teeth') for biting and scraping, attached to the dental plate, which is attached to anterior end of dental muscle; single tooth on roof of mouth. No paired fins and no dorsal fin; midventral finfold present on trunk; caudal fin extends onto dorsal and ventral surfaces of tail. Single nostril above mouth, which leads respiratory water through the nasopharyngeal duct to pharynx and gill pouches. Pharyngocutaneous duct (PCD) on the left side confluent with (or posterior to) the last gill aperture. Three pairs of barbels on head: the first 2 pairs adjacent to nostril, the 3rd pair immediately adjacent to oral cavity. Vestigial eyes embedded in flesh. Body naked, with mucus-secreting slime glands, in segmentally arranged row, with pores opening along ventrolateral sides of body and tail. Endoskeleton cartilaginous; cranium a simple trough below brain, and postcranial skeleton comprising a large unconstricted notochord without centra or ribs. No separate larval phase. Attains 127 cm TL.

Hagfishes have an entirely marine distribution. Usually burrow in mud-bottom habitats, and found from surface to >5 000 m deep; however, some species appear to be closely associated with deepwater rocky areas, coral reefs, or hydrothermal vent habitats. Can produce copious amounts of slime or mucus, which might have a gill-clogging effect on predatory fish. Hagfishes have an important role in their benthic ecosystems: as scavengers, they are one of the most important mechanisms for the rapid clean-up and processing of carrion-falls; likewise, in areas subject to intensive commercial fisheries, they probably play a key role in the removal and recycling of discarded bycatch.

Six genera and 87 species; 2 genera and 4 species in WIO.



#### **KEY TO GENERA**

1a	External gill apertures 5–8 pairs	Eptatretus
1b	External gill apertures 1 pair	Myxine

# GENUS **Eptatretus** Cloquet 1819

External gill apertures 5–14 on each side of body; nostril short and not tubular. Distributed in all oceans, except eastern North Atlantic and Southern Ocean. About 53 species, 3 in WIO.

#### **KEY TO SPECIES**

1a	External gill apertures 5 pairs; prebranchial pores 12–15;
	tail pores 15–17 <i>E. profundus</i>

- 2a External gill apertures usually 6 pairs; anterior unicusps 8–10; total cusps 44–49; trunk pores 53–60..... *E. hexatrema*
- 2b External gill apertures usually 8 pairs; anterior unicusps 7; total cusps 38–40; trunk pores 63–68...... *E. octatrema*

## Eptatretus hexatrema (Müller 1836)

Sixgill hagfish

PLATE 1

- *Bdellostoma hexatrema* Müller 1836: 79 (Table Bay, Cape of Good Hope, South Africa).
- *Bdellostoma heterotrema* Müller 1836: 79, Pls. 1, 3, 6–9 (Table Bay, Cape of Good Hope, South Africa).

Bdellostoma forsteri var. heterotrema: Müller 1838; Paepke & Schmidt 1988. Bdellostoma forsteri var. hexatrema: Müller 1838; Paepke & Schmidt 1988. Bdellostoma cirrhatum: Günther 1870 [in part], 1880;

Putnam 1874 [in part].

Homea cirrhata: Garman 1899 [in part]; Dean 1904 [in part]; Fowler 1964 [in part].

Heptatretus hexatrema: Regan 1912; Barnard 1947\*; Smith 1950\*.

Bdellostoma (Eptatretus) hexatrema: Holly 1933\*.

*Eptatretus hexatrema*: Strahan 1975\*; SSF No. 1.1\*; Bianchi *et al*. 1999\*; Knapp *et al*. 2011; Mincarone 2017\*. Gill pouches 6 (rarely 7); external gill apertures 6 (rarely 7). Total cusps 44–49: anterior cusps 3 + 8–10, posterior cusps 2 + 9–10. Total slime pores 93–107: prebranchial pores 21–29, branchial pores 5–7, trunk pores 53–60, and tail pores 12–14. Percentage TL: prebranchial length 27–33%, branchial length 5–7%, trunk length 48–58%, tail length 12–14%, body depth at PCD 4–7%, body depth at cloaca 4–7%, tail depth 6–8%.

Body dark brown dorsally, paler ventrally; barbels usually with white tip; eyespots conspicuous; mouth, gill apertures and ventral finfold with pale margins; slime pores and caudal finfold the same colour as body. Attains 80 cm TL.

**DISTRIBUTION** Southern Africa: Namibia (Walvis Bay) in southeastern Atlantic, to South Africa (KwaZulu-Natal) in WIO.

**REMARKS** Common throughout its range, from shallow waters (usually associated with rocky reefs) to 400 m. Caught incidentally by baited traps, hooks and bottom trawls.

#### Eptatretus octatrema (Barnard 1923)

Eightgill hagfish

Heptatretus octatrema Barnard 1923: 439 (Agulhas Bank, off Cape St Blaize, South Africa); Barnard 1947.
Bdellostoma (Eptatretus) octatrema: Holly 1933.
Bdellostoma octatrema: Adam & Strahan 1963.
Homea octatrema: Fowler 1964.
Eptatretus octatrema: SSF No. 1.2; Knapp et al. 2011; Mincarone 2017\*.

Gill pouches usually 8 (rarely 7); external gill apertures usually 8 (rarely 7). Total cusps 38-40: anterior cusps 3 + 7, posterior cusps 2 + 7-8. Total slime pores 104-117: prebranchial pores 22-26, branchial pores 7-13, trunk pores 63-68, and tail pores 11-14. Percentage TL: prebranchial length 22-25%, branchial length 7-9%, trunk length 55-58%, tail length 10-13%, body depth at PCD 3-4%, body depth at cloaca 3-6%, tail depth 4-7%.

Body dark brown dorsally, paler ventrally; some specimens with whitish ventral band along the body; barbels with white tip; eyespots inconspicuous; rostrum, mouth, branchial apertures, cloaca and caudal fin with pale margin. Attains at least 42 cm TL.



Eptatretus octatrema, diagram of lateral view.

PLATE 1

**DISTRIBUTION** South Africa (Western Cape). In coastal waters of Kommetjie (near Cape Town) in Atlantic, and off Mossel Bay in WIO.

**REMARKS** May be naturally rare. Known only from a few specimens, caught in relatively shallow water, at 49–66 m. IUCN Red List conservation status Critically Endangered; but see Mincarone (2017) for further information.

# Eptatretus profundus (Barnard 1923)

Fivegill hagfish

PLATE 1

*Heptatretus profundus* Barnard 1923: 439 (off Cape Point, South Africa). Barnard 1947.

Bdellostoma (Eptatretus) profundum: Holly 1933.

Bdellostoma profundum: Adam & Strahan 1963.

Homea profunda: Fowler 1964.

Eptatretus profundum: Hardisty 1979.

*Eptatretus profundus*: Strahan 1975\*; SSF No. 1.3; Knapp *et al.* 2011; Mincarone 2017\*.

Gill pouches 5; external gill apertures 5. Total cusps 42–46: anterior cusps 3 + 8–9, posterior cusps 2 + 8–9. Total slime pores 81–86: prebranchial pores 12–15, branchial pores 4–5, trunk pores 48–52, and tail pores 15–17. Percentage TL: prebranchial length 19–28%, branchial length 5–7%, trunk length 53–69%, tail length 17–24%, body depth at PCD 7–10%, body depth at cloaca 6–9%, tail depth 7–11%.

Body purplish dark brown; eyespots conspicuous; tip of barbels and margin of mouth pale; gill apertures and ventral finfold variably with pale margins; caudal finfold the same colour as body. Attains 70 cm TL.

**DISTRIBUTION** South Africa: Lambert's Bay (Western Cape) in southeastern Atlantic to Cape Agulhas (Western Cape) in WIO.

**REMARKS** Poorly known; so far collected from 490–1 150 m. May be caught incidentally in bottom trawls.

# GENUS Myxine Linnaeus 1758

One pair of external gill apertures: efferent branchial ducts coalesce to form a common aperture on each side. Occurs in all oceans, except eastern Indian Ocean and western South Pacific. About 25 species, 1 in WIO.

## Myxine capensis Regan 1913

Cape hagfish

PLATE 1

*Myxine capensis* Regan 1913: 398 (Cape of Good Hope, South Africa); Fernholm 1981; Lloris 1986\*; SSF No. 1.4; Villanueva 1993; Bianchi *et al.* 1999\*; Knapp *et al.* 2011; Mincarone *et al.* 2011\*.

*Myxine glutinosa* (non Linnaeus 1758): Adam & Strahan 1963 [in part]; Fowler 1964; Hardisty 1979 [in part].

Gill pouches 7; external gill aperture 1 pair. Total cusps 40–44: anterior cusps 2 + 7–10, posterior cusps 2 + 7–9. Total slime pores 93–111: prebranchial pores 26–37, trunk pores 55–66, and tail pores 9–13. Percentage TL: prebranchial length 27–33%, trunk length 54–63%, tail length 11–14%, body depth at cloaca 4–6%, tail depth 3–6%.

Body pinkish grey; barbels, face, slime pores, and ventral surface of prebranchial region sometimes paler; ventral finfold pale. Attains 53 cm TL.

**DISTRIBUTION** Southern Africa: Namibia (Walvis Bay) in southeastern Atlantic, to South Africa and Mozambique (off Maputo) in WIO.

**REMARKS** Females mature at ~32–33 cm TL. Considered common; known from 88–675 m. Incidentally caught by baited traps, hooks and in bottom trawls; this species accounted for ~14% of the diet in 90 specimens of *Octopus magnificus* caught in its range.



# CLASS CHONDRICHTHYES





# **ORIGIN AND EVOLUTION OF THE CHONDRICHTHYES**

#### Marcelo R de Carvalho

Sharks, rays (batoids), and chimaeras (ratfishes, ghost sharks) form the class Chondrichthyes, the cartilaginous fishes referred to as 'sharks' - a group of jawed vertebrates (gnathostomes) whose origins extend back in time to the early Late Ordovician period, some 455 million years ago (MYA) (see geological timescale, Figure 1), as evidenced by very small dermal denticles probably of chondrichthyan affinity (Andreev et al. 2016; Sansom & Andreev 2019). Other fossil denticles attributed to sharks from the lower Silurian of Mongolia (Andreev et al. 2016) and China (Andreev et al. 2020) are only some 25 million years younger. The minute dermal denticles of these early sharks, collectively known as 'mongolepids', were deposited within shallow coastal marine environments. Their classification as sharks is, however, still debated and would be reinforced by the discovery of associated skeletons (Janvier & Pradel 2016; cf. Sansom & Andreev 2019).

Fossil shark teeth, among the most common vertebrate fossils, are known from the beginning of the Early Devonian (*Leonodus*, ~400 MYA). The earliest known shark skeletons appeared in the fossil record in the late Early Devonian, a little later than teeth (*Doliodus*; Maisey *et al.* 2019). Sharks are, therefore, one of the first groups of gnathostomes to have become established in early, warm Palaeozoic seas, even though the oldest 'fishes' (stem craniates, like the diminutive *Haikouichthys* and *Myllokunmingia* from China) appeared in the Early Cambrian some 65 million years earlier (Shu *et al.* 1999, 2003).

Presently, there are about 1 267 living species of chondrichthyans worldwide, comprised of some 540 species of sharks, 675 of rays (skates, guitarfishes, sawfishes, stingrays and electric rays), and 52 species of chimaeras (Last *et al.* 2016; Ebert & Dando 2021). However, a handful of new species (principally rays) are described every year, and dozens of 'known' undescribed species await scientific description. More than 20% of all valid chondrichthyan species were described in the past 20 years alone, indicating that much of their diversity may still be undetected. By comparison, the number of species of chondrichthyans described from fossils is close to twice the number of living species, with the vast majority of these known from isolated remains such as teeth, denticles or spines.

Extant sharks and rays and their immediate fossil relatives are grouped together as elasmobranchs, a group with greater modern diversity compared to any single time in their past, while living and fossil chimaeras, collectively known as holocephalans, were significantly more diverse in the Palaeozoic. Most fossil chondrichthyans are placed in either the Elasmobranchii or Holocephali, but many fossils (called stem sharks), including some very early ones, are phylogenetically placed at the base of the chondrichthyan phylogeny, before its divergence into elasmobranchs and holocephalans.

But what, exactly, is a chondrichthyan? The answer depends on whether one is referring to living species and their more closely related fossils (crown group), which includes elasmobranchs and holocephalans, or to the entire lineage, including the stem fossils, and which stretches back in time from the earliest recognisable fossil sharks to their origins.





Answering this question is difficult because many fragmentary and transitional gnathostome, shark or shark-like fossils display a mosaic of characters, especially those discovered in the past 30 years. Phylogenetically informative characters evolved intermittently, as opposed to 'all at once', and hence defining characters present in crown chondrichthyans evolved at different times on the stem. Add to this the scarce depositional environments associated with early sharks worldwide, and the fact that their skeletons have no significant endochondral ossification (i.e., are not made of true bone; as it turns out, a unique feature of sharks as bone is present in armoured jawless fishes), and one begins to get the picture that early, somewhat complete or even partially preserved sharks are hard to come by.



**Figure 2** Time-calibrated phylogenetic relationships of early chondrichthyans (modified from Janvier & Pradel 2016). Current phylogenies differ in the relationships of stem chondrichthyans (as a larger monophyletic group here, from *Pucapampella* to symmoriiforms), as there is much disagreement concerning early chondrichthyan evolution. Note that acanthodians are placed outside Chondrichthyes as an outgroup but current understanding is that the many acanthodian genera are scattered on the chondrichthyan stem below sharks. † denotes an extinct taxor; **1** Chondrichthyes; **2** Elasmobranchii + Holocephali. See Janvier & Pradel (2016) and the text for explanations.

There is still much debate around unequivocally defining early chondrichthyans, acanthodians and placoderms, the first gnathostome lineages dating from the Ordovician and Silurian (see Long et al. 2019; Maisey et al. 2019 - Figure 2). These three traditionally recognised early gnathostome lineages shared many generalised (plesiomorphic) features, muddling their distinctiveness. This is apparent, for example, in Doliodus (an early Devonian shark, known from portions of the head and pectoral-fin skeleton) that is phylogenetically placed low on the chondrichthyan stem even though it had acanthodian-like fin spines (Maisey et al. 2017). Early stem taxa may also have had specialisations that belie their supposed primitiveness, such as in stem sharks of the family Pucapampellidae from South America and Africa (Maisey et al. 2019). Phylogenetic studies attempting to answer questions about the origins of early gnathostomes have repeatedly shown that acanthodians and placoderms do not form natural groups. Acanthodian genera are scattered on the base of the chondrichthyan tree, and placoderms occupy basal positions on the gnathostome stem below the divergence of Chondrichthyes and Osteichthyes (Zhu et al. 2013; Brazeau & Friedman 2015; Coates et al. 2017; Long et al. 2019; but note that Long et al. 2015 and King et al. 2016 maintain placoderms as a natural group).

Current notions that acanthodians are phylogenetically scattered on the chondrichthyan stem (at the base of the chondrichthyan phylogeny), and the fact that many early shark fossils are known only from partial remains, have implications for the definition of Chondrichthyes. Given this scenario, chondrichthyans without acanthodians (the 'euchondrichthyans' of Janvier & Pradel 2016) are best characterised by tessellated mineralisation of the skeleton (Dean & Summers 2006; Janvier & Pradel 2016), in which mineral deposits form a tile-like mosaic overlying the cartilaginous matrix. This feature is present in crown- and stem-group chondrichthyans, but is not present in acanthodians phylogenetically positioned on the chondrichthyan stem (Maisey et al. 2021). Crown-group chondrichthyans also have pelvic claspers derived from radial pelvic-fin elements, easily observed in any living male shark, ray or chimaera, but many early fossil sharks do not have preserved pelvic fins or claspers so we cannot know exactly when this feature first evolved on the chondrichthyan stem. Claspers have also been found in placoderms, but these are differently structured and positioned and are therefore not homologous with shark claspers (Long et al. 2015; Trinajstic et al. 2015).

Many Palaeozoic sharks that are placed low on the chondrichthyan stem are known from three-dimensional remains, and their anatomy is actually better known than many Mesozoic sharks. Recognised stem sharks vary among phylogenetic studies, as do their relationships to each other. According to Janvier & Pradel (2016), these sharks include *Cladoselache* from the Devonian, the 'symmoriiforms' (e.g., *Stethacanthus, Falcatus* [see Figure 3], *Damocles, Akmonistion* [see Figure 4], *Cobelodus*, the South African *Dwykaselachus*, and the recently described *Kawichthys*, *Ozarcus* and *Ferromirum*), the xenacanths (such as *Orthacanthus*, *Triodus*, *Xenacanthus*), the 'ctenacanths' (e.g., *Cladodoides*, *Tamiobatis*, *Dracopristis*), and the earliest sharks known from neurocranial remains (*Doliodus*, *Antarctilamna*, *Pucapampella* and *Gydoselache*), among a few others. But Coates *et al.* (2017, 2018) and Frey *et al.* (2020) regard *Cladoselache* and symmoriiforms as stem holocephalans, and Hodnett *et al.* (2021) don't recognise a chondrichthyan stem at all and place these sharks as stem elasmobranchs. Hence, there is still much uncertainty concerning the early evolution of chondrichthyans.

Stem chondrichthyans are mostly Devonian and Carboniferous, but some (such as *Triodus* – see Figure 5) extended into the Triassic, contemporaneous with more modern elasmobranchs and holocephalans. Most of these stem sharks were demersal or benthic littoral forms, and also



Figure 3 A Falcatus falcatus from the Lower Carboniferous Bear Gulch limestone of Montana, USA. A greatly modified symmoriiform shark. Notice the enlarged, forward projecting cephalic spine in specimen (indicative of a male); claspers are also present in male. **B** Reconstructed adult male specimen; notice greatly developed lower lobe of caudal fin. **C** Artist impression of a male and female possibly in a mating ritual. (A H Zell, CC BY-SA 3.0; **B** from Lund 1985; **C** Smokeybjb, CC BY-SA 3.0)

rather small, measuring only up to one metre in length, but some were very large (Orthacanthus measured up to three metres and one ctenacanth from Texas may have been the size of a white shark). Almost all stem sharks were marine, but xenacanths lived predominantly in fresh and brackish waters. Many of these stem sharks deviated from the 'conventional' shark design, i.e., streamlined or fusiform with a heterocercal caudal fin, large, falcate pectoral fins, and multiple rows of large, sharp teeth, and some were downright bizarre compared to modern species. Stethacanthids had strange spine-brush complexes on or behind the head in mature males (in Akmonistion it was a massive, anvil-shaped structure larger than the head - see Figure 4), sometimes associated with enlarged denticles, possibly used either to ward off predators or to attract females; these sharks also had a whip-like extension trailing their pectoral fins. Xenacanths were eel-like, with



**Figure 4** A *Akmonistion zangerli* from the Lower Carboniferous of Scotland. This symmoriiform shark has also been considered a stem holocephalan in some phylogenetic studies. Specimen is male as per the large spine-brush complex and claspers. Notice also the slender head and trunk, large caudal fin (with massive ventral lobe), and elongate projections from pectoral fins. **B** Reconstruction. **C** Artist impression. (**A** USNM PAL508523; © Smithsonian Institution; **B & C** from Coates & Sequiera 2001)

elongate bodies and a low dorsal fin, sometimes with a double anal fin and diphycercal caudal fin, backward projecting cephalic or dorsal-fin spines with retrorse barbs (like a stingray tail sting), and unusual bicuspid teeth. Many Palaeozoic sharks did not survive the fateful Permian–Triassic extinction, but a few lineages that evolved in the Devonian, Carboniferous and Permian (e.g., hybodonts) flourished in the Mesozoic.

Palaeontological evidence of *bona fide* Palaeozoic holocephalans points to a Late Devonian (c. 380 million years) divergence between elasmobranchs ('euselachians' of Janvier & Pradel 2016 – see Figure 6) and holocephalans ('euchondrocephalans' of Grogan & Lund 2004; see also Grogan *et al.* 2012). The distinction between living sharks and rays (as a group) and chimaeras is morphologically straightforward, as evidenced by many characters in the neurocranium, mandibular and gill arches, claspers, sensory systems, and reproduction (Janvier 1996; Didier *et al.* 2012).

Elasmobranchs include all modern sharks and rays but also certain fossils, mostly Mesozoic, that are placed lower on the elasmobranch phylogeny and are therefore usually not classified within lower-level living groups. These sharks share with modern species features of the neurocranium, membranous labyrinth, gill arches and even fused pelvic girdles, and include the diverse hybodontiforms (e.g., *Tristychius, Onychoselache, Hybodus* [Figure 7], *Egertonodus*; Maisey *et al.* 2004; Coates & Gess 2007) that ranged from the Late Devonian to the Cretaceous, and the mostly Mesozoic (Jurassic and Cretaceous) 'synechodontiforms'.

Synechodontiforms (e.g., *Palidiplospinax*, *Paraorthacodus*, *Synechodus*) were closer to living forms than hybodonts as they had fully calcified vertebral centra (Maisey *et al.* 2004). They are often referred to as neoselachians together with



**Figure 5 A** A well-preserved specimen of a xenacanth shark, *Triodus* cf. *sessilis*, 60 cm TL, from the freshwater Permian outcrops of the Saar-Nahe basin, southwest Germany. Notice the eel-like, elongated body and dorsal fin, acute cephalic spine, claspers (specimen is an adult male), diphycercal caudal fin, and slender double anal fin. **B** Reconstruction. (**A** © D Herr-Heidtke; **B** adapted from Brignon 2015)

living sharks and rays, and some synechodontiforms may have been closer to living galeomorph sharks (Maisey *et al.* 2004; cf. Cappetta 2012). Hybodonts comprise a morphologically diverse group that were the dominant sharks of the Triassic and Jurassic. They ranged in size from a few metres (*Asteracanthus ornatissimus*, one of the largest Jurassic sharks) to only about 15 centimetres (*Lissodus*), and occupied diverse niches from large pelagic predators to smaller bottom-feeding durophagous



**Figure 6** Time-calibrated phylogenetic relationships of Elasmobranchii (modified from Janvier & Pradel 2016). Only major groups of fossil elasmobranchs are represented, and fossils that are nested within living groups are not shown for simplicity. This phylogeny summarises current understanding based on molecular characters, in which sharks are a natural group (but note that Echinorhiniformes is not recognised as a separate order here, and squatiniforms and pristiophoriforms are usually considered to be more closely related to each other than either is to squaliforms). In contrast, morphological phylogenies place the rays ('Batomorphs' in the phylogeny) within squalomorphs, and more closely related to squatiniforms and pristiophoriforms.

† denotes an extinct taxon; **1** Elasmobranchii; **2** Neoselachi; **3** crown-group or living elasmobranchs; 4 Squalomorphi; 5 Galeomorphi. See Janvier & Pradel (2016) and the text for explanations.

forms (*Tribodus*), both freshwater and marine. Hybodonts had massive jaws, large dorsal fins with ornate fin spines (with retrorse denticles and smooth ribs), conspicuous cephalic spines in males, and are among the best-known fossil sharks.

Crown-group elasmobranchs include all living sharks and rays and fossil relatives usually placed within living groups (at the ordinal, familial or generic levels). Researchers working on living elasmobranchs usually recognise nine orders of living sharks (Heterodontiformes, Orectolobiformes, Lamniformes, Carcharhiniformes, Hexanchiformes, Echinorhiniformes, Squaliformes, Squatiniformes and Pristiophoriformes) and four orders of living rays (Rhinopristiformes, Torpediniformes, Rajiformes and Myliobatiformes) (Last *et al.* 2016; Ebert & Dando 2021). Phylogenetic studies conducted over the past 25 years, based on robust morphological and molecular datasets, vary in the recognition or placement of certain orders (e.g., in recognising a separate order for the bramble sharks, *Echinorhinus*), but more fundamentally differ in



**Figure 7** Specimens of hybodont sharks. **A** Photograph of *Hybodus fraasi* from the Lower Jurassic of southern Germany (caudal fin not preserved); notice enlarged dorsal-fin spines. **B** Illustrations of the Late Carboniferous *Hamiltonichthys mapesi*, male, from Kansas, USA. Reconstruction in lateral view (top) showing massive jaws, dorsal-fin spines, cephalic spines and claspers; bottom image depicting anatomy in ventral view (note differences between male and female).

(A from Kriwet & Klug 2004; B from Maisey 1989)

recovering two conflicting patterns of relationships centred on the evolution of the rays. Analyses of morphological datasets conclude that rays are phylogenetically positioned high among squalomorph sharks (hexanchiforms, echinorhiniforms, squaliforms, squatiniforms and pristiophoriforms), and hence sharks are not monophyletic without their inclusion. Rays and pristiophoriforms (sawsharks) are considered to be each other's closest relative, and their next closest relative are the squatiniforms (angelsharks); this is known as the hypnosqualean hypothesis (Shirai 1992, 1996; Carvalho 1996; Carvalho & Maisey 1996).

Molecular data, however, point to an earlier derivation of rays from sharks on the elasmobranch stem and recognise living shark orders as a monophyletic group divided into two superorders, the galeomorphs (heterodontiforms, orectolobiforms, lamniforms and carcharhiniforms) and squalomorphs, equivalent in rank to batoids (Maisey *et al.* 2004). The conflicting results, therefore, centre on *when* batoids diverged from sharks—either before the split into the two extant shark superorders (molecular phylogenetic hypothesis), or after the appearance of squalomorphs (morphological data). As characterised by the great American zoologist William King Gregory, rays are indeed 'winged sharks' (Gregory 1935).

The oldest fossils of extant elasmobranch orders are teeth from the Early Jurassic assigned to heterodontiforms, hexanchiforms, orectolobiforms and batoids (Cappetta 2012; Maisey 2012). Articulated skeletons are known from the Middle (batoids) and Late Jurassic (squatiniforms, carcharhiniforms, heterodontiforms, hexanchiforms and lamniforms; Maisey et al. 2004; Thies & Leidner 2011; Maisey 2012), and there is an undescribed Early Jurassic holomorphic batoid from Holzmaden (Germany). Many articulated Jurassic, Cretaceous and early Cenozoic elasmobranchs are essentially modern in morphology and easy to assign to living suprafamilial groups (e.g., stingrays such as Asterotrygon, Heliobatis, Promyliobatis [Carvalho et al. 2004]; electric rays, e.g., Titanonarke; angelsharks, e.g., Pseudorhina; sixgill sharks, e.g., Notidanoides), whereas other more complete fossils are placed on the stem leading to certain modern orders (e.g., the morphologically 'transitional' Protospinax, a Late Jurassic squalomorph positioned on the stem leading to angelsharks and sawsharks [and batoids in the hypnosqualean hypothesis; Carvalho & Maisey 1996]; Sphenodus, on the stem of carcharhiniforms and lamniforms; Maisey et al. 2004; cf. Cappetta 2012). Clearly the Mesozoic record concerning articulated remains referable to living orders is still somewhat incomplete. Articulated basal neoselachians (synechodontiforms) are also known from the Early Jurassic, such as the morphologically unique and enigmatic Ostenoselache from Italy. However, essentially no Triassic fossils, teeth or otherwise, can be assigned to any living elasmobranch order.
The holocephalans were far more diverse in the Palaeozoic. The three living families pale in comparison to the Carboniferous holocephalan heyday when diverse and morphologically aberrant forms were common, especially in the Bear Gulch lagerstätte of Montana, an ancient warm water embayment at the time (Grogan & Lund 2004) (Figure 8). Fossil tooth plates and some articulated remains from the Mesozoic can be assigned to the three living families (Callorhinchidae, e.g., the Jurassic holomorphic *Ischyodus*; Chimaeridae, Cretaceous; Rhinochimaeridae, Triassic), but articulated Palaeozoic specimens are assigned to several dozen genera. Some of these were eel-like with elongated cephalic tenacula (the chondrenchelyid Hapagofututor - see Figure 9), or were shaped like frogfishes of the genus Antennarius (petalodontids), but the most bizarre stem holocephalans were the eugeneodontids with symphyseal, vertical tooth-whorls



**Figure 8** Time-calibrated phylogenetic relationships of Holocephali (modified from Janvier & Pradel 2016). Note that recent authors differ in the phylogenetic relationships of stem holocephalans, and that only a few groups of fossil holocephalans are represented (fossils that are nested within living groups are not shown for simplicity).

 $\dagger$  denotes an extinct taxon;  ${\bf 1}$  Holocephali;  ${\bf 2}$  Chimaeroidea. See Janvier & Pradel (2016) and the text for explanations.

on their lower jaws that perhaps acted to remove soft nautiloid animals from their shells. The Carboniferous genera of the Bear Gulch represent some of the most unusual fossil chondrichthyans known (iniopterygians had dorsally articulating pectoral fins), but *Echinochimaera*, also from Bear Gulch, clearly resembles modern holocephalans in overall appearance (Figure 10).

When we consider all the morphologically peculiar to outlandish Palaeozoic sharks and holocephalans, especially in the Carboniferous, it becomes easy to dismiss the popular notion that 'sharks have not evolved or changed in the past 400 million years'; clearly, modern chondrichthyans resemble their closer Mesozoic relatives much more. We can summarise chondrichthyan evolution by noting that, whether they originated in the Late Ordovician or Early Silurian (denticlebased forms), they began diversifying in the Devonian but truly exploded in diversity in the Carboniferous when dozens of families of sharks and holocephalans were the dominant fishes in many environments. The 'age of amphibians' was also the 'golden age of sharks'. This dominance was perhaps facilitated by major extinction events at the end of the Devonian that eliminated many competing lineages of fishes, such as placoderms, including the giant predator Dunkleosteus,



**Figure 9** *Harpagofututor volsellorhinus*, a small (up to 25 cm in length) and morphologically unique holocephalan from the Early Carboniferous Bear Gulch limestone of Montana, USA. **A** Female (top) and male (bottom) specimens. Arrows point to enlarged cephalic appendages in the male (with small hooks, believed to have been used to grasp females in copulation – see **B** for more detail). **C** Eel-like body in female. (**A** from D Chure, http://qvcproject.blogspot.com/2012/05/; **B & C** from Lund 1982)

and many acanthodians and armoured jawless fishes. Extinction played a role again a little later during the Permian– Triassic extinction, but in a less favourable way to sharks and holocephalans as many were eliminated along with some 95% of marine life. Some chondrichthyan lineages survived into the Mesozoic, however, when sharks again underwent gradual but significant diversification especially with the evolution of rays sometime in the Early Jurassic (rays are more diverse than sharks at the species level even though there are fewer ray orders). As noted, the beginning of the Jurassic saw the rise of some modern orders of sharks, and almost all elasmobranch orders were established by the end of this period and the early to mid-Cretaceous (the sole exception are the electric rays that date from the Paleocene).

Holocephalan diversity gradually became reduced to the three modern families toward the end of the Mesozoic, with most fossils being massive, hypermineralised tooth plates. There are, however, some remarkable Mesozoic holocephalan articulated fossils, all from Europe, such as the Early Jurassic Squaloraja (a flattened, guitarfish-like chimaera with long median and lateral rostral cartridges that formed a huge, triangular snout, a very large and upward-curved cephalic spine [or frontal clasper], huge eyes and short neurocranium see Figure 11) and Acanthorhina (with a sturgeon-like neurocranium and a very long dorsal fin spine). These articulated fossils, along with Myriacanthus (also known from Early Jurassic skeletons), are closely related to the living families (Stahl 1999). Ischyodus, however, known from some three dozen species, including I. quenstedti from articulated remains from the Late Jurassic of Bavaria, is classified as a basal



**Figure 10** *Echinochimaera meltoni*, a small holocephalan from the Early Carboniferous Bear Gulch limestone of Montana, USA. Even though similar to modern chimaeras, their bodies were covered with small dermal denticles. Specimens reached about 24 cm in length. Both male (**A**) and female (**B**) specimens are shown (redrawn from Lund 1988).

callorhinchid (Callorhinchus has three living species).

Many elasmobranchs did not survive the end-Cretaceous extinction (e.g., sclerorhynchid sawfishes), but representatives of living orders obviously did, and elasmobranch diversity steadily increased to present-day levels shortly after in the Cenozoic. Some of the more diverse extant groups date from earlier, in the Late Cretaceous (rajiforms, scyliorhinoids, dasyatids, triakids, many squaliform families), but other species-rich families are from the Eocene (e.g., carcharhinids, narcinids) or even as recent as from the Miocene (potamotrygonins). These records, however, only indicate earliest occurrences in the fossil record and not actually when these groups achieved their present-day diversity.

In terms of modern diversity, elasmobranchs show interesting trends. For example, much speciation has occurred in cold, deep waters and not just in tropical, warm-water regions. The three most diverse families predominantly occur in cold deeper waters of the continental slope and outer shelf regions: skates (Rajidae, ~160 spp.), softnose skates (Arhynchobatidae, 111 spp.), and deepwater catsharks (Pentanchidae, ~110 spp.). These three families alone account for close to one-third of all living elasmobranch diversity. Significant speciation has also occurred in warmer, more shallow seas, as seen in the families Dasyatidae (100 spp.) and Carcharhinidae (57 spp.), and relatively rapid speciation rates are also a factor in Neotropical freshwater ecosystems (potamotrygonin stingrays; 38 spp.).

The Cenozoic rise in diversity is presently being counterbalanced by threats not from catastrophic extinction events, such as recorded at the end of the Cretaceous, but from climate change brought on by global warming, habitat degradation, and overexploitation—in other words, by factors associated with unsustainable development. Having survived the past four global mass extinctions dating back to the Devonian, chondrichthyans now face an entirely different plight.



Figure 11 A Squaloraja polyspondyla, an Early Jurassic, morphologically modified holocephalan with a very enlarged and flattened head endowed with a head spine in males (frontal clasper).
B Photo of specimen from Osteno, Italy.
(A Illustration modified from Patterson 1965; B from Stahl 1999)

#### GLOSSARY

**acanthodian** – a class of early jawed vertebrates that resembled sharks in overall appearance, characterised by fin spines on their dorsal and paired fins (sometimes in between these), divided into three or four orders, and presently considered not to be monophyletic; acanthodians were entirely Palaeozoic ranging from the early Silurian to the Permian. **batoids** – refers to the monophyletic group that contains all of the orders of rays.

**cephalic tenacula** – appendage(s) on the head in male chimaeroids, both fossil and living, used to help grasp the female during copulation (at least in living species; function in fossils possibly the same). **crown chondrichthyans** – present-day sharks, rays, skates and holocephalans and their immediate fossil relatives.

**ctenacanths** – a group of mostly Palaeozoic sharks characterised by specific dentitions and fin spines, usually phylogenetically placed on the chondrichthyan stem below the elasmobranch/holocephalan divide.

**demersal** – living just above the bottom of the sea or a lake. **diphycercal** – a type of caudal fin with a subtriangular shape in which both dorsal and ventral lobes are symmetrical and vertebrae continue towards the extremity of the fin.

**durophagous** – eating of hard-shelled (exoskeleton- bearing) organisms as a major part of the diet.

endochondral ossification – a type of ossification (formation of bony tissue) in which osteoblasts replace hyaline cartilage with bone. galeomorphs – a diverse group of sharks that includes the orders Heterodontiformes (bullhead sharks), Orectolobiformes (carpet sharks), Lamniformes (mackerel sharks) and Carcharhiniformes (ground sharks). gnathostomes – vertebrates that have true jaws, or simply 'jawed vertebrates'; includes placoderms, acanthodians, chondrichthyans and osteichthyans.

**holomorphic fossil** – a fossil that is more or less 'completely' preserved; fossils that include a good portion of the skeleton and body outline. **homologous** – characters or features of organisms that have the same origins in phylogenetic history, and hence usually the same general position and structure (but not necessarily the same function, as in the swim bladder in fishes and lungs in tetrapods).

**hybodonts** – a diverse group of Palaeozoic and Mesozoic sharks that ranged significantly in size and morphology and that occupied many diverse habitats and niches, characterised by small cephalic spines in males, massive jaws and shoulder girdles, and characteristic fin spines and dentitions. Hybodonts are phylogenetically placed usually on the elasmobranch stem below neoselachians.

**hypermineralised tooth plates** – referring to the teeth of chimaeroids that undergo significant hardening by the excessive deposition of minerals to facilitate a durophagous diet.

**lagerstätte** – a sedimentary deposit that has extraordinarily preserved fossils, usually in abundance, and sometimes even with preserved soft tissues.

**membranous labyrinth** – the fluid-filled, adjoining canals of the inner ear housed in the bony labyrinth of the skull and responsible for the sense of equilibrium.

**mongolepids** – refers to an order of early fossil sharks known from isolated dermal denticles from the Ordovician or Silurian of Colorado (USA), Mongolia and China; includes the genera *Mongolepis, Solinalepis, Rongolepis, Teslepis, Shiqianolepis, Xinjiangichthys*, and Sodolepis.

**monophyletic group** – a group of organisms (or a clade) comprising the most recent common ancestor and all its descendants; a natural group; a group that is formally recognised in classifications. **natural group** – see *monophyletic group*.

**nautiloid** – refers to the mostly extinct cephalopod molluscs of the group Nautiloidea, with shells that are concave (appearing coiled) and that flourished in the Palaeozoic seas; a few genera, such as *Nautilus*, are extant.

**neurocranial** – the skeletal structure surrounding the brain and cranial sensory organs.

neurocranium - skull.

**pelvic claspers** – organs present in males for introducing semen directly into the reproductive tracts of females.

**phylogenetic studies** – studies that aim to reveal the evolutionary relationships (i.e. the historical patterns of descent) of fossil and living organisms.

**phylogenetically informative characters** – characters that reveal the historical patterns of phylogenetic relationships among organisms. **placoderms** – refers to a Palaeozoic (Silurian to Devonian) gnathostome group of armoured fishes with dermal bony plates on head and trunk, and which occupied numerous habitats and niches from large pelagic predators to small benthic forms; presently

considered to not be monophyletic and placed on the stem below the divergence between osteichthyans and chondrichthyans.

**plesiomorphic** – an ancestral (also referred to as 'generalised', underived or 'primitive') character state shared by groups of organisms (homologous for the group). Such characters cannot be used to define a group of organisms. For example, the presence of vertebrae cannot be used to define sharks.

retrorse (spines) - pointing or curved backward.

**stem chondrichthyan** – refers to a fossil shark that is placed very low on the chondrichthyan phylogeny, below the divergence of elasmobranchs and holocephalans.

**stem elasmobranch** – refers to a fossil shark that is placed very low on the elasmobranch phylogeny below the divergence of 'neoselachians', the level of more modern sharks and rays. **stem holocephalan** – refers to a fossil holocephalan that is placed very low on the holocephalan phylogeny below the divergence of more modern chimaeroids.

**symmoriiforms** – a diverse group of Palaeozoic sharks (or possibly basal holocephalans) known from many morphologically unique genera displaying strong sexual dimorphism.

**symphyseal teeth** – teeth at the junction between the two halves of the lower jaw.

**synechodontiforms** – an extinct order of sharks, mostly Mesozoic, which are phylogenetically close to living orders of elasmobranchs (sharks and rays) as they share, for example, calcified vertebral centra. **xenacanths** – a group of fossil, mostly Palaeozoic, sharks phylogenetically positioned on the chondrichthyan stem according to many authors, known for their morphological peculiarity and living mostly in freshwater habitats.

## COASTAL CHONDRICHTHYANS OF THE WESTERN INDIAN OCEAN

#### David A Ebert and James DS Knuckey

The class Chondrichthyes contains the cartilaginous fishes, which includes the sharks, batoids and chimaeras. The chondrichthyans are one of the most successful groups of fishes and are found in most marine ecosystems. Most of the species (~55%) occur on continental shelves, from the intertidal zone to 200 m deep, and to a lesser extent on insular shelves (Compagno 1990; Ebert & Winton 2010). The diversity of the shelf species is greatest in the tropics and lower in temperate seas. However, knowledge of regional chondrichthyan biodiversity is relatively uneven, with some areas, such as Australia, Europe and North America, being relatively well known, while some regions considered biodiversity hotspots, are poorly known. One of these regions is the Western Indian Ocean (WIO), which has the second most diverse chondrichthyan fauna in the world (at least 304 known species), exceeded only by Australia (with 322 species), although the former region has been comparatively little surveyed relative to the latter region (DA Ebert, personal database). The total number of WIO chondrichthyans represents about 24% of all known species in the class, with 164 shark, 131 batoid, and at least 9 chimaera species present.

Knowledge of the fauna within WIO is relatively uneven, with the number of chondrichthyan species widely differing across the five large marine ecosystems (Agulhas Current, Somali Current, Arabian Sea, Red Sea and Sri Lankan waters) as well as across the Indian Ocean offshore ecosystem. The southwestern Indian Ocean environment, primarily that affected by the Agulhas Current (including Comoros, Geyser Reef, Zélée Bank, Madagascar, Mozambique and South Africa), is the best known, with ~205 species, followed by the Arabian Sea ecosystem (including Persian/Arabian Gulf but not the Red Sea), with ~142 species (Jabado & Ebert 2015; Jabado et al. 2017). Faunal surveys conducted primarily off South Africa and southern Mozambique, and more recently off India, have increased our knowledge of these areas. The three other ecosystems (Somali Current, Red Sea and Sri Lankan waters) have fewer species reported to date: Somali Current has ~119 species (R Bennett, pers. comm.), Red Sea has 77 species (Bonfil & Abdallah 2004; Golani & Fricke 2015), and Sri Lankan waters (Ebert et al. 2017) have 124 species. The lower numbers reported for these marine ecosystems may be due to research constraints in these regions.

Despite limited scientific sampling, the coast of East Africa from Mozambique to the Arabian Sea, including the many islands and island chains in the region, has produced a remarkable number of new species. From 2001–2020 at least 50 new chondrichthyan species from WIO type localities have been described and named, and 2 new genera, with at least another dozen species awaiting formal description. The number of new species described compares well to the at least 88 species described from Australian waters during this same period. No other regions come close to the total number of species described over the past decade from Australian waters and the WIO. Furthermore, endemism is relatively high, especially among sharks and batoids, with respectively ~18.5% and 36.6% of these elasmobranchs known only from WIO. These numbers may increase with improved taxonomic resolution in some groups, especially the Myliobatiformes (devilrays, eagle rays and stingrays), Squalidae (dogfishes) and Pentanchidae (deepwater catsharks). The high degree of endemism also suggests that some of these species, especially coastal ones, may be at a high conservation risk.

Among elasmobranch species in WIO, approximately 16% have been assessed by the IUCN Red List of Threatened Species (http://www.iucnredlist.org/) as being Data Deficient, with another 2.4% not yet assessed. At the other end of the scale, at least 39 species (13.5%) are listed as Endangered and 24 species (8.3%) are considered Critically Endangered. However, the conservation assessments for many species at potential risk, especially those not considered to be charismatic, may be underestimated relative to some higherprofile, more appealing species. Many of the threatened species have a relatively restricted geographic range, which may be subject to intense anthropomorphic disturbances. Two case studies emphasise this point: Pliotrema warreni (sixgill sawshark) and Holohalaelurus favus (honeycomb catshark). Pliotrema warreni has a relatively restricted range, primarily from Cape Point, South Africa, to southern Mozambique, with its main distribution on the Agulhas Bank, an area of intense commercial trawl fisheries. Basic information on the biology and population structure of this species and the potential impacts of fishing, if any, are virtually unknown. Another species that may be of even more concern is H. favus, which was not formally named until 2006, but this once common species has not been seen in surveys since 1972.

Higher classification within the chondrichthyans has undergone considerable debate by taxonomists this century, especially with regard to the taxonomic and phylogenetic relationships of various groups (Shirai 1992; Carvalho 1996; Naylor *et al.* 2005, 2012; Aschliman *et al.* 2012; Straube *et al.* 2015). Recent classifications have suggested that the batoids are a sister group to the modern shark orders Pristiophoriformes

and Squatiniformes, and share a common ancestry with the Squaliformes (e.g., the hypnosqualean hypothesis). The two hypotheses break down largely between traditional morphological dichotomy of all modern sharks and batoids (Compagno 1973, 1977, 2001, 2005; Shirai 1992, 1996; Carvalho 1996) and newer molecular evidence (Douady et al. 2003; Naylor et al. 2005, 2012). In this book the classification of orders of sharks and rays is based on phylogenetic studies mostly carried out by Naylor et al. (2012, 2016) in the framework of the Tree of Life program which recognises 9 orders of sharks with 37 families, and the Rays of the World book (Last et al. 2016), both of which have been adopted in Eschmeyer's Catalog of Fishes. The batoid classification used here recognises 4 orders and 26 families (DA Ebert, personal database). The higher classification of chimaeras follows Didier et al. (2012) in recognising 1 order and 3 families. Within WIO, all 13 chondrichthyan orders, and ~33 shark, 19 batoid, and 3 chimaera families occur.

The **Hexanchiformes** (cow sharks and frilled sharks) comprises 2 families, 4 genera, and 7 species of morphologically unique sharks in that they are the only group to possess six or seven paired gill slits, a single dorsal fin, and an anal fin. Members of this group of moderately sized to very large sharks mostly occupy a benthopelagic habitat. Both families, all 4 genera, and 5 species occur in WIO. The Chlamydoselachidae (frilled sharks) are sporadically distributed off South Africa, but also with scattered records from islands and seamounts, including the Maldives and Madagascar Ridge. The Hexanchidae (cow sharks) are also wide-ranging but scattered, with records throughout WIO. The hexanchoids occur mostly along the outer continental shelf and upper slope, with the exception of *Notorynchus cepedianus*, a primarily coastal continental-shelf species.

The **Echinorhiniformes** (bramble sharks) is a small group consisting of the family, Echinorhinidae, with a single genus, and 2 wide-ranging benthopelagic species with a scattered worldwide distribution. The single WIO species, *Echinorhinus brucus*, is spottily distributed over a wide range, from South Africa, Mozambique, Maldives, Gulf of Aden, Oman and India.

The **Squaliformes** (dogfish sharks) is the third largest group of chondrichthyans, with 6 families, 23 genera, and at least 143 described species. The squaloids are a morphologically diverse group that contains some of the smallest and largest known species of sharks. Morphological characteristics of this order include 2 small to moderately large dorsal fins, which may be preceded by a fin-spine in some species, the absence of an anal fin, eyes without a nictitating membrane, and five paired gill slits located anterior of the pectoral fins. In WIO, members of this group inhabit both shallow and deepwater environs, but are generally replaced by Carcharhinidae (requiem sharks) and Sphyrnidae (hammerhead sharks) in warm-temperate to tropical seas. All 6 families, 17 genera and at least 42 species occur in WIO. The members of this group occupy a wide variety of habitats and are found from inshore to depths of 4 000+ m. Individual species may be demersal or epipelagic, with some species exhibiting pronounced ontogenetic changes in habitat during their lifetime. In WIO, 36 of the species in this group occur within 4 families: Squalidae, Centrophoridae, Etmopteridae and Somniosidae. The remaining 6 species in this group are in 2 relatively small families: Dalatiidae and Oxynotidae.

The Squalidae is a wide-ranging, benthopelagic family with 2 genera and 40 species recognised worldwide; both genera and 10 species occur in WIO. This family shows a high degree of endemism, with only 1 species (Squalus acanthias) known to occur in multiple geographic locations. The Centrophoridae is a taxonomically challenging group, with 2 genera and 19 species recognised worldwide, although the genus Centrophorus is currently undergoing an extensive taxonomic revision by the lead author (DA Ebert) and WT White (CSIRO, Hobart, Australia). Both genera and at least 9 species of this family have been recorded in WIO. This group of benthopelagic sharks is most diverse off South Africa and Mozambique, with 8 species reported from that region; the Gulf of Aden has 3 species, and the Aldabra and Seychelles islands have 4 species. The Etmopteridae is the largest family of squaloid sharks, with 5 genera and 52 species worldwide. Members are benthopelagic, mostly in deep water at 200-1 500 m, though some species may range to 4 500 m (Compagno et al. 2005). Two genera (Centroscyllium and Etmopterus) and 11 species occur in WIO. Most species in this family are regional endemics, except for 3 species (Etmopterus bigelowi, E. granulosus and E. pusillus) that are known from scattered records throughout the WIO region. The Somniosidae is one of the widest-ranging groups, occurring in most seas from the tropics to polar seas. Members of this group are benthopelagic, although some are pelagic. The group consists of 7 genera and 17 species, and contains some of the smallest and largest known species of sharks; 4 genera and 6 species occur in WIO. The Oxynotidae is a morphologically distinct group of 5 small- to medium-sized sharks in a single genus. Members are benthic-dwelling, at 50-650 m; only 1 species (Oxynotus centrina) is found in WIO <200 m. The Dalatiidae is a small group of 7 genera and 10 species of dwarf to medium-sized, wide-ranging sharks. Members of this group are mostly pelagic or benthopelagic and occur from near surface to >1 500 m deep; 5 genera, each with a single species, are represented in WIO.

The **Pristiophoriformes** (sawsharks) is a small group of poorly known, morphologically distinct sharks within a single family (Pristiophoridae), with 2 genera and 10 described species, with 4 species occurring in WIO. These slender-bodied sharks have a long flattened saw-like snout, lateral rostral teeth, a pair of long string-like barbels in front of the nostrils, two dorsal fins, and no anal fin. Most species have 5 paired gill slits, though the WIO genus *Pliotrema* has six pairs. These benthopelagic sharks occur primarily on continental shelves in temperate seas and usually along the upper continental slope in deeper tropical seas. Both genera occur in WIO; *Pliotrema warreni* is known off South Africa and Mozambique, *P. annae* from Zanzibar, and a deepwater species *P. kajae* from the Mascarene Plateau and off Madagascar; the African dwarf sawshark *Pristiophorus nancyae* is known off southern Mozambique only in deep water (>200 m).

The **Squatiniformes** (angel sharks) is a single family (Squatinidae) and genus (*Squatina*) of small-sized, rather morphologically similar, dorso-ventrally flattened sharks which are often misidentified; they are benthic inhabitants and often seen partially buried in the sediment. There are approximately 20 described species worldwide, occurring mostly in temperate to subtropical seas, with 1 endemic species (*Squatina africana*) in WIO.

The **Heterodontiformes** (bullhead or horn sharks) is a small group with 1 family (Heterodontidae) and single genus (*Heterodontus*) of 9 similar-looking species; 2 species occur in WIO. These are the only living sharks having both a spine preceding each dorsal fin and also an anal fin. Members of this group are primarily benthic-dwelling and nocturnal, tending to rest in caves and rocky crevices during the day but becoming quite active at night. They are found mostly nearshore, from the intertidal zone to ~100 m deep, but at least 1 species (*H. ramalheira*) is known to 275 m.

The **Orectolobiformes** (carpet sharks) comprises 7 families, 13 genera and 45 species worldwide. Members of this group can be distinguished by the presence of nasal barbels, two dorsal fins which are not preceded by a spine, and an anal fin. They are primarily benthic, occurring in tropical to warmtemperate seas, from the intertidal zone to ~200 m deep, usually on rocky or coral reefs, but 1 notable species (the whale shark *Rhincodon typus*) is oceanic. Four families (Hemiscylliidae, Ginglymostomatidae, Stegostomatidae and Rhincodontidae), 5 genera and 9 species are known in WIO, with the highest diversity found in the Arabian Sea and off India and Sri Lanka.

The Lamniformes (mackerel sharks) is a small but diverse group of 8 very distinctive families, 10 genera and 15 species. Although each of these families appears to be morphologically distinct, they are united by several unique features, including: a pointed snout, no dorsal-fin spine, a similar type of dentition, and oophagous reproduction. These sharks occupy a wide range of nearshore, coastal, deepsea and pelagic habitats, in temperate to tropical seas. All but one species have been recorded in WIO. These generally large-sized, wide-ranging sharks are most diverse off South Africa and Madagascar and in the Persian Gulf. The Lamnidae are mostly large, actively swimming, voracious sharks, and are among the most mediapublicised species. The 3 species of Alopiidae, with their distinctively long tail, and the monotypic Carchariidae, with the coastal raggedtooth shark Carcharias taurus, are common throughout the region, while Odontaspididae (2 species)

and the monotypic families Cetorhinidae, Megachasmidae, Mitsukurinidae and Pseudocarchariidae are less common.

The **Carcharhiniformes** (ground sharks) is the most diverse and largest group of shark-like cartilaginous fishes, comprising 10 families, 52 genera and 300 species. These sharks, despite a wide range in external morphological appearances, all have two spineless dorsal fins, an anal fin, a long mouth that extends behind the eyes which are protected by nictitating lower eyelids, and usually no oronasal grooves. Nine families (Carcharhinidae, Galeocerdonidae, Hemigaleidae, Pentanchidae, Proscylliidae, Pseudotriakidae, Scyliorhinidae, Sphyrnidae and Triakidae), 35 genera and 89 species are known in WIO. Most of these (54 species) are members of either the Carcharhinidae (30 species) or Pentanchidae (28 species), and may be very common locally.

The Pentanchidae (deepwater catsharks) is the largest family of sharks, comprising 11 genera and at least 111 harmless species worldwide; 5 genera and 28 species are reported from WIO of which 16 are deepwater species. This group may be subdivided into the colourful and mostly shallower species, and the uniformly coloured deepsea forms, which mostly lack any distinctive colour patterns. The number of catshark species is likely to increase with increased exploration, especially in deepsea habitats. The Scyliorhinidae (catsharks) has 10 genera and 71 species; 4 genera and 7 species in WIO. The Proscylliidae (finback catsharks) with 3 genera and 6 species, and Pseudotriakidae (false catsharks) with 3 genera and 5 species worldwide, both have 2 genera with 3 and 2 species respectively, in WIO, where they are known only from a few scattered, mostly deepsea locations. The Triakidae (houndsharks) is a large family consisting of 9 genera and at least 47 species worldwide, of which 6 genera and 9 species are found in WIO. These are mostly coastal species, several of which are regional endemics, plus there may be numerous others that are undescribed. The Hemigaleidae (weasel sharks) is a small group, with 4 genera and 8 species, with all genera and 5 species in WIO; most species in this family, except for the snaggletooth shark Hemipristis elongata, are relatively uncommon and poorly known. The Carcharhinidae (requiem sharks) is one of the largest shark families, comprising 11 genera and at least 59 species worldwide. The family is most diverse in coastal warm-temperate to tropical seas; 9 genera and 30 species are known to occur in WIO. The Sphyrnidae (hammerhead sharks) is a small group, with 2 genera and 8 species, which are mostly tropical to warm-temperate; both genera and 4 species found in WIO. The Galeocerdonidae (tiger sharks) is a family with one distinctive monotypic genus; it is circumglobal in warm-temperate to tropical seas, but not present in the Mediterranean Sea.

The **Torpediniformes** (electric rays) is a group of specialised batoids characterised by a thick, flabby, oval to round disc; a short, truncated snout; soft, loose skin without dermal denticles; a relatively short, thick tail; 0–2 dorsal fins; and a

well-developed caudal fin. Five families and 15 genera are recognised, of which 3 families and 7 genera occur in WIO; 2 of these genera (*Electrolux* and *Heteronarce*) are WIO endemics, and as currently known have very restricted geographic ranges. Worldwide there are ~69 species, with 19 known in WIO. Limited molecular data suggest this traditional classification is likely to change once more sampling has been undertaken.

The Rhinopristiformes (sawfishes, giant guitarfishes, guitarfishes, wedgefishes and banjo rays) is comprised of 5 families and 61 species; 4 families and 29 species known in WIO. The Pristidae (sawfishes) is a small group of very large batoids, having a flattened, greatly elongated rostrum with large transverse teeth along each side; 2 genera and 5 species occur worldwide, with both genera and 3 species found in the WIO. A recent global revision of the family has revealed the genus Pristis to have only 5 species rather than the 7 usually noted in the literature (Faria et al. 2013). This is likely the most critically endangered group of elasmobranchs, as their coastal habitat has been impacted from anthropogenic changes in many areas, and they are severely affected by fishing. Pristis are now considered locally extinct in several countries, including South Africa. The morphologically distinct Glaucostegidae (giant guitarfishes) has a single genus and 6 species worldwide, with 5 species in WIO; all species with IUCN Red List conservation status Critically Endangered. Glaucostegidae was formerly classified in Rhinobatidae, from which it differs in head shape and nasal flap configuration. The Rhinobatidae (guitarfishes) range in size from small to large, with 3 genera and 33 species worldwide, 2 genera and at least 16 species in WIO. The more shark-like, medium- to large-sized Rhinidae (wedgefishes) has 3 genera with 10 species, 2 genera and at least 4 species in WIO.

Four families are recognised within the order **Rajiformes** (skates), the Anacanthobatidae (legskates), Arhynchobatidae (softnose skates), Gurgesiellidae (pygmy skates) and Rajidae (hardnose skates). Members of this group usually have skin with prickles or thorns, often in a row along the midback; 0–2 dorsal fins; and long slender claspers in males. Worldwide, 39 genera are recognised within this order as presently arranged, with all families and 15 genera represented in WIO. This is the largest group of batoids, with 298 species, of which 32 occur in WIO, 9 of which are from deep water.

The **Myliobatiformes** (stingrays, devilrays and eagle rays) is the second-largest group of batoids and are characterised by their enlarged, expanded pectoral fins being completely fused to the head and trunk (except for the Myliobatidae), a disc ranging from diamond-shaped to subrhombic or lozenge-shaped; the presence or absence of a single dorsal fin; and a long whip-like tail in many species, with one or more stinging spines at the base. Worldwide there are 12 families, 56 genera and 244 species recognised; of these, 8 families, 22 genera and 51 species occur in WIO. It is likely that the numbers of species may increase with improved taxonomic resolution of some of these groups. In WIO, the family Dasyatidae (stingrays) is

most diverse, with 27 species, followed by the Myliobatidae (eagle rays) with 6 species and Mobulidae (devilrays) with 7 species. The families Gymnuridae (butterfly rays) and Rhinopteridae (cownose rays) have 4 species and 2 species, respectively, in WIO; the family Aetobatidae (pelagic eagle rays) has 1 genus with 5 species, 3 species in WIO; while the families Hexatrygonidae (sixgill stingray) and Plesiobatidae (deepwater stingray) each have a single wide-ranging species, including in WIO.

The **Chimaeriformes** (chimaeras, ratfishes or silver sharks) is a small group of unusual chondrichthyans comprised of 3 families, 6 genera, and 57 described species. The chimaeras are readily distinguished by their elongated tapering body, filamentous whip-like tails, smooth scales, large venomous dorsal-fin spine, broad wing-like pectoral fins, single external gill opening on each side, and open lateral-line canals, which appear as grooves on the head and along the trunk. Chimaeras are sluggish-swimming species usually found along outer continental shelves and upper slopes, although some species (especially members of the Callorhinchidae) are common in nearshore coastal waters. All 3 families, 6 genera and 9 or 10 species occur in WIO. However, of the Callorhinchidae (elephantfishes) only one species is represented in WIO at depths of <200 m (Callorhinchus capensis, off South Africa). The Chimaeridae (shortnose chimaeras or ratfishes) is the most species-rich family, with possibly 3 of 40 described species in 2 genera occurring in WIO, but all at depths exceeding 200 m, although there are several species in this region awaiting formal descriptions. The family Rhinochimaeridae (longnose chimaeras) is a wide-ranging group, with 3 genera and 5 species found in WIO, 2 species in 2 genera known from depths of <200 m, and a 3rd species likely.

#### ACKNOWLEDGEMENTS

We thank the following individuals for assistance and discussions on various aspects of this study: JJ Bizzarro, RH Bennett (WCS), M Bougaardt, LJV Compagno, PD Cowley, D van Beuningen (WCS), RW Leslie, GJP Naylor, N Straube and WT White. DAE acknowledges support from Moss Landing Marine Laboratories and The David and Lucile Packard Foundation.

#### GLOSSARY

hypnosqualean hypothesis – a hypothesis that proposes that the batoids (rays and skates) are derived sharks, based on many morphological phylogenetic studies.
nictitating membrane or nictitans – a clear, moveable inner eyelid found in sharks, and some reptiles and birds.
benthopelagic – fishes that live and feed near the bottom as well as in midwater.

## **ANATOMY OF SHARKS, RAYS AND CHIMAERAS**

The Chondrichthyes, comprising the sharks, batoids (rays and skates) and chimaeras, are characterised by a skeleton that is entirely cartilaginous. In adults the skeleton becomes more or less hardened by calcification. Shark skeletons also have far fewer components than the skeletons of bony fishes. The cranium is a single cartilaginous box to which the gill arches and jaws are not directly attached. The vertebral column consists of a series of vertebrae below an arch that protects the spinal cord. It runs from the posterior part of the skull to the tip of the upper caudal-fin lobe. The fins are variously supported by cartilaginous structures that articulate distally with keratinous, hair-like filaments, called ceratotrichia.

#### **BODY FORM AND FINS**

The torpedo-shaped body of most sharks and chimaeras has remained nearly the same for millions of years. They vary in length from the nearly 19-m whale sharks (Rhincodontidae) with a confirmed length of 18.8 m (McClain *et al.* 2015) to little lanternsharks (Etmopteridae), that are less than 30 cm. Rays and skates have very flattened (depressed) bodies.

The body of sharks and chimaeras is divided into three parts: head, trunk and tail. In batoids head and trunk are a single unit, their large pectoral fins stretch from the head, and tails are long, frequently thin, and sometimes very small. Several of the larger, fast-swimming sharks also have a fleshy keel on each side of the caudal peduncle to strengthen it and reduce turbulence.

Most sharks have two rigid dorsal fins, the first usually larger than the second, and several families have a stout spine in front of each dorsal fin. The second dorsal fin is absent in cow sharks (Hexanchidae) and frilled sharks (Chlamydoselachidae). Most sharks also have an anal fin. An asymmetrical caudal fin, in which the upper lobe is longer than the lower, is the main propulsion unit. In fast-swimming sharks, like the mako and the white shark (Lamnidae), the caudal fin is almost symmetrical. The paired pectoral and pelvic fins are nearly rigid and are used to provide lift (like the wings on an aeroplane) and control pitch (the 'nose up' or 'nose down' of an aeroplane). In the batoids (rays and skates) and in chimaeras the flexible pectoral fins are used for swimming.

















Ventral and lateral views of a generalised catshark to show morphology and measurements



Morphology and measurements of a generalised batoid



Morphology and measurements of a generalised chimaeroid

## BUOYANCY

Sharks and other chondrichthyans do not have a swimbladder to compensate for a heavy bony skeleton. Many sharks store lightweight oil, called squalene, in their (large) livers for buoyancy, while species such as the raggedtooth shark swallow air at the surface and use the stomach to provide buoyancy. Most batoids are demersal and have no buoyancy requirements. Large pelagic rays, e.g., mantas, use their pectoral fins like wings to provide lift when swimming.

#### THE SKIN



The skin of most sharks is covered with minute placoid scales or denticles, although some stingrays and electric rays lack these.

Scanning electron micrograph of shark scales (denticles) from a houndshark (Family Triakidae).

Unlike the scales of typical bony fishes, the dermal denticles of elasmobranchs (chondrichthyans that exclude the chimaeras) are essentially similar to their teeth, with a hard outer layer of enameloid over a core of dentine. Also, like the teeth, most denticles do not increase in size after they are completely formed; individual denticles and teeth are continually being replaced by newly formed and slightly larger ones during the lifespan of the shark. There is considerable variation in the shape and size of denticles. The surface of the denticles has series of tiny ridges and furrows, with the points of the denticles directed backwards. This accounts for the roughness of a shark's skin if stroked toward the head. The pattern and shape of denticles can often be used to identify certain species. However, the most important function of the ridged surface of the denticles is to produce laminar flow of water over the skin. This minimises the surface drag (or friction) of the water, and thus maximises swimming efficiency. Denticles also protect against parasites and fouling.

The skin of chimaeras is smooth and mostly covered with placoid scales.

In many skates the denticles are modified as thorns of various sizes, and occur in species-specific patterns on the dorsal surface of the disc, and along the tail. In stingrays (Dasyatidae) one or two thorns have become modified into sharp, serrated, venomous stinging spines (caudal sting), situated near the base of the tail.



#### Thorn patterning terminology and positions

## **HEAD MORPHOLOGY**

The mouth of nearly all elasmobranchs is situated on the underside of the head, with a pair of nostrils of varying form in front of the mouth. In whale and basking sharks the very large mouth is situated at the front of the head. Each nostril has a (generally) anterior incurrent and a posterior excurrent aperture, separated by an anterior nasal flap, which directs water into the nostril over highly sensitive olfactory membranes. In some sharks the nostrils are small, in some large, often with a barbel of varying length on the anterior margin.



The underside of the snout and sides of the head carry many ampullae of Lorenzini, visible as small black dots, which are extremely sensitive detectors of electrical fields generated by other animals.



## Teeth

The teeth are thought to be modified dermal denticles. Sharks and rays have a lifetime supply of teeth, as new ones are continually being formed on the inner surface of the jaws. The teeth are firmly anchored to a layer of tough



connective tissue, not fixed in sockets in the jaws (as in most bony fishes). When the outer teeth are lost, the next one in the row moves forward to take its place. In the few species in which tooth replacement rate has been measured, the outer teeth are replaced every one to two weeks; thus, a species that lives for 20 to 30 years produces thousands of teeth during a lifetime, and which is reflected in the numerous teeth in the fossil record. Chimaeras, which feed on molluscs and other invertebrates, have three pairs of large, permanent grinding plates that are worn down as they are used, and grow continuously.



The outer row (1) of shark teeth anchored at the front of the jaw cartilage eventually falls out to be replaced by row 2. Teeth rows follow the line of the outer jaw, while a series (or file) of teeth are at right angles to the jaw.



Cross-section through the jaw of *Carcharias taurus* (Family Carchariidae) to show a single file of teeth, where older teeth drop off and are continuously replaced by new teeth forming inside the mouth/jaw. Source: CFSA

Shark teeth are a useful taxonomic character, as their number and shape are more or less distinctive. A species may sometimes be identified with certainty from a single tooth. There are three basic types of teeth with three main types of function (cutting, grabbing and crushing): broad, triangular, blade-like teeth, as in white and tiger sharks, which are well adapted for slicing pieces from large prey; slender awl-like teeth as in raggedtooth sharks, which are suited for catching and holding bite-sized fishes and squid; and the blunt, molariform, pavement-like teeth of smoothhound sharks, rays and skates, which are generally used to crush hard-shelled molluscs and crustaceans.



#### Feeding

The common belief that all sharks are scavengers is incorrect. Many species are selective feeders: many species of Carcharhinidae feed largely on fishes but the tiger and Zambezi (or bull) shark are omnivorous; whereas *Heterodontus* species (Heterodontidae) mostly eat sea urchins. However, many species are opportunistic and will feed on other prey when it is available. The megamouth, basking and whale sharks are the only sharks that feed on plankton but may also ingest small fish.

The cookiecutter shark (*Isistius brasiliensis*) ambushes an unsuspecting prey, bites it and then rolls over, removing an almost circular chunk of skin and muscle tissue from the victim. The tawny nurse shark (*Nebrius ferrugineus*) uses its pharynx as a suction pump to suck prey out of nooks and crannies in the reef. Most rays and skates are benthic animals and feed on shrimps, crabs, shellfish and other invertebrates. Skates have small, sharklike teeth while rays have plate-like teeth adapted for crushing prey. All devil and manta rays (Mobulidae) are filter feeders and rely on plankton as their main food source.

Chimaeras, elephantfishes and ghost sharks are opportunistic feeders, feeding on small fishes and benthic invertebrates such as crabs, molluscs, marine worms and sea-urchins. Like sharks they use electroreception to locate prey in the dark. Their teeth are formed into two pairs of ever-growing tooth plates in the upper jaw and one pair in the lower jaw.

All elasmobranchs (and some primitive bony fishes) have a spiral valve in their intestine. In most species the valve is like a spiral staircase; in others it looks like a series of elongated cones, one inside the other, and, in the requiem and hammerhead sharks, it resembles a scroll of rolled paper. The spiral valve provides a much greater area of absorptive intestinal surface than a straight tube.



## Gill slits and spiracles

All sharks have five pairs of gill slits of varying size, except for Chlamydoselachidae with 6 pairs, and Hexanchidae with 6 or 7 pairs of gill slits. In fast-swimming sharks, such as the Lamnidae, the gill slits are situated in front of the pectoral fins and are long, sometimes extending to below the fins. In demersal and generally slower-moving sharks the gill slits are shorter and sometimes lie partially above the pectoral fins.

Batoids also have five pairs of gill slits (except a single deepwater family, the Hexatrygonidae, with six pairs), which are on the ventral surface of the head. Batoids and sharks have an opening of varying size situated behind the eye, known as a spiracle. In pelagic species (Lamnidae, Carcharhinidae and Sphyrnidae) the spiracle is absent or greatly reduced, but is well developed in benthic (bottom-living) sharks and batoids. In rays it is the major inlet for water to be pumped over the gills, for the mouth of these sedentary fishes is usually submerged in sand or mud, and a respiratory current of water coming in through the mouth would soon foul the gills.

Chimaeras have only a single pair of gill slits in front of the pectoral fins and no spiracle.



## Ears and eyes

As in other fishes, the ear of sharks and other chondrichthyans lies behind the eye and consists of three semi-circular canals at right angles to each other and connected to a sac-like structure, the sacculus. The ear is connected to the surface of the skin by a pore, and sensitive to gravity and vibrations (sound waves) in the water.

The eye of most sharks is round, with a round pupil. The pupil can also be crescent-shaped or a vertical, diagonal or horizontal slit. Some sharks have a moveable inner lower eyelid called a nictitating membrane or nictitans, which is variously developed in different species. In sharks with a large spiracle, the nictitans is absent or represented by a rudimentary groove along the lower edge of the orbit; while those without a spiracle (or a very small one) have a well-developed nictitans that can be pulled up to cover the eye completely. The white shark and whale shark lack a nictitans and protect their eyes by rotating them in the socket, exposing the tough sclera covering to protect the eye.

In benthic dwelling rays and skates (except the Mobulidae) the eyes are located more dorsally on the head and tend to look upwards. Chimaeras have prominent, large eyes on the sides of the head, also with a nictitans.

The retina of chondrichthyans contain both cones (for colour vision) and rods, which are sensitive to low light levels. They also have a reflective layer, a tapetum, behind the retina that bounces the light around inside the eye, thus improving their vision in low light and causing the eyes to glow in the dark.

## **REPRODUCTIVE ORGANS**

All chondrichthyans fertilise eggs internally. Males have a pair of modified pelvic fin radials that serve as intromittant organs, called claspers. These deposit sperm in a female's cloaca during mating. The claspers of chimaeras, which lie in front of the pelvic fins, can be retracted. Male chimaeras also have an additional, retractable sexual appendage on the forehead, called a tenaculum, which is thought to aid in holding onto a female during mating.

## **KEY TO FAMILIES OF CHONDRICHTHYANS**





9a	Rostrum soft and fla	bby, flexible ARHYNCHOE	ATIDAE (Softnose skates)
		and the second second	
9b	Rostrum semi-rigid,	to rigid and stiff, infle	exible 10
10a	Tail thin and filamentous, dorsal fins absent; pelvic fins with 2 lobes, anterior lobes usually longest ANACANTHOBATIDAE (Leaskates)		
10b	Tail thin but not filan with single lobe, or c length	nentous, 0–2 dorsal 1 deeply incised with 2 <b>GURGES</b>	fins; pelvic fins lobes of similar <b>IELLIDAE (Pygmy skates)</b>
10c	Tail not slender or file pelvic fins weakly to usually shortest	amentous, with 2 do deeply incised, with <b>R/</b>	rsal fins; anterior lobes AJIDAE (Hardnose skates)
	anterior pelvic- fin lobe leg- like and long	pelvic-fin lobes of similar length	anterior pelvic-fin lobe shorter than posterior lobe
	10a	10b	10c
11a	Disc thick and flabby organs visible throug	v, with prominent kic gh skin on underside	Iney-shaped electric 12
11b	Disc relatively thin, n kidney-shaped elect	ot flabby, without pr ric organs	rominent <b>14</b>
12a			
	Disc truncated or em	narginate anteriorly; 	rostrum reduced PEDINIDAE (Torpedo rays)
	Disc truncated or em	narginate anteriorly; 	rostrum reduced PEDINIDAE (Torpedo rays)
12b	Disc truncated or en	narginate anteriorly; TORI	rostrum reduced PEDINIDAE (Torpedo rays)
12b 13a	Disc truncated or en	narginate anteriorly; TORI orly; rostrum evident long and protractibi 	rostrum reduced PEDINIDAE (Torpedo rays) 



17a Head elevated above disc; eyes and spiracles on 17b Head not elevated above disc; eyes and spiracles on 18a Head with paired horn-like flaps at front MOBULIDAE (Devilrays) (Cownose rays) 20a Spiracle openings almost completely visible in dorsal view; rear edge of internasal flap deeply notched . AETOBATIDAE (Pelagic eagle rays) 20b Spiracle openings not visible in dorsal view; rear edge of internasal flap almost straight ...... MYLIOBATIDAE (Eagle rays) 21a Disc width >1.5 times disc length; tail short, slender, less than disc length; stinging spine relatively short GYMNURIDAE (Butterfly rays)

21b Disc width <1.5 times disc length; tail long and whip-like, greater than disc length; stinging spine relatively long ..... 22

 22a
 Caudal fin absent
 DASYATIDAE (Stingrays)

 22b
 Caudal fin present
 PLESIOBATIDAE (M) (Deepwater stingray)









- 24b Body low, nearly cylindrical in cross-section; dorsal fins low  $\ldots~25$



 25b
 Teeth dissimilar in both jaws, with prominent single cusp, possibly flanked by lateral cusplets; caudal fin with subterminal notch
 26





26bFlanks, underside of body and caudal fin without conspicuous<br/>black markings or photophores27







28b Head narrow and rounded conically; dorsal-fin spines absent from most genera (except *Squaliolus*) ...... DALATIIDAE (Kitefin sharks)



- 29a
   Gill openings 6 or 7 pairs; 1 dorsal fin
   30

   29b
   Gill openings 5 pairs; 2 dorsal fins
   31



30b Body fusiform; mouth subterminal; teeth not tricuspidate, dissimilar on upper and lower jaws; 6 or 7 paired gill openings, with first not extending across throat HEXANCHIDAE (Cow sharks)

~ 5



(M) = Monotypic

39a Caudal fin about as long as trunk of body. ALOPIIDAE (Thresher sharks) 39b Caudal fin much shorter than trunk of body ...... 40 40a Caudal fin heterocercal, not crescent-shaped; peduncle with weak lateral keels or without keels ..... 41 **40b** Caudal fin crescent-shaped; peduncle with strong lateral keels 43 40a 40b 41a Body stout; eyes relatively small; anal fin broad-based, not pivoting; peduncle with upper precaudal pit, but without 41b Body slender; eyes relatively large; anal fin narrow-based, pivoting; peduncle with upper and lower precaudal pits, and with weak lateral keels ...... PSEUDOCARCHARIIDAE (M) (Crocodile shark) \)))/ 42a Snout slightly depressed; dorsal fins subequal; 1st dorsal fin closer to pelvic-fin bases than to pectoral-fin bases ...... CARCHARIIDAE (M) (Raggedtooth shark) 42b Snout conical; 1st dorsal fin larger than 2nd; 1st dorsal fin closer to pectoral-fin bases than to 111// 42a 42b Teeth large, triangular, blade-like; gill slits large, but not 43a extending onto dorsal surface of head or nearly across throat, and without internal gill rakers ..... LAMNIDAE (Mackerel sharks)

- 43b Teeth minute, hook-shaped, and not blade-like; gill slits extremely large, extending onto surface of head and the first nearly extending across the throat, and with internal gill rakers ...... CETORHINIDAE (M) (Basking shark) Head with distinctive lateral, blade- or hammer-like 44a 45a First dorsal-fin base opposite or behind pelvic-fin bases ..... 51 46a No precaudal pits; caudal-fin dorsal margin without **46b** Precaudal pits present; caudal-fin dorsal margin wavy or not wavy ...... 49 precaudal pit 47a Labial furrows relatively long, with upper furrows extending partway or all the way anterior to level of jaw symphysis; rear teeth on dental bands not comb-like (III) 47b Labial furrows absent or very short and confined to mouth 48a Snout bell-shaped in dorsoventral profile, with deep groove in front of eyes; 1st dorsal fin more or less elongated, base closer to pectoral fins than to pelvic fins ..... PSEUDOTRIAKIDAE (False catsharks) 48b Snout rounded-parabolic or subangular in dorsoventral profile, without deep groove in front of eyes; 1st dorsal fin short, base closer to pelvic fins than to pectoral fins ..... PROSCYLLIIDAE (Finback catsharks)



- 49c Eyes round; upper lip folds reaching front edge of eyes; all fins not falcate; spiracles present; caudal peduncle with low lateral keel on either side ......GALEOCERDONIDAE (M) (Tiger shark)
- 50a Snout recurved and plough-shaped; 2nd dorsal fin falcate, short and high; caudal fin heterocercal CALLORHINCHIDAE (Elephantfishes)

- 50bSnout straight, not recurved or plough-shaped; 2nd dorsal fin<br/>long and low; caudal fin not heterocercal52
- 51a Narrow shelf-like cartilage extending from the chondrocranium to just above the eyes, forming a supraorbital crest (usually detected by touching the area) ....... SCYLIORHINIDAE (Catsharks)



51b No supraorbital crest present ......PENTANCHIDAE (Deepwater catsharks)



52a Snout short and blunt ..... CHIMAERIDAE (Chimaeras) [known in WIO only from >300 m]

52b Snout elongated and tapering ...... RHINOCHIMAERIDAE (Longnose chimaeras)

(M) = Monotypic

# ORDER HEXANCHIFORMES

## FAMILY CHLAMYDOSELACHIDAE

## Frilled sharks

David A Ebert

Medium-sized, distinctively elongate and slender-bodied; snout extremely short with truncated tip. Six paired gill slits with enlarged frilly interbranchial septa; ventral edge of 1st gill slit extends across throat as gular flap. Nostrils opposite jaw symphyses. Mouth terminal; teeth and jaw edges exposed when mouth closed; upper lip not expanded below level of tooth bases to form prominent flange and groove; lower lip not expanded anteriorly and laterally to the teeth series. Teeth alike in both jaws, with 3 cusps and 2 cusplets on crown; no small granular posterior teeth. Dermal denticles on lateral trunk with spike-like crowns; enlarged monocuspidate denticles at edges of mouth. One dorsal fin, very low, rounded and elongated, its insertion just before upper caudal origin. Pectoral fins rounded and smaller than pelvic fins. Pelvic fins with broadly rounded anterior margin and apex, inner margin not forming expanded clasper sheath in males. Pair of longitudinal ridges or tropeic folds on ventral surface of abdomen. Anal fin broad-based and rounded, larger than dorsal fin, its insertion ending at lower caudal origin. Peduncle short, compressed. Caudal fin with vestigial subterminal notch, and ventral lobe essentially absent.

Generally considered a deepsea group, although members are well known to ascend to within ~20 m of surface. No directed fisheries for these sharks. One genus with 2 species: one species wide-ranging, but patchy, in most seas, and the other appearing more restricted to the southwestern Indian Ocean and southeastern Atlantic.

## GENUS Chlamydoselachus Garman 1884

Diagnosis as for family. Two similar-looking species with differing vertebral counts. Unconfirmed whether both species co-occur off African coast, as records of *Chlamydoselachus* species prior to 2009 were given as if a single wide-ranging species.

#### **KEY TO SPECIES**

- 1aVertebrae 147; monospondylous-diplospondylous transition<br/>at about 18th vertebrae, just behind pectoral-fin tips; spiral<br/>valve turns 26–28; HL 17–18% TLC. africana

## Chlamydoselachus africana

Ebert & Compagno 2009

#### Southern African frilled shark

PLATE 2

*Chlamydoselachus africana* Ebert & Compagno 2009: 3, Figs. 1–4, 6 (off Cunene River, Namibia); Ebert 2013; Ebert *et al.* 2013\*. *Chlamydoselachus anguineus*: Smith 1951, 1967\*; Bass *et al.* 1975; Compagno *et al.* 1989\*, Compagno *et al.* 1991.

Chlamydoselachus sp. A: Ebert 1990\*; Compagno et al. 2005\*.



Chlamydoselachus africana, 102 cm TL, immature female (Namibia). Source: Bass et al. 1975

Slender, eel-like, with body compressed beyond pectoral-fin region; 6 paired gill slits. One dorsal fin. Mouth terminal; teeth tricuspid, similar in both jaws, with 3 long, slender, smooth-edged cusps, and small pointed cusplet between each cusp; upper jaw with paired medial teeth, shaped similar to anterolateral teeth, but reduced; lower jaw with single medial tooth undifferentiated from anterolateral teeth; teeth in both jaws curved inwards and set on broad base projecting behind and interlocking with tooth base behind it; tooth count 24–30/23–27. Dermal denticles on lateral trunk lanceolate, single-cusped, with flattened bases; crowns widely spaced and slightly projecting above body, with 4 longitudinal ridges from base to cusp. Total vertebrae 147, precaudal vertebrae 94, monospondylous vertebrae 18; spiral valve turns 26–28.

Body dark brown, but with thin membranous mucous covering, which upon being rinsed off after death reveals dark grey colour; preserved specimens pale brown to grey. Attains ~125 cm TL.

**DISTRIBUTION** Southern Africa: Angola to Namibia in southeastern Atlantic, and scarce records from South Africa and around Cape Peninsula in WIO.

**REMARKS** Externally morphologically similar to common frilled shark C. anguineus, but smaller, with proportionally longer head, greater head height and width, and proportionally longer pre-pectoral and predorsal-fin lengths. Viviparous with yolk sac; litters of 3 or more; size at birth 40-45 cm TL. Males mature at 92 cm TL, but female size at maturity unknown (117 cm TL for an immature female). Little-known; occupies benthic, epibenthic, and possibly pelagic habitat on upper continental slope, at 100-1 400 m. Hydrographic and substratum data collected on a specimen caught off southern Namibia found it occurred in a low-oxygen, high-nutrient zone with soft substratum; one record of note is of a frilled shark caught off a pier in South Africa (Eastern Cape). Feeds mostly on other sharks, squaloids and scyliorhinids, bony fishes and cephalopods; capable of swallowing relatively large prey (measuring 1/3-1/2 their own body length) in much the same way snakes are able to consume large prey items. Likely caught as bycatch in bottom-trawl and longline fisheries.

## Chlamydoselachus anguineus Garman 1884

Frilled shark

PLATE 2

*Chlamydoselachus anguineus* Garman 1884: 47, Fig. (Japanese seas [probably southeastern Honshu]); Ebert 1990, 2003, 2013\*; Compagno *et al.* 2005\*; Ebert & Compagno 2009; Ebert *et al.* 2013\*. Long, slender, eel-like, with compressed body beyond pelvic-fin region; 6 paired gill slits. One dorsal fin. Mouth terminal and snake-like; teeth tricuspid in both jaws. Tooth count 19–28/21–29; total vertebrae 160–171, precaudal vertebrae 93–102, monospondylous vertebrae 72–75; spiral valve turns 35–49.

Body uniformly dark brown, brownish grey or brownish black (closely resembles *C. africana*, but differs in counts and measurements). Attains 196 cm TL.

**DISTRIBUTION** North Atlantic and Pacific oceans (wide-ranging, patchy); unconfirmed in southwestern Indian Ocean.

**REMARKS** Viviparous with yolk sac; litters of 2–15 (average 6); size at birth ~55 cm TL. Males mature at 118–163 cm TL, and females at 130–150 cm TL. Reproduces all year in deep water off Japan, with mating thought to occur in springtime, from March to June; gestation period long, probably 1–2 years. Feeds on a variety of other sharks, bony fishes, and epipelagic and deepwater cephalopods. Occupies benthic, epibenthic and occasionally pelagic habitat, over offshore continental and insular shelves and upper slopes, at 20–1 440 m, but occasionally taken at surface. Harmless species of little importance to fisheries, but taken as a bycatch, especially in bottom trawls and midwater shrimp trawls; used for fishmeal and human consumption. Has been kept in aquaria in Japan.

## FAMILY HEXANCHIDAE

#### Cow sharks David A Ebert

Medium- to very large-sized (~1.4–5 m TL), elongated, moderately slender to stout cylindrical body and flattened head; snout moderately long, pointed to broadly rounded. Six or 7 paired gill slits. Eyes about opposite symphyses of mouth. Mouth subterminal; teeth dissimilar in upper and lower jaws: anterolateral teeth in lower jaw much larger, compressed and comb-like, with more cusplets than teeth in upper jaw, and posterior teeth smaller, granular. One spineless dorsal fin, relatively high, angular, short; its insertion well before upper caudal origin by at least length of dorsal-fin base. Pectoral fins angular, larger than pelvic fins. Pelvic-fin anterior margin nearly straight and apex narrowly rounded. Anal fin narrowbased and angular, its insertion well anterior to lower caudal origin. Peduncle moderately long and cylindrical. Caudal fin with well-developed subterminal notch, and lower lobe weak to moderately developed. No longitudinal keels on abdomen.

Reproductive mode yolk-sac viviparity. Mostly deepwater inhabitants of outer continental shelf and upper slope, insular shelves and slopes, and submarine canyons, to at least 2 500 m deep, and found near bottom or well above it; however, some species also occur in shallow bays, close inshore and near surface.

Worldwide in boreal and cold-temperate to tropical seas. Three genera and 4 species, all in WIO.

#### **KEY TO GENERA**

1a	Six paired gill slits	Hexanchus
1b	Seven paired gill slits	2
2a	Eyes very large; head extremely narrow and pointed; body plain, without spots; medium-sized, maximum	
	~1.4 m TL	leptranchias
2b	Eyes small; head broad and rounded; body usually w scattered small black spots and sometimes white spots	ith ots;

## GENUS Heptranchias Rafinesque 1810

Head acutely pointed, compressed, and rounded or vertically oval in section at eyes. Seven paired gill slits. Eyes large. Mouth very narrow and angular-parabolic. Lower jaw with 5 rows of long and low, comb-like anterolateral teeth, with a few short mesial cusplets and an abruptly high cusp, and up to 7 or 8 distal cusplets in adults, increasing and then decreasing in size distal to cusp. Peduncle elongated. Lateral-line canal closed. No spots on body; dorsal fin and caudal-fin upper lobe with black tips (may be faded or absent in adults but prominent in young). One species.

#### Heptranchias perlo (Bonnaterre 1788)

Sharpnose sevengill shark

PLATE 2

Squalus perlo Bonnaterre 1788: 10 (France, Mediterranean Sea). Heptranchias perlo: Bass et al. 1975\*; SSF No. 2.1\*; Compagno et al. 1989\*; Ebert 1990\*, 2013\*; Ebert et al. 2013\*.

Diagnosis as for genus. Medium-sized, with narrow head and large eyes. Tooth count 23–43/20–33; total vertebrae 141–159, precaudal vertebrae 72–94 (calcified and with obvious double cones); spiral valve turns 18–22.

Body uniformly pale grey to olive dorsally, paler to white ventrally; no spots on body; dorsal fin with black blotch and caudal-fin upper lobe black-tipped (prominent in young, faded or absent in adults). Attains 137 cm TL.

**DISTRIBUTION** All tropical and temperate seas (wideranging, somewhat sparse), except eastern North Pacific. WIO: South Africa (Western Cape to KwaZulu-Natal), southern Mozambique, Comoros, Madagascar, Mascarene Seamounts, Mauritius to Seychelles, Aldabra, Maldives, Gulf of Aden, southwestern India (Kollam), Sri Lanka and southwestern Mediterranean Sea.



Heptranchias perlo, 68 cm TL, immature male (S Mozambique). Source: Bass et al. 1975

**REMARKS** Litters of 6–20; size at birth ~26 cm TL. Males mature at 70-78 cm TL, and females at 89-90 cm TL. Appears reproductively active during entire year off Japan and in Mediterranean Sea. Relatively uncommon; primarily deepwater, benthic and epibenthic, over continental and insular shelves and upper slopes, at 27-720 m (mostly deeper than 100 m, and possibly to ~1 000 m), but occasionally close inshore. Probably strong-swimming as indicated by firm skin and muscles and by vigorous, agile, snake-like activity when captured. Feeds on a wide variety of small to moderately large demersal and pelagic bony fishes, small elasmobranchs such as catsharks (Pentanchidae and Scyliorhinidae), lanternsharks (Etmopterus, Etmopteridae), skates (Rajidae), cephalopods and crustaceans. Narrow jaws and teeth with prominent, narrow, needle-sharp cusps and cusplets suggest they are well equipped for grabbing, holding and swallowing small soft-bodied prey, but less capable of dismembering large-bodied tough-skinned prey. Of minor importance commercially, but affected by a wide variety of demersal fisheries and generally caught as bycatch of bottom-trawl fisheries or deepwater fisheries using bottom longlines. IUCN Red List conservation status Near Threatened as population declines may have occurred in some areas, such as southern Mozambique, where deepwater demersal trawling for shrimp and bony fishes has been operational during the past few decades.

#### GENUS Hexanchus Rafinesque 1810

Head narrowly or broadly parabolic in dorsoventral view, depressed and transversely oval in section at eyes. Six paired gill slits. Eyes small or large. Mouth moderately to very wide, parabolic or arcuate. Lower jaw with 5 or 6 rows of comb-like anterolateral teeth, long and low in adults but higher in young; mesial edges smooth in young but with serrations in adults; low to moderately high cusp, and 8–10 distal cusplets in adults, decreasing in size distal to cusp. Peduncle short to elongated. Lateral-line canal closed. Spiral valve turns 22–39. Body without spots or with irregular brown spots; fins not blacktipped. Size at maturity 123 cm to  $\sim$ 5 m TL. Two species, both in WIO.

#### **KEY TO SPECIES**

- 1b Snout long, pointed and narrow; eyes large; lower jaw usually with 5 rows of large, comb-like anterolateral teeth; distance between dorsal-fin insertion and upper caudal origin much greater than length of dorsal-fin base; medium-sized, maximum ~1.8 mTL

#### Hexanchus griseus (Bonnaterre 1788)

Bluntnose sixgill shark

PLATE 3

*Squalus griseus* Bonnaterre 1788: 9 (France, Mediterranean Sea); Forster *et al.* 1970.

*Hexanchus griseus*: Bass *et al.* 1975\*; Bass 1978; SSF No. 2.2\*; Compagno *et al.* 1989\*; Ebert 1990\*, 1994, 2002, 2003\*, 2013\*; Anderson & Ahmed 1993; Ebert *et al.* 2013\*.

Stout, heavy-bodied and broad-headed; snout bluntly rounded to rounded-angular in dorsoventral view; preoral length shorter, 4.3–5.4% TL. Eyes small. Mouth broad, U-shaped, its width more than twice length. Lower jaw with 6 rows of blade-like, comb-shaped anterolateral teeth. Pectoral fins broadly triangular. Peduncle short and stout. Caudalfin postventral margin weakly concave to straight and not subdivided, and lower lobe poorly developed at all stages. Tooth count 26–46/20–36; vertebrae 118–148; spiral valve turns 35–39.

Body grey or tan to blackish, with conspicuous paler lateral line; sometimes with darker spots on flanks; underside often paler than dorsal surface in newborns, and colouration more uniform in larger juveniles and adults. Attains ~5 m TL.



**DISTRIBUTION** Wide-ranging in boreal, temperate and tropical seas, except possibly Arctic and Antarctic oceans. WIO: Somalia, Mozambique, South Africa (Western Cape to KwaZulu-Natal), Madagascar, Comoros, Aldabra, Réunion, Maldives, India and Sri Lanka.

**REMARKS** Litters of 47–108 (relatively large); size at birth 61-74 cm TL. Males mature at ~3.1 m TL, and females at ~4.2 m TL. Pupping grounds apparently occur on upper slope and outer continental shelf, with one such area known from southern Namibia. Based on size, may be long-lived but it has yet to be aged. Mostly deepwater, benthic and pelagic, over continental and insular shelves and slopes and off seamounts and underwater ridges, found close to or well off bottom. Occurs at surface in tropics, and close inshore near beaches, at heads of submarine canyons, and in bays in cold-temperate waters, but down to at least 2 500 m on upper continental slope. May show equatorial submergence in tropics as with some other deepwater sharks, and may not normally penetrate warm tropical inshore waters, although it has been known to rise to surface offshore in response to fishing operations. Young of this species often found close inshore, occasionally in enclosed bays; adults, especially males, often found in water >200 m deep in temperate areas, although adults and subadults may enter shallow water in open or enclosed bays with adjacent deepwater canyons. Often associated with areas of upwelling and high biological productivity. Off southern Africa, smaller individuals (<120 cm TL) feed primarily on cephalopods, and secondarily on bony fishes and some chondrichthyans, while larger individuals (~120-200 cm TL) feed primarily on cephalopods, bony fishes and chondrichthyans, plus a small marine mammal component. Very large individuals (>200 cm TL) feed mostly on marine mammals (Cape fur seals and cetaceans) and large pelagic teleosts, with a smaller component of cartilaginous fishes and cephalopods. Apparently feeds on the bottom and well above it; may be able to take large active prey, such as eared seals, cetaceans and large pelagic bony fishes, by means of stealthy stalking, which is possibly aided by its cryptic colouration. Caught largely as a bycatch of other fisheries, but once targeted for human consumption off the Maldives

where a fishery for this species peaked but subsequently collapsed. Used fresh, frozen, and dried and salted for human consumption, and for fishmeal and oil. Implicated in attacks on humans, but also the subject of ecotourism dives in some areas. IUCN Red List conservation status Near Threatened; assumed to be vulnerable to overfishing and not able to sustain intensive targeted fisheries.

#### Hexanchus nakamurai Teng 1962

Bigeyed sixgill shark

Hexanchus griseus nakamurai Teng 1962: 30,

Fig. 5 (Keelung [Cheng-gong], Taiwan).

*Hexanchus vitulus* Springer & Waller 1969: 160, Figs. 1–4 (near Bimini, Bahamas); Forster *et al.* 1970; Bass *et al.* 1975\*; Compagno 1984\*; SSF No. 2.3\*; Compagno *et al.* 1989.

Hexanchus nakamurai: Ebert 1990\*, 2013\*; Compagno et al. 2005\*; Ebert et al. 2013\*.

Moderate-sized, slender-bodied and narrow-headed; snout narrowly rounded. Eyes large. Mouth V-shaped, narrow, its width ~1.5 times length. Lower jaw with 5 rows of bladelike, comb-shaped anterolateral teeth. Pectoral fins narrowly triangular and semi-falcate. Anal fin smaller than dorsal fin and set just behind it. Peduncle long and slender. Caudal-fin postventral margin deeply notched in adults and subdivided into upper and lower parts; lower lobe short and strong in adults but weak in young. Tooth count 25–33/9–32; vertebrae 155; spiral valve turns 22–28.

Body pale grey to brown, darker dorsally and paler ventrally; fins sometimes dusky and usually with trailing white edges. Attains 180 cm TL.

**DISTRIBUTION** Sparsely but widely distributed in warmtemperate and tropical seas, except possibly eastern Pacific. WIO: Kenya, South Africa (KwaZulu-Natal), northern Madagascar, Nazareth Bank, Aldabra, Mauritius, Geyser and Zélée coral banks (NE Mozambique Channel) and southwestern Mediterranean Sea (rare).



Hexanchus nakamurai, 155 cm TL, mature male (South Africa). Source: Bass et al. 1975

PLATE 3

**REMARKS** Litters of 13–26; size at birth 40–43 cm TL. Males mature at 123–157 cm TL, and females at 142–178 cm TL; fecundity is much lower in this species than in *H. griseus*. Little-known, primarily deepwater, over continental and insular shelves and slopes, at 90–621 m, usually on or near bottom, but occasionally moving to near surface or inshore in tropics. Likely feeds on small- to medium-sized bony fishes, cephalopods and crustaceans. Uncommonly taken on line gear and in trawls; of relatively minor importance to fisheries, probably used for human consumption and as fishmeal. Apparently uncommon to rare, thus affected by a variety of bottom fisheries; often misidentified as juveniles of *H. griseus*. IUCN Red List conservation status Near Threatened.

## GENUS Notorynchus Ayres 1855

Head broad, bluntly pointed, depressed and transversely oval in cross-section at eyes. Seven paired gill slits. Eyes small. Mouth wide and arcuate. Lower jaw with 6 rows of combshaped anterolateral teeth, with low mesial serrations and cusplets in young, and with serrations, low cusp, and 5 or 6 distal cusplets in adults, decreasing in size distal to cusp. Peduncle short. Precaudal vertebrae not calcified and without obvious double cones. One species.

#### Notorynchus cepedianus (Péron 1807)

Sevengill shark

PLATE 4

Squalus cepedianus Péron 1807: 337 (Adventure Bay, Tasmania, Australia). Notorynchus cepedianus: Bass et al. 1975\*; SSF No. 2.4\*; Compagno et al. 1989\*; Ebert 1990\*, 1991, 1991, 1996, 2002, 2003\*; Ebert et al. 2013\*.

Diagnosis as for genus. Tooth count 21–42/20–37; vertebrae 123–130; spiral valve turns 14–17.

Body olive brown to grey, usually covered with numerous small black spots and sometimes with white spots, but occasionally plain; dorsal fin and caudal-fin dorsal lobe with black tips in young individuals, but lost in larger specimens. Attains 3 m TL. **DISTRIBUTION** Wide-ranging in most temperate seas (antitropical), but not in North Atlantic and Mediterranean Sea. WIO: South Africa (Western Cape and Eastern Cape).

**REMARKS** Litters of 80–100; size at birth 34–45 cm TL. Males mature at 150-180 cm TL, at 4-5 years; females mature at ~220 cm TL, at 11-21 years; estimated maximum lifespan 30 years. Reproductive cycle may take 2 years, with gestation period ~1 year plus a year's recovery period during which the next batch of eggs mature. Females give birth to young in shallow bays in springtime. Neritic and benthic, over continental and insular shelves, from surfline and intertidal zone to at least 136 m deep, but mostly above 50 m, and often less than 1 m deep and at surface. Common in shallow bays and close inshore, with larger individuals ranging into deeper water offshore and deep channels in bays. Apparently coordinates its movements in bays with tidal cycle by moving in with a tidal rise and out with its fall, and moves out of shallow bays and estuaries during rainy winter season when salinities drop. Powerful, high-level predator and rather broad-spectrum in its feeding habits, concentrating on marine vertebrates (primarily chondrichthyans, bony fishes and marine mammals). Active and strong-swimming, often found cruising steadily and slowly near bottom but sometimes at surface. Apparently social, often found in groups or aggregations; most active at night, in early morning before sunrise and on overcast days, and tends to move into shallower water in low light and retreat into deeper water with increasing light. In South African waters it may aggregate in patches of turbid brown water caused by phytoplankton blooms, runoff or sediment disturbance. Its large size, local abundance, and high-quality flesh have made this species the subject of fisheries. A small-scale fishery occurred in Namibia (Lüderitz Lagoon) where it was common in the late 1980s and early 1990s. Also caught as a bycatch of various inshore fisheries, including demersal trawls targeting bony fishes and gillnet shark fisheries. Can respond vigorously and assertively when provoked; has been involved in a few unprovoked attacks on divers and swimmers, and has behaved aggressively towards spearfishers. In recent years, an ecotourism industry has sprung up in South Africa (False Bay) where sevengill sharks congregate in relatively clear water.



Notorynchus cepedianus, 209 cm TL, mature female (South Africa). Source: Bass et al. 1975

# ORDER ECHINORHINIFORMES

## FAMILY ECHINORHINIDAE

#### Bramble sharks

David A Ebert

Large-sized with heavy cylindrical body, covered with coarse spiky denticles or enlarged tack-like denticles; head flattened; snout short and broad. Spiracles very small and far behind eyes. Fifth gill slit much larger than preceding four. Nostrils wide-spaced, internarial width much greater than nostril width. Mouth large, broadly arched, with thin non-papillose lips; lip furrows short, confined to mouth corners, not elongated posteriorly into post-oral grooves or anteriorly into preoral grooves. Teeth in both jaws compressed, low-crowned, broad, blade-like, forming saw-like edge, not arranged in quincunx pattern but in single, flat, non-overlapping series. Two small, spineless, posteriorly placed dorsal fins; 1st dorsalfin origin over or behind pelvic-fin origins. Pectoral fins low, broadly rounded, with anterior margin moderately large and about equal to or smaller than prespiracular length, and rear tip rounded and not elongated. Pelvic fins as large as or slightly larger than pectoral fins, and more than twice size of dorsal fins. No anal fin. Peduncle compressed, very short; no lateral keels or precaudal pits.

Found from inshore to deep water; circumglobal but sparse, in cold-temperate to tropical seas. One genus with 2 species.

## GENUS **Echinorhinus** Blainville 1816

Diagnosis as for family. Body and fins with large, whitish, scattered and conspicuous thorn-like denticles. First dorsal-fin

base above pelvic fins. Tooth counts 20–26/20–26; vertebrae 86–102; spiral valve turns 8–16. Body pale to medium grey, grey-brown, brownish or blackish dorsally, often paler ventrally, sometimes with red or black spots or blotches on body, and fin edges blackish. Size range ~1.5–4.5 m TL or more. Two wide-ranging species, 1 in WIO.

#### Echinorhinus brucus (Bonnaterre 1788)

#### Bramble shark

Squalus brucus Bonnaterre (ex Broussonet) 1788: 11 (L'Ocean [North Atlantic]).

*Echinorhinus obesus* Smith 1838 (Cape of Good Hope, South Africa). *Echinorhinus brucus*: SFSA No. 44\*; Nair & Lal Mohan 1971; Silas & Selvaraj 1972; Bass *et al.* 1976\*; Compagno 1984\*; SSF No. 6.1\*; Compagno *et al.* 1989; Anderson & Ahmed 1993\*; Akhilesh *et al.* 2013; Ebert 2013\*; Ebert *et al.* 2013\*.

Diagnosis as for genus. Dermal denticles on body and fins (in adults and juveniles >90 cm TL) varying from small to very large, with many large, widely spaced thorn- or bucklerlike denticles, with bases not stellate and >1 cm wide; some groups of denticles may fuse to form plates >2.5 cm across. Ventral surface of snout and area around mouth with closeset small denticles in young, and large conspicuous denticles or thorns in larger juveniles and adults; newborns densely covered with small and high denticles with narrow semistellate bases. Tooth count 20–26/21–26; vertebrae 102; spiral valve turns 12–16.

Body dark purplish grey, sometimes with red or black blotches on back and sides. Attains at least 3.2 m TL, possibly 3.9 m TL.



*Echinorhinus brucus*, 122 cm TL, female (Namibia). Source: Bass *et al.* 1976

PLATE 4

**DISTRIBUTION** Eastern Atlantic, Indo-Pacific and Mediterranean Sea (wide-ranging but sparse). WIO: South Africa (Eastern Cape and KwaZulu-Natal), southern Mozambique, Gulf of Aden, Oman, India (southwestern coast and Gulf of Mannar) and Maldives.

**REMARKS** Viviparous with yolk sac; litters of 10–52; size at birth 40–54 cm TL. Males mature at 150–198 cm TL, and females at 189–230 cm TL; does not appear to have defined breeding season in Indian waters. Diet includes benthic and littoral bony fishes, smaller sharks and crustaceans. Sluggish and bottom-dwelling, rarely encountered, usually singly. Found in shallow (<18 m) water inshore in cold-temperate areas and places with upwelling, and possibly intertidal

zone, but primarily in deep water on continental and insular shelves and upper slopes, at 200–900 m. Along cold-temperate west coast of South Africa and Namibia it moves inshore up submarine canyons and is regularly caught by shore anglers on rod and reel and by small-scale commercial fishers. Relatively unimportant to fisheries, except off southwestern India where it is taken in large numbers; fished in mid-19th century off Namibia in small-scale targeted line fishery; occasionally caught off South Africa as bycatch of demersal trawls. Sporadic and largely unused bycatch of other fisheries, including those for other sharks, but sometimes used for fishmeal and the liver oil used medicinally by traditional healers in South Africa. Has never been known to bite people in the water. Seldom kept in captivity. IUCN Red List conservation status Endangered.

# ORDER SQUALIFORMES

## FAMILY SQUALIDAE

#### Dogfishes

David A Ebert

Small- to moderately large-sized with cylindrical body, and abdomen with or without inconspicuous lateral ridges; head moderately broad and flattened; snout flat and broadly to narrowly rounded or angular. Mouth nearly transverse, short, with thin non-papillose lips; lip furrows short, confined to mouth corners, extending to below eyes and elongated posteriorly into post-oral grooves. Teeth in both jaws compressed, low-crowned, broad and blade-like, forming saw-like edge and deeply overlapping series; upper teeth slightly smaller than lower teeth; tooth count 21-29/19-27. Two dorsal fins, each with strong ungrooved spine; 1st dorsalfin origin opposite or slightly behind pectoral fins; 2nd dorsal fin strongly falcate, its origin opposite or well behind pelvic-fin inner margins; interdorsal space elongate, greater than length of 1st dorsal-fin base. Pectoral fins moderately high, angular, falcate or subtriangular; anterior margin moderately large and subequal or slightly greater than prespiracular length; free rear tip rounded to angular but not greatly elongated. Pelvic fins smaller than pectoral fins and 1st dorsal fin, and subequal to or smaller than 2nd dorsal fin. No anal fin. Peduncle with low, strong keels. Caudal fin without subterminal notch; upper precaudal pit present in Squalus and absent or faintly indicated in Cirrhigaleus. Dermal denticles small to moderately large, with leaf-shaped, tricuspidate, lanceolate or cross-shaped crowns.

Reproductive mode yolk-sac viviparity. Occur in intertidal zone and near surface of epipelagic zone to ~1 500 m on deep slopes, with most species occurring on continental shelf and uppermost slope (usually to ~600 m deep). Found close inshore and in shallow bays in cool-temperate waters, but often epipelagic well offshore in the tropics, where they tend to be replaced in inshore habitats by small species of Carcharhinidae, Sphyrnidae and other carcharhinoid families. Maximum size ~180 cm TL, except one species (*Squalus acanthias*) reportedly reaching 200 cm TL.

Almost circumglobal, in boreal, temperate and tropical seas (except tropical eastern Pacific), often in association

with continents, islands, submarine peaks and ridges. Two genera and 40 species, although several may represent speciescomplexes, with additional species likely to be described; both genera and at least 10 species in WIO.

#### **KEY TO GENERA**





## GENUS Cirrhigaleus Tanaka 1912

Heavy-bodied with short blunt snout, and short anterior nasal flaps with broad-based, stubby to elongated medial barbels. Low blade-like cutting teeth in both jaws. Dorsal fins each with stout, high, ungrooved spine; 2nd dorsal fin about as large as 1st. Peduncle with lateral keels, but precaudal pit poorly developed or absent. Fins with conspicuous white rear margins but no black markings. Three species, 1 in WIO.

## Cirrhigaleus asper (Merrett 1973)

Roughskin spurdog

PLATE 5

*Squalus asper* Merrett 1973: 94, Pl. 1b, Fig. 1 (off Aldabra, equatorial WIO); Bass *et al.* 1976\*; Compagno 1984\*; SSF No. 5.25\*;

Compagno et al. 1989\*.

Cirrhigaleus asper: Compagno et al. 2005\*; Ebert 2013\*; Ebert et al. 2013\*.

Anterior nasal flaps with large medial barbel, usually as broad as distance from its base to inner edge of nostrils, but not elongated behind flaps and not reaching mouth. Obliquecusped cutting teeth; tooth count 27/23. Dorsal fins subequal; origin of 1st dorsal-fin spine behind pectoral-fin free rear tips.

Body dark grey or brown dorsally; no white spots on flanks, but prominent white-edged fins; pale ventrally. Attains 118 cm TL.

**DISTRIBUTION** Peculiar disjunct distribution; known in western Atlantic, central Pacific and WIO, suggesting it may occur elsewhere. WIO: southern Mozambique, South Africa (Eastern Cape and KwaZulu-Natal), Aldabra, Comoros, Saint-Paul and Amsterdam Is., and possibly India (west and east coasts).

**REMARKS** Litters of 18–22 (relatively large); size at birth 25–28 cm TL. Males mature at 85–90 cm TL, and females at 89–118 cm TL. Distinctive but little-known; occurs on upper continental and insular slopes and outer continental shelf in warm-temperate to tropical seas, on or near bottom, at 73–600 m. Often caught well offshore on upper slopes at ~73–110 m, but sometimes found near bays and in river mouths (e.g., in Eastern Cape and KwaZulu-Natal, South Africa). Feeds mostly on bony fishes and squid. Interest

to fisheries minimal, but probably caught incidentally and possibly a utilised bycatch of bottom-trawl and deep longline fisheries; occasionally caught by inshore line fisheries in South Africa (Eastern Cape).

#### GENUS **Squalus** Linnaeus 1758

Slender body with flat rounded head, angular to conical in dorsoventral view; snout flattened, rounded to narrowly pointed, short to elongate. Anterior nasal flaps short, with small, narrow medial barbels; length of upper lip furrow greater than nostril width. Teeth blade-like, with cutting edge, similar in both jaws. Skin smooth; dermal denticles small, crowns of lateral trunk denticles in adults usually <0.5 mm long, with low, pedicellate, lanceolate or tricuspid and triple-ridged, flat, leaf-shaped crowns. Pectoral-fin free rear tips narrowly rounded to acutely angular. Pelvic fins low and obtusely triangular, with anterior margin  $\sim \frac{1}{3} - \frac{1}{2}$  length of pectoral-fin anterior margin. Dorsal-fin spines stout; 2nd dorsal fin smaller than 1st, its base ~3/4 length of 1st dorsal-fin base; 2nd dorsal-fin origin usually behind pelvic-fin rear tips, but occasionally over them. Peduncle elongated, with lateral keels and precaudal pit. Caudal fin slightly elongated, upper lobe narrower, lower lobe strong, and postventral margin usually deeply concave in adults. Body grey to pale brown, some species with white spots on flanks; fins plain or with conspicuous white or black margins or tips, but usually without continuous broad white rear margins. Worldwide, at least 24 species, some of which appear to be species-complexes, which may result in several additional species descriptions; at least 9 species in WIO, 4 species too late for inclusion.



Cirrhigaleus asper, 38 cm TL, female (South Africa). Source: Bass et al. 1976

#### **KEY TO SPECIES**

- 1b First dorsal-fin origin usually in front of pectoral-fin free rear tips; origin of 1st dorsal-fin spine usually over pectoralfin inner margins and ahead of pectoral-fin free rear tips (but varying from just behind to well ahead of tips); medial barbel on anterior nasal flaps well-developed; no white spots on flanks



Distance from snout tip to inner edge of nostril shorter than distance from inner edge of nostril to front of uppe	r
lip furrow	
Distance from snout tip to inner edge of nostril longer than distance from inner edge of nostril to front of upper lip furrow	r 4
Anterior nasal flaps not bifurcate; total vertebrae 93–95,	
	Distance from snout tip to inner edge of nostril shorter than distance from inner edge of nostril to front of uppe lip furrow

- precaudal vertebrae 67–69
  Anterior nasal flaps bifurcate; total vertebrae 102–110, precaudal vertebrae 78–84 *S. acutipinnis*

## Squalus acanthias Linnaeus 1758

#### Spotted spiny dogfish

Squalus acanthias Linnaeus 1758: 233 (Mediterranean Sea and northeastern Atlantic); Barnard 1925; Smith 1949\*, 1965\*;
Bass et al. 1976\*; Compagno 1984\*; Myagkov & Kondyurin 1986;
SSF No. 5.24\*; Compagno, Ebert & Smale 1989\*; Compagno,
Ebert & Cowley 1991; Ebert, Compagno & Cowley 1992;
Compagno et al. 2005; Ebert et al. 2013\*.

*Squalus acanthias africana* Myagkov & Kondyurin 1986: 564 (SE Atlantic) (junior homonym of *Squalus africanus* Gmelin 1788 = *Poroderma africana* [Scyliorhinidae]).

Moderate- to large-sized and slender-bodied with moderately long, narrow, angular or subangular snout. Preoral length 1.1–1.4 mouth width; distance from snout tip to excurrent nostril aperture slightly greater than or approximately same as distance from excurrent aperture to front edge of upper lip furrow. Oblique-cusped cutting teeth in both jaws. Pectoral fins narrow and falcate or semifalcate, with straight to concave rear margin. First dorsal fin relatively long and low; its spine slender and short, and spine origin always behind pectoral-fin free rear tips. No subterminal notch on caudal fin. Tooth count 26–29/20–27; vertebrae 109–116; spiral valve turns 12 or 13.

Body grey, with series of white spots on flanks and dorsal surface, paler ventrally; dorsal-fin edges dusky in adults, and black in young; caudal web dusky, no blackish bar along caudal base and caudal-fin lower lobe not black. Attains ~180 cm TL (highly variable depending on population).



Squalus acanthias, 46 cm TL, female (South Africa). Source: Bass et al. 1976

**DISTRIBUTION** Nearly circumglobal, except for tropics, with apparently isolated allopatric populations or subpopulations, some of which are of uncertain taxonomic status; replaced by *S. suckleyi* (Girard 1855) in North Pacific (Ebert *et al.* 2010). WIO: South Africa (Western Cape to Eastern Cape).

**REMARKS** Litters of 1–32 (usually 2–12 in southern Africa); gestation period long, reported as 18-24 months, possibly varying regionally. Size at maturity varies between populations, those in southern Africa at the lower end of the range, at 52-77 cm TL for males, and 66-100 cm TL for females. Slowgrowing: age at maturity may vary markedly between regions and populations, and is variously reported as 10-20 years for females and 11+ years for males. Long-lived: estimated maximum age 25–30 years, but possibly as long as 75 years. Found in boreal to warm-temperate seas, from inshore and offshore; mainly demersal but also apparently epipelagic over continental and insular shelves, and nearer surface in oceanic waters; mostly above ~600 m deep over continental shelf and upper slope (usually at 124–515 m in southern Africa), but to 1 446 m on deep slopes. Also found in open bays and estuaries, and often caught over soft substrates, and reported from brackish estuaries and freshwater of Europe and eastern United States. Most stocks are highly migratory in dense aggregations. Powerful voracious predator, feeds primarily on bony fishes and capable of dismembering largish prey with its strong jaws and clipper-like teeth; invertebrate prey includes cephalopods, crustaceans, polychaete worms, jellyfish and comb jellies. Can inflict toxins with its tail. Predators on this species include larger bony fishes, sharks and cetaceans. Naturally abundant and heavily fished in many parts of its range, especially North Atlantic and Mediterranean Sea; in WIO, mainly caught incidentally in bottom-trawl hake fisheries and usually discarded. IUCN Red List conservation status Vulnerable globally.

#### Squalus acutipinnis Regan 1908

#### Shortnose spurdog

PLATES 5 & 6

*Squalus acutipinnis* Regan 1908: 248, Pl. 37 (SW Indian Ocean); Regan 1921; Barnard 1925, 1947\*; Viana & Carvalho 2016\*. Squalus megalops (non Macleay 1881): Bass et al. 1976\*; Compagno 1984\*; Myagkov & Kondyurin 1986\*; SSF 5.26\*; Compagno, Ebert & Smale 1989\*; Muñoz-Chapuli & Ramos 1989\*; Compagno, Ebert & Cowley 1991; Ebert, Compagno & Cowley 1992;
Watson & Smale 1998; Compagno et al. 2005\*; Ebert 2013; Ebert et al. 2013\*.

Small-sized, moderately slender, with wide head and short, broad, rounded-parabolic snout; preoral length 1.2–1.4 mouth width. Anterior nasal flaps with small medial bifurcate barbel. Dorsal fins unequal-sized; 1st dorsal fin moderately high, its origin over pectoral-fin inner margins; both dorsal-fin spines slender, 1st spine low and tapering. Pectoral fins broad and falcate, with moderately concave rear margin. No subterminal notch on caudal fin. Vertebrae 107–111.

Body pale grey-brown to dark brown dorsally, paler ventrally; no white spots on flanks; dorsal fins with black tips, and white rear margins and free rear tips (but this colouration often inconspicuous or lacking in adults). Attains 77 cm TL.



*Squalus acutipinnis*, 40 cm TL, male, ventral head (South Africa). Source: Bass *et al.* 1976

**DISTRIBUTION** Southeastern Atlantic to southwestern Indian Ocean. WIO: Mozambique to South Africa (Western Cape), Madagascar and Mauritius.

**REMARKS** Litters of 1–6 (usually 2 or 3); size at birth 23–28 cm TL. In populations off South Africa there appears to be no defined reproductive cycle as adult females continuously reproduce without a gap between pregnancies; estimated gestation period 2 years. Males mature at ~46 cm TL, and females at ~54 cm TL. Common to abundant in temperate



Squalus acutipinnis, 56 cm TL, female (South Africa). Source: SSF

and tropical seas, on inner and outer continental shelves and upper slopes. Generally found on or near bottom, from intertidal zone and close inshore, to ~732 m deep. Off South Africa, newborns are pelagic on pupping grounds over outer continental shelf of Agulhas Bank. Feeds on a variety of bony fishes and occasionally other elasmobranchs, as well as invertebrates, including crustaceans and cephalopods. Often abundant and of considerable interest to fisheries; taken in large quantities in bottom trawls but also on line gear; a common bycatch of trawl fisheries off southern Africa, but largely discarded.

### Squalus blainville (Risso 1827)

#### Longnose spurdog

PLATE 6

*Acanthias blainville* Risso 1827: 133, 478, Pl. 3, Fig. 6 (France, northwestern Mediterranean Sea).

Squalus blainville: Compagno 1984\*; Myagkov & Kondyurin 1986\*; Parin 1987; Muñoz-Chapuli & Ramos 1989\*; Ebert *et al.* 2002; Compagno *et al.* 2005\*; Ebert 2013\*; Ebert *et al.* 2013\*.

Large-sized, stocky to moderately slender, with broad head; oblique distance from snout tip to eyes less than interorbital space; mouth moderately long, rounded-parabolic, preoral length 1–1.3 mouth width. Oblique-cusped cutting teeth in both jaws. Dorsal fins unequal-sized, each with high ungrooved spine; 1st dorsal fin very high and short, with tall slender spine, and spine origin over pectoral-fin inner margins. Pectoral fins broad, semifalcate, with slightly concave rear margin.

Body greyish brown, without white spots on flanks; dorsal and caudal fins white-edged and without conspicuous dark markings or dusky edges or tips; caudal fin without conspicuous dark markings. Attains 100 cm TL.

**DISTRIBUTION** Undefined; possibly globally wide-ranging but may be species-complex with several undescribed species represented. WIO: South Africa (KwaZulu-Natal).

**REMARKS** Details of its biology are uncertain because of taxonomic confusion. Litters of 3–8. Males mature at ~56 cm TL, but female adult size undefined due to confusion with similar-looking *Squalus* species. Females attain ~100 cm TL, and males ~75 cm TL; a female, possibly of this species, caught off KwaZulu-Natal, South Africa, measured 96 cm TL and had 8 near-term embryos. Nominally recorded from continental shelves and upper slopes, on or near bottom, at 16–600 m, but probably occurs deeper. Feeds on bony fishes, cephalopods and crustaceans. Interest to fisheries unclear, but at minimum taken as a bycatch of other fisheries. IUCN Red List conservation status Data Deficient due to uncertain taxonomy.

## Squalus lalannei Baranes 2003

#### Seychelles spurdog

Squalus lalannei Baranes 2003: 42, Figs. 6–12 (Alphonse I., Seychelles).

Moderately slender-bodied with short head and snout; snout rounded; prenarial length 2–2.3 preoral length. Anterior nasal flaps not bifurcate. Mouth rounded, upper jaw concave; mouth width 1.3–1.4 preoral length; upper lip furrow longer than lower lip furrow, with groove extending to level below posterior edge of spiracle. Teeth unicuspidate, bladelike, with cusps directed laterally; teeth similar in both jaws, but upper teeth slightly larger than lower teeth; tooth count 24–26/22–24. First dorsal-fin origin over pectoral-fin free rear tips; 2nd dorsal fin smaller than 1st, with origin over pelvicfin bases. No interdorsal ridge. Pectoral fins moderate-sized. Peduncle with lateral keels originating below 2nd dorsal-fin free rear tip and reaching past caudal-fin origin. Caudal-fin terminal margin moderately convex; tip of lower lobe narrowly rounded. Vertebrae 93–95.

Body uniformly grey, with blackish dorsal fins. Attains at least 79 cm TL.



Squalus blainville, 96 cm TL, female (South Africa).

**REMARKS** Known only from two specimens from the Seychelles (79-cm-TL female; 62-cm-TL male), collected from ~1 000 m. Closely resembles Australian *S. megalops*, but differs in lower number of precaudal vertebrae (67–69 versus 78–84), non-bifurcated anterior nasal flaps, and rounded pectoral-fin free rear tips (cf. more angular tips in *S. megalops*). Presumed viviparous with yolk sac. Possibly taken in deepwater longline fisheries.

#### Squalus mitsukurii Jordan & Snyder 1903

Shortspine spurdog

PLATE 6

Squalus mitsukurii Jordan & Snyder in Jordan & Fowler 1903: 629,

Fig. 3 (Misaki, Japan); Barnard 1925; Smith 1949\*, 1965\*; Bass *et al.* 1976\*; Compagno 1984\*; SSF No. 5.27\*; Compagno, Ebert & Smale 1989\*; Muñoz-Chapuli & Ramos 1989\*; Compagno, Ebert & Cowley 1991; Ebert, Compagno & Cowley 1992;

Compagno *et al.* 2005<sup>\*</sup>; Ebert 2013<sup>\*</sup>; Ebert *et al.* 2013<sup>\*</sup>.

*Squalus probatovi*: Myagkov & Kondyurin 1986.

Medium- to large-sized, moderately stout-bodied, with broad head; snout moderately long and rounded. Anterior nasal flaps with small medial barbel; distance from snout tip to excurrent aperture of nostril slightly greater than from excurrent aperture to front edge of upper lip furrows. Teeth broad-based, with single smooth-edged triangular cusp, and similar in both jaws; tooth count 25–29/22–25. First dorsal fin moderately high, more anteriorly situated, with origin over pectoralfin inner margins and just behind pectoral-fin insertions. Peduncle with upper precaudal pit. Vertebrae 118–127.

Body grey or grey-brown dorsally, white or paler grey ventrally; no white spots on flanks; pectoral fins dusky above, with white tips and rear margins. Attains 125 cm TL (variable depending on population).

**DISTRIBUTION** Wide-ranging in cold-temperate to tropical seas, but range only approximately known because of widespread nominal records of similar-looking species. WIO: Tanzania, Mozambique, South Africa (Eastern Cape and KwaZulu-Natal), Madagascar and India.

**REMARKS** Males mature at 47–85 cm TL, at 4–11 years, and attain ~18 years; females mature at 50–100 cm TL, at 15–20 years, and attain ~27 years. Litters of 2–15, correlating to size of mother; size at birth 21–30 cm TL; gestation period possibly up to 2 years. Occurs on continental and insular shelves, upper slopes, submarine ridges and seamounts, on or near bottom, at 4–954 m (mostly 100–500 m). Common to abundant, sometimes in large aggregations. Powerful predator, feeds primarily on bony fishes, cephalopods and crustaceans. Of considerable interest to fisheries in some regions; commonly caught incidentally in demersal fisheries off southern Africa.



Squalus mitsukurii, 43 cm TL, male (South Africa). Source: Bass et al. 1976



Squalus mitsukurii, 81 cm TL, female (S Mozambique). Source: SSF

## FAMILY CENTROPHORIDAE

## Gulper sharks

Small- to moderate-sized (~40-170 cm TL) with cylindrical or slightly compressed trunk, without lateral ridges on abdomen, and moderately broad to narrow and somewhat flattened head. Snout short or long, flattened and narrowly rounded to elongate-rounded in dorsoventral view. Nostrils wide-spaced, internarial width greater than nostril width, with simple anterior nasal flaps and no medial barbels. Spiracles large, set close behind large oval eyes. Teeth in both jaws with compressed cusp, distal blade and sometimes medial blade, and no cusplets; upper cusps narrow, erect to oblique, and broad-based; lower cusps oblique to semi-erect; tooth counts 22-45/24-35. Two dorsal fins, each with strong grooved spine; 1st dorsal-fin origin usually opposite pectoral-fin bases or inner margins or sometimes just behind free rear tips; 2nd dorsal fin not falcate, its origin usually opposite pelvic-fin bases or inner margins, but exceptionally somewhat behind pelvic-fin rear tips. Interdorsal space elongated and usually greater than length of 1st dorsal-fin base (subequal to or slightly longer than fin base in some species). Pectoral fins low, angular or rounded, and not falcate; anterior margins moderately large. Pelvic fins smaller than pectoral fins and 1st dorsal fin, and subequal to or smaller than 2nd dorsal fin. No anal fin. No keels or precaudal pit on peduncle. Caudal fin heterocercal, lower lobe poorly to strongly developed in adults, and subterminal notch strong. Dermal denticles small to large and variable in shape, with leaf-shaped, tricuspidate or polycuspidate crowns and slender pedicels, high pitchforkshaped erect crowns on high pedicels, or low-ridged sessile crowns. Total vertebrae 106-131; spiral valve turns 10-25. Body plain or with pale or dark markings on fins; no black photophore markings on flanks or tail.

Reproductive mode yolk-sac viviparity, with litters of 1-12. Primarily bottom-dwelling deepwater inhabitants of continental and insular slopes, but also found on submarine ridges and seamounts, at mostly 200–1 500 m (one has been photographed on ocean floor below 4 000 m). Rarely, these sharks also occur offshore to ~50 m deep over continental and insular shelves. Geographic and bathymetric ranges are imperfectly known for most *Centrophorus* species, a result of uneven sampling of deepwater slope-dwelling sharks as well as problems in identifying these species. Social, forming small to huge aggregations, making them among the most common deep-benthic sharks in temperate and tropical seas, although their behaviour and biology are little known. Feed mostly

on bony fishes and cephalopods but also crustaceans, small sharks, batoids and chimaeras. For fisheries purposes, one of the most important families of deep-benthic slope-dwelling sharks, although catches are relatively smaller and more localised than sympatric species of squalid dogfishes. In WIO (especially off India) and elsewhere, these sharks are a common component of targeted deepwater shark fisheries and also form an important bycatch of deepwater fisheries for bony fishes; however, separate fisheries statistics are seldom reported for the family or for individual species. Used for human consumption as dried-salted or fresh, for fishmeal, and for their oily squalene-rich livers, which are extremely large. The conservation status of gulper sharks is of concern because of expanding deepwater fisheries, limited knowledge of their biology, and possibly more extreme limits in life-history parameters than what occurs among well-known and more abundant species of squalid dogfishes.

Almost circumglobal in cold-temperate to tropical seas, in association with landmasses, including continents, islands, submarine peaks and ridges. Generally absent from very high latitudes, and most diverse in warm-temperate waters and the tropics. Several species are wide-ranging, whereas a few may be localised endemics. The greatest known diversity of this family occurs in the Indo-Pacific. Two genera and 15 species; both genera and at least 9 species in WIO.

#### **KEY TO GENERA**



## GENUS Centrophorus Müller & Henle 1837

Body cylindrical, with very tough skin; snout moderately long and broad; eyes huge and iridescent green. Dermal denticles large and leaf-like, thorn-like or pebble-shaped; lateral trunk denticles with low, flat, ridged crowns, varying from leafshaped and with low pedicels and posterior cusps, to cuspless, block-shaped and without pedicels. Teeth similar in both jaws, blade-like and without cusplets, but lower teeth imbricated (overlapping) and much larger than upper teeth; tooth counts 30–45/24–35. Lip furrows not extended anteromedially as elongated preoral grooves. Pectoral-fin free rear tips more or less angular to attenuated. Vertebrae 106–127. Body pale grey or grey-brown to blackish grey, sometimes paler below; fin webs dusky or with pale and dark bars.

This group is extremely problematic taxonomically, with many species poorly known, and with undescribed species possibly occurring in the WIO, especially in deep waters. Worldwide, 17 species; 6 species in WIO, but 2 of these (*C. isodon* [Chu, Meng & Liu 1981] and *C. seychellorum* Baranes 2003) are known only from very deep water (off Maldives, Seychelles, South Africa, and possibly southern Mozambique), thus given the rarity of these deepwater *Centrophorus* species they are included here only in the key.

#### **KEY TO SPECIES**

- Lateral trunk denticles with flat sessile crowns on denticle bases, without separate pedicels, and crowns usually with or sometimes without posterior medial cusp but no lateral cusps
   2







- 2b Second dorsal fin larger, nearly as high as 1st dorsal fin, with origin of 2nd dorsal-fin spine usually over pelvic-fin inner margins
   3

Continued . . .

#### **KEY TO SPECIES**

3a	Snout elongated, narrowly parabolic in dorsoventral view, narrowly cuneate in lateral view; preoral length 1.4–1.8 mouth width, and 0.9–1.1 head width at mouth corners; mouth narrower, its width 5.2–7.3% TL; body usually slender
3b	Snout short and broad, broadly parabolic in dorsoventral view, broadly wedge-shaped in lateral view; preoral length 0.9–1.5 (usually <1.3) mouth width, and 0.6–0.9 head width at mouth corners; mouth broader, its width 6.6–9.5% TL; body usually stout
4a	Dorsal-caudal space relatively short, <7% TL; flank denticles block-like, widely spaced
4b	Dorsal-caudal space relatively long, >8% TL; flank denticles not block-like, more pointed, close-set <i>C. seychellorum</i> [deep water]
5a	Fins of adults and subadults prominently marked with black tips and margins, including pectoral fins, both dorsal fins, both caudal-fin lobes, and sometimes pelvic fins; maximum <100 cm TL
5b	Fins of adults and subadults not prominently marked, their colour pale to dusky; maximum ~170 cm TL <i>C. granulosus</i>

#### Centrophorus atromarginatus Garman 1913

Blackfin gulper shark

PLATE 7

*Centrophorus atromarginatus* Garman 1913: 200, Pl. 13, Figs. 1–4 (Suruga Gulf, Japan); Compagno *et al.* 2005\*; Ebert 2013\*; Ebert *et al.* 2013\*.

Snout moderately long and thick. Teeth blade-like, monocuspidate in both jaws, with lower teeth much larger than upper teeth; tooth count 40–42/29–30. Pectoral-fin free rear tips narrowly angular and greatly elongated. First dorsal fin short, higher than 2nd; base of 2nd dorsal-fin spine over pelvic-fin inner margins or rear tips. Vertebrae 107–114; spiral valve turns 10–12.

Body grey or grey-brown dorsally, paler ventrally; prominent blackish markings on fins. Attains 94 cm TL.



Centrophorus atromarginatus, 85 cm TL (Japan). Source: Garman 1913

**DISTRIBUTION** Sparse records from eastern Atlantic, western Pacific and Indian oceans, but probably wider-ranging. WIO: Somalia (Gulf of Aden), India and Sri Lanka.

**REMARKS** Often confused with *C. granulosus* and *C. moluccensis*. Litters of 1 or 2; size at birth 30–36 cm TL. Males mature at ~56 cm TL, and females at ~75 cm TL. Occupies moderately deep water along outer continental shelf, upper slope and insular shelves, at 150–450 m. Feeds on small bony fishes and shrimps. Mostly caught as bycatch in WIO; elsewhere localised fisheries occur for its squalene-rich liver oil.

#### Centrophorus granulosus (Bloch & Schneider 1801)

Gulper shark

PLATE 7

Squalus granulosus Bloch & Schneider 1801: 135 [Canary Is.].

*Centrophorus lusitanicus* Barbosa du Bocage & De Brito Capello 1864: 260, Fig. 1 (Portugal, North Atlantic); Compagno 1984 [in part: southern Africa]; SSF No. 5.2\*; Compagno *et al.* 1989\*.

Centrophorus granulosus: Bass et al. 1976\*; SSF No. 5.1\*;

Compagno *et al.* 1989\*; Compagno *et al.* 1991; Ebert *et al.* 1992; Compagno *et al.* 2005\*; Ebert 2013\*; Ebert *et al.* 2013\*; White *et al.* 2013\*.

Snout moderately long and thick. Teeth blade-like, monocuspidate, similarly shaped in both jaws, but lower teeth larger than upper teeth; tooth count 36–43/28–32. Pectoral-fin free rear tips narrowly angular and greatly elongated. First dorsal fin higher than 2nd; base of 2nd dorsalfin spine over pelvic-fin inner margins. Vertebrae 113–125; spiral valve turns 11–15.

Body dark grey or grey-brown above, slightly paler below; fins with dark grey or blackish webs, but without prominent black tips or margins. Attains 170 cm TL.

**DISTRIBUTION** Circumglobal, including Mediterranean Sea, southeastern Atlantic, central and western Pacific (excluding eastern Pacific), and Indian Ocean. WIO: Gulf of Aden, Mozambique, South Africa (Mozambique/KwaZulu-Natal border to Orange River, Northern Cape Province), Madagascar, southern Madagascar Ridge, Aldabra, Maldives and possibly India.

**REMARKS** Often confused with *C. atromarginatus* and *C. moluccensis*. Litters of 1–6; size at birth 30–45 cm TL. Males mature at 110–128 cm TL, and females at >130 cm TL; size at maturity varies by region. Occurs in deep water along outer continental shelves and upper slopes, and insular shelves, at 50–1 440 m (usually 200–600 m). Feeds on bony fishes, cephalopods and crustaceans. Fished primarily in North Atlantic, and taken mostly as a bycatch in WIO; used for its meat and squalene-rich liver oil. IUCN Red List conservation status Critically Endangered in North Atlantic and Mediterranean, but Vulnerable in WIO.



Centrophorus granulosus, 85 cm TL, male (S Mozambique). Source: Bass et al. 1976


Centrophorus moluccensis, 78 cm TL, mature male (S Mozambique). Source: Bass et al. 1976

## Centrophorus moluccensis Bleeker 1860

#### Smallfin gulper shark

PLATE 7

*Centrophorus moluccensis* Bleeker 1860: 3 (Ambon I., Moluccas, Indonesia); Compagno 1984\*; Compagno *et al.* 1989\*, 2005\*; Ebert 2013\*; Ebert *et al.* 2013\*.

Centrophorus scalpratus McCulloch 1915: 97, Pl. 13, Figs. 2–7 (off Gabo I., Victoria coast, Australia); Bass et al. 1976\*; SSF No. 5.3\*.

Atractophorus armatus Gilchrist 1922: 48, Pl. 7, Fig. 3 (KwaZulu-Natal, South Africa); SFSA No. 47\*.

Snout moderately long and flat. Teeth blade-like, similar in both jaws, but lower teeth much larger than upper teeth; tooth count 36–45/29–35. Pectoral-fin free rear tips elongated into very narrow angular lobe, reaching well behind base of 1st dorsal-fin spine. First dorsal fin much higher and larger than 2nd; base of 2nd dorsal-fin spine usually behind pelvic-fin rear tips. Caudal-fin postventral margin conspicuously notched. Vertebrae 113–131; spiral valve turns 20–29.

Body grey-brown or grey dorsally, slightly paler ventrally; pectoral fins with conspicuously paler rear margins. Attains at least 100 cm TL.

**DISTRIBUTION** Indo-Pacific (scattered); Indian subcontinent records of *C. atromarginatus* may be of *C. moluccensis*, thus this species likely has a more extensive range than indicated

by available records. WIO: South Africa (KwaZulu-Natal) and Mozambique.

**REMARKS** Litters of 2; size at birth 31–37 cm TL; gestation period possibly 2 years. Males mature at ~70 cm TL, and females at ~89 cm TL. Common; occurs in deep water over outer continental and insular shelves and upper slopes, on or near bottom, at 128–823 m. Feeds primarily on bony fishes, other dogfish sharks (Order Squaliformes), cephalopods and crustaceans. Of minor to moderate fisheries importance off South Africa and Australia, along with other dogfish sharks, but little utilised; mostly taken as a bycatch of fisheries targeting shrimp and bony fishes (populations off the east coast of southern Africa may have declined markedly as a result).

## Centrophorus squamosus (Bonnaterre 1788)

#### Leaf-scale gulper shark

*Squalus squamosus* Bonnaterre 1788: 12 [probably eastern North Atlantic]. *Encheiridiodon hendersoni* Smith 1967: 129, Pls. 24–27 (Algoa Bay, South Africa).

Centrophorus squamosus: Forster et al. 1970; Bass et al. 1976\*; Compagno 1984\*; SSF No. 5.4\*; Compagno et al. 1989\*; Muñoz-Chapuli & Ramos 1989; Compagno et al. 1991\*; Ebert et al. 1992\*; Compagno et al. 2005\*; Last & Stevens 2009\*; Ebert 2013\*; Ebert et al. 2013\*.



Centrophorus squamosus, 93 cm TL, male (South Africa). Source: Bass et al. 1976

Large-sized and relatively stocky; snout short, thick and somewhat flattened. Teeth blade-like, similar in both jaws, but lower teeth much larger than upper teeth; tooth count 30–38/24–32. Lateral trunk denticles high, rough, leaf-shaped, tricuspidate or multicuspidate. Pectoral-fin free rear tips hardly angular and not greatly elongated, forming angular corner to very short, narrow, angular lobe that ends well in front of 1st dorsal-fin spine. First dorsal fin long and low, usually slightly lower but larger than 2nd; base of 2nd dorsalfin spine usually opposite pelvic-fin inner margins or rear tips. Vertebrae 106–120; spiral valve turns 12–14.

Body grey, grey-brown or reddish brown dorsally, usually similar ventrally; fin webs and margins dusky, but no prominent fin markings. Attains 165 cm TL.

**DISTRIBUTION** Atlantic and Indo-Pacific (wide-ranging). WIO: South Africa (Eastern Cape and KwaZulu-Natal), southern Madagascar Ridge, Seychelles (Aldabra and Assumption) and India.

**REMARKS** Litters of 4–9; size at birth 30–40 cm TL. Males mature at ~100 cm TL, at ~30 years; females mature at ~120 cm TL, at ~35 years; estimated maximum age 70 years. Occurs in deep water over continental slope, at 229–2 359 m, usually on or near bottom, but has been found in epipelagic and mesopelagic zones, between the surface and ~1 250 m over water to ~3 940 m deep. (One specimen was collected by a spearfisher in water ~4 m deep off east coast of South Africa, but this shark does not normally stray onto the continental shelf.) Feeds on bony fishes, chimaeras, small dogfish sharks, cephalopods and crustaceans. Interest to fisheries limited, mostly taken as a bycatch. IUCN Red List conservation status Endangered globally.

# GENUS Deania Jordan & Snyder 1902

Body cylindrical or compressed, with fine rough skin and large erect denticles with pitchfork-shaped crowns and 3 sharp cusps. Snout very long (>50% HL) and broad, spatulate in dorsoventral

view, and appearing as thin elongated wedge in lateral view. Lip furrows elongated anteriorly as preoral grooves. Teeth without cusplets, similar in both jaws; lower teeth imbricated and somewhat larger than upper teeth; tooth counts 22–36/24–33. Pectoral-fin free rear tip rounded, not angular or attenuated. Two dorsal fins, each with strong grooved spine; 2nd dorsal-fin spine much larger than 1st. Second dorsal fin about as large as, or slightly larger than, 1st dorsal fin, its base ≤¾ length that of 1st; 2nd dorsal-fin origin about over middle of pelvic-fin bases, with free rear tip reaching caudal-fin origin. No anal fin. Caudal fin with strong subterminal notch. Vertebrae 118–128. Body pale grey or grey-brown to blackish; fins dusky and without conspicuous markings; eyes huge and iridescent green or yellowish. Four species, 3 in WIO.

#### **KEY TO SPECIES**



## Deania calceus (Lowe 1839)

#### Birdbeak dogfish

PLATE 8

Acanthidium calceus Lowe 1839: 92 (Madeira, North Atlantic). Deania calcea: Bass et al. 1976\*; Compagno 1984\*; Compagno et al. 1989\*; Compagno et al. 1991; Ebert et al. 1992; Compagno et al. 2005\*; Last & Stevens 2009\*; Ebert 2013\*; Ebert et al. 2013\*. Deania calceus: SSF No. 5.8\*.



Deania calceus, 84 cm TL, male (South Africa). Source: Bass et al. 1976 Snout extremely long and flattened. First dorsal fin long and low; 2nd dorsal-fin spine much higher than 1st. No subcaudal keel on underside of peduncle. Tooth count 25–35/27–33; vertebrae 118–127.

Body varying from uniformly pale or dark grey or grey-brown to dark brown; fins darker, and fin webs dusky to blackish; juveniles also with darkened dorsal-fin margins, caudal-fin lobes, and areas above eyes and gills. Attains 122 cm TL.

**DISTRIBUTION** Eastern Atlantic, southeastern and western Pacific, and Indian oceans (patchy). WIO: South Africa and southern Madagascar Ridge.

**REMARKS** Litters of 6–17; size at birth ~30 cm TL. Males mature at ~80 cm TL, at ~14 years; females mature at ~105 cm TL, at ~22 years; estimated maximum age 35 years. Occurs in deep water over outer continental and insular shelves and upper, middle and lower slopes, on bottom to well above it, at 73–1 471 m. Common, sometimes found in large groups. Diet includes bony fishes, cephalopods and crustaceans. Interest to fisheries moderate, usually taken as a bycatch; used for its meat and squalene-rich liver oil. IUCN Red List conservation status Near Threatened.

# Deania profundorum (Smith & Radcliffe 1912)

#### Arrow-head dogfish

Nasisqualus profundorum Smith & Radcliffe in Smith 1912: 681; Pl. 53,

Fig. 3 (between Leyte and Mindanao Is., Philippines). *Acanthidium natalense* Gilchrist 1922: 49, Pl. 7, Fig. 2 (off Illovo River,

KwaZulu-Natal, South Africa).

Deania profundorum: Bass et al. 1976\*; Compagno 1984\*; SSF No. 5.9\*; Compagno et al. 1989\*; Compagno et al. 1991; Ebert et al. 1992; Compagno et al. 2005\*; Ebert 2013\*; Ebert et al. 2013.

Snout long and flattened. First dorsal fin short, moderately high; 2nd dorsal-fin spine higher than 1st dorsal-fin spine. Distinctive subcaudal keel on underside of peduncle.

Body uniformly grey or grey-brown. Attains 97 cm TL.

**DISTRIBUTION** Atlantic, western Pacific (Philippines), and Indian oceans (patchy). WIO: South Africa, southern Madagascar Ridge, Gulf of Aden and India.

**REMARKS** Litters of 5–7; size at birth ~31 cm TL or more. Males mature at ~43 cm TL, and females at ~60 cm TL. Little-known; occurs in deep water over upper continental and insular slopes, on or near bottom, at 205–1 800 m (Weigmann 2016). Diet includes bony fishes, cephalopods and crustaceans. No interest to fisheries, but taken as bycatch and often caught in huge aggregations.

# Deania quadrispinosa (McCulloch 1915)

#### Longsnout dogfish

PLATE 8

Acanthidium quadrispinosum McCulloch 1915: 100, Pl. 14, Figs. 5–8 (Great Australian Bight, between Gabo I. and Cape Everhard). Deania quadrispinosum: Bass et al. 1976\*; Compagno et al. 1989;

Compagno *et al.* 1991; Compagno 1999; Ebert 2013\*; Ebert *et al.* 2013\*. *Deania quadrispinosus*: SSF No. 5.10\*.

Deania quadrispinosa: Compagno 1984\*; Last & Stevens 2009.



PLATE 8

Deania profundorum, 33 cm TL, male syntype of Acanthidium natalense (South Africa). Source: Bass et al. 1976



Deania quadrispinosa, 110 cm TL, female (S Mozambique). Source: Bass et al. 1976

Snout extremely long and flattened. First dorsal fin relatively short, moderately high; 2nd dorsal-fin spine much higher than 1st dorsal-fin spine. No subcaudal keel on underside of peduncle. Tooth count 28–33/29–31; vertebrae 118–127.

Body uniformly brown, grey, grey-brown or black; sometimes with white-edged fins. Attains 118 cm TL.

**DISTRIBUTION** Southeastern Atlantic, western Pacific and Indian oceans. WIO: South Africa (Eastern Cape), southern Mozambique, and seamounts south of Madagascar.

**REMARKS** Litters of 5–17; size at birth ~24 cm TL. Males mature at ~80 cm TL, and females at ~100 cm TL. Little-known; occurs in deep water over outer continental shelves and slopes, and around seamounts, on or near bottom, at 150–1 360 m. Feeds on bony fishes. Fisheries interest minimal; a discarded bycatch of trawl fisheries off South Africa. IUCN Red List conservation status Near Threatened, because of its limited bathymetric and geographical distribution and its possible vulnerability to overfishing due to low fecundity.

# FAMILY ETMOPTERIDAE

Lanternsharks

James DS Knuckey and David A Ebert

Dwarf to medium-sized (often <100 cm TL) with cylindrical to slightly depressed body; snout short or long, flat or conical, and narrow to broadly rounded. Fifth gill slit not much enlarged relative to first 4 slits (although widths may increase slightly from 1st to 5th slits). Mouth varying from broadly arched or U-shaped and elongated to nearly transverse and very short. Teeth similar in both jaws or differentiated between jaws; upper teeth with medial cusp, sometimes flanked by smaller lateral cusplets; lower teeth similar to upper teeth (*Centroscyllium*) or compressed and blade-like with cusp (*Etmopterus*). Dermal denticles small to moderately large, variable in shape, with slender to stout, pointed, wedgeshaped or hooked erect crowns without pedicels, or with low concave sessile crowns. Dorsal fins each with strong grooved spine. First dorsal fin and spine usually smaller than 2nd, its origin varying from opposite pectoral-fin inner margins to somewhat behind their free rear tips. Second dorsal fin falcate or not, its origin often opposite pelvic-fin bases or inner margins. No anal fin. Caudal fin with subterminal notch moderately strong to lacking. Body and fin bases with photophores (light organs), inconspicuous and diffuse or in black patches on ventrolateral surfaces.

Reproductive mode where known is yolk-sac viviparity. Primarily deepwater and bottom-dwelling, at ~70–2 250 m (possibly deeper); a few species are semi-oceanic in epipelagic and mesopelagic zones. Some species form schools. Of little importance to fisheries but often taken as bycatch. Circumglobal in boreal, austral, temperate and tropical seas, mostly in association with landmasses, including continents, continental and oceanic islands, or submarine peaks and ridges.

The largest family of Squaliformes, with with 4 genera and >50 species recognised; 2 genera and at least 11 species in WIO (4 of these species from deep water: *Centroscyllium ornatum* [Alcock 1889], *Etmopterus gracilispinis* Krefft 1968, *E. sculptus* Ebert, Compagno & De Vries 2011 and *E. viator* Straube 2011).

#### **KEY TO GENERA**

1a	Lower teeth similar t blade-like, and not o mouth moderately lo	o upper teeth, not compressed or verlapping or abutting one another; ong and arcuate <b>Centroscyllium</b>
1b	Lower teeth dissimila blade-like, and with abutting one anothe transverse	ar to upper teeth, compressed and adjacent teeth overlapping or r; mouth short, nearly straight and from deep water in Arabian Sea and Bay of Bengal]
	1a М. М.	

# GENUS Centroscyllium Müller & Henle 1841

Head broad and flattened, wider than deep. Snout short to moderately long, moderately rounded or slightly pointed; preoral length 0.5-1 mouth width. Teeth similar in both jaws, comb-like, with conical straight cusps and 1 or 2 pairs of cusplets, not compressed and blade-like and not imbricated. Dermal denticles on lateral trunk with bluntly conical, thornor bristle-like cusps and stellate bases; denticles usually sparse and well-spaced. First dorsal-fin base much shorter than interdorsal space; 1st dorsal-fin origin usually about opposite or just behind pectoral-fin free rear tips. Second dorsal fin usually larger than 1st. Photophores denser on ventral surface than on dorsal surface or else absent from dorsal surface; usually no conspicuous black photophore patches on underside of head and abdomen or on flanks and tail. Body greyish or blackish brown; fin webs varying from as dark as fin bases to abruptly white or with black and white markings; no naked patch of white skin on edge of upper eyelid. Seven species, 2 in WIO (one of which is only known from deep water).

#### **KEY TO SPECIES**

# Centroscyllium fabricii (Reinhardt 1825)

#### Black dogfish

PLATE 9

*Spinax fabricii* Reinhardt 1825: 3 (Julianehåb [Qaqortoq], Greenland, North Atlantic).

Centroscyllium fabricii: Bass et al. 1976\*; SSF No. 5.5\*;

Compagno *et al.* 1989\*; Compagno *et al.* 1991; Ebert *et al.* 1992; Compagno *et al.* 2005\*; Ebert *et al.* 2013\*. Body moderately stout and compressed. Mouth moderately arched; teeth in both jaws with narrow cusps and cusplets. Skin firm; denticles high, conical and sharp-cusped, dense and numerous on dorsal and ventral surfaces. Pectoralfin apex when laid back ending well before origin of 1st dorsal-fin spine. First dorsal fin low, subtriangular, with low subangular fin web. Second dorsal fin larger than 1st. Dorsalfin spines grooved; 1st dorsal-fin spine ending below fin apex; 2nd dorsal-fin spine longer than 1st dorsal-fin spine, and extending to below or about height of 2nd dorsal-fin apex. Vertebrae 87–97; spiral valve turns 7–10.

Body uniformly blackish brown, without white markings on fins or discrete black photophores on body. Attains at least 107 cm TL.

**DISTRIBUTION** Western and eastern North Atlantic, including West Africa and southern Africa, from Orange River mouth (Namibia/South Africa) in southeastern Atlantic, and just rounding Cape Agulhas (South Africa) into WIO.

**REMARKS** Viviparous with yolk sac; size at birth 15–20 cm TL. Matures at ~46 cm TL. Abundant deepwater schooling shark of outermost continental shelves and slopes, at 180–2 250 m (mostly deeper than 275 m). May move up to near surface, especially during winter and when darkest at night. Feeds primarily on crustaceans, cephalopods, jellyfish and bony fishes. Interest to fisheries limited, commonly taken as a bycatch. IUCN Red List conservation status Least Concern.

## GENUS **Etmopterus** Rafinesque 1810

Head broad and flattened, wider than deep; gill slits very short. Snout moderately long, broadly rounded to slightly pointed, flattened or subconical; preoral length 0.9–1.7 mouth width. Teeth strongly differentiated in upper and lower jaws; upper teeth small, not fang-like, with strong, conical, nearly straight cusps, and 1 to several pairs of cusplets; lower-jaw dentition compressed and blade-like, with flattened cusp, no cusplets,



Centroscyllium fabricii, 64 cm TL, female (NW Atlantic). Source: Bass *et al.* 1976, drawn from Bigelow & Schroeder 1948 and distal blade. Dermal denticles on lateral trunk with thornor bristle-like, conical or hooked cusps, or flat, truncate and without cusps; denticles usually closely spaced. First dorsalfin base much shorter than interdorsal space; 2nd dorsal fin noticeably larger than 1st. Dorsal-fin spines usually large and strongly posteriorly curved; 2nd dorsal-fin spine usually much longer than 1st, its tip extending to 2nd dorsal-fin apex. Photophores denser on ventral surface than on dorsal surface; conspicuous black photophore patches often on underside of head and abdomen, and on flanks and tail, but obscure or absent in some species. Body varying from blackish to tan dorsally, and often black ventrally; fin webs slightly or abruptly paler than fin bases; semicircular or elongated patch of white skin on edge of upper eyelid in some species. At least 37 species; at least 8 species in WIO at depths near 200 m and 3 species from deep water (see family account).

#### **KEY TO SPECIES**





2b

#### **KEY TO SPECIES**

- 3b Upper tooth cusps slender; lower tooth cusps do not become erect with growth; 1st dorsal fin more posteriorly set; distance from pectoral-fin insertion to 1st dorsal-fin base
   <3 in interdorsal space; 1st dorsal-fin spine short and slender, its length equal to or less than fin base; spiral valve turns</li>



# Etmopterus bigelowi Shirai & Tachikawa 1993

#### Blurred lanternshark

*Etmopterus bigelowi* Shirai & Tachikawa 1993: 487, Figs. 1b–d, 2b, 5b (off Angola); Compagno *et al.* 2005; Ebert *et al.* 2013\*.

Body firm, slightly compressed and slender. Head broad and flattened, width about equal to preoral length. Snout long, broad and spatulate. Upper teeth with 1–3 or occasionally 4 pairs of lateral cusplets, with stout expanded cusps; lower teeth cusps become erect with maturity. Dermal denticles on lateral trunk flat and robust with sessile low crowns (not thornor bristle-like), fairly wide-spaced and not overlapping, giving skin smooth texture, and denticles not in regular longitudinal rows; no rows of enlarged denticles on flanks above pectoral fins; denticles on 2nd dorsal fin densely covering fin base but sparse or absent on fin web. First dorsal-fin origin opposite or just behind pectoral-fin free rear tips; 1st dorsal-fin spine stout and slightly lower than fin apex. Second dorsal fin larger than



Continued

*Etmopterus bigelowi*, 46 cm TL, mature male (Angola). Source: Shirai & Tachikawa 1993 1st, fin apex rounded-angular, and rear margin deeply concave; 2nd dorsal-fin spine stout, with tip posteriorly curved to become diagonally vertical. Caudal fin long. Vertebrae 79–91; spiral valve turns 16–19.

Body dark brown or blackish dorsally, with underside of snout and abdomen grading to black. Flank and caudal photophores inconspicuous; flank photophore base before 2nd dorsal-fin spine, with anterior branch long and broad, and no posterior branch. Attains at least 72 cm TL.

**DISTRIBUTION** Tropical and subtropical waters of Atlantic and Indo-Pacific. WIO: South Africa (Cape Town to Mossel Bay, and open ocean off KwaZulu-Natal).

**REMARKS** Viviparous with yolk sac. Males mature at 41–67 cm TL, and females at 50–65 cm TL. Deep water, over continental shelves and slopes, island slopes and submarine ridges, at 163–1 000 m; also partly epipelagic and occurring near surface in open ocean, at 110–700 m. Feeds primarily on fishes and cephalopods. Of no importance to fisheries, although probably a discarded bycatch of commercial fisheries.

## Etmopterus compagnoi Fricke & Koch 1990

Brown lanternshark

PLATE 9

*Etmopterus compagnoi* Fricke & Koch 1990: 2, Figs. 1–2 (off Cape Town, South Africa); Ebert *et al.* 2013\*.

*Etmopterus spinax*: Gilchrist 1922; Barnard 1925, 1947\*; Smith 1949\*; Compagno 1984\*.

Etmopterus gracilispinis: Karrer 1973; Compagno 1984\*;

Compagno et al. 1989; Compagno et al. 1991; Ebert et al. 1992. Etmopterus sp.: SSF No. 5.16\*.

Etmopterus unicolor: Compagno et al. 2005\*.

Body moderately firm, cylindrical, heavy and stout. Head broad and flattened, width about 1.1–1.3 preoral length; snout thick but broadly spatulate, not bulbous. Upper teeth generally with 1 or 2 pairs of cusplets, and expanded cusps. Denticles on 2nd dorsal fin densely covering base; lateral trunk denticles slender, close-set, and with very slender, hooked, conical crowns, giving skin velvety texture. First dorsal-fin origin well behind pectoral-fin free rear tips; 1st dorsal-fin spine stout and usually lower than fin apex. Second dorsal fin larger than 1st, fin apex more or less pointed or rounded-angular, and rear margin broadly concave; 2nd dorsal-fin spine stout with tip posteriorly curved. Caudal fin short. Vertebrae 82–90; spiral valve turns 9–12.

Body dark grey, dark brown or grey-brown dorsally; underside of snout and abdomen grading to blackish brown; flanks above and behind pelvic fins and underside of caudal fin with indistinct darker markings. Photophores on flank and caudal area inconspicuous, photophore base mostly behind 2nd dorsal-fin spine, with anterior branch long and slender, and longer than posterior branch. Attains 79 cm TL.

**DISTRIBUTION** Subtropical waters of southeastern Atlantic and Western Indian Ocean. WIO: South Africa (Western Cape to KwaZulu-Natal).

**REMARKS** Viviparous with yolk sac; litters of 2–21 (average 12); size at birth ~17 cm TL. Males mature at 48–68 cm TL, and females at 53–79 cm TL. Occurs in deep water over continental and insular slopes, often on bottom but sometimes well off it, at 465–1 500 m; has also been caught in epipelagic zone of open ocean in 120 m. Broadly sympatric with larger *E. granulosus*, with which it has been caught in experimental trawls on slopes off South Africa, at 383–1 300 m. Feeds on cephalopods, penaeid shrimp and fishes. Limited or no interest to fisheries, although probably an incidental bycatch of deepwater trawl fisheries.



Etmopterus compagnoi, 32 cm TL, male (South Africa). Source: SSF



Etmopterus granulosus, 31 cm TL, male holotype (Chile). Source: SSF

# Etmopterus granulosus (Günther 1880)

Southern lanternshark

Spinax granulosus Günther 1880: 19, Pl. 2c (SW coast of South America). Etmopterus baxteri: Compagno et al. 2005\*.

Etmopterus granulosus: Bass et al. 1976\*; SSF No. 5.12\*; Compagno et al. 1989\*; Compagno et al. 1991; Ebert et al. 1992; Ebert 2013\*; Ebert et al. 2013\*.

Body moderately large, firm and slightly compressed. Head broad and flattened, width greater than preoral length. Gill slits very small. Upper teeth with medial cusp and 2 or 3 pairs of lateral cusplets. No rows of enlarged dermal denticles on flanks above pectoral fins; denticles in regular longitudinal rows on peduncle, caudal fin and sometimes flanks, but no denticles on head and 2nd dorsal fin; lateral trunk denticles large and robust, with low-hooked, stout, conical crowns, and widespaced, giving skin very rough texture. First dorsal-fin origin well behind pectoral-fin free rear tips; 1st dorsal-fin spine stout and usually lower than fin apex. Second dorsal fin much larger than 1st, fin apex rounded-truncated, rear margin shallowly concave; 2nd dorsal-fin spine stout with tip posteriorly curved to become horizontal in adults. Caudal fin short. Vertebrae 86-94; spiral valve turns 9-13.

Body dark brown or blackish, with underside of snout and abdomen blackish, but dorsal surface not conspicuously paler than ventral surface. Flank and caudal photophores inconspicuous; flank photophore base well forward of 2nd

dorsal-fin spine, with anterior branch elongated and narrow, much longer than posterior branch. Attains 88 cm TL.

**DISTRIBUTION** Southeastern Pacific to southwestern Atlantic (Chile to Argentina), and Indian Ocean. WIO: South Africa (Western Cape to Eastern Cape), Madagascar Ridge (south of Madagascar); elsewhere near Marion I. (sub-Antarctic Indian Ocean).

**REMARKS** Viviparous with yolk sac; litters of 6–16; size at birth 16-17 cm TL. May breed all year long. Males mature at 46-68 cm TL, at ~20 years; females mature at 75-86 cm TL, at ~30 years; maximum estimated ages 48 and 57 years for males and females, respectively. Common to abundant in deep water over upper continental and insular slopes, on or near bottom, at 220-1 500 m (Weigmann 2016) (mostly deeper than 600 m). Feeds primarily on cephalopods, crustaceans and other invertebrates, and bony fishes. Of minor interest to fisheries. IUCN Red List conservation status Least Concern, although deepwater trawl and line fisheries occur in its range and habitat.

# Etmopterus pusillus (Lowe 1839)

#### Smooth lanternshark

Acanthidium pusillum Lowe 1839: 91 (Madeira, North Atlantic). Etmopterus pusillus: Bass et al. 1976\*; SSF No. 5.14\*; Compagno et al. 1989\*; Compagno et al. 1991; Ebert et al. 1992; Compagno et al. 2005\*; Ebert et al. 2013\*.



Etmopterus pusillus, 33 cm TL, female (South Africa). Source: SSF

PLATE 10

PLATE 9

PLATE 11

Body moderately firm, slightly compressed, and moderately slender. Head moderately broad and flattened, not deep and conical; head width 0.9-1.4 preoral length; snout flattened and broadly rounded, not bulbous. Upper teeth generally with 1 or 2 pairs of cusplets, with slender cusps. Dermal denticles on lateral trunk flat and robust, close-set but not overlapping, with sessile low crowns (not thorn- or bristle-like), giving skin smooth texture, and denticles not in regular longitudinal rows; no rows of enlarged dermal denticles on flanks above pectoral fins; denticles on 2nd dorsal fin densely covering base but absent from rear margin. First dorsal-fin origin just behind pectoral-fin free rear tips; 1st dorsal-fin spine slender, short and usually lower than fin apex. Second dorsal fin much larger than 1st, fin apex more or less pointed or narrowly rounded, rear margin usually deeply concave; 2nd dorsal-fin spine stout with tip posteriorly curved to become diagonally vertical. Caudal fin long. Vertebrae 82-88; spiral valve turns 10-13.

Body pale or dark brown to blackish dorsally; ventral surface conspicuously darker, and underside of snout and abdomen abruptly black. Flank and caudal photophores inconspicuous; flank photophore base opposite and behind 2nd dorsal-fin spine, with anterior branch long and broad, and no posterior branch. Attains 50 cm TL.

**DISTRIBUTION** Tropical and subtropical waters of Atlantic and Indo-Pacific. WIO: Mozambique, South Africa (KwaZulu-Natal) and Sapmer Seamount.

**REMARKS** Viviparous with yolk sac; litters of 1–6. Males mature at 31–48 cm TL, at 5–9 years; females mature at 38–50 cm TL, at 8–11 years. Size at maturity may vary regionally. Found on continental slopes, submarine ridges and seamounts, on or near bottom, at 0–1 500 m (Weigmann 2016). Consumes primarily fish eggs, fishes and other small dogfish sharks, and squid. Interest to fisheries limited, but often captured as a bycatch.

## Etmopterus sentosus

Bass, D'Aubrey & Kistnasamy 1976

#### Thorny lanternshark

*Etmopterus sentosus* Bass, D'Aubrey & Kistnasamy 1976: 22, Figs. 15, 18a (southern Mozambique); SSF No. 5.15\*; Compagno *et al.* 1989\*; Compagno *et al.* 2005\*; Ebert *et al.* 2013\*.

Body small, moderately firm, cylindrical, not compressed, and slender. Head somewhat depressed, flattened and subconical; head width ~2.4 preoral length; snout flattened-conical, not bulbous. Upper teeth generally with 2–4 pairs of cusplets, with cusps greatly expanded. Two or 3 horizontal rows of greatly enlarged hook-like denticles on flanks above and slightly behind pectoral fins. Denticles on head not in longitudinal rows, but denticles on flanks and tail arranged in regular or semi-regular longitudinal rows; scattered denticles on base of 2nd dorsal fin. First dorsal-fin origin over pectoral-fin inner margins; 1st dorsal-fin spine stout, lower than fin apex. Second dorsal fin much larger than 1st, fin apex rounded-angular, and rear margin deeply concave; 2nd dorsal-fin spine stout with tip posteriorly curved to become diagonally vertical. Vertebrae 83; spiral valve turns 8 or 9.

Body greyish above and below, dorsal surface paler but ventral surface not conspicuously dark, except underside of snout and abdomen blackish; caudal-fin tip with conspicuous black bar. Flank and caudal photophores conspicuous; flank photophore base just behind 2nd dorsal-fin spine, with anterior branch long and broad, much longer than posterior branch. Attains 27 cm TL.

**DISTRIBUTION** WIO: Kenya to South Africa (KwaZulu-Natal) and Madagascar (off Toliara).

**REMARKS** Viviparous with yolk sac. Males mature at 22–27 cm TL, and females at 25–27 cm TL. Little known, dwarf-sized; found in deep water at or near bottom, at 200–500 m. No interest to fisheries but apparently caught as a bycatch. IUCN Red List conservation status Least Concern, although bottom-trawl fisheries occur in its range.



*Etmopterus sentosus,* 27 cm TL, male holotype (S Mozambique). Source: Bass *et al.* 1976

# FAMILY SOMNIOSIDAE

## Sleeper sharks

Jenny M Kemper and David A Ebert

Small-sized to gigantic (a few >6 m TL, but most <170 cm TL) with moderately broad head and short to long, flattened snout; trunk cylindrical or slightly compressed, and abdomen often with lateral ridges. Spiracles large and close behind eves. Nostrils wide-spaced (internarial width greater than nostril width) and with simple anterior nasal flaps. Mouth short, nearly transverse, with thin lips; lip furrows short to greatly elongated, encircling mouth or not. Upper teeth small, lanceolate, high-crowned, needle-like, and with narrow erect to semi-oblique cusps and no cusplets or blades; lower teeth large, compressed, high-crowned, narrow and blade-like, with compressed erect to oblique cusp, a distal blade, and no cusplets. Dermal denticles moderate-sized and pedicellate, with flattened, narrow to broad-keeled or smooth leaf-shaped, round or narrow thorn-like crowns, slender pedicels and low bases. Two small, broad dorsal fins, with or without spines; 1st dorsal-fin origin anterior to pelvic-fin origins; 2nd dorsal fin smaller than or subequal to 1st, its origin over to slightly anterior to pelvic-fin insertions; interdorsal space greater than length of 1st dorsal-fin base. Pectoral fins low, angular or rounded, not falcate, and free rear tips rounded and short. Pelvic fins subequal to or larger than pectoral fins and 1st dorsal fin, and subequal to or smaller than 2nd dorsal fin. No anal fin. Peduncle slightly compressed, short to moderately elongated. Caudal fin heterocercal, lower lobe weakly to strongly developed in adults, with strong subterminal notch. Body without photophores (light organs).

Reproductive mode yolk-sac viviparity, with litters of 4–59 pups. Diet includes bony fishes, chondrichthyans, cephalopods and other molluscs, crustaceans, seals, whale meat, carrion, sea birds, echinoderms and jellyfish. Targeted and taken as a bycatch in deepwater shark fisheries and caught by line gear and trawls; used for human consumption, fishmeal, and their squalene-rich liver oil. Mainly deepwater and bottom-dwelling, on continental and insular slopes, but also on submarine ridges and seamounts, at ~200–3 675 m (possibly deeper), but mainly <1 500 m. Some species are oceanic. Circumglobal in most seas, including polar to tropical regions. Seven genera and 17 species; 5 genera and 6 species in WIO.

#### **KEY TO GENERA**

1a	Dorsal fins with spines, although sometimes partially or	
	wholly covered by skin	
1b	No dorsal-fin spines	

Continued ...

#### **KEY TO GENERA**





- 3a Lower teeth with relatively low and more-or-less oblique cusps



- 4b Dorsal-fin spines fairly stout, tips prominent; pectoral fins large, body strongly tapering beyond pectoral fins; pectoral-fin apex when laid back approximately reaches 1st dorsal-fin spine
  Scymodon

# GENUS Centroscymnus

Barbosa du Bocage & De Brito Capello 1864

Snout flattened, short to moderately long; preoral length varying from about equal to much less than distance from mouth to pectoral-fin origins. Body not tapering beyond pectoral fins. Anterior nasal flaps short; lips thick but not pleated or suctorial. Upper teeth very slender, with acute cusps; lower teeth blade-like, interlocked, with short oblique cusps and distal blades. Dermal denticles with low, pedicellate, flat, ovoid crowns, varying from triple-ridged and tricuspid to smooth and acuspidate in adults, and tripleridged and tricuspidate in young. Dorsal fins each with small inconspicuous spine, sometimes covered with skin. First dorsal-fin origin varying from over pectoral-fin bases to behind their free rear tips; 1st dorsal-fin insertion anterior to pelvic-fin origins and closer to pectoral- than to pelvic-fin bases. Second dorsal fin as large as or slightly smaller than 1st, its origin about over middle of pelvic-fin bases. Pectoral-fin free rear tips short, broadly rounded; fin apex when laid back not reaching 1st dorsal-fin spine. Caudal fin asymmetric, upper lobe long and lower lobe short but well-developed, with strong subterminal notch. Two species, both in WIO.

#### **KEY TO SPECIES**

- 1a
   Snout short, preoral length ~3 distance from mouth to

   1st gill slit and less than mouth width; abdomen without

   lateral ridges

   *C. coelolepis*

## Centroscymnus coelolepis

Barbosa du Bocage & De Brito Capello 1864

#### Portuguese dogfish

PLATE 11

*Centroscymnus coelolepis* Barbosa du Bocage & De Brito Capello 1864: 263, Fig. 4 (Portugal, North Atlantic); Compagno 1984\*;

Compagno *et al.* 1989\*; Compagno *et al.* 1991; Ebert *et al.* 1992; Last & Stevens 2009\*; Ebert 2013\*; Ebert *et al.* 2013\*.

*Centroscymnus fuscus* Gilchrist & Von Bonde 1924: 2 (off St Helena Bay, South Africa, SE Atlantic); Barnard 1925; Smith 1949, 1965.

Scymnodon melas Bigelow, Schroeder & Springer 1953: 233,

Fig. 5 (off Georges Bank, North Atlantic); Krefft & Tortonese 1973; Compagno 1984; McEachran & Branstetter *in* Whitehead *et al.* 1984; Castro 2011.

Centrophorus squamosus: Hulley 1971\*; Bass et al. 1976\*.

Centroscymnus macrops Hu & Li in Chu et al. 1982: 305, Fig. 4 (South China Sea); Compagno 1984; White & Last 2013.

Body stout, not strongly tapering after pectoral fins. Snout short, preoral length much less than distance from mouth to 1st gill slit and less than mouth width; lip furrows very short. Upper teeth lanceolate; lower teeth blade-like, with short oblique cusps. Dermal denticles on lateral trunk large, with smooth circular acuspidate crowns in adults and subadults. Dorsal fins about equal in size and height; each fin with very small spine with tip protruding. First dorsal-fin base not extending forward as prominent ridge; fin origin behind pectoral fins. Second dorsal-fin base shorter than space between fin insertion and upper caudal origin; free rear tip well in front of upper caudal origin. Pectoral fins moderately large; fin apex when laid back not reaching 1st dorsal-fin spine. Pelvic-fin rear tips extend beyond 2nd dorsal-fin insertion. Tooth count 43-68/29-42; vertebrae 102-114; spiral valve turns 16-21.

Body uniformly pale to blackish or golden brown. Attains 122 cm TL.

**DISTRIBUTION** Atlantic and Indo-Pacific (widespread). WIO: South Africa (Eastern Cape) and submarine seamounts, including Madagascar Ridge (south of Madagascar) and southern East Indian range (between Africa and Australia).

**REMARKS** Litters of 1–29 (mostly 12–14); size at birth 30–35 cm TL. Males mature at ~85 cm TL, and females at ~100 cm TL. Appears to be depth-segregated by size, sex and stage of maturity. Common on or near bottom on continental slopes and upper and middle rises, at 128–3 675 m (commonly >400 m). Prey includes bony fishes, other sharks, benthic invertebrates, and cetacean and seal meat. Limited interest to fisheries; caught by bottom trawls, fixed nets and line gear in targeted deepwater shark fisheries and as bycatch in other deepwater demersal fisheries. IUCN Red List conservation status Near Threatened globally.



Centroscymnus coelolepis, 93 cm TL, male (South Africa). Source: Bass et al. 1976



*Centroscymnus owstonii,* 58 cm TL, female (New Zealand). Source: Bass *et al.* 1976

## Centroscymnus owstonii Garman 1906

#### Roughskin dogfish

PLATE 11

Centroscymnus owstonii Garman 1906: 207 (Sagami Bay, Japan); Garman 1913\*; Compagno 1984\*; Anderson et al. 1998;

Last & Stevens 2009\*; Ebert 2013; Ebert *et al*. 2013\*.

Centroscymnus cryptacanthus Regan 1906: 437 (Madeira, eastern Atlantic).

Body stout, not strongly tapering after pectoral fins. Snout moderately long, preoral length about equal to distance from mouth to 1st gill slit. Upper lip furrows very short. Upper teeth lanceolate; lower teeth blade-like. Dorsal fins each with spine covered by skin or tip barely exposed. Dermal denticles on lateral trunk large, with mostly smooth, circular, cuspidate and acuspidate crowns in adults. First dorsal fin long and low; fin origin over or behind pectoral-fin bases. Second dorsal fin taller than 1st. Pectoral fins moderately large; fin apex when laid back falling before 1st dorsal-fin spine. Pelvic-fin rear tips below or slightly before 2nd dorsal-fin insertion. Tooth count 36–39/32–40; vertebrae 96–114.

Body pale grey or brown to dark brown or black. Attains 120 cm TL.

**DISTRIBUTION** Atlantic and Indo-Pacific. WIO: South Africa (southern portion of Western Cape) and Madagascar.

**REMARKS** Litter size unknown, but 34 eggs have been observed in a female; size at birth ~30 cm TL. Males mature at 70–84 cm TL, and females at 100–104 cm TL. Found on outer continental shelves and upper continental slopes, on or near bottom, at ~150–1 500 m (commonly deeper than 600 m), sometimes in schools. Prey includes bony fishes and cephalopods. Limited and localised interest to fisheries, mainly as a bycatch.

### GENUS Centroselachus Garman 1913

Moderately sized, fairly slender with body not strongly tapering beyond pectoral fins. Snout greatly elongated; preoral length about equal to distance from mouth to pectoral-fin origins. Lips not thick and fleshy; upper lip furrows very long, their length greater than distance between their anterior ends. Lower teeth with moderately long and semi-oblique cusps, and moderately high and fairly broad roots; upper teeth lanceolate. Dermal denticles on lateral trunk moderately large, with anteriorly smooth but posteriorly ridged, oval cuspidate crowns. Dorsal fins about equal in size and height; each with very small spine with tip protruding. First dorsal-fin base extends forward as prominent ridge; fin origin over pectoralfin bases. Second dorsal-fin base longer than space between fin insertion and upper caudal origin; free rear tip nearly reaching upper caudal origin. Pectoral fins moderately large; fin apex when laid back not reaching 1st dorsal-fin spine. Pelvic-fin rear tips extend to about opposite 2nd dorsal-fin insertion. Tooth count 36-51/30-36; vertebrae 105-119. One species.

# Centroselachus crepidater

Longnose velvet dogfish

(Barbosa du Bocage & De Brito Capello 1864)

PLATE 12

*Centrophorus crepidater* Barbosa du Bocage & De Brito Capello 1864: 262, Fig. 3 [Portugal, North Atlantic].

Centrophorus rossi Alcock 1898: 143 (off Travancore coast, India).

*Centrophorus jonssonii* Saemundsson 1922: 192, Pl. 5, Figs. 1–2 (Iceland, North Atlantic).

Centroscymnus furvescens De Buen 1960: 20, Fig. 7 (off Valparaíso, Chile). Centroscymnus crepidater: Bass et al. 1976; Compagno 1984\*; SSF No. 5.6\*;

Compagno *et al.* 1989\*; Compagno *et al.* 1991; Ebert *et al.* 1992. *Centroselachus crepidater*: Compagno *et al.* 2005\*; Irvine *et al.* 2006; Last & Stevens 2009\*; Ebert 2013\*; Ebert *et al.* 2013\*; Ebert & Stehmann 2013\*; White *et al.* 2014.



Centroselachus crepidater, 90 cm TL, female (New Zealand). Source: Bass et al. 1976, drawn from Garrick 1959

Diagnosis as for genus. Snout very long; lip furrows nearly encircling small mouth. First dorsal-fin base extends forward as prominent ridge.

Body black or blackish brown, with narrow paler fin margins. Attains 105 cm TL.

**DISTRIBUTION** Eastern Atlantic and Indo-Pacific. WIO: South Africa, seamounts of the Madagascar Ridge (south of Madagascar), Aldabra Is., Seychelles and India (Travancore coast).

**REMARKS** Formerly placed in genus *Centroscymnus*. Litters of 1–9; size at birth 28–35 cm TL. Males mature at 60–68 cm TL, and females at 77–82 cm TL. Found on upper continental and insular slopes, on or near bottom, at 200–1 500 m (mainly below 500 m). Prey includes bony fishes and cephalopods. Limited interest to fisheries; taken as a bycatch in bottom trawls.

# GENUS Scymnodon

Barbosa du Bocage & De Brito Capello 1864

Body robust, fusiform, and tapering after pectoral fins. Snout short to moderately long. Upper lip furrows very short. Lower teeth with short oblique cusps, and moderately high and fairly broad roots. Dermal denticles on lateral trunk large, with triple ridges and cusps. Dorsal fins subequal or 2nd slightly higher than 1st; each fin with small but prominent spine. Second dorsal-fin base about as long as space between fin insertion and upper caudal origin; free rear tip well in front of upper caudal origin. Pectoral fins large; fin apex when laid back nearly reaching 1st dorsal-fin spine. Pelvic-fin rear tips fall well in front of 2nd dorsal-fin insertion. Four species, 1 in WIO.

# Scymnodon macracanthus (Regan 1906)

Largespine velvet dogfish

PLATE 12

Centroscymnus macracanthus Regan 1906: 436 (Strait of Magellan). Centrophorus plunketi Waite 1910: 384, Pl. 37 (South Island, New Zealand);

Compagno 1984\*.

Centroscymnus plunketi: Compagno 1984\*.

Proscymnodon plunketi: Compagno et al. 2005\*; Last & Stevens 2009\*; Ebert 2013\*; Ebert et al. 2013\*.

Scymnodon plunketi: White et al. 2015\*.

Scymnodon macracanthus: Weigmann 2016.

Large-bodied with very short snout. Upper teeth lanceolate and dagger-like; lower teeth with short oblique cusps. Dermal denticles on lateral trunk moderately large. Dorsal fins about equal in size and height; each fin with very small spine with tip slightly protruding. First dorsal-fin base extends forward as prominent ridge; fin origin behind pectoral fins. Pectoral fins moderately large. Tooth count 48/32–35; vertebrae 114–115.

Body uniformly dark grey-brown. Attains 170 cm TL.



**DISTRIBUTION** Southwestern Atlantic and Indo-Pacific. WIO: Madagascar Ridge and Melville Ridge (south of Madagascar).

**REMARKS** Litters of up to 36; size at birth 32–36 cm TL. Males mature at 100–131 cm TL, and females at 129–170 cm TL. Inhabits continental and insular slopes of temperate seas, near bottom, at 219–1 550 m (commonly 550–732 m). Occurs in large schools, segregated by size and sex. Prey includes bony fishes and cephalopods. Caught by longlines and demersal trawls; discarded or utilised. IUCN Red List conservation status Near Threatened.

## GENUS Somniosus Le Sueur 1818

Moderately large-sized to gigantic with snout short to fairly long, broadly rounded to pointed, and somewhat flattened. Anterior nasal flaps short; lips thin, and not fringed, pleated or suctorial. Upper teeth small, with narrow, acute, erect cusps; lower teeth much larger, blade-like, interlocked, with low to moderately high oblique or semi-erect cusps and distal blade, edges serrated or not. Dermal denticles with oblique to erect, hooked, cuspidate narrow crowns. Dorsal fins spineless; 1st dorsal fin set on midback and behind pectoral fins, its insertion before pelvic-fin origins; 2nd dorsal fin slightly smaller than 1st, its origin over front half of pelvic-fin bases to posterior of their rear tips. Pectoral fins with short, narrowly to broadly rounded free rear tips and inner margins. Caudal fin semisymmetrical and paddle-shaped, with relatively short upper lobe and long lower lobe, and strong subterminal notch. Five species, 1 in WIO.

## Somniosus antarcticus Whitley 1939

#### Southern sleeper shark

Somniosus antarcticus Whitley 1939: 242 (Macquarie I., south of Tasmania); Yano et al. 2004\*, 2007; Compagno et al. 2005\*;
Murray et al. 2008; Last & Stevens 2009\*; Ebert 2013; Ebert et al. 2013.
Somniosus microcephalus: Bass et al. 1976\*; Duhamel & Hureau 1982; Compagno 1984\*; Duhamel & Compagno 1985; Gushchin et al. 1986;

Compagno 1984°; Dunamei & Compagno 1985; Gushchin *et al.* 1986; Compagno *et al.* 1989; Compagno 1990\*.

Gigantic, heavy cylindrical body; snout short, rounded; precaudal fins very small. Upper teeth lanceolate; lower teeth with short, low, strongly oblique cusps. Dermal denticles on lateral trunk with narrow erect crowns and hooked cusps, giving skin rough texture. Equal-sized dorsal fins; 1st dorsal fin very low, slightly closer to pelvic fins than to pectoral fins; interdorsal space greater than distance from snout to 2nd gill slit. Peduncle short, with lateral keels. Tooth count 37–48/ 49–59; vertebrae 36–38; spiral valve turns 36–41.

Body uniformly grey to pinkish. Attains ~6 m TL.

**DISTRIBUTION** Southern Ocean to Antarctica, and spotty records from southernmost Indian Ocean. WIO: Madagascar Ridge (south of Madagascar), South Africa and Kerguélen Is. and Heard I. (between Madagascar and Antarctica).

**REMARKS** Size at birth ~40 cm TL; males mature at ~4 m TL, and females at ~4.3 m TL. Coastal and epibenthic, on continental and insular shelves and upper slopes, to ~1 440 m deep. Prey includes fishes, pinnipeds, cetaceans and cephalopods. Not commercially important, but taken as a bycatch in trawl and longline fisheries.



Somniosus antarcticus, 4.4 m TL, female (South Africa). Source: FSO

PLATE 12



Zameus squamulosus, 59 cm TL, female (NW Africa). Source: Bass et al. 1976 (drawn from Vaillant 1888)

# GENUS Zameus Jordan & Fowler 1903

Small- to moderate-sized and slender, with low flattened head, and long to short but narrow snout. Eyes moderately large. Mouth short, narrow, transverse; upper lip furrows short. Upper teeth small and spear-like; lower teeth larger, highly erect, and knife-like. Dermal denticles on lateral trunk with or without tricuspid ridges. First dorsal-fin spine relatively large; 2nd dorsal-fin spine slightly shorter than 1st; length of 1st dorsal-fin base greater than 2nd. Pectoral fins small, narrow to moderately broad, leaf-shaped to round. Pelvic fins small, subequal to 2nd dorsal fin. Peduncle long; length of 2nd dorsal-fin base about equal to distance from fin insertion to upper caudal origin. Caudal fin with strong subterminal notch and short lower lobe. Tooth count 42–60/28–38; vertebrae 93–105; spiral valve turns 12–16. One species. The only bioluminescent sleeper shark known at present.

# Zameus squamulosus (Günther 1877)

#### Velvet dogfish

PLATE 12

Centrophorus squamulosus Günther 1877: 433 (off Enoshima, Japan). Centroscymnus obscurus: Vaillant 1888; SSF No. 5.7\*.

Scymnodon ?obscurus: Bass et al. 1976\*

Scymnodon obscurus: Compagno 1984\*.

Scymnodon niger: Chu & Meng 1982.

Scymnodon squamulosus: Compagno et al. 1989\*; Compagno et al. 1991; Ebert et al. 1992.

Zameus squamulosus: Compagno et al. 2005; Last & Stevens 2009\*; Ebert 2013\*; Ebert et al. 2013\*; White et al. 2015.

Small-sized, slender-bodied, with fairly long, narrow snout, and post-oral grooves very long, much longer than upper lip furrows. Dermal denticles on lateral trunk with cross-ridges on crowns. Second dorsal fin slightly larger than 1st, and subequal to pelvic fins. Pectoral fins narrow to moderately broad and leaf-shaped; fin apex when laid back falling well in front of 1st dorsal-fin spine. Tooth count 47–60/32–38; vertebrae 93–105; spiral valve turns 16.

Body uniformly black to dark brown. Attains 89 cm TL.

**DISTRIBUTION** Wide-ranging but patchy in most seas. WIO: South Africa (KwaZulu-Natal), Melville Ridge south of Madagascar, Réunion, Mauritius and India.

**REMARKS** Size at birth ~20 cm TL; males mature at 47–51 cm TL, and females at 59–69 cm TL. Inhabits continental and insular slopes, on or near bottom, at 0–1 511 m (Weigmann 2016). Prey includes bottom-dwelling fishes and invertebrates. Minor interest to fisheries as a bycatch in bottom fisheries.

# FAMILY OXYNOTIDAE

## Roughsharks or prickly dogfishes David A Ebert

Unmistakable small- to moderate-sized (up to ~150 cm TL) with strongly compressed body, extremely rough skin, 2 saillike, spined dorsal fins, and no anal fin. Body humpbacked, very high and triangular in cross-section; lower abdomen with strong lateral ridges between pectoral fins and pelvic fins. Head moderately broad and somewhat flattened; snout blunt. Spiracles large to enormous. Gill slits about subequal. Mouth nearly transverse, very short, small, with thick papillose lips, and more or less encircled by lip folds. Teeth with dignathic heterodonty well-developed; upper teeth spear-like, much smaller than lower teeth; lower teeth compressed, forming saw-like edge; tooth counts 9-18/9-15. Dermal denticles on lateral trunk large and close-set, pedicellate with flattened, broad-keeled, erect or semi-erect trident-like crowns with medial and lateral cusps, slender tall pedicels, and low bases. Dorsal fins very large, broad, angular, high, and often falcate, each with strong stout spine nearly completely buried in fin but with smooth exposed tip. Pectoral fins high, narrow, distally lanceolate, leaf-shaped, subangular or falcate; length of anterior margins varying from about equal to prespiracular length to equal to head length; free rear tips rounded and not elongated. Pelvic-caudal space short, subequal to less than twice length of pelvic-fin bases. Peduncle slightly compressed, short; no lateral

keels or precaudal pit. Caudal fin with subterminal notch. Vertebrae 84–95; spiral valve turns 9–11.

Mostly bottom-dwelling, on outer continental shelf, upper continental slope and insular slopes, at 40–1 067 m. Wideranging but disjunct, in temperate to tropical seas. One genus, *Oxynotus* Rafinesque 1810, with 5 species, 1 in WIO.

## Oxynotus centrina (Linnaeus 1758)

#### Angular roughshark

PLATE 13

Squalus centrina Linnaeus 1758: 233 (Mediterranean Sea).

*Oxynotus shubnikovi* Myagkov *in* Gubanov, Kondyurin & Myagkov 1986: 171, Fig. 59b (Cunene River mouth, Namibia).

*Oxynotus centrina*: Bass *et al.* 1976\*; SSF No. 5.21\*; Compagno *et al.* 1989\*, 1991, 2005\*; Ebert *et al.* 2013\*.

Snout short and blunt; knob-like ridges over eyes, covered with enlarged denticles; spiracles vertically elongated. Upper teeth lanceolate; lower teeth blade-like; tooth count 10/9. First dorsal-fin spine inclined forward.

Body uniformly grey or grey-brown, with darker blotches on head and flanks (sometimes obscure, particularly in adults); paler horizontal line separating dark areas on head and another crosses cheeks below eyes. Attains 150 cm TL (most records <100 cm TL).

**DISTRIBUTION** Eastern Atlantic (Norway to England and Mediterranean Sea, and southward to South Africa [Cape Town]); possibly Mozambique in WIO (needs confirmation).

**REMARKS** Bass *et al.* (1976) noted that records of *O. centrina* from Angola, Namibia and South Africa may be of an undescribed species, differing in having a much shorter interdorsal space. Myagkov (in Gubanov, Kondyurin & Myagkov 1986) named the Namibian rough shark as a new species, *O. shubnikovi*, which was recognised by Compagno *et al.* (1991). However, subsequent examination

of southern African specimens of Oxynotus and comparison with specimens from Namibia and the North Atlantic indicates the supposed differences are due to allometric and ontogenetic changes and individual variation (Yano & Matsuura 2002). Biology sketchily known, despite being widely known in European waters. Viviparous with yolk sac; litters of 7–23; size at birth 21-24 cm TL. In the Mediterranean Sea, mating reportedly occurs in February, with parturition possibly 3-12 months later. Males mature at ~60 cm TL, and females at ~65 cm TL. Bottom-dwelling on continental shelf and upper slope, at 50–660 m (mostly deeper than 100 m), usually over coralline algal and muddy bottoms. Appears to be a weak swimmer and a suction feeder, based on a diet that includes primarily polychaetes. A minor bycatch in bottomtrawl fisheries in North Atlantic. IUCN Red List conservation status Vulnerable because of its occurrence in areas with intense demersal fisheries.

# FAMILY **DALATIIDAE**

Kitefin sharks

Kelsey C James and David A Ebert

Small- to moderately large-sized (to ~180 cm TL) with cylindrical body, narrow conical head and snout, and no anal fin. Spiracles large, close behind eyes. Nostrils widely to narrowly spaced, with simple anterior nasal flaps; internarial width equal to or much greater than nostril width. Mouth nearly transverse, short, with lips thin and smooth or thickened and fringed or pleated; lip folds short to moderately long, thin or thickened, and not encircling mouth. Upper teeth small, lanceolate, with narrow erect cusps; lower teeth highly compressed, broad and bladelike. Dermal denticles small, usually with low-ridged sessile crowns. Two dorsal fins, small- to moderate-sized, angular or rounded-angular, most often without spines. First dorsal fin small- to medium-sized, its base usually over pectoral–pelvic space or partially over pelvic fins. Second dorsal fin subequal



*Oxynotus centrina*, 55 cm TL, female (Namibia). Source: Bass *et al.* 1976

to, or slightly or much larger than 1st dorsal fin; 2nd dorsal-fin base over or just behind pelvic-fin bases. Pectoral-fin anterior margin rounded-angular or rounded-lobate, and mostly shorter than or sometimes subequal to prespiracular length; pectoralfin free rear tip short and broadly rounded. Pelvic fins subequal to or smaller than pectoral fins. Claspers with or without lateral spine. Caudal fin markedly heterocercal to almost diphycercal, and subterminal notch weak to strong. Body usually dark brown or blackish, with pale fins and fin margins; ventral surface with luminous organs (photophores) in some species.

Reproductive mode yolk-sac viviparity, with litters of 6–16, although reproduction is not known for all species. Diet includes bony and cartilaginous fishes, crustaceans, cephalopods, polychaetes, siphonophores and tunicates. One species (*Isistius brasiliensis*) is ectoparasitic on larger pelagic marine vertebrates. Limited fisheries interest exists for these sharks due to their small sizes, but one species (*Dalatias licha*) is caught by longlines and trawls and used for human consumption, fish meal, leather and its squalene-rich liver oil.

Primarily oceanic and mesopelagic, but with some benthic deepwater members, over continental and insular shelves and slopes, at 20–1 800 m (mostly 200–1 000 m). Some species seem to be diel vertical migrators. Nearly circumglobal in warm-temperate to tropical seas. Seven genera and 10 species; 5 genera and 5 species in WIO.

#### **KEY TO GENERA AND SPECIES**



# GENUS Dalatias Rafinesque 1810

Moderately sized with short, broadly conical snout. Gill slits moderately broad and about equal in width. Anterior nasal flaps short, not expanded into barbels. Lips thick, and fringed or pleated. Upper teeth small, with narrow, hooked, needleshaped cusps and no cusplets; lower teeth very large, bladelike, interlocked, with broad, erect, triangular cusps, small distal blades and serrated edges. Dermal denticles with low, flat, ridged, unicuspid crowns. Dorsal fins without spines; 2nd dorsal fin slightly larger than 1st, its base <1.5 length of 1st. First dorsal-fin origin behind pectoral-fin free rear tips, and fin insertion before pelvic-fin origins. Second dorsal-fin origin midway over pelvic-fin bases. No precaudal pit or lateral keels on peduncle. Caudal fin asymmetric, upper lobe long and lower lobe very short or virtually absent; subterminal notch well-developed. Cloaca without luminous gland. One species.



## Dalatias licha (Bonnaterre 1788)

#### Kitefin shark

PLATE 13

*Squalus licha* Bonnaterre (ex Broussonet) 1788: 12 (Cape Breton I., Nova Scotia, Canada).

Scymnorhinus brevipinnis: Smith 1936.

Scymnorhinus licha: SSF No. 5.22\*.

Dalatias licha: Bass et al. 1976\*; Compagno et al. 1989\*; Compagno et al. 2005\*; Ebert 2013\*; Ebert et al. 2013\*.

Diagnosis as for genus. Tooth count 16-21/17-20.

Body greyish to black or blackish brown, sometimes violet with black spots; posterior margins of most fins translucent. Attains 159 cm TL, possibly 182 cm TL.

**DISTRIBUTION** Mediterranean Sea, Atlantic, western and central Pacific, and Indian oceans. WIO: South Africa (Eastern Cape and KwaZulu-Natal) and Mozambique.

**REMARKS** Litters of 3–16; size at birth 30–40 cm TL. Males mature at ~100 cm TL, and females at ~120 cm TL. Primarily solitary with wide depth range, found on outer continental and insular shelves and slopes, at 37–1 800 m (mainly >200 m). Prey includes deepwater bony fishes, some elasmobranchs, cephalopods, crustaceans, polychaetes and siphonophores. Has been found with pieces of large fish in its stomach, suggesting that it bites chunks from live fish prey in a 'cookie-cutter' fashion. Of some interest to commercial fisheries. IUCN Red List conservation status Vulnerable globally, Endangered in European waters.

# GENUS **Euprotomicrus** Gill 1865

444 COASTAL FISHES OF THE WESTERN INDIAN OCEAN | VOLUME 1

Small-sized with cylindrical body and ventral photophores. Snout moderately long and bulbously conical; anterior nasal flaps very short. Gill slits very small and uniform. Lips thin, not fringed, pleated or suctorial. Upper teeth small, not blade-like, with narrow, acute, erect cusps and no cusplets; lower teeth much larger, blade-like, interlocked, with high, broad, nearly erect cusp and distal blade, edges not serrated. Dermal denticles flat and block-like, no posterior cusps on flat depressed crowns. Dorsal fins without spines; 1st dorsalfin tiny, fin origin far behind pectoral-fin free rear tips, fin insertion closer to pelvic fins than to pectoral fins; 2nd dorsal-fin base much longer than 1st, its origin over rear of pelvic-fin bases. Pectoral fins with short, broadly rounded free rear tips and inner margins, not expanded, and acute or lobate. No precaudal pit; peduncle with low lateral keels and no midventral keel. Caudal fin nearly symmetrical, paddleshaped, with short strong upper lobe and long lower lobe; subterminal notch well-developed. One species.

# Euprotomicrus bispinatus

(Quoy & Gaimard 1824)

#### Pygmy shark

*Scymnus bispinatus* Quoy & Gaimard 1824: 197, Pl. 44 (Mauritius, Mascarenes).

*Euprotomicrus bispinatus*: Hubbs *et al.* 1967\*; Bass *et al.* 1976\*; SSF No. 5.18\*; Compagno *et al.* 1989\*; Compagno *et al.* 2005\*; Ebert 2013\*; Ebert *et al.* 2013\*.

Diagnosis as for genus. Tiny, with bulbous snout and large eyes; peduncle with low lateral keels; ventral photophores; tooth count 19–21/19–23.

Body blackish, with conspicuously pale-edged fins. Attains 27 cm TL.

**DISTRIBUTION** Oceanic and amphitemperate, throughout central South Atlantic, central South and North Pacific, and southern Indian oceans. WIO: South Africa (KwaZulu-Natal), Madagascar and Mauritius.

PLATE 13



**REMARKS** Litters of ~8; size at birth 6–10 cm TL. Males mature at 17–19 cm TL, and females at 22–23 cm TL. Pelagic, from 1 829–9 938 m, but occurs at or near surface at night, apparently descending to midwater or well below 1 500 m during daytime. Prey includes deepwater squid, bony fishes and some crustaceans.

# GENUS Heteroscymnoides Fowler 1934

Small-sized with very long bulbously conical snout. Anterior nasal flaps very short. Gill openings very small and uniform. Lips thin, unpleated. Upper teeth small, with narrow, acute, erect cusps and no cusplets; lower teeth much larger, bladelike, interlocked, with high, moderately broad semi-erect cusp and distal blade, edges not serrated. Dermal denticles flat, but with pedicels with lanceolate, ridged, wedge-shaped, monocuspidate crowns. Dorsal fins without spines; 1st dorsal fin set far forward, fin origin over pectoral-fin bases, fin insertion closer to pectoral fins than to pelvic fins; 2nd dorsal fin slightly larger than 1st but with equal base length, its origin midway over pelvic-fin bases. Pectoral fins with short, narrowly rounded free rear tips and inner margins, not expanded, and acute or lobate. No precaudal pit or lateral or midventral keels on peduncle. Caudal fin semi-symmetrical, almost paddleshaped, with moderately long upper lobe and well-developed lower lobe; subterminal notch strong. One species.

# Heteroscymnoides marleyi Fowler 1934

#### Longnose pygmy shark

PLATE 14

Heteroscymnoides marleyi Fowler 1934: 240, Fig. 4 (Durban, KwaZulu-Natal, South Africa); Bass et al. 1976\*; SSF No. 5.19\*; Compagno et al. 1989\*; Compagno et al. 2005\*; Ebert 2013\*; Ebert et al. 2013\*.

Diagnosis as for genus. Tooth count 22/23.

Body dark brown, with conspicuous dark-and-pale-banded fin margins. Attains at least 36.5 cm TL.

**DISTRIBUTION** Likely circumglobal, but limited, in cold sub-Antarctic waters and current systems of Southern Hemisphere: records from southeastern Atlantic (Walvis Ridge), southeastern Pacific (Chile) and southwestern Indian Ocean (KwaZulu-Natal, South Africa).

**REMARKS** Rare, known only from six specimens. Presumably viviparous with yolk sac; probably has few young; size at birth ~13 cm TL. Oceanic and epipelagic, from surface to ~502 m deep, over waters 830–4 000 m deep. Likely feeds on pelagic fishes and invertebrates.



Heteroscymnoides marleyi, 13 cm TL, female holotype, ventral view and dentition (South Africa). Source: Bass *et al.* 1976

## GENUS Isistius Gill 1865

Cigar-shaped body with long abdomen and short tail. Snout short and bulbously conical. Anterior nasal flaps very short. Gill slits small and uniform. Lips expanded, fleshy and suctorial. Upper teeth small, with narrow, acute, erect cusps and no cusplets; lower teeth very large, blade-like, interlocked, with high, broad, erect cusp, edges not serrated; tooth counts 29-43/17-31. Dermal denticles flat and block-like, no posterior cusps on flat depressed crowns. Dorsal fins without spines; 1st dorsal fin set far back, its origin far behind pectoral fins; 2nd dorsal fin slightly larger than 1st, its origin over pelvic-fin rear tips. Pectoral fins with short rounded free rear tips and inner margins, not expanded, and acute or lobate. No precaudal pit; peduncle with low lateral keels and no midventral keel. Caudal fin asymmetrical to nearly symmetrical, with short upper lobe and short to long lower lobe; strong subterminal notch. Two species, 1 in WIO.

# Isistius brasiliensis (Quoy & Gaimard 1824)

Cookiecutter shark

PLATE 14

Scymnus brasiliensis Quoy & Gaimard (ex Cuvier) 1824: 198 (Brazil). Isistius brasiliensis: Bass et al. 1976\*; SSF No. 5.20\*; Jahn & Haedrich 1987; Compagno et al. 1989\*; Compagno et al. 2005\*; Ebert 2013\*; Ebert et al. 2013\*.

Small-sized, with large anteriorly set eyes and suctorial lips. Pectoral fins subquadrate. Pelvic fins larger than dorsal fins. Second dorsal-fin height about equal to 1st; space between 2nd dorsal-fin insertion and caudal-fin origin long. Caudal fin large and nearly symmetrical, with long lower lobe. Tooth count 30–37/25–31.

Body medium grey or grey-brown, with pale-edged fins; prominent dark collar-marking around branchial region; tips of caudal-fin lobes blackish. Photophores on lower abdomen allow it to glow bright green. Attains 56 cm TL. **DISTRIBUTION** Circumtropical (patchy), in Atlantic, Pacific and Indian oceans. WIO: South Africa, Madagascar and Mauritius.

**REMARKS** Little known. Litters of ~6–12; size at birth 14–15 cm TL. Males mature at 31–37 cm TL, and females at 38–44 cm TL. Inhabits epipelagic and bathypelagic zones in tropical regions, at 85–3 500 m. Poor swimmer; makes diel vertical migrations, ascending to midwater or near surface at night. Prey includes squid, gonostomatid fishes and crustaceans; an ecoparasite on large marine organisms, attaching itself to sides of elasmobranchs, bony fishes, phocid seals, dolphins and other cetaceans, and then twisting to cut out conical plug of flesh. Of little interest to fisheries; occasionally caught in bottom trawls.

# GENUS Squaliolus Smith & Radcliffe 1912

Head cylindrical, snout long and slightly pointed. Upper margin of orbit nearly straight to broadly arched or angular and chevron-shaped. Anterior nasal flaps very short. Gill slits very small and uniform. Lips thin, and not fringed, pleated or suctorial; upper lip with or without paired lateral papillae. Upper teeth small, with narrow, acute, erect cusps and no cusplets; lower teeth much larger, blade-like, interlocked, with high, moderately broad, nearly erect cusp and distal blade, edges not serrated. Dermal denticles on lateral body flat, blocklike, with no posterior cusps on flat depressed crowns. First dorsal fin with small spine; fin anteriorly set, its origin about opposite pectoral-fin inner margins or free rear tips. Second dorsal-fin base about twice length of 1st; fin origin over front half of pelvic-fin bases. Pectoral fins with short, narrowly rounded free rear tips and inner margins. No precaudal pit; peduncle with low lateral keels but no midventral keels. Caudal fin nearly symmetrical, paddle-shaped; strong subterminal notch. Cloaca normal, not expanded as luminous organ. These are among the smallest known species of living sharks. Two species, 1 in WIO.



Isistius brasiliensis, 43 cm TL, female (Gulf of Mexico). Source: Bass et al. 1976, drawn from from Bigelow & Schroeder 1948

# Squaliolus laticaudus Smith & Radcliffe 1912

#### Spined pygmy shark

PLATE 14

*Squaliolus laticaudus* Smith & Radcliffe *in* Smith 1912: 684, Pls. 50, 54, Fig. 4 (Batangas Bay, Luzon I., Philippines); Compagno *et al.* 2005\*; Ebert 2013; Ebert *et al.* 2013\*.

Eyes large, upper margin of orbit nearly straight. Upper lip without pair of lateral papillae; tooth count 22–23/16–21. First dorsal-fin spine sometimes covered by skin. Vertebrae 55–62.

Body blackish or blackish brown, with conspicuously pale fin margins; ventral surface densely covered with photophores. Attains  $\sim$ 28 cm TL.

**DISTRIBUTION** Nearly circumtropical, in Atlantic and Indo-Pacific. WIO: Somalia.

**REMARKS** Little known. Size at birth <10 cm TL; litter size probably small, but 12 mature eggs observed in single ovary of a mature female. Males mature at ~15 cm TL, and females at 17–20 cm TL. Epipelagic, near continental and island land masses, at 200–500 m; possibly occurs in aggregations as well as solitarily. Makes diel vertical migrations, ascending at night only to the upper limit of its depth range (avoids surface). Prey includes deepwater squid and bony fishes.



Squaliolus laticaudus, 14 cm TL, female (Philippines). Source: Bass et al. 1976

# ORDER PRISTIOPHORIFORMES

# FAMILY **PRISTIOPHORIDAE**

### Sawsharks

David A Ebert

Distinctive small- to medium-sized (up to ~150 cm TL, but a few <70 cm TL) with slender, cylindrical body; head greatly flattened and somewhat expanded laterally; snout greatly elongated, flattened, laterally expanded, with close-set lateral and ventral 'sawteeth' and pair of string-like ventral rostral barbels before nostrils. Eyes dorsolateral on head; no nictitans, secondary lower eyelids or subocular pouches, but upper eyelid not fused to eyeball. Spiracles large and set just behind eyes. Five or 6 paired gill slits (in Pristiophorus and Pliotrema, respectively), lateral on head, posteriormost just before pectoral-fin origins. Nostrils longitudinal on snout and separate from mouth; no barbels, oronasal grooves or circumnarial grooves; anterior nasal flaps short and not reaching mouth. Mouth small, subterminal, broadly arched and short, extending slightly behind eyes; lip folds short on both jaws; teeth weakly differentiated, no enlarged teeth. Dermal denticles covering entire body, but not enlarged as thorns or spines. Two spineless dorsal fins; 1st dorsal-fin origin behind pectoral-fin free rear tips and well before pelvic-fin bases. Pectoral fins rather large, not fused to head. Pelvic fins small, and vent continuous with their inner margins. Peduncle with long, thick lateral ridges. No anal fin. Caudal-fin axis nearly straight, with long upper lobe and no lower lobe. Total vertebrae 132-157, precaudal vertebrae 90-108; spiral valve turns 6-10.

Reproductive mode yolk-sac viviparity, with litters of 5–17. Food habits of this group are poorly known, but known prey includes small fishes, crustaceans and squid. No directed fisheries for these sharks in WIO, but taken incidentally in groundfish trawl fisheries; elsewhere, especially in Australia, directed sawshark fisheries occur. Sometimes found in aggregations. Harmless to people, unlike the larger sawfishes (Pristidae).

Found in temperate and tropical benthic and epibenthic waters, over continental and insular shelves and upper slopes, from close inshore to ~950 m deep. Some temperate-water species occur in shallow bays and estuaries, near intertidal

zone or on offshore sand and gravel banks, down to upper continental slope; tropical species occur mainly on upper continental and insular slopes. Sporadic, disjunct distribution in western North Atlantic, eastern South Atlantic, Indian, and western Pacific oceans. Fossil records indicate a formerly almost worldwide distribution during the Tertiary, including into the eastern Pacific and eastern Atlantic oceans (including the Tethys-Mediterranean Sea). Two genera and 10 species; both genera and 4 species occur in WIO in depths <200 m (*Pristiophorus nancyae* Ebert & Cailliet 2011 only known from deep water and not included here).

# GENUS Pliotrema Regan 1906

Six paired gill slits. Barbels posteriorly situated on snout, snout length 64–67% preoral length. Larger rostral sawteeth with 2–8 mesially directed barbs along posterior edges; teeth with transverse ridges on basal ledges. Dermal denticles on lateral trunk widely spaced and only semi-imbricated in large specimens, usually separated by distance greater than crown lengths. Three species, one described here; see key (from Weigmann *et al.* 2020) for *P. annae* (from 20–35 m; too late for inclusion of species account), and a 3rd species *P. kajae* from deep water.

#### **KEY TO SPECIES**

- barbers about natiway norm ostrantip to mouth, prebarbel length subequal to distance from barbel origin to upper-jaw symphysis; rostrum constricted between barbel origin and nostrils; dorsal surface with or without 2 longitudinal stripes

Continued ...

#### **KEY TO SPECIES**

- 2b Snout short, head length 34.2–34.5% TL, preorbital length 21.7–22.0% TL, preoral length 24.6–25.1% TL, prebarbel length 12.6–12.7% TL, barbel origin to upper-jaw symphysis 12.1–12.3% TL; 16–17 large lateral rostral teeth; 35–37 upper-jaw tooth rows, jaw teeth without basal folds; medium to dark brown dorsally without longitudinal stripes, white ventrally with some indistinct dark blotches on belly, posterior fin margins conspicuously white-edged ...... *P. annae* [Tanzania (Zanzibar); Plate 15]

## Pliotrema warreni Regan 1906

Sixgill sawshark

PLATE 15

Pliotrema warreni Regan 1906: 1, Pl. 1 (KwaZulu-Natal, South Africa;
False Bay, South Africa); Bass et al. 1975\*; Compagno 1984\*;
SSF No. 20.1\*; Compagno, Ebert & Smale 1989\*; Compagno,
Ebert & Cowley 1991; Compagno, Dando & Fowler 2005\*; Ebert 2013\*;
Ebert et al. 2013\*.

Small-sized with 6 paired gill slits; barbs on posterior edges of larger rostral teeth, and barbels positioned closer to mouth as

compared with other species; teeth in mouth small, unicuspid. Tooth count 36–43/31–38; vertebrae 146–157; spiral valve turns 7 or 8.

Body pale brown dorsally, paler to white ventrally. Attains at least 136 cm TL.

**DISTRIBUTION** Southern Africa: southeastern Atlantic (Table Bay, South Africa) and eastwards in WIO, from South Africa (False Bay to KwaZulu-Natal) to southern Mozambique and southeastern Madagascar.

**REMARKS** Litters of 5–17: size at birth 35–37 cm TL. Males mature at ~83 cm TL, and females at ~110 cm TL. Benthic and epibenthic in offshore areas; partially segregates by depth, with young in shallower water. Neonates, small juveniles, and females with full-term foetuses have been collected in WIO off South Africa, suggesting pupping grounds there; larger individuals, including adult males, appear wider-ranging. Most records from warm-temperate and subtropical waters along southern Cape coast of South Africa, at 60-290 m, but recorded to 500 m. Feeds on small fishes, crustaceans and squid. Larger individuals may have pale-coloured parallel cuts and scratches, suggestive of combat scars inflicted by the oral and rostral teeth of other sawsharks. Predators include the tiger shark Galeocerdo cuvier. Taken as bycatch of demersal fisheries off South Africa, but not utilised. Has not been kept in captivity. This species requires study as it is endemic to southern Africa with a relatively restricted geographic, bathymetric and possibly habitat distribution, and displays morphological vulnerability to net gear while intensive offshore trawl fisheries occur in its known range.



Pliotrema warreni, 36 cm TL, immature male (South Africa). Source: Bass et al. 1975

# ORDER SQUATINIFORMES

# FAMILY SQUATINIDAE

# Angel sharks

David A Ebert

Medium-sized sharks (~90-240 cm TL) with uniquely shaped flattened and batoid-like body with pectoral fins not attached to head but the triangular anterior portion of fins covering underside of head in area of gill slits. Head greatly depressed and laterally expanded; snout extremely short and truncated, without lateral sawteeth or rostral barbels. Five paired gill slits, laterally placed, posteriormost before pectoral-fin origins. Nostrils terminal on snout and separate from mouth, but anterior nasal flaps overlap mouth; anterior nasal flaps with barbels; no oronasal grooves or circumnarial grooves. Eyes dorsal on head; spiracles large. Mouth large, terminal, broadly arched and subangular, terminating about opposite or slightly behind eyes; lip folds very large, on both jaws. Teeth moderately differentiated, with slightly enlarged anterior teeth and no enlarged molars, and no gap or small intermediate teeth between anterior and lateral teeth in upper jaw. Dermal denticles covering entire body or absent from underside; some denticles on head and midline of back more or less enlarged as thorns or spines. Pectoral fins very large, expanded and ray-like; pelvic fins very large, inner margins lateral to vent. Two spineless dorsal fins set rearwards on tail; 1st dorsal-fin origin over or behind pelvic-fin rear tips. No anal fin. Peduncle with short thick keels on caudal base, but no elongated lateral dermal ridges. Caudal fin with high rounded upper lobe and expanded lower lobe. Vertebrae 119-146; intestinal valve of conicospiral type.

Reproductive mode yolk-sac viviparity, with litters of 1–25. Some species are heavily patterned dorsally, and most are uniformly pale ventrally. Angel sharks (also known as monkfish) occur in cool-temperate to tropical continental waters, from close inshore to ~1 300 m depth. Ambush predators that lie mostly buried in sand and mud bottoms with only eyes and spiracles protruding; many species are intensively fished.

Globally wide-ranging throughout Atlantic and Pacific oceans, but absent from most of Indian Ocean and central Pacific. One genus.

# GENUS Squatina Duméril 1806

Diagnosis as for family. At least 20 species, several of these undescribed; 1 species known in WIO (specimens from East Africa should be compared with southern African material to determine whether a single species is represented).

# Squatina africana Regan 1908

African angel shark

PLATE 16

*Squatina africana* Regan 1908: 248, Pl. 38 (Durban Bay, KwaZulu-Natal, South Africa); Bass *et al.* 1975\*; Compagno 1984; SSF No. 21.1\*; Compagno *et al.* 1989\*; Compagno *et al.* 2005\*; Ebert 2013\*; Ebert *et al.* 2013\*.

Head greatly depressed and interorbital space concave. Spiracles with 14 pseudobranchial folds; spiracle width 1.2–1.4 eye length. Nasal barbels simple, flat, with fairly narrow, tapering or spatulate tips and weakly fringed bases. Anterior nasal flaps smooth or moderately fringed, and tips expanded as simple narrow-based lobe; posterior nasal flaps greatly enlarged and moderately fringed. Moderately large thorns on snout and interorbital space above and medial to eyes, and pair of thorns between spiracles in young individuals, but clusters of numerous large denticles in adults; no enlarged predorsal thorns on midline of back; adult males with enlarged alar thorn patch and a few thorns on apices of pelvic fins. Dermal denticles closely spaced but not imbricated, forming rough surface; denticles with slender (in young) to thick (in adults) posteriorly hooked cusp, and with 4 ridges and distal scalloping; ventral surface smooth. Pectoral-fin apex angular, posterior margin somewhat concave, and free rear tip usually closer to pelvic-fin origin than to pelvic-fin apex (sometimes equidistant). First dorsal-fin origin about opposite or slightly after pelvic-fin rear tips. Caudal-fin postventral margin oblique. Vertebrae 121-143; tooth count 20-22/18-20.

Body greyish or reddish brown dorsally, white ventrally; numerous pale and dark spots on dorsal surface, often with large ocelli with granular centres, and symmetrical darker bands or blotches. Attains 122 cm TL. **DISTRIBUTION** Tropical and warm-temperate waters of WIO: South Africa (Western Cape to KwaZulu-Natal [commonest]), Mozambique, southern Madagascar, Mauritius (Mascarenes), Tanzania and possibly Socotra.

**REMARKS** Benthic and epibenthic on continental shelf and uppermost slope, from surfline to ~494 m deep, but mostly 60–300 m. Litters of 1–12 (average 6); size at birth 28–30 cm TL; gestation period probably ~12 months. Males mature at 77–95 cm TL, and females at 82–107 cm TL. Mating appears to take place in February and March, with embryos of various sizes present from April through January, and the largest embryos typically found in October. Off KwaZuluNatal, South Africa, it occurs inshore along open beaches, near river mouths, and on offshore banks, but not in estuaries; the Thukela Bank appears to be a nursery ground as neonates are commonly taken as a bycatch in the prawn trawl fishery. Feeds on small bony fishes (mainly Carangidae, Haemulidae and Sparidae), cephalopods and large shrimp. Limited interest to fisheries, often captured as bycatch of bottom trawls and in anti-shark gillnets; not used as far as known. IUCN Red List conservation status Near Threatened; population trends should be monitored because of its occurrence in areas off South Africa and Mozambique subjected to intensive demersal trawling.



Squatina africana, 44 cm TL, immature male (South Africa). Source: Bass et al. 1975

# ORDER HETERODONTIFORMES

# FAMILY HETERODONTIDAE

# Bullhead sharks

Kelsey C James and David A Ebert

Small- to medium-sized and stout-bodied with blunt piglike snout. Eyes slightly elongated, with very prominent supraorbital ridge; spiracles small, set below and behind eyes. Anterior nasal flap J-shaped, encircling most of nostril, or lobate. Mouth small; lip folds moderately long to long. Teeth similar in both jaws; anterior teeth small and raptorial with 0–3 cusp pairs, and posterior teeth molariform. Two dorsal fins, each with stout spine. First dorsal fin triangular, medium to large, elongated in some species, and larger than 2nd dorsal fin; 1st dorsal-fin base well ahead of pelvic fins and sometimes over pectoral-fin bases. Anal fin midway between pelvic fins and caudal fin. No precaudal pit. Caudal-fin lower lobe moderately or well developed. Vertebrae 103–123; spiral valve turns 7. Body colouration includes dark saddles, spots or intricate stripes.

Mostly coastal in warm and temperate seas, but also found over outer continental shelf, to ~245 m deep, and almost always closely associated with the bottom. Commonly found in rocky habitats or on sandy bottom, most bullhead sharks are nocturnal and hide in crevices and caves during daytime. Reproductive mode oviparity, with large screw-shaped egg cases laid in pairs. Oviposition can be seasonal, in spring or summer, or year round; gestation period 6-12 months, depending on water temperature. Most species mature at >55 cm TL. Most feed predominantly on hard benthic invertebrates, including sea urchins, crustaceans and molluscs, and occasionally on small demersal fishes. No species are important to fisheries but several are a common bycatch in trawl nets, bottom gillnets and traps; catches are generally discarded but occasionally used for human consumption, fishmeal, or for their skin. Some species keep well in public aquariums, and small individuals are sometimes caught for the aquarium trade. None of the species are dangerous to people.

One genus, *Heterodontus* Blainville 1816, with 9 species, 2 in WIO.

#### **KEY TO SPECIES**

# Heterodontus omanensis Baldwin 2005

Oman bullhead shark

PLATE 17

Heterodontus omanensis Baldwin 2005: 262, Figs. 1–2 (Gulf of Masira, Oman, Arabian Sea).

*Heterodontus* sp. A: Randall 1995\*; Compagno 2001\*, 2005\*; Ebert *et al.* 2013\*.

Medium-sized with moderately low supraorbital ridges. Anterior teeth with cusp and pair of cusplets in adults; posterior molars unknown. Lateral trunk denticles fairly rough. First dorsal-fin low, rounded-angular, with free rear tip; fin origin before pectoral-fin insertion, and fin insertion well forward of pelvic-fin origin. Second dorsal fin nearly as large as 1st. Anal fin subangular. Vertebrae 106.

Body tan or brown, with 4 or 5 broad, diffuse-edged brown saddles distributed from snout tip to caudal-fin origin (no pale or dark spots or harness-type marking); dorsal fins, caudal fin and pectoral fins with black tips, and dorsal-fin tips also with white spot; single dark blotch under eyes. Colour of hatchlings possibly differs from adults. Attains 61 cm TL.

DISTRIBUTION WIO: Arabian Sea (Oman).

**REMARKS** Apparently common in northern Indian Ocean, but little known. Egg cases screw-shaped with two full turns and long tendrils extending from base. Males mature at ~52 cm TL, and females at ~61 cm TL. Caught by commercial trawlers off southern Oman: the type specimen was taken at ~72 m over soft bottom.

# Heterodontus ramalheira (Smith 1949)

#### Whitespotted bullhead shark

PLATE 17

*Gyropleurodus ramalheira* Smith 1949: 367, Fig. 1 (off Inhambane, Mozambique).

Heterodontus ramalheira: SFSA No. 4a\*; Pinchuk 1969; SSF No. 4.1\*; Van der Elst & Vermeulen 1986; Compagno *et al.* 1989\*; Compagno 2001\*; Compagno *et al.* 2005\*; Ebert 2013\*; Ebert *et al.* 2013\*.

Medium-sized with moderately high supraorbital ridges. Anterior teeth in adults with cusp and pair of cusplets; posterior molars strongly ridged and not greatly expanded or rounded. Lateral trunk denticles large and rough. First dorsalfin triangular, its origin far before pelvic fins. Second dorsal fin smaller than 1st, its origin over pelvic fins. Anal fin angular and falcate. Vertebrae 104–116. Body dark red-brown with white spots; hatchlings paler and without dark harness-type marking but with darker, indistinct saddles. Attains at least 85 cm TL.

**DISTRIBUTION** WIO: South Africa (KwaZulu-Natal) to south-central Mozambique, Somalia, eastern Arabian Peninsula and southern Oman.

**REMARKS** Apparently uncommon or rare and little-known; benthic on outer continental shelf and upper slopes of East Africa. An unusually deepwater species for the family, at 40–275 m, with most records from >100 m. Egg cases not described, but hatchlings ~18 cm TL. Males mature at ~60 cm TL, and females at 75–83 cm TL. Diet mostly unknown, but crabs found in stomach of two individuals. Occasionally taken as a bycatch in commercial bottom trawls, such as for shrimp, off Mozambique and South Africa.



Heterodontus ramalheira, 64 cm TL, male (S Mozambique). Source: Bass et al. 1975

# ORDER ORECTOLOBIFORMES

# FAMILY HEMISCYLLIIDAE

# Bamboo sharks

Brett A Human

Small-sized (to ~110 cm TL) and slender with cylindrical to slightly depressed body, with or without dorsal or lateral ridges, and with very long tail. Head length 12-14% TL; 5 paired gill slits, small and lateral on body, with 5th slit overlapping 4th; snout broadly to acutely rounded, not depressed, and with nasal barbels, circumnarial folds and grooves, and oronasal groove. Eyes dorsolateral, with strong subocular ridges, and no nictitans; spiracles large, set just below eyes. Mouth small, transverse or nearly so, subterminal to inferior, with welldeveloped lip folds fused medially into a complete post-oral fold. Teeth similar in both jaws, with strong median cusp and 1 or no lateral cusps; tooth counts 26-35/21-32. Fins rounded to angular or weakly falcate. Two spineless equal-sized dorsal fins, set far back; 1st dorsal fin over or slightly behind pelvicfin bases. Pectoral fins small and rounded, gill slits extending well over pectoral-fin bases; pelvic fins subequal to pectoral fins. Anal fin low, moderate to long, smaller than 2nd dorsal fin, and more or less contiguous with caudal fin but separated by notch; anal-fin origin after 2nd dorsal-fin insertion. Peduncle long, cylindrical, without ridges or pits; distance from cloaca to anal fin >26% TL. Caudal fin elongated, weakly heterocercal, and without lower lobe. Vertebrae 151-192; spiral valve turns 12-20. Juveniles and adults typically with very different colour patterns.

Strongly associated with coral and rocky reefs. Also known as longtailed carpetsharks, as they possess muscular pectoral and pelvic fins that allow them to clamber over reef bottoms. Often seen in very shallow water and tidal pools, their preferred habitat makes them highly susceptible to humaninduced habitat degradation. Oviparous where known, with conflicting reports as to whether oviparity is single or multiple. Common to rare, and all species are poorly known biologically and taxonomically; poorly represented in fish collections and more specimens are needed to clarify taxonomic identities and distributions. Although not targeted in any fishery they can be common bycatch species and are often exploited in the aquarium trade due to their colourful patterns and hardiness. Two genera and 15 species, plus several undescribed species; one genus and at least 5 species in WIO.

#### **KEY TO GENERA**

- 1b
   Nostrils terminal; snout length <3% TL; mouth closer to snout tip than to eyes</th>
   Hemiscyllium



# GENUS Chiloscyllium Müller & Henle 1837

Predorsal and interdorsal ridges variably developed, and lateral body ridges present or not. Mouth closer to eyes than to snout tip. Eyes and supraorbital ridges weakly elevated. Length of nasal barbels >1.3% TL. Cloaca to anal fin <38% TL. No large dark spot over pectoral fins. Found mostly in shallow water, but ranging to 100 m deep. Indo-Pacific distribution, primarily tropical but with a few species extending into subtropical or temperate waters. At least 7 species, 5 in WIO.

#### **KEY TO SPECIES**

1a 1b	Lateral body ridges present	
2a	Length of anal-fin base equal to or greater than distance from lower caudal origin to subterminal notch; anal-fin origin far behind 2nd dorsal-fin free rear tip	
2b	Length of anal-fin base less than distance from lower caudal origin to subterminal notch; anal-fin origin behind 2nd dorsal-fin free rear tip	
3a	First dorsal-fin origin above or behind rear half of pelvic-fin bases	
3b	First dorsal-fin origin before or above front half of pelvic-fin bases	

#### **KEY TO SPECIES**

# Chiloscyllium arabicum Gubanov 1980

#### Arabian bamboo shark

#### PLATE 18

Chiloscyllium arabicum Gubanov in Gubanov & Shleib 1980: 14,

Figs. 6–7 (Persian/Arabian Gulf); Compagno 1984\*, 2001\*; Randall 1995\*; Carpenter *et al.* 1997\*; Al-Shuaily & Henderson 2003\*; Compagno *et al.* 2005\*; Ebert *et al.* 2013\*.

*Chiloscyllium confusum*: Dingerkus & DeFino 1983\*; Goto 2001; Eschmeyer 2010.

Predorsal and interdorsal ridges well-developed. First dorsalfin base shorter than 2nd dorsal-fin base, rarely equal; 1st dorsal-fin origin above rear half of pelvic-fin bases. Length of anal-fin base less than distance from lower caudal origin to subterminal notch; anal-fin origin not far behind 2nd dorsal-fin free rear tip. Cloaca to tail tip 61–68% TL. Vertebrae 141–175.

Adults uniformly tan dorsally, paler ventrally; juveniles identical to adult colouration but with pale spots on fins. Attains 70 cm TL.

**DISTRIBUTION** WIO: Arabian Sea, Persian/Arabian Gulf, Oman, Pakistan and India.

**REMARKS** Goto (2001) and Eschmeyer (2010) recognised *C. confusum* as a valid species, but without supporting evidence; Compagno (2001) found *C. confusum* agreed with *C. arabicum*, however, and this has been followed by most subsequent authors. Confusion with *C. griseum* may have resulted in some erroneous distribution and abundance data. Lays up to 4 egg cases on coral reefs, hatching after 70–80 days; also found on rocky shores and in mangrove estuaries and lagoons. Has reproduced in captivity. Feeds on shelled molluscs, squid, crustaceans and fishes. IUCN Red List conservation status Near Threatened.

# Chiloscyllium griseum Müller & Henle 1838

#### Grey bamboo shark

PLATES 18 & 19

*Chiloscyllium griseum* Müller & Henle 1838: 19, Pl. 4 (Puducherry, India; Japan); Smith 1949\*; Dingerkus & DeFino 1983\*; Compagno 1984\*, 2001\*; Carpenter *et al.* 1997\*; Morón *et al.* 1998; Al-Shuaily & Henderson 2003\*; Compagno *et al.* 2005\*; Ebert *et al.* 2013\*. *Chiloscyllium indicum*: Thompson 1914.

No predorsal or interdorsal ridges. First dorsal-fin base longer than 2nd dorsal-fin base; 1st dorsal-fin origin above rear half of pelvic-fin bases. Length of anal-fin base less than distance from lower caudal origin to subterminal notch; anal-fin origin not far behind 2nd dorsal-fin free rear tip. Tail long, thick; cloaca to tail tip 58–64% TL. Vertebrae 156–170; spiral valve turns 15–19.

Adults pale brown dorsally, paler ventrally; juveniles with distinct pale to dark grey-brown bands without black margins. Attains 77 cm TL.



Chiloscyllium arabicum, ~40 cm TL, subadult (Persian/Arabian Gulf).



Chiloscyllium griseum, adult (WIO). Composite

**DISTRIBUTION** Indo-Pacific. WIO: Arabian Sea, Persian/ Arabian Gulf, Oman, Pakistan, India and Sri Lanka; elsewhere to Indonesia and southern Japan.

**REMARKS** Müller & Henle's (1838) illustration on Pl. 4, labelled *C. griseum*, is actually based on a specimen referable to *C. punctatum*. Westernmost records of this species may be confused with *C. arabicum*, and easternmost records may be partly based on *C. hasseltii*. Very poorly known despite being reportedly common within its distribution. Occupies inshore rocky habitat and lagoons; lays oval-shaped egg cases. Assumed to feed mostly on invertebrates. Commonly caught in fisheries and used for human consumption in Pakistan, India and Thailand, and also exploited in the aquarium trade. IUCN Red List conservation status Vulnerable.

# Chiloscyllium indicum (Gmelin 1789)

Slender bamboo shark

PLATE 19

Squalus indicus Gmelin 1789: 1503 (East Indies [Indian Ocean]).

Chiloscyllium indicum: Gilchrist 1902; Thompson 1914; Barnard 1925, 1927\*; Smith 1949\*; Dingerkus & DeFino 1983\*; Compagno 1984\*,

2001\*; Compagno *et al.* 2005\*; Ebert *et al.* 2013\*.

Very slender body, with well-developed predorsal and interdorsal ridges. First dorsal-fin base longer than 2nd dorsalfin base; 1st dorsal-fin origin above rear half of pelvic-fin bases. Length of anal-fin base equal to or greater than distance from lower caudal origin to subterminal notch; anal-fin origin far behind 2nd dorsal-fin free rear tip. Cloaca to tail tip 62–67% TL. Vertebrae 166–170; spiral valve turns 14 or 15.

Adults pale brown dorsally, with many small dark spots, saddles and bars; juveniles with many large dark saddles and bands without dark margins, which fade and separate to create adult patterning. Attains 65 cm TL. **DISTRIBUTION** Indo-Pacific. WIO: Pakistan, India and Sri Lanka; elsewhere to Thailand, Indonesia, China and Japan.

**REMARKS** Despite a long history in southern African literature, this species has not been definitively recorded from there; the confusion seems to have arisen from specimens probably collected in India or Asia, deposited in South Africa, and mislabelled in the early 19th century. Biology virtually unknown, but commonly caught in fisheries in India, Sri Lanka and Thailand and used for human consumption. Found inshore on bottom; also reported from brackish water in Malaysia. IUCN Red List conservation status Vulnerable.

# Chiloscyllium plagiosum

(Anonymous [Bennett] 1830)

#### Whitespotted bamboo shark

PLATE 19

- *Scyllium plagiosum* Anonymous [Bennett] 1830: 694 (Java Sea, Sumatra, Indonesia).
- Chiloscyllium indicum: Gilchrist 1902; Thompson 1914.
- *Chiloscyllium caerulopunctatum*: Bass *et al.* 1975\*; Bauchot & Bianchi 1984\*; Compagno 1984\*; SSF No. 7.1\*.
- *Chiloscyllium plagiosum*: Dingerkus & DeFino 1983\*; Compagno 1984\*, 2001\*; Compagno *et al.* 2005\*; Ebert *et al.* 2013\*.

Predorsal and interdorsal ridges well-developed. First dorsal-fin base longer than 2nd dorsal-fin base; 1st dorsal-fin origin above rear half of pelvic-fin bases. Length of anal-fin base less than distance from lower caudal origin to subterminal notch; anal-fin origin behind 2nd dorsal-fin free rear tip. Cloaca to tail tip 61–67% TL. Vertebrae 161–185; spiral valve turns 16 or 17.

Adults with blotchy, pale to medium brown background, prominent dark saddles and many dark and white or bluish spots; juveniles dark, almost black, or with patterning similar to adults. Attains 95 cm TL.



Chiloscyllium indicum, 32 cm TL, juvenile female. Source: Dingerkus & DeFino 1983



Chiloscyllium indicum, 43 cm TL, adult female. Source: Dingerkus & DeFino 1983



Chiloscyllium plagiosum, 67 cm TL, female (S Madagascar). Source: Bass et al. 1975

**DISTRIBUTION** Indo-Pacific. WIO: Madagascar, India and Sri Lanka; elsewhere to Indonesia, Philippines, Japan and New Guinea.

**REMARKS** *C. plagiosum* recorded from Madagascar is probably a distinct species; however, it is synonymised here due to a lack of published diagnostic characters (although blue spots are noted as present only on specimens from Madagascar). Common, but biology virtually unknown. Nocturnally feeds on bony fishes and crustaceans. Frequently caught in fisheries in Madagascar, India, Thailand and China, and used for human consumption. A popular aquarium species that has reproduced in captivity. IUCN Red List conservation status Near Threatened.

## Chiloscyllium punctatum Müller & Henle 1838

Brownbanded bamboo shark

PLATES 19 & 20

*Chiloscyllium punctatum* Müller & Henle 1838: 18, Pl. 3 (off Jakarta, Java, Indonesia); Dingerkus & DeFino 1983\*; Compagno 1984\*, 2001\*; Compagno *et al.* 2005\*; Ebert *et al.* 2013\*.

Predorsal and interdorsal ridges not well developed. Dorsal-fin rear margins concave, and free rear tips elongated. First dorsalfin base longer than 2nd dorsal-fin base; 1st dorsal-fin origin above or before front half of pelvic-fin bases. Length of anal-fin base less than distance from lower caudal origin to subterminal notch; anal-fin origin below posterior half of 2nd dorsal-fin base. Tail long, thick; cloaca to tail tip 61–64% TL. Vertebrae 136–170; spiral valve turns 20.

Adults with pale to medium brown background, paler ventrally, and usually without other markings, but with pale gill-slit margins; juveniles with prominent broad dark bands on very pale background, fading with maturity to small dark spots on darker background. Attains 132 cm TL.

**DISTRIBUTION** Indo-Pacific. WIO: Arabian Sea and southern India; elsewhere to Thailand, Indonesia, Philippines, Japan, New Guinea and Australia.

**REMARKS** Biology virtually unknown despite relatively wide distribution and abundance. Lays rounded egg cases. Diet includes invertebrates and fishes. Common but cryptic, regularly taken in fisheries in India and South East Asia for human consumption. A common species in aquaria, where it has reproduced. IUCN Red List conservation status Near Threatened.



Chiloscyllium punctatum, 63 cm TL, adult female (Indian Ocean).

# FAMILY STEGOSTOMATIDAE

Zebra shark

David A Ebert

Distinctively large-bodied, somewhat stout, with prominent longitudinal ridges; head broad, conical and somewhat flattened; snout broadly rounded. Nostrils short, with small pointed barbels; oronasal groove long and well-developed. Eves lateral, without strong subocular ridges; spiracles large. Mouth small, nearly transverse. Five paired short gill slits, the last three behind pectoral-fin origins; 4th and 5th gill slits very close, often giving the impression of one slit. First dorsal fin larger than 2nd, its origin well before pelvic-fin origins. Pectoral fins large, broadly rounded. Anal-fin origin about opposite upper caudal origin. Caudal fin greatly elongated, broad, equal to half TL, with strong terminal lobe and subterminal notch. Body with distinct spotting, it is also known as the leopard shark.

Monotypic genus Stegostoma Müller & Henle 1837.

## Stegostoma tigrinum (Forster 1781)

Zebra shark

Squalus tigrinus Forster 1781: 24, Fig. 2, Pl. 13 (Sri Lanka). Stegostoma tigrinum Barnard 1937; Dahl et al. 2019. Stegostoma fasciatum: SFSA No. 30\*; Compagno 1984\*; SSF No. 7.4\*; Compagno et al. 1989\*; Compagno 1999; Compagno 2001\*;

Heemstra & Heemstra 2004\*; Ebert et al. 2013\*; Ebert & van Hees 2015; Weigmann 2016.

Stegostoma varium: Bass et al. 1975\*.

Diagnosis as for family. Teeth small and tricuspidate, similar in both jaws; tooth count 28-33/22-34. Vertebrae 207-243; spiral valve turns 18.

Body yellowish with distinctive pattern of dark brown spots in adults, or with dark saddles and black spots alternating with vertical white or yellowish stripes in juveniles. Attains 250 cm TL (possibly >3.5 m TL).

**DISTRIBUTION** Indo-Pacific (widespread). WIO: Persian/ Arabian Gulf, Red Sea, Gulf of Aden to South Africa (Eastern Cape and KwaZulu-Natal), Madagascar, Mauritius, Maldives, India and Sri Lanka; elsewhere to Indonesia, southern Japan, Marshall Is., Australia New Caledonia and Samoa.

**REMARKS** Oviparous, with large dark brown to purplish black egg cases with fine hair-like fibres that anchor to substrate; egg dimensions approximately 13-17 cm long, 8 cm wide, 5 cm thick. Litter size likely >2, as multiple egg cases have been found in a single oviduct. Inshore shark of tropical continental and insular shelves, from intertidal zone, coral reefs and offshore sediments, to ~62 m deep. Usually solitary, but also found in large aggregations (up to ~50) over sand near reefs, but young seldom seen. Slow but strong-swimming; probably nocturnal, feeds mostly on molluscs (gastropods and bivalves) but also small bony fishes and crustaceans. Commonly taken throughout its range in nearshore fisheries by bottom trawls or longlines, and often sold in local fish markets; captured in smaller numbers by prawn trawls in Australia. Also kept in public aquaria. IUCN Red List conservation status Endangered due to fishing and coral reef degradation, but assessed as Least Concern in Australia where it is abundant.



PLATE 20

Stegostoma tigrinum, 128 cm TL, immature male (South Africa). Source: Bass et al. 1975



Stegostoma tigrinum, 37 cm TL, immature male (South Africa). Source: Bass et al. 1975



Stegostoma tigrinum, 51 cm TL, juvenile (South Africa). Source: SFSA

# FAMILY **GINGLYMOSTOMATIDAE**

Nurse sharks Brett A Human

Moderately sized (up to ~3 m TL) and robust with thick cylindrical or depressed body; head short, broad and depressed (HL 19-27% TL). Five paired gill slits, small, lateral on body, with 4th nearly overlapping 5th. Eyes tiny, lateral or dorsolateral on head, set posterior to mouth; no nictitans; spiracles minute, posterior to and level with eyes, <<sup>1</sup>/<sub>3</sub> eye diameter. Mouth transverse, subterminal, not extending behind eyes, with well-developed lip folds. Teeth moderately sized, similar in both jaws, with strong median cusp and 1-7 lateral cusps; tooth counts 24-38/22-32. Nasal flaps small, with short to moderately long nasal barbels, and well-developed oronasal grooves. Fins rounded or falcate. Two spineless dorsal fins; 1st dorsal fin over or slightly anterior to pelvic fins; 2nd dorsal fin subequal to 1st. Pectoral fins rounded or falcate, fin origin under 4th gill slit. Pelvic fins smaller than pectoral fins. Anal fin large and high, equal in size to 2nd dorsal fin; analfin origin under or slightly posterior to 2nd dorsal-fin origin. Peduncle short, compressed, without ridges or pits. Tail large, elongated, slightly elevated, heterocercal and lower lobe poorly developed. Vertebrae 135–195; spiral valve turns 15–24.

Reproductive mode yolk-sac viviparity, except *Pseudoginglymostoma* which is possibly oviparous. Distributed throughout tropical and subtropical oceans, found inshore and close to bottom, from intertidal zone to at least 70 m deep, and abundant where recorded. Nurse sharks are a common catch in artisanal and commercial fisheries, and used for human consumption, liver oil and leather products.

Three monotypic genera; 2 genera and 2 species in WIO.

#### **KEY TO GENERA AND SPECIES**

- **1b** Fin tips rounded; dorsal fins equal-sized; body cylindrical; tail length <20% TL ...... *Pseudoginglymostoma brevicaudatum*

# GENUS Nebrius Rüppell 1837

Head U-shaped in dorsal view; snout moderately elongated, preoral length ~63% mouth width; eye diameter >1% TL. Nasal barbels elongated, reaching past mouth; teeth with 3 or more lateral cusps either side of median cusp; lower lip trilobate. One species.

# Nebrius ferrugineus (Lesson 1831)

Tawny nurse shark

PLATES 20 & 21

- *Scyllium ferrugineum* Lesson 1831: 95 (New Ireland, Bismarck Archipelago; Waigeo I., Moluccas, Indonesia).
- *Nebrodes macrurus* Garman 1913: 58, Pl. 8, Figs. 7–10 (Mauritius, Mascarenes).
- Nebrius concolor: Bass et al. 1975\*; Dingerkus 1986; SSF No. 7.3\*; Taniuchi & Yanagisawa 1987.
- Nebrius ferrugineus: Compagno 1984\*, 2001\*; Stevens 1984; Compagno et al. 1989\*; Randall & Anderson 1993; Randall 1995\*; Carpenter et al. 1997\*; Marine Research Section [Maldives] 1997; Winterbottom & Anderson 1997; Al-Shuaily & Henderson 2003\*; Compagno et al. 2005\*; Ebert et al. 2013\*.

Diagnosis as for genus. Fins angular; 1st dorsal fin larger than 2nd, its base over pelvic-fin bases. Nasal barbels fairly long. Tooth count 29–33/26–28; vertebrae 189–195; spiral valve turns 23 or 24.

Body uniformly brown, but may slowly change to yellowish or greyish depending on habitat, and paler ventrally; albinism recorded. Attains 3.2 m TL. **DISTRIBUTION** Tropical and subtropical waters of Indo-Pacific (widespread). WIO: Persian/Arabian Gulf, Red Sea, Gulf of Aden, Arabian Sea, Mozambique, South Africa (KwaZulu-Natal), Madagascar, Aldabra, Mauritius, Seychelles, Chagos, Maldives, India and Sri Lanka; elsewhere to Indonesia, southern Japan, Australia, New Caledonia, Society Is. and Marquesas Is.

**REMARKS** Its wide distribution has given rise to multiple synonyms, with those pertaining to WIO discussed in Bass et al. (1975). Viviparous with yolk sac; the only orectolobiform shark known to be intrauterine oophagous. Found on continental and insular shelves, on or near bottom, mainly at 5-30 m. Maintains limited home range; nocturnal and often found resting during daylight hours, sometimes with individuals positioned atop others, although active during day in some regions. Large pharynx and small mouth capable of powerful suction for feeding on invertebrates (crustaceans, cephalopods, echinoderms) and reef fishes; may squirt water from mouth when caught and reportedly makes a grunting sound. Caught frequently in commercial and artisanal nearshore fisheries. Individuals of this species from Japan and Taiwan have been reported missing their 2nd dorsal fin, possibly due to teratogenic pollution. Docile and popular with divers, and common in public aquaria. IUCN Red List conservation status Vulnerable due to high fishing pressure and habitat loss, but assessed as Least Concern in Australia.



Nebrius ferrugineus, 118 cm TL, immature male (N Mozambique). Source: Bass et al. 1975



# GENUS Pseudoginglymostoma

#### Dingerkus 1986

Head broadly rounded and parabolic in dorsal view; snout bluntly rounded, preoral length 34.5–42% mouth width; eye diameter <1% TL. Nasal barbels very short, not reaching mouth; teeth with 1 or 2 lateral cusps either side of median cusp; lower lip not trilobate. Tail length <¼ TL. This genus (and the subfamily Pseudoginglymostomatinae, Compagno 2001) may warrant separation from Ginglymostomatidae, based on morphology, and could be placed in its own family pending a review. One species.

# Pseudoginglymostoma brevicaudatum (Günther 1867)

#### Shorttail nurse shark

PLATE 21

*Ginglymostoma brevicaudatum* Günther *in* Playfair & Günther 1867: 141, Pl. 21 (Zanzibar, Tanzania); Garman 1913; Bass *et al.* 1975\*; Compagno 1984\*; SSF No. 7.2\*.

*Pseudoginglymostoma brevicaudatum*: Dingerkus 1986; Compagno 2001\*; Ebert *et al.* 2013\*.

Diagnosis as for genus. Fins rounded; dorsal fins equal-sized. Tooth count 24–27/22–27; vertebrae 135–143; spiral valve count 15.

Body uniformly dark brown, but paler ventrally. Attains 75 cm TL. **DISTRIBUTION** WIO: Kenya to Mozambique, Madagascar, and possibly Seychelles and Mauritius.

**REMARKS** Reportedly common off East Africa, but poorly known. Assumed oviparous, as one female that lived for at least 33 years in an aquarium laid infertile egg cases. Nocturnal in captivity; diet presumably similar to other members of the family. Probably landed in inshore artisanal and commercial fisheries.

# FAMILY RHINCODONTIDAE

Whale shark David A Ebert

Daviu A Ebert

Enormous, heavy-bodied and fusiform, with wide depressed head and huge terminal transverse mouth. Gill slits huge and internally modified for filter-feeding; nostrils rudimentary; spiracles larger than eyes. Teeth hook-shaped with single cusp; ~300 tooth rows in each jaw. First dorsal fin very large, posteriorly set, with insertion over pelvic fins; 2nd dorsal fin smaller, with insertion over anal-fin base; pectoral fins falcate; caudal fin crescent-shaped, unnotched. Prominent ridges along flanks, with lowest ending in keel on peduncle.

Monotypic.

## Rhincodon typus Smith 1828

Whale shark

PLATES 21 & 22

*Rhincodon typus* Smith 1828: 2 (Table Bay, South Africa); Bass *et al.* 1975\*; SSF No. 8.1\*; Compagno *et al.* 1989\*; Anderson & Ahmed 1993; Beckley *et al.* 1997; Wintner 2000; Ebert 2003\*; Ebert *et al.* 2013\*.

Diagnosis as for family. Vertebrae 153–174; spiral valve turns 68–73.

Body with prominent pattern of white or yellow spots between vertical and horizontal stripes reminiscent of checkerboard pattern, offset by background colour of dark grey, bluish or brown above, becoming white below. The largest living fish, attains at least 18 m TL.

**DISTRIBUTION** Circumglobal in tropical to temperate seas, including Red Sea. WIO: South Africa (Western Cape) to Sri Lanka and northwards to Pakistan.

**REMARKS** Viviparous with yolk sac, without placenta; litters large, up to 300 embryos; size at birth 55-64 cm TL; gestation period and pupping grounds unknown. Males mature at 7-9 m TL, at ~20 years; females mature at ~10 m TL, at 27 years or more. Diet of this huge filter-feeding shark includes plankton, copepods, jellyfish, and pelagic crustaceans such as crabs. Gulps and feeds, often while hanging vertically, by drawing in water at high velocity by expansion of gill cavity, unlike the more passive method of basking sharks. Oceanic, to >700 m deep, but usually found at or near the surface, from close inshore, including bays and lagoons, to far out at sea. Prefers areas of mixing layers of cool upwelled oceanic waters. Highly migratory, often making long-term transoceanic migrations. A docile species that has at times been the subject of regional artisanal fisheries; has become a popular attraction for ecotourism in recent years. IUCN Red List conservation status Endangered; currently protected under CITES.



*Rhincodon typus*, 4.4 m TL, immature female; lateral and dorsal tooth from 8-m-TL specimen, and natural appearance of teeth with all but tips covered by skin (South Africa). Source: Bass *et al.* 1975
# ORDER LAMNIFORMES

# FAMILY CARCHARIIDAE

#### Raggedtooth shark Brett A Human

Large-sized (to at least 3.2 m TL) with stout cylindrical body and long head (HL ~25% TL); snout short, slightly depressed and pointed; snout length less than mouth length; preorbital length 0.3-0.5 times mouth width; eye diameter 0.9-1.4% TL. Five paired gill slits, large, lateral and not extending dorsally, with all slits before pectoral-fin origins. Eyes small, to ~1.4% TL, set dorsolaterally on head over mouth, with prominent lateral head ridge underneath; no nictitating eyelid; spiracles minute. Mouth parabolic, subterminal, extending behind eyes, with lip folds. Teeth large, similar in both jaws, although identifiable as symphysials, anteriors, intermediates, laterals and posteriors; teeth erect with elongated medium cusp and a much shorter lateral cusplet either side of median cusp, becoming compressed posteriorly; tooth counts 40-42/38-42. Vertebrae 156–186. Nasal flaps small, without nasal barbels; no oronasal grooves. Fins angular and broad; dorsal fins, pelvic fins and anal fin subequal. Two spineless low dorsal fins, 1st dorsal-fin base closer to pelvic-fin base than to pectoralfin base. Pectoral fins medium-sized and broad; pectoral-fin origin behind 5th gill slit. Anal-fin base under 2nd dorsal fin. Peduncle compressed, without lateral keels, and with upper crescentic precaudal pit, but no lower pit. Caudal fin heterocercal, its dorsal margin <1/3 precaudal length. Intestinal valve of ring type.



Reproductive mode oophagous, the only shark species where adelphophagy has been verified. Near global distribution in inshore tropical and temperate waters, except for central and eastern Pacific. Occur close inshore to oceanic, generally littoral or epipelagic, to ~230 m, but reported to 500 m. Common to rare, they have proven to be largely innocuous, are popular in public aquaria, and have become the subject of dive ecotourism. Monotypic genus *Carcharias* Rafinesque 1810.

# Carcharias taurus Rafinesque 1810

#### Raggedtooth shark

PLATE 22

*Carcharias taurus* Rafinesque 1810: 10, Pl. 14, Fig. 1 (Sicily, Mediterranean Sea); Barnard 1925, 1927\*; Smith 1949\*; D'Aubrey 1964\*, 1969; Forster *et al.* 1970; Compagno & Follett 1986; Compagno *et al.* 1989\*; Cliff & Wilson 1994\*; Randall 1995\*; Pradervand 2000; Compagno 2001\*; Smale 2002; Al-Shuaily & Henderson 2003\*; Bonfil & Abdallah 2004\*; Heemstra & Heemstra 2004\*; Compagno *et al.* 2005; Dicken *et al.* 2007; Moore *et al.* 2007\*; Ebert *et al.* 2013\*; Weigmann 2016; Stone & Shimada 2019\*.

*Odontaspis americanus*: Gilchrist 1902; Thompson 1914. *Carcharias tricuspidatus*: Smith 1949\* [in part]; Compagno 1984\* [in part]. *Odontaspis taurus*: Bass *et al.* 1975\*; Buxton *et al.* 1984; Wallace *et al.* 1984. *Eugomphodus taurus*: Compagno 1984\*, 1986\*; SSF No. 19.1\*.



Diagnosis as for family.

Body pale brown to grey, often darker brown or bronzy dorsally, with large dark blotches spread over body; abruptly white ventrally. Attains at least 3.2 m TL, possibly 4.3 m TL.

**DISTRIBUTION** Warm-temperate and tropical coastal waters of Atlantic, Indo-Pacific and western Mediterranean Sea. WIO: northern India, Pakistan, Gulf of Oman, Red Sea, Tanzania (Zanzibar), Mozambique and South Africa (KwaZulu-Natal to southeastern Atlantic).

**REMARKS** Some literature cites Pl. 44 in Rafinesque's original description as an illustration of this species; however, it is Pl. 14 that illustrates the species. Able to gulp air into its stomach and hover motionlessly, it is often found aggregating in so-called shark gutters, demonstrating philopatry for those sites, sometimes numbering dozens of individuals. More active at night, feeds on a wide range of fishes and invertebrates. Known to migrate seasonally in South Africa, Australia and United States, apparently between mating and pupping grounds. Historical persecution by spearfishers has resulted in dramatically reduced populations in eastern Australia, and to a lesser extent in South Africa; commercially fished throughout its range, particularly in Pakistan and India. Historically common in many parts of its range, and a familiar shark due to its popular use in public aquaria as well as supporting dive tourism operations globally. Not usually dangerous to people. IUCN Red List conservation status Vulnerable to Critically Endangered, depending on population, but the health of most populations is unknown. Given the declines reported in most areas where the population status of the species is known, as well as its philopatric habit, it can be assumed to be Near Threatened.

# FAMILY ODONTASPIDIDAE

#### Sandtiger sharks

Brett A Human

Large-sized (to at least 4.5 m TL) with stout cylindrical to compressed body and long head (HL ~25% TL); snout moderately elongate, slightly depressed and bluntly conical; 5 paired gill slits, large, lateral and not extending dorsally, with all slits before pectoral-fin origins. Eyes moderately large, 1.6–2.8% TL, set laterally on head over mouth without lateral head ridges underneath; no nictitating eyelid; spiracles minute. Mouth parabolic, subterminal, extending behind eyes, with lip folds. Teeth large, similar in both jaws, although identifiable as symphysials, anteriors, intermediates, laterals and posteriors; teeth erect with long cusps and 1–3 lateral cusplets either side of median cusp, becoming compressed posteriorly; tooth counts 34–56/36–46. Nasal flaps small, without nasal barbels; no oronasal grooves. Fins angular and weakly falcate or rounded. Two spineless dorsal fins; 2nd dorsal fin smaller than 1st. Pectoral fins medium-sized and broad; pectoral-fin origin behind 5th gill slit. Pelvic fin smaller than 1st dorsal fin. Anal fin subequal to 2nd dorsal fin; anal-fin base slightly posterior to 2nd dorsal-fin base. Peduncle compressed, without lateral keels, and with upper crescentic precaudal pit, but no lower pit. Caudal fin heterocercal, its dorsal margin <½ precaudal length.

Reproductive mode presumed yolk-sac viviparous with oophagy. Patchily distributed throughout tropical and temperate oceans. Occur close inshore to oceanic, generally littoral or epipelagic, but known to 1 600 m deep. Common to rare. One genus and 2 species, both in WIO.

## GENUS Odontaspis Agassiz 1838

Snout relatively long, preorbital length 0.8–1.2 times mouth width. Fins angular and weakly falcate or rounded; 1st dorsal fin larger than 2nd dorsal fin, pelvic fins and anal fin; 1st dorsal-fin origin closer to pectoral fins than to pelvic fins. Teeth erect, with elongated median cusp and 1–3 much shorter cusplets on either side. Scattered distribution in tropical and temperate waters. Two species, both in WIO.

#### **KEY TO SPECIES**

464 COASTAL FISHES OF THE WESTERN INDIAN OCEAN | VOLUME 1



# Odontaspis ferox (Risso 1810)

Smalltooth sandtiger

PLATE 23

*Squalus ferox* Risso 1810: 38 (Nice, France, Mediterranean Sea). *Odontaspis herbsti*: Bass *et al.* 1975\*.

Odontaspis ferox: D'Aubrey 1969\*; Bauchot & Bianchi 1984\*; Compagno 1984\*, 2001\*; Gubanov 1985; SSF No. 19.2\*; Compagno *et al.* 1989\*; Randall & Anderson 1993; Marine Research Section [Maldives] 1997\*; Winterbottom & Anderson 1997; Adam *et al.* 1998\*; Morón *et al.* 1998; Heemstra & Heemstra 2004; Compagno *et al.* 2005\*; Ebert 2013\*; Ebert *et al.* 2013\*; Stone & Shimada 2019.

Diagnosis as for genus. Teeth erect, with elongated median cusp and 2 or 3 pairs of lateral cusps; 2 rows of symphysial teeth in upper and lower jaws, and 2–5 rows of intermediate teeth in upper jaw; gill filaments not protruding from gill slits; tooth count 46–56/36-46. Vertebrae 177–183; spiral valve turns 32–34.

Body grey or grey-brown dorsally, paler ventrally; variably with or without large darker spots; no pale blotch on 1st dorsal fin; extreme piebald colour variant recorded in Mediterranean Sea. Attains ~4.5 m TL.

**DISTRIBUTION** Probably circumglobal, fragmented populations, in warm-temperate to tropical deep water of open ocean, including Mediterranean Sea. WIO: Tanzania (Pemba I.), South Africa (KwaZulu-Natal and possibly Transkei region), Melville Bank, southern Madagascar, Chagos, Maldives and Sri Lanka. **REMARKS** Uncommon to rare, and rarely encountered. Mostly known from deep inshore coastal waters, on or near bottom and near reef gullies, but possibly epipelagic. Feeds on small bony fishes, shrimp and squid. Incidentally caught in commercial fisheries by gillnets, longlines and trawl gear, particularly in Mediterranean Sea, Japan and Australia. The subject of ecotourism dives off Colombia (Malpelo I.) and Lebanon; has been exhibited in public aquaria with limited success. IUCN Red List conservation status Vulnerable, but listed as possibly Endangered in Mediterranean Sea, and protected in parts of Australia.

# Odontaspis noronhai (Maul 1955)

Bigeye sandtiger

PLATE 23

*Carcharias noronhai* Maul 1955: 3, Figs. 1–4 (Madeira, North Atlantic). *Odontaspis noronhai*: Compagno 1984\*, 2001\*; Morón *et al.* 1998; Compagno *et al.* 2005\*; Ebert 2013; Ebert *et al.* 2013\*; Stone & Shimada 2019.

Diagnosis as for genus. Large eyes. Teeth erect with elongated central cusp and pair of lateral cusps; 1 or no rows of symphysial teeth in upper jaw, and 2–4 rows of symphysial teeth in lower jaw; 1 or 2 rows of intermediate teeth in upper jaw; tooth count 34–43/37–46. Spiral valve turns 30.

Body dark reddish brown, sometimes greyish or purplish to black, without darker spots; sometimes with white blotch on 1st dorsal-fin apex; bright red gill filaments may protrude from 1st gill slit. Attains ~4.2 m TL.



Odontaspis noronhai © Food and Agriculture Organization of the United Nations, Original Scientific Illustrations Archive; reproduced with permission

**DISTRIBUTION** Possibly circumglobal in deep warm waters, but with disjunct populations: scattered records from Atlantic (Brazil and Mexico), central Pacific (including Hawaii) and Indian oceans. WIO: India, Sri Lanka and possibly Seychelles.

**REMARKS** Rare and poorly known from very few records. Oceanic; primarily mesopelagic, but also epipelagic to demersal. May undergo diurnal vertical migrations, inhabiting deep water by day and near surface at night; some evidence of seasonality in Brazilian waters, indicating the species may also be migratory. Very rare bycatch of oceanic longline fisheries. IUCN Red List conservation status Data Deficient, although a prohibited catch in the United States.

# FAMILY MITSUKURINIDAE

# Goblin shark

David A Ebert

Body soft, somewhat flabby, and slender, with unmistakable snout flattened as long blade-like rostrum; head nearly as long as body. Eyes small, spiracles very small; 5 paired gill slits, broad, with exposed filaments. Mouth large, broadly arched, with highly protrusible jaws; anterior teeth very long and slender with single cusp; tooth count 35–53/31–62. Dermal denticles small, with erect narrow cusps, making skin rough. Two dorsal fins, small, rounded, nearly equal in size, and smaller than pelvic fins and anal fin; pectoral fins small and broad; caudal-fin dorsal margin elongate, but <½ body length, and no lower lobe.

Monotypic.

## Mitsukurina owstoni Jordan 1898

#### Goblin shark

Mitsukurina owstoni Jordan 1898: 200, Pls. 11–12 (Sagami Sea, Japan); Bass, D'Aubrey & Kistnasamy 1975\*; Piotrovsky & Prut'ko 1980; SSF No. 17.1\*; Compagno *et al.* 1989\*; Compagno 2001\*; Compagno *et al.* 2005\*; Yano *et al.* 2007; Last & Stevens 2009\*; Ebert 2013\*; Ebert *et al.* 2013\*.

Diagnosis as for family. Vertebrae 122–125; spiral valve turns 18–23.

Colour in life spectacular pinkish white, with bluish fins; preserved specimens uniformly grey or brown. Attains at least 5.5 m TL, possibly 6.2 m TL.

**DISTRIBUTION** Circumglobal in temperate to tropical seas (patchy), with records from western and eastern Atlantic, western Pacific (Taiwan, Japan, Australia, New Zealand), eastern Pacific (southern California), and Indian Ocean. WIO: South Africa (Western Cape to KwaZulu-Natal) and Mozambique seamount range.

**REMARKS** Very poorly known; occurs over outer continental shelf and upper slope, to at least 1 300 m deep, and off seamounts. Its soft body and the prey items it consumes suggest it is poor-swimming and mesopelagic. Reproduction presumed oophagous; gestation period and litter size unknown; size at birth 80–90 cm TL. Males mature at ~265 cm TL, and females at >400 cm TL. Diet includes small pelagic fishes, pelagic crabs and cephalopods. Infrequent bycatch of deepwater fisheries, the jaws are sought by collectors.



PLATE 24

PLATE 24

# FAMILY **PSEUDOCARCHARIIDAE**

# Crocodile shark

Brett A Human

Small-sized (diminutive among lamniforms), with slender, cylindrical body and long head (HL ~1/3 TL); snout long and broadly conical. Nostrils small, with small nasal flaps. Five paired gill slits, broad and lateral on body, extending onto dorsal surface of head, with all gill slits anterior to pectoralfin origin. Eyes huge (diameter 3.6-4.9% TL), lateral on head, above front of mouth; no nictitans; spiracles very small, level with eyes and approximately over angle of mouth. Mouth large, subterminal, parabolic and extending well behind eyes, with highly protrusible jaws. Teeth large, monocuspidate, and similar in both jaws, although identifiable as anteriors, intermediates and laterals; anterior teeth elongate, becoming short and compressed posteriorly; teeth sexually dimorphic: females with longer and broader teeth. Fins small, low and broadly angular. Two spineless dorsal fins; 1st dorsal fin low, larger than 2nd, and about equally spaced between pectoral and pelvic fins. Pectoral fins much shorter than HL, origin behind 5th gill slit. Pelvic fins smaller than pectoral fins; pelvic-fin bases before 2nd dorsal-fin origin. Anal fin minute, smaller than 2nd dorsal fin; anal-fin origin opposite 2nd dorsal-fin insertion. Peduncle short, cylindrical to slightly depressed, with poorly developed lateral keel, and weak upper and lower precaudal pits (but upper pit better developed). Caudal fin heterocercal; upper lobe long, but dorsal margin <<sup>1</sup>/<sub>3</sub> TL. Monotypic genus Pseudocarcharias Cadenat 1963.

## Pseudocarcharias kamoharai (Matsubara 1936)

#### Crocodile shark

*Carcharias kamoharai* Matsubara 1936: 380 (Kôti fish market, Japan); D'Aubrey 1964\*.

Odontaspis kamoharai: Bass et al. 1975\*.

Pseudocarcharias kamoharai: Compagno 1984\*, 2001\*; SSF No. 18.1\*; Compagno et al. 1989\*; Romanov & Samorov 1994\*; Anderson et al. 1998; Morón et al. 1998; Compagno et al. 2005\*.

Diagnosis as for family. Tooth count 22–29/21–26; vertebrae 146–158; spiral valve turns 24–27.

Body dark grey, brown or lilac above, paler below, and snout sometimes greyish underneath; fin margins white and translucent; white blotch between mouth and gill slits in some Pacific specimens. Attains 120 cm TL.

**DISTRIBUTION** Circumtropical to subtropical in all seas, although patchily recorded. WIO: South Africa, Mozambique, Madagascar, Seychelles, Chagos, Maldives and Sri Lanka; not known from Red Sea and Persian/Arabian Gulf.

**REMARKS** D'Aubrey (1964) compared a specimen captured off Cape Town, South Africa, to all other known synonyms for this species, concluding that only a single species exists worldwide. Biology poorly known; its morphology suggests a strong-swimming, active, predatory shark. Reproduction mode yolk-sac viviparity and oophagy, potentially adelphophagy given low litter size of 2–4. Diet includes small pelagic fishes, shrimp and squid. Uncommon (except previously locally common in Mozambique Channel); predominately oceanic, epipelagic and possibly mesopelagic, seldom found close inshore; recorded to 590 m deep. A bycatch of pelagic longlines.



Pseudocarcharias kamoharai, 81 cm TL, mature female. Source: Bass et al. 1975

# FAMILY **MEGACHASMIDAE**

# Megamouth shark

Brett A Human

Gigantic filter-feeding shark with cylindrical, relatively flabby body tapering towards caudal fin, and massive head (HL ~1/3 TL). Snout short and broadly rounded, giving head bulbous appearance; nostrils small, with small nasal flaps; 5 paired gill slits, moderately large and lateral on body; gill rakers finger-like, cylindrical, on internal gill arches. Eyes moderately large, diameter <1/3 length of largest gill slit, set laterally on head, above and before corners of mouth; no nictitans; spiracles minute. Mouth huge, terminal, broadly arched, and extending behind eyes. Teeth small, numerous, hook-shaped, unicuspid, similar in both jaws, with >100 rows in each jaw, but with toothless gap at symphyses. Pectoral fins long and narrow, but shorter than HL, origin under 4th gill slit. Other fins small: 2 spineless, triangular dorsal fins; 1st dorsal fin nearly twice size of 2nd, and closer to pectoral fins than to pelvic fins; pelvic fins smaller than 1st dorsal fin; anal fin behind and smaller than 2nd dorsal fin. Peduncle short, compressed, with upper precaudal pit and no ridges. Caudal fin hypercercal: upper lobe very long, <1/2 body length, and lower lobe short but well-developed.

Monotypic.

# Megachasma pelagios Taylor, Compagno &

Struhsaker 1983

#### Megamouth shark

PLATE 24

Megachasma pelagios Taylor, Compagno & Struhsaker 1983: 96, Figs. 2–5 (off Oahu I., Hawaii); Compagno 1990\*, 2001\*; Nelson *et al.* 1997\*; Smale *et al.* 2002\*; Ebert 2003\*; Compagno *et al.* 2005\*; Last & Stevens 2009\*; Ebert *et al.* 2013\*.

Diagnosis as for family. Vertebrae 139–151; spiral valve turns 23 or 24.

Body grey to dark grey, paler below; lower jaw often with numerous large dark spots, and luminous bar visible above upper lip when jaw extended; pale margins on tips and posterior edges of pectoral fins and pelvic fins. Attains 5.5+ m TL.



Megachasma pelagios, 190 cm TL, juvenile (Atlantic Ocean).

**DISTRIBUTION** All tropical and some temperate seas, but few records, mostly from the Pacific (Taiwan, Philippines, Japan and Hawaii). WIO: one specimen found washed ashore in South Africa (Nature's Valley, Western Cape).

**REMARKS** Reproduction assumed yolk-sac viviparity, possibly with intrauterine cannibalism. One of three species of filter-feeding sharks; jaw morphology suggests it uses gulping action to feed on plankton, mainly euphausiids, copepods and jellyfish; a band of luminescent tissue on the upper lip may attract prey. Oceanic and coastal over very deep water; primarily tropical but may stray into temperate water following warm ocean currents. Epipelagic, with evidence that it migrates vertically in diurnal cycle following migrations of euphausiid shrimps (krill), which migrate from 5-40 m at night to ~600 m during the day. Specimens often found scarred by attacks from cookiecutter shark Isistius brasiliensis. Known since 1976, rarely seen and presumed rare throughout its (possibly fragmented) range; an extremely rare incidental bycatch of various coastal and high-seas fisheries, and some specimens found washed ashore.

# FAMILY ALOPIIDAE

Thresher sharks Brett A Human

Large- to very large-sized (to ~6 m TL) with stout, cylindrical body; HL <<sup>1</sup>/<sub>8</sub> TL; snout moderately long and conical; 5 paired gill slits, short and lateral, not extending onto dorsal surface of head, with 4th and 5th gill slits over pectoral fins. Eyes large, 1.8–4.3% TL, lateral to dorsolateral and above mouth; no nictitans; spiracles minute, level with eyes, and posterior to eyes by about twice snout length. Nostrils small, with small nasal flaps and no oronasal groove. Mouth small, subterminal, parabolic to broadly arching, extending behind eyes; and lip folds present or absent. Teeth small to moderate, similar in both jaws, although identifiable as symphysials (absent in *Alopias superciliosus*), anteriors, intermediates and

PLATE 25

laterals; unicuspid, erect, slender and elongated to oblique and compressed, with small secondary cusp posterior to primary cusp; tooth counts 19–52/20–50. Fins large or small and falcate. Two spineless dorsal fins; 1st dorsal fin high and falcate, 2nd dorsal fin minute. Pectoral fins large, long and narrow, broadly to narrowly pointed. Pelvic fins moderately large, falcate, approximately size of 1st dorsal fin. Anal fin low, minute, equivalent in size to 2nd dorsal fin; anal-fin base behind 2nd dorsal-fin base. Peduncle short, slightly compressed, without lateral keels, and with crescentic upper and lower precaudal pits. Caudal fin strongly heterocercal; upper lobe extremely long, equal to body length; lower lobe moderately developed.

Wide-ranging in tropical to cold-temperate seas. Reproductive mode yolk-sac viviparity with oophagy. All are active swimmers and popular game fish. Use their enormous tails to corral and stun smaller fish for feeding, behaviour that is confirmed by the high numbers of thresher sharks caught by their tails in longline fisheries due to the sharks whipping the live bait on the hooks. One genus, *Alopias* Rafinesque 1810, with 3 species, all in WIO.

#### **KEY TO SPECIES**



## Alopias pelagicus Nakamura 1935

Pelagic thresher

*Alopias pelagicus* Nakamura 1935: 2, Pl. 1, Fig. 1 (Taiwan); Gubanov 1972; Bass *et al.* 1975\*; Compagno 1984\*, 1986\*, 2001\*; SSF No. 16.1\*; Compagno *et al.* 1989\*; Cliff & Wilson 1994\*; Randall 1995\*; Anderson *et al.* 1998; Morón *et al.* 1998; Bonfil & Abdallah 2004\*; Compagno *et al.* 2005\*; Ebert *et al.* 2013\*.

Pectoral fins nearly straight and narrowly rounded. Mouth parabolic, without lip folds; teeth oblique, with small secondary cusp posterior to primary cusp, and with symphysial teeth; tooth count 41–45/37–48. Vertebrae 453–477.

Body metallic pale blue to dark blue or grey, above and on fins, changing to white below level of pectoral fins. Attains 3.8 m TL.

**DISTRIBUTION** Tropical to temperate waters of Indo-Pacific. WIO: Gulf of Oman, Red Sea, Gulf of Aden, Arabian Sea, Kenya, Tanzania, South Africa (KwaZulu-Natal), Maldives and Sri Lanka.

**REMARKS** Historically confused with thresher shark *A. vulpinus* in the Indian Ocean, resulting in few reliable records of this species from the region. Oceanic, epipelagic, and apparently rare in inshore waters of Indian Ocean. Behaviour and biology poorly known, and virtually unknown in WIO. Reproduction in the Pacific is possibly aseasonal; gestation <1 year, with litters of 2; the Gulf of Aden may be used as pupping grounds. Taken as a bycatch in longline fisheries where it occurs. IUCN Red List conservation status Endangered globally.



Alopias pelagicus, 2.8 m TL, immature female (South Africa). Source: Bass et al. 1975



Alopias superciliosus, 1.6 m TL, immature female (South Africa). Source: Bass et al. 1975

#### Alopias superciliosus (Lowe 1841)

#### Bigeye thresher

PLATE 25

Alopecias superciliosus Lowe 1841: 39 (Madeira, North Atlantic).
Alopias superciliosus: Gubanov 1972, 1978; Bass et al. 1975\*; Gruber & Compagno 1981; Compagno 1984\*, 1986\*, 2001\*; SSF No. 16.2\*;
Compagno & Smale 1986; Compagno et al. 1989\*; Randall & Anderson 1993; Cliff & Wilson 1994\*; Marine Research Section [Maldives] 1997\*;
Morón et al. 1998; Bonfil & Abdallah 2004\*; Heemstra & Heemstra 2004; Compagno et al. 2005\*; Ebert et al. 2013\*.

Pectoral fins weakly falcate and somewhat pointed. Eyes huge, with deep nuchal groove above eye to over gill slits. Mouth broadly rounded, without lip folds. Teeth erect and unicuspid, slender and elongate (as compared with thresher shark *A. vulpinus*), without secondary cusps, and no symphysial teeth; tooth count 22–27/20–24. Vertebrae 219–319.

Body dark grey, varying from purplish to brownish dorsally and on fins, changing to white below level of pectoral-fin bases. Attains 4.8 m TL.

**DISTRIBUTION** Circumglobal in tropical to temperate seas, including western Mediterranean Sea. WIO: Red Sea, Gulf of Aden to Tanzania, South Africa (KwaZulu-Natal; Cape Peninsula), Madagascar, Maldives and Sri Lanka.

**REMARKS** Readily identifiable from other thresher sharks by the arrangement of the eyes which give this species dorsal binocular vision. Reproduction probably aseasonal, with litters of 2–4. Epipelagic and neritic, oceanic to close inshore, and more common in inshore waters than pelagic thresher *A. pelagicus*. Feeds largely on pelagic fishes, but also demersal fishes and squid. Caught as a bycatch primarily in longline fisheries, but also by gillnets and trawls. IUCN Red List conservation status Vulnerable globally.

## Alopias vulpinus (Bonnaterre 1788)

Thresher shark

PLATE 25

Squalus vulpinus Bonnaterre 1788: 9 (Mediterranean Sea). Alopecias vulpes: Bleeker 1878; Sauvage 1891; Gilchrist 1902. Alopias vulpes: Thompson 1914; Barnard 1925. Alopias pelagicus: D'Aubrey 1964\*.

Alopias vulpinus: Smith 1949\*; Gubanov 1972, 1978; Bass et al. 1975\*; Compagno 1984\*, 1986\*, 2001\*; SSF No. 16.3\*; Compagno et al. 1989\*; Compagno et al. 1991; Randall & Anderson 1993; Cliff & Wilson 1994\*; Marine Research Section [Maldives] 1997\*; Winterbottom & Anderson 1997; Bonfil & Abdallah 2004\*; Heemstra & Heemstra 2004\*; Compagno et al. 2005\*; Ebert et al. 2013\*.



Pectoral-fin tips acute; 1st dorsal-fin height greater than peduncle depth; caudal-fin upper lobe nearly equal to body length. Mouth parabolic, with lip folds. Teeth erect, unicuspid, relatively broad-based and compressed (as compared with bigeye thresher *A. superciliosus*), without secondary cusps, and with symphysial teeth; tooth count 32–52/25–50. Vertebrae 230–364.

Body dark grey or bluish dorsally and on fins, flanks sometimes silvery or coppery, and white below; white on at least part of gill region and over pectoral-fin bases; pectoralfin tips sometimes white dorsally. Attains at least 5.8 m TL, possibly 6.3 m TL.

**DISTRIBUTION** Nearly circumglobal in tropical to coldtemperate seas, including throughout Mediterranean Sea. WIO: Pakistan, Gulf of Oman, Red Sea, Gulf of Aden to Tanzania, South Africa (Western Cape to KwaZulu-Natal), Madagascar, Mauritius, Chagos, Maldives, India and Sri Lanka.

**REMARKS** Litters of 2–4 (occasionally 6); gestation period (in NE Pacific) 9 months. Wide-ranging, oceanic, epipelagic to neritic, although most common near land and in temperate waters. Behaviour poorly known, but populations in various oceans may be distinct with differing demographics; horizontal and vertical sexual segregation occurs in northern WIO, and seasonally migratory in NE Pacific. Diet includes small pelagic and schooling fishes, some demersal and benthic fishes, cephalopods, shrimp and pelagic crabs; uses its tail to corral and stun fish prey. A bycatch of pelagic longlines and gillnets; in WIO, commonly caught 350–800 km off Somalia and NE of Maldives. IUCN Red List conservation status Vulnerable globally, with several differing regional assessments.

# FAMILY **CETORHINIDAE**

#### Basking shark David A Ebert

Unmistakably huge, stout, fusiform body; head moderately long; snout large, pointed and conical; gill slits enormous, nearly encircling head. Eyes small; spiracles very small. Mouth huge, subterminal; teeth minute, with single smooth-edged hook-shaped cusps, and >200 tooth rows on upper and lower jaws. Dermal denticles large, with hook-shaped cusps, very rough to touch. First dorsal fin very large, triangular, high and erect, its origin behind pectoral-fin free rear tips; 2nd dorsal fin smaller, <½ height of 1st dorsal fin, its origin before anal fin. Pectoral fins very large and broad, but their length less than HL. Pelvic fins smaller, less than 1st dorsal-fin height, but larger than 2nd dorsal fin. Peduncle depressed, with strong lateral keels, and with upper and lower crescent-shaped precaudal pits. Caudal fin large, lunate. Monotypic genus *Cetorhinus* Blainville 1816.

## Cetorhinus maximus Gunnerus 1765

Basking shark

PLATE 26

*Cetorhinus maximus* Gunnerus 1765: 33, Pl. 2 (northern Norway); Bass *et al.* 1975\*; SSF No. 15.1\*; Compagno *et al.* 1989\*; Compagno *et al.* 2005\*; Natanson *et al.* 2008; Ebert *et al.* 2013\*.

Diagnosis as for family. Vertebrae 107–116; spiral valve turns 47–51.

Body colour dark and variable, often with mottling on flanks and back; white blotches under head. Attains >10 m TL.

**DISTRIBUTION** Circumglobal in all warm- to cold-temperate seas, known to make transoceanic migrations. WIO: South Africa (Western Cape to southern KwaZulu-Natal).



Cetorhinus maximus, ~7.5 m TL, adult female (South Africa). Source: SSF; dorsal view of head, and teeth (South Africa). Source: SFSA

**REMARKS** Oophagous, with litters of 1–6; size at birth ~1.5-2 m TL; gestation period unknown. Exceeded in size only by the whale shark (*Rhincodon typus*); males mature at ~4-5 m TL, and females at ~8-9 m TL. Juveniles >3 m TL rarely encountered, suggesting pupping and nursery grounds located in planktonic-rich oceanic waters far from populated coastal areas. Previous age estimates for this species are known to be erroneous as vertebral bands are associated with growth and not age. Oceanic, coastal and pelagic, will dive to several hundred metres and remain at depth as they migrate between hemispheres. Docile and often seen cruising slowly ('basking') at surface in cooler waters, these enormous filter-feeding sharks consume vast quantities of zooplankton, including copepods and planktonic larvae. When feeding, adults swim at ~2 knots and will pass ~2 000 tons of water over the gills per hour as they close their mouth every 30-60 seconds to ingest filtered plankton trapped in their gill rakers; an individual may have half a ton of food in its stomach at any time. Sometimes found swimming in large groups. A target fishery for this species has occurred in the northeastern Atlantic and North Pacific; occasionally indirectly caught in other fisheries. IUCN Red List conservation status Endangered globally, due to regional fishing pressure.

# FAMILY LAMNIDAE

Mackerel sharks Brett A Human and David A Ebert

Large- to very large-sized (to 6+ m TL) with slender to stout fusiform body, and conical head (HL 20-30% TL). Five paired gill slits, lateral on body and extending dorsally, with all gill slits anterior to pectoral-fin origins. Snout relatively long, bluntly to sharply conical. Eyes lateral or dorsolateral on head, over mouth; eye diameter 0.7-3.1% TL; no nictitans; spiracles minute or absent. Nasal flaps small, without nasal barbels or oronasal grooves. Mouth broadly arched, subterminal, extending behind eyes, with poorly developed lip folds. Teeth large to very large, similar in both jaws, although identifiable as anteriors, intermediates, laterals and posteriors; teeth varying from long and elongate with strong median cusp, with or without lateral cusplets, to compressed, triangular and bladelike with serrated margins; tooth counts 22-31/20-29. Fins triangular or rounded to falcate. Two spineless dorsal fins; 1st dorsal fin high, much larger than 2nd, its origin over pectoralfin bases or rear margins. Pectoral fins long and narrow; pectoral-fin origin behind 5th gill slit. Pelvic fins much smaller than pectoral fins. Anal fin tiny, equivalent in size to 2nd dorsal fin; anal-fin base under or slightly posterior to 2nd dorsal-fin base. Peduncle moderately long and strongly depressed, with 1 or 2 well-developed lateral keels, and with upper and lower crescentic precaudal pits. Caudal fin lunate. Vertebrae 153–197; spiral valve turns 38–55.

Reproductive mode yolk-sac viviparity, with oophagy. All lamnids are warm-blooded and maintain a body temperature higher than the ambient water temperature, facilitating their highly active and predatory habits, with the family containing some of the swiftest fishes known. Found close inshore to oceanic, generally littoral or epipelagic, but known to ~1 280 m deep. Lamnid species are either common, uncommon or endangered, with some particularly susceptible to high-seas longline fisheries; all are regarded as game fish. Distributed throughout all tropical, temperate and subpolar oceans (absent only from icepack regions in polar oceans).

Three genera and 5 species; all genera and 4 species in WIO.

#### **KEY TO GENERA**



# GENUS Carcharodon Smith 1838

Body cylindrical and moderately stout; peduncle with single well-developed lateral keel; eye diameter 0.7–1.8% TL. Teeth triangular, compressed, broad, and with serrations. One species.

## Carcharodon carcharias (Linnaeus 1758)

#### White shark

PLATE 26

*Squalus carcharias* Linnaeus 1758: 235 (western and northeastern Atlantic, and Mediterranean Sea).

*Carcharodon capensis* Smith 1849: no page no., Pl. 4 (Cape Seas, South Africa).

Carcharodon rondeletii: Gilchrist 1902; Thompson 1914.

*Carcharodon carcharias*: Barnard 1925; Fowler 1925; Smith 1949\*; Farquhar 1963; D'Aubrey 1964\*; SFSA No. 26\*; Bass *et al.* 1975\*; Compagno 1984\*, 1986\*, 2001\*; SSF No. 14.1\*; Compagno *et al.* 1989\*; Cliff & Wilson 1994\*; Smale & Heemstra 1997\*; Cliff *et al.* 2000; Zuffa *et al.* 2002; Heemstra & Heemstra 2004\*; Compagno *et al.* 2005\*; Ebert *et al.* 2013\*.

Diagnosis as for genus. First dorsal-fin origin over pectoral-fin inner margins; 2nd dorsal fin minute. Tooth count 20–28/20–28; vertebrae 170–187.

Body grey to dark blue-grey, sometimes brownish, blackish or bronzy dorsally, and abruptly white ventrally; eyes very black; pectoral fins black-tipped underneath, with or without black axillary spot on body near rear edge of pectoral fins; albinism recorded. Attains 6.4 m TL, but most adults <6 m TL.

**DISTRIBUTION** Worldwide in most temperate and tropical seas. WIO: Kenya to South Africa, Madagascar, Seychelles, Réunion, Mauritius, southern India and Sri Lanka, and possibly Red Sea; records from Arabian Sea and Persian/Arabian Gulf need confirmation.

**REMARKS** One of the greatest known distributions for a shark; this wide geographic and habitat distribution has given rise to multiple synonyms. Generally uncommon but locally common in some areas; principally pelagic, but known

from surfline to >1 250 m deep. Individuals are known to make transoceanic movements: in the Indian Ocean, tagged white sharks have travelled from South Africa to Australia and back within a span of months. Litters of 2-13; size at birth 110–160 cm TL; gestation period assumed >1 year. Feeds on a variety of bony fishes and elasmobranchs, other vertebrates (seabirds, pinnipeds and cetaceans), cephalopods and crustaceans. Larger individuals are a spectacular ambush predator of marine mammals, patrolling close to bottom and then launching upwards to strike prev close to surface, sometimes becoming airborne in the process. Relentlessly sensationalised in the media, particularly regarding infrequent but dangerous interactions with humans; paradoxically now the subject of dive tourism. IUCN Red List conservation status Vulnerable; a protected species in South Africa and in other countries outside the region due to historical persecution and its highly valued jaws and teeth.

## GENUS Isurus Rafinesque 1810

Body slender to robust, fusiform, and with single welldeveloped lateral keel on peduncle; eye diameter 1–3% TL. Teeth long, slender, and somewhat protrude from mouth; anterior teeth enlarged, with single, large, robust, smoothedged cusp; upper and lower teeth similarly shaped. Two species, both in WIO.

#### **KEY TO SPECIES**



Carcharodon carcharias, jaws of a large specimen (South Africa).



Carcharodon carcharias, 177 cm TL, immature female (South Africa). Source: Bass et al. 1975



Isurus oxyrinchus, 250 cm TL, female; ventral view, ~100 cm TL, immature female (both South Africa). Source: SSF (lateral view); Bass et al. 1975 (ventral view)

#### Isurus oxyrinchus Rafinesque 1810

#### Shortfin mako

PLATE 26

*Isurus oxyrinchus* Rafinesque 1810: 12, Pl. 13, Fig. 1 (Sicily, Mediterranean Sea); Smith 1957\*; Bass *et al.* 1975\*; Compagno 1984\*, 1986\*, 2001\*; SSF No. 14.2\*; Compagno *et al.* 1989\*; Cliff *et al.* 1990; Anderson & Ahmed 1993\*; Randall & Anderson 1993; Cliff & Wilson 1994\*; Randall 1995\*; Marine Research Section [Maldives] 1997; Winterbottom & Anderson 1997; Ebert 2001, 2003\*; Al-Shuaily & Henderson 2003; Bonfil & Abdallah 2004; Heemstra & Heemstra 2004\*; Compagno *et al.* 2005\*; Last & Stevens 2009\*; Ebert *et al.* 2013\*.

Lamna glauca: Gilchrist 1902; Thompson 1914.

Isurus glauca: Barnard 1925.

*Isurus glaucus*: Smith 1949\*, 1957\*; D'Aubrey 1964\*. *Isurus tigris africanus*: Smith 1957\*.

Body fusiform and relatively robust; snout long, acutely pointed; eye diameter <33% snout length. Teeth with single, large, smooth-edged cusps, similar in both jaws. First dorsalfin origin behind pectoral-fin free rear tips; pectoral fins shorter than HL; anal-fin origin under midbase of 2nd dorsal fin; caudal fin crescent-shaped. Peduncle with strong keel. Tooth count 24–26/22–32; vertebrae 183–196; spiral valve turns 47–48.

Body brilliant blue or purplish dorsally, becoming paler blue along flanks and white ventrally. Attains  $\sim$ 4 m TL, but most adults <3 m TL.

**DISTRIBUTION** Widespread in most tropical to warmtemperate seas, including Mediterranean Sea, Red Sea and coastal and oceanic waters throughout WIO.

**REMARKS** Litters of 4–25; size at birth 60–70 cm TL. Males mature at ~195 cm TL, at ~4 years; females mature at 270-300 cm TL, at 7-8 years. Reproductive cycle may be up to 3 years, including 15-18 month gestation period followed by 18 month resting stage; juveniles grow rapidly during first 2 years, nearly doubling their length. Coastal and oceanic, from surface to at least 500 m deep. Possibly the fastest swimming shark, known for its spectacular jumping ability when feeding on schools of fish or hooked by an angler. Feeds primarily on fishes, including other sharks, rays, mackerel, vellowtail, sea bass and swordfish, as well as squid; larger individuals will feed on sea turtles and small cetaceans. Has been implicated in attacks on people, particularly when provoked. The subject of directed fisheries worldwide and a popular big-game fish; highly valued for its meat and fins. IUCN Red List conservation status of most populations Endangered; Critically Endangered in Mediterranean Sea.

#### Isurus paucus Guitart Manday 1966

#### Longfin mako

PLATE 27

*Isurus paucus* Guitart Manday 1966: 3, Figs. 1–3 (Cuba, western Atlantic); Compagno 1984\*, 2001\*; SSF No. 14.3\*; Ebert 2001\*, 2003\*; Compagno *et al.* 2005\*; Last & Stevens 2009\*; Ebert *et al.* 2013\*.

Body fusiform and relatively slender; snout narrow to bluntly pointed; eye diameter >33% snout length. Teeth with single, large, robust, smooth-edged cusp; shape similar in both jaws. Pectoral-fin length subequal to HL. Anal-fin origin slightly behind 2nd dorsal-fin origin. Peduncle with long, strong keel. Tooth count 24–26/22–24; vertebrae 195–197; spiral valve turns 54.

PLATE 27

Body blue above and below, except for white midventral undersurface; underside of snout distinctly dusky, except in small juveniles. Attains 4.3 m TL.



Isurus paucus, 123 cm TL, immature male. Drawn from Garrick 1967

**DISTRIBUTION** Circumglobal in tropical to warm-temperate seas, but poorly recorded; commonly western Atlantic and central Pacific. WIO: Mozambique Channel, Madagascar and possibly off South Africa, but probably occurs throughout the region.

**REMARKS** Poorly known and infrequently encountered species, often misidentified as the better-known shortfin mako *I. oxyrinchus*. Litters of 2–8; size at birth 92–97 cm TL. Males mature at ~190 cm TL, and females at ~245 cm TL. Probably epipelagic, off continental shelf and over deep water in open ocean, but depth range unknown. Feeds primarily on schooling fishes and pelagic cephalopods. Despite its large size, has never been implicated in attacks on people. Of little commercial value as the flesh is soft and of poor quality.

#### GENUS Lamna Cuvier 1816

Body stout and fusiform, with 2 well-developed lateral keels on peduncle. Eyes 1.6–2.8% TL. Teeth elongate, with single small lateral cusps either side of median cusp. Global, in temperate to subpolar seas. Two species, 1 in WIO.

#### Lamna nasus (Bonnaterre 1788)

#### Porbeagle

*Squalus nasus* Bonnaterre 1788: 10, Pl. 85, Fig. 350 (Cornwall, England). *Lamna nasus*: Smith 1949\*; Farquhar 1963; Nakaya 1971\*; Bass *et al.* 1975\*; Compagno 1984\*, 1986\*, 1990\*, 2001\*; SSF No. 14.4\*; Compagno *et al.* 1989\*; Compagno *et al.* 2005\*; Last & Stevens 2009\*; Ebert *et al.* 2013\*.

Diagnosis as for genus. First dorsal-fin origin over pectoral-fin bases or inner margins. Tooth count 28–31/26–29; vertebrae 150–162.

Body grey to dark grey, sometimes bluish or blackish, abruptly changing to white ventrally, but no white patch above pectoral fins; 1st dorsal fin with distinctive white free rear tip; underside of head dusky in adults; abdomen blotched in some Southern Hemisphere adults as compared with no blotching and with white underside of head in Northern Hemisphere specimens. Attains 3.5 m TL.

**DISTRIBUTION** Antitropical and amphitemperate. In Northern Hemisphere: mostly North Atlantic and Mediterranean Sea, and absent from North Pacific. In Southern Hemisphere: most commonly found south of the region covered here and apparently only strays into the region. WIO: South Africa (south coast) and eastwards, but perhaps circumglobal in southern temperate seas.

**REMARKS** Swift and active shark, although its behaviour is poorly known. Coastal and oceanic on continental shelf, in littoral and epipelagic habitats, but recorded to 700 m deep. Migratory, moving inshore to surface waters in summer and offshore to deeper waters in winter. Mediterranean Sea appears to be a nursery ground. Feeds primarily on small pelagic and demersal bony fishes, but also sharks and squid. Heavily caught in both hemispheres as secondary target or in directed fisheries, especially by longline. Not dangerous to people. IUCN Red List conservation status Vulnerable globally, due to dramatic population declines.



Lamna nasus, 94 cm TL, immature male (South Africa); ventral head; teeth from larger specimen. Source: FSO (lateral view); Bigelow & Schroeder 1948 (head); SSF (teeth)

# ORDER CARCHARHINIFORMES

# FAMILY **PENTANCHIDAE**

# Deepwater catsharks

Brett A Human and David A Ebert

Generally small-sized, long and slender (most WIO species <69 cm TL), but a few are relatively stocky. Head length typically 20% TL; 5 pairs of gill slits, small to greatly enlarged, the last gill slit over or behind pectoral-fin origins. Snout broadly to acutely rounded, not depressed to strongly depressed and laterally expanded compared to rest of head, and tip upturned or not. Anterior nasal flaps not expanded, or short or extending to mouth, and separate or fused; nasal barbels absent; oronasal grooves present or absent. Eyes slit-like and dorsolateral, with rudimentary nictitans and strong subocular ridges, but no supraorbital crest; spiracles moderately large, set just behind eyes. Mouth inferior, long and arched, and with or without lip folds. Teeth multicuspidate (except monocuspidate in adult males of one species of Apristurus not known from WIO) and in multiple functional rows in both jaws. Fins rounded or weakly falcate. Two spineless dorsal fins (except one dorsal fin in Pentanchus from western Pacific), usually subequal; 1st dorsal fin above or behind pelvic-fin bases. Pectoral fins moderately sized and paddle-shaped. Pelvic fins moderately long and low. Anal fin long and low to high, its base usually longer than 2nd dorsal-fin base. Peduncle and caudal fin slender; peduncle without pits or keels; caudal fin heterocercal, lower lobe poorly developed. Sexual dimorphism of TL absent to extreme (with males attaining greater lengths than females) and also apparent in dentition of some species. Tooth counts 28-130/28-111; vertebrae 85-183; spiral valve turns 5-23.

Reproductive mode mostly oviparous, with females depositing egg cases on the substrate where the embryos develop and eventually hatch out. However, for some species the egg cases are retained *in utero* until the embryos are nearly completely developed. A few species are viviparous with a yolk sac whereby the embryos fully develop internally prior to the mother giving live birth to fully developed, free-swimming neonates. Deepwater catsharks are found worldwide in tropical to cold-temperate seas, with preferred temperatures likely contributing to a high degree of endemism. Most shallowwater species occur in tropical to warm-temperate regions and tend to be brightly coloured, whereas deep-sea forms tend to be more uniformly coloured. Sluggish, poor swimmers, and generally deep-dwelling on or near bottom, although some deep-sea species are known to migrate far off the bottom into the water column; in the WIO they are found from intertidal zone to deep-sea (>1 500 m), over soft bottom, on rocky or coral reefs and in kelp beds, and are rarely locally abundant. Some species may have become recently extirpated from parts of the WIO. Harmless to people. Generally not targeted in fisheries but often comprise a bycatch where abundant; some of the more colourful species are displayed and bred in aquariums.

A large family of diverse sharks, with 11 genera and ~111 species; 5 genera (2 in >200 m) and 16 species in WIO in <200 m, with several additional deep-sea species awaiting formal description. The genera *Apristurus* Garman 1913 and *Bythaelurus* Compagno 1988 are both deep-sea groups, and the former is included here only in the key. Previous records of a species of *Parmaturus* Garman 1906 from the southwestern Indian Ocean are now known to be a *Bythaelurus* species (see Ebert [2013] for more information on the deep-sea genera).

#### **KEY TO GENERA**



Continued ...

#### **KEY TO GENERA**



2b Body flaccid; snout strongly depressed and laterally expanded as compared to rest of head ...... Apristurus [deep-sea group]



- 3a Nasal flaps extend to mouth, fused ...... Haploblepharus



# GENUS Bythaelurus Compagno 1988

Body usually slender; head moderately depressed. Eyes not noticeably elevated. Snout broadly rounded and not coming to a point, not strongly depressed, and tip not upturned. Upper and lower lip folds short. Nasal flaps weakly expanded, separate and not fused; no nasal barbels. Fins more or less rounded. Dorsal and anal fins more or less subequal. Pelvic fins more or less fused but never forming pelvic curtain; claspers stout to elongate, outer length 5–6 times width. No crest of enlarged denticles along upper peduncle or dorsal caudal margin. Stomach not inflatable. Tooth counts 62–99/63–101; vertebrae 117–142; spiral valve turns 7–10. Presumed size range 29–76 cm TL.

Genus was separated from *Halaelurus*. Patchily distributed in all oceans; recorded from 220–1 262 m. At least 12 species, with 9 species in WIO, but all recorded at depths of >200 m. *Bythaelurus lutarius* is included here nonetheless as it is likely to be trawled from shallower depths in the future. The other deepwater WIO species are: *B. alcockii* (Garman 1913) (resembling *B. hispidus*, but only tentatively placed in this genus), *B. bachi* Weigmann *et al.* 2016, *B. clevai* (Séret 1987) (Plate 27), *B. hispidus* (Alcock 1891), *B. naylori* Ebert & Clerkin 2015 (Plate 28), *B. stewarti* Weigmann *et al.* 2018, *B. tenuicephalus* Kaschner *et al.* 2015, and *B. vivaldii* Weigmann & Kaschner 2017.

# Bythaelurus lutarius (Springer & D'Aubrey 1972)

#### Mud catshark

Halaelurus lutarius Springer & D'Aubrey 1972: 6, Fig. 1 (off Maputo Bay, Mozambique); Bass 1973; Bass *et al.* 1975\*; Springer 1979\*; Compagno 1984\*, 1988; SSF No. 11.6\*; Compagno *et al.* 1989\*; Howe & Springer 1993; Sommer *et al.* 1996\*.

Bythaelurus lutarius: Compagno 1999; Compagno et al. 2005\*.

Anal-fin base twice length of 2nd dorsal-fin base; prenarial length equal to internarial distance. Tooth count 74/86; vertebrae 132; spiral valve turns 10.

Body uniformly drab grey, but sometimes with dusky saddles beneath dorsal-fin bases, and slightly paler ventrally. Attains 39 cm TL.

**DISTRIBUTION** WIO: Mozambique (patchy), Somalia, Socotra and Oman.

**REMARKS** Reportedly viviparous with yolk-sac placenta; litters of 2; size at birth 10–14 cm TL. Abundant where caught in Oman; recorded from 220–813 m, on continental slope, over mud bottom. Feeds on cephalopods, crustaceans and small bony fishes. Probably a bycatch of trawl fisheries.



Bythaelurus lutarius, 24 cm TL, immature male (S Mozambique). Source: Bass et al. 1975

# GENUS Halaelurus Gill 1862

Body slender; head moderately depressed. Eyes noticeably elevated. Snout acutely rounded and tip pointed, upturned or not upturned, and with or without knob; upper and lower lip folds short to minuscule; nasal flaps somewhat expanded, separate; no nasal barbels. Fins rounded to weakly falcate. Dorsal fins and anal fin more or less subequal. Pelvic fins sometimes partially fused, but never forming pelvic curtain; claspers stout to elongate, outer length 5–6 times width. No crest of enlarged denticles along upper peduncle and dorsal caudal margin. Stomach not inflatable. Tooth counts 54–60/41–55; vertebrae 117–142; spiral valve turns 7–10. Abundant to rare on continental shelf, inshore or offshore, in Indo-Pacific. At least 7 species, 4 in WIO.

#### **KEY TO SPECIES**

1a 1b	Snout tip upturned Snout tip not upturned	
		uh la
	Id	a
2a	Mouth width 6–7% TL; sade edges, and numerous smal	dle markings dusky with thin dark I dark spots on body <b>H. lineatus</b>
2b	Mouth width 7–9% TL; sade dark edges, and no spots o	dle markings prominent with broad n body <b>H. natalensis</b>
3a 3b	Snout tip without knob Snout tip with knob	H. boesemani H. quagga

## Halaelurus boesemani Springer & D'Aubrey 1972

PLATE 32

#### Speckled catshark

Halaelurus boesemani Springer & D'Aubrey 1972: 11, Fig. 1 (Somalia); Springer 1979\*; Compagno 1984\*, 1988; Bonfil & Abdallah 2004\*; Compagno et al. 2005\*; White et al. 2007. Scyliorhinus buergeri: Regan 1908. Scyliorhinus quagga: Norman 1939\*.

Head broad; snout tip pointed (but not upturned) and without knob; mouth width 6–8% TL, mouth length 3–4% TL. Tooth count 57/55; vertebrae 135–139; spiral valve turns 6.

Body pale brown to yellowish brown background dorsally, with ~8 darker brown bands or saddle markings; numerous small dark spots scattered over body, dorsal and caudal fins, and large dark blotches on dorsal fins and caudal fin; paler ventrally and on pelvic and anal fins. Attains 48 cm TL.

**DISTRIBUTION** Indo-Pacific (range uncertain). WIO: Somalia and Gulf of Aden.

**REMARKS** Poorly known; previously confused with the western Pacific *H. buergeri* until it was separated as distinct by Springer & D'Aubrey (1972). Its presence beyond WIO needs to be reassessed as species recently described from Australia and Indonesia/Philippines were previously recorded as this species, and thus records of *H. boesemani* from Australia are erroneous and its presence in Indonesia is now questionable. Retained oviparity, with up to 4 egg cases per uterus. Probably uncommon with restricted range, on continental and insular shelves, at 37–91 m. Presumably subject to bycatch in demersal trawl and artisanal net fisheries.



Halaelurus boesemani (Somalia). Source: Springer & D'Aubrey 1972



Halaelurus lineatus, 48 cm TL, mature female (top); 13 cm TL, immature male paratype (bottom) (both South Africa). Source: Bass *et al.* 1975

# Halaelurus lineatus Bass, D'Aubrey & Kistnasamy 1975

Banded catshark

PLATES 28 & 32

Halaelurus lineatus Bass, D'Aubrey & Kistnasamy 1975: 12, Figs. 8, 20e
(Durban shoreline, KwaZulu-Natal, South Africa); Springer 1979\*;
Compagno 1984\*, 1988; SSF No. 11.5\*; Compagno et al. 1989\*;
Heemstra & Heemstra 2004; Compagno et al. 2005\*; White et al. 2007.
Scylliorhinus natalensis: Gilchrist & Thompson 1911; Gilchrist 1922.
Scyliorhinus natalensis: Fowler 1934.

Halaelurus natalensis: Smith 1950.

Head narrow; snout tip pointed and upturned with knob; mouth width 6–7% TL, mouth length 2% TL. First dorsal fin slightly larger than 2nd. Caudal-fin postventral margin concave. Tooth count 56–60/53–55; vertebrae 124–142; spiral valve turns 7.

Body pale brown background dorsally, with ~15 thin, prominent dark brown bands arranged in pairs outlining dusky saddles, and blanketed with many small dark spots; cream ventrally. Attains 56 cm TL.

**DISTRIBUTION** WIO: South Africa (Eastern Cape) to Mozambique (Beira).

**REMARKS** Relatively common, although with restricted range; previously confused with *H. natalensis*. Retained oviparity, with up to 8 egg cases per uterus; hatched in captivity at ~8 cm TL after 23–36 days, although young may be live born in the wild. Segregated, with gravid females predominating in

northern regions of its range, and a larger proportion of males and juveniles occurring southwards and by depth. Found on continental shelf and upper slope, from surf zone to ~290 m deep. Feeds mostly on crustaceans, but also small bony fishes and cephalopods. A frequent bycatch of prawn trawl fishery.

# Halaelurus natalensis (Regan 1904)

#### Tiger catshark

PLATES 28 & 32

*Scyllium natalense* Regan 1904: 128 (KwaZulu-Natal, South Africa). *Scyliorhinus natalensis*: Regan 1908; Fowler 1934, 1935. *Scylliorhinus natalensis*: Gilchrist & Thompson 1911, 1914; Thompson 1914; Gilchrist 1922; Von Bonde 1923; Barnard 1925.

*Halaelurus natalensis*: Smith 1949\*, 1950; Bass *et al.* 1975\*; Springer 1979\*; Compagno 1984\*, 1986\*, 1988\*; SSF No. 11.7\*; Compagno *et al.* 1989\*; Compagno *et al.* 1991; Heemstra & Heemstra 2004\*; Compagno *et al.* 2005; Human *et al.* 2006; White *et al.* 2007.

Head broad; snout tip pointed and upturned with knob; mouth width 7–9% TL, mouth length 2–3% TL. First dorsal fin slightly larger than 2nd. Tooth count 54/41; vertebrae 123–133.

Body yellow-brown background dorsally, with ~10 conspicuous broad red-brown saddles with thick darker brown margins, and no spots; creamy ventrally. Attains 50 cm TL.

**DISTRIBUTION** WIO: endemic to South Africa (Cape Agulhas to southern KwaZulu-Natal).



**REMARKS** Although originally described from KwaZulu-Natal, South Africa, large-scale trawl surveys since the mid-1980s have shown this species to be most common south of East London in the Eastern Cape. Also, historical confusion between this species and *H. lineatus* makes it difficult to attribute records from KwaZulu-Natal to *H. natalensis*, and records from Mozambique are almost certainly attributable to *H. lineatus* instead. Retained oviparity, with 6–11 egg cases per uterus; embryos ~4.5 cm TL when eggs are laid. Found on continental shelf, from surf zone to at least 172 m deep (most commonly <100 m). Feeds predominantly on bony fishes and crustaceans, but also cephalopods and other small sharks. Frequently caught by recreational anglers, in commercial crayfish traps, and a discarded bycatch of bottom trawlers.

## Halaelurus quagga (Alcock 1899)

Quagga catshark

PLATES 29 & 32

*Scyllium quagga* Alcock 1899: 17 (Laccadive Sea [Lakshadweep], Malabar coast, India).

Scyliorhinus quagga: Regan 1908.

Halaelurus quagga: Springer & D'Aubrey 1972\*; Springer 1979\*; Compagno 1984\*, 1988\*; White et al. 2007; Akhilesh et al. 2011.

Small-sized and extremely slender. Head narrow; snout tip pointed and with knob, but not upturned; mouth width 6–9% TL, mouth length 4% TL. Second dorsal-fin base longer than 1st. Tooth count 55–59/54–55; vertebrae 123–125.

Body pale brown dorsally, with 20+ narrow, equally spaced dark brown bands (those under dorsal fins forming slight saddle markings) and no spots; paler ventrally. Attains at least 35 cm TL.

**DISTRIBUTION** WIO: Somalia, Socotra, Maldives, Lakshadweep and India (Kerala and southwards).

**REMARKS** Poorly known from only a few specimens. Differences in colour patterns (Springer 1979) between specimens from India and Somalia may indicate population substructuring. Uncommon; occurs in tropical offshore waters of outer continental shelf, at 54–220 m. Presumably taken as bycatch of demersal fisheries.

# GENUS Haploblepharus Garman 1913

Body slender to moderately stocky; head not depressed. Eyes moderately elevated or not elevated. Snout broadly rounded, or narrowly rounded and coming to a point, and tip not upturned; nostrils very large; nasal flaps expanded, fused and extending to mouth; oronasal grooves deep; no nasal barbels. Upper and lower lip folds short. Fins rounded. Dorsal fins and anal fin more or less equally sized. Pelvic fins not fused; claspers long and robust, outer length 3–8 times width. No crest of enlarged denticles along upper peduncle and dorsal caudal margin. Stomach not inflatable. Tooth counts 40–90/46–92; vertebrae 93–141; spiral valve turns 6–9. Inshore, abundant to rare, found primarily in kelp forests and on rocky reefs, but also along sandy shores; all members are poorly represented in museum collections. At least 4 species, all in WIO.

#### **KEY TO SPECIES**

[ <b>Ex</b> ( 1a	cludes juveniles] Body slender, compressed to moderately depressed; sourt acutaly rounded tooth in upper jaw <70 in total
1b	Body stocky, moderately to greatly depressed; snout broadly rounded, not acutely pointed; teeth in upper jaw $\geq$ 75 in total
2a	Head width ≥1.3 times head height at pectoral-fin origins <i>H. edwardsii</i>
2b	Head width ≤1.3 times head height at pectoral-fin origins H. kistnasamyi
3a	Saddle markings and sparse white spots obvious (these markings usually occur together)
3b	Saddle markings obscure, and black or white spots inconspicuous or absent (saddles and spots rarely occur together)
	occur together)



Halaelurus quagga, 35 cm TL, adult male (Somalia). Source: Springer 1979



Haploblepharus edwardsii, 49 cm TL, mature male (top right); 13 cm TL, juvenile, dorsal and lateral views (bottom left); 53 cm TL, female, dorsal view (bottom right) (all South Africa). Source: Bass et al. 1975 (male); Smith 1953 (juvenile); SFSA (female)

#### Haploblepharus edwardsii (Schinz 1822)

#### Happy Eddie

PLATE 29

Squalus edwardsii Schinz 1822: 504 (False Bay, Western Cape, South Africa).

Catulus edwardii: Smith 1837.

Scyllium edwardsii: Müller & Henle 1838\*; Gilchrist 1902.

Scyliorhinus edwardsii: Regan 1908.

*Scylliorhinus edwardsi*: Thompson 1914; Gilchrist & Thompson 1916; Barnard 1925.

Scylliorhinus edwardsii: Gilchrist 1922.

Haploblepharus edwardsii: Smith 1950\*; Bass 1973; Bass et al. 1975\*; Springer 1979\*; Compagno 1984\*, 1986\*, 1988; SSF No. 11.8\*; Compagno et al. 1989\*; Van Oijen 2001; Heemstra & Heemstra 2004; Compagno et al. 2005\*; Human & Compagno 2006\*; Human et al. 2006; Human 2007\*.

Trunk moderately depressed; head rounded, head height at rear edge of orbit 5–7% TL; snout short. Eyes moderately elevated. Claspers slender, inner length 3–6 times width. Tooth count 51–68/46–60; vertebrae 121–139; spiral valve turns 7.

Body pale brown to pale or dark grey-brown background dorsally, with numerous small white spots, and 8–10 prominent saddles with brilliant orange or orange-brown centres and dark brown to almost black margins; paler ventrally. Attains 60 cm TL.

**DISTRIBUTION** WIO: endemic to South Africa (Langebaan Lagoon in Western Cape to Algoa Bay in Eastern Cape), with possibly severely fragmented populations.

**REMARKS** Historically confused with other *Haploblepharus* species (particularly H. pictus), resulting in confused distributional and biological reports, and probably leading to overestimations in its abundance and range. Oviparous, pairs of egg cases laid aseasonally, hatching after 3 months. Matures at ~7 years; maximum age at least 22 years. Occurs over continental shelf and slope, at 30-130 m; found in kelp forests and along rocky shores (western part of its range), and on sandy bottom in deeper water (eastern part of its range); possibly separate populations in deeper water attain greater sizes than inshore populations. Social, frequently aggregating and resting in groups; if disturbed these sharks will curl up with their caudal fin over their eyes, hence the common name shyshark. Feeds on small bony fishes, worms, crustaceans, squid and octopus. Treated as a nuisance catch by recreational anglers; also used in aquariums. IUCN Red List conservation status Endangered.

# Haploblepharus fuscus Smith 1950

#### Brown shyshark

PLATES 29 & 32

Haploblepharus fuscus Smith 1950: 883, Fig. 2 (East London, Eastern Cape, South Africa); Bass et al. 1975\*; Springer 1979; Compagno 1984\*, 1986\*, 1988\*; SSF No. 11.9\*; Compagno et al. 1989\*; Compagno et al. 2005\*; Human & Compagno 2006; Human et al. 2006; Human 2007\*.
Haploblepharus edwardsii: Smith 1949\*.



Haploblepharus fuscus, 43 cm TL, immature male (South Africa). Source: Bass *et al.* 1975 Body stocky; head broad, head height at rear edge of orbit 5–8% TL; nostrils large. Eyes moderately elevated. Claspers stout, inner length 3–4 times width. Tooth count 40–90/48–92; vertebrae 128–141; spiral valve turns 9.

Body uniformly brown or dull grey-brown dorsally, with slightly darker saddle markings variably present (more conspicuous in smaller specimens); rarely with inconspicuous black spots or inconspicuous white spots (never both); white ventrally. Attains 73 cm TL.

**DISTRIBUTION** WIO: endemic to South Africa (site-specific, fragmented population: Storms River mouth, Western Cape, to near Durban, KwaZulu-Natal).

**REMARKS** Biology unknown, assumed singly oviparous. Very restricted, close inshore range, from surf zone to ~35 m deep. Adults are locally common over rocky reefs; juveniles possibly use unknown habitat as nursery areas, and are rare in collections. Presumably feeds on small bony fishes and crustaceans. Treated as a nuisance catch by rock and surf anglers. IUCN Red List conservation status Vulnerable.

## Haploblepharus kistnasamyi

Human & Compagno 2006

#### Happy chappie

PLATE 32

Haploblepharus kistnasamyi Human & Compagno 2006: 44, Figs. 2–5 (Park Rynie, KwaZulu-Natal, South Africa); Human 2007\*.

Haploblepharus edwardsii: Smith 1949 [in part], 1950\* [in part]; Bass et al. 1975\* [in part]; Compagno 1984\* [in part], 1988 [in part]; SSF No. 11.8\* [in part]; Compagno et al. 1989\* [in part].

Haploblepharus sp. A: Compagno et al. 2005\*; Human et al. 2006.

Stocky body, trunk not depressed; head slightly depressed, head height at rear edge of orbit 6–11% TL. Nasal flaps with slight median indentation and reaching to upper lip. Claspers stout, inner length 5 times width. Tooth count 55–81/53–74; vertebrae 121–133; spiral valve turns 6. Body with 8 or 9 dark brown, loosely H-shaped saddle markings, the centres darker than background, and white spots only on these saddles (giving blotchy effect over body and fins); saddle markings in juveniles conspicuous, with orange-brown centres and dark brown margins; interspaces between saddles and on fins with dark mottling on brown background; abruptly white ventrally. Attains 50 cm TL.

**DISTRIBUTION** WIO: endemic to southern Mozambique and South Africa (eastern Western Cape to KwaZulu-Natal).

**REMARKS** Superficially resembles *H. edwardsii*; first recognised as distinct by Bass *et al.* (1975) who described it as the Natal form and *H. edwardsii* proper as the Cape form. Biology unknown, presumably singly oviparous. Rarely seen, with very a restricted range and possibly a single population: so far, adults found only from southern Mozambique (Techobanine) (to ~100 m deep) to KwaZulu-Natal (Park Rynie), South Africa (collected from 40–50 m), and juveniles tentatively assigned to this species only found further south, to Mossel Bay in South Africa. Usually close inshore, from surf zone to ~30 m deep. IUCN Red List conservation status Vulnerable.

## Haploblepharus pictus (Müller & Henle 1838)

Pretty happy

PLATE 32

- *Scyllium pictum* Müller & Henle 1838: 4 (Cape of Good Hope, South Africa).
- Scyliorhinus edwardsii: Regan 1908.
- *Scylliorhinus edwardsi:* Thompson 1914; Gilchrist & Thompson 1916. *Haploblepharus edwardsii:* Smith 1950.

Haploblepharus pictus: Bass et al. 1975\*; Springer 1979\*; Compagno 1984\*, 1988\*; SSF No. 11.10\*; Compagno et al. 1989\*, 2005\*; Van Oijen 2001;

Heemstra & Heemstra 2004; Human & Compagno 2006\*;

Human et al. 2006; Human 2007\*.



Haploblepharus kistnasamyi, 48 cm TL, mature female (South Africa). Source: Bass et al. 1975



Head and trunk moderately depressed; head broad, height at rear edge of orbit 6–8% TL. Eyes moderately elevated. Claspers stout, inner length 3–5 times width. Tooth count 45–83/47–75; vertebrae 93–133.

Body pale to dark brown (tawny coloured) or almost black, grey, grey-brown or red-brown background dorsally; 6–8 faint saddle markings, the centres variously orange, brown or dark brown to almost black, and the margins often indistinct; irregular large white spots absent or present on saddles or on and between saddles (never just between saddles); abruptly white or creamy ventrally. Attains 60 cm TL.

**DISTRIBUTION** Endemic from Namibia in southeastern Atlantic, to South Africa (Storms River mouth, Eastern Cape) in WIO.

**REMARKS** Long considered a synonym of *H. edwardsii* and largely ignored until it was resurrected by Bass *et al.* (1975), and hence recognised as *H. pictus* by subsequent authors. However, misidentification with *H. edwardsii* and other species continued largely due to its highly variable patterning. Oviparous, single egg case laid aseasonally; hatched in captivity after 3.5–10 months. Matures at ~15 years; maximum age at least 25 years. Has a greater range than previously thought; common to abundant in shallow inshore waters, in kelp forests and along rocky reefs and shores, to ~35 m deep. Feeds on small bony fishes, worms, crustaceans, cephalopods, molluscs (snails), echinoderms and occasionally algae. A common bycatch in recreational fishing and generally treated as a nuisance catch; regularly seen by divers.

## GENUS Holohalaelurus Fowler 1934

Body slender; head strongly depressed; tail long; some with enlarged denticles on mid-back, but no crest of denticles on dorsal caudal margin. Eyes dorsolateral on head, very broad subocular ridges below eyes. Snout short, broadly rounded; no lip folds; nasal flaps short, separate; no nasal barbels. Dorsal fins subequal and higher than anal fin, but anal fin much longer than dorsal-fin lengths. Pelvic fins partially fused but never forming pelvic curtain; claspers elongate, their outer length 4–8 times width. Anal fin long and low. Tail long and slender. Stomach not inflatable. Tooth counts 40–68/38–78; vertebrae 85–132; spiral valve turns 7. All species have conspicuous black sensory pores ventrally (fresh specimens). Males appear to attain greater sizes than females.

Although *Holohalaelurus* sharks have long been known from East Africa, their biology is poorly known; their taxonomy was revised by Human (2006), although they remain poorly represented in collections. Historically abundant but now mostly rare, over the continental shelf and slope; some species and populations from southwestern WIO are possibly extinct in the region. Reports of *Holohalaelurus* from Somalia are likely valid but require confirmation. At least 5 species, all in WIO. Of these, *H. melanostigma* is known only from four male specimens collected from 607–658 m, off Tanzania and Kenya, and is distinguished by a tear-line marking on the snout forward of the eyes.

#### **KEY TO SPECIES**

#### [Excludes juveniles]

1a	No enlarged denticles on midback; small white spot at origin of both dorsal fins, and C- or V-shaped mark on each dorsal fin
1b	Denticles on dorsal midline, moderately or greatly enlarged; small white spot absent or variably present at origin of either dorsal fin (never at origin of both fins)
2a 2b	Anal-fin base ≤1.5 times pelvic-fin base; club-shaped papillae on tips of claspers; several white spots distributed on upper body, and short horizontal bar on each dorsal fin <i>H. grennian</i> Anal-fin base >1.8 times pelvic-fin base; no papillae on tips of claspers; no white spots anywhere on body
	Cartinual

Continued ...

#### **KEY TO SPECIES**

3a	No enlarged denticles on dorsal surface of snout, and no enlarged denticles on dorsal surface of pectoral fins in adults; 'tear-line' markings present from front edge of eyes to anterior margin of snout	
3b	Enlarged denticles present on dorsal surface of shout and on dorsal surface of pectoral fins in adults; no 'tear-line' markings from front edge of eyes to anterior margin of shout	
4a	Markings on body resembling honeycomb pattern; at most 1 symphysial tooth in each jaw	
4b	Irregular horseshoe-shaped markings on body; up to 6 symphysial teeth in each jaw	

## Holohalaelurus favus Human 2006

Honeycomb Izak

PLATE 32

*Holohalaelurus favus* Human 2006: 36, Figs. 11–12 (KwaZulu-Natal, South Africa); Human 2007, 2010.

Holohalaelurus regani (non Gilchrist 1922): Bass et al. 1975\*; Compagno 1984; SSF No. 11.12\*; Compagno et al. 1989; Richardson et al. 2000; Compagno et al. 2005\*.

Head broad; 1st dorsal fin rounded, 2nd dorsal fin angular. Slightly enlarged denticles on midback from snout to 2nd dorsal-fin, and denticles on dorsal surface of pectoral fins greatly enlarged in adults. Buccal papillae large and prominent on upper and lower surfaces in mouth; no papillae on tips of claspers. Tooth count 65–68/53–70; vertebrae 104–124. Body yellowish brown background dorsally, covered with relatively regular checkerboard-like spots resembling honeycomb pattern; rarely with inconspicuous white spots above pectoral fins; paler grey-brown ventrally. Attains ~52 cm TL.

**DISTRIBUTION** WIO: South Africa (KwaZulu-Natal) to southern Mozambique.

**REMARKS** Long confused with *H. regani* (sometimes referred to as the north-eastern subspecies) and illustrated several times under that name. First recognised as distinct by Bass *et al.* (1975) who referred to it as the Natal form rather than the Cape form of true *H. regani*, and subsequent authors recognised the two forms. Oviparous, probably pairs of egg cases laid. Possibly a single population, with estimated depth range of 200–1 000 m on upper to mid slope, with juveniles preferring deeper water than adults. Probably feeds on small bony fishes, cephalopods and crustaceans. Previously a common bycatch off South Africa (KwaZulu-Natal) and southern Mozambique, but now rarely recorded as bycatch in the KwaZulu-Natal prawn trawling fishery. IUCN Red List conservation status Endangered.

# Holohalaelurus grennian Human 2006

Grinning Izak

PLATE 30

*Holohalaelurus grennian* Human 2006: 44, Figs. 14–16 (off Ras Ngomeni, Kenya); Human 2007, 2010\*.

Holohalaelurus punctatus (non Gilchrist 1914): Bass et al. 1975;
Springer 1979; Compagno 1984, 1988\*; SSF No. 11.11\*;
Compagno et al. 1989; Compagno et al. 1991.
Holohalaelurus sp. A: Compagno et al. 2005\*.



*Holohalaelurus favus*, 13 cm TL, immature male (South Africa). Source: Bass *et al.* 1975

Holohalaelurus favus, 52 cm TL, holotype (South Africa). © BA Human



Head broad; dorsal fins subequal. Slightly enlarged denticles on midback from above pectoral-fin origins to 1st dorsal fin, and denticles on dorsal surface of pectoral fins not enlarged. Buccal papillae inconspicuous; club-shaped papillae on tips of claspers. Tooth count 40-43/38-41; vertebrae 90-112.

Body yellow-brown to grey-brown background dorsally, with numerous small brown spots; characteristic horizontal dark bar or C-shaped mark on both dorsal fins; white spot absent or variably present at origin of either dorsal fin (never at origin of both fins); large pale or white spots above pectoral fins; paler ventrally. Attains at least 27 cm TL.

**DISTRIBUTION** WIO: northern Tanzania, Zanzibar, southern Kenya (few records) and southern Mozambique.

**REMARKS** Smallest of the *Holohalaelurus* sharks. A single female specimen, along with several male specimens, was included in the original description of *H. melanostigma*, with differences between male and female specimens attributed to sexual dimorphism. However, additional male specimens were later collected and readily identifiable with the female specimen previously collected, thus justifying the separation of this species from *H. melanostigma*. Very rare, known only from five museum specimens; biology unknown, presumably singly oviparous. Possibly a single population, recorded from 238–353 m, on upper slope.

#### Holohalaelurus punctatus (Gilchrist 1914)

#### White-spotted Izak

PLATE 30

- *Scylliorhinus punctatus* Gilchrist 1914: 129, Fig. (off Bazaruto, Mozambique); Thompson 1914; Gilchrist 1922; Barnard 1925; Norman 1939.
- *Scyliorhinus (Halaelurus) polystigma* Regan 1921: 413 (KwaZulu-Natal, South Africa); Norman 1939\*.

Halaelurus punctatus: Fowler 1934, 1935\*.

Holohalaelurus punctatus: Smith 1949\*; Bass 1973; Bass et al. 1975\*; Springer 1979\*; Compagno 1984\*, 1988\*; SSF No. 11.11\*; Séret 1987; Compagno et al. 1989\*; Compagno et al. 1991; Compagno et al. 2005\*; Human 2006\*, 2007, 2010.

Head broad; dorsal fins angular. No enlarged denticles on midback or dorsal surface of pectoral fins in adults. Buccal papillae large and prominent on upper and lower surfaces of mouth; no papillae on tips of claspers. Tooth count 42–52/40–47; vertebrae 85–120.

Body golden brown to dark brown background dorsally, densely covered with irregularly shaped dark brown spots (but no reticulations, blotches or horizontal stripes); few white spots scattered on back and at origins of both dorsal fins; C- or V-shaped marking on each dorsal fin; paler ventrally, and sometimes with tiny dark dots and dashes under head. Attains 34 cm TL.



Holohalaelurus punctatus, 24 cm TL, immature male (South Africa). Source: Bass *et al.* 1975 **DISTRIBUTION** WIO: South Africa (KwaZulu-Natal) to Mozambique and Madagascar; its occurrence off Cape Point, South Africa (as stated in literature), is highly unlikely.

**REMARKS** Oviparous, pairs of egg cases laid. May be comprised of two distinct populations, one off South Africa and Mozambique, and the other off Madagascar. Estimated depth range of 219–420 m off Mozambique and South Africa, and 280–600 m off Madagascar; juveniles may live in deeper water than adults. Probably feeds on bony fishes, crustaceans and cephalopods. Previously a common bycatch in fisheries off South Africa and Mozambique, but now rarely recorded off KZN; records require confirmation due to confusion with *H. favus*; the status of the Madagascan population is unknown. IUCN Red List conservation status Endangered; Data Deficient in Madagascar.

## Holohalaelurus regani (Gilchrist 1922)

Izak

PLATES 30, 31 & 32

Scylliorhinus regani Gilchrist 1922: 45 (SE of Hondeklip Bay, Northern Cape, South Africa); Barnard 1925.

Holohalaelurus regani: Smith 1949\*; Bass 1973; Bass et al. 1975\*; Springer 1979\*; Compagno 1984\*, 1986\*, 1988\*; SSF No. 11.12\*; Compagno et al. 1989\*; Compagno et al. 1991; Ebert et al. 1996; Richardson et al. 2000\*; Compagno et al. 2005\*; Human 2006\*, 2007, 2010; Human et al. 2006.

Snout short, mouth long. Slightly enlarged denticles on midback from snout to 2nd dorsal fin, and denticles on dorsal surface of pectoral fins greatly enlarged in adults. Buccal papillae large and prominent on upper and lower surfaces of mouth; no papillae on tips of claspers. Tooth count 55–72/ 27–78; vertebrae 106–132; spiral valve turns 7.

Body pale yellow to yellow-brown background dorsally, with bold reticulations and irregular horseshoe-shaped markings and virtually no spots, and never with white spots in adults; whitish ventrally; juveniles darker, with irregularly shaped larger dark brown spots, and also row of white spots along each side. Attains 69 cm TL (males reach larger sizes than females).



Holohalaelurus regani, 61 cm TL, mature male (South Africa). Source: SFSA

**DISTRIBUTION** Southern Africa: Namibia in southeastern Atlantic, to South Africa (Eastern Cape, possibly KwaZulu-Natal and southern Mozambique) in WIO.

**REMARKS** Oviparous and highly fecund, with pairs of egg cases laid aseasonally; size at hatching 8-10 cm TL. Found over continental shelf, at 40-910 m (mostly 100-500 m). Abundant in southern and western parts of its distribution, where continental shelf is broadest. Most records of H. regani in the eastern part of its distribution are referable to H. favus (once considered a subspecies). Minor seasonal migrations occur in at least part of its range, and depth segregation is apparent, with females dominant above 200 m and males occurring deeper; juveniles most commonly found in shallow water. Males attain ~20% greater length than females, and juveniles are much more elongate than adults (possibly true for all Holohalaelurus species). Active and demersal, feeds on small bony fishes, crustaceans, cephalopods, polychaetes, kelp and possibly fishing discards. A regular discard of demersal hake trawl fishery throughout its range.



Holohalaelurus regani, 57 cm TL, mature male (South Africa). Source: Bass et al. 1975

# FAMILY SCYLIORHINIDAE

#### Catsharks

Brett A Human and David A Ebert

Generally small-sized, long and slender (most WIO species <80 cm TL), but a few are relatively large and stocky. Head length typically 20% TL; 5 pairs of gill slits, small to greatly enlarged, the last gill slit over or behind pectoral-fin origins. Snout broadly to narrowly rounded, not depressed to moderately depressed and laterally expanded compared to rest of head, and tip not upturned. Anterior nasal flaps short or extending to mouth, and separate or fused; nasal barbels present or absent; oronasal grooves absent. Eyes slit-like and dorsolateral, with rudimentary nictitans and strong subocular ridges; spiracles moderately large, set just behind eyes. Supraorbital crest present (usually detected by touch), and is the main character separating the catshark families.



Mouth inferior, long and arched, and with or without lip folds. Teeth multicuspidate (except monocuspidate in one species of *Schroederichthys* not known from WIO) and in multiple functional rows in both jaws. Fins rounded, angular or weakly falcate. Two spineless dorsal fins, usually subequal; 1st dorsal fin above or behind pelvic-fin bases. Pectoral fins moderately sized and paddle-shaped. Pelvic fins moderately long and low. Anal fin long and low to high, its base usually longer than 2nd dorsal-fin base. Peduncle and caudal fin slender; peduncle without pits or keels; caudal fin heterocercal, lower lobe poorly developed. Sexual dimorphism in TL absent to extreme (with males attaining greater lengths than females) and also apparent in dentition of some species. Tooth counts 40–116/38–110; vertebrae 100–183; spiral valve turns 6–14.

Reproductive mode oviparous, with females depositing egg cases on the substrate where the embryos develop and eventually hatch out. However, for some species the egg cases are retained *in utero* until the embryos are nearly completely developed. Scyliorhinids are found worldwide in tropical to cold-temperate seas, with preferred temperatures likely contributing to a high degree of endemism. Most shallowwater species occur in tropical to warm-temperate regions and tend to be brightly coloured, whereas deep-sea forms tend to be more uniformly coloured. Sluggish, poor swimmers, and generally deep-dwelling (<1 000 m) on or near bottom, although some deep-sea species are known to migrate far off the bottom into the water column; in the WIO they are found from intertidal zone to deep-sea (<700 m), over soft bottom, on rocky or coral reefs and in kelp beds, and may be rarely seen to locally abundant. Harmless to people. Generally not targeted in fisheries but often comprise a bycatch where abundant; some of the more colourful species are displayed and bred in aquariums.

This family has 7 genera and ~50 species; 4 genera and 7 species in the WIO, with one of the species from deep water.



Typical catshark egg case. Source: SFSA

#### GLOSSARY

**supraorbital crest** – narrow cartilaginous ridge from the chondrocranium extending above the eyes in catsharks of the family Scyliorhinidae. The crests can usually be felt by touch without needing any dissection.

#### **KEY TO GENERA**





## GENUS Atelomycterus Garman 1913

Body slender; head narrow and slightly depressed. Eyes not noticeably elevated. Supraorbital crest present. Snout narrowly rounded and coming to a point, not depressed, and tip not upturned; upper and lower lip folds long; nasal flaps long, extending to mouth, and separate or fused; no nasal barbels. Fins rounded to weakly falcate. Dorsal fins subequal, and dorsal fins much larger than anal fin. Pelvic fins not fused; claspers elongate, outer length 5–11 times width. No crest of enlarged denticles along upper peduncle or dorsal caudal margin. Stomach not inflatable. Tooth counts 56–89/50–74; vertebrae 147–183; spiral valve turns 11–14. Predominantly Australasian distribution; strongly associated with coral reefs and frequently found in very shallow water, otherwise poorly known. At least 5 species, 1 in WIO.

# Atelomycterus marmoratus

(Anonymous [Bennett] 1830)

Coral catshark

PLATE 32

Scyllium marmoratum Anonymous [Bennett] 1830: 693

(Sumatra, Indonesia); Smith 1837.

Scyllium maculatum: Müller & Henle 1838; Day 1878\*.

Scyliorhinus marmoratus: Regan 1908.

Atelomycterus marmoratus: Springer 1979\*; Compagno 1984\*, 1988\*; Yamakawa *et al.* 1995\*; Compagno *et al.* 2005\*; White *et al.* 2005; Jacobsen & Bennett 2007.

Tooth count 74/74; vertebrae 157–162; spiral valve turns 7.

Body grey to dark grey background dorsally, with many large white spots and dash and bar marks over body and fins, some bordered by thick black margins and spots, giving mottled appearance but no clear saddle markings; both dorsal fins conspicuously white-tipped, and pectoral and pelvic fins with white margins; abruptly pale ventrally. Attains 70 cm TL.

**DISTRIBUTION** Indo-Pacific. WIO: Pakistan to southern India; elsewhere to Malaysia, Indonesia, New Guinea, Philippines, Taiwan and southern China.

**REMARKS** Wide-ranging, but poorly known. Oviparous, egg cases laid in pairs; size at hatching 10–13 cm TL. Inshore, common on coral reefs, often found in crevices and holes; nocturnal. Feeds on crustaceans and small fishes. Common bycatch of artisanal fisheries, and a hardy aquarium species. IUCN Red List conservation status Near Threatened due to habitat degradation.



Atelomycterus marmoratus, 65 cm TL (Philippines). PC Heemstra © NRF-SAIAB

PLATES 31 & 32

# GENUS Cephaloscyllium Gill 1862

Body moderate to stocky; head moderately depressed. Eyes not elevated, dorsolateral on head, broad subocular ridges present below eyes. Supraorbital crest present. Snout broadly to narrowly rounded, tip not upturned; no lip folds; nasal flaps generally short and lobate, separate, and not reaching mouth (except in *C. silasi*); no nasal barbels. Fins rounded to weakly falcate. First dorsal fin larger than 2nd. Pelvic fins not fused; claspers short and stubby to elongate, outer length 4–7 times width. Anal fin subequal to 1st dorsal fin and larger than 2nd dorsal fin. No crest of enlarged denticles on dorsal caudal margin. Stomach inflatable (often highly inflatable). Tooth counts 48–116/45–110; vertebrae 100–140; spiral valve turns 7–12.

Members of this genus are also referred to as swellsharks because of their ability to swallow water or air when aggravated, thus distending their abdomen as an anti-predator tactic. Generally with restricted ranges, but common to uncommon locally and found in a wide range of habitats in Indo-Pacific. Recent descriptions of new species have more than doubled the number of species in the genus, with ~18 species now recognised, plus possibly additional species yet to be described; at least 2 species in WIO. *Cephaloscyllium silasi* (Talwar 1974), a dwarf species (attains 36 cm TL), collected from ~300 m deep off Kollam, India, in WIO (specimens from Andaman Sea off Myanmar may represent a different species), is distinguishable from *C. sufflans* by nasal flaps that reach to mouth.

#### **KEY TO SPECIES**

- 1a
   Dorsal colour pattern with indistinct dusky saddles

   present or absent
   C. sufflans

# Cephaloscyllium sufflans (Regan 1921)

Balloon shark

*Scyliorhinus (Cephaloscyllium) sufflans* Regan 1921: 413 (KwaZulu-Natal, South Africa); Norman 1939.

Scylliorhinus sufflans: Gilchrist 1922; Barnard 1925.

*Cephaloscyllium sufflans*: Fowler 1935\*; Smith 1949\*; Bass *et al.* 1975\*; Springer 1979; Compagno 1984\*; SSF No. 11.3\*; Compagno *et al.* 1989\*; Compagno *et al.* 2005\*.

Body robust. Ridges over eyes. Nasal flaps end well before mouth. Vertebrae 124–140.

Body pale grey background dorsally, with dusky grey-brown saddle markings which fade with age and may be absent in adults; paler and unspotted ventrally. Attains 110 cm TL.

**DISTRIBUTION** Indian Ocean: South Africa (KwaZulu-Natal), Mozambique and Comoros in WIO (patchy); records from Kenya and Gulf of Aden may be of a separate species (cf. Springer 1979; Heemstra *et al.* 2006); elsewhere, records from Vietnam appear to be of an undescribed species.

**REMARKS** Oviparous, pairs of egg cases laid; size at hatching ~20–22 cm TL; matures at 70–80 cm TL. Common, found on continental shelf and upper slope, over sand and mud bottom, at 40–600 m. Ontogenetic segregation, with immature individuals reported to be a relatively common bycatch in KwaZulu-Natal prawn trawls in South Africa; adults may live deeper or north of the trawl grounds. Feeds primarily on crustaceans (shrimp and lobsters) and cephalopods, but also bony fishes and elasmobranchs. Known prey item of the coelacanth (*Latimeria chalumnae*) in Comoros. A discarded bycatch of bottom trawlers.



Cephaloscyllium sufflans, 42 cm TL, immature male (South Africa). Source: Bass et al. 1975

# GENUS Poroderma Smith 1837

Body moderate to stocky; head only slightly depressed. Eyes moderately elevated. Supraorbital crest present. Snout broadly to narrowly rounded, and not strongly depressed; nasal flaps separate; nasal barbels well developed. Mouth short; upper and lower lip folds short. Dorsal fins posteriorly set; 1st dorsal fin larger than 2nd. Pelvic fins not fused; claspers short and stubby, their outer length 2–4 times their width. No crest of enlarged denticles along upper peduncle and dorsal caudal margin. Stomach not inflatable. Tooth counts 38–57/28–51; vertebrae 102–137; spiral valve turns 8–13. Common to abundant, inshore, often in kelp beds and on rocky reefs. Two species, both in WIO.

#### **KEY TO SPECIES**

- 1a Nasal barbels <1/2 preoral snout and not reaching mouth; prenarial length slightly less than mouth length; body with several dark longitudinal stripes and no spots ...... P. africanum

## Poroderma africanum (Gmelin 1789)

Pyjama shark

PLATES 33 & 34

Squalus africanus Gmelin 1789: 1494 (South Africa).

Scyllium africanum: Smith 1849\*; Bleeker 1878; Sauvage 1891; Gilchrist 1902, 1914.

Scyliorhinus africanus: Regan 1908.

*Scylliorhinus africanus*: Thompson 1914; Gilchrist & Thompson 1916; Gilchrist 1922; Barnard 1925, 1927, 1948.

Poroderma africanum: Fowler 1925, 1934; Smith 1949\*; Bass 1973;
Bass et al. 1975\*; Springer 1979\*; Compagno 1984\*, 1986\*, 1988\*;
SSF No. 11.13\*; Compagno et al. 1989\*; Heemstra & Heemstra 2004\*;
Compagno et al. 2005\*; Human 2006\*; Human et al. 2006.

Body stocky; snout narrowly rounded. Nasal barbels prominent but short, not reaching mouth. Tooth count 41–49/30–47; vertebrae 115–137; spiral valve turns 9–12.

Body creamy to grey background, with several striking longitudinal dark brown to black stripes and no spots; albinism reported. Attains 109 cm TL.

**DISTRIBUTION** Endemic to South Africa, from Northern Cape in southeastern Atlantic to southern Cape and KwaZulu-Natal (rare) in WIO; records from Madagascar, Seychelles and Mauritius are erroneous.

**REMARKS** Oviparous, pairs of large  $(5 \times 10 \text{ cm})$  egg cases laid aseasonally, hatching after 4–5+ months. Matures at ~11 years; maximum age at least 22 years. Readily identifiable, common, inshore waters of continental shelf to upper slope, from intertidal zone to ~100 m deep, often along rocky shores, in caves and in kelp beds, with individuals apparently keeping a confined home range. Generally nocturnal. Feeds mostly on squid and octopus, but also bony fishes, other sharks and egg cases, hagfishes, molluscs, crustaceans, worms and echinoderms. Observed lying in squid egg beds to ambush mating squid during their spawning season. A common bycatch in various commercial, unregulated and sport fisheries; sometimes used for lobster bait; also a minor target of aquarium trade and hardy in captivity.



Poroderma africanum, 72 cm TL, mature female (South Africa). Source: Bass et al. 1975

# Poroderma pantherinum (Müller & Henle 1838)

#### Leopard catshark

PLATES 33 & 34

*Scyllium pantherinum* Müller & Henle (ex Smith) 1838: 14 (Cape of Good Hope, South Africa).

Poroderma pantherinum Smith 1838: 85 [name only]; Fowler 1925, 1934;
Bass et al. 1975\*; Springer 1979\*; Compagno 1984\*, 1986\*, 1988\*;
SSF No. 11.15\*; Compagno et al. 1989\*; Heemstra & Heemstra 2004;
Compagno et al. 2005\*; Human 2006\*; Human et al. 2006.

Poroderma submaculatum Smith 1838: 85 [name only].

Poroderma variegatum: Smith 1838.

*Scyllium leopardinum* Müller & Henle 1838: 13 [name only].

*Scyllium maeandrinum* Müller & Henle (ex Von Rapp) 1838: 13 [name only]. *Scyllium variegatum* Müller & Henle (ex Smith) 1838: 14 (Cape of Good

Hope, South Africa); Bleeker 1878; Sauvage 1891.

- *Scyllium pantherinum* Smith 1845: no page number, Pl. 25, Fig. 3 (near Algoa Bay, Eastern Cape, South Africa); Bleeker 1878; Sauvage 1891.
- Scyliorhinus pantherinus: Regan 1908.
- Scylliorhinus pantherinus: Thompson 1914; Gilchrist & Thompson 1916; Gilchrist 1922; Barnard 1925\*, 1927\*.
- *Scyliorhinus regani* [misidentification of *Poroderma pantherinum*]: Fowler 1925\*, 1926\*.
- Poroderma marleyi Fowler 1934: 234 (KwaZulu-Natal, South Africa); Smith 1949\*; Bass *et al.* 1975\*; Springer 1979; Compagno 1984\*, 1986\*; SSF No. 11.14\*.

Scyliorhinus leopardus: Fowler 1935.

Body moderate; snout broadly rounded. Nasal barbels reach mouth. Tooth count 38–57/28–51; vertebrae 102–124; spiral valve turns 8–13.

Body generally pale grey to charcoal grey background dorsally, with dense leopard-like spots or rosettes in irregular longitudinal rows; paler ventrally (and sometimes strongly demarcated). Colour pattern highly variable within a continuum of four colour morphs: 1) 'typical' morph with large black blotches, whole or broken rosettes, with spots sometimes fused into short thin broken lines to long thin lines; 2) 'salt and pepper' morph usually pale grey dorsal background with numerous small black spots; 3) 'marleyi' morph with pale grey dorsal background and few large black spots; and 4) 'melanistic' morph with faint longitudinal stripes and/or spots, or without patterning, and glossy jet black. Attains 84 cm TL.

**DISTRIBUTION** Endemic to South Africa (possibly fragmented populations), from Northern Cape in southeastern Atlantic, to KwaZulu-Natal in WIO, and possibly Mozambique; records from Madagascar and Mauritius need confirmation.

**REMARKS** Highly variable colour patterning in this species has led to taxonomic confusion, resulting in numerous synonyms. In particular, P. marleyi has been shown to be an extreme colour variant of adults of this species. Interestingly, nearly all hatchlings emerge with the 'marleyi' patterning, which usually develops into one of the other colour morphs, depending in part on its geographic range, possibly indicating multiple populations. While the 'typical' morph and its intermediates are found throughout its range, the 'salt and pepper' and 'marleyi' morphs are found only in the Eastern Cape of South Africa, and the 'melanistic' morph only in False Bay, Western Cape. Oviparous, pairs of egg cases laid aseasonally. Matures at ~10 years; maximum age ~19 years. Common, inshore, with biology and habitat similar to P. africanum, but also caught near sandy shores; recorded from shore to ~50 m deep. Taken by inshore commercial line fisheries and by recreational fishers and often persecuted by them; also heavily targeted for the aquarium trade; frequently seen by divers.



Poroderma pantherinum, 56 cm TL, immature female (top); 53 cm TL, immature male, ventral view (bottom left); 34 cm TL, immature male (right) (all South Africa). Source: Bass et al. 1975

## GENUS Scyliorhinus Blainville 1816

Body slender to stocky; head moderately depressed. Eyes not elevated. Supraorbital crest present. Snout broadly to narrowly rounded and not depressed; lip folds on lower jaw only; nasal flaps separate or fused; nasal barbels absent to moderately developed. Fins rounded or angular, but not falcate. First dorsal fin larger than 2nd. Pelvic fins partially to completely fused to form pelvic curtain; claspers short and stubby, their outer length 4–8 times width. Anal fin more or less subequal to 1st dorsal fin and larger than 2nd dorsal fin. No crest of enlarged denticles on dorsal caudal margin. Stomach not inflatable. Tooth counts 40–70/38–65; vertebrae 108–144; spiral valve turns 6–12. Estimated size range 32–162 cm TL. Rare to abundant; occur mainly in the Atlantic and are infrequent in the Indo-Pacific. At least 15 species, 2 in WIO.

#### **KEY TO SPECIES**

- 1a Preoral length <5% TL; nasal flaps small, separate .... *S. capensis*

#### Scyliorhinus capensis (Müller & Henle 1838)

Yellowspotted catshark

PLATE 33

*Scyllium capense* Müller & Henle 1838: 11 (Cape of Good Hope, South Africa); Day 1875\*; Gilchrist 1902.

*Scyliorhinus capensis*: Regan 1908; Smith 1949\*; Bass 1973; Bass *et al.* 1975\*; Springer 1979\*; Compagno 1984\*, 1986\*, 1988\*; SSF No. 11.16\*; Compagno *et al.* 1989\*; Compagno *et al.* 1991\*; Ebert *et al.* 1996; Richardson *et al.* 2000; Compagno *et al.* 2005\*; Ebert *et al.* 2006\*; Human *et al.* 2006.

*Scylliorhinus capensis*: Thompson 1914; Gilchrist 1922; Von Bonde 1922; Barnard 1927.

Relatively large-bodied. Nasal flaps small, separate, and end before mouth; no oronasal grooves. Pelvic fins completely fused to form pelvic curtain. Tooth count 65–70/55–65; vertebrae 130–144; spiral valve turns 8–12.

Body grey-brown background dorsally, with 8 or 9 darker brown or grey-brown saddle markings (sometimes poorly defined) and numerous small bright yellow or whitish spots (no dark spots); cream ventrally. Attains 122 cm TL.

**DISTRIBUTION** Southern Africa: Namibia in southeastern Atlantic, to South Africa (KwaZulu-Natal) in WIO; records from India are of a separate species.

**REMARKS** Oviparous, pairs of egg cases laid, possibly only during winter; hatches at 25–27 cm TL. Males mature at ~84 cm TL, and females at ~75 cm TL; males attain larger sizes than females. Bottom-dwelling over continental shelf and upper slope, at 26–695 m (mostly 200–400 m), and ventures deeper in warmer water; prefers rocky reef habitat. Coils tightly when disturbed or caught. Feeds on bony fishes and various invertebrates, especially crustaceans and worms. Common bycatch of commercial trawls, typically discarded. IUCN Red List conservation status Near Threatened.



Scyliorhinus capensis, 86 cm TL, adult male (South Africa). Source: Bass et al. 1975



Scyliorhinus comoroensis, 47 cm TL, adult male holotype (Comoros). Source: Compagno 1988

#### Scyliorhinus comoroensis Compagno 1988

Comoro catshark

PLATE 33

Scyliorhinus comoroensis Compagno 1988: 609, Figs. 1, 3–8 (Moroni, Grande Comore I.); Compagno *et al.* 2005\*; Heemstra *et al.* 2006.

Holotype: snout broadly rounded; nasal flaps large, fused and reach to mouth; no nasal barbels; no oronasal groves. Pelvic fins partially fused to form pelvic curtain. Tooth count 50/50; vertebrae 137; spiral valve turns 8.

Body pale grey-brown background dorsally, with distinct broad darker grey-brown saddles, and scattered with small to moderate-sized white spots (no small dark spots); dark bar under eyes; paler ventrally. Attains at least 60 cm TL.

#### DISTRIBUTION WIO: Comoros.

**REMARKS** Described from one specimen captured at ~400 m, but other individuals were subsequently photographed from a submersible in coelacanth (*Latimeria chalumnae*) habitat, on insular slope off Grande Comore I., at ~270 m deep. Bottom-dwelling; estimated depth range 200–400 m.

# FAMILY **PROSCYLLIIDAE**

#### Finback catsharks

Jenny M Kemper and David A Ebert

Dwarf- to small-sized with elongated fusiform body, narrowly rounded head; eyes slit-like with rudimentary nictitans, but no deep groove in front of eyes; spiracles moderately large. Nostrils without barbels or oronasal grooves; internarial space 0.5–1.9 nostril width; anterior nasal flaps broadly angular. Mouth long and angular, arched, and reaching vertical at front edge of eyes; small papillae inside mouth and on edges of gill arches; lip furrows very short or absent. Teeth small, with acute narrow cusps, often with lateral cusplets, and strong basal ledges and grooves, not blade-like, and similar in both jaws; posterior teeth comb-like. Two spineless dorsal fins, each much shorter than caudal fin; 1st dorsal-fin base well ahead of pelvic fins and usually closer to pelvic-fin bases than to pectoral-fin bases. Pectoral fins with radials confined to fin bases. Anal fin small. Caudal-fin lower lobe very weak or absent, no undulations or ripples on dorsal margin, and no precaudal pits.

Poorly known; bottom-dwelling on outer continental and insular shelves and upper slopes, at 50–766 m. Reproductive mode appears to be viviparity with yolk-sac placenta (except for oviparous *Proscyllium habereri* from western Pacific); most species feed on small fishes and invertebrates. A few species are taken as bycatch in commercial bottom trawls, but their small adult size (15–65 cm TL) makes them unsuitable for fisheries utilisation. Disjunct distributions, mainly in Indo-Pacific but also tropical northwestern Atlantic.

Three genera and at least 6 species; 2 genera and 3 species in WIO.

#### **KEY TO GENERA**

1a	Caudal fin broad, not ribbon-like, its length subequal	
	to HL; colour pattern of spots, saddles and bars on body	
	and fins Ctenacis	
1b	Caudal fin narrow, ribbon-like, and longer than head; no colour pattern except vertical barring on caudal fin	

## GENUS Ctenacis Compagno 1973

Relatively stout body and stout tail; head and snout not bellshaped in dorsoventral view; preoral length  $\sim^{2}/_{3}$  mouth width. Anterior nasal flaps small, with rear edges not reaching mouth; internarial space  $\sim$ 1.2 nostril width. Mouth triangular, with short lip furrows. Dorsal fins subequal; 1st dorsal-fin origin before pectoral-fin free rear tips. Anal-fin origin slightly behind 2nd dorsal-fin origin. Caudal fin relatively broad, short, not ribbon-like, its dorsal margin  $\sim$ 23% TL. One species.

## Ctenacis fehlmanni (Springer 1968)

#### Harlequin catshark

*Triakis fehlmanni* Springer 1968: 614, Figs. 1–4 (SW of Cape Guardafui, Somalia).

Ctenacis fehlmanni: Bass et al. 1975\*; Compagno 1984\*, 1988\*; SSF No. 10.1\*; Henderson & Reeve 2011; Ebert 2013\*; Ebert et al. 2013\*.

Diagnosis as for genus. Tooth count 86/88; vertebrae 136; spiral valve turns 10.

Body with unique pattern of large, reddish brown, irregular dorsal saddle blotches, interspersed with smaller round spots and vertical bars, as well as spots on fins. Attains at least 52 cm TL.

DISTRIBUTION WIO: Somalia and Oman.

**REMARKS** Females mature at ~52 cm TL (large, thin-walled egg cases in each uterus of the holotype suggests the species is viviparous with yolk-sac placenta); size at birth uncertain, but smallest free-swimming specimen was 17 cm TL.

Bottom-dwelling in tropical waters of outer continental shelf, usually deeper than 300 m. Likely feeds on very small invertebrates.

# GENUS **Eridacnis** Smith 1913

Small-sized, slender body, with narrow and moderately long head. Head and snout not bell-shaped in dorsoventral view; preoral length subequal to  $\sim \frac{2}{3}$  mouth width. Anterior nasal flaps small, with rear edges well in front of mouth; internarial space 0.7–1.3 times nostril width. Lip furrows extremely short to vestigial or absent. Teeth exhibit sexual heterodonty: males with slightly larger, lanceolate teeth. First dorsal-fin origin well anterior to slightly posterior to pectoral-fin free rear tips. Pectoral fins with straight or convex posterior margin. Anal-fin origin slightly anterior to, under, or slightly posterior to 2nd dorsal-fin origin. Caudal fin narrow, long and ribbon-like, its dorsal margin 25–30% TL. Tooth counts 55–78/63–77; vertebrae 113–144; spiral valve turns 6–8. Three species, 2 in WIO.

#### **KEY TO SPECIES**

- 1b
   Preoral length <1.5 mouth length; lateral dermal denticles narrow, with narrow, long cusps
   *E. radcliffei*



Ctenacis fehlmanni, 46 cm TL, adult female (Somalia). Source: Springer 1968; © Proc. Biol. Soc. Wash., Allen Press Publ. Services



*Eridacnis radcliffei*, 19 cm TL, male (Philippines). Source: SSF

PLATE 35

## Eridacnis radcliffei Smith 1913

Pygmy ribbontail catshark

PLATE 34

*Eridacnis radcliffei* Smith 1913: 599, Figs. 1–3 (Jolo I., Sulu Archipelago, Philippines); Nair & Appukuttan 1973, 1974; Nair & Lal Mohan 1973; Compagno 1984\*, 1988\*; SSF No. 10.2\*; Ebert 2013\*; Ebert *et al.* 2013\*.

Preoral length <1.5 mouth length; mouth triangular, lip furrows rudimentary or absent. Lateral dermal denticles narrow-crowned and with long, narrow cusps. Dorsal fins subequal, moderate and high; 1st dorsal-fin anterior margin at low angle to body axis. Anal-fin height <½ height of dorsal fins. Caudal fin narrow, long and ribbon-like; junction of caudal-fin pre- and post-ventral margins broadly rounded. Tooth count 72–78/65–77; vertebrae 113–130; spiral valve turns 6–8.

Body brown, with dark markings on dorsal fins, and prominent dark banding on caudal fin. Attains 24 cm TL.

**DISTRIBUTION** Indo-Pacific (patchy). WIO: Tanzania, Gulf of Aden (scattered) and southern India (common).

**REMARKS** Litters of 1 or 2; size at birth ~11 cm TL. Males mature by 18–19 cm TL, and females at 15–16 cm TL. Wideranging in tropical waters on upper continental and insular slopes and outer shelves, often over mud bottom, at 71–766 m. Feeds on small bony fishes and crustaceans, with squid a lesser component of diet. Interest to fisheries minimal; taken in commercial bottom trawls in Philippines.

## Eridacnis sinuans (Smith 1957)

African ribbontail catshark

*Neotriakis sinuans* Smith 1957: 262, Fig. 2 (off Durban, KwaZulu-Natal, South Africa).

*Eridacnis sinuans*: Bass *et al.* 1975\*; Compagno 1984\*, 1988\*; SSF No. 10.3\*; Compagno *et al.* 1989\*; Ebert 2013; Ebert *et al.* 2013\*.

Preoral length more than twice mouth length; mouth triangular, lip furrows very short. Lateral dermal denticles broad-crowned and with short broad cusps. Dorsal fins subequal, moderately large and high; 1st dorsal-fin anterior margin at high angle to body axis. Anal-fin height <½ height of dorsal fins. Caudal fin narrow, long and ribbon-like; junction of caudal-fin pre- and post-ventral margins broadly rounded. Tooth count typically 64/50; vertebrae 132–144; spiral valve turns 7.

Body brownish grey, with pale margins on dorsal fins, and faint dark banding on caudal fin. Attains 37 cm TL.

**DISTRIBUTION** WIO: Tanzania to South Africa (KwaZulu-Natal).

**REMARKS** Litters of 2; size at birth 15–17 cm TL. Males mature at 29–30 cm TL, and females at ~37 cm TL. Bottom-dwelling in warm-temperate and tropical waters of upper continental slope and outer shelf, at 180–500 m. Apparent geographic or bathymetric segregation of populations by sex occurs, as most specimens taken off South Africa (KwaZulu-Natal) are males. Prey includes small bony fishes, crustaceans and cephalopods. Of no importance to fisheries but sometimes taken as bycatch of bottom-trawl fisheries in parts of its range.



*Eridacnis sinuans*, 28 cm TL, immature male (South Africa). Source: Bass *et al.* 1975

# FAMILY **PSEUDOTRIAKIDAE**

False catsharks David A Ebert

Distinctive small- to large-sized with slender body, narrowly rounded, depressed head, with more or less elongated bellshaped snout, and deep groove in front of elongated slit-like eyes. Nictitans rudimentary; spiracles large. Anterior nasal flaps broadly angular and not barbel-like; internarial space >1.5 times nostril width. Mouth long, angularly arched, and extending posteriorly to below spiracles; no papillae inside mouth or on edges of gill arches; lip folds short and always present. Teeth small, similar in both jaws, with acute narrow cusps, lateral cusplets, and strong basal ledges and grooves, and not blade-like; posterior teeth comb-like; tooth rows numerous, 94-294/81-335. Pectoral fins with radials confined to bases. First dorsal fin elongated, base closer to pectoralfin bases than to pelvic-fin bases. Caudal-fin lower lobe weak or absent; no undulations or ripples on dorsal margin; no precaudal pits. Vertebrae 140-186; spiral valve turns 11-17. Body usually grey to brown or blackish.

Small group of poorly known deepwater sharks, found on outer continental and insular shelves and slopes, on or near bottom, at ~130–1 890 m. At least two species are oophagous. Three genera and at least 5 species; 2 genera and at least 2 species in WIO, but only 1 of these at depths of ~200 m. The pygmy false catshark *Planonasus parini* Weigmann, Stehmann & Thiel 2013, was described from off Socotra I. (Arabian Sea), collected at 1 000–1 120 m.

# GENUS **Pseudotriakis** De Brito Capello 1868

Large-sized, bulky and soft-bodied, with short snout and wide angular mouth. Anterior nasal flaps broadly angular, not barbel-like. Teeth numerous, small, cuspidate, with >200 rows

in each jaw. First dorsal fin long, low and keel-like; 2nd dorsal fin higher than 1st; anal fin small; caudal-fin lower lobe weak, no undulations on dorsal margin, and no precaudal pits. One species.

#### Pseudotriakis microdon De Brito Capello 1868

False catshark

PLATE 35

*Pseudotriakis microdon* De Brito Capello 1868: 316, Pl. 5, Fig. 1 (Setúbal, Portugal); Forster *et al.* 1970; Compagno 1984\*; SSF No. 12.1\*; Compagno *et al.* 2005\*; Ebert 2013\*; Ebert *et al.* 2013\*.

Diagnosis as for genus. Vertebrae 180–186; spiral valve turns 17.

Body dark brown to blackish, with darker fins. Attains ~3 m TL.

**DISTRIBUTION** All seas (patchy) except southern Atlantic and eastern Pacific. WIO: Aldabra and Seychelles.

**REMARKS** Poorly known; exhibits a unique form of oophagy: in early development the embryos are nourished by a yolk sac, but oophagy commences as development proceeds. Litters of 2, possibly more; smallest free-swimming specimen measured 96 cm TL. Large body cavity, soft fins, and soft skin and musculature suggest it is relatively inactive and sluggish and can hover over the bottom at virtually neutral buoyancy. Wide-ranging, with scattered records; found on continental and insular slopes at 200-1 890 m, but occasionally wandering onto continental shelf and into shallower water; seemingly uncommon to rare wherever it occurs. Diet consists mostly of fishes, including other elasmobranchs, and cephalopods and crustaceans: small, sharp, cuspidate teeth, combined with very large mouth may allow it to capture considerably large prey items. Interest to fisheries minimal, taken incidentally in surveys and in fisheries by deep longlines and less commonly bottom trawls, but likely not impacted.

Pseudotriakis microdon, 93 cm TL, immature specimen (Atlantic). Drawn from Bigelow & Schroeder 1948

# FAMILY TRIAKIDAE

#### Houndsharks

Kelsey C James and David A Ebert

Small- to medium-sized with fusiform body. Eyes elongated and horizontally oval, or slit-like; nictitans external, transitional or internal; spiracles small to large. Anterior nasal flaps elongated to lobe-like or vestigial; lip folds moderate to very long. Teeth small to moderately large, with acute and narrow to moderately large cusps, and with lateral cusplets in some species but these structures reduced or absent in others; cuspidate and not blade-like or compressed, or else blade-like or thickened and molariform; teeth usually similar in both jaws, but differentiated in a few species; tooth counts 18-42/27-106. Two medium to large spineless dorsal fins; 1st dorsal fin large to very large but not keel-like, usually much shorter than caudal fin, and its base well ahead of pelvic-fin bases, usually closest to pectoral-fin bases. Caudalfin lower lobe well-developed to absent; no undulations on dorsal margin, and no precaudal pits. Spiral valve turns 4-11. Some species with spots or variegated patterns, and others uniformly coloured.

Reproductive mode viviparity with yolk sac or yolk-sac placenta, with litter sizes ranging from 1 or 2 to 52. Many are fairly strong swimmers, occurring in small to large schools or singly, and others rest on the bottom. Feed primarily on bottom-dwelling and midwater bony fishes and invertebrates, with some species taking mainly fishes, others largely crustaceans, and a few primarily cephalopods. Houndshark genera that are especially important to fisheries include Galeorhinus, Iago, Mustelus and Triakis; all these are fairly common to very abundant in coastal waters where they occur but are relatively small-sized, and hence important mostly to small commercial and artisanal fisheries which are limited in their gear and movements, as well as to sports fisheries; used primarily for their meat, but also for liver oil, fishmeal, and in the shark-fin trade. None of the species are dangerous to people.

Although a motley assemblage, one of the largest families of sharks. Occur in warm- to cold-temperate seas, most often on continental and insular shelves, from shoreline and intertidal areas, to outermost shelf, often close to bottom; a few deepwater species occur on continental slope to considerable depths, possibly >2 000 m. None are coral-reef specialists or oceanic, but mainly inhabit sandy, muddy or rocky inshore areas, including enclosed bays. A few species have been recorded from river mouths, but apparently none can tolerate freshwater for an extended period. Nine genera and at least 47 species, plus many undescribed species; 6 genera and 9 species in WIO.

#### **KEY TO GENERA**



- **1b** Anterior nasal flaps moderately large to very small, wellseparated and not reaching mouth; no oronasal grooves .....**2**



- 3b Mouth broadly arched; 2nd dorsal fin subequal to anal fin; caudal-fin terminal lobe ~½ length of caudal-fin dorsal margin.



- 4a Eyes lateral, subocular ridges rudimentary; 1st dorsal-fin origin anteriorly set, over pectoral-fin bases ...... *lago*
- 4b Eyes dorsolateral, subocular ridges strong; 1st dorsal-fin origin posteriorly set, over or behind pectoral-fin inner margins ..... 5



- 5b Snout parabolic to subangular in dorsoventral view; mouth angular, lower jaw with straight or nearly straight edges



## GENUS Galeorhinus Blainville 1816

Slender-bodied with snout moderately long and parabolic in dorsoventral view, preoral length about equal to mouth width. Eyes horizontally oval and lateral; subocular ridges obsolete. Anterior nasal flaps vestigial, formed as small, low, angular points; no oronasal grooves. Mouth broadly arched and long; lip folds moderately long, upper furrows ending well behind level of upper symphysis. Teeth blade-like, compressed and cuspidate, similar in both jaws; anteroposterior teeth with oblique cusps and cusplets; medial teeth well-differentiated from anteroposterior teeth. First dorsal fin moderately large, its origin over or slightly behind pectoral-fin free rear tips; 2nd dorsal fin much smaller, <½ height of 1st. Anal fin about equal to 2nd dorsal fin. Caudal-fin lower lobe strong; terminal lobe very long, ~2 times into caudal-fin dorsal margin. One species.

## Galeorhinus galeus (Linnaeus 1758)

Tope

PLATES 36 & 38

Squalus galeus Linnaeus 1758: 234 (Mediterranean Sea and NE Atlantic). Galeorhinus galeus: Bass et al. 1975\*; SSF No. 9.20\*; Compagno et al. 1989\*;

Compagno et al. 1991; Heemstra & Heemstra 2004\*; Compagno et al. 2005\*; Ebert et al. 2013\*.

Diagnosis as for genus. Nictitans internal; no interdorsal ridge. Tooth count 32–46/31–38; vertebrae 123–146; spiral valve turns 4–6.

Body greyish above, white below; fins plain, but with black markings in young. Attains 195 cm TL.

**DISTRIBUTION** Worldwide in temperate seas, including Mediterranean Sea. WIO: South Africa (Western Cape and Eastern Cape).

**REMARKS** Viviparous with yolk sac; litters of 6–52; size at birth 30–40 cm TL, with pups born in late spring and early summer in enclosed bays and estuaries. Males mature at 120–170 cm TL, at ~8 years; females mature at 130–185 cm TL, at ~11 years; maximum age possibly 60 years. Coastal-pelagic, in temperate continental and insular waters, often near bottom, at 2–471 m; strong-swimming and seasonally migratory. Feeds on large variety of demersal bony fishes and midwater schooling fishes, also cephalopods. High fisheries interest throughout its distribution, with its meat excellent for human consumption; the principal target species of the South African directed shark fishery. Different stocks may occur worldwide. IUCN Red List conservation status Critically Endangered.

# GENUS Hypogaleus Smith 1957

Slender-bodied with snout moderately long, broadly pointed and parabolic in dorsoventral view; preoral length about equal to mouth width. Eyes horizontally oval and lateral; subocular ridges obsolete. Anterior nasal flaps vestigial, well separated from each other and mouth; no oronasal grooves. Mouth angular and long; lip folds moderately long, upper furrows ending well behind level of upper symphysis. Teeth blade-like, compressed and cuspidate, similar in both jaws; anteroposterior teeth with oblique cusps and cusplets; medial teeth well differentiated from anteroposterior teeth. First dorsal fin moderately large, its origin over or slightly behind pectoralfins free rear tips; 2nd dorsal fin smaller than 1st. Anal fin smaller than 2nd dorsal fin. Caudal-fin lower lobe strong, and terminal lobe relatively short. One species.



Galeorhinus galeus, 134 cm TL, mature male (South Africa). Source: Bass et al. 1975


# Hypogaleus hyugaensis (Miyosi 1939)

#### Blacktip tope

PLATE 38

*Eugaleus hyugaensis* Miyosi 1939: 91, Fig. 1 (off Hyūga Nada, Miyazaki Prefecture, Japan).

Hypogaleus zanzibarensis: Smith 1957\*.

Hypogaleus hyugaensis: Bass et al. 1975\*; SSF No. 9.22\*;

Compagno et al. 1989\*; Compagno et al. 2005\*; Ebert et al. 2013\*.

Diagnosis as for genus. Nictitans internal; no interdorsal ridge. Tooth count ~50/44.

Body bronzy to grey-brown above, paler below; dorsal and caudal fins with dusky tips (especially in young). Attains 130 cm TL.

**DISTRIBUTION** Indo-Pacific (patchy). WIO: Persian/Arabian Gulf, South Africa (KwaZulu-Natal) to Tanzania (Zanzibar) and Kenya.

**REMARKS** Viviparous with yolk-sac placenta; litters of 2–15 (average ~10); size at birth 33–35 cm TL; gestation period <12 months. Matures at ~95 cm TL. Found in relatively deep tropical and warm-temperate seas, along continental shelf and upper slope, at 40–230 m. Diet includes bony fishes and cephalopods. A bycatch of numerous fisheries, especially the trawl fisheries in WIO region.

GENUS lago Compagno & Springer 1971

Slender-bodied with snout moderately long and subangular in dorsoventral view. Eyes horizontally oval; vestigial lateral subocular ridges. Anterior nasal flaps formed as low, rounded or angular lobes, well-separated from each other and from mouth; no oronasal grooves. Mouth angular and moderately long; lip folds short to moderately long. Teeth blade-like, compressed and cuspidate; anteroposterior teeth with oblique cusps and cusplets or blades; medial teeth well-differentiated from anteroposterior teeth. First dorsal fin moderately large, anteriorly set, its origin over pectoral-fin bases; 2nd dorsal fin somewhat smaller than 1st. Anal fin considerably smaller than 2nd dorsal fin. At least 2 species, 1 in WIO.

## lago omanensis (Norman 1939)

### Bigeye houndshark

PLATE 38

*Eugaleus omanensis* Norman 1939: 11, Fig. 3 (Gulf of Oman). *Iago omanensis*: Compagno & Springer 1971\*; Compagno 1984\*; Compagno *et al.* 2005\*; Ebert 2013\*; Ebert *et al.* 2013\*.

Snout moderately broad; gill slits large. Eyes large, length nearly equal to longest gill slit. Lip folds relatively short, with upper furrows falling well behind level of lower symphysis.



*lago omanensis*, ~40 cm TL (Red Sea).

Body brownish or greyish above, paler below; no conspicuous markings, but dorsal-fin margins sometimes darker. Attains 58 cm TL.

**DISTRIBUTION** WIO: Red Sea, Gulf of Oman, and Pakistan to southwestern India.

**REMARKS** Viviparous with yolk-sac placenta; litters of 2–10; size at birth ~17 cm TL; females attain larger sizes than males. Found in deep, tropical waters of continental shelf and slope, at 110–1 000 m or more (particularly in Red Sea); tolerates warmer, oxygen-poor conditions. Segregates by sex, with females occurring in shallower waters. Feeds primarily on bony fishes (especially lanternfishes) and cephalopods. Limited fisheries interest; taken in gillnet fisheries in India and in small-scale and longer-range fisheries in Red Sea.

## GENUS Mustelus Linck 1790

Slender-bodied with long, parabolic, subangular snout. Eyes horizontally elongated or oval and dorsolateral; subocular ridges strong. Anterior nasal flaps elongated and lobate, wellseparated from each other and mouth; no oronasal grooves. Mouth angular and moderately long; lip folds moderately long, upper folds ending well behind level of upper-jaw symphysis. Teeth forming a pavement, similar in both jaws, but varying from compressed and with short erect cusps and cusplets, to rounded, molariform and without cusps and cusplets. First dorsal fin moderately large, its origin over pectoral-fin inner margins or slightly behind free rear tips; 2nd dorsal fin subequal to 1st. Anal fin much smaller than 2nd dorsal fin. Caudal-fin lower lobe poorly developed to short and strong. About 27 species, 4 in WIO.

#### **KEY TO SPECIES**

1a 1b	Small white spots on dorsal surface and flanks
2a 2b	Pectoral fins larger, rear margin 12–16% TL
3a	Denticles on tongue extend to level of 1st gill-arch lower limb; 2nd dorsal-fin base to caudal-fin origin 1.7–2 times length of caudal-fin upper lobe; precaudal vertebrae 62–76
3b	Denticles on tongue not extending to level of 1st gill-arch lower limb; 2nd dorsal-fin base to caudal-fin origin 1.9–2.4 times length of caudal-fin upper lobe; precaudal vertebrae 81–95

## Mustelus manazo Bleeker 1854

Starspotted smoothhound

PLATE 36

Mustelus manazo Bleeker 1854: 126 (Nagasaki, Japan); Compagno et al. 2005\*; Ebert et al. 2013\*.

Medium-sized with slender body, short head, and snout moderately long and bluntly angular in lateral view; internarial space narrow. Eyes large; interorbital space narrow. Mouth short, upper lip folds longer than lower folds. Teeth molariform and asymmetric, with cusps reduced to low point. Paired fins moderately sized. Trailing edges of dorsal fins denticulate, without bare ceratotrichia. First dorsal fin broadly triangular, with posteroventrally sloping rear margin, and midbase closer to pelvic-fin bases than to pectoral-fin bases. Anal-fin to caudal-fin space greater than 2nd dorsal-fin height. Caudal-fin lower lobe not falcate in adults. Vertebrae 129–142.

Body grey or grey-brown above, paler below; usually many white spots on dorsal surface, but no dark spots or bars. Attains 117 cm TL.



**DISTRIBUTION** Northwestern and western central Pacific and Indian oceans. WIO: Kenya.

**REMARKS** Viviparous with yolk sac; litters of 1–22 (mostly 2–6), varying with geographic region and mother's size; size at birth 20–35 cm TL; gestation period 10–12 months. Matures at 60–80 cm TL, depending on region. Wide-ranging, but reports may represent more than one species. Occurs in temperate to tropical continental waters, from intertidal and subtidal areas, and commonly close inshore, especially over mud or sand bottoms. Feeds on bottom-dwelling crustaceans, fishes, polychaetes and sipunculid worms. Considerable fisheries interest, especially off Japan; used for its meat and oil.

### Mustelus mosis Hemprich & Ehrenberg 1899

### Hardnose smoothhound

#### PLATES 36 & 38

Mustelus mosis Hemprich & Ehrenberg 1899: 8, Pl. 7, Fig. 3a–d (Red Sea); Van der Elst 1981\*; SSF No. 9.26\*; Compagno *et al.* 1989\*; Fricke 1999; Bonfil & Abdallah 2004; Heemstra & Heemstra 2004; Compagno *et al.* 2005\*; Ebert *et al.* 2013\*.

Large-sized with fairly slender body, short head, and snout moderately long, bluntly angular in lateral view, with expanded bulbous tip in adults (hypercalcified rostral cartilages). Eyes large; interorbital space narrow. Internarial space broad. Mouth moderately long; upper lip folds about equal to lower folds. Teeth molariform and asymmetric, with cusp reduced to low point. Trailing edges of dorsal fins without bare ceratotrichia. First dorsal fin somewhat falcate, with rear margin nearly vertical from apex, midbase closer to pectoral-fin bases than to pelvic-fin bases. Pectoral fins medium-sized. Pelvic fins small. Anal-fin to caudal-fin space greater or subequal to 2nd dorsalfin height. Caudal-fin lower lobe more or less falcate in adults. Vertebrae 122. Body tan to greyish-tan above, paler below; no white or dark spots or dark bars, but South African specimens generally have white-tipped 1st dorsal fin and black-tipped 2nd dorsal fin and caudal fin. Attains 150 cm TL.

**DISTRIBUTION** Indian Ocean. WIO: South Africa (KwaZulu-Natal) to Red Sea, Persian/Arabian Gulf, Pakistan, India, Mascarenes, Maldives and Sri Lanka; elsewhere to east coast of India and Bay of Bengal.

**REMARKS** Viviparous with yolk-sac placenta; litters of 6–10. Males mature at 63–67 cm TL. Occurs in continental waters and common in its range; bottom-dwelling, found inshore and offshore, occasionally on coral reefs. Feeds mostly on small bottom-dwelling fishes, molluscs and crustaceans. Caught in multiple gears; targeted in fisheries off Pakistan and India.

### Mustelus mustelus (Linnaeus 1758)

### Smoothhound

Squalus mustelus Linnaeus 1758: 235 (Mediterranean Sea).

*Mustelus mustelus*: SSF No. 9.27\*; Compagno *et al.* 1989\*; Compagno *et al.* 1991; Heemstra & Heemstra 2004\*; Compagno *et al.* 2005\*; Ebert *et al.* 2013\*.

Large-sized with short slender head, and snout moderately long and bluntly angular in lateral view. Eyes fairly large; interorbital space narrow. Internarial space broad. Mouth fairly short; upper lip folds usually slightly longer than lower furrows. Teeth molariform and asymmetric, with cusps reduced to low point; cusplets absent except in very young individuals. Buccopharyngial denticles confined to tongue tip and extreme anterior end of palate. Crowns of lateral dermal denticles lanceolate or weakly tricuspidate, with longitudinal ridges extending their entire length. Paired fins moderately large. Trailing edges of dorsal fins denticulate, without



PLATE 38

bare ceratotrichia. First dorsal fin falcate, with rear margin almost vertical, midbase closer to pectoral-fin bases than to pelvic-fin bases. Caudal-fin lower lobe more or less falcate. Vertebrae 131–137.

Body grey to grey-brown above, paler below; no white spots or dark bars, but occasionally few irregular, small dark spots. Attains 164 cm TL.

**DISTRIBUTION** North Sea, Mediterranean Sea, eastern Atlantic (British Isles to West Africa), to South Africa (KwaZulu-Natal) in WIO.

**REMARKS** Viviparous with yolk-sac placenta; litters of 4–15; size at birth ~39 cm TL; gestation period 10–11 months. Males mature at 70–74 cm TL, and females at ~80 cm TL. Occurs on continental shelf and uppermost slope, most commonly in shallow depths of ~5–50 m, but also intertidal areas and ranging to at least 350 m deep; swims near bottom but sometimes in midwater. Feeds primarily on crustaceans, but also cephalopods and bony fishes. Considerable fisheries interest in European waters, Mediterranean Sea and off West Africa; taken in bottom trawls, fixed bottom nets and with line gear; used for human consumption. IUCN Red List conservation status Vulnerable.

# Mustelus palumbes Smith 1957

### Whitespotted smoothhound

PLATES 37 & 38

Mustelus palumbes Smith 1957: 358, Fig. 1e–f (Knysna Estuary, Western Cape, South Africa); SSF No. 9.28\*; Compagno *et al.* 1989\*; Compagno *et al.* 1991; Heemstra & Heemstra 2004; Compagno *et al.* 2005\*; Ebert *et al.* 2013\*. Slender-bodied with short head, and snout moderately long and bluntly angular in lateral view. Eyes fairly large; interorbital space moderately narrow. Internarial space broad. Mouth short, subequal to eye length; upper lip folds longer than lower folds. Teeth molariform and asymmetric, with cusps reduced to low point. Buccopharyngial denticles covering almost entire palate and floor of mouth. Trailing edges of dorsal fins denticulate, without bare ceratotrichia. Interdorsal space moderately long; 1st dorsal fin broadly triangular, with posteroventrally sloping rear margin, and midbase closer to pectoral-fin bases than to pelvic-fin bases. Pectoral fins large. Pelvic fins high and broadly rounded. Anal-fin to caudal-fin space greater than 2nd dorsal-fin height. Caudal-fin lower lobe somewhat expanded and sometimes semifalcate.

Body grey or grey-brown above, paler below; numerous small white spots, distinct or faint. Attains 120 cm TL.

**DISTRIBUTION** Southern Africa: Namibia in southeastern Atlantic, to South Africa and southern Mozambique in WIO.

**REMARKS** Viviparous with yolk sac; litters of 3–8. Males mature at 76–88 cm TL, and females at 79–102 cm TL. Occurs in temperate seas, on continental shelf and upper slope, inshore and offshore, on or near bottom, from intertidal to at least 360 m deep; prefers sand or gravel bottoms. Feeds on crabs and probably other invertebrates. Limited fisheries interest, often taken by sports anglers and as bycatch of commercial bottom trawls.



Mustelus palumbes, 88 cm TL, mature male holotype, ventral head; lateral view of 99-cm-TL specimen (both South Africa). Source: SSF

# GENUS Scylliogaleus Boulenger 1902

Medium-sized with short and broadly rounded snout. Eyes horizontally elongated with strong subocular ridges. Anterior nasal flaps triangular and overlapping mouth; oronasal grooves broad and shallow; internarial width half nostril width. Mouth broadly arched, short, with long lip folds. Teeth small, bluntcrowned, molariform and without cusps and cusplets. First dorsal fin moderately large, its origin over pectoral-fin inner margins or free rear tips; 2nd dorsal fin as large as 1st, its height at least three-quarters as high as 1st dorsal fin. Anal fin smaller than 2nd dorsal fin. One species.

### Scylliogaleus quecketti Boulenger 1902

### Flapnose houndshark

PLATE 37

Scylliogaleus quecketti Boulenger 1902: 51, Pl. 4 (off Umkomaas, KwaZulu-Natal, South Africa); Bass et al. 1975\*; SSF No. 9.34\*; Compagno et al. 1989\*; Compagno et al. 2005\*; Ebert et al. 2013\*.

Diagnosis as for genus. Distinguished by large fused nasal flaps expanded over mouth; nictitans transitional. Interdorsal ridge present. Caudal-fin lower lobe very short in adults, barely developed in young; caudal-fin terminal lobe moderately long. Tooth count 51/50; vertebrae 139–149; spiral valve turns 7.

Body grey above, paler below; fins plain in adults, but median fins with white rear edges in newborns. Attains 102 cm TL.

# **DISTRIBUTION** WIO: endemic to South Africa (Eastern Cape and KwaZulu-Natal).

**REMARKS** Little known and uncommon; occurs in warmtemperate or subtropical waters of continental shelf, inshore, from surfline and intertidal areas to close offshore. Viviparous, but presence or absence of placenta unknown; litters of 2–4; size at birth ~34 cm TL; gestation period 9–10 months. Feeds primarily on crustaceans, also squid. Minimal fisheries interest; occasionally caught by shore anglers. IUCN Red List conservation status Vulnerable.

### GENUS Triakis Müller & Henle 1838

Stout-bodied with short, broadly rounded snout. Eyes horizontally elongated with strong subocular ridges. Anterior nasal flaps elongated and lobate; no oronasal grooves. Mouth broadly arched and moderately short; lip folds long, upper furrows nearly reaching level of upper symphysis. Teeth semiblade-like, somewhat compressed, with erect or oblique cusps variably developed, and similar in both jaws; medial teeth not differentiated from anteroposterior teeth. First dorsal fin moderately large, its origin over pectoral-fin inner margins; 2nd dorsal fin nearly as large as 1st. Anal fin smaller than 2nd dorsal fin. Five species, 1 in WIO.



Scylliogaleus quecketti, 94 cm TL, mature female (South Africa). Source: Bass et al. 1975

# Triakis megalopterus (Smith 1839)

### Spotted gullyshark

PLATES 37 & 38

Mustelus megalopterus Smith 1839: 4, Pl. 2 (Cape Town, South Africa). Mustelus natalensis: Steindachner 1866\*; Smith 1957\*. Mustelus nigropunctatus: Smith 1952\*.

*Triakis megalopterus*: Bass *et al.* 1975\*; Van der Elst 1981\*; SSF No. 9.36\*; Compagno *et al.* 1989\*; Heemstra & Heemstra 2004\*; Compagno *et al.* 2005\*; Ebert *et al.* 2013\*.

Anterior nasal flaps lobate, not reaching mouth and far separated from each other; upper lip folds reaching level of lower symphysis of mouth. Teeth with strong erect cusps on at least the middle two-thirds of dental band, cusplets low or absent on almost all teeth, semi-molariform but not blade-like. Unusually high interdorsal ridge; 1st dorsal fin with abrupt vertical rear margin. Pectoral fins broadly falcate in adults. Caudal-fin lower lobe barely developed in young, but short and strong in adults; caudal-fin terminal lobe moderately long. Tooth count 48–52/44; vertebrae 162–166; spiral valve turns 6.

Body grey, with few to numerous small black spots in

adults, and few or no spots in young, although plain adults have been recorded. Attains 208 cm TL.

**DISTRIBUTION** Southern Africa: southern Angola in southeastern Atlantic, to South Africa (Cape Peninsula and eastwards, but rarely to southern KwaZulu-Natal) in WIO.

**REMARKS** Viviparous with yolk sac; litters of 5–15; size at birth 40–45 cm TL; gestation may last 19–21 months, with reproductive cycle of 2 or 3 years, depending on time between pregnancies. Males mature at 125–135 cm TL, and females at 145 cm TL. Inshore, bottom-dwelling, in temperate continental waters, usually in shallow depths to surfline. Prefers rocky reefs at <10 m. Congregates in schools in summer (False Bay, South Africa). Diet changes with growth: small sharks <100 cm consume primarily Cape rock crabs (*Plagusia chabrus*), sharks 100–140 cm consume cephalopods in addition to Cape rock crabs, and larger sharks feed mostly on bony fishes. Hardy and keeps well in captivity. Limited fisheries interest; commonly caught by shore anglers.



Triakis megalopterus, 145 cm TL, mature female (South Africa). Source: Bass et al. 1975

# FAMILY HEMIGALEIDAE

### Weasel sharks

James DS Knuckey and David A Ebert

Small- to moderate-sized, with horizontally oval eyes, internal nictitans and small spiracles. Mouth short to long, arched and reaching past anterior edge of eyes; lip folds moderately long; no oronasal grooves or barbels. Teeth not compressed; upper teeth small to moderately large and blade-like, lower teeth more or less cuspidate. Two moderately-sized spineless dorsal fins; 1st dorsal fin much shorter than caudal fin, its base well ahead of pelvic-fin bases; 2nd dorsal fin ~<sup>2</sup>/<sub>3</sub> size of 1st. Anal fin subequal to 2nd dorsal fin. Caudal-fin lower lobe well-developed, acute; caudal-fin dorsal margin with lateral undulations or ripples; precaudal pits present. Spiral valve turns 4–6.

Reproductive mode viviparity with yolk-sac placenta. Commonly occurring in tropical coastal waters, over continental and insular shelves, to ~100 m deep but usually shallower. Of modest importance to artisanal and small commercial inshore and near offshore fisheries where they occur, and used mostly for their meat for human consumption, although larger species are used for their liver oil and in the shark-fin trade.

Limited to the eastern Atlantic and continental Indo-Pacific, and not far into the central Pacific. Four genera and ~8 species; all genera and 5 species in WIO.

#### **KEY TO GENERA**

1a	Lower anterolateral teeth with long hooked cusps; lower teeth protrude prominently when mouth closed
1b	Lower anterolateral teeth with short, straight or weakly hooked cusps; lower teeth concealed or protrude only slightly when mouth closed
2a	Snout obtusely wedge-shaped in dorsoventral view; lower jaw rounded at symphysis; diastema at midline of both jaws; mesial edges of teeth smooth, sometimes a few cusplets on mesial edges of lower teeth; pectoral fins not falcate; rear margins of 2nd dorsal fin and anal fin moderately concave;

Continued ....

#### **KEY TO GENERA**

# GENUS Chaenogaleus Gill 1862

Snout moderately long and wedge-shaped in dorsoventral view. Gill slits long, about twice or more eye length. Mouth parabolic and very long, its length 66–82% width; lower teeth protrude prominently when mouth closed; ends of upper lip folds behind rear edge of eyes; lower jaw rounded at symphysis. No diastema at midline of jaws. Upper anterolateral teeth with smooth mesial edges and very long cusps; lower anterolateral teeth with very long, stout, strongly hooked cusps, and no cusplets; lower crown feet and roots deeply arched, giving teeth inverted Y-shape. Second dorsal fin at least three-fifths height of 1st dorsal fin. Pectoral fins not falcate. One species.

# Chaenogaleus macrostoma (Bleeker 1852)

Hooktooth shark

PLATE 38

Hemigaleus macrostoma Bleeker 1852: 46, Pl. 2, Fig. 10 (Coromandel coast, India).

Chaenogaleus macrostoma: Compagno et al. 2005\*; Ebert et al. 2013\*.

Diagnosis as for genus. Tooth count 33–38/34–36; vertebrae 131–139; spiral valve turns 5 or 6.

Body pale grey or bronzy, and without prominent markings, but sometimes black-tipped 2nd dorsal fin and caudal-fin terminal lobe. Attains ~100 cm TL.



Chaenogaleus macrostoma (Oman). © M Harris

**DISTRIBUTION** Indo-Pacific. WIO: Persian/Arabian Gulf to Pakistan, India and Sri Lanka.

**REMARKS** Common but little known; found in tropical inshore waters of continental and insular shelves, to at least 59 m deep. Litters of 4; size at birth at least 20 cm TL; males mature at 68–97 cm TL, and females at ~86 cm TL. Probably feeds on small fishes, cephalopods and crustaceans. Commonly landed in inshore and offshore artisanal fisheries, with drifting bottom gillnets, longlines and other line gear; meat used fresh for human consumption, and offal processed into fishmeal. IUCN Red List conservation status Vulnerable.

# GENUS Hemigaleus Bleeker 1852

Snout rounded in dorsoventral view. Gill slits short. Mouth broadly arched and very short, its length 31–43% width; lower teeth not protruding when mouth closed; ends of upper lip folds extend below eyes; lower jaw rounded at symphysis. No diastema at midline of jaws. Upper anterolateral teeth with smooth mesial edges and very short cusps; lower anterolateral teeth with short, slender, unhooked cusps and no cusplets; lower crown feet and roots deeply arched, giving teeth inverted Y-shape; tooth counts 25–34/37–52. Second dorsal fin 0.4–0.6 times height of 1st dorsal fin. Dorsal fins, pectoral fins, pelvic fins and caudal-fin lower lobe strongly falcate. Vertebrae 112–150. Body pale grey to brown or bronzy dorsally, paler to white ventrally. Two species, 1 in WIO.

# Hemigaleus microstoma Bleeker 1852

Sicklefin weasel shark

PLATE 39

*Hemigaleus microstoma* Bleeker 1852: 46, Pl. 2, Fig. 6 (Jakarta, Java, Indonesia); Compagno *et al.* 2005\*; Ebert *et al.* 2013\*.

Small-sized with rounded, moderately long snout. Gill slits short, ~1.3 times eye length in adults. Mouth very short, arched, with teeth not protruding. Upper teeth with distal cusplets but no serrations; lower teeth inverted Y-shape, with short, straight, smooth-edged cusps, no cusplets, and highly arched roots. Fins distinctly falcate. Second dorsal fin about two-thirds size of 1st dorsal fin. Anal fin smaller than 2nd dorsal fin; no preanal ridges. Precaudal pits crescentic. Tooth count 25–34/37–43; vertebrae 137–150; spiral valve turns 4–6.

Body pale grey or bronzy dorsally, paler ventrally; sometimes with pale dorsal-fin margins and tips, and white spots on sides. Attains 111 cm TL.

**DISTRIBUTION** Indo-Pacific. WIO: Red Sea, and southern India to Sri Lanka.

**REMARKS** Appears naturally uncommon; found in tropical inshore waters of continental shelf, on or near bottom, at 12–167 m. Litters of 4–14; size at birth 26–28 cm TL; males mature at 72–91 cm TL, and females at ~85 cm TL. Specialist feeder on cephalopods, but also crustaceans and echinoderms. Taken regularly in inshore artisanal fisheries in gillnets and probably on line gear; meat used fresh for human consumption. IUCN Red List conservation status Vulnerable.

# GENUS Hemipristis Agassiz 1843

Moderately large-sized and fairly slender, with snout broadly rounded in dorsoventral view. Gill slits very long. Mouth trapezoidal-parabolic and long, its length 50–70% width; lower teeth protrude prominently when mouth closed; ends of upper lip folds behind rear edge of eyes; lower jaw truncated at symphysis. Diastema at midline of both jaws. Upper anterolateral teeth with short cusps and serrated mesial edges (but smooth edges in young); lower anterolateral teeth with very long, stout, strongly hooked cusps, and serrations and cusplets variably developed on crown feet; lower crown feet and roots deeply arched, giving teeth inverted Y-shape. Second dorsal fin <sup>2</sup>/<sub>5</sub>–<sup>3</sup>/<sub>5</sub> height of 1st dorsal fin. Pectoral fins strongly falcate. Rear margins of 2nd dorsal fin, pectoral fins, pelvic fins and anal fin deeply concave. Precaudal pits crescentic, transverse. One species.



*Hemigaleus microstoma*, 90 cm TL.

© Food and Agriculture Organization of the United Nations, Original Scientific Illustrations Archive; reproduced with permission



Hemipristis elongata, 137 cm TL, immature female; ventral head (Tanzania). Source: Bass et al. 1975 (lateral view); © NRF-SAIAB (head)

# Hemipristis elongata (Klunzinger 1871)

### Snaggletooth shark

PLATE 39

*Dirrhizodon elongatus* Klunzinger 1871: 665 (Al-Qusayr, Egypt, Red Sea). *Hemipristis elongatus*: Bass *et al.* 1975\*; Compagno *et al.* 1989\*;

Compagno et al. 2005\*; Ebert et al. 2013\*.

Hemipristis elongata: SSF No. 9.21\*.

Diagnosis as for genus. Tooth count 26-30/30-36.

Body bronzy to pale grey-brown above, paler below; no prominent markings, but sometimes tips of pectoral fins, pelvic fins, anal fin and caudal fin prominently white, and 2nd dorsal fin, anal fin, and caudal-fin terminal lobe with dusky blotch, especially in juveniles. Attains 240 cm TL.

**DISTRIBUTION** Indo-Pacific. WIO: Persian/Arabian Gulf, Pakistan to India, Red Sea to South Africa (KwaZulu-Natal), Madagascar and Maldives.

**REMARKS** Found in tropical coastal waters, inshore and offshore on continental and insular shelves, at 1–132 m. Litters of 2–11, with larger females having larger litters; size at birth 45–52 cm TL; males mature at 110–145 cm TL, and females at 110–170 cm TL. Feeds on prawns, shrimp, cephalopods and a variety of fishes. Moderate fisheries interest and a common catch in some areas; regularly taken in artisanal and commercial fisheries, with floating and fixed bottom gillnets, floating longlines, trawls and probably on hook and line; used fresh for its meat, the liver processed for vitamin oil, the fins in the shark-fin trade, and offal and spoiled carcasses for fishmeal. IUCN Red List conservation status Vulnerable.

# GENUS Paragaleus Budker 1935

Snout rounded or slightly pointed in dorsoventral view. Gill slits short. Mouth broadly arched or parabolic and short to moderately long, its length 44-64% width; lower teeth not protruding or else slightly protruding when mouth closed; ends of upper lip folds reach below eyes; lower jaw rounded at symphysis. No diastema at midline of jaws. Upper anterolateral teeth with smooth mesial edges and moderately long cusps; lower anterolateral teeth with short, fairly stout, unhooked or slightly hooked cusps with distal cusplets; roots and crown feet hardly arched, giving teeth inverted T-shape; tooth counts 26-30/27-33. Pectoral fins narrow and falcate. Interdorsal ridge present; 2nd dorsal fin at least three-fifths height of 1st dorsal fin. Body pale to dark grey or grey-brown above, whitish below; some species variously with paired blackish marks or dusky blotches on underside of snout, longitudinal bands, or fins with pale rear margins. Four species, 2 in WIO.

#### **KEY TO SPECIES**

- 1a
   Five or more rows of lower anterolateral teeth with distal cusplets; lower anterior teeth mostly with oblique cusps
   *P. randalli P. randalli P. randalli P. randalli*
- 1b Two or 3 rows of lower anterolateral teeth with distal cusplets; lower anterior teeth mostly with erect cusps.......*P. leucolomatus*

# Paragaleus leucolomatus Compagno & Smale 1985

Whitetip weasel shark

PLATE 39

Paragaleus leucolomatus Compagno & Smale 1985: 9, Figs. 2–8 (SE of Kosi Bay, KwaZulu-Natal, South Africa); SSF No. 9.30\*; Compagno et al. 2005\*; Ebert et al. 2013\*.

Compagno et al. 2005^; Ebert et al. 2013^.

Slender-bodied, with long snout with bluntly rounded tip in lateral view. Eyes oval; gill slits twice eye length. Mouth long; jaw symphyses extending well anterior to upper lip folds; teeth small and serrated. Lower jaw relatively deep (prominently visible in lateral view); lower anterolateral teeth mostly erectcusped and without distal cusplets, except for last 2 or 3 rows. Pectoral fins strongly falcate. Dorsal fins and anal fin with deeply concave rear margins. Tooth count 28/30; vertebrae 180; spiral valve turns 5.

Body dark grey dorsally, white ventrally, but underside of prenarial snout with broad dusky patches; 2nd dorsal fin with black apical spot; most fins with conspicuous white tips and rear margins. Attains at least 103 cm TL.

**DISTRIBUTION** WIO: Southern Mozambique to northeastern South Africa (type locality); records of *Paragaleus* species from Madagascar may be this or a different species.

**REMARKS** Known from one formally described female specimen, with 1 pup in each uterus, taken from shallow, tropical water, at ~20 m; plus photographs from Mozambique (Plate 39). Assumed viviparous with yolk-sac placenta. The species may be subject to bycatch by coastal line fishers. IUCN Red List conservation status Vulnerable.

# Paragaleus randalli

Compagno, Krupp & Carpenter 1996

#### Slender weasel shark

*Paragaleus randalli* Compagno, Krupp & Carpenter 1996: 393 (Bahrain, Persian/Arabian Gulf); Compagno *et al.* 2005\*; Ebert *et al.* 2013\*.

PLATE 40

Snout with narrowly rounded tip in lateral view. Gill slits about equal to eye length. Mouth long, lower jaw relatively deep (prominently visible in lateral view); jaw symphyses extending well anterior to upper lip folds. Upper and lower teeth not strongly differentiated; lower anterolateral teeth mostly oblique-cusped, with 1 or 2 distal cusplets on 5 or more rows. Pectoral fins falcate. Dorsal fins and anal fin with deeply concave rear margins. Tooth count 28–30/28–33; vertebrae 165–187; spiral valve turns 4 or 5.

Body grey-brown dorsally (head, trunk and caudal fin), becoming paler on flanks, and whitish ventrally (from snout to caudal-fin origin); underside of prenarial snout with pair of narrow black lines (but no dark patches); most fins with dark web and inconspicuous pale rear margin (but no abruptly white or black fin tips). Attains at least 81 cm TL.

**DISTRIBUTION** Indian Ocean: Persian/Arabian Gulf, Gulf of Oman, India and Sri Lanka in WIO; elsewhere to Bay of Bengal (India and possibly Myanmar).

**REMARKS** Poorly known; patchy records from shallow inshore tropical waters of continental shelf, to at least 18 m deep. Probably litters of 1 or 2; size at birth ~29 cm TL; males mature at 62–72 cm TL, and females at 70–80 cm TL. IUCN Red List conservation status Near Threatened.



Paragaleus leucolomatus, 96 cm TL, gravid female holotype, upper and lower teeth from middle of right side of jaw (South Africa). Source: Compagno & Smale 1985

# FAMILY GALEOCERDONIDAE

Tiger shark

Megan V Winton and David A Ebert

Body fairly stout, and head large and broad; snout short and bluntly rounded. Eyes fairly large, without posterior notch; spiracles relatively large, slit-like. Anterior nasal flaps short, broadly triangular, and not tubular. Mouth large; lip folds very long, and upper folds at least twice as long as lower folds and reaching forward to nearly below eyes. Teeth in both jaws characteristically cockscomb-shaped and heavily serrated (sawlike). Pectoral fins moderately broad and semifalcate. Interdorsal ridge prominent; 2nd dorsal fin much smaller than 1st; dorsalfin free rear tips long. Anal fin approximately subequal to 2nd dorsal fin; preanal ridges short; rear margin deeply concave. Peduncle with low lateral keels; upper precaudal pit transverse and crescentic. Caudal fin slender with acutely pointed tip. Viviparous without yolk-sac placenta. One species.

# GENUS Galeocerdo Müller & Henle 1837

Diagnosis as for family. One species, previously placed in the family Carcharhinidae.

### Galeocerdo cuvier (Péron & Lesueur 1822)

Tiger shark

PLATE 49

- *Squalus cuvier* Péron & Lesueur *in* Lesueur 1822: 351 (northwestern Australia).
- *Galeocerdo cuvier*: Bass *et al.* 1975\*; Van der Elst 1981\*; SSF No. 9.19\*; Compagno *et al.* 1989\*; Cockcroft 1991; Smale & Cliff 1998; Dudley *et al.* 2000; Ebert 2003\*; Compagno *et al.* 2005\*; Ebert *et al.* 2021.

Diagnosis as for family. Tooth count 18–26/18–25; vertebrae 216–234.

Body grey dorsally, with characteristic pattern of dark grey to black vertical bars and spots, present as bold reticulations in young and progressively fainter in adults; white ventrally. Attains 7.4 m TL but individuals larger than 5 m TL rarely seen.

**DISTRIBUTION** Circumglobal in warm-temperate to tropical seas, but not known from Mediterranean Sea. WIO: South Africa (Eastern Cape) to Red Sea, Persian/Arabian Gulf, India and Sri Lanka, as well as Socotra, Madagascar, Comoros and Seychelles.

**REMARKS** Relatively large litters of 10–82 (mean ~30–35); size at birth 51-76 cm TL; gestation period 13-16 months with a triennial reproductive cycle in some regions. Fastgrowing in early life; males mature at 226-290 cm TL, aged 7-10 years; females mature at 250-350 cm TL, aged 8-10 years. Maximum age possibly 27-37 years. Common worldwide, on or adjacent to continental and insular shelves; coastal-pelagic, from surface and intertidal zone to at least 140 m deep. Often occurs in estuaries, harbours, coral atolls and lagoons, and may be resident or semi-resident around oceanic islands, preferring turbid areas near volcanic islands and large island groups where a large volume of freshwater runoff may contribute to higher densities of prey organisms. Strong swimmer known to make excursions far offshore into open ocean and capable of long-distance migrations over short periods, with seasonal movements noted in continental waters. A formidable predator and true scavenger; perhaps the most omnivorous of sharks, feeds on a huge variety of bony fishes, elasmobranchs (including tiger shark pups), marine reptiles, sea birds, marine mammals, invertebrates, carrion and even garbage. Caught in numerous coastal and offshore fisheries, as target species or bycatch, and by recreational anglers. One of the potentially most dangerous sharks, with more confirmed attacks on divers, swimmers and boats recorded for this species than for any other shark except the white shark (Carcharodon carcharias), and thus a target of many shark control programmes, yet often nonaggressive when encountered underwater and valuable for dive tourism. IUCN Red List conservation status Near Threatened.



*Galeocerdo cuvier*, 107 cm TL, immature female (S Mozambique). Source: Bass *et al.* 1975

# FAMILY CARCHARHINIDAE

### Requiem sharks

Megan V Winton and David A Ebert

Small- to large-sized (~70 cm to 7 m TL, but most 1-3 m TL) with long arched mouth reaching past front edge of eyes, 2 spineless dorsal fins, and anal fin well separated from caudal fin. Five pairs of gill slits, lateral on body, the last over or behind pectoral-fin origin. Eves circular, vertically oval, or horizontally oval; nictitans internal, and moveable lower eyelid well-developed; spiracles usually absent or minute. Nostrils without oronasal grooves or barbels; anterior nasal flaps vary from vestigial or weakly expanded to tube-shaped. Mouth long and arched; lip folds moderately long and conspicuous to short and confined to corners of mouth. Teeth small to large, with acute and narrow to moderately broad cusps, sometimes with lateral cusplets, and basal ledges and grooves low to absent; teeth variably differentiated in both jaws, but posterior teeth not comb-like; upper teeth often more compressed and bladelike, lower teeth often cuspidate and not compressed; tooth counts 18-60/18-56. Pectoral fins with radials extending into distal web of fins. First dorsal fin moderately sized to very large, its base well ahead of pelvic-fin bases. Second dorsal fin usually much smaller than 1st. Upper and lower precaudal pits present; peduncle usually without lateral keels. Caudal fin non-lunate, but lower lobe distinct and strong; undulations on upper caudal margin. Vertebral centra with strong, wedgeshaped intermedial calcifications. Intestinal valve scrolled. Colour variable and generally without distinctive patterns.

All species for which reproductive information is available are viviparous with yolk-sac placenta; litter sizes range from 1 to 135. Generally strong, active swimmers, occurring singly, in loose aggregations, or socially schooling. Major predators, feeding on a wide variety of prey, including bony fishes, elasmobranchs, cephalopods and crustaceans, as well as sea birds, turtles, sea snakes, marine mammals, benthic invertebrates, carrion and flotsam. By far the most important shark family for fisheries in the tropics, with numerous species prominent in artisanal, commercial and sports fisheries; used for their meat and often highly valued fins, and also the hide and liver oil. Several of the species are among the most dangerous sharks to humans, yet most of the species pose relatively little danger to swimmers and divers, while several are popular subjects of ecotourism, and some are popularly displayed in public aquaria.

One of the largest groups of living sharks. Found in all warm and temperate seas, and the dominant sharks in tropical waters. Many species are common and wide-ranging. Mainly neritic, over continental and insular shelves, coral reefs and reef flats; only a few species are truly oceanic. Several carcharhinid species regularly enter brackish-water river mouths, estuaries or lagoons, and a very few are the only shark species able to live in freshwater for an extended period. None are truly specialised deepwater sharks, although some may be found below ~400 m. Eleven genera and at least 59 species; 9 genera and 30 species in WIO.

#### **KEY TO GENERA**



#### **KEY TO GENERA**

- 3a Snout short, preoral length less than mouth width; upper and lower teeth with narrow smooth-edged cusps ....... *Negaprion*





- Head conical to slightly depressed; pectoral fins narrow, distance from fin origin to free rear tip at most four-fifths length of anterior margin; 1st dorsal-fin free rear tip usually before pelvic-fin origins; anal-fin rear margin concave to deeply concave
- 5a Second dorsal-fin origin well behind anal-fin origin (but sometimes over or slightly before it); preanal ridges long and prominent, and subequal to or greater in length than anal-fin base; anal-fin rear margin straight or shallowly concave .......7

# GENUS Carcharhinus Blainville 1816

Fairly slender to very stout-bodied; snout very short to long, narrowly parabolic or subangular to bluntly rounded or nearly truncate in dorsoventral view. Eyes small to large; no spiracles. Nostrils small and wide-spaced; anterior nasal flaps short or vestigial, and narrowly or broadly triangular, but not tubular. Lip folds confined to corners of mouth. Teeth highly variable, and anteroposterior teeth similar or strongly differentiated in both jaws; upper teeth usually with more or less erect, broad to narrow cusps, with variably developed cusplets or blades, and usually with serrations; lower teeth with oblique to erect cusps, serrations, and blades present or absent, and no cusplets; tooth counts 24-37/23-35. Pectoral fins moderately broad and semifalcate to narrow and falcate or broad-tipped, origins under 3rd to 5th gill slits; distance from fin origin to free rear tip  $\sim \frac{1}{3}-\frac{2}{3}$  length of anterior margin. Interdorsal ridge absent,

- **6a** Rear edge of eyes with distinct notch; lip folds short and confined to corners of mouth; 1st dorsal-fin base 2–3 times into distance between pectoral- and pelvic-fin bases ... *Loxodon*



low or prominent. First dorsal-fin origin over or slightly before pectoral-fin insertions to slightly behind their free rear tips; midbase closer to pectoral-fin bases than to pelvic-fin bases but sometimes equidistant between them; free rear tip usually well in front of pelvic fins but occasionally opposite their origins. Second dorsal fin much smaller than 1st (at most twofifths height of 1st dorsal fin); origin about opposite anal-fin origin. Anal fin as large as to much larger than 2nd dorsal fin; preanal ridges short or absent; rear margin concave to deeply concave. No keels on peduncle; upper precaudal pit transverse and crescentic. Body grey, bronzy or brownish dorsally, usually without distinct colour patterns, but pale and dark fin markings and pale band on flanks variably present. Small- to very large-sized, <1 m to ~4 m TL. Thirty species, 21 in WIO.

#### **KEY TO SPECIES**



7a	Interdorsal ridge present
	interoorban nuge
7b	No interdorsal ridge
8a	Second dorsal fin, pectoral fins, and caudal-fin lower lobe strikingly black-tipped
8b	Fins plain or with dusky tips, but not strongly black-tipped
9a	Second dorsal fin low, with long inner margin over twice fin height; upper anterolateral teeth with strongly serrated cusps; usually only 12 rows of upper anteroposterior teeth <i>C. sorrah</i>
9b	Second dorsal fin high, with shorter inner margin (margin ~1.5 times fin height); upper anterolateral teeth with weakly serrated or smooth-edged cusps; 14 or 15 rows of upper anteroposterior teeth
10a	First dorsal-fin origin well behind pectoral-fin free rear tips; 2nd dorsal-fin inner margin very long, usually >2 times fin height (but down to 1.6 times fin height); crown feet of upper anterolateral teeth relatively narrow and with coarse serrations or small cusplets
10b	First dorsal-fin origin over or before pectoral-fin free rear tips; crown feet of upper anterolateral teeth with small (not coarse) serrations; 2nd dorsal-fin inner margin short, usually <2 times fin height
11a	First dorsal-fin origin above or slightly after pectoral-fin insertions, but nearer to insertions than to free rear tips 12
11b	First dorsal-fin origin above or slightly before pectoral-fin free rear tips, but closer to free rear tips than to insertions 13
	11a 11b

#### **KEY TO SPECIES**

- 14a Entire dorsal and ventral caudal margin with narrow but obvious black edge; 1st dorsal fin and caudal-fin lower lobe with obvious black tips, and smaller black tips to all other fins
  C. melanopterus
- 14bMargin of caudal fin not black, or only partly dusky or black;<br/>fins black-tipped or not15
- 15aSnout short and broadly rounded; internarial space usually<br/>less than preoral length; upper anterolateral teeth with<br/>broad, triangular cusps16









- 19aTeeth in both jaws with smooth edges, except some upper<br/>teeth weakly and irregularly serrated in adultsC. leiodon





Carcharhinus albimarginatus, 83 cm TL, male (central Mozambique). Source: Bass et al. 1973

# Carcharhinus albimarginatus (Rüppell 1837)

Silvertip shark

PLATES 40 & 41

*Carcharias albimarginatus* Rüppell 1837: 64, Pl. 18, Fig. 1 (Ras Muhammad, Sinai, Egypt, Red Sea).

*Carcharhinus albimarginatus*: Bass *et al.* 1973\*; Garrick 1982\*; SSF No. 9.1\*; Compagno *et al.* 1989\*; Compagno *et al.* 2005\*; Last & Stevens 2009\*; Voigt & Weber 2011\*; Ebert *et al.* 2021.

Large-sized, fusiform body; snout moderately long and broadly rounded. Upper lip folds inconspicuous. Upper teeth broad, with erect to moderately oblique triangular cusps, with strong serrations and coarser serrations basally; lower teeth broad but with erect, more slender cusps, and no serrations. Pectoral fins large, semifalcate. Interdorsal ridge present. First dorsal fin moderately large, semifalcate; origin usually over or slightly before pectoral-fin free rear tips. Second dorsal-fin origin over or slightly behind anal-fin origin. Tooth count 12–14/12–14; vertebrae 216–231.

Body bronzy, dark grey or brown dorsally; inconspicuous white band on flanks; all fins (except 2nd dorsal fin) with distinctive white tips and rear margins; white ventrally. Attains 3 m TL.

**DISTRIBUTION** Indo-Pacific (widespread but fragmented). WIO: Red Sea, Kenya to South Africa (KwaZulu-Natal), Madagascar, Comoros, Aldabra, Seychelles, Mascarenes and Chagos.

**REMARKS** Litters of 1–11 (average 6); size at birth 63–81 cm TL; gestation period ~12 months; biennial reproductive cycle. Males mature at 160–180 cm TL, and females at 160–199 cm TL. Occurs in tropical waters over

continental and insular shelves, from surface to ~800 m deep. Has localised movements, with strong preference for offshore islands, coral reefs and offshore banks, but also found in lagoons and well offshore; young are restricted to shallower water close to shore. Feeds on a variety of midwater and bottom fishes and cephalopods. A known bycatch in areas where it occurs, especially around the WIO islands where it is abundant. Potentially dangerous to humans (although few, if any, attacks have been attributed to it). IUCN Red List conservation status Vulnerable.

# Carcharhinus altimus (Springer 1950)

#### Bignose shark

*Eulamia altima* Springer 1950: 9 (off Cosgrove Reef, Key West, Florida). *Carcharinus radamae* Fourmanoir 1961: 24, Figs. 14–17, Pls. 6–7 (west coast of Madagascar).

*Carcharhinus altimus*: Bass *et al.* 1973\*; Compagno 1984\*; SSF No. 9.2\*; Compagno *et al.* 1989\*; Heemstra & Heemstra 2004; Compagno *et al.* 2005\*; Fricke *et al.* 2007; Last & Stevens 2009\*; Voight & Weber 2011\*; Ebert 2013\*; Ebert *et al.* 2021\*.

Large-sized, fairly slender cylindrical body; snout moderately long and bluntly pointed. Eyes circular and moderately large. Anterior nasal flaps short, broadly triangular. Upper lip folds short and inconspicuous. Upper teeth high, triangular, usually with fine serrations and no cusplets; lower teeth with erect, narrow cusps, with serrations. Gill slits moderately long. Pectoral fins large, straight, with pointed or narrowly rounded apices. Interdorsal ridge prominent. First dorsal fin relatively tall, falcate; apex bluntly pointed; origin over pectoralfin insertions to midlength of inner margins, fin height

PLATE 41



*Carcharhinus altimus,* 130 cm TL, female (South Africa). Source: Bass *et al.* 1973

<1/2 predorsal length. Second dorsal-fin origin slightly before anal-fin origin. Tooth count 15/15; vertebrae 194–206.

Body pale grey, sometimes bronzy dorsally; faint white marking on flanks; fin tips (except pelvic fins) dusky; white ventrally. Attains 3 m TL.

**DISTRIBUTION** Circumglobal in all warm-temperate to tropical seas (patchy), including western Mediterranean Sea (rare). WIO: Red Sea, South Africa (Eastern Cape), Madagascar, Maldives, southern India and Sri Lanka.

**REMARKS** Litters of 3–15; size at birth 60–90 cm TL; biennial reproductive cycle. Males mature at 190–267 cm TL, and females at 205–282 cm TL. Common in deep water near edge of continental and insular shelves and on upper slope, at 80–430 m, but young are occasionally found in shallower water (up to 25 m depth). Diurnally vertically migrating: mostly bottom-dwelling by day and nearer surface at night. Feeds on a variety of bony fishes, elasmobranchs and cephalopods (cuttlefish). Interest to fisheries localised; often a utilised bycatch in pelagic longline, gillnet and bottom-trawl fisheries. Not regarded as dangerous to humans due to its deepwater habitat. IUCN Red List conservation status Near Threatened.

# Carcharhinus amblyrhynchoides (Whitley 1934)

Graceful shark

PLATE 40

*Gillisqualus amblyrhynchoides* Whitley 1934: 189, Fig. 4 (Queensland, Australia).

Carcharhinus amblyrhynchoides: Compagno et al. 2005\*;

Last & Stevens 2009\*; Voight & Weber 2011\*; Ebert et al. 2021\*.

Moderately large-sized and stout-bodied; snout fairly short and pointed, wedge-shaped; internarial width 1–1.2 times preoral length. Eyes circular and moderately large. Upper lip folds short and inconspicuous. Teeth in both jaws with erect, narrow cusps, with serrations and no cusplets. Gill slits long. Pectoral fins rather large, falcate. No interdorsal ridge. First dorsal fin rather large, broadly triangular, semifalcate; origin over or slightly after pectoral-fin insertions. Second dorsal-fin origin over or slightly before anal-fin origin. Tooth count 15/15; vertebrae 168–193.

Body grey dorsally; conspicuous white band on flanks; most fins black-tipped, and caudal-fin upper lobe usually with dusky edges (dark fin markings may fade in adults); paler ventrally. Attains at least 178 cm TL.



Carcharhinus amblyrhynchoides, 70 cm TL, male (Pakistan). © HB Osmany

**DISTRIBUTION** Indo-Pacific. WIO: Gulf of Aden, Pakistan, southwestern India and Sri Lanka (but possibly continuous from Somalia to India, including Red Sea and Persian/Arabian Gulf).

**REMARKS** Poorly known. Litters of 1–9 (average 3); size at birth 50–60 cm TL. Both sexes mature at 110–115 cm TL; gestation period ~9–10 months. Coastal-pelagic in tropical waters on continental and insular shelves. Feeds primarily on fishes, but also cephalopods and crustaceans. Taken in gillnet and longline fisheries where it occurs, but not targeted. Considered harmless to humans. IUCN Red List conservation status Near Threatened.

# Carcharhinus amblyrhynchos (Bleeker 1856)

Grey reef shark

PLATES 41 & 42

*Carcharias (Prionodon) amblyrhynchos* Bleeker 1856: 467 (Java Sea near Solombo I., Indonesia).

?Carcharhinus wheeleri Garrick 1982: 111, Figs. 50-51 (Red Sea).

Carcharhinus amblyrhynchos: Bass et al. 1973\*; Compagno 1984\*; SSF No. 9.3\*; Compagno et al. 2005\*; Last & Stevens 2009\*; Voight & Weber 2011\*; Ebert et al. 2021\*.

Moderate-sized, fairly stocky body; snout moderately long and broadly rounded. Eyes usually round and fairly large. Anterior nasal flaps weakly expanded, low triangular lobes. Upper lip folds short and inconspicuous. Upper anteroposterior teeth with narrow cusps, with serrations, and crown feet with coarser serrations and often low distal cusplets. Gill slits moderate-sized. Pectoral fins moderately large, narrow and falcate. Interdorsal ridge weak or absent. First dorsal fin relatively tall and semifalcate; origin usually over or just before pectoral-fin free rear tips. Second dorsal-fin origin roughly over anal-fin origin. Tooth count 28–30/27–29 (usually 28/28); vertebrae 211–221, precaudal vertebrae 110–119.

Body grey dorsally; 1st dorsal fin plain or irregularly whiteedged; pectoral fins, 2nd dorsal fin, pelvic fins and anal fin with dusky tips; caudal-fin rear margin with conspicuous broad black edge; white ventrally. Attains 255 cm TL.

**DISTRIBUTION** Indo-Pacific. WIO: Red Sea, Gulf of Aden, Kenya to South Africa, Madagascar, Seychelles, Mauritius and India.

**REMARKS** Litters of 1–6; size at birth 45–60 cm TL; gestation period 12-14 months. Males mature at 130-145 cm TL, aged 7-9 years; females mature at 122-137 cm TL, aged 7.5-11 years; maximum age 25 years or more. Coastalpelagic, on continental and insular shelves and in adjacent oceanic waters, from surface and intertidal zone to at least 100 m. Common on coral reefs, often in deeper areas near drop-offs to open sea, in atoll passes, and in shallow lagoons adjacent to areas of strong current. Active, strong swimming, social species found in groups or individually; prefers clear water and shows site fidelity, forming daytime schools or aggregations in favoured areas. Feeds mostly on small bony fishes, but also cephalopods (squid and octopus) and crustaceans. Can be aggressive to humans, yet a very popular subject of dive ecotourism. Caught in multi-species tropical fisheries. IUCN Red List conservation status Endangered.



Carcharhinus amblyrhynchos, 68 cm TL, female (NW Madagascar). Source: Bass et al. 1973



Carcharhinus amboinensis, 144 cm TL, female (South Africa). Source: Bass et al. 1973

### Carcharhinus amboinensis (Müller & Henle 1839)

Pigeye shark

PLATE 41

Carcharias (Prionodon) amboinensis Müller & Henle 1839: 40, Pl. 19 [teeth] (Ambon I., Maluku Is., Indonesia). Triaenodon obtusus Day 1878: 720, Pl. 189, Fig. 3 (Karachi, Pakistan). Carcharhinus amboinensis: D'Aubrey 1964, 1971; Bass et al. 1973\*; Compagno 1984\*; SSF No. 9.4\*; Compagno et al. 1989\*; Bonfil &

Abdallah 2004\*; Heemstra & Heemstra 2004; Compagno *et al.* 2005\*; Last & Stevens 2009\*; Voight & Weber 2011\*; Ebert *et al.* 2021\*.

Large-sized, stocky body and thick head; snout short and bluntly rounded, internarial width 0.9–1 in preoral length. Eyes circular and small. Anterior nasal flaps short, broadly triangular. Upper lip folds short and inconspicuous. Upper teeth broadly triangular, with serrations and no cusplets; lower teeth slightly narrower, with serrations and arched roots. Gill slits moderately long. Pectoral fins large and broad, triangular to semifalcate, with narrow, pointed apices. No interdorsal ridge. First dorsal fin large, broadly triangular or somewhat falcate, with pointed or sharply rounded apex; origin over or just behind pectoral-fin insertions; its height >3 times 2nd dorsal fin. Second dorsalfin origin before anal-fin origin. Tooth count 11–13/10–12; vertebrae 185–195, precaudal vertebrae 89–95.

Body grey dorsally; inconspicuous white band on flanks; fin tips dusky, especially in young; pale ventrally. Attains 280 cm TL.

**DISTRIBUTION** Northeastern Atlantic (Nigeria) and Indo-Pacific (patchy). WIO: Pakistan, Gulf of Aden, Persian/ Arabian Gulf, South Africa (KwaZulu-Natal), Madagascar and Sri Lanka.

**REMARKS** Similar in appearance to bull shark *C. leucas*. Litters of 3–13; size at birth 60–75 cm TL; gestation period ~12 months. Males mature at 195–210 cm TL, and females at 198–223 cm TL. Occurs near bottom in shallow inshore waters over continental and insular shelves, from near surfline to ~60 m deep, and occasionally enters brackish water; usually solitary. Feeds primarily on bottom fishes, crustaceans, cephalopods and molluscs. Caught in small numbers in longline and gillnet fisheries. Considered potentially dangerous to humans. IUCN Red List conservation status Data Deficient globally; Near Threatened for southwestern Indian Ocean subpopulation.

# Carcharhinus brachyurus (Günther 1870)

Bronze whaler

PLATE 41

*Carcharias brachyurus* Günther 1870: 369 (off Wanganui, New Zealand). *Carcharhinus improvisus* Smith 1952: 761, Fig. 1 (Algoa Bay, South Africa). *Carcharhinus brachyurus*: Bass *et al.* 1973\*; Van der Elst 1981\*; Compagno 1984\*; SSF No. 9.5\*; Compagno *et al.* 1989\*; Walter & Ebert 1991; Heemstra & Heemstra 2004\*; Compagno *et al.* 2005\*; Last & Stevens 2009\*; Voight & Weber 2011\*; Ebert *et al.* 2021\*.

Large-sized; snout moderately broad, long and bluntly pointed, internarial width 1.1–1.4 in preoral length. Eyes circular and moderately large. Anterior nasal flaps low and poorly developed. Upper lip folds short and inconspicuous. Upper anterolateral teeth with narrow bent cusps, with serrations and no cusplets; teeth in both jaws with transverse notch at root. Gill slits moderately long. Pectoral fins moderately large and falcate. Usually no interdorsal ridge. First dorsal fin large, falcate; origin over or slightly before pectoral-fin free rear tips. Second dorsal-fin origin over or slightly after anal-fin origin. Tooth count 14–16/14–15; vertebrae 179–203.

Body bronzy grey to olive-grey dorsally; fairly prominent white band on flanks; most fins with inconspicuous darker edges and dusky to black tips; white ventrally. Attains 295 cm TL.



Carcharhinus brachyurus, 143 cm TL, male (South Africa). Source: Bass et al. 1973

**DISTRIBUTION** Atlantic (Gulf of Mexico and West Africa), Pacific and Indian oceans, and Mediterranean Sea. WIO: South Africa (Cape of Good Hope to KwaZulu-Natal).

**REMARKS** Often confused with other large *Carcharhinus* species. Litters of 7–24; size at birth 59–70 cm TL; gestation period 15–21 months. Males mature at 200–229 cm TL, age 13–19 years; females mature at 240–292 cm TL, age 19–20 years; maximum age 25–30 years. Found in subtropical and temperate waters, close inshore and occasionally offshore, from surfline to at least 100 m deep, and enters lower reaches of large rivers; occurs singly and in loose schools. Apparently migratory in northern parts of its range, moving northwards in spring and summer and southwards in fall and winter; also segregates by size and sex, and uses inshore bays and open coastline as nursery grounds. Feeds on a wide variety of bottom-dwelling and pelagic fishes and cephalopods. Taken as bycatch and in directed fisheries, mostly by gillnets and bottom

longlines, as well by recreational anglers and in anti-shark nets. Several provoked and unprovoked encounters with swimmers and divers have been ascribed to this species. IUCN Red List conservation status Vulnerable globally.

## Carcharhinus brevipinna (Valenciennes 1839)

Spinner shark

PLATE 41

*Carcharias (Aprion) brevipinna* Valenciennes *in* Müller & Henle 1839: 31, Pl. 9 (Java, Indonesia).

- *Carcharinus johnsoni* Smith 1951: 88, Figs. 1–2 (Port Elizabeth, Eastern Cape, South Africa).
- *Carcharhinus brevipinna*: Bass *et al.* 1973\*; Compagno 1984\*; SSF No. 9.6\*; Compagno *et al.* 1989\*; Allen & Cliff 2000; Allen & Wintner 2002; Heemstra & Heemstra 2004\*; Compagno *et al.* 2005\*; Fricke *et al.* 2009; Last & Stevens 2009\*; Voight & Weber 2011\*; Ebert *et al.* 2021\*.



Carcharhinus brevipinna, 179 cm TL, male (South Africa). Source: Bass et al. 1973

PLATE 41

Large-sized, fairly slender body; snout long and pointed, internarial width 1.5–1.8 in preoral length. Eyes small and circular. Anterior nasal flaps low and inconspicuous. Upper lip folds usually long and conspicuous. Teeth fairly small; upper anterolateral teeth mostly erect, with narrow cusps, serrated or partly serrated, and no cusplets; lower teeth with narrow cusps and no serrations. Gill slits long. Pectoral fins falcate. No interdorsal ridge. First dorsal fin relatively large, semifalcate; height >2.2 in interdorsal space; origin over or slightly after pectoral-fin free rear tips. Second dorsal-fin origin over or slightly after anal-fin origin. Tooth count 15–18/14–17 (usually 16/16); vertebrae 155–185.

Body greyish to bronzy dorsally; inconspicuous white band on flanks; all fins (except pelvic fins) black-tipped in large juveniles and adults, and plain in young (<80 cm TL); white ventrally. Attains 3 m TL.

**DISTRIBUTION** Circumglobal, including Mediterranean Sea, except not eastern Pacific. WIO: Pakistan, Gulf of Oman, Gulf of Aden, Red Sea, Mozambique, South Africa (southwestern Cape), Madagascar, Comoros (Mayotte I.), Amirante Bank, Seychelles, Réunion, India and Sri Lanka; not known from Persian/Arabian Gulf.

**REMARKS** Litters of 3–15; size at birth 60–75 cm TL; gestation period 12–15 months; biennial reproductive cycle. Males mature at 159–203 cm TL, and females at 170–200 cm TL, aged 8–10 years for both sexes; maximum age 17–19 years. Common, coastal-pelagic, in warm-temperate to tropical waters, close inshore off beaches, in bays and off river mouths, from surface to at least 75 m deep offshore (but usually above 30 m). Some segregation by size and sex. Active, schooling shark that often leaps 'spinning' from water as part of its frenzied feeding run. Feeds mostly on a variety of fishes, but also cephalopods. Captured frequently in commercial and recreational fisheries. Not considered dangerous to people, but possibly troublesome to divers when spearfishing. IUCN Red List conservation status Vulnerable.

# Carcharhinus dussumieri (Valenciennes 1839)

#### Whitecheek shark

Carcharias (Prionodon) dussumieri Valenciennes in Muller & Henle 1839: 47, Pl. 19 [teeth] (Puducherry, India).

*Carcharhinus dussumieri*: Fowler 1941; Compagno 1984\*; Compagno *et al.* 2005\*; Last & Stevens 2009\*; Voigt & Weber 2011\*; White 2012\*; Ebert *et al.* 2021\*.

Small-sized; snout moderately long and rounded. Eyes fairly large, horizontally oval. Anterior nasal flaps conspicuous, triangular. Mouth width 6.4–8.3% TL; upper lip folds short and inconspicuous. Upper teeth with oblique triangular cusps, with serrations and cusplets; lower teeth smaller, with erect narrow cusps. Pectoral fins small and semifalcate, with narrow angular apices. Interdorsal ridge low, if present. First dorsal fin moderately large, broadly triangular, not strongly falcate; origin over rear half of pectoral-fin inner margins. Second dorsal-fin origin over or slightly after anal-fin origin. Tooth count 12–14/11–15 (usually 13/13); vertebrae 109–150, precaudal vertebrae 54–74.

Body grey, grey-brown or bronzy dorsally; inconspicuous pale band on flanks; 2nd dorsal fin with black or dusky tip, other fins with pale trailing edges; paler ventrally. Attains 100 cm TL.

**DISTRIBUTION** Indo-Pacific. WIO: Persian/Arabian Gulf, Gulf of Oman, Pakistan, India and Sri Lanka.

**REMARKS** Common but poorly known. Litters of 1–4 (usually 2); size at birth 28–38 cm TL. Males mature at 65–70 cm TL, and females at 70–75 cm TL. Occurs in inshore tropical waters, over continental and insular shelves, to ~170 m deep. Feeds mostly on small bony fishes, but also cephalopods and crustaceans. Commonly caught in inshore fisheries, including artisanal and small-scale commercial fisheries; used for its meat and fins. Harmless to humans. IUCN Red List conservation status Endangered globally.



Carcharhinus dussumieri. © Food and Agriculture Organization of the United Nations, Original Scientific Illustrations Archive; reproduced with permission

# Carcharhinus falciformis (Müller & Henle 1839)

#### Silky shark

PLATE 41

*Carcharias (Prionodon) falciformis* Müller & Henle (ex Bibron) 1839: 47 (Cuba, western Atlantic).

Carcharhinus falciformis: Compagno 1984\*; SSF No. 9.7\*;

Compagno *et al.* 2005\*; White *et al.* 2006\*; Last & Stevens 2009\*; Voigt & Weber 2011\* [as *falciformes*]; Ebert *et al.* 2021\*.

Large-sized, fairly slender body; snout moderately long and rounded. Eyes circular and moderately large. Upper lip folds short and inconspicuous. Teeth in both jaws with erect to semi-oblique, fairly narrow, triangular cusps; upper teeth with strong serrations, and lower teeth smooth-edged. Pectoral fins long and narrowly falcate. Interdorsal ridge narrow. First dorsal fin low, falcate, with moderately rounded apex and curved rear margin; origin behind pectoral-fin free rear tips. Second dorsal fin low; inner margin very long, usually more than twice fin height; origin over or slightly after anal-fin origin. Skin smooth, with tightly packed, small, overlapping denticles. Tooth count 14–16/13–17; vertebrae 199–215.

Body dark bronzy grey-brown dorsally, sometimes nearly blackish; inconspicuous white band on flanks; tips of all fins (except 1st dorsal fin) dusky but not black-tipped; white ventrally. Attains 3.7 m TL.

**DISTRIBUTION** Circumglobal in tropical seas (widespread). WIO: Mozambique to Red Sea, Persian/Arabian Gulf, Pakistan, India and Sri Lanka.

**REMARKS** Litters of 2–18; size at birth 65–81 cm TL; gestation period ~12 months. Males mature at 180–230 cm TL, age 5–13 years; females mature at 180–246 cm TL, age 6–15 years. Oceanic and coastal-pelagic; common near edge of continental and insular shelves, but also found far from land

in open sea, from surface to at least 500 m. Segregates by size; young occur in coastal nursery areas and move offshore as subadults. Feeds mostly on bony fishes, but also cephalopods and crustaceans (pelagic crabs). Dominates as a bycatch or target species in tropical pelagic fisheries; often associated with schools of tuna. Active, swift and inquisitive, and can be aggressive to divers. IUCN Red List conservation status Vulnerable.

# Carcharhinus galapagensis

(Snodgrass & Heller 1905)

Galapagos shark

PLATE 41

*Carcharias galapagensis* Snodgrass & Heller 1905: 343 (Galápagos Is.). *Carcharhinus galapagensis*: Bass *et al.* 1973\*; Compagno 1984\*; SSF No. 9.8\*; Compagno *et al.* 2005\*; Last & Stevens 2009\*; Voight & Weber 2011\*; Ebert *et al.* 2021\*.

Large-sized, fairly slender body; snout moderately long and broadly rounded, internarial width 1–1.3 in preoral length. Eyes circular and moderately large. Anterior nasal flaps low and poorly developed. Upper lip folds short and inconspicuous. Upper teeth with broad, triangular, erect to semi-oblique cusps, with strong serrations; lower teeth with erect and narrow cusps. Pectoral fins large and long, narrowly falcate. Interdorsal ridge low. First dorsal fin moderately large, falcate; origin over slightly before pectoral-fin free rear tips. Second dorsal-fin inner margin long (cf. *C. obscurus*); origin over or slightly before anal-fin origin. Tooth count usually 14/14; vertebrae 200–215, precaudal vertebrae 103–109.

Body brownish grey to dark grey dorsally; inconspicuous white band on flanks; fin tips may be dusky but not black or white; white ventrally. Attains 3.7 m TL.



*Carcharhinus falciformis,* 94 cm TL, male (S Mozambique). Source: Bass *et al.* 1973



Carcharhinus galapagensis, 114 cm TL, female (Walters Shoals). Source: Bass et al. 1973

**DISTRIBUTION** Circumglobal in tropical to warm-temperate seas: patchy records from widely separated oceanic islands and some coastal sites. WIO: Walters Shoals (south of Madagascar).

**REMARKS** This shark and the dusky shark *C. obscurus* may prove to be the same species as they are currently differentiated only by their habitat. Litters of 4–16 (average 9); size at birth 57–80 cm TL; gestation period ~12 months. Males mature at 170–250 cm TL, and females at ~205–250 cm TL. Coastal-pelagic, from close inshore to well offshore near or on insular or continental shelves, especially around offshore islands, from surface to at least 285 m. Can be locally abundant; favours rugged substrates and clear water, and possibly strong currents; swims near bottom, frequently in aggregations. Feeds primarily on bottom fishes and cephalopods. Potentially aggressive to divers.

### Carcharhinus hemiodon (Valenciennes 1839)

Pondicherry shark

PLATE 41

*Carcharias (Hypoprion) hemiodon* Valenciennes *in* Müller & Henle 1839: 35, Pl. 19 [teeth] (Puducherry, India).

*Carcharhinus hemiodon*: Garrick 1985\*; Compagno *et al.* 2005\*; Voigt & Weber 2011\*; Ebert *et al.* 2013\*; Ebert *et al.* 2021\*.

Small-sized, somewhat stocky body; snout moderately long and bluntly pointed, internarial width 1–1.4 in preoral length. Eyes circular and moderately large. Anterior nasal flaps low and poorly developed. Upper lip folds short and inconspicuous. Upper teeth with narrow, oblique, high cusps, with low cusplets; lower teeth with semierect, narrow cusps, with no serrations. Pectoral fins moderately small and broad. Interdorsal ridge present. First dorsal fin large, falcate; origin just behind pectoral-fin insertions. Second dorsal-fin inner margin short, 1.4–1.6 times fin height; origin over or slightly after anal-fin origin. Tooth count 14 or 15/13 or 14; vertebrae 147–155.

Body grey dorsally; conspicuous white band on flanks; black tips on pectoral fins, 2nd dorsal fin, and caudal-fin upper and lower lobes; white ventrally. Attains at least 102 cm TL.



**DISTRIBUTION** Indo-Pacific. WIO: Gulf of Oman to Pakistan, India, Sri Lanka and Lakshadweep.

**REMARKS** Rare, possibly extinct; last recorded in India in 1979, and now known only from ~20 museum specimens collected from heavily fished, widely separated sites. Size at birth <32 cm TL. Coastal, over continental and insular shelves; also reported from river mouths and upriver in freshwater, but these records may have been based on species of *Glyphis* or other *Carcharhinus*. IUCN Red List conservation status Critically Endangered.



*Carcharhinus humani*, 91 cm TL, female (South Africa). Source: Bass *et al*. 1973

# Carcharhinus humani White & Weigmann 2014

Human's whaler shark

PLATE 42

*Carcharhinus humani* White & Weigmann 2014: 71, Fig. 1 (Socotra). *Carcharias sealei* Pietschmann 1913: 172, Pl. 1 (Sandakan fish market, Sabah, Borneo, Indonesia).

Carcharhinus sealei: Bass et al. 1973\*; Van der Elst 1981\*; SSF No. 9.16\*; Compagno et al. 1989\*; Compagno et al. 2005\*; Voigt & Weber 2011\*; White 2012\*; Ebert et al. 2021\*.

Small-sized body; snout moderately long and rounded. Anterior nasal flaps moderately elongate and broadly triangular. Eyes large and oval. Mouth width 4.2–6.6% TL; upper lip folds short and inconspicuous. Teeth in both jaws with oblique cusps, with coarse serrations or low cusplets basally. Pectoral fins small and strongly falcate; distance from origin to free rear tip 1.7–2 into length of anterior margin. Interdorsal ridge low, if present. First dorsal fin falcate; origin over or slightly after pectoral-fin free rear tips. Second dorsalfin origin after anal-fin origin. Tooth count 24–28/23–27; vertebrae 148–167, precaudal vertebrae 74–85.

Body grey or tan dorsally; inconspicuous pale band on flanks; 2nd dorsal fin with conspicuous black or dusky tip, other fins with pale rear edges and no dark markings; paler ventrally. Attains 95 cm TL.

**DISTRIBUTION** Indo-Pacific. WIO: west coast of Madagascar, Seychelles, South Africa (KwaZulu-Natal) to Somalia, Socotra, Oman to Pakistan, India and Sri Lanka. **REMARKS** Litters of 1 or 2; size at birth 33–45 cm TL; gestation period ~9 months. Males mature at 70–80 cm TL, and females at 68–75 cm TL, at ~1 year for both sexes; maximum age at least 5 years. Occurs on continental and insular shelves, in shallow water, from surf line and intertidal zone to ~40 m deep. Feeds on small fishes, cephalopods (squid) and crustaceans (prawns). Commonly taken with line gear and gillnets in artisanal and small-scale commercial fisheries and by sport anglers where it occurs. Harmless to humans.

# Carcharhinus leiodon Garrick 1985

### Smoothtooth blacktip shark

*Carcharhinus leiodon* Garrick 1985: 11, Figs. 7–8 (Gulf of Aden [probably at Yemen]); Manilo & Bogorodsky 2003; Compagno *et al.* 2005\*; Voigt & Weber 2011\*; Moore *et al.* 2011, 2013; Ebert *et al.* 2021\*.

Body somewhat stocky; snout moderately long and bluntly pointed. Eyes large and circular. Upper lip folds short and inconspicuous. Upper teeth narrow, smooth-edged, erect near centre of mouth and slightly oblique laterally; lower teeth narrow, erect, and smooth-edged. Gill slits relatively long, 3rd gill slit 4.5% TL and >½ length of 1st dorsal-fin base. Pectoral fins relatively small, weakly falcate, and apices pointed. No interdorsal ridge. First dorsal fin moderately high, triangular, not falcate, with pointed apex; origin over midlength of pectoral-fin inner margins. Second dorsal fin relatively large and high, height ~4% TL; origin above anal-fin origin. Both dorsal fins with short free rear tips. Upper precaudal pit strongly developed, and lower pit weakly developed. Tooth count 35/33; vertebrae 198, precaudal vertebrae 115 (holotype).

Body greyish green to greenish yellow dorsally, paler to whitish ventrally; all fins with conspicuous black tips. Attains 165 cm TL.

**DISTRIBUTION** WIO: Gulf of Aden to Oman and Persian/ Arabian Gulf.

**REMARKS** Similar in appearance to the grey reef shark *C. amblyrhynchoides* (which has serrated upper teeth). Until recently only known from holotype (immature male, 75 cm TL). Litters of 4–6; size at birth ~35–53 cm TL. Males mature at ~120 cm TL, and females at ~131 cm TL. Feeds on bony fishes. Habitat likely inshore. IUCN Red List conservation status Endangered.

### Carcharhinus leucas (Valenciennes 1839)

Bull shark

PLATE 43

*Carcharias (Prionodon) leucas* Valenciennes *in* Müller & Henle 1839: 42 (Antilles, western Atlantic).

*Carcharias (Prionodon) zambezensis* Peters 1852: 276 (Zambezi River, Mozambique); Bonfil & Abdallah 2004\*; Voigt & Weber 2011\*.

*Carcharhinus vanrooyeni* Smith 1958: 12, Fig. (St Lucia, KwaZulu-Natal, South Africa).

Carcharhinus leucas: D'Aubrey 1964, 1971; Bass et al. 1973\*;

Van der Elst 1981\*; SSF No. 9.9\*; Compagno *et al.* 1989\*; Cockcroft 1991; Wintner *et al.* 2002; Heemstra & Heemstra 2004\*; Compagno *et al.* 2005\*; Last & Stevens 2009\*; Voigt & Webber 2011\*; Ebert *et al.* 2021\*.

Large-sized, heavy-bodied; snout very short and bluntly rounded, internarial width 0.7–1 in preoral length. Eyes circular and small. Upper lip folds short and inconspicuous. Upper teeth large, broad and triangular, and saw-like with serrations and no cusplets; lower teeth triangular and narrower, erect to slightly oblique, and with weak serrations. Pectoral fins large and broad, triangular to semifalcate. No interdorsal ridge. First dorsal fin large and broadly triangular, its height up to 3 times 2nd dorsal fin; origin over or slightly after pectoral-fin insertions (sometimes nearer their free rear tips). Second dorsal fin relatively high; origin before anal-fin origin. Inconspicuous caudal keels. Tooth count 25–29/25–29; vertebrae 198–227, precaudal vertebrae 101–123.

Body grey to grey-brown dorsally; inconspicuous white band on flanks; juveniles with dusky to black fin tips, and adults with indistinct fin markings; caudal-fin upper lobe with dusky anterior and posterior margins, whitish between. Attains 3.4 m TL.

**DISTRIBUTION** Cosmopolitan in most tropical seas and commonly enters freshwater rivers and lakes. WIO: Persian/ Arabian Gulf, Somalia to South Africa (Eastern Cape), Zanzibar (Tanzania), east coast of Madagascar, Seychelles, Lakshadweep and India; regional range includes great distance up Zambezi River (Mozambique).



*Carcharhinus leucas*, 166 cm TL, male (South Africa). Source: Bass *et al*. 1973

**REMARKS** Often confused with the pigeye shark C. amboinensis or Ganges shark Glyphis gangeticus. Litters of 1–13 (mostly 6–8); gestation period 10–11 months; size at birth 56-81 cm TL. Males mature at 157-226 cm TL, age ~25 years; females mature at 180-230 cm TL, aged 14-22 years; maximum age 29-32 years, possibly >50 years. Coastal, estuarine, riverine, and lacustrine tropical and subtropical; usually close inshore, from surfline to usually <30 m but recorded to 152 m. Commonly occurs in hyposaline and hypersaline lagoons and bays, river mouths, and near surfline; the only wide-ranging shark that penetrates far into freshwater and is able to exist there for long periods. Migrates seasonally in some portions of its range; uses estuaries and river mouths as pupping grounds. A versatile and opportunistic predator with a varied diet comprising bony fishes, elasmobranchs, sea turtles, birds, dolphins, whale offal, terrestrial mammals and invertebrates. Caught in fisheries throughout its range, but rarely targeted. Lives well in public aquaria displays, for which there is increasing demand. May be the most dangerous species of tropical shark (particularly in warm turbid waters), and one of the three most dangerous sharks by number of recorded attacks on people. IUCN Red List conservation status Near Threatened.

# Carcharhinus limbatus (Valenciennes 1839)

#### Blacktip shark

PLATES 43 & 44

*Carcharias (Prionodon) limbatus* Valenciennes *in* Müller & Henle 1839: 49, Pl. 19 [teeth] [Martinique I., West Indies].

*Carcharhinus limbatus*: Bass *et al.* 1973\*; Van der Elst 1981\*; SSF No. 9.10\*; Compagno *et al.* 1989\*; Dudley & Cliff 1993; Wintner & Cliff 1996; Heemstra & Heemstra 2004; Compagno *et al.* 2005\*; Last & Stevens

2009\*; Voigt & Weber 2011\*; Ebert *et al.* 2021\*.

Large-sized, fairly stocky body; snout long, narrow and pointed. Anterior nasal flaps low, triangular. Upper lip folds short and inconspicuous. Upper anterolateral teeth with narrow, mostly erect cusps, with serrations and no cusplets; lower teeth with narrow cusps, usually with fine serrations. Gill slits long. Pectoral fins moderately large and falcate. No interdorsal ridge. First dorsal fin large, falcate, its height 1.5–2 in interdorsal space; origin just after pectoral-fin insertions (exceptionally nearer pectoral-fin free rear tips). Second dorsal-fin origin over or slightly before anal-fin origin; free rear tip long. Tooth count 29–35/27–33; vertebrae 174–203.

Body bronzy to grey-brown dorsally (fades to grey after death); conspicuous white band on flanks; black tips usually present on pectoral fins, dorsal fins and caudal-fin lower lobe, and sometimes on pelvic fins and anal fin (but anal fin often plain in individuals >80 cm TL), and black edges usually present on 1st dorsal fin (from tip to posterior margin) and caudal-fin upper lobe (anterior margin to tip); juveniles with stronger fin markings, and adults with less distinct fin markings; white ventrally. Attains 255 cm TL.

**DISTRIBUTION** Circumglobal in tropical to warm-temperate seas (widespread), including Mediterranean Sea. WIO: Persian/Arabian Gulf, Red Sea, Arabian Sea, Socotra to South Africa (Eastern Cape), Madagascar, Comoros, Seychelles and Sri Lanka.

**REMARKS** Litters of 1–11 (usually 4–7); size at birth 38–72 cm TL; gestation period 10–12 months, reproductive cycle biennial. Males mature at 135–180 cm TL, aged 4–5 years; females mature at 120–190 cm TL, aged 6–7 years; maximum age at least 12 years. Common close inshore and rarely deeper than 30 m; found off river mouths and in



*Carcharhinus limbatus*, 115 cm TL, male (South Africa). Source: Bass *et al.* 1973 estuaries (not far into freshwater), in shallow muddy bays, the more saline parts of mangrove swamps, island lagoons, and along coral reef drop-offs, as well as offshore around some oceanic islands. Seasonally migratory in some parts of its range. Active and fast-swimming, often in loose aggregations near surface, and may leap from the water while feeding (often confused with the spinner shark, *C. brevipinna*, for this behaviour). Feeds mostly on bony fishes, also cephalopods, crustaceans and small elasmobranchs. Caught in recreational, artisanal and directed commercial fisheries with a variety of gear; its meat and fins are highly marketable. Poses little threat to humans unless stimulated by food, with only a few attacks recorded. IUCN Red List conservation status Near Threatened.

### Carcharhinus longimanus (Poey 1861)

Oceanic whitetip shark

PLATE 44

Squalus longimanus Poey 1861: 338, Pl. 19, Figs. 9-10 (Cuba).

*Pterolamiops magnipinnis* Smith 1958: 132, Pl. 1, Fig. 1 (off Port Elizabeth, Eastern Cape, South Africa).

Pterolamiops budkeri Fourmanoir 1961: 76 (Mozambique Channel).

*Carcharhinus longimanus*: Bass *et al.* 1973\*; SSF No. 9.11\*; Compagno *et al.* 1989\*; Ebert 2003\*; Compagno *et al.* 2005\*; Last & Stevens 2009\*; Voigt & Weber 2011\*; Ebert *et al.* 2021\*.

Body stocky; snout short and blunt. Upper lip folds short and inconspicuous. Upper teeth broad and triangular, erect to slightly oblique, and with strong serrations; lower teeth somewhat smaller and narrower, erect to slightly oblique, and with serrations and no cusplets. Pectoral fins distinctly long and broadly rounded (paddle-shaped), length of anterior margin 19–29% TL. Interdorsal ridge low, if present. First dorsal fin very long and large, with distinctly broadly rounded apex; origin just before pectoral-fin free rear tips. Second dorsal-fin origin over or slightly before anal-fin origin. Inconspicuous caudal keels. Tooth count 27–32/27–33; vertebrae 228–244.

Body grey-bronze dorsally; inconspicuous white band on flanks; white mottling usually present on fins (particularly pectoral fins, 1st dorsal fin, pelvic fins, and caudal-fin tips); young with additional black blotches or tips on most fins, as well as black saddles at 2nd dorsal-fin origin, upper caudal-fin origin, and sometimes between dorsal fins, and fading in adults; white ventrally. Attains 3.9 m TL, but most adults <3 m TL.

**DISTRIBUTION** Circumglobal in tropical to warm-temperate seas (widespread), including Mediterranean Sea. WIO: South Africa (KwaZulu-Natal) to Red Sea, and eastwards to India and Sri Lanka, including Socotra, Zanzibar, Madagascar, Comoros, Seychelles, Mascarenes, Chagos, Maldives and Lakshadweep; not known from Persian/Arabian Gulf.

**REMARKS** Litters of 1–15; size at birth 55–77 cm TL; gestation period ~12 months; reproductive cycle likely biennial. Males mature at 168–198 cm TL, and females at 175–224 cm TL; matures at 4–7 years. Oceanic and epipelagic, from surface to at least 152 m over deep water, but occasionally coastal in shallow water off oceanic islands or where continental shelf is very narrow. Feeds mostly on oceanic bony fishes and cephalopods, but also stingrays, crustaceans, sea birds, turtles, marine mammals and flotsam. Caught in large numbers as bycatch and in directed oceanic shark fisheries, particularly by longline and driftnet; the fins have high value in international trade. Slow-moving near surface, yet very active and inquisitive; dangerous to humans, but sought by ecotourism divers in the Red Sea and Indo-Pacific. IUCN Red List conservation status Critically Endangered globally.



# Carcharhinus macloti (Müller & Henle 1839)

#### Hardnose shark

PLATE 44

*Carcharias (Hypoprion) macloti* Müller & Henle 1839: 34, Pl. [10] (New Guinea).

Carcharhinus macloti: Compagno 1984\*; SSF No. 9.12\*; Compagno et al. 2005\*; Last & Stevens 2009\*; Voigt & Weber 2011\*; Ebert et al. 2021\*.

Small-sized slender body; snout long, pointed, and noticeably stiff (rostrum hypercalcified, easily detectable by pinching or cutting into snout). Eyes large and circular. Anterior nasal flaps short and nipple-like. Upper lip folds short and inconspicuous. Upper anterolateral teeth with large basal serrations; lower teeth with oblique, moderately high, narrow cusps, with no serrations. Pectoral fins small and falcate. No interdorsal ridge. First dorsal fin moderate-sized, falcate; origin over or slightly before pectoral-fin free rear tips. Second dorsal fin low; origin near anal-fin midbase. Inner margin of both dorsal fins extremely long, at least two-thirds length of fin bases. Tooth count 29–32/26–29; vertebrae 151–156.

Body bronzy grey-brown dorsally; inconspicuous pale band on flanks; 2nd dorsal fin and upper caudal fin sometimes dark-edged, and pectoral fins and lower caudal fin sometimes pale-edged, but fins not conspicuously marked; white ventrally. Attains 110 cm TL.

**DISTRIBUTION** Indo-Pacific. WIO: Tanzania to Gulf of Aden, Socotra, Oman, Pakistan, India, Lakshadweep and Sri Lanka; not known from Persian/Arabian Gulf.

**REMARKS** Litters of 1 or 2; size at birth 45–50 cm TL (relatively large); gestation period ~12 months. Males mature at ~69 cm TL, and females at 76–89 cm TL. Coastal, in tropical waters over continental and insular shelves, from close inshore

to 170 m deep. Forms large aggregations segregated by sex. Feeds primarily on small bony fishes, also cephalopods and crustaceans. Regularly caught in subsistence, artisanal and commercial fisheries, using longlines, gillnets and trawls. Harmless to humans. IUCN Red List conservation status Near Threatened globally; Least Concern in Australia.

# Carcharhinus melanopterus

(Quoy & Gaimard 1824)

Blacktip reef shark

PLATE 45

*Carcharias melanopterus* Quoy & Gaimard 1824: 194, Pl. 43, Figs. 1–2 (Papua Barat, Indonesia).

*Carcharhinus melanopterus*: Bass *et al.* 1973\*; Compagno 1984\*; SSF No. 9.13\*; Compagno *et al.* 1989\*; Compagno *et al.* 2005\*; Last & Stevens 2009\*; Voigt & Weber 2011\*; Ebert *et al.* 2021\*.

Moderate-sized; snout short and bluntly rounded, internarial width 0.9–1.1 in preoral length. Eyes horizontally oval. Anterior nasal flaps nipple-shaped. Upper lip folds short and inconspicuous. Upper teeth narrowly triangular, erect to oblique, and with coarse serrations basally; lower teeth erect, with narrower cusps, with fine serrations. Pectoral fins moderately large, narrow, and falcate. No interdorsal ridge. First dorsal fin large, falcate; origin usually over pectoral-fin free rear tips. Second dorsal fin relatively large; origin about over anal-fin origin. Tooth count 23–28/21–27; vertebrae 193–214.

Body pale brownish grey dorsally; conspicuous white band on flanks; 1st dorsal fin and caudal-fin lower lobe with large black apical blotch and brilliantly highlighted proximally with white, and other fins generally with less prominent black tips or thin edges; white ventrally. Attains 180 cm TL.



*Carcharhinus macloti,* 77 cm TL (Kenya). Source: SSF



*Carcharhinus melanopterus*, 70 cm TL, male (St Brandon Shoals). Source: Bass *et al.* 1973

**DISTRIBUTION** Indo-Pacific. WIO: South Africa (KwaZulu-Natal) to Red Sea, Persian/Arabian Gulf, Pakistan, India and Sri Lanka, as well as Madagascar, Comoros, Seychelles, Mauritius, St Brandon Shoals, Maldives and Lakshadweep; possibly enters eastern Mediterranean Sea through Suez Canal.

**REMARKS** Litters of 2–4; size at birth 33–52 cm TL. Males mature at 91–100 cm TL, and females at 96–112 cm TL; biennial reproductive cycle. Occurs in tropical and subtropical waters; common in very shallow inshore areas on or near coral reefs and reef flats, but also found in mangroves, near reef drop-offs and occasionally close offshore. Occurs singly or in small groups or aggregations, but not strongly schooling. Feeds on small fishes, also crustaceans, molluscs and cephalopods. Predators of this species include large groupers and other sharks. Not a major target but regularly caught in fisheries where it occurs. Not regarded as dangerous to humans yet has bitten people swimming or wading in the vicinity of reefs; a popular subject of ecotourism and aquarium displays. IUCN Red List conservation status Vulnerable.

# Carcharhinus obscurus (Lesueur 1818)

Dusky shark

PLATE 45

*Squalus obscurus* Lesueur 1818: 223, Pl. 9 [east coast of USA]. *Carcharhinus iranzae* Fourmanoir 1961: 40, Pl. 13, Fig. C (west coast of Madagascar).

*Carcharhinus obscurus*: Bass *et al.* 1973\*; Van der Elst 1981\*; Compagno 1984\*; SSF No. 9.14\*; Compagno *et al.* 1989\*; Cockcroft 1991; Natanson & Kohler 1996; Ebert 2003\*; Heemstra & Heemstra 2004\*; Compagno *et al.* 2005\*; Last & Stevens 2009\*; Voigt & Weber 2011\*; Ebert *et al.* 2021\*.

Large-sized; snout fairly short and broadly rounded. Anterior nasal flaps low and poorly developed. Upper lip folds short and inconspicuous. Upper anterolateral teeth broad, triangular, rather low, erect to semi-oblique, with fine serrations and no cusplets; lower teeth erect, with narrow cusps. Gill slits long. Pectoral fins large, falcate. Interdorsal ridge low. First dorsal fin moderate-sized, semifalcate; origin over or slightly after pectoral-fin free rear tips. Second dorsal fin small and erect; inner margin moderately long, 1.6–2 times fin height



*Carcharhinus obscurus*, 165 cm TL, male (South Africa). Source: Bass *et al.* 1973

(cf. *C. galapagensis*); origin roughly over anal-fin origin. Tooth count 29–33/27–32; vertebrae 173–194, precaudal vertebrae 89–95.

Body grey or grey-brown dorsally; inconspicuous white band on flanks; most fin tips dusky but not black or white; white ventrally. Attains 4 m TL.

**DISTRIBUTION** Circumglobal in tropical to warm-temperate seas (patchy), including western Mediterranean Sea. WIO: Red Sea to South Africa (strays to southwest coast, but mostly KwaZulu-Natal and Eastern Cape), as well as Socotra and Madagascar.

**REMARKS** Notably slow-growing and late-maturing. Litters of 2-18; size at birth 70-100 cm TL; gestation period 16-22 months, reproductive cycle biennial or triennial. Males mature at 265-290 cm TL, age 17-24 years; females mature at 257-310 cm TL, age 21-32 years; maximum age 40-53 years. Coastal-pelagic, over continental and insular shelves and adjacent oceanic waters, from surf zone to far out to sea, from surface to 400 m. Adults undertake long temperature (seasonal) migrations. Feeds throughout water column on bony fishes, elasmobranchs, cephalopods, crustaceans, and other marine vertebrates and invertebrates; the young are preyed upon by other large sharks. Regularly caught as a bycatch and target species by longline, hook and line, and set bottom nets; the fins are highly valued and among the most sought-after for shark fin soup. Large individuals considered dangerous to humans. IUCN Red List conservation status Endangered.

# Carcharhinus plumbeus (Nardo 1827)

#### Sandbar shark

*Squalus plumbeus* Nardo 1827: 26, 35; also Nardo 1827: 477, 483 (Adriatic Sea).

*Carcharhinus plumbeus*: Bass *et al.* 1973\*; SSF No. 9.15\*; Compagno *et al.* 1989\*; Heemstra & Heemstra 2004; Compagno *et al.* 2005\*; Last & Stevens 2009\*; Voigt & Weber 2011\*; Ebert *et al.* 2021\*.

Medium-sized, stout body; snout short and rounded, shortest distance from nostrils to mouth >2.4 in mouth width. Anterior nasal flaps short, nipple-like. Upper lip folds short and inconspicuous. Upper teeth broad and triangular with high cusps, with serrations and no cusplets; lower teeth smaller, with narrow, erect, serrated cusps. Pectoral fins large, semifalcate. Interdorsal ridge narrow. First dorsal fin very large and triangular, semifalcate; origin over or slightly before pectoral-fin insertions. Second dorsal-fin inner margin short, 1–1.6 times fin height; origin over or slightly before anal-fin origin. Tooth count 27–32/25–32; vertebrae 152–189.

Body grey-brown dorsally; inconspicuous white band on flanks; fin tips and rear edges often dusky; white ventrally. Attains 240 cm TL.

**DISTRIBUTION** Circumglobal in tropical to warm-temperate seas (widespread), including Mediterranean Sea, but possibly not eastern Pacific. WIO: South Africa (Eastern Cape) to Red Sea, Gulf of Oman, Persian/Arabian Gulf, Pakistan and India, as well as Socotra, Madagascar, Seychelles, Mauritius and Lakshadweep.



*Carcharhinus plumbeus*, 118 cm TL, male (South Africa). Source: Bass *et al.* 1973

PLATE 45

**REMARKS** Litters of 1–14; size at birth 56–75 cm TL; gestation period 9-12 months, reproductive cycle possibly every 2 or 3 years. Males mature at 131-178 cm TL, age ~14 years; females mature at 144–183 cm TL, age ~16 years; maximum age 19-25 years. Coastal-pelagic, over continental and insular shelves, adjacent oceanic waters and oceanic banks, from intertidal zone to 280 m deep (typically above 100 m). Also common in shallow muddy or sandy bays, harbours and estuaries but does not enter freshwater; tends to avoid sandy beaches, the surf zone, coral reefs, rough bottom and the surface. Seasonally migratory in some portions of its range. Feeds near seabed on relatively small bony fishes, elasmobranchs, cephalopods, molluscs and crustaceans. A large component of coastal shark fisheries, caught with longlines, hook and line, set bottom nets, and rod-andreel. Not considered dangerous to humans. IUCN Red List conservation status Vulnerable.

### Carcharhinus sorrah (Valenciennes 1839)

Spottail shark

PLATE 46

*Carcharias (Prionodon) sorrah* Valenciennes *in* Müller & Henle 1839: 45, Pl. 16 (Java, Indonesia).

*Carcharhinus sorrah*: Bass *et al.* 1973\*; Compagno 1984\*; SSF No. 9.17\*; Compagno *et al.* 1989\*; Anderson & Ahmed 1993; Compagno *et al.* 2005\*; Fricke *et al.* 2009; Last & Stevens 2009\*; Voigt & Weber 2011\*; Ebert *et al.* 2021\*. Moderately-sized spindle-shaped body; snout moderately long and rounded. Eyes large and circular. Anterior nasal flaps nipple-shaped. Upper lip folds short and inconspicuous. Upper teeth with oblique, moderately narrow cusps, with strong serrations basally, usually only 24–26 rows of upper anteroposterior teeth; lower teeth slightly oblique, narrower, smooth cusps. Pectoral fins fairly small and falcate. Interdorsal ridge low. First dorsal fin moderate-sized, falcate; origin from slightly before to after pectoral-fin free rear tips. Second dorsal fin small and low; inner margin very long, 0.9–1.1 times fin height; origin about opposite anal-fin base. Tooth count 24–26/22–25; vertebrae 153–160.

Body grey dorsally; conspicuous white band on flanks; conspicuous black tip on pectoral fins, 2nd dorsal fin and caudal-fin lower lobe, but 1st dorsal fin with black edge at most; white ventrally. Attains 160 cm TL.

**DISTRIBUTION** Indo-Pacific (patchy). WIO: Persian/Arabian Gulf, Red Sea to South Africa (KwaZulu-Natal), Madagascar, Seychelles, Mascarenes and Maldives.

**REMARKS** Litters of 1–8; size at birth 35–45 cm TL; gestation ~10 months with a reproductive periodicity of 1 year. Males and females mature at 2–3 years. Common; occurs on continental and insular shelves in tropical waters. Feeds mostly on bony fishes and cephalopods (octopus). Regularly caught with line gear and gillnets by artisanal and small-scale commercial fisheries where it occurs. Not dangerous to humans. IUCN Red List conservation status Near Threatened.



*Carcharhinus sorrah*, 111 cm TL, female (W Madagascar). Source: Bass *et al*. 1973

# GENUS **Glyphis** Agassiz 1843

Large-sized stocky body; snout short and broadly rounded, preoral length much less than mouth width. Eyes tiny; no spiracles. Nostrils small; anterior nasal flaps short, triangular. Lip folds short, confined to corners of mouth. Teeth strongly differentiated in upper and lower jaws; upper teeth usually erect, broad, triangular, with finely serrated cusps and no cusplets; lower teeth with or without cusplets or blades, but with variably oblique to erect long cusps, generally with no serrations, and cusps prominently protruding when mouth closed; tooth counts 29-37/28-34. Pectoral fins moderately broad, falcate or semifalcate. No interdorsal ridge. First dorsal-fin origin far anterior, over rear half of pectoral-fin bases, its free rear tip well before pelvic fins. Second dorsal fin smaller than 1st, its origin slightly before anal-fin origin. Anal fin slightly smaller than 2nd dorsal fin; preanal ridges very short or absent; rear margin concave to deeply concave. No lateral keels on peduncle; upper precaudal pit longitudinal and not crescentic. Body grey or brownish dorsally, white ventrally, and without conspicuous markings. Found in turbid brackish freshwater of rivers, estuaries and possibly adjacent marine areas. Possibly 5 species, all with small, fragmented populations in Indo-Pacific; 1 species in WIO.

### Glyphis gangeticus (Müller & Henle 1839)

### Ganges shark

Carcharias (Prionodon) gangeticus Müller & Henle 1839: 39, Pl. 13 (Ganges River at Hooghly, India [Lower Sundarbans, Bangladesh]). Glyphis gangeticus: Compagno 1984\*; Compagno et al. 2005\*;

Ebert et al. 2021\*.

Preorbital snout rather long; preoral snout much shorter than mouth width. First dorsal-fin origin over rear of pectoral bases. Second dorsal fin  $\sim \frac{1}{2}$  height of 1st dorsal fin. First few anterior teeth in lower jaw with cutting edges along entire cusp, giving cusps claw-like shape, and with low cusplets. Tooth count 30-37/31-34; vertebrae 169.

Body grey dorsally, without markings; white ventrally. Attains at least 204 cm TL.

**DISTRIBUTION** India (east coast) and Bangladesh (lower reaches of Ganges-Hooghly river system, West Bengal), but may occur in other river systems or shallow estuaries in the area, and possibly in Pakistan (vicinity of Karachi) and northern India in WIO.



**REMARKS** Biology little known, but probably viviparous; newborn specimens 56–61 cm TL. Rare, elusive; freshwater riverine and possibly inshore marine and estuarine habitat. Apparently fished in the Ganges-Hooghly river system. Reputed to have attacked humans, but such incidents most likely involved the Zambezi shark (*Carcharhinus leucas*) which occurs in the same area. IUCN Red List conservation status Critically Endangered.

# GENUS Lamiopsis Gill 1862

Moderate-sized, rather stocky body; snout moderately long, preoral length nearly equal to or slightly greater than mouth width. Eyes fairly small; no spiracles. Nostrils small; anterior nasal flaps short, broadly triangular. Lip folds confined to corners of mouth. Teeth generally differentiated in upper and lower jaws; upper anteroposterior teeth with more or less erect, broad, triangular cusps, with irregular serrations and no cusplets or blades; lower teeth with oblique to erect, long, hooked cusps, with no serrations, and cusps slightly protruding when mouth closed. Pectoral fins very broad and triangular; origins under 4th or 5th gill slits; distance from origin to free rear tip ><sup>3</sup>/<sub>4</sub> length of anterior margin. No interdorsal ridge. First dorsal-fin origin over pectoral-fin inner margins; midbase slightly closer to pectoral-fin bases than to pelvic-fin bases; free rear tip over or slightly before or after pelvic-fin origins. Second dorsal fin >3/4 height of 1st; origin about opposite anal-fin origin. Anal fin smaller than 2nd dorsal fin; preanal ridges very short or absent; rear margin slightly concave. No lateral keels on peduncle; upper precaudal pit longitudinal and not crescentic. Tooth counts 29-37/28-34. Two species, 1 in WIO.



Lamiopsis temminckii. © Food and Agriculture Organization of the United Nations, Original Scientific Illustrations Archive; reproduced with permission

# Lamiopsis temminckii (Müller & Henle 1839)

#### Broadfin shark

*Carcharias (Prionodon) temminckii* Müller & Henle 1839: 48, Pl. [18] (India).

Lamiopsis temminckii: Setna & Sarangdhar 1949; Compagno 1984\* [as temminicki]; Manilo & Bogorodsky 2003 [as temminicki]; Compagno et al. 2005\*; White et al. 2010\*; Ebert et al. 2021\*.

Fifth gill slit ~<sup>1</sup>/<sub>2</sub> length of 1st gill slit. Upper teeth with broad triangular cusps, with uneven serrations, and usually 5–7 posterior rows of molariform teeth; lower teeth hookshaped, with narrow cusps and no serrations; tooth count 29–37/28–34.

Body pale grey or brownish dorsally; pale ventrally. Attains 168 cm TL.

**DISTRIBUTION** Indo-Pacific. WIO (patchy): Pakistan, India, Lakshadweep and Maldives, but not known from off Sri Lanka in Gulf of Mannar.

**REMARKS** Litters of 4–8; size at birth 40–60 cm TL; gestation period 8 months. Occurs mostly close inshore, in brackish and marine waters. Likely feeds on small fishes and invertebrates. Rare throughout most of its range, but once common off west coast of India. Taken by local fishermen in bottom and floating gillnets and with line gear; used for its meat, fins and liver oil. Not dangerous to humans. IUCN Red List conservation status Endangered.

# GENUS Loxodon Müller & Henle 1838

Small-sized, slender body with distinctly long snout. Head fairly narrow and moderately depressed; snout narrowly parabolic in dorsoventral view, preoral length greater than internarial space and mouth width. Eyes large, with posterior notch; spiracles minute. Anterior nasal flaps very short and narrowly triangular. Lip folds short, confined to corners of mouth. Pectoral fins relatively short, moderately broad and triangular, slightly falcate; distance between pectoraland pelvic-fin bases 2–3 times that of 1st dorsal-fin base. Interdorsal ridge absent or rudimentary. First dorsal-fin origin behind pectoral-fin free rear tips, its midbase roughly equidistant between pectoral- and pelvic-fin bases, and its free rear tip usually before pelvic-fin origins (but occasionally over them). Second dorsal fin much smaller than 1st (<¼ height of 1st dorsal fin); origin roughly over anal-fin insertion. Anal fin larger than 2nd dorsal fin; preanal ridges long; rear margin concave. No keels on peduncle; upper precaudal pit transverse and crescentic. One species.

### Loxodon macrorhinus Müller & Henle 1839

Sliteye shark

PLATE 46

Loxodon macrorhinus Müller & Henle 1839: 61, Pl. [25] [probably Indian Ocean]; Springer 1964\*; Bass *et al.* 1975\*;

Compagno 1984\*; Compagno *et al.* 1989\*; Compagno *et al.* 2005\*; Last & Stevens 2009\*; Ebert *et al.* 2021\*.

Scoliodon ceylonensis: Setna & Sarangdhar 1946; Bonfil & Abdallah 2004\*.

Diagnosis as for genus. Teeth smooth-edged, similar in both jaws; anteroposterior teeth with fairly slender, highly oblique cusps, with distal blades but no cusplets or serrations. Tooth count 25–28/24–28; vertebrae 148–191.

Body grey to brownish dorsally; 1st dorsal fin and caudal fin with thin black margins, and pectoral fins with pale posterior margins; pale ventrally. Attains 99 cm TL.



Loxodon macrorhinus, 75 cm TL, adult male (S Mozambique). Source: Bass et al. 1975

**DISTRIBUTION** Indo-Pacific. WIO: South Africa (KwaZulu-Natal) to Red Sea, Oman, Pakistan, India and Sri Lanka, as well as Socotra, Madagascar, Seychelles and Maldives; not known from Persian/Arabian Gulf.

**REMARKS** Litters of 2–4; size at birth 40–45 cm TL. Males mature at 62–66 cm TL, and females at 79–99 cm TL. Occurs in moderately shallow, clear waters of continental and insular shelves, at 7–100 m. Feeds on small bony fishes, crustaceans (shrimp) and occasionally cephalopods (cuttlefish). Commonly caught in artisanal and small-scale commercial fisheries by gillnets and longlines; used for its meat and fins. Harmless to people.

### GENUS Negaprion Whitley 1940

Large-sized, moderately stout, fusiform body; snout short, broadly rounded or angular; preoral length subequal to internarial space and much less than mouth width. Eyes small; no spiracles. Lip folds confined to corners of mouth. Teeth strongly differentiated in upper and lower jaws; upper anteroposterior teeth with more or less erect, moderately slender cusps, with mostly smooth edges and generally low cusplets; lower teeth with mostly erect, fairly long, slightly hooked cusps, with smooth edges and no cusplets, and cusps not protruding when mouth closed. Pectoral fins broad and triangular or falcate; distance from origin to free rear tip  $>\frac{2}{3}$  length of anterior margin. No interdorsal ridge. First dorsal-fin origin over or behind pectoral-fin free rear tips; midbase closer to pelvic-fin bases than to pectoral-fin bases; free rear tip more or less over pelvic-fin origins. Second dorsal fin at least four-fifths height of 1st dorsal fin; origin about opposite anal-fin origin. Anal fin somewhat smaller than 2nd dorsal fin; preanal ridges hardly developed; rear margin deeply concave. No lateral keels on peduncle; upper precaudal pit longitudinal and not crescentic. Tooth counts 27–33/27–33. Two species, 1 in WIO.

# Negaprion acutidens (Rüppell 1837)

Sharptooth lemon shark

*Carcharias acutidens* Rüppell 1837: 65, Pl. 18, Fig. 3 (Jeddah, Saudi Arabia, Red Sea).

PLATE 46

*Odontaspis madagascariensis* Fourmanoir 1961: 15, Pl. 3 (Madagascar). *Negaprion acutidens*: Gohar & Mazhar 1964; Bass *et al.* 1975\*;

SSF No. 9.29\*; Compagno *et al.* 1989\*; Compagno *et al.* 2005\*; Last & Stevens 2009\*; Ebert *et al.* 2021\*.

Snout broad and blunt. Pectoral fins falcate. Tooth count 28–32/28–30; vertebrae 217–229.

Body yellowish brown to pale brown or grey dorsally; whitish ventrally. Attains 3.1 m TL.



Negaprion acutidens, 243 cm TL, mature male (South Africa). Source: Bass et al. 1975

**DISTRIBUTION** Indo-Pacific. WIO (widespread): South Africa (KwaZulu-Natal) to Red Sea, Pakistan, India and Sri Lanka, as well as Socotra, Madagascar, Seychelles, Mauritius, Maldives and Lakshadweep; not known from Persian/Arabian Gulf.

**REMARKS** Litters of 1–14; size at birth 45–80 cm TL; gestation period 10–11 months. Matures at 220–240 cm TL. Occurs on continental and insular shelves in tropical waters, on or near bottom, from intertidal zone to at least 30 m deep. Prefers bays, estuaries, sandy plateaus, outer-reef shelves at moderate depths, reef lagoons and mangrove estuaries; young commonly found on very shallow reef flats. Sluggish, slow swimmer, often found in turbid, still water; shy, but potentially dangerous to humans if provoked. Feeds on benthic bony fishes, elasmobranchs (stingrays) and cephalopods. Caught in Pakistan and India with longlines and gillnets; used for its meat, fins and liver oil. IUCN Red List conservation status Vulnerable globally; Southeast Asian subpopulation Endangered.

### GENUS Prionace Cantor 1849

Large-sized, slender body, with long head and very long, conical snout; preoral length greater than internarial space and mouth width. Eyes large; no spiracles. Nostrils small; internarial space ~2.5–3 times nostril width; anterior nasal flaps very short and broadly triangular, and not tubular. Lip folds minute. Pectoral fins long and narrow (scythe-like); origins under 3rd and

4th gill slits. No interdorsal ridge. First dorsal fin well behind pectoral-fin free rear tips, its midbase closer to pelvic-fin bases, and its free rear tip slightly before pelvic-fin origins. Second dorsal fin much smaller than 1st (<½ height of 1st dorsal fin). Anal fin slightly larger than 2nd dorsal fin; preanal ridges short; rear margin deeply concave. Peduncle with weak keels; upper precaudal pit transverse and crescentic. Caudal fin narrow, and lower lobe long. One species.

# Prionace glauca (Linnaeus 1758)

Blue shark

PLATE 47

Squalus glaucus Linnaeus 1758: 235 (Mediterranean Sea and NE Atlantic). Prionace glauca: Fourmanoir 1961; Bass et al. 1975\*; Gubanov & Grigor'yev 1975; Compagno 1984\*; SSF No. 9.32\*; Compagno et al. 1989\*; Ebert 2003\*; Heemstra & Heemstra 2004\*; Compagno et al. 2005\*; Last & Stevens 2009\*; Ebert et al. 2021\*.

Diagnosis as for genus. Teeth well-differentiated in upper and lower jaws; upper anteroposterior teeth triangular, curved, with erect to oblique cusps, with serrations and no blades or cusplets (except in very young specimens); lower teeth with slender cusps, no blades or cusplets, and serrations variably developed, and cusps not prominently protruding when mouth closed. Tooth count 24–31/25–34.

Body deep blue dorsally, with bright blue to silvery blue flanks (fades to grey soon after death), and pure white ventrally; pectoral-fin tips dusky. Attains 4 m TL, possibly more.



Prionace glauca, 279 cm TL, mature male (South Africa). Source: Bass et al. 1975

**DISTRIBUTION** Cosmopolitan in tropical to temperate seas, including Mediterranean Sea and throughout WIO, but not known from Red Sea and Persian/Arabian Gulf.

**REMARKS** Litters of 4–135 (average ~35); size at birth 35-60 cm TL; gestation period 9-12 months. Males mature at 183-218 cm TL, and females at 183-221 cm TL; fecund and fast-growing, matures at 4-8 years; maximum age ~20 years. Females have much thicker skin and often bear bite wounds as mating scars. Possibly the most wide-ranging shark, with complex movements; oceanic and pelagic, from surface to ~1 000 m deep, but occasionally ventures inshore (where continental shelf is narrowest); also makes long migrations by swimming slowly with major transoceanic currents. Graceful swimmer, observed cruising at surface with dorsaland caudal-fin tips out of the water, and may form large aggregations to feed. Feeds on relatively small bony fishes and sharks, cephalopods (squid) and other invertebrates (pelagic red crabs), and occasionally seabirds and cetacean carrion. Heavily fished and used mostly for its fins; taken mainly as a bycatch with pelagic longlines, driftnets, and occasionally bottom trawls. Often timid, but potentially dangerous to humans. IUCN Red List conservation status Near Threatened.

# GENUS Rhizoprionodon Whitley 1929

Small-sized, fairly slender to moderately stout, fusiform body; head moderately depressed; snout long, narrowly to broadly parabolic or obtusely wedge-shaped in dorsoventral view. Eyes large, without posterior notch; no spiracles. Nostrils small, wide-spaced; preoral length greater than internarial space and mouth width; anterior nasal flaps very short, narrowly triangular. Lip folds short to rather long; upper and lower folds unequal in length. Teeth similar in both jaws; anteroposterior teeth with relatively slender, oblique cusps, and with distal blades, no cusplets, and serrations variably developed. Pectoral fins relatively short, moderately broad and triangular, slightly falcate. Interdorsal ridge absent or rudimentary. First dorsalfin origin over pectoral-fin free rear tips; midbase about equidistant between pectoral- and pelvic-fin bases or closer to pectoral fins; free rear tip slightly before pelvic-fin origins. Second dorsal fin much smaller than 1st (<1/3 height of 1st dorsal fin); origin just before or over anal-fin insertion. Anal fin larger than 2nd dorsal fin; preanal ridges very long; rear margin shallowly to deeply concave. No keels on peduncle; upper precaudal pit transverse and crescentic. Body pale grey, yellowish or brownish grey dorsally, pale ventrally, and without colour patterns. Tooth counts 23-31/21-28; vertebrae 121-170. Seven species, 2 in WIO.

### **KEY TO SPECIES**

1a	Upper lip folds long and rather prominent (usually as long as or longer than lower folds); tooth rows more numerous on
	average: mostly >25/24
1b	Upper lip folds reduced and often inconspicuous (usually shorter than lower folds); tooth rows fewer on average: mostly <25/24
# Rhizoprionodon acutus (Rüppell 1837)

#### Milk shark

PLATE 47

*Carcharias acutus* Rüppell 1837: 65, Pl. 18, Fig. 4 (Jeddah, Saudi Arabia, Red Sea).

Rhizoprionodon acutus: Springer 1964; Bass et al. 1975\*; Van der Elst 1981\*; Compagno 1984\*; SSF No. 9.33\*; Compagno et al. 1989;
Heemstra & Heemstra 2004\*; Compagno et al. 2005\*; Last & Stevens 2009\*; Ebert et al. 2021\*.

Upper lip folds long and prominent (as long as or longer than lower folds); prenarial snout long, 4–5.4% TL; usually >8 enlarged hyomandibular pores on each side of head just behind angle of mouth. Teeth generally smooth-edged, but with some irregular low cusplets. Tooth count 23–27/23 or 24; vertebrae 121–162.

Body greyish or bronzy dorsally; pectoral fins with pale margins, and 1st dorsal fin and caudal-fin upper lobe sometimes dark-edged; white ventrally. Attains 178 cm TL.

**DISTRIBUTION** Eastern Atlantic, Mediterranean Sea (one record off Italy) and Indo-Pacific (widespread). WIO: South Africa (KwaZulu-Natal) and Madagascar to Red Sea, Socotra, Persian/Arabian Gulf, Pakistan, India, Sri Lanka, Lakshadweep and Maldives.

**REMARKS** Litters of 1–8 (usually 2–5); size at birth 25–39 cm TL; gestation period ~12 months. Males mature at 68–72 cm TL, and females at 70–81 cm TL, both sexes at

~2 years; maximum age at least 8 years. Abundant, inshore and offshore over continental shelf, from shore to ~200 m deep; often found off sandy beaches, at midwater or near bottom, and sometimes in estuaries, but does not tolerate low salinities or range into freshwater. Feeds primarily on bony fishes, but also squid and crustaceans. Often preyed on by larger sharks. One of the most common sharks in inshore waters where it occurs; taken by artisanal and small-scale commercial fisheries and offshore fishing fleets by longline, hook and line and bottom trawl; used for its meat and fins, and as fish meal. Harmless to people. IUCN Red List conservation status Vulnerable.

# Rhizoprionodon oligolinx Springer 1964

Grey sharpnose shark

PLATE 48

*Rhizoprionodon oligolinx* Springer 1964: 621, Pl. 2, Figs. 12–13 (Gulf of Thailand); Nair *et al.* 1974; Compagno *et al.* 2005\*; Last & Stevens 2009\*; Ebert *et al.* 2021\*.

Interdorsal ridge sometimes present. Upper lip folds often inconspicuous, usually shorter than lower folds; prenarial snout short, 3.7–4.7% TL; 3–8 enlarged hyomandibular pores on each side of head just behind angle of mouth. Teeth small, with oblique cusps, with no serrations; tooth count 23–25/ 21–24. Vertebrae 151–164, precaudal vertebrae 84–91.

Body grey to bronzy grey dorsally; fins with inconspicuous dusky edges; pale ventrally. Attains 70 cm TL.



Rhizoprionodon acutus, 99 cm TL, mature female (South Africa). Source: Bass et al. 1975



*Rhizoprionodon oligolinx.* © Food and Agriculture Organization of the United Nations, Original Scientific Illustrations Archive; reproduced with permission

**DISTRIBUTION** Indo-Pacific. WIO: Arabian Sea and Persian/ Arabian Gulf to Pakistan, India, Sri Lanka and Maldives.

**REMARKS** Common but poorly known. Litters of 3–5; size at birth 21–26 cm TL. Males mature at 29–38 cm TL, and females at 32–41 cm TL. Abundant, inshore and offshore, over continental and insular shelves. Likely feeds on small bony fishes, cephalopods and crustaceans. Probably commonly taken in subsistence, artisanal and commercial fisheries throughout its range, by gillnet, longline and trawl; used for its meat and fins.

GENUS Scoliodon Müller & Henle 1837

Small-sized, stocky body with compressed head; snout long and flattened, laterally expanded and trowel-shaped, preoral length greater than internarial space and mouth width. Eyes

small; no spiracles. Anterior nasal flaps short and narrowly

triangular. Lip folds short to rudimentary. Teeth similar in

oblique cusps, with distal blades but no cusplets or serrations.

both jaws; anteroposterior teeth small, with very slender,

Pectoral fins very short, broad and triangular. Interdorsal

ridge absent or rudimentary. First dorsal-fin origin over or behind pectoral-fin free rear tips; midbase closer to pelvic-fin bases than to pectoral-fin bases; free rear tip approximately over pelvic-fin midbase. Second dorsal fin much smaller than 1st (<<sup>1</sup>/<sub>3</sub> height of 1st dorsal fin); origin behind anal-fin midbase. Anal fin much longer than 2nd dorsal fin; preanal ridges short; rear margin straight or slightly concave. No keels on peduncle; upper precaudal pit transverse and crescentic. Two species, 1 in WIO.

# Scoliodon laticaudus Müller & Henle 1838

Spadenose shark

PLATE 48

Scoliodon laticaudus Müller & Henle 1838: 27, 28, Pl. [8] (India);
 Müller & Henle 1839\*; Compagno 1984\*; Compagno et al. 2005\*;
 White et al. 2006\*; Ebert et al. 2021\*.

No interdorsal ridge; 1st dorsal-fin origin behind pectoralfin free rear tips. Anal fin very long, and rear margin nearly straight. Tooth count 25–33/24–34; vertebrae 148–171.

Body pale grey, yellowish or bronzy grey dorsally; white ventrally. Attains 74 cm TL.



Scoliodon laticaudus. © Food and Agriculture Organization of the United Nations, Original Scientific Illustrations Archive; reproduced with permission

**DISTRIBUTION** Indo-Pacific. WIO: Tanzania to Somalia, Socotra, Arabian Sea, Persian/Arabian Gulf, Pakistan, India, Lakshadweep and Sri Lanka; not known to enter Gulf of Aden and Red Sea.

**REMARKS** Litters of 1–14; size at birth 12–15 cm TL. Males mature at 24–36 cm TL, and females at 33–35 cm TL, at age 1–2 years for both sexes; maximum age 5–6 years. Occurs close inshore over continental and insular shelves, often in large schools; enters upper reaches of rivers in parts of Southeast Asia. Feeds on small pelagic schooling or benthic bony fishes, crustaceans (mantis shrimp, crabs) and cephalopods (cuttlefish). Abundant and heavily fished; commonly taken in Pakistan and India in artisanal and commercial fisheries by trawl lines, longlines, gillnets and traps; used for its meat. IUCN Red List conservation status Near Threatened.

## GENUS Triaenodon Müller & Henle 1837

Moderate-sized, fairly slender body with relatively broad head; snout short and bluntly rounded, preoral length subequal to internarial space and much less than mouth width. Spiracles usually absent (may be present as minute pores or slit-like openings). Nostrils small; internarial space ~3 times nostril width; anterior nasal flaps slightly elongated, distally truncated, and forming distinct tubes with mesonarial flaps. Lip folds confined to corners of mouth. Pectoral fins fairly broad and triangular. No interdorsal ridge. First dorsal-fin origin well behind pectoral-fin free rear tips; midbase much closer to pelvic-fin bases than to pectoral-fin bases; free rear tips end slightly after pelvic-fin origins. Second dorsal fin large but distinctly smaller than 1st (½–¾ height of 1st dorsal fin); origin about opposite anal-fin origin. Anal fin approximately subequal to 2nd dorsal fin; preanal ridges short; rear margin deeply concave. No lateral keels on peduncle; upper precaudal pit transverse and crescentic. One species.

# Triaenodon obesus (Rüppell 1837)

Whitetip reef shark

PLATE 48

*Carcharias obesus* Rüppell 1837: 64, Pl. 18, Fig. 2 (Jeddah, Saudi Arabia, Red Sea).

*Triaenodon obesus*: Klausewitz 1960\*; Fourmanoir 1961; Gohar & Mazhar 1964; Bass *et al.* 1975\*; Compagno 1984\*; SSF No. 9.35\*; Compagno *et al.* 1989\*; Compagno *et al.* 2005\*; Ebert *et al.* 2021\*.

Diagnosis as for genus. Mouth slanted downward in lateral view. Eyes horizontally oval. Teeth similar in both jaws, with erect to semi-oblique, triangular central cusps, flanked by pair of conspicuous cusplets or pointed blades, and no serrations; tooth count 42–50/42–48. Vertebrae 208–214.

Dorsal and lateral surfaces of head, trunk and tail greyish brown, becoming paler ventrally; flanks usually with few scattered dark spots; conspicuous white tip on 1st dorsal fin and caudal-fin upper lobe, and sometimes 2nd dorsal fin and caudal-fin lower lobe, as well as underside of pectoral-fin tips. Attains 213 cm TL.



Triaenodon obesus, 139 cm TL, mature male (South Africa). Source: Bass et al. 1975

**DISTRIBUTION** Indo-Pacific. WIO: South Africa (KwaZulu-Natal) to Red Sea, Gulf of Oman, Pakistan, India and Sri Lanka, as well as Madagascar, Comoros, Aldabra, Seychelles, Maldives and Lakshadweep; not known from Persian/Arabian Gulf.

**REMARKS** Litters of 1–5; size at birth 52–60 cm TL. Matures at 104–109 cm TL, with mating having been observed in the wild; maximum age at least 25 years. Occurs on continental and insular shelves, to ~330 m deep (commonly <40 m); associated with reefs and coral lagoons, in clear waters, on or near bottom and resting in caves and crevices, and occupies a small home range. Feeds on small bony fishes, cephalopods (octopus) and crustaceans. Caught in line and net trawl fisheries near shallow reef areas; used for its fins. Popular subject of ecotourism, may be curious and approach divers, and occasionally aggressive towards humans when food stimulus present. IUCN Red List conservation status Vulnerable.

# FAMILY SPHYRNIDAE

## Hammerhead sharks

Neil C Aschliman and Dave A Ebert

Small- to large-sized with distinctive laterally expanded head shaped like double mallet in profile. Eyes circular or nearly so, with internal nictitans; no spiracles. Anterior nasal flaps short and triangular, not barbel-like; internarial width usually 7–14 times nostril width (but <1.3 times nostril width in Eusphyra, which has greatly expanded nostrils). Lip folds vestigial or absent. Teeth small to moderately large, more or less bladelike, with acute and narrow to moderately broad cusps and no lateral cusplets; teeth weakly differentiated in upper and lower jaws; tooth counts 24-37/25-37. First dorsal fin moderate to large, tall, not keel-like, and much shorter than caudal fin; 1st dorsal-fin base ahead of pelvic-fin bases, varying from equidistant between pectoral- and pelvic-fin bases to closer to pectoral fins. Second dorsal fin much smaller than 1st, its size equal to or slightly smaller than anal fin. Pectoral fins with transverse radials extending into distal web. Caudal-fin lunate, lower lobe strong; undulations or ripples in caudal-fin dorsal margin; precaudal pits present. Intestinal valve scrolled.

The unique flattened and laterally expanded prebranchial head (cephalofoil) of hammerhead sharks has been interpreted as a bow plane that greatly increases their manoeuverability and may also increase their sensory capacity, as the wide-spaced eyes enhance binocular vision, the expanded nasal capsules allow more acute and directional olfaction, and the increased head area allows more extensive lateralline canals and ampullae of Lorenzini for pressure and electromagnetic senses.

Reproductive mode viviparity with yolk-sac placenta. Size range ~1.5–6 m TL. Found from surface, including surfline and intertidal zone, to at least 275 m deep. Common and wideranging in all warm-temperate to tropical seas, in continental and insular waters, on or adjacent to continental shelf, but none truly oceanic; some may enter estuaries. Considered potentially dangerous to people, but rare unprovoked attacks have been attributed to only a few of the species; most are shy and difficult to approach. Two genera and 8 species; both genera and 4 species in WIO.

#### **KEY TO GENERA**



Drawing 1b sourced from Compagno 1984, © FAO

# GENUS **Eusphyra** Gill 1862

Head immensely laterally expanded and arrow-shaped in dorsoventral view, with bumps along front margin opposite nostrils; lateral blades of head narrow, wing-like; head width 40–50% TL. Nostrils enlarged. Teeth small, smooth-edged, oblique and similar in both jaws. First dorsal fin anteriorly set, its origin over pectoral-fin insertions. Anal-fin rear margin shallowly concave. Upper precaudal pit longitudinal, not crescentic. One species.



Eusphyra blochii © Food and Agriculture Organization of the United Nations, Original Scientific Illustrations Archive; reproduced with permission

# Eusphyra blochii (Cuvier 1816)

#### Winghead shark

PLATE 49

*Zygaena blochii* Cuvier 1816: 127 [no locality given] [based on Bloch 1785: Pl. 117 and brief description].

Eusphyra blochii: Compagno 1984\*; Compagno et al. 2005\*.

Diagnosis as for genus. Nostril width 0.8–0.9 times internarial width and nearly twice mouth width. Tooth count 30–31/29; vertebrae 108–124.

Body grey or grey-brown dorsally, paler ventrally; no dark fin markings. Attains 186 cm TL.

**DISTRIBUTION** Indo-Pacific. WIO: Persian/Arabian Gulf to India and Sri Lanka.

**REMARKS** Common, distinctive, but poorly known. Litters of 6–25; size at birth 32–45 cm TL; gestation period 8–11 months. In Indian waters, young are born just before monsoon season, in April and May, and mating appears to occur from June to August. Males mature at ~108 cm TL, and females at ~120 cm TL. Found in shallow tropical waters of continental and insular shelves. Diet includes small fishes, cephalopods and crustaceans. Commonly fished throughout its range; caught with gillnets, stake nets, seines, floating and bottom longlines, and probably on hook and line; exploited for its meat and fins. Relatively harmless to people. IUCN Red List conservation status Endangered.

# GENUS **Sphyrna** Rafinesque 1810

Moderately broad, spade-, mallet- or axe-shaped head in dorsoventral view, and without bumps along front margin; lateral blades of head broad, not wing-like; head width ¼-⅓ TL. Nostrils short, nostril width 7–14 times internarial width and less than half mouth width. First dorsal-fin origin over or behind pectoral-fin insertions. Upper precaudal pit transverse and crescentic. Circumglobal in all warm-temperate and tropical seas. Seven species, 3 in WIO.

#### **KEY TO SPECIES**





Drawings sourced from CFSA



*Sphyrna lewini*, 47 cm TL, immature female (South Africa). Source: Bass *et al*. 1975

## Sphyrna lewini (Griffith & Smith 1834)

#### Scalloped hammerhead

PLATE 50

*Zygaena lewini* Griffith & Smith 1834: 640, Pl. 50 (south coast of New Holland [southern Australia]).

Sphyrna lewini: Bass et al. 1975\*; Compagno 1984\*; SSF No. 13.1\*; Compagno et al. 1989\*; Smale & Cliff 1998.

Large-sized, with front margin of head curved, and with prominent median and lateral indentations; rear margins of head long, angled posterolaterally, and generally broader than mouth width. Preoral snout length 1/5-1/3 head width; welldeveloped prenarial grooves in front of nostrils. Rear edge of eyes slightly ahead of upper symphysis of mouth. Anterior teeth with moderately long, smooth or occasionally weakly serrated cusps; posterior teeth mostly cuspidate and not keeled or molariform. First dorsal fin moderately falcate, its origin above or slightly behind pectoral-fin insertions, and free rear tip anterior to pelvic-fin origins. Second dorsal fin low, less than anal-fin height, with shallowly concave rear margin, and inner margin long, about twice fin height, and ending slightly before upper caudal origin. Pelvic fins not falcate, with straight to slightly concave rear margins. Anal fin larger than 2nd dorsal fin, and moderately long, its base 4.3-6.4% TL; anal-fin origin well ahead of 2nd dorsal-fin origin, and rear margin deeply concave. Tooth count 30-36/30-35; vertebrae 174-209.

Body pale grey-brown to bronzy dorsally, white ventrally; pectoral-fin tips dusky to black. Attains 4.2 m TL.

**DISTRIBUTION** Circumglobal in warm-temperate to tropical seas. WIO: South Africa (KwaZulu-Natal) northwards to Red Sea, Persian/Arabian Gulf and Gulf of Oman, and eastwards to Seychelles, Maldives, India and Sri Lanka.

**REMARKS** Genetic evidence suggests multiple subpopulations. Litters of 15-31; size at birth 42-55 cm TL. Males mature at 140-165 cm TL, at ~10 years; females mature at ~212 cm TL, at ~15 years; maximum age ~35 years. Coastalpelagic and semioceanic, over continental and insular shelves and in adjacent deep water, from intertidal zone and surface to at least 275 m deep, but often approaching close inshore and entering enclosed bays and estuaries. Forms large schools at different stages of its lifecycle, though solitary individuals of both young and adults also occur. Feeds on a wide variety of fishes, but also invertebrates (especially cephalopods). Perhaps the most common hammerhead shark in the tropics and once abundantly available to inshore artisanal and small commercial fisheries as well as offshore operations; caught as a bycatch and in target fisheries (including illegally targeted) with a variety of gear; used less often for its meat than for its highly valued fins. Young and sometimes adults are displayed at large aquaria. IUCN Red List conservation status Critically Endangered globally, due to heavy fishing pressure in many regions.

# Sphyrna mokarran (Rüppell 1837)

Great hammerhead

PLATE 50

*Zygaena mokarran* Rüppell 1837: 66, Pl. 17, Fig. 3 (Massawa, Eritrea, Red Sea).

Sphyrna mokarran: Bass et al. 1975\*; Compagno 1984\*; SSF No. 13.2\*; Compagno et al. 1989\*; Cliff 1995; Smale & Cliff 1998.

Large-sized to gigantic, with front margin of head nearly straight in adults (but broadly arched in young), and with prominent median and lateral indentations; rear margins of head long, and transverse in adults but inclined posterolaterally in young, and about as broad as mouth width. Preoral snout length <1/3 head width; prenarial grooves absent or weakly developed. Rear edge of eyes before upper symphysis of mouth. Anterior teeth with moderately long, stout cusps and strongly serrated edges. First dorsal fin strongly falcate, origin over or slightly behind pectoral-fin insertions, free rear tip before pelvic-fin origins. Second dorsal fin moderately high, about equal to anal-fin height, with strongly concave rear margin, and short inner margin (about equal to fin height) ending slightly in front of upper caudal origin. Pelvic fins strongly falcate, with strongly concave rear margins. Anal fin moderately long, about as large as 2nd dorsal fin, its base 5.6-7.3% TL; anal-fin origin ahead of 2nd dorsal-fin origin, and rear margin deeply concave. Tooth count 36-37/34-35; vertebrae 197-212.

Body bronzy to grey-brown dorsally, pale ventrally; no fin markings except 2nd dorsal fin dark-tipped in juveniles. Attains 6.1 m TL, possibly more. **DISTRIBUTION** Circumtropical and some warm temperate waters. WIO: South Africa (KwaZulu-Natal) to Red Sea, Persian/Arabian Gulf, Gulf of Oman, Madagascar, Comoros, Seychelles, Mauritius, Maldives, India and Sri Lanka.

**REMARKS** Litters of 6–42; gestation period at least 7 months, with birth occurring in late spring or summer in the Northern Hemisphere; size at birth 50-70 cm TL. Males mature at 234–269 cm TL, and females at 250–300 cm TL. Most adults of either sex do not exceed ~3.6 m TL, but a small percentage of the population (mainly or entirely females) attain a size much greater than the adult average. Apparently nomadic and seasonally migratory, and generally regarded as solitary. Coastal-pelagic and semi-oceanic, found from close inshore to well offshore, over continental shelf and island terraces, in passes and lagoons of coral atolls, and over deep water near land; often favours continental and insular coral reefs, from near surface to >80 m deep. Feeds on a variety of prey, especially stingrays and other batoids, groupers and sea catfishes. Interest to fisheries more limited than occurs for other large hammerhead sharks; although less abundant than S. lewini (which forms large schools) it is regularly caught in the tropics, especially by longline, and is favoured for its largesized fins. Believed to be potentially more dangerous to people than other hammerhead sharks, though few if any unprovoked attacks have been definitely attributed to it because of the difficulty in distinguishing the species involved in encounters with people. IUCN Red List conservation status Critically Endangered, due to fishing pressure in some regions.



Sphyrna mokarran, 187 cm TL, immature male (South Africa). Source: Bass et al. 1975

# Sphyrna zygaena (Linnaeus 1758)

Smooth hammerhead

PLATE 51

*Squalus zygaena* Linnaeus 1758: 234 (Mediterranean Sea and North Atlantic).

*Sphyrna zygaena*: Bass *et al.* 1975\*; Compagno 1984\*; SSF No. 13.3\*; Compagno *et al.* 1989\*; Cliff 1995; Smale & Cliff 1998.

Large-sized with front margin of head curved forward, and with prominent lateral indentations but no median indentation; rear margins of head long, angled posterolaterally, and generally broader than mouth width. Preoral snout 1/5-1/3 head width; well-developed prenarial grooves anteromedially to nostrils. Rear edge of eyes slightly behind upper symphysis of mouth. Teeth relatively small; anterior teeth with moderately long, very stout cusps, and smooth or weakly serrated edges. First dorsal fin moderately falcate, origin over pectoral-fin insertions, and free rear tip well anterior to pelvic-fin origins. Second dorsal fin low, less than anal-fin height, with shallowly concave rear margin and long inner margin (about twice fin height) ending in front of upper caudal origin. Pelvic fins not falcate; rear margins straight or slightly concave. Anal fin long, slightly larger than 2nd dorsal fin, base 4.3–5.7% TL; anal-fin origin slightly ahead of 2nd dorsal-fin origin, and rear margin deeply concave. Tooth count 30-32/29-30; vertebrae 193-206.

Body dark grey-brown or olive dorsally, white ventrally; pectoral-fin tips dusky. Attains 4 m TL.

**DISTRIBUTION** Circumglobal in mostly warm-temperate and occasionally tropical seas. WIO: South Africa (Western Cape to KwaZulu-Natal) to southern Mozambique, Persian/ Arabian Gulf, southern India to Sri Lanka and Maldives.

**REMARKS** Widespread, but little known of its biology. Litters of 20–50, born in spring or early summer; size at birth 50-61 cm TL; gestation period 10-11 months. Matures at 210-240 cm TL; females attain greater size than males. Coastalpelagic and semi-oceanic, with a wider range than other members of its family; found close inshore and in shallow water, over continental and insular shelves, to well offshore, from surface to at least 200 m deep, and probably much deeper. Enormous schools of young sharks may occur in some areas (such as off Eastern Cape of South Africa). Feeds on a variety of mostly bony fishes, but also on small sharks, skates and stingrays, as well as invertebrates (mostly squid). Common to abundant, taken in a variety of artisanal and commercial fisheries and in numerous shark fisheries with a wide variety of gears; targeted mostly for its highly valued fins. IUCN Red List conservation status Vulnerable.



Sphyrna zygaena, 71 cm TL, immature female (South Africa). Source: Bass et al. 1975

# ORDER TORPEDINIFORMES

# FAMILY TORPEDINIDAE

## Torpedo rays

Marcelo R de Carvalho

Small- to moderately large-sized (to ~180 cm TL, but mostly <100 cm TL) with greatly expanded pectoral fins fused with head and trunk to form depressed and nearly circular disc; pectoral-fin margins thick and fleshy; well-developed electric organs; and tail stout and shark-like, shorter than disc length. Anterior margin of head straight, truncated or only slightly arched; snout reduced. Eyes small and bulging or somewhat depressed; spiracles smooth or with papillae on rear and lateral margins, and with pseudobranchial folds on inside anterior wall. Five pairs of small gill slits on front half of underside of head. Nostrils subcircular and relatively large; oronasal grooves present; internasal flap with smooth posterior edge, and sometimes with small medial indentation or lobe. Mouth moderately sized, highly arched, without lip folds but sometimes with grooves at corners. Teeth monocuspidate, relatively sharp, small and numerous (adults with 25-70 tooth rows in upper and lower jaws), set in quincunx along each jaw. Skin smooth, devoid of dermal denticles, thorns, spines and tubercles. Pair of electric organs lateral to head at mid-disc, kidney-shaped and visible through skin on underside, with irregular honeycomb appearance due to stacked electric plates.



Position of electric organ. Source: SSF

Pelvic fins fleshy, broad and rounded, comprised of a single lobe. Tail slightly depressed in cross-section, with lateral skin folds, and no caudal-region electric organs. Two prominent dorsal fins, fin apices rounded to angular; 1st dorsal fin larger than 2nd, and 1st dorsal-fin base partially or totally above pelvic-fin bases. Caudal fin subtriangular, larger than each dorsal fin, with more or less equal-sized upper and lower lobes and continuous around end of vertebral column, but lobes sometimes separated by small indentation. Body typically uniformly brown, dark grey, reddish brown or blackish background dorsally, often with various pale and dark markings (ocelli, spots, mottling, vermiculation); usually paler or white ventrally, but disc margins and pelvic fins often darker.

Reproductive mode yolk-sac viviparity. Worldwide in temperate and tropical seas, on continental shelf in shallow coastal regions or open waters of pelagic zone, often to ~100 m deep; some species may also inhabit estuaries and/or deeper waters of continental slope, but none enter freshwater. Slowswimming, on or well off the bottom, and often found resting on mud or sandy substrates. Some larger pelagic species may undertake long migrations. The powerful electric organs can discharge up to 45 volts, depending on size of the individual ray, and are used to stun prey and for defence against predation. Feed on bony fishes and benthic invertebrates and can ingest large prey items, close to their own disc length, as their jaws and mouths have a wide gape and are highly protrusile. Prey items are grasped by their discs which aid in pushing them into their gaping mouths, and then swallowed whole. Caught principally as a bycatch in trawls, but also by hook and line, and by longlines; not regularly consumed as food. Harmless to people unless interfered with, and able to stun through strong shocks if handled alive after capture. The IUCN Red List conservation status of Torpedo adenensis is Endangered, but all other WIO species are Data Deficient.

Two genera and 18 species recognised, but many are in need of revision, and several undescribed species have been recently discovered (including one in the Red Sea). Furthermore, distributions among members of this species-complex may be more restricted than so far noted, with the possibility that high endemicity will be discovered in the course of revisions or new species descriptions. Large series of preserved specimens are scarce, however, and thus consistent, useful variation in specimens is difficult to find. Both genera and 6 species in WIO.

#### **KEY TO GENERA**

1a	Spiracle margins smooth, without any knobs
	or papillae
1b	Spiracle margins with a few to many knobs
	or papillae

# GENUS Tetronarce Gill 1862

Large-sized rays (to ~180 cm TL) that appear very soft, flabby and oily when captured. Distinguished from *Torpedo* by spiracles with smooth margins, devoid of papillae or similar structures. Disc round or transversely oval, and large; snout truncate or emarginate. Mouth markedly arcuate; no lip folds or circumoral grooves; teeth small, sharp, monocuspidate. Tail short and strong, with 2 prominent dorsal fins; caudal fin broad, more or less symmetrical, fan-shaped, and larger than dorsal fins. Circumglobal, in warm-temperate to temperate waters, including off distant and isolated oceanic islands and over seamounts. Primarily pelagic hunters, swimming in water column, usually at night, where they over-swim prey items that they can quickly immobilise with an electric discharge.

This genus is in need of revision, having been treated as a subgenus of *Torpedo* or else its species included in *Torpedo* without distinct supraspecific recognition. *Tetronarce* is considered a full genus here based on its consistent morphological distinctiveness. Past authors have relied on relative distances between the dorsal fins and between the 2nd dorsal fin and caudal fin as taxonomically reliable characters, but caution is necessary as these ratios are variable and not necessarily helpful. Currently, 8–10 species recognised, but many remain poorly characterised, and intermediate and unidentified specimens abound in collections; 1 species in WIO.

## Tetronarce cowleyi Ebert, Haas & Carvalho 2015

#### Cowley's torpedo ray

PLATE 52

*Torpedo hebetans*: Thompson 1914; Von Bonde & Swart 1923. *Narcobatus nobilianus*: Barnard 1925, 1947\*; Norman 1935. *Torpedo nobiliana*: Fowler 1936\*, 1941; Smith 1949\*, 1965\*; Lloris 1986\*;

Turon *et al.* 1986; Compagno *et al.* 1989\*; Compagno *et al.* 1991; SSF No. 23.2\* [1995].

*Tetronarce cowleyi* Ebert, Haas & Carvalho 2015: 237, Figs. 2–6 (South Africa, SE Atlantic); Carvalho *et al.* 2016.

Disc broad, subcircular, wider than long and thick anteriorly; snout short with rounded median protuberance. Eyes and spiracles small and close together; spiracle margins smooth, without papillae. Mouth greatly arched; teeth small and sharp, in up to 32 rows. Dorsal fins rounded to broadly oval; 1st dorsal fin taller than 2nd. Tail short, thick; caudal fin huge, paddle-shaped, with straight or rounded posterior margin. Electric organs clearly visible in ventral view. Claspers with distal integumental flap.

Disc mostly uniformly purplish black, dark brown or dark grey dorsally, but disc margins and pelvic fins sometimes darker; whitish ventrally. Attains at least 68 cm TL.



Tetronarce cowleyi, 42 cm TL (South Africa). Source: SSF

**DISTRIBUTION** Southern Africa: Namibia (Walvis Bay) in southeastern Atlantic, to South Africa (Algoa Bay, Eastern Cape) in WIO.

**REMARKS** Previously recognised in the region as *Torpedo nobiliana* (the occurrence of which needs verification in WIO). Known from 110–457 m. Feeds on (relatively large) bony fishes and possibly small sharks. Capable of delivering a powerful shock even after capture. Occasionally caught as bycatch, but typically discarded.

# GENUS Torpedo Houttuyn 1764

Small- to medium-sized rays (to ~150 cm TL) distinguished from *Tetronarce* by spiracles with margins that are rugose or with papillae of varying dimensions and arrangement (but papillae often less conspicuous in larger specimens). Disc round or transversely oval, and large; snout usually more or less straight across, sometimes with small median bulge or even truncate. Mouth markedly arcuate; no lip folds or circumoral grooves; teeth small, sharp, monocuspidate. Tail short and strong, with 2 prominent dorsal fins; caudal fin broad, more or less symmetrical, fan-shaped, and larger than dorsal fins. Mostly benthic, sluggish rays, covering themselves with sand or mud to ambush prey (mostly fishes) which are stunned and immobilised by discharges from their electric organs. Occur predominantly in insular waters of continental shelf (in contrast to *Tetronarce*, which can occur on deeper slopes); most diverse in tropical waters of eastern Atlantic Ocean.

As with *Tetronarce*, *Torpedo* has been referred to at the subgeneric level; however, it is considered here as a full genus and separate from *Tetronarce*. At least 15 species, plus several recently discovered undescribed species; at least 5 species in WIO.

#### **KEY TO SPECIES**

1a	Disc uniformly reddish brown or orange-brown dorsally, without distinctive spots, blotches, reticulations or other patterns
1b	Disc not uniformly reddish brown or orange-brown dorsally, but with specific patterns formed by spots, blotches or reticulations
2a	Dorsal background grey to brown, and with closely packed brownish black spots, but no whitish or cream-coloured patterns; dorsal fins brownish black
2b	Dorsal background reddish, greyish or brown, and with paler patterns of irregular spots, rosettes or vermiculations
3a	Dorsal surface of disc with few but large (greater than eye diameter) brown spots
3b	Dorsal surface of disc with pale patterns of irregular spots, rosettes or vermiculations, but no large brown spots
4a	Dorsal surface includes small whitish spots that do not form broad vermiculate pattern
4b	Dorsal surface of disc, pelvic fins, and tail with strong creamy to whitish and dense vermiculate pattern, and anterior and lateral disc regions with many irregular whitish spots (no larger than eve diameter)

## Torpedo adenensis Carvalho, Stehmann & Manilo 2002

#### Aden torpedo

*Torpedo adenensis* Carvalho, Stehmann & Manilo 2002: 5, Figs. 1–7 (close to shore of Yemen, Gulf of Aden); Manilo & Bogorodsky 2003; Carvalho *et al.* 2016.

Disc broad, slightly wider than long; anterior disc margin relatively straight, with small median protuberance. Eyes and spiracles close together; spiracle margins with a few (3–8) inconspicuous knob-like papillae. Teeth small, in up to 47/39 rows. Dorsal-fin apices rounded; 1st dorsal fin larger than 2nd, its base extending just posterior to level of pelvic-fin axil (not entirely positioned over pelvic-fin bases); distance between 2nd dorsal fin and caudal fin greater than distance between dorsal fins. Males with fleshy skin flap in clasper glans region. Disc uniformly reddish brown or orange-brown dorsally, without any distinctive spots, blotches or reticulations; whitish ventrally. Attains at least 41 cm TL.



Torpedo adenensis, 41 cm TL, male holotype (Yemen). Drawn from photograph

#### DISTRIBUTION WIO: Gulf of Aden.

**REMARKS** Known only from a few specimens collected from three distinct but adjacent localities in eastern Gulf of Aden, close to Yemen; recorded from 26–140 m. Presumably a bycatch of shrimp trawl fisheries. IUCN Red List conservation status Endangered, due to its restricted known range.

# Torpedo fuscomaculata Peters 1855

## Blackspotted torpedo

PLATE 53

*Torpedo fuscomaculata* Peters 1855: 466 (Ibo, Mozambique); Playfair & Günther 1866; Fraser-Brunner 1949; Smith 1961\*; Smith & Smith 1963; Wallace 1967\*; Compagno *et al.* 1989\*; Debelius 1993\*; SSF No. 23.1\* [1995]; Sommer *et al.* 1996; Carvalho *et al.* 2002\*; Manilo & Bogorodsky 2003; Carvalho *et al.* 2016.

*Torpedo smithii* Günther 1870: 451 [probably South Africa]. *Torpedo* sp.: Fourmanoir 1963.

Disc slightly wider than long, widest near midlength; anterior margin truncated, sometimes with median bulge. Teeth small, in up to 50 rows. Dorsal fins rounded at apices; 1st dorsal fin larger than 2nd. Tail short, stout; lateral skin fold on tail usually ridge-like, extending from about midbase of 2nd dorsal fin to lower peduncle. Caudal fin broad, with slightly sloping upper lobe. Males without skin flap in clasper glans region.

Disc grey, yellowish to reddish brown background dorsally, with close-set brownish black spots; dorsal fins brownish black with white edges. (Colour variation in this species needs more study, as the number, size and arrangement of darker spots on the dorsal surface vary, such as no spots in specimens from South Africa, but smaller and more regularly arranged spots in specimens from Mauritius.) Attains 65 cm TL.



Torpedo fuscomaculata, 27 cm TL (South Africa). Source: Whitfield 1998

**DISTRIBUTION** WIO: South Africa (Eastern Cape) to southern Mozambique, Tanzania (Zanzibar), Madagascar, Seychelles and Mauritius.

**REMARKS** Specimens ~25 cm TL almost sexually mature. Recorded from intertidal zone to ~440 m deep, and also from estuaries in South Africa. Feeds on fishes and cuttlefish.

# Torpedo panthera Olfers 1831

Leopard torpedo

PLATE 53

*Torpedo panthera* Olfers 1831: 15–16 (Red Sea); Blegvad & Løppenthin 1944; Qureshi 1972\*; Dor 1984; Compagno *et al.* 1989\*; Baranes & Golani 1993; Debelius 1993\*; Randall 1995\*; Sommer *et al.* 1996\*; Carpenter *et al.* 1997\*; Hennemann 2001\*; Carvalho *et al.* 2002\*; Manilo & Bogorodsky 2003; Carvalho *et al.* 2016.

Disc broadly circular, slightly wider than long, and widest just before mid-disc; anterior margin usually straight. Eyes and spiracles similarly sized, but separated by wide gap; spiracles with ~7 short papillae. First dorsal fin larger than 2nd and rounder at apex; interdorsal distance roughly equal to distance between 2nd dorsal fin and caudal fin. Tail short and moderately stout. Caudal-fin upper lobe larger and more sloping than lower lobe. Males with median skin flap in clasper glans region.

Disc brown, reddish brown or tan background dorsally; clusters of more or less isolated and sometimes blurry whitish spots over disc, pelvic fins, tail and even dorsal fins, with spots varying in density, size and arrangement. Attains ~60 cm TL.



Torpedo panthera, 35 cm TL (Red Sea).

**DISTRIBUTION** Indian Ocean. WIO: Red Sea, Gulf of Oman and shallow areas of Arabian Sea, to Bay of Bengal; reports from Persian/Arabian Gulf and south of Somalia probably erroneous.

**REMARKS** Biology virtually unknown other than males mature by ~28 cm TL, and feeds on invertebrates and fishes. Occurs on continental shelf, over muddy and sandy bottoms, usually to ~110 m deep.

# Torpedo sinuspersici Olfers 1831

## Marbled torpedo

PLATE 53

*Torpedo sinuspersici* Olfers 1831: 15, 17 [presumably Persian/Arabian Gulf]; Fowler 1941, 1956; Wallace 1967\*; Bianchi 1985; SSF No. 23.3\*; Compagno *et al.* 1989\*; Van der Elst 1993\*; Debelius 1993\*, 1999\*; Randall 1995\*; Sommer *et al.* 1996\*; Carpenter *et al.* 1997\*; Hennemann 2001\*; Carvalho *et al.* 2002\*; Manilo & Bogorodsky 2003; Heemstra & Heemstra 2004\*; Carvalho *et al.* 2016. *Torpedo polleni:* Bleeker 1865.

Disc broad, slightly wider than long, fleshy at margins. Eyes and spiracles subequal, separated by clear gap. Spiracles rounded, their margins usually with 9 or 10 short papillae. First dorsal fin with rounded apex, larger than 2nd; distance between 2nd dorsal fin and caudal fin usually greater than interdorsal distance. Caudal-fin upper lobe larger and more sloping than lower lobe; fin margin straight or slightly rounded. Distinguished by dorsal surface with strong vermiculate colour pattern. Disc russet to blackish background dorsally, and brightly patterned with strong creamy to whitish and rather thick vermiculations or rosettes over entire disc, pelvic fins and tail, and decreasing in size towards disc margins, plus many creamcoloured and irregular spots (no larger than eye diameter) on anterior and lateral regions of disc; whitish ventrally. Attains 130 cm TL (commonly ~80 cm TL).



Torpedo sinuspersici, 26 cm DW (Mozambique). O Alvheim © IMR

**DISTRIBUTION** Indian Ocean. WIO: South Africa (Eastern Cape), Mozambique to Red Sea, Persian/Arabian Gulf, Gulf of Oman to India and Sri Lanka.

**REMARKS** Litters of 9–22; matures at ~40 cm TL. Found over sandy bottom and near reefs, to ~300 m deep. Feeds on invertebrates and fishes. Seemingly widespread in WIO (often seen by divers), but probably a species-complex. Has been misidentified as *T. fuscomaculata* from Madagascar and Seychelles, and as *T. marmorata* from Lakshadweep; Smith & Smith's (1963) *T. fuscomaculata* from Seychelles may have also included *T. sinuspersici* (see Carvalho *et al.* 2002). Jones & Kumaran (1980) illustrated a specimen from Lakshadweep labelled *T. marmorata* but which resembles *T. sinuspersici*; other misidentifications of this species as *T. marmorata* occur in the literature.

## Torpedo suessii Steindachner 1898

#### Red Sea torpedo

PLATE 54

Torpedo suessii Steindachner 1898: 784, Pl. 2 (Yemen, Red Sea); Carvalho et al. 2002\* [as suessi]; Carvalho et al. 2016. ?Torpedo sp. 1: Michael 1993\*. Small-sized with ornate colour pattern. Disc broadly circular, slightly wider than long, and widest near mid-disc; snout short, anterior margin usually straight. Spiracle margins with short papillae. First dorsal fin larger and more broadly rounded at apex than 2nd. Caudal-fin upper lobe larger and more slanted than lower lobe. Claspers slender, elongate, and without skin flap in clasper glans region (versus skin flap present in *T. adenensis* and *T. panthera*).

Dorsal background bright brown, with large dark brown spots with pale outlines (ocelli), slender reticulations over disc and disc edges, and smaller dark spots on front of disc, pelvic fins and dorsal-fin bases. Attains at least 35–40 cm TL.



Torpedo suessii, ~30 cm TL (Yemen). Composite

#### DISTRIBUTION WIO: southern Red Sea.

**REMARKS** Biology virtually unknown; inhabits shallow water and probably feeds on fishes and benthic invertebrates. Almost all authors have placed this species as a junior synonym of *T. sinuspersici*, but Carvalho *et al.* (2002) presents evidence for its recognition. Its colouration easily separates it from congeners in the Red Sea and adjacent gulfs.

# FAMILY NARCINIDAE

# Numbfishes

Marcelo R de Carvalho

Small-sized rays (to ~86 cm TL, commonly <50 cm TL) with oval, rounded, heart-shaped or distinctively shovel-shaped disc; stout shark-like tail equal to or greater than disc length or width. Electric organs large, bean-shaped, paired at pectoralfin bases, and visible through skin on underside. Trunk and head depressed; head relatively broad. Snout moderately elongated, broadly rounded to angular. Eyes small or minute

(vestigial in *Benthobatis*), and larger or smaller than spiracles; spiracles circular to oval, margins with or without papillae, and with or without elevated rims. Five pairs of small gill slits on underside near front half of pectoral-fin bases. Nostrils circular, small; oronasal grooves broad; internasal flap overlaps mouth slightly. Mouth relatively small and transverse, surrounded by prominent groove and with strong lip folds. Teeth small, rounded-oval, with short cusps, and in 12-45 rows in each jaw. Skin smooth, without dermal denticles or thorns. Pectoralfin insertions opposite pelvic-fin origins. Pelvic fins low, subangular or rounded, not divided into anterior and posterior lobes. Tail broad but abruptly narrower than trunk, moderately depressed and long, and usually with lateral skin folds. Two dorsal fins, moderately large and clearly separated, usually subequal (1st dorsal fin slightly larger than 2nd in some species of Narcine), and angular or rounded-angular; 1st dorsal-fin origin just behind mid-total length and pelvic-fin insertions. Caudal fin usually large and broadly rounded, but upper lobe slightly more angular. Dorsal surface varies from whitish, yellowish, brownish, grey-brown, greenish, reddish or black (blackish in deepwater species), either without spots or with dark spots, blotches, bars or lines, or with white spots and lines sometimes forming complex ocelli on disc and pectoral fins.

Reproductive mode yolk-sac viviparity, as far as known. Feed mostly on benthic invertebrates; their jaws and mouths are highly protrusile, forming a tubular mechanism that enables them to extract prey organisms from the substrate, especially polychaetes. Slow-swimming bottom-dwellers of upper continental slope, over sandy, muddy or occasionally rocky substrates, including off sandy beaches, in muddy enclosed bays and on coral reefs, from intertidal zone to deeper than 1 000 m; apparently unable to penetrate freshwater to any extent. Caught as minor bycatch of small local inshore fisheries and offshore trawl fisheries. Numbfishes can deliver a moderate shock when disturbed or captured, but are otherwise harmless to people. The IUCN Red List conservation status of most species is Data Deficient or Least Concern; one species from the Caribbean Sea is listed as Critically Endangered.

Almost circumglobal in tropical to warm-temperate seas, but conspicuously absent from Mediterranean Sea, eastern Atlantic, eastern South Pacific and Pacific Plate region, and so far unrecorded from the Red Sea and Persian/Arabian Gulf in WIO, despite good collections from those waters. Four genera and ~36 species recognised, but many species are in need of revision and several species are undescribed; 2 genera and 7 species in WIO (the one genus and species, *Benthobatis moresbyi* [Plate 54], known only from 524–1 071 m in the Arabian Sea and off SW India).

#### **KEY TO GENERA**

## GENUS Narcine Henle 1834

Disc oval, somewhat rounded to shovel-shaped; greatest width at ~<sup>3</sup>/<sub>3</sub> disc length. Snout rounded to broadly angular; preorbital snout ~<sup>1</sup>/<sub>4</sub> disc length. Eyes larger than spiracles (and bulging in fresh specimens); spiracles circular to ovoid, usually with thick elevated rims; some species (not in WIO) with warty papillae on entire spiracular margin. Nostrils small and circular; internasal flap wider than long, reaching upper tooth band. Mouth protrusile. Electric organs bean-shaped, extending on underside from opposite eyes to rear fifth of disc. Tail stout at base, its length greater than disc width or length; lateral skin folds usually prominent. Dorsal fins similarly shaped, usually subequal (either fin may be slightly taller than other). Caudal fin usually tall, fan-shaped, with broadly rounded ventral margin and slightly angular upper lobe.

Distinguished from other narcinid genera by internasal flaps with relatively straight posterior edge (except trilobed in *Discopyge*); nostrils not subdivided into two distinct compartments by bridge of stiff integument; both upper and lower tooth bands exposed when mouth closed; eyes functional and evident anterior to spiracles, usually about same size as spiracles; pelvic fins posteriorly separated and not joined to form continuous pelvic curtain; claspers laterally joined to pelvic fins, and not dorsally covered and concealed by pelvic fins; and lateral skin folds on tail generally welldeveloped, extending from level of 1st dorsal fin to peduncle (much reduced, ridge-like folds in *Benthobatis*).

Common inshore component of Indian Ocean fauna (less so in southwestern Indian Ocean, but very common in eastern Arabian Sea). Relatively common off Pakistan, but apparently these species have not dispersed into the Persian/Arabian Gulf and are also absent from the Red Sea. Probably fairly stationary with limited individual ranges. A revision of this genus is needed, especially of species occurring in the northern Indian Ocean. Previous accounts of batoids in the Arabian Sea have usually misidentified numbfish species. With the exception of *N. lingula* and *N. timlei*, all WIO species are known from only a few specimens. Furthermore, many doubts linger concerning the number of valid species of *Narcine* from this region. For example, *N. maculata* is known from the Bay of Bengal and eastwards; it may also be present in the Arabian Sea but has not yet been found west of Sri Lanka, and thus is not included here. About 20 species; at least 6 species (possibly 2 others undescribed) in WIO.

#### **KEY TO SPECIES**

1a	Internasal flap longer than wide					
1b	Internasal flap wider than long					
	internasal flap					
	······································					
Za	reticulations or other markings					
2b	Dorsal colouration composed of reticulations, spots or other					
	mottled patterns, not uniformly pale or purplish brown 3					
3a	Preorbital snout length about equal to preoral snout					
	length; 2nd dorsal fin about equal to or taller than					
	1st dorsal fin					
3b	Preoral snout length greater than preorbital snout length;					
	Devel colouration compand of our evolution and developed					
4a	Dorsal colouration composed of numerous small dark spots					
4b	Dorsal colouration devoid of small dark spots					
	· ·					
5a	Mean disc width and length <45% TI : branchial basket					
	length (distance between 1st and 5th gill slits) clearly					
	greater than transverse distance between last pair of					
	gill slits N. oculifera					
5b	Mean disc width and length ~48% IL; branchial basket					
	pair of gill slits					
	r J					

## Narcine atzi Carvalho & Randall 2003

#### Atz's numbfish

*Narcine atzi* Carvalho & Randall 2003: 60, Figs. 1–2 (off Khalil, Oman, Gulf of Oman); Carvalho & Last 2016.

Disc subcircular to broadly oval, widest just posterior to its midlength; preorbital snout length shorter than preoral snout length. Spiracles wider than long, with elevated, smooth rims without papillae. Upper and lower tooth bands broadly circular and subequal in width (upper tooth band wider than lower band), visible with closed mouth; teeth in up to 20 rows. Dorsal fins large, with rounded apices, and 1st dorsal-fin height and base greater than that of 2nd dorsal fin; caudal fin tall, rounded at apex and posterior margin; lateral skin folds wide.

Disc tan dorsally, with numerous small dark brown spots (equal to or smaller than eye diameter) over disc and tail and within spiracular walls, coupled with fewer faint, large blotches on disc; creamy white ventrally, with grey margins on disc and pelvic fins, and blotches on tail. Attains at least 36 cm TL.



Narcine atzi, ~30 cm TL, female (Oman). Composite

**DISTRIBUTION** Northern Indian Ocean: Gulf of Oman in WIO, to Bay of Bengal and Andaman Sea.

**REMARKS** Poorly known from only a few specimens, collected in shallow water (~30 m). Size at birth ~11 cm TL; females mature by ~31 cm TL. Incidental bycatch in trawls.

## Narcine insolita Carvalho, Séret & Compagno 2002

#### Madagascar numbfish

Narcine insolita Carvalho, Séret & Compagno 2002: 140, Figs. 1–5 (NW Madagascar); Carvalho & Randall 2003\*; Carvalho & Last 2016.

Disc oval or heart-shaped, its greatest width just posterior to its midlength; disc fleshy, with thick margins. Eyes large and bulging; spiracles large, conspicuously rounded, with elevated rims all around; interspiracular distance much smaller than interorbital distance. Teeth in up to 24 rows; upper and lower tooth bands equal in width and broadly circular in outline. Internarial flap short and wide. First dorsal fin very large, conspicuously taller and with longer base than 2nd dorsal fin; caudal fin with well-developed upper and lower lobes, and rounded-oval apex; lateral skin folds wide.

Disc yellowish brown dorsally, with irregular darker brown to reddish brown blotches on disc margins, posterior disc and front margin of snout; darker brown blotches on front part of dorsal fins and caudal-fin apex, and laterally on tail beneath dorsal fins; creamy white ventrally. Attains at least 27 cm TL.



DISTRIBUTION WIO: Madagascar (west coast).

**REMARKS** Known only from seven specimens collected from two localities off northwestern and southwestern Madagascar, at 150–175 m. Another specimen from southern Mozambique, collected at ~69 m, is very close in morphology to *N. insolita* but with dorsal fins of subequal proportions, and may represent an undescribed species. Occurs predominantly over muddy substrates. An unutilised bycatch of shrimp trawl fisheries.

# Narcine lingula Richardson 1846

#### Chinese numbfish

PLATES 54 & 55

Narcine lingula Richardson 1846: 196 (China Seas, off Canton; Indian Ocean, off Chennai, India); Carvalho & Randall 2003\*;

Carvalho & Last 2016.

Narcine timlei: Qureshi 1972\*; Carvalho et al. 1999\*.

Disc oval-rounded, widest near its midlength. Preorbital length about equal to preoral length, and much greater than prenarial length. Eyes and spiracles moderately sized, subequal; spiracles with slender, elevated rims. Internarial flap short and wide. Tooth bands rounded, broad, subequal in width; up to 22 rows of teeth. Dorsal fins subequal, rounded-oval at apex; caudal fin rounded, slightly larger than dorsal fins; lateral skin folds slender.

Disc brown dorsally, with dark brown spots (usually larger than eye diameter); whitish ventrally. Attains probably 33 cm TL.



Narcine lingula (W India).

**DISTRIBUTION** Indo-Pacific. WIO: Arabian Sea (Pakistan and India); occurrence in Persian/Arabian Gulf needs confirmation; elsewhere to east coast of India, Indonesia (Java) and East China Sea.

**REMARKS** Very similar to *N. maculata* from western Pacific Ocean, thus the separation of these species is provisional. Matures at ~25 cm TL. Found inshore and offshore, in shallow tropical continental waters, but depth range unknown. A bycatch of demersal trawl fisheries.

# Narcine oculifera Carvalho, Compagno & Mee 2002

Big-eye numbfish

PLATE 55

Narcine oculifera Carvalho, Compagno & Mee 2002: 137, Figs. 1–4 (Muscat, Oman, Gulf of Oman); Carvalho & Randall 2003\*; Manilo & Bogorodsky 2003; Carvalho & Last 2016.

Narcine indica: Norman 1939; Baranes & Randall 1989. Narcine sp.: Debelius 1993\*; Randall 1995\*.

Disc oval to heart-shaped, widest posterior to its midlength, with thick margins. Eyes large, bulging; spiracles large and circular with elevated rims; interorbital distance greater than interspiracular distance. Internarial flap short and wide. Teeth with small lateral cusplets; upper and lower tooth bands roughly equal in width, broadly rounded in profile. Dorsal fins tall, with rounded to oval apices, and 1st dorsal-fin height slightly greater than 2nd; caudal fin tall, with oval apex; lateral skin folds wide.

Striking dorsal colouration with reticulated pale brown to reddish brown patterning over disc, pelvic fins and tail, demarcating white or creamy oval, reniform and circular spots of variable sizes over disc, and with small white spots on dorsal fins and caudal fin; white to yellowish white ventrally. Attains at least 35 cm TL.



Narcine oculifera, 24 cm TL (Oman).

**DISTRIBUTION** WIO: Gulf of Oman (off Oman) and Gulf of Aden (off Somalia).

**REMARKS** Distinctive and not easily confused with other numbfishes. Males mature by ~24 cm TL, and females by ~32 cm TL. Known from few specimens collected mostly from shallow water (<25 m), but recorded to deeper than 100 m. Observed resting on rocky surfaces. A rare bycatch of trawls.

#### Narcine rierai (Lloris & Rucabado 1991)

Mozambique numbfish

PLATE 55

Heteronarce rierai Lloris & Rucabado 1991: 327, Figs. 1–3 (Mozambique); SSF No. 24a.1\* [1995].

Narcine rierai: SSF No. 24a.1\* [1995]; Carvalho & Randall 2003\*;

Compagno & Heemstra 2007; Carvalho & Last 2016.

Disc markedly shovel-shaped; tail elongated, much longer than disc length. Eyes larger than spiracles; spiracles elliptical, without elevated rims or papillae. Internasal flap longer than wide, and with pores; nostrils slit-like. Mouth small. Tooth bands slender and subtriangular in outline; up to 16 rows of teeth. Dorsal fins wide-set, subequal; caudal fin low and elongate; lateral skin folds low ridges.

Disc uniformly medium or pale brown to golden-brown or reddish brown dorsally; uniformly whitish ventrally. Attains at least 30 cm TL.



Narcine rierai, 11 cm DW, 31 cm TL (Mozambique).

**DISTRIBUTION** WIO: Somalia to Mozambique and possibly South Africa.

**REMARKS** Size at birth >7 cm TL; matures at 23–27 cm TL. Occurs over lower continental shelf, with most specimens collected from 169–255 m, but one specimen from Tanzania collected from 300–500 m.

## Narcine timlei (Bloch & Schneider 1801)

Brown numbfish

PLATE 55

*Raja timlei* Bloch & Schneider 1801: 359 (Tharangambadi, India, Bay of Bengal).

Narcine indica Henle 1834: 315, Pl. 2 (Tharangambadi coast, India). Narcine microphthalma Duméril (ex Valenciennes) 1852: 275

(Malabar coast, India).

Narcine macrura Valenciennes in Duméril 1852: 277 ('Sea of the Indies'). Narcine brunnea Annandale 1909: 45, Pl. 3a (Bay of Bengal); Garman 1913; Qureshi 1972\*; Carvalho *et al.* 1999\*; Manilo & Bogorodsky 2003. Narcine timlei: Carvalho & Randall 2003\*; Carvalho & Last 2016.

Disc oval to subtrapezoidal (usually oval in preserved specimens); snout broadly rounded. Eyes very small, smaller than spiracles; spiracles rounded, with smooth low rims. Internarial flap much wider than long. Tooth bands wide, rounded; teeth in up to 25 rows. Pelvic fins wide; dorsal fins small, rounded, subequal; caudal-fin margin usually rounded; lateral skin folds low.

Disc uniformly purplish brown, pale or dark brown or tan dorsally, and devoid of any specific markings; creamy white ventrally. Attains at least 38 cm TL.



Narcine timlei, male (India).

**DISTRIBUTION** Indo-Pacific. WIO: Pakistan, India and Sri Lanka; elsewhere to Bay of Bengal, Indonesia and South China Sea.

**REMARKS** Males mature >24 cm TL; free-swimming by ~5.5 cm TL. Inshore and offshore in tropical continental waters. Probably frequently captured by trawls.

# FAMILY NARKIDAE

### Sleeper rays

Marcelo R de Carvalho

Small-sized (~8–52 cm TL; most attain <35 cm TL) with disc proportionally large, circular to oval, and with thick margins in some species. Head depressed, broad, not raised above disc; snout broadly rounded, short to moderately elongate. Pectoralfin origins lateral to snout, ending just behind pelvic-fin origins. Pair of large bean-shaped electric organs visible through skin on underside at pectoral-fin bases. Eyes generally small (vestigial or externally absent in *Typhlonarke*), relatively close together and contiguous to spiracles; spiracles subcircular, margins with papillae (*Electrolux*) or without papillae (*Heteronarce, Narke*), usually with low rims. Five pairs of very small gill slits on ventral front half of disc; no gill rakers on internal gill slits. Nostrils slender, elongated or circular, slightly oblique, just in front of mouth and connected to it through broad oronasal grooves; internasal flap overlapping mouth, usually studded with pores, and posterior edge with or without small knob (trilobed). Mouth narrow and transverse, nearly straight across, with strong lip folds and weak peripheral grooves or pleats. Teeth small, similar in shape, oval, and with low transverse ridge or single cusps on crowns, not laterally expanded and plate-like; maximum ~25 tooth rows in each jaw. Skin smooth, completely devoid of denticles, spines, thorns or tubercles. Pelvic fins elongate, low, broadly rounded or subangular, and not subdivided into distinct lobes (except anterior lobes separate and leg-like in Typhlonarke). Tail abruptly narrower than trunk, stout and shark-like, slightly depressed, and with or without lateral skin folds. Tail either with 1 dorsal fin (Narke, Typhlonarke), 2 usually subequal-sized dorsal fins (Electrolux, Heteronarce), or no dorsal fin (Temera); dorsal fin(s) rounded to angular, with confluent apex and margin; 1st dorsal-fin origin behind pelvicfin insertions or above rear part of pelvic fins. Caudal fin broadly rounded to oval, nearly symmetrical, and larger than dorsal fins, but not shark-like; lower lobe small. Dorsal colour usually brownish, greyish or reddish brown, either plain or with few markings and usually without complex patterns (except ornate in *Electrolux*); white, greyish or brownish ventrally, sometimes with darker outer disc margins or pelvic fins. Many narkids have their proportions distorted by post-mortem bloating and desiccation, usually compounded after fixation; typically, the disc, pelvic fins and eyes shrink significantly, and many other proportions such as dorsal-fin shape are also affected.

Reproductive mode yolk-sac viviparity. Shallow inshore to deepwater offshore rays of outer continental shelf and upper slope, usually found over soft bottom, to ~330 m deep; not known from freshwater. Slow-swimming bottomdwellers which feed on small benthic invertebrates (especially polychaetes); their small mouths restrict their diet, and whether sleeper rays prey on fishes is unknown. Caught as a minor bycatch of shallow trawl fisheries and some inshore artisanal net fisheries, but typically not used. All species can deliver a moderately strong shock when disturbed or captured. IUCN Red List conservation status of most species is Data Deficient, but several are considered Vulnerable.

Occur in temperate to tropical waters of Indo-Pacific (surprisingly absent from around Australia). Six genera and ~12 species; 3 genera and 6 species in WIO.

#### **KEY TO GENERA**



Continued ...

#### **KEY TO GENERA**



# GENUS **Electrolux** Compagno & Heemstra 2007

Disc subcircular (about as long as wide), disc length roughly half TL, and anterior margin broadly rounded; head small; snout very short and nearly straight. Eyes well-developed, close to front edge of disc, but mostly hidden by loose skin; eyes and spiracles contiguous; spiracle length slightly longer than eye diameter; spiracle margins with relatively large stiff papillae. Mouth and nostrils project ventrally from disc in lateral view; lower lip folds and grooves short, ending far lateral to mouth. Pelvic fins not divided into anterior and posterior lobes; claspers in adult males short and flat, not extending beyond pelvic-fin rear tips. Tail stout, with broad lateral skin folds. Two dorsal fins; 1st dorsal-fin origin over pelvic-fin rear tips, and only slightly larger than 2nd dorsal fin. One species.

### Electrolux addisoni Compagno & Heemstra 2007

Ornate sleeper ray

PLATE 55

*Electrolux addisoni* Compagno & Heemstra 2007: 22, Figs.1–9, 11–19 (off Margate, KwaZulu-Natal, South Africa); Carvalho 2016.

Diagnosis as for genus; distinguished from other narkid species by its spiracles, oronasal region, and colouration. Spiracle margins with low ridge-like rims, 5 or 6 long, slender, stiff papillae, and 2 or 3 short soft papillae, including 1 minute papilla on eye; incurrent apertures of nostrils circular with broad trumpet-like flaps; internasal flap deeply incised, with prominent lateral lobes and very small medial lobe (trilobed). Tooth count 15 or 16/17 or 18.

Unique elaborate colour pattern of dark brown or greenish brown background, and small yellowish spots over dorsal surface of disc and fins (spots largest on middle region of disc; spots white in preserved specimens), and with several curved, somewhat concentric black stripes on disc (less evident in preserved specimens); ventral surface abruptly white in centre of disc and at pelvic-fin bases, forming pear-shaped white blotch. Attains at least 52 cm TL.



*Electrolux addisoni*, 51 cm TL, male (South Africa). Source: Compagno & Heemstra 2007

**DISTRIBUTION** WIO: endemic to South Africa (Coffee Bay, Eastern Cape to Virginia Beach, KwaZulu-Natal).

**REMARKS** Apparently rare but long known and photographed by divers before it was described; the original description is based on specimens collected from a short strip of South African coastline, from <50 m deep. Inhabits sandy and gravelly patches of shallow-water reefs. Relatively largesized; only adult male specimens collected to date; males mature by 50 cm TL. Diurnal forager; apparently feeds on small crustaceans and polychaetes. The striking colour pattern may represent an aposematic adaptation as these rays seem to enact threat displays. This species is protected in South Africa.

## GENUS Heteronarce Regan 1921

Disc subcircular (slightly longer than wide), anterior margin oval to broadly rounded, head somewhat small, and snout relatively short (longest in *H. garmani*). Eyes and spiracles contiguous; spiracles circular, and margins with low ridgelike rim without papillae. Incurrent apertures of nostrils slitlike, elongated; internasal flap usually without pores (pores in *H. bentuviai*), and with median groove but posterior edges nearly straight. Lower lip folds and grooves relatively short. Tooth count 10–16/10–16. Claspers in adult males extend slightly beyond pelvic-fin rear tips (less so in *H. bentuviai*). Two dorsal fins, usually subequal (1st dorsal fin much larger than 2nd in *H. bentuviai*); 1st dorsal-fin origin over pelvic-fin rear tips; 2nd dorsal-fin apex usually more rounded than that of 1st. Lateral skin folds on tail only moderately broad. Colour usually greyish, reddish or brownish dorsally, sometimes with darker undefined markings or more defined spots, and usually creamy ventrally.

Poorly known genus, but easily distinguished from other narkid genera by the combination of two dorsal fins and plain dorsal colouration. Specimens of *Heteronarce* are rare in collections, which precludes a better understanding of species limits. A review by Talwar (1981) recognised *H. mollis*, *H. garmani* and *H. prabhui* (*H. bentuviai* was described later), but was based on few specimens from a small area (off Kerala, India). Even the separation of *H. garmani* and *H. mollis* (including *H. prabhui*) is in need of revision. Four species recognised in the literature, but the validity of *H. prabhui* is not accepted here; 3 species in WIO.

#### **KEY TO SPECIES**

Dorsal colouration with dark spots anterodorsal to pectoral-fin
insertions and on 1st dorsal fin and caudal-fin upper lobe; 1st
dorsal fin triangular H. bentuviai
Dorsal colouration generally uniformly grey, brownish red or pale brown, without conspicuous dark spots on pectoral-fin insertions, 1st dorsal fin or caudal-fin upper lobe; 1st dorsal fin
broadly rounded
Eyes and spiracles similar in size, with spiracles relatively small (interspiracular distance about 3 times spiracle length or width): body mostly creamy white ventrally <b><i>H. aarmani</i></b>

## Heteronarce bentuviai (Baranes & Randall 1989)

Eilat electric ray

PLATE 56

Narcine bentuviai Baranes & Randall 1989: 87, Pl. 1, Figs. 1–9 (Gulf of Aqaba, Red Sea); Carvalho 2016.

Heteronarce bentuviai: Hennemann 2001\*; Compagno & Heemstra 2007.

Disc broadly rounded anteriorly, disc length ~½ TL; spiracles larger than eyes. Internasal flap with numerous pores, and more or less straight posterior margin. Teeth in up to 11 rows, not visible externally. Pelvic fins posteriorly rounded, broad; 1st dorsal-fin origin over pelvic-fin midlength, much larger than 2nd dorsal fin and apex more angular; caudal fin larger than dorsal fins; tail short and broad. Males with relatively short claspers (compared to *H. garmani*).

Conspicuous, bold colour pattern, including clearly delimited dark brown to blackish spots or blotches just anterodorsal to pectoral-fin insertions and covering 1st dorsal fin and caudal-fin upper lobe, and with poorly defined darker streaks dorsally over region of electric organs. Attains at least 19 cm TL.



Heteronarce bentuviai, 15 cm TL (Red Sea).



Heteronarce bentuviai, 18 cm TL, male paratype (Red Sea).

# **DISTRIBUTION** WIO: northern Red Sea, including Gulf of Aqaba.

**REMARKS** Males mature by ~15 cm TL. Small-sized, conspicuous-looking narkid known only from Red Sea, at 80–200 m (specimens from off Socotra in Gulf of Aden were morphologically similar to *H. bentuviai* but differed in details of dorsal colouration and dorsal-fin proportions). Found on sandy and muddy bottoms; probably feeds on soft bottom-dwelling invertebrates, such as polychaetes and crustaceans, as do other narkids. A discarded bycatch of trawl and gillnet fisheries.

## Heteronarce garmani Regan 1921

Natal electric ray

Heteronarce garmani Regan 1921: 414 (off Umvoti River, KwaZulu-Natal, South Africa); Gilchrist 1922; Barnard 1925, 1927; Fowler 1941; SFSA No. 90\*; Wallace 1967\*; Hulley 1972\*; ?Talwar 1981; SSF No. 24.1\*; Compagno *et al.* 1989\*; Lloris & Rucabado 1991; Compagno & Heemstra 2007\*; Ebert 2014\*; Carvalho 2016.

Heteronarce regani Von Bonde & Swart 1923: 14, Pl. 22, Fig. 2 (KwaZulu-Natal, South Africa).

Narcine natalensis Fowler 1925: 198, Fig. 2 (KwaZulu-Natal, South Africa).

Disc subcircular, shovel-shaped, slightly longer than wide; anterior margin broadly oval or subcircular; preorbital snout length slightly greater than interspiracular distance; snout slightly longer than in congeners. Eyes and spiracles contiguous, relatively small, and subequal in diameter; spiracles with slender elevated rims. Posterior margin of internasal flap broadly rounded to straight, slightly fringed. Teeth in up to 12 rows, barely visible externally. Pelvic fins and tail slender; 2nd dorsal fin slightly larger and more rounded than 1st; 1st dorsal-fin origin over pelvic-fin free rear tips; caudal fin larger than dorsal fins.

Disc pale to dark brown dorsally, sometimes with small indistinct darker markings; creamy white ventrally, but disc margin sometimes brown posteriorly. Attains at least 27 cm TL.



Heteronarce garmani, 20 cm TL (South Africa). Source: SFSA

**DISTRIBUTION** WIO: South Africa and Mozambique.

**REMARKS** Males mature by ~17 cm TL. Occurs on outer shelf and upper slope, at 154–329 m. Specimens are rare; records from the Arabian Sea and off Kerala, India (Talwar 1981), probably refer to *H. mollis*; specimens identified as *H. garmani* from Madagascar are *Narcine insolita*  (Narcinidae). Wallace (1967) considered *H. garmani* distinct from *H. mollis* by its longer snout and smaller eyes and spiracles, and proportions of the mouth, nostrils and internasal flap appear to be greater in *H. mollis*, yet its separation from *H. mollis* requires further clarification. IUCN Red List conservation status Near Threatened.

## Heteronarce mollis (Lloyd 1907)

#### Soft electric ray

PLATE 56

Narcine mollis Lloyd 1907: 8 (Gulf of Aden); Annandale 1909\*; Lloyd 1909\*; Garman 1913.

?Heteronarce prabhui Talwar 1981: 149, Fig. 1 (off Kollam, India). Heteronarce mollis: Norman 1939; Fowler 1941; Talwar 1981; Lloris & Rucabado 1991; Compagno & Heemstra 2007\*; Ebert 2014\*; Carvalho 2016.

Disc subcircular, broad, slightly wider than long; snout broadly rounded, relatively short, preorbital snout length less than twice interspiracular distance (in preserved specimens). Spiracle diameter clearly greater than eye diameter. Internasal flap about equal in length and width, with a small notch at mid-width, and sometimes with pores. Teeth small, in up to 16 rows, not visible externally. Pelvic fins broad; dorsal fins broadly rounded, subequal or else 2nd dorsal fin slightly larger than 1st; 1st dorsal-fin origin over pelvic-fin free rear tips; lateral skin folds moderately broad; caudal fin larger than dorsal fins, with broadly rounded posterior margin.

Body dark brown to greyish dorsally, sometimes with small irregular paler and darker markings; pale brown, tan or greyish ventrally. Attains at least 25 cm TL.



Heteronarce mollis (Yemen). Source: Annandale 1909

**DISTRIBUTION** WIO: Somalia, Socotra, Gulf of Aden and India.

**REMARKS** Known from very few specimens collected mostly in Gulf of Aden from 73-346 m. Males mature at 17-20 cm TL. Recognition of H. prabhui (Quilon electric ray) as a synonym of H. mollis needs further scrutiny as the former has been accepted as valid but without supporting evidence (cf. Compagno 1999, 2005; Compagno & Heemstra 2007), and Talwar's (1981) original description of H. prabhui (collected from ~300 m) is remarkably similar to Lloyd's (1909) original illustration of H. mollis and to material examined from Somalia (moreover, the characters used to separate the species are features that typically vary with preservation: slight differences in disc width and length, and the relative proportions of spiracles and eyes). Lloyd (1909) provided the first illustration of this species (in entire dorsal aspect and ventral anterior disc), and Annandale (1909) illustrated the mouth and teeth, but no new data have been published since Norman's (1939) brief record (also from Gulf of Aden). Thus, a conservative approach is adopted here, and H. prabhui is therefore regarded as a junior synonym of H. mollis. And, with the synonymy of H. prabhui with H. mollis, its range can be extended to the eastern Arabian Sea.

## GENUS Narke Kaup 1826

Disc circular to subcircular (slightly longer than wide); anterior margin broadly rounded to slightly convex; head region small, and eyes close to front edge of disc. Eyes and spiracles closely adjacent; spiracles circular, slightly greater than eye diameter, and margins with low ridge-like rims (least prominent in N. dipterygia) and without papillae. Incurrent apertures of nostrils slit-like, elongated; internasal flap usually without pores, and with median groove, and posterior edge nearly straight. Lower lip folds and grooves relatively short. One dorsal fin, its origin over pelvic-fin rear tips. Tail with moderately broad lateral skin folds. Claspers in adult males extend beyond pelvic-fin rear tips. Body usually greyish, reddish or brownish dorsally, without markings (N. capensis) or with darker spots and irregular blotches on disc margins (N. japonica), or with paired white spots on disc posteriorly and white bars on tail laterally (N. dipterygia); usually creamy white or grey ventrally.

Easily separated from other narkid genera in the region by its single dorsal fin and pelvic fins not bilobed. Biology poorly known, even as the best-known genus in the family. Specimens of *Narke* are rare in collections, impeding revisionary studies. Almost nothing known concerning variation in species. *Narke dipterygia*, however, shows significant variation over its extensive range and may require subdivision. Three species, 2 in WIO.

#### **KEY TO SPECIES**

1a	Dorsal colour uniformly yellowish brown to brown, without conspicuous spots or other markings; yellowish or creamy ventrally, but outer margins brownish; spiracle margins elevated as low ridge-like rims; pelvic fins broad and slightly rounded <b><i>N. capensis</i></b>
1b	Dorsal colour uniformly brownish to reddish brown, with pair of large white spots on disc posteriorly, and white bars on tail laterally and extending to above pelvic-fin bases; uniformly white to creamy ventrally; spiracle margins faintly or not distinctly elevated as rims above disc; pelvic fins angular

## Narke capensis (Gmelin 1789)

Cape sleeper ray

PLATE 56

Raja capensis Gmelin 1789: 1512 (Cape of Good Hope, South Africa). Astrape capensis: Müller & Henle 1841; Günther 1870. Narke capensis: Garman 1913; Von Bonde & Swart 1923; Barnard 1925;

SFSA No. 89\*; Smith 1964\*; Wallace 1967\*; SSF No. 24.2\*; Compagno *et al.* 1989\*; Compagno & Heemstra 2007; Carvalho 2016.

Disc circular,  $\sim \frac{3}{5}$  TL; anterior margin usually broadly rounded; snout region short. Spiracles circular, slightly larger than eyes, margins with prominent rims and  $\sim 3$  small papillae. Internasal flap short, with few pores. Teeth small, usually in 10-12/11-13 rows. Pelvic fins broad and long; dorsal-fin origin slightly posterior to pelvic-fin insertions; dorsal fin and caudal fin relatively low and broadly rounded; caudal fin larger than dorsal fin;

Disc brown dorsally, and pelvic fins with darker margins; yellowish or creamy ventrally, with brownish outer margins; some specimens with yellowish, brownish or whitish spots towards end of tail. Attains 38 cm TL.

PLATE 56



*Narke capensis*, 23 cm TL, adult female (South Africa). Source: Compagno & Heemstra 2007



Narke capensis, 25 cm TL, adult female (South Africa). Source: SSF

**DISTRIBUTION** Southern Africa: Namibia in southeastern Atlantic, to South Africa (Cape Point to KwaZulu-Natal) and possibly Mozambique and Madagascar in WIO.

**REMARKS** Males mature at 17.5–23 cm TL, and females at ~16 cm TL. Occurs on continental shelf, to ~180 m deep (usually in <100 m), most commonly on muddy bottom and in bays along south Cape coast of South Africa. Feeds primarily on polychaetes. A bycatch of inshore trawls.

# Narke dipterygia (Bloch & Schneider 1801)

## Spottail sleeper ray

*Raja dipterygia* Bloch & Schneider 1801: 359 (Tharangambadi, India). *Bengalichthys impennis* Annandale 1909: 48, Fig. 6, Pl. 3a (Balasore Bay, Odisha coast, India); Annandale 1910; Rao 1974\*.

*Astrape dipterygia*: Müller & Henle 1841; Günther 1870; Annandale 1909\*. *Narke dipterygia*: Garman 1913; Fowler 1941; Munro 1955\*; Bianchi 1985\*; Compagno & Last 1999\*; Kapoor *et al.* 2002; Compagno & Heemstra 2007; Carvalho 2016.

Disc circular (usually longer than wide in preserved specimens); anterior margin usually broadly rounded; snout short. Spiracles larger than eyes, with low rims. Internasal flap short and wide, with pores. Teeth small, in  $\sim$ 16–18 rows, not visible externally. Dorsal-fin origin over pelvic-fin insertions; dorsal fin and caudal fin broadly rounded, caudal fin larger than dorsal fin.

Body brownish, reddish brown or dark brown dorsally, with pair of whitish spots on disc posteriorly, and creamy blotches on disc above pelvic-fin bases and laterally on base of tail; creamy to whitish ventrally. Attains 35 cm TL.

**DISTRIBUTION** Indo-Pacific. WIO: Oman to India and Sri Lanka; elsewhere to Malaysia and Japan.

**REMARKS** Biology mostly unknown; litters of 4–6, and males mature at 15–18 cm TL. Occurs inshore and offshore on continental shelf, in tropical to temperate waters, over sandy and muddy bottoms; reportedly locally common and widespread at just 5–36 m deep. A bycatch in bottom-trawl and gillnet fisheries, but probably not used.

# ORDER RHINOPRISTIFORMES

# FAMILY **PRISTIDAE**

## Sawfishes

Marcelo R de Carvalho and Bernard Séret

Distinctive, extremely large-sized rays (up to 5 m TL; possibly >7 m TL) with blade-like hypertrophied rostrum (rostral saw) that extends forward from head and reaches 1/5-1/3 TL; body shark-like, stout, and moderately depressed; gill openings, mouth and nostrils entirely ventral; 5 small paired gill slits on underside of head (at front half of pectoral-fin bases). No paired ventral barbels on snout. Rostrum with perpendicular lateral rostral teeth (modified dermal denticles) that grow continuously but are not replaced when broken or lost (unlike the much smaller deepwater sawsharks of family Pristiophoridae), and without smaller rostral teeth between large ones; posteriormost rostral teeth well ahead of nostrils. Eves dorsolateral on head and well anterior to spiracles. Mouth transverse and straight, without knobs or depressions. Nostrils anteriorly set, well-separated from each other and completely separate from mouth, without oronasal grooves; anterior nasal flaps short, not connected to each other, and not reaching mouth. Teeth small, rounded-oval, in numerous (up to 60) rows in each jaw and set in pavement-like bands, and without cusps. Pectoral fins relatively small, somewhat disjunct from head and not forming complete or continuous disc, inner rear tips free; pectoral-fin origins well posterior to mouth, attached to rear part of head over gill slits, and fins ending well ahead of pelvic-fin origins. Pelvic fins angular, not subdivided into anterior and posterior lobes. Two large, equal-sized and widely separated dorsal fins, tall, with similar angular or rounded-angular shape and with distinct apices and anterior, posterior and inner margins, and well-developed free rear tips varying from triangular to strongly falcate; 1st dorsal fin over or partially in front of pelvic fins. No anal fin. Peduncle moderately depressed and with keels. Tail not abruptly narrower than trunk. Caudal fin large, angular, shark-like and strongly asymmetrical, with vertebral axis raised above body axis; caudal-fin lower lobe well-developed or poorly developed. Body yellowish, brownish, grey-brown or greenish dorsally and on flanks, and pale or white ventrally, and without prominent markings although fins may be slightly dusky.

Reproductive mode yolk-sac (lecithotrophic) viviparity, with litters of 1–23 (usually 6–9), but the reproductive cycle of most species is poorly known; some species reach maturity

only after ~20 years. Relatively sluggish; benthic, found predominantly over muddy and sandy bottoms. Widely distributed in tropical and subtropical inshore coastal waters, to only ~10 m deep, and frequently present in estuaries, river mouths and inland freshwater (some populations may breed in freshwater). Feed on benthic organisms and small schooling bony fishes, using the saw-like rostrum to dislodge benthic prey items or to slash at schooling fishes. Mostly caught as a bycatch of small local inshore and freshwater fisheries and by recreational anglers. Sawfishes can inflict injuries to their captors when struggling individuals are removed from nets or line gear; occasional, but largely provoked, attacks on people have occurred in shallow water. One of the most at-risk groups of sharks or rays due to the degradation of the limited inshore marine, brackish and inland freshwater habitats they occur in, and extreme vulnerability to gillnets and bottom trawls, with numerous local extinctions having already occurred. IUCN Red List conservation status of all species is Endangered or Critically Endangered globally, and all species are listed on Appendix I of CITES, banning their trade.

Sawfishes are sometimes confused with sawsharks of family Pristiophoridae, but can be easily distinguished from them by a much larger size, ventrally situated gill slits, and lacking long barbels on underside of rostrum. Two genera and 5 species currently recognised as valid (of 23 available nominal species names, as these are usually applied solely on a geographical basis); both genera and 3 species in WIO.

#### **KEY TO GENERA**



# GENUS Anoxypristis White & Moy-Thomas 1941

Rostral teeth in 16–29 pairs, and not present on basal portion of rostrum, irregularly spaced, greatly flattened, blade-like, narrowly triangular, and with single posterior edges. Rostrum relatively slender, narrowing slightly near base. Lateral edges of head and incurrent apertures of nostrils connected by broad incurrent grooves on underside of snout. Nostrils narrow, long, oblique. Pectoral-fin bases narrow, and fins shark-like. First dorsal fin posteriorly situated, its origin slightly behind pelvic-fin insertions. Peduncle with secondary keel below main keel. Caudal fin with shallow subterminal notch, lower lobe prominent, and posterior margin deeply concave. One species currently recognised.

# Anoxypristis cuspidata (Latham 1794)

Narrow sawfish

PLATE 51

Pristis cuspidatus Latham 1794: 279, Pl. 26, Fig. 3 (Malabar, India); Klunzinger 1871; Annandale 1909\*; Blegvad & Løppenthin 1944\*; Munro 1955\*; Qureshi 1972\*.

*Squalus semisagittatus* Shaw 1804: 361 (Visakhapatnam, India) [based on Russell 1803: Pl. 13].

Anoxypristis cuspidata: Debelius 1993\*; Randall 1995\*; Compagno & Last 1999\*; Last & Stevens 2009\*; Faria et al. 2013; Last et al. 2016.

Diagnosis as for genus. Regional and individual variation noted in pairs of rostral teeth (males usually with lower tooth counts than females); rostrum length >12 times its width at level of posteriormost lateral teeth.

Adults greyish, greyish blue or slightly greenish dorsally, and young darker grey; paler ventrally; fins dusky or pale. Attains 4.7 m TL, possibly 6.1 m TL.

**DISTRIBUTION** Indo-Pacific. Historic range uncertain; current populations are rare and fragmented. WIO: Red Sea, Gulf of Aden, Persian/Arabian Gulf to India and Sri Lanka.

**REMARKS** Viviparous with large yolk sac retained until about birth; uterine eggs enclosed in soft egg cases, and uterine wall frilly, glandular, and reported to produce histotroph (uterine secretions). Litters of 6-23 (average 12-15); size at birth (free-swimming neonates) 50-60 cm TL; gestation period 4-5 months. Males mature at ~4 years, and females at ~5 years, at 2.5-4.7 m TL. Apparently the most productive sawfish species. Now rare, but formerly common inshore, in shallow water down to at least 40 m (possibly ~100 m), also in brackish water in some river deltas, estuaries and shallow bays; appears to penetrate some rivers (in India, Myanmar and Thailand), but freshwater occurrences need verification. Relatively active (as compared with more benthic Pristis species) as evidenced by its more streamlined shark-like form, smaller pectoral fins, stouter tail with secondary keel on peduncle, and semi-lunate caudal fin. Feeds on fishes, cuttlefish (Sepiidae) and prawns. Highly vulnerable to bottom-trawl and gillnet fisheries. IUCN Red List conservation status Endangered (previously Critically Endangered).



Anoxypristis cuspidata, ~80 cm TL (W India). Composite

# GENUS Pristis Linck 1790

Rostral teeth in 14–34 pairs (usually 20–28), and extend to base of rostrum, triangular in cross-section and with double posterior edges. Rostrum robust, widest at its base, ~5–7 times longer (*Pristis pristis*) or 7–10 times longer (*Pristis pectinata* group) than its greatest width. Nostrils short, broad, transverse, and with large anterior nasal flaps. Underside of snout without broad incurrent grooves connecting lateral edges of head and incurrent apertures of nostrils. Pectoral fins somewhat low, relatively long, with broad base. First dorsal fin anteriorly situated, its origin anterior to, over or posterior to pelvic-fin origins. Peduncle without secondary keel. Caudal fin without subterminal notch.

Two groups of species in the genus: *Pristis pristis*, with proportionally larger and heavier rostrum, usually fewer rostral teeth, and distinct caudal-fin lower lobe (formerly including other species such as *P. microdon*, now a single widespread species recognised); and *Pristis pectinata* group, with proportionally smaller and less heavy rostrum, usually more rostral teeth, and caudal-fin lower lobe not well-defined. *Pristis pristis* with definitive freshwater records. Four valid species, at least 2 in WIO.

#### **KEY TO SPECIES**

- 1a First dorsal-fin origin in front of pelvic-fin origins .......*P. pristis*1b First dorsal-fin origin above or behind

## Pristis pristis (Linnaeus 1758)

#### Largetooth sawfish

*Squalus pristis* Linnaeus 1758: 235 ('Europa' [Mediterranean Sea; western Atlantic; Indian Ocean]).

Pristis microdon Latham 1794: 280, Pl. 26, Fig. 4 [no locality given];
SFSA No. 58; Munro 1955\*; Wallace 1967\*; Qureshi 1972\*;
Van der Elst 1988\*; Compagno *et al.* 1989\*; SSF No. 22.1\* [1995];
Compagno & Last 1999\*; Heemstra & Heemstra 2004\*; White *et al.* 2006\*; Last & Stevens 2009\*.

Pristis pectinatus: Barnard 1925\*; SFSA No. 59\*.

Pristis annandalei Chaudhuri 1908: 391, Fig. (Elephant Point, Myanmar). Pristis perotteti: Annandale 1909\*; Barnard 1925 [as perrotteti]. Pristis pectinata: Compagno et al. 1989\*.

Pristis pristis: Faria et al. 2013; Last et al. 2016.

Rostrum broad and stout, not significantly tapering anteriorly; rostrum length ~5–6 times its width at posteriormost lateral teeth. Pairs of rostral teeth 14–23 (with known variation between individuals, and males frequently with higher tooth count than females), moderately flattened, with double posterior edges, and a groove on posterior edge of lateral teeth at all stages. First dorsal-fin origin well ahead of pelvic-fin origins, and its insertion close behind pelvic-fin origins.

Body yellowish brown to greyish dorsally, white ventrally; fin webs yellowish. Attains 6.5 m TL, possibly 7 m TL.

**DISTRIBUTION** Circumglobal, with numerous suspected local extinctions. WIO: South Africa (Eastern Cape and KwaZulu-Natal), Mozambique, Madagascar, Réunion, Tanzania, Somalia, Oman, Pakistan and India.



Pristis pristis, 357 cm TL, mature male (South Africa). Source: SSF 1995

PLATE 51

**REMARKS** Species identity for 'largetooth sawfishes' is confined to a single wide-ranging species with distinct subpopulations, which are considered ecologically (rather than genetically) different. Pristis microdon is a former name most commonly applied to records from the Indo-Pacific, from South Africa to northern Australia. Many local records of this species are of juveniles from freshwater and upper estuaries, as large individuals are now rarely seen. Viviparous with volk sac; litters up to 13; size at birth 70-90 cm TL; gestation period ~5 months. Slow-growing; matures at 2.5-3 m TL, at ~8-9 years. Inshore, intertidal to commonly <10 m deep, and spending part of its lifecycle in freshwater systems. Feeds on small fishes and benthic invertebrates. Formerly common, its global abundance has declined sharply due to intense human pressure and vulnerability to entanglement in all types of fishing gear. IUCN Red List conservation status Critically Endangered globally.

## Pristis zijsron Bleeker 1851

#### Green sawfish

PLATE 52

Pristis zijsron Bleeker 1851: 417, 442 (Bandjarmasin, Borneo, Indonesia);
Munro 1955\*; Qureshi 1972\*; Compagno et al. 1989\*; Randall 1995\*;
SSF No. 22.3\* [1995]; Compagno & Last 1999\*; Bonfil & Abdallah 2004\*;
Last & Stevens 2009\*; Faria et al. 2013; Everett et al. 2015.
Pristis zysron: Annandale 1909; Blegvad & Løppenthin 1944\*.
Pristis pectinatus: Annandale 1909\*; Barnard 1925\*; Smith 1949\*.
Pristis pectinata: Compagno et al. 1989\*; Debelius 1993\*; SSF No. 22.2\*

[1995]; Compagno & Last 1999\*; Last & Stevens 2009\*.

Rostrum relatively slender, with sides nearly parallel near base; rostrum width at base  $\frac{1}{10}-\frac{1}{7}$  its length. Pairs of rostral teeth 23–34; lateral teeth moderately flattened, elongated, with double posterior edges and groove on posterior edges in adults. First dorsal-fin origin somewhat posterior to pelvic-fin origins but ahead of pelvic-fin insertions. Peduncle with no secondary keel below main keel.

Body greenish or greyish brown to olive dorsally, pale or white ventrally. Attains at least 5.4 m TL, possibly 7.3 m TL.

**DISTRIBUTION** Indo-Pacific, with localised extinctions probable. WIO: extant in southwestern Red Sea to Gulf of Aden, Persian/Arabian Gulf to Gulf of Oman, and off Pakistan, India and Sri Lanka; also East Africa off Kenya, Mozambique and Madagascar; presumed extinct from South Africa (KwaZulu-Natal) and Mascarenes (Everett *et al.* 2015).

**REMARKS** Probably the largest sawfish species, but large individuals have become rare. Life history poorly known; presumably more tolerant of cooler waters than other *Pristis* species. Lecithotrophic viviparity, with litters of ~12; size at birth ~80 cm TL. Matures at 3–4.3 m TL, at ~9 years. Inhabits inshore coastal waters, estuaries and freshwater river systems, most common in shallow waters, but to ~70 m deep. Feeds on small schooling fishes, crustaceans and other benthic invertebrates. Presumed extirpated from many areas of its native range due to intensive inshore gillnet and trawl fisheries and habitat loss. IUCN Red List conservation status Critically Endangered.



Pristis zijsron, 109 cm TL, immature male (South Africa). Source: SSF 1995

# FAMILY RHINIDAE

# Shark ray and wedgefishes

David A Ebert, Neil C Aschliman and Brett A Human

Medium to large shark-like rays (up to ~3.1 m TL), with depressed robust body, head either thick and broadly rounded, or flattened and wedge-shaped; and long tail, robust or flattened. No stinging spine on tail and no electric organs. Thorns variably developed above eyes, on each shoulder, and along dorsal midline; dermal denticles on dorsal surface otherwise granular, minute. Spiracles large, with or without skin folds at rear margins. Five paired gill slits on underside. Nostrils large, oblique; nostril length more than one-half mouth width. Large triangular pectoral fins, separate, not joined to body to form disc. Two large, upright to falcate, spineless dorsal fins; 1st dorsal fin larger than 2nd, its origin over or slightly before pelvic-fin mid-bases or origins. Caudal fin large, posterior margin concave; upper lobe longer than lower lobe.

Reproductive mode yolk-sac viviparity where known; at least one with supplementary uterine fluid. Wedgefishes and shark rays have been observed perched on their pectoral-fin tips with the anterior part of their body off the bottom. A common bycatch in artisanal and commercial fisheries and recognised as strong gamefish. Their high-quality flesh, highlyvalued large fins, and susceptibility to a range of fishing gear, combined with an inshore habitat (from intertidal zone to ~90 m deep), raises conservation concerns.

Found in tropical to warm-temperate waters of eastern Atlantic, western Pacific and Indian oceans. Three genera and 10 species; 2 genera and at least 4 species in WIO. Although there are substantial morphological and anatomical character differences between the 2 genera such that they formerly comprised separate families, a recent molecular analysis indicated that they are sister groups and combined them into family Rhinidae (Last, Séret & Naylor 2016).

#### **KEY TO GENERA**

# GENUS **Rhina** Bloch & Schneider 1801

Unmistakable bulky, shark-like ray with domed head and robust tail. Heavy ridges of spiky thorns above eyes, on central dorsal midline, and on shoulder. Spiracles without skin folds. Mouth broadly arched; lower jaw trilobed; numerous small blunt teeth in undulating rows and visible when mouth closed; tooth count 47/50. One species.

# Rhina ancylostomus Bloch & Schneider 1801

Shark ray

PLATE 57

*Rhina ancylostomus* Bloch & Schneider 1801: 352, Pl. 72 (Coromandel, India).

Rhina ancylostoma: SFSA No. 59a\*; Wallace 1967\*; SSF No. 27.1\*; Compagno, Ebert & Smale 1989\*; Manilo & Bogorodsky 2003; Heemstra & Heemstra 2004\*; Last & Stevens 2009\*; Last *et al.* 2016.

#### Diagnosis as for genus.

Body bluish grey to brownish dorsally, with numerous pale spots scattered over body, fins and tail (markings most pronounced in juveniles); mostly white ventrally. Attains 270 cm TL.



Rhina ancylostomus, 54 cm TL, juvenile (Oman).



Rhina ancylostomus, 130 cm TL, female (South Africa). Source: SSF

**DISTRIBUTION** Indo-Pacific (widespread). WIO: South Africa (KwaZulu-Natal) to Arabian Sea, Red Sea, Persian/ Arabian Gulf, Pakistan, India, Sri Lanka, Seychelles, Madagascar, Réunion and Mauritius; elsewhere to east coast of India, Philippines, Taiwan, southern Japan, Australia and New Caledonia.

**REMARKS** Viviparous with yolk sac and supplementary uterine fluid; litters of 2–11; size at birth 46–48 cm TL. Males mature at 150–175 cm TL, females mature at ~180 cm. Uncommon but widely scattered; coastal, demersal, found close inshore on sandy and muddy bottoms and around coral reefs, possibly to ~90 m deep. Diet includes crustaceans (crabs) and other shellfish. A target species and bycatch in multiple artisanal and commercial fisheries throughout its range; traded in Asia for its fins. Harmless to people and a popular exhibit animal in public aquariums. IUCN Red List conservation status Critically Endangered globally; Near Threatened in Australian waters.

## GENUS Rhynchobatus Müller & Henle 1837

Head depressed and not elevated above disc, anterior margins not indented; snout acutely pointed, triangular and depressed, and rostrum not forming a 'saw.' Mouth opening weakly undulating to nearly straight; circumoral grooves and lip folds present. Tail with well-developed lateral ridges. Pectoral fins rather small, originating posterior to nostrils and confluent with head and body, apices angular, and forming short narrow wedge-shaped disc; free rear tips ending well before pelvic-fin origins. Spiracles with 2 skin folds on rear margin. Supraorbital and scapular thorns inconspicuous, and males without alar or malar thorn patches; thorns in dorsal midline in front of 1st dorsal fin usually small, blunt; interdorsal thorns inconspicuous, and no thorns between 2nd dorsal fin and caudal fin. Tooth counts 31–52/37–48; vertebrae 144–211.

This genus is in need of revision to improve our understanding of the number of species and their distribution. Until recently, *Rhynchobatus djiddensis* (commonly known as the giant wedgefish) was thought to be the only representative in the Indian Ocean, ranging to the western Pacific. However, there may be at least 3 similar-looking species in the Indian Ocean, and *R. djiddensis* may not occur beyond WIO. Thus, the synonymies and distributions provided here are highly provisional. Eight species, plus an unknown number of undescribed species; 3 species, plus at least 1 undescribed species in WIO.

As a taxonomically problematic group, a key to species cannot be presented due to the lack of critical species descriptions. Characters and colouration listed for the species below are highly provisional for WIO representatives, as detailed redescriptions of all putative species are needed.

# Rhynchobatus australiae Whitley 1939

Bottlenose wedgefish

PLATE 57

*Rhynchobatus djiddensis australiae* Whitley 1939: 245, Fig. 14 (off Manning River mouth, New South Wales, Australia). *Rhynchobatus australiae*: Compagno & Last 1999\*, 2008;

White & Dharmadi 2007; Last & Stevens 2009\*; Bineesh *et al.* 2014.

Disc margin beside eyes concave; thorns not set on raised ridges; 1st dorsal-fin origin slightly behind pelvic-fin origins. Vertebrae 144–164.

Body pale grey to yellowish brown dorsally and ventrally; dorsal surface with few large white spots loosely arranged in 2 rows on each side of trunk, and well-defined dark spot surrounded by 4 white spots at base of each pectoral fin. Attains 3 m TL.

**DISTRIBUTION** WIO records: Mozambique to Arabian Sea, Red Sea, Persian/Arabian Gulf, Pakistan, India, Sri Lanka, Madagascar, and Seychelles. Indo-Pacific: tropical eastern Indian Ocean and western Pacific Ocean.

**REMARKS** Probably a species complex of 3 or 4 difficult-todistinguish species; smaller individuals are easily misidentified with *R. laevis*. Very little known about its behaviour and biology. Litters of 7–19; size at birth 46–50 cm TL. In the Pacific, males mature at 110–130 cm TL, and females at ~150 cm TL. IUCN Red List conservation status Critically Endangered.

# Rhynchobatus djiddensis (Forsskål 1775)

Whitespotted wedgefish

PLATE 57

Raja djiddensis Forsskål in Niebuhr 1775: 18, viii (Red Sea).
Rhynchobatus djeddensis: Sauvage 1891; Barnard 1925\*, 1927\*;
Smith 1959\*; SFSA No. 60\*; Wallace 1967\*.
?Rhinobates rueppelli: Swainson 1838.
Rhynchobatus djiddensis: Morrow 1954; Compagno & Smale 1986;
SSF No. 27.7\*; Compagno & Randall 1987; Compagno et al. 1989\*;
Randall & Anderson 1993; Cliff & Wilson 1994\*;
?Al-Abdessalaam 1995\*; ?Randall 1995\*; ?Branch et al. 1999;
Compagno & Last 1999, 2008; Bonfil & Abdallah 2004\*;

Heemstra & Heemstra 2004\*; Letourneur et al. 2004;

Last & Stevens 2009.

Disc margin slightly concave in front of eyes; 1st dorsal-fin origin above or before pelvic-fin origins. Tooth count 31–42/37–48; vertebrae 204–211.

Body dark olive-green or yellowish brown dorsally and ventrally; dorsal surface with 3 or more rows of loosely arranged large white spots on each side of trunk, and welldefined dark spot surrounded by 4 or 5 white spots at each pectoral fin base; sometimes with dark polygonal mark between eyes. Attains 3.1 m TL.



Rhynchobatus djiddensis, 146 cm TL (South Africa). Source: CFSA

**DISTRIBUTION** Extent of global distribution difficult to ascertain due to taxonomic confusion; records from western Pacific and eastern Indian oceans probably represent a species complex. WIO: South Africa (Eastern Cape and KwaZulu-Natal) to Red Sea, Gulf of Aden, and to Persian/Arabian Gulf, Madagascar, Mauritius, Réunion, Seychelles and Maldives.

**REMARKS** The most commonly cited species of Rhynchobatus, yet poorly described. Previously considered wide-ranging in tropical to warm-temperate waters of Indo-Pacific, recent literature restricts it to the coast of East Africa and the Red Sea, being replaced by a similar-looking species from approximately Pakistan (Arabian Sea) and eastwards, and probably sympatric with at least one other possibly undescribed species in WIO (photographs of Rhynchobatus from Oman have a similar colour pattern to R. djiddensis from East Africa). Colouration reported to change with undisturbed and disturbed states, resulting in more prominent patterning in the latter, and probably adding to taxonomic confusion. Viviparous with yolk sac; litters of probably 4-7; size at birth 55-67 cm TL. Seasonal occurrence in South Africa during the summer is associated with pupping. Inshore over mud and sand bottoms, from surf line to ~30 m deep, and also caught in estuaries. Diet includes crabs, bivalve molluscs, squid and fishes. Common bycatch in nearshore artisanal and commercial fisheries where it occurs, and valued as a gamefish; highly valued for its fins and palatability of its meat. IUCN Red List conservation status Critically Endangered.

# Rhynchobatus laevis (Bloch & Schneider 1801)

#### Smoothnose wedgefish

Rhinobatus laevis Bloch & Schneider 1801: 354, Pl. 71 (Japan). Rhinobatos laevis: Bloch & Schneider 1801\*. Rhynchobatus djeddensis: Day 1878\*; Bianchi 1985\*; Morón et al. 1998. Rhinobatis djeddensis: Annandale 1909. Rhynchobatus djiddensis: Carpenter et al. 1997\*. Rhynchobatus cf. laevis: Compagno & Last 1999\*. Rhynchobatus laevis: Compagno & Last 2008; Last & Stevens 2009\*.

PLATE 57

Diagnosis as for family. Disc margin slightly concave beside spiracles; 1st dorsal-fin origin over or slightly behind pelvic-fin origins. Vertebrae 144–149.

Body dark greyish brown to almost black dorsally, with few large white spots loosely arranged in 2 rows on each side of trunk; sometimes with a poorly defined dark spot and 3 white spots near it (not surrounding it) at each pectoral-fin base; paler ventrally. Attains possibly 200 cm TL.



Rhynchobatus laevis, 71 cm TL (Thailand). © JE Randall, Bishop Museum

**DISTRIBUTION** Indo-Pacific. WIO: Mozambique, Madagascar, Oman, possibly Persian/Arabian Gulf, Pakistan, India and Sri Lanka; possibly more wideranging, but unconfirmed; elsewhere to Thailand, Indonesia, Japan and Australia.

**REMARKS** Occurs in tropical waters of northern Indian Ocean, perhaps along with *R. australiae* or a similar-looking possibly undescribed species, and generally mistaken for *R. djiddensis* although probably allopatric to that species. Behaviour and biology poorly known, especially in WIO; litters of 7–26 (average 15) are reported elsewhere. Feeds mostly on benthic invertebrates. Taken heavily in gillnet fisheries in India, as well as in other fisheries where it occurs. Valued for its meat and fins, and a prized gamefish in Australia. IUCN Red List conservation status Critically Endangered.

# FAMILY RHINOBATIDAE

## Guitarfishes and shovelnose rays

Bernard Séret and Marcelo R de Carvalho

Small- to medium-sized rays (up to ~3 m TL) with wedgeshaped disc, depressed shark-like trunk, and thick tail. Head flattened and pectoral fins fused with head and trunk; snout more or less elongated, acutely to obtusely angular, and supported by stout rostral cartilage. Spiracles close to eyes, usually large, and with 1 or 2 skin folds along rear edge. Five pairs of gill slits on ventral side. Nostrils rather short (subequal to or slightly longer than internasal space), very oblique, without nasal curtain; anterior nasal flaps more or less well-developed, with medial extension of the inner margins onto internasal space defining genera. Two large, nearly equal-sized dorsal fins; 1st dorsal fin well behind pelvic fins. Pelvic fins large, not forming separate lobes, and origins under posterior margins of pectoral fins. Caudal fin strongly asymmetrical with well-developed upper lobe and small or absent lower lobe; no electric organs. Skin covered with granulations, and often with small thorny tubercles around eyes and spiracles and/or blunt tubercles in median dorsal row. Teeth in jaws numerous, small and blunt. Colour variable, either plain dorsally or with distinctive pattern of more or less symmetrically arranged spots, blotches, ocelli or bands; usually white ventrally, sometimes with dark blotch on underside of snout.

Reproductive mode of all species yolk-sac viviparity. Circumglobal and widely distributed, in warm-temperate to tropical waters, mostly inshore over continental shelf, although some occur in deeper water, to ~400 m. Some also enter estuaries and closed bays. Mainly bottom-dwelling, often resting on soft mud or sandy bottom, with shallow sandy habitat important as nursery areas. Feed mainly on benthic invertebrates and small bony fishes. Taken mostly as a bycatch of artisanal and commercial benthic trawl, gillnet, and beachseine fisheries; appreciated in many countries for their meat, but fins usually too small for the shark-fin trade. IUCN Red List conservation status of the species varies from Endangered to Near Threatened.

The large genus *Rhinobatos* has been typically understood to comprise three subgenera: *Rhinobatos* Linck 1790, *Glaucostegus* Bonaparte 1846, and *Acroteriobatus* Giltay 1928 (cf. Last *et al.* 2004). The subgenus *Glaucostegus* was elevated to generic level by Compagno *et al.* (2005), and has now been placed in its own family, Glaucostegidae (Last *et al.* 2016). The other two groups left as subgenera of *Rhinobatos*, *Acroteriobatus* and *Rhinobatos*, are now elevated to full genera. Three genera and 33 species; 2 genera and 16 species in WIO, plus at least 2 other undescribed species.

#### **KEY TO GENERA**



# GENUS Acroteriobatus Giltay 1928

Distinguished from other rhinobatid genera in the region by well-developed anterior nasal flaps, inner edge of nostrils extending medially well onto internasal space and almost joining at snout midline; snout tip broadly acute; and rostral cartilage ridges widely separated but with slight constriction midway. Ten species, all in WIO, with 2 recently described species, *Acroteriobatus andysabini* and *A. stehmanni* Weigmann, Ebert & Séret 2021 in the key.

#### **KEY TO SPECIES**

1a	Snout broadly pointed; disc wider, anterolateral margins conspicuously convex
1b	Snout more or less acutely pointed; disc narrower, anterolateral margins straight to slightly convex or slightly concave
2a	Dorsal surface plain brown (but with some small whitish spots in young)
2b	Dorsal surface pale brown and with dark-edged pale blue blotches on pectoral fins and pelvic fins
3a	Dorsal surface plain coloured; medium-sized, to ~200 cm TL
3b	Dorsal surface with spots, blotches or bands
4a	Snout with 2 or 3 brown bands on each side; pectoral fins and pelvic fins with blue variegated markings
4b	No dark bands on snout5

Continued ...

#### **KEY TO SPECIES**

5a	Blotches on dorsal surface not dark-edged, but sometimes edged by rings formed of dots; large bluish grey blotches on snout, greyish brown blotches on pectoral fins and pelvic fins, and symmetrically paired faint brown blotches on back of trunk
5b	Blotches on dorsal surface usually dark-edged, and rings continuous and not formed of dots
6a 6b	Numerous blotches on dorsal surface   7     Fewer blotches on dorsal surface   9
7a	Dorsal surface with numerous large blue-grey ocellus-like blotches with narrow dark brown edges, sometimes incomplete
7b	Dorsal surface with numerous relatively small brown blotches or dark-edged ocelli
8a	Centre of ocelli consisting of a small dark spot, or dorsal surface with brown blotches
8b	Centre of ocelli consisting of a small white spots A. omanensis
9a	Body brownish with orange tinge, blue-grey spots more numerous, ventral snout with V-shaped dark mark <i>A. andysabini</i> [Madagascar; Plate 58]
9b	Body brownish with greenish tinge, blue-grey spots fewer, ventral snout without dark marking

## Acroteriobatus annulatus (Smith 1841)

Lesser guitarfish

PLATE 58

Rhinobatus (Syrrhina) annulatus Smith (ex Müller & Henle) 1841: 116 (Cape of Good Hope, South Africa); Wallace 1967\*.

Rhinobatos annulatus: SSF No. 27.2\*; Compagno et al. 1989\*; Heemstra & Heemstra 2004\*.

Acroteriobatus annulatus: Last et al. 2016.

Disc moderately broad, anterolateral margins slightly convex; disc length ~39% TL, disc width ~1.15 in disc length. Snout triangular, bluntly pointed and short, preorbital length ~14% TL. Eyes slightly larger than spiracles; 2 prominent folds at rear edge of spiracles. Small tubercles around eyes and spiracles, and ~35 tubercles in median dorsal row.

Body pale brown background dorsally, with distinctive pattern of numerous dark-edged ocelli, with central dark spot surrounded by pale ring or else centre simply brownish; white ventrally. Attains 140 cm TL.



Acroteriobatus annulatus, 53 cm TL, male (South Africa). Source: SSF

**DISTRIBUTION** Southern Africa: Angola in southeastern Atlantic, to South Africa and southern Mozambique in WIO.

**REMARKS** Recent genetic analyses show no differences between the Atlantic Ocean and WIO (KwaZulu-Natal) forms. Litters of 2–10. Matures relatively quickly, males at ~59 cm TL and females at 68–89 cm TL; maximum age ~7 years. Most abundant rhinobatid in southern Africa; occurs from surfline to ~100 m deep, and in estuaries and enclosed bays, with shallow sandy areas serving as important nursery habitat. Feeds on small benthic invertebrates (mainly crustaceans) and small bony fishes. Caught by recreational shore anglers and in commercial gillnet, beach-seine and benthic trawl fisheries.

# Acroteriobatus blochii (Müller & Henle 1841)

Bluntnose guitarfish

PLATE 58

Rhinobatus (Syrrhina) blochii Müller & Henle 1841: 115, Pl. 37 (Cape of Good Hope, South Africa).
Rhinobatos blochii: SSF No. 27.3\*; Compagno et al. 1989\*.
Acroteriobatus blochii: Last et al. 2016.

Disc moderately broad and moderately long; anterolateral margins slightly convex, posterior margins entirely rounded; disc length ~42% TL, disc width ~1.1 in disc length. Snout acutely rounded and very short, preorbital length ~11% TL. Eyes slightly larger than spiracles; only 1 prominent fold at rear edge of spiracles. Minute tubercles around eyes, and small tubercles in median dorsal row (reducing with growth).

Body plain brown dorsally, but young with few white spots; white ventrally. Attains 96 cm TL.



Acroteriobatus blochii. Source: SSF, composite

PLATE 58

**DISTRIBUTION** Southern Africa: Namibia (Walvis Bay) in southeastern Atlantic, to Cape of Good Hope (South Africa) in WIO.

**REMARKS** Often misidentified with *A. annulatus* where they overlap in distribution. Rare, biology unknown; occurs close inshore, in shallow bays and off sandy beaches. Occasionally taken in recreational shore fisheries.

## Acroteriobatus leucospilus (Norman 1926)

Greyspotted guitarfish

PLATE 58

Rhinobatus leucospilus Norman 1926: 966, Fig. 18 (Durban, KwaZulu-Natal, South Africa); Wallace 1967\*.

Rhinobatos leucospilus: SSF No. 27.5\*; Compagno et al. 1989\*;

Heemstra & Heemstra 2004.

Acroteriobatus leucospilus: Last et al. 2016.

Disc moderately broad, anterolateral margins almost straight to slightly convex; disc length ~38% TL, disc width ~1.2 in disc length. Snout bluntly triangular and short, preorbital length ~12% TL. Eyes slightly larger than spiracles; 2 folds at rear edge of spiracles, outer fold more prominent. Very small tubercles around eyes and spiracles and on shoulders, and up to 50 tubercles in median dorsal row.

Body sandy brown background dorsally, with symmetrically arranged faint brown blotches across disc and trunk, and bluish spots on snout and pectoral- and pelvic-fin margins; dorsal fins and caudal fin with fainter grey-brown blotches and bluish spots; margins of pectoral fins and pelvic fins with bluish grey edges; white ventrally. Attains 120 cm TL.



Acroteriobatus leucospilus, 68 cm TL, female (South Africa). Source: SSF

**DISTRIBUTION** WIO: South Africa (Transkei region and KwaZulu-Natal) to Mozambique.

**REMARKS** Litters of 2–4; matures at ~56 cm TL. Apparently rare and poorly known; occurs from surfline to ~100 m deep. Feeds on benthic invertebrates (bivalves, gastropods, crabs, shrimp) and small bony fishes. Possibly taken by recreational shore anglers and in gillnet fisheries.

# Acroteriobatus ocellatus (Norman 1926)

#### Maculate guitarfish

*Rhinobatus ocellatus* Norman 1926: 967, Fig. 20 (Bird Island, Algoa Bay, South Africa); Wallace 1967\*.

*Rhinobatos ocellatus*: SSF No. 27.6 (not illustration); Compagno *et al.* 1989\*; Heemstra & Heemstra 2004.

Acroteriobatus ocellatus: Last et al. 2016.

Disc moderately narrow, anterolateral margins straight to slightly concave; disc length ~36% TL, disc width ~1.2 in disc length. Snout bluntly triangular and relatively short, preorbital length 12–13% TL; 2 prominent folds at rear edge of spiracles. Small tubercles around eyes and spiracles and on shoulders and dorsal midline in young (becoming lost with growth and maturity).

Body brownish background dorsally, with numerous large ocellus-like blotches, with blue-grey centre and narrow dark brown edges, dark rims sometimes incomplete; blotch size decreases towards margins. Attains 81 cm TL.



Acroteriobatus ocellatus (South Africa). Composite

**DISTRIBUTION** WIO: South Africa (south coast) to southern Mozambique.

**REMARKS** Often misidentified with *A. annulatus* and *A. zanzibarensis*. Rare and little known from few specimens. Found along sandy coastal areas, usually close inshore, at 60–185 m. Likely taken as a bycatch in trawl and net fisheries.

## Acroteriobatus omanensis

Last, Henderson & Naylor 2016

#### Oman guitarfish

Acroteriobatus sp.: Henderson et al. 2016.

Acroteriobatus omanensis Last, Henderson & Naylor 2016: 277, Figs. 1–7 (off Muscat, Gulf of Oman). Disc broadly wedge-shaped, anterolateral margins slightly undulated; disc length 41.4–43.3% TL, disc width 1.3–1.4 in disc length. Snout bluntly angular and short, preorbital length 12.9–15.3% TL. Eye length 1.3–1.6 times spiracle length; 2 well-developed spiracular folds. Denticles minute (skin smooth to touch); some enlarged denticles on snout over orbits, on shoulders, on dorsal midline from nape to first dorsal origin, and between dorsal fins.

Dorsal surface medium greenish brown, paler along pectoral and pelvic posterior margins, with symmetrically arranged faint dark brown blotches and numerous small ocelli consisting of white central spot surrounded by a dark, diffuseedged brownish rim; dorsal fins with small white spots, caudal fin with two large dark blotches, one each side, and a pair of dark saddles on caudal peduncle. Ventral surface whitish, with diffuse dark marking on snout tip; margins of nasal flaps vivid red or orange. Attains at least 60 cm TL.

DISTRIBUTION WIO: Gulf of Oman.

**REMARKS** Biology unknown. Described from three adult females and a male collected from Muscat fish market.

## Acroteriobatus salalah (Randall & Compagno 1995)

Salalah guitarfish

Rhinobatos salalah Randall & Compagno 1995: 293, Fig. 4 (Salalah fish market, Oman, Arabian Sea); Randall 1995\*; Manilo & Bogorodsky 2003;

Bonfil & Abdallah 2004\*.

Acroteriobatus salalah: Last et al. 2016.

Disc heart-shaped, somewhat roundish; anterolateral margins usually convex, posterior margins strongly convex; disc length ~37% TL, disc width ~1.1 in disc length. Snout tip bluntly pointed, snout broad and short; preorbital length ~11% TL; 2 prominent folds at rear edge of spiracles. Small tubercles around eyes and spiracles, and up to 55 blunt tubercles in median dorsal row.

Body tan dorsally, with dark-edged pale bluish spots on snout, pectoral fins and pelvic fins; white ventrally. Attains 78 cm TL.

**DISTRIBUTION** WIO: Arabian Sea (Oman and Pakistan).

**REMARKS** Known only from a few specimens collected from a fish market in Salalah, Oman, and a harbour in Karachi, Pakistan. Presumably occurs along sandy inshore areas. Likely taken in local fisheries.

## Acroteriobatus variegatus (Nair & Lal Mohan 1973)

#### Stripenose guitarfish

Rhinobatos variegatus Nair & Lal Mohan 1973: 77, Fig. 4 (Gulf of Mannar, India); Manilo & Bogorodsky 2003. Acroteriobatus variegatus: Last et al. 2016.

PLATE 59

Disc relatively narrow, anterolateral margins almost straight; disc length ~37% TL, disc width ~1.3 in disc length. Snout triangular, pointed, and moderately long, preorbital length ~13% TL; 2 developed folds at rear edge of spiracles. Minute tubercles in front of eyes, and ~36 small tubercles in median dorsal row.

Body brownish grey dorsally; pectoral fins and pelvic fins with blue variegated markings; snout with 2 or 3 brown longitudinal stripes on each side of rostral cartilage, and snout tip slightly darker; white ventrally. Attains at least 65 cm TL.



Acroteriobatus variegatus, 28 cm TL (SW India). KV Akhilesh © CMFRI

**DISTRIBUTION** WIO: Gulf of Mannar (India).

**REMARKS** Known from few specimens in 10–40 m. Deep records (to 366 m) are probably erroneous.

# Acroteriobatus zanzibarensis (Norman 1926)

#### Zanzibar guitarfish

PLATE 59

*Rhinobatus zanzibarensis* Norman 1926: 966, Fig. 19 (Zanzibar, Tanzania). *Acroteriobatus zanzibarensis*: Last *et al.* 2016.

Disc rather narrow; anterolateral margins almost straight to slightly concave. Snout bluntly triangular and moderately long, preorbital length ~12% TL; 2 well-developed folds at rear edge of spiracles. Small tubercles around eyes and spiracles, and in two groups on shoulders; small blunt tubercles in median dorsal row.

Body plain coloured dorsally, white ventrally. Attains at least 75 cm TL.



*Acroteriobatus zanzibarensis*, male holotype (Zanzibar). Drawn from photograph

DISTRIBUTION WIO: Tanzania (Zanzibar).

**REMARKS** Known only from two specimens (75 cm TL male and 20.5 cm TL female; depth of capture unknown); very similar if not identical to *A. ocellatus*, except for its plain colour and slightly concave anterolateral margins. IUCN Red List conservation status Near Threatened.

GENUS **Rhinobatos** Linck 1790

Distinguished from other rhinobatid genera in WIO by weakly developed anterior nasal flaps, inner margins extending to level of inner corner of nostrils; snout triangular and relatively acute; and rostral ridges well separated and more or less parallel to each other. Nineteen species, 6 in WIO, plus probably 2 undescribed species from Oman.

#### **KEY TO SPECIES**

1a	Dorsal surface with pattern of prominent white spots or brown blotches
1b	Dorsal surface plain or with faint blotches
2a	Dorsal surface with widely scattered white spots R. punctifer
2b	Dorsal surface with symmetrically arranged brown blotches
3a	Skin mostly smooth, with no thorns on dorsal disc, and tail and unpaired fins generally naked, except for small patch of denticles across peduncle and caudal-fin lower lobe
	caudal fin denticle patches

Continued . .

#### **KEY TO SPECIES**

# Rhinobatos austini Ebert & Gon 2017

Austin's guitarfish

PLATE 59

*Rhinobatos austini* Ebert & Gon 2017: 205 (near Port Shepstone, KwaZulu-Natal, South Africa).

Disc broadly wedge-shaped, anterolateral margins almost straight to slightly concave; disc length 42.0–43.8% TL, disc width ~1.3 in disc length. Snout bluntly angular and moderately long, preorbital length 13.6–14.6% TL. Eye length 1.33–1.58 times spiracle length; 2 distinct folds at rear edge of spiracles. Denticles minute (skin smooth to touch), slightly coarser on dorsal midline from nape to first dorsal origin, between dorsal fins and on caudal peduncle.

Dorsal surface light to medium brown, with a pattern of symmetrical dark brown blotches and spots, some aligned on saddle-like darker bands. Ventral surface mostly white, with a prominent tear-shaped blackish blotch on snout tip. Attains at least 115 cm TL.



Rhinobatos austini, 103 cm TL (S Mozambique). O Alvheim © IMR

**DISTRIBUTION** WIO: South Africa (KwaZulu-Natal) to Mozambique.

**REMARKS** This species was previously considered a variant of *Rhinobatos holcorhynchus*. Described from four female specimens only. Biology unknown. Coastal, from shore to about 100 m depth. Likely to be caught in local fisheries.

## Rhinobatos holcorhynchus Norman 1922

Slender guitarfish

PLATE 59

*Rhinobatus holcorhynchus* Norman 1922: 318 (KwaZulu-Natal, South Africa); Wallace 1967\*.

Rhinobatos holcorhynchus: SSF No. 27.4\*; Compagno et al. 1989\*; Heemstra & Heemstra 2004.

Disc rather narrow and moderately long, anterolateral margins almost straight to slightly concave; disc length 41–42% TL, disc width ~1.3 in disc length. Snout triangular, sharply pointed and long, preorbital length 15–16% TL. Eyes about twice size of spiracles; 2 distinct folds at rear edge of spiracles. Denticles minute (skin smooth to touch), but with small tubercles on rostrum, around eyes and spiracles, and 1 or 2 on each shoulder; larger, irregular tubercles in median dorsal row.

Body plain dark olive-brown; margins of pectoral fins and pelvic fins somewhat dusky; snout noticeably translucent on each side of rostral cartilage; white ventrally, but with conspicuous dark blotch on underside of snout. Attains at least 127 cm TL.



Rhinobatos holcorhynchus, 45 cm TL, male (South Africa). Source: SSF

**DISTRIBUTION** WIO: South Africa (KwaZulu-Natal) to Kenya.

**REMARKS** Biology unknown. Rare; occurs offshore, at 75–253 m. Likely taken in local fisheries.

## Rhinobatos lionotus Norman 1926

#### Smoothback guitarfish

*Rhinobatus lionotus* Norman 1926: 961, Fig. 14 (East Channel, mouth of Hooghly River, India).

Rhinobatos lionotus: Compagno & Randall 1987; Talwar & Jhingran 1991.

Disc broad, anterolateral margins slightly concave. Snout bluntly triangular and moderately long, preorbital length 12% TL; 2 strongly developed folds at rear edge of spiracles. Denticles minute (skin smooth to touch), except small tubercles around eyes and spiracles, 1 on each shoulder, and in median dorsal row.

Body plain brownish grey to olive-green dorsally; white ventrally. Attains at least 53 cm TL.



Rhinobatos lionotus (India). Source: Norman 1926

**DISTRIBUTION** Indian Ocean: known only from the holotype, collected from estuary of Hooghly River, West Bengal, but possibly also in the Arabian Sea and Sri Lanka.

**REMARKS** Taken at ~73 m deep. The status of this species is doubtful and should be re-examined. Biology unknown; apparently inhabits brackish water of river mouths.

# Rhinobatos nudidorsalis

Last, Compagno & Nakaya 2004

Bare	back	ŚŚ	hove	eln	ose	ray
------	------	----	------	-----	-----	-----

PLATE 59

*Rhinobatos nudidorsalis* Last, Compagno & Nakaya 2004: 154, Figs. 2–6 (Saya de Malha Bank, Mascarene Ridge).

Disc narrow and moderately long, anterolateral margins almost straight to slightly concave; disc length ~41% TL, disc width ~1.4 in disc length. Snout bluntly triangular and moderately long, preorbital length ~15% TL. Spiracles smaller than eyes; 2 strongly compressed folds at rear edge of spiracles, outer fold
larger. No thorns on dorsal surface of disc; tail and unpaired fins almost entirely naked, except small patch of denticles across peduncle and caudal-fin lower lobe.

Body plain brownish pink dorsally; short dark streak on each side of snout near tip; pinkish white ventrally. Attains 50 cm TL.



*Rhinobatos nudidorsalis*, 50 cm TL, mature male, denticle patches on dorsal body shown by stippled areas (Mascarene Ridge). Drawn from Last *et al.* 2004

**DISTRIBUTION** Known only from the holotype, a mature male, collected from Saya de Malha Bank.

**REMARKS** Taken from relatively deep water (~125 m). IUCN Red List conservation status Near Threatened.

### Rhinobatos punctifer Compagno & Randall 1987

Spotted guitarfish

PLATE 60

Rhinobatos punctifer Compagno & Randall 1987: 336, Fig. 1 (Gulf of Aqaba, Red Sea); Randall 1995\*; Randall & Compagno 1995\*; Bonfil & Abdallah 2004\*.

Disc moderately broad and relatively long, anterolateral margins almost straight to slightly concave; disc length ~44% TL, disc width ~1.1 in disc length. Snout bluntly triangular and moderately long, preorbital length ~13% TL; 2 prominent folds at rear edge of spiracles. No tubercles around eyes and spiracles or on shoulders; small blunt tubercles in median dorsal row.

Body brownish grey dorsally, with conspicuous small white spots (widely scattered or more or less arranged in rows); creamy white ventrally, but with dusky blotch on underside of snout. Attains at least 88 cm TL.



Rhinobatos punctifer, 71 cm TL (Gulf of Aqaba).

**DISTRIBUTION** WIO: Red Sea (including Gulf of Aqaba and Gulf of Suez) and off Oman (Muscat).

**REMARKS** Litters of up to 7; size at birth ~25 cm; matures at ~62 cm TL. Not rare; occurs inshore on continental shelf, at 10–70 m. Feeds mainly on small crustaceans. Regularly taken in local fisheries. Variants of *R. punctifer* are very similar to *R. annandalei* Norman 1926, from India, and are difficult to distinguish from their colour pattern and morphology; however, molecular analysis suggest divergences that need further investigation.

### Rhinobatos schlegelii Müller & Henle 1841

#### Brown guitarfish

*Rhinobatus* (*Rhinobatus*) *schlegelii* Müller & Henle 1841: 123, Pl. 42 (Japan).

Rhinobatos schlegelii: Compagno & Last 1999\*; Bonfil & Abdallah 2004\*.

Disc narrow and moderately long, anterolateral margins almost straight to slightly concave; disc length ~41% TL, disc width ~1.3 in disc length. Snout triangular, broadly pointed, and moderately long, preorbital length ~12% TL; 2 weak folds on rear edge of spiracles. Minute tubercles around eyes and spiracles, and on shoulders; small blunt tubercles in median dorsal row.

Body tan background dorsally, with symmetrical pattern of faint brown botches over entire disc and trunk; white ventrally, but with black longitudinal bar on snout tip. Attains 100 cm TL.

**DISTRIBUTION** Indo-Pacific, though a known speciescomplex; records from Oman and India in WIO most likely represent a species within this complex, but records from South Africa are misidentifications. **REMARKS** Taxonomy uncertain: critical examination of the forms of this species is needed. Occurs inshore on continental shelf, to ~200 m deep, on sand and mud bottom. Feeds mainly on invertebrates (shrimp, cephalopods) and bony fishes.

# FAMILY **GLAUCOSTEGIDAE**

### Giant guitarfishes

Bernard Séret and Marcelo R de Carvalho

Large to very large rays (2–3 m TL) with wedge-shaped to spade-like disc, depressed shark-like trunk, and thick tail. Head flattened and pectoral fins fused with head and trunk; snout relatively elongated and acutely triangular, with tip acutely to bluntly rounded, or knob-like, and supported by stout rostral cartilage. Small spiracles close to small eyes, and with 1 or 2 skin folds along rear edge. Five pairs of gill slits on ventral side. Nostrils long (subequal to or noticeably longer than internasal space), oblique to transverse, without nasal curtain; anterior nasal flaps weakly developed, forming small triangular lobe inserted on front margin of nostril.



Glaucostegus head ventral view.

Two large, nearly equal-sized dorsal fins; 1st dorsal fin well behind pelvic fins. Pelvic fins large, not forming separate lobes, and origins under posterior margins of pectoral fins. Caudal fin strongly asymmetrical with well-developed upper lobe and small or absent lower lobe; no electric organs. Skin covered with fine granulations, often with small thorny tubercles near eyes, on shoulders and snout, and/or blunt tubercles in median dorsal row. Teeth in jaws numerous, small and blunt. Colour plain dorsally with pale translucent snout sharply demarcated by rostral cartilage and anterior skull; usually white ventrally, with snout faintly translucent, sometimes with dark blotch on underside of snout.

Reproductive mode of all species yolk-sac viviparity. Circumglobal and widely distributed, in warm-temperate to tropical waters, mostly inshore over continental shelf, although some occur in deeper water, to ~400 m. Some also enter estuaries and closed bays, and one species has been recorded in freshwater. Mainly benthic, often resting on soft mud or sandy bottom, with shallow sandy habitat important as nursery areas. Feed mainly on benthic invertebrates and small bony fishes. Taken mostly as a bycatch of artisanal and commercial benthic trawl, gillnet, and beach-seine fisheries; appreciated in many countries for their meat, and the large fins have high value in the shark-fin trade. IUCN Red List conservation status of WIO species is Vulnerable.

The subgenus *Glaucostegus* (formerly in *Rhinobatos*) was elevated to generic level by Compagno *et al.* (2005), but is now a recently erected family (Last *et al.* 2016), with one genus and 6 species; at least 5 species in WIO (*Glaucostegus cemiculus* appears unlikely in WIO).

### GENUS Glaucostegus Bonaparte 1846

See family account.

#### **KEY TO SPECIES**

1a 1b	Snout tip expanded into club-like knob <i>G. thouin</i> Snout tip not expanded into club-like knob 2
2a 2b	Snout longer and pointed, preorbital length <5 in TL
3a 3b	Internarial space wide, subequal to nostril length <i>G. granulatus</i> Internarial space narrow, ~½ nostril length
4a	Snout moderately long, preorbital length 5–7 in TL
4b	Snout short, preorbital length >7 in TL
5a	Anterolateral margins of disc almost straight; 2 folds on rear edge of spiracles; distinct median dorsal row of thorns
5b	Anterolateral margins of disc convex; 1 weak fold on rear edge of spiracles; no distinct median dorsal row of thorns

### Glaucostegus cemiculus (Geoffroy Saint-Hilaire 1817)

Blackchin guitarfish

PLATE 60

Rhinobatus cemiculus Geoffroy Saint-Hilaire 1817: Pl. 27,

Fig. 3 (Lake Menzilah, Alexandria, Egypt, Mediterranean Sea). Rhinobatos cemiculus: Whitehead et al. 1984\*. Glaucostegus cemiculus: Compagno et al. 2005.

Disc moderately broad, anterolateral margins almost straight and slightly undulating; disc length ~39% TL, disc width ~1.1 in disc length. Snout triangular and moderately long, preorbital length ~14% TL; 2 folds at rear edge of spiracles, outer fold more prominent. Small orbital and interspiracular thorns, 1–3 scapular, and in median dorsal row.

Body plain beige to pale brown dorsally; outer margins of disc and fins sometimes reddish brown; white ventrally, but with conspicuous black blotch on underside of snout tip (mainly in juveniles). Attains 265 cm TL.



Glaucostegus cemiculus, 2.1 m TL (Ghana). O Alvheim © IMR

**DISTRIBUTION** Recorded from Madagascar as *Rhinobatos petiti*; elsewhere known from Mediterranean Sea and eastern Atlantic.

**REMARKS** *Glaucostegus cemiculus* probably does not occur in WIO. However, the nominal species *Rhinobatos petiti* Chabanaud 1929 (known only from holotype) from Madagascar was synonymised with it in Séret & McEachran (1986): pending the collection of further specimens, the synonymy with *G. cemiculus* is maintained. Litters of up to 20 (usually 4–6); size at birth ~34 cm TL. Males mature at 138–154 cm TL, and females at 153–164 cm TL. Coastal, mainly in shallow water but to ~100 m deep, and also enters estuaries and coastal lagoons; rests partly buried on sandy or muddy bottom. An active hunter, feeds on small benthic invertebrates and small fishes. Targeted for its fins in artisanal fisheries in West Africa. IUCN Red List conservation status Endangered.

### Glaucostegus granulatus (Cuvier 1829)

Sharpnose guitarfish

PLATE 60

Rhinobatus granulatus Cuvier 1829: 396 (Puducherry, India). Rhinobatos granulatus: Randall 1995\*; Randall & Compagno 1995\*; Compagno & Last 1999\*; Bonfil & Abdallah 2004\*. Glaucostegus granulatus: Compagno et al. 2005.

Disc relatively narrow and long; anterolateral margins concave at level of snout tip, then almost straight; disc length almost half of TL, disc width ~1.4 in disc length. Snout triangular, acutely pointed and long, preorbital length ~22% TL. Eyes smaller than spiracles; 2 weakly developed folds at rear edge of spiracles, inner fold rudimentary. Conspicuous tubercles on rostral ridges, around eyes and spiracles, and on shoulders; strong and compressed tubercles in median dorsal row.

Body greyish brown dorsally, and margins usually reddish; each side of snout noticeably translucent; white ventrally. Attains 280 cm TL.



Glaucostegus granulatus, 39 cm TL (Persian/Arabian Gulf).

**DISTRIBUTION** Indo-Pacific (range poorly defined). WIO: Persian/Arabian Gulf to India and Sri Lanka; records from Red Sea and Oman need confirmation; elsewhere possibly to Indonesia, Philippines, China, New Guinea and Australia.

**REMARKS** Often misidentified with *G. typus*, which is common in the Indo-Pacific. Coastal, inshore and offshore, to ~119 m deep, but mainly in shallow water and also enters estuaries and coastal lagoons. Biology little known, other than litters of 6–10. Directly and indirectly fished throughout its range; once moderately abundant, now irregularly caught.

### Glaucostegus halavi (Fabricius 1775)

#### Halavi guitarfish

PLATE 60

*Raja halavi* Fabricius *in* Niebuhr (ex Forsskål) 1775: 19, viii (Jeddah, Saudi Arabia, Red Sea).

*Rhinobatos halavi:* Randall 1995\*; Randall & Compagno 1995\*; Compagno & Last 1999\*; Bonfil & Abdallah 2004\*.

Glaucostegus halavi: Compagno et al. 2005.

Disc narrow and moderately long, anterolateral margins slightly convex to almost straight; disc length ~40% TL, disc width ~1.4 in disc length. Snout bluntly triangular and somewhat short, preorbital length ~13% TL; 2 folds at rear edge of spiracles, inner fold rudimentary. Small tubercles on rostral ridges, and around eyes and spiracles; large blunt tubercles in median dorsal row.

Body plain tan dorsally, and margins slightly paler; large translucent area on each side of snout; white ventrally. Attains 171 cm TL.



Glaucostegus halavi, ~1 m TL, mature male (Red Sea).

**DISTRIBUTION** WIO: Red Sea, Gulf of Oman, and probably Gulf of Aden and other areas of northeastern Arabian Sea; reports from Mediterranean Sea and elsewhere in Indo-Pacific need confirmation.

**REMARKS** Litters of up to 10; size at birth ~29 cm TL; males mature at ~83 cm TL. Coastal, in shallow inshore waters and sheltered bays, to ~40 m deep; young observed in very shallow water washing onto beach with breakers and then sliding back into water. Aggregates during spawning. Feeds mostly on small crustaceans. A target species and utilised bycatch mainly in artisanal fisheries.

### Glaucostegus obtusus (Müller & Henle 1841)

#### Widenose guitarfish

Rhinobatus (Rhinobatus) obtusus Müller & Henle 1841: 122, Pl. 37 (Puducherry, India). Rhinobatos obtusus: Compagno & Last 1999\*; Sujatha 2002\*;

Manilo & Bogorodsky 2003.

Glaucostegus obtusus: Compagno et al. 2005.

Disc moderately broad and moderately long, anterolateral margins slightly convex; disc length ~41% TL, disc width ~1.1 in disc length. Snout triangular, obtusely angular and short, preorbital length ~14% TL; 1 weak fold at rear edge of spiracles. Small tubercles on rostral ridges, around eyes, on shoulders, and in median dorsal row; tubercles stronger in young and reduced or absent in adults (often limited to series of few small ones on midback).

Body plain brown to greenish brown dorsally, and margins reddish brown; white ventrally. Attains 93 cm TL.

**DISTRIBUTION** Indo-Pacific. WIO: Pakistan to India and Sri Lanka; reports from South Africa (KwaZulu-Natal) are probably misidentifications; elsewhere to Indonesia.

**REMARKS** Biology unknown, other than litters of 2–16. Coastal, inshore and offshore over continental shelf. Previously moderately abundant, now irregularly taken in local fisheries.

### Glaucostegus thouin (Anonymous [Lacepède] 1798)

Clubnose guitarfish

PLATE 60

PLATE 60

*Raja thouin* Anonymous [Lacepède] 1798: 287, 677, 685, Pl. 1, Figs. 2–4 [probably Indonesia].

Rhinobatos thouin: Compagno & Last 1999\*; Bonfil & Abdallah 2004\*; Yano et al. 2005\*; White et al. 2006\*.

Glaucostegus thouin: Compagno et al. 2005; Last et al. 2010\*.

Disc rather narrow and moderately long, anterolateral margins almost straight; disc length ~42% TL, disc width ~1.3 in disc length. Snout acutely triangular and long, tip distinctly

extended as club-like knob; preorbital length (including knob) ~18% TL; 2 weak folds at rear edge of spiracles. Small tubercles around eyes and spiracles, 2 on each shoulder, and larger irregular tubercles in median dorsal row.

Body plain greyish brown to olive-brown dorsally, and pectoral- and pelvic-fin margins paler; entire snout translucent on each side of rostral cartilage, but snout tip sometimes greyish; white ventrally. Attains 3 m TL.



Glaucostegus thouin (SW India). KK Bineesh © CMFRI

**DISTRIBUTION** Indo-Pacific. WIO: Red Sea and southwestern India; elsewhere, Thailand, Vietnam, Indonesia and New Guinea.

**REMARKS** Biology unknown. Coastal, inshore, mainly in shallow water <60 m deep; juveniles inhabit sand flats and mangrove estuaries. Feeds primarily on shellfishes. Previously moderately abundant, now irregularly taken in artisanal and commercial fisheries.

### Glaucostegus typus (Anonymous [Bennett] 1830)

Giant shovelnose ray

PLATE 61

*Rhinobatus typus* Anonymous [Bennett] 1830: 694 (Sumatra, Indonesia; India).

Rhinobatos typus: Compagno & Last 1999\*; Bonfil & Abdallah 2004\*; White *et al.* 2006\*.

*Glaucostegus typus*: Compagno *et al.* 2005; Last & Stevens 2009\*; Last *et al.* 2010\*; Kottelat 2013. Disc rather narrow and moderately long, anterolateral margins almost straight to slightly concave; disc length ~41% TL, disc width ~1.3 in disc length. Snout bluntly triangular and moderately long, preorbital length ~17% TL; 2 weak folds at rear edge of spiracles. Small tubercles around eyes and spiracles, and larger irregular tubercles in median dorsal row.

Body plain greyish brown to olive-brown dorsally, and pectoral- and pelvic-fin margins paler; snout tip sometimes darker grey; white ventrally. Attains at least 270 cm TL, probably more.

**DISTRIBUTION** Indo-Pacific (widespread). WIO: southern India and Sri Lanka; elsewhere to Thailand, Indonesia, Philippines and northern Australia.

**REMARKS** Size at birth 30–40 cm TL; matures at 150–180 cm TL. Juveniles often found in mangroves, estuaries and coastal lagoons; adults occur inshore, over continental shelf, to ~100 m deep, and also the only rhinobatid reportedly able to live and breed in freshwater. Feeds primarily on crustaceans (crab, prawns) and also small bony fishes. Previously moderately abundant, now irregularly taken in local fisheries.

# ORDER RAJIFORMES

# FAMILY RAJIDAE

### Hardnose skates

Neil C Aschliman and David A Ebert

Small- to large-sized skates (up to ~250 cm TL). Head and body depressed, with wing-like pectoral fins forming flattened rhomboidal disc; snout pointed, short and rounded to long and angular. Five pairs of gill slits on underside of head. Mouth small, with numerous small teeth in powerful jaws. Pelvic fins deeply notched, divided into anterior and posterior lobes. Tail rather slender but not whip-like, with usually 2 small dorsal fins near tail tip (rarely 1 or none), and reduced caudal fin; most species with one or more rows of enlarged denticles or thorns from mid-dorsal disc to tail, but no stinging spine. Trunk vertebrae 22–44, predorsal caudal vertebrae 29–81.

Speciose and morphologically diverse but poorly defined with no unambiguous characters uniting current members of the family. However, the snout is typically rigid (unlike softnose skates, family Arhynchobatidae) and supported by stiff cartilage, and in several genera the rostral bar is attenuated and extended. The distribution and size of spinules, enlarged denticles or thorns on the dorsal surface of the disc and tail, are important characters for distinguishing species (see thorn patterning diagram in the chapter on **Anatomy of sharks**, **rays and chimaeras**). While external morphology is difficult to use in distinguishing hardnose skates from softnose skates, anatomical characters defining (but not exclusive to) Rajidae include: scapulocoracoid lacking anterior bridge, clasper expanded distally (3–9 terminal cartilages), and clasper glans with component rhipidion.

Reproductive mode oviparity; females may deposit an estimated 50–300 or more dark brown, keratinous egg cases annually. Adult males typically have small hooked malar spines or thorns, used to clasp females. Found in estuaries and inshore habitats on continental shelf as well as on continental slope to >4 000 m deep. Mostly feed on bottom-living invertebrates and fishes. Skates have been the subject of target and non-targeted fisheries worldwide; mostly taken as bycatch in WIO. Some species are of commercial interest as skate 'wings' are considered good eating.

Worldwide in all seas. Currently the largest family of batoids, with 16 genera and at least 158 valid species; 8 genera and at least 23 species in WIO, of which 5 are from deep water.

#### **KEY TO GENERA**

1a 1b	Tip of snout firm and not flexible; anteriormost extension of pectoral fins distinctly separated from snout by semi- translucent area
2a	Snout moderately to distinctly elongated; anterolateral margins of disc concave (except in <i>Okamejei heemstrai</i> , but colour pattern distinguishes this species from <i>Dipturus</i> and <i>Rostroraja</i> ); internarial width usually <70% of distance from tip of snout to front edge of nostrils
2b	Snout generally not elongated; anterolateral margins of disc straight to moderately convex; internarial width usually >70% of distance from tip of snout to front edge of nostrils
3a 3b	Pores on ventral surface of disc darkly pigmented <i>Dipturus</i> Pores on ventral surface of disc not darkly pigmented
4a 4b	Triangular thorn patch over nuchal and scapular areas
5a	Thorns similarly sized in 1 to several rows along mid-dorsal disc and tail in juveniles and adults
5b	Thorns in several rows along mid-dorsal disc and tail, and median row either smaller than lateral row(s) or absent in large juveniles and adults
6a 6b	Pores on ventral surface of disc darkly pigmented <i>Okamejei</i> Pores on ventral surface of disc not darkly pigmented
7a	Snout very short and broad; tooth rows in upper jaw <61
7b	Snout moderately elongate and angular; tooth rows

### GENUS Dipturus Linnaeus 1758

This genus is in need of revision as most characters currently used to distinguish *Dipturus* from *Okamejei* are not useful. Disc quadrangular, moderately broad to broad; pectoral-fin apex angular to slightly rounded. Snout acutely triangular and long to very long (generally >60% HL). Eyes relatively small, eye diameter less than interorbital space. Pelvic-fin anterior lobes moderately long to long and approaching length of posterior lobes. Tail relatively short to moderately long, moderately stout and often slightly broader at midlength. Disc with patterns of few small to moderately small thorns, otherwise smooth.

#### **KEY TO SPECIES**

1a 1b	Tail relatively long and slender, width at 1st dorsal-fin origin<½ width at baseTail not long and slender, width at 1st dorsal-fin origin½ width at base3
2a	Pelvic-fin anterior lobes not extending to rear edge of posterior lobes; tail length from 2nd dorsal-fin insertion to tail tip <5% TL <i>D. lanceorostrata</i> [deep water, from 430–439 m]
2b	Pelvic-fin anterior lobes extending to or beyond rear edge of posterior lobes; tail length from 2nd dorsal-fin insertion to tail tip >5% TL
3a	Ventral surface of disc broadly covered with denticles
	D. springeri
3b	Ventral surface of disc not broadly covered with denticles, although enlarged denticles may occur on snout and along anterior margins

### Dipturus campbelli (Wallace 1967)

#### Blackspot skate

PLATE 61

Raja campbelli Wallace 1967: 24, Fig. 12 (KwaZulu-Natal, South Africa). Raia pullopunctata Smith 1964: 285, Pl. 25a [in part].

*Raja pullopunctata*: SSF No. 25.15\*.

Raja (Dipturus) campbelli: Séret 1989.

Dipturus campbelli: McEachran & Dunn 1998; Compagno 1999, 2005;

Heemstra & Heemstra 2004; Compagno & Ebert 2007\*; Ebert *et al.* 2008; Last 2008.

Thorns (when present): 1 preorbital and ~9 orbital, 1 to several scapular and/or nuchal, and 1 median row in males and 3–5 median and lateral rows in females. Trunk vertebrae 30–35, predorsal caudal vertebrae 42–72. Egg cases >6.5 cm long excluding horns. Disc generally brown to almost black dorsally, and plain or vaguely patterned; pale grey to greyish brown ventrally, with darkly pigmented pores. Size up to ~200 cm TL. Worldwide, except for northeastern and tropical western Pacific, and most diverse in southeastern Atlantic (off southern Africa) and in South Pacific (off Australia). Depth range mostly ~40–1 000 m. About 50 species, 8 in WIO (2 of the species are from deep water and have no species accounts).

Disc rather narrow, DW <60% TL; pectoral-fin apex weakly angular. Snout acutely triangular and moderately extended; length 17–20% TL. Teeth in upper jaw 45. Pelvic-fin anterior lobes moderately long, not reaching rear edge of posterior lobes. Tail relatively short, slightly shorter than distance from snout tip to rear edge of cloaca, and moderately wide over its length or slightly broader at midlength than at its base. Ventral surface with enlarged denticles on anterior margins of disc and snout tip. Dorsal surface with ~7 orbital thorns, 1 or 2 nuchal, in median row on mid-dorsal disc to 1st dorsal fin, 1 interdorsal, and short lateral row on tail. Disc brown to medium grey dorsally, with scattered small dark spots; grey ventrally, with conspicuous black pores. Attains 66 cm TL.



*Dipturus campbelli*, 66 cm TL, female holotype (South Africa). Source: Wallace 1967

**DISTRIBUTION** WIO: possibly endemic to South Africa (KwaZulu-Natal) and southern Mozambique.

**REMARKS** Size at hatching <18 cm TL. Males mature by 56 cm TL, and females by 63 cm TL. Poorly known with patchy, limited distribution; found over outer continental shelf to upper slope, at 137–403 m. Probably caught incidentally by trawlers. IUCN Red List conservation status Near Threatened.

### Dipturus doutrei (Cadenat 1960)

Javelin skate

PLATE 61

*Raja doutrei* Cadenat 1960: 294, Figs. 1–11, 13, 15 (Senegal); Hulley 1970\*. *Raja (Dipturus) doutrei*: Hulley 1972\*; Lloris 1982; SSF No. 25.11\*;

Séret 1989; Stehmann 1990, 1995; Compagno *et al.* 1991; Bianchi *et al.* 1993.

Dipturus doutrei: McEachran & Dunn 1998; Compagno 1999, 2005; Compagno & Ebert 2007\*; Ebert *et al.* 2008; Last 2008.

Disc angular and broad, DW 72–75% TL; pectoral-fin apex narrowly rounded. Snout distinctly extended, acutely triangular and long, ~20% TL. Teeth in upper jaw 27–35. Pelvic-fin anterior lobes moderately long, not extending to rear edge of posterior lobes. Tail relatively short, shorter than distance from snout tip to rear edge of cloaca, and relatively broad at base and slightly broader at midlength. Ventral surface with enlarged denticles along anterior margins of disc and on snout. Dorsal surface with 6 or 7 small orbital thorns, 13–26 in median row on tail to 1st dorsal fin, and 1 or 2 interdorsal; females with lateral row of thorns on tail. Predorsal caudal vertebrae 43–49.

Disc dark brown dorsally, with scattered irregular darker blotches, and blackish towards tail tip; brown ventrally, with black pores and scattered paler patches. Attains ~115 cm TL; males mature at ~101 cm TL (Last *et al.* 2016).



Dipturus doutrei, 62 cm DW (Namibia). Source: SSF

**DISTRIBUTION** Eastern Atlantic (Mauritania to South Africa), and just rounding Cape of Good Hope (South Africa) into WIO.

**REMARKS** Little known; found over outer continental shelf to deep slope, at 163–1 200 m (usually 450–600 m). Some records may be misidentifications of *D. springeri*. A bycatch of deepwater trawl fisheries for bony fishes. Diet includes benthic bony fishes, shrimp and crabs.

### Dipturus johannisdavisi (Alcock 1899)

Travancore skate

PLATE 62

*Raja johannisdavisi* Alcock 1899: 21 (Laccadive Sea, off India); Alcock 1900 [Fig.].

Raja (Dipturus) johannisdavisi: Stehmann 1976; Séret 1986, 1989.

Dipturus johannisdavisi: McEachran & Dunn 1998; Compagno 1999, 2005; Manilo & Bogorodsky 2003; Last 2008.

Small-sized with moderately broad disc, DW 55–66% TL; pectoral-fin apex slightly angular. Snout distinctly extended and moderately long, length 18–19% TL. Teeth in upper jaw 36–42. Pelvic-fin anterior lobes long and extending beyond rear edges of posterior lobes. Tail relatively long, equal to or greater than distance from snout tip to rear edge of cloaca, and narrow at base and slightly attenuated distally. Ventral surface with enlarged denticles along anterior margins of disc and on snout. Dorsal surface with 2 or 3 preorbital and 1 or 2 postorbital thorns, in median row on tail, and 1 interdorsal.

Disc uniformly dark brownish black dorsally and ventrally. Attains at least 54 cm TL (Last *et al.* 2016).



Dipturus johannisdavisi (India). Source: Alcock 1900

**DISTRIBUTION** WIO: Gulf of Aden (Yemen), Lakshadweep and India (Travancore coast); a record from Tanzania (Zanzibar) is questionable.

**REMARKS** Found over continental slope; known from 220–660 m (Last *et al.* 2016).

### Dipturus pullopunctatus (Smith 1964)

Slime skate

PLATE 62

*Raia pullopunctata* Smith 1964: 285, Pl. 25a (Algoa Bay, Eastern Cape, South Africa).

Raja pullopunctata: Hulley 1966\*, 1970\*; Wallace 1967\*.

Raja (Dipturus) pullopunctata: Hulley 1972\*; SSF No. 25.15\*; Séret 1989; Compagno et al. 1991; Ebert et al. 1991; Bianchi et al. 1993;

Stehmann 1995; Walmsley-Hart *et al.* 1999. Dipturus pullopunctatus: McEachran & Dunn 1998; Compagno 1999, 2005.

Dipturus pullopunctata: Heemstra & Heemstra 2004; Compagno &

Ebert 2007\*; Ebert et al. 2008\*; Last 2008.

Disc angular, DW 1.2–1.4 times disc length and 76–78% TL; pectoral-fin apex broadly rounded. Snout moderately extended, acutely triangular and long, length 15–17% TL and 3.4–4 times interorbital space; angle anterior to spiracles 92°–108°. Jaws moderately large and strong. Teeth in upper jaw 53–58; teeth sharp and pointed in males, rounded in females. Pelvic-fin anterior lobes moderately long, not extending to

rear edge of posterior lobes. Tail moderately long, about equal to distance from snout tip to rear edge of cloaca, and narrow at base and slightly attenuated distally. Thorns in juveniles: 2 preorbital and 1 post-orbital, 1 enormous nuchal, 8–12 in median row on mid-dorsal disc to 1st dorsal fin, and 1 interdorsal. Thorns in adults: 3–8 small orbital, 1 or 2 typically extremely large nuchal, 8–12 in median row on mid-dorsal disc to 1st dorsal fin, and 1 or 2 interdorsal; ventral surface with spinules on snout tip, internasal area and anterior margins of disc. Egg cases very large, ~13 cm long excluding horns; surface striated and covered with dense fibres; broad lateral keels; horns tapering, with anterior horns hook-like and slightly longer than inward-curving posterior horns. Predorsal caudal vertebrae 50–58.

Disc tan or pale brown dorsally, with widely spaced small dark spots (especially in juveniles, maybe lost in adults), and sometimes surrounded by paler ring or small white spots (especially in adults); usually with very large dark blotch at each pectoral-fin base; greyish ventrally, with black pores. Attains 125 cm TL (females reach larger sizes than males).



*Dipturus pullopunctatus*, 14 cm DW, juvenile holotype (top); 52 cm DW, mature male (bottom; both South Africa). Source: SSF

**DISTRIBUTION** Southern Africa: Namibia (Lüderitz) in southeastern Atlantic, to Eastern Cape, Agulhas Bank (off South Africa) and possibly to Mozambique (Bazaruto) in WIO.

**REMARKS** Size at hatching <19 cm TL. Males mature at 88–96 cm TL; females' maturation and maximum size vary widely between populations: off the west coast of South Africa, females mature at 100–111 cm TL and attain 125 cm, and off the south coast, females mature at 78–82 cm TL and attain only 103 cm. Occurs over shallow continental shelf to upper slope, at 15–457 m (usually 100–300 m). Becomes extremely slippery when handled. Feeds mostly on benthic fishes, crabs, mysids, mantis shrimp, krill, bivalves and cuttlefish. Commonly taken in hake-directed longline and trawl fisheries but not in large quantities.

### Dipturus springeri (Wallace 1967)

#### Roughbelly skate

PLATES 62 & 63

Raja springeri Wallace 1967: 18, Figs. 9–10 (KwaZulu-Natal, South Africa). Raja (Dipturus) springeri: Hulley 1972\*; SSF No. 25.20\*; Séret 1989;

Compagno *et al.* 1991; Ebert *et al.* 1991; Bianchi *et al.* 1993; Stehmann 1995.

*Dipturus springeri*: McEachran & Dunn 1998; Compagno 1999, 2005; Compagno & Ebert 2007\*; Ebert *et al.* 2008; Last 2008.

Large-sized with very broad disc, DW ~1.4 times disc length and 80–85% TL; pectoral-fin apex acutely angled. Snout triangular and long, length 19–23% TL and 4.8–5.1 times interorbital space; angle anterior to spiracles ~85°. Teeth sharp, cuspidate; 35–40 in upper jaw. Pelvic-fin anterior lobes long, extending to or beyond rear edge of posterior lobes. Tail relatively short, shorter than distance from snout tip to rear edge of cloaca, and narrow at base and not attenuated distally or else slightly broader at midlength. Ventral surface densely covered with small denticles. Dorsal surface with 2–7 small orbital thorns, no nuchal thorns, ~15 in median row on tail to 1st dorsal fin, and 2 interdorsal. Predorsal caudal vertebrae 53–56.

Disc dark grey dorsally and ventrally; pores on ventral surface black. Attains 190 cm TL (females reach larger sizes than males).



Dipturus springeri, 27 cm DW, 34 cm TL (Kenya). Source: SSF

**DISTRIBUTION** Southern Africa: Namibia (Lüderitz) in southeastern Atlantic, to South Africa, Mozambique, Kenya and off Madagascar in WIO.

**REMARKS** Size at hatching <27 cm TL; slow-growing. Males mature at 125–134 cm TL, and females at 171–190 cm TL. Uncommon, in scattered localities; occurs on outer continental shelf to slope, at 50–970 m. Diet includes fishes (rattails, round herring), crustaceans and squid.

### Dipturus stenorhynchus (Wallace 1967)

Prownose skate

PLATE 63

*Raja stenorhynchus* Wallace 1967: 23, Fig. 11 (east of False Bay, South Africa).

Raja (Dipturus) stenorhynchus: Hulley 1972\*; SSF No. 25.21\*; Séret 1989. Dipturus stenorhynchus: McEachran & Dunn 1998; Compagno 1999, 2005; Compagno & Ebert 2007\*; Ebert *et al.* 2008; Last 2008.

Disc broad, DW ~1.1 times disc length and 60–66% TL; pectoral-fin apex angular. Snout acutely triangular and greatly elongated, length 21–24% TL and ~5.5 times interorbital width; angle anterior to spiracles ~64°. Teeth in upper jaw 38. Pelvic-fin anterior lobes moderately long, almost reaching rear edge of posterior lobes. Tail moderately short, shorter than distance from snout tip to rear edge of cloaca, and moderately narrow at base, often slightly broader at midlength, and triangular in cross-section. Ventral surface with enlarged denticles along anterior margins of disc and near front of snout. Dorsal surface with cluster of small denticles on snout tip, 9–11 small orbital thorns, 1 nuchal thorn, ~35 thorns in median row on tail, 4 or 5 laterally thorns near base of tail, and 1 interdorsal thorn. Predorsal caudal vertebrae 49.

Disc dark grey dorsally and ventrally; pores on ventral surface black. Attains 60+ cm DW, 105 cm TL.



*Dipturus stenorhynchus*, 60 cm DW, female holotype (S Mozambique). Source: SSF

**DISTRIBUTION** WIO: South Africa (False Bay to KwaZulu-Natal) and central Mozambique.

**REMARKS** Poorly known from few specimens. Size at hatching <24 cm TL. Males mature at 83–98 cm TL, and females at >50 cm TL. Occurs over edge of continental shelf and slope, at 253–761 m; may be more widespread in deeper water.

### GENUS *Leucoraja* Malm 1877

Disc heart-shaped and moderately broad to broad, DW usually >52% TL; pectoral-fin apex narrowly to broadly rounded. Snout moderately short and broad, with tip slightly produced as small oblique process. Eyes moderately sized, eye diameter slightly less to slightly more than interorbital space. Pelvic-fin anterior lobes usually much shorter than (rarely about as long as) posterior lobes. Tail relatively short, length generally <60% TL, and relatively broad at base and attenuated distally. Dorsal fins rather large, and not separated at bases or separated by distance <½ length of 1st dorsal-fin base. Dorsal surface sparsely to densely covered with coarse denticles, generally denser in juveniles than in adults; moderate-sized thorns on snout tip, along orbital rim, medial to spiracles, in triangular patch on

nuchal and scapular area, and along mid-dorsal disc to tail in several fairly regular rows, with median row reduced or absent in large juveniles and adults. Body tan to dark grey or brown dorsally, and either plain or patterned with spots, bars or ocelli; pale ventrally, and plain or with dark blotches. Predorsal caudal vertebrae 64–81. Size up to ~150 cm TL. Primarily limited to North Atlantic, tropical and eastern South Atlantic and Indian oceans. Found on continental shelf and upper slope, from shoreline to ~600 m deep. Thirteen species, 2 in WIO, and 1 undescribed species known from Madagascar Ridge in 750–1 050 m (S Weigmann, pers. comm.).

#### **KEY TO SPECIES**

- 1b Pelvic-fin anterior lobes much shorter than posterior lobes; dorsal fins separated by small interspace; teeth in upper jaw 59–69

### Leucoraja compagnoi (Stehmann 1995)

Tigertail skate

PLATE 63

*Raja (Leucoraja) compagnoi* Stehmann 1995: 43, Fig. 10 (Strandfontein, Western Cape, South Africa).

*Leucoraja compagnoi*: McEachran & Dunn 1998; Compagno 1999, 2005; Compagno & Ebert 2007\*; Ebert *et al.* 2008; Last *et al.* 2008.

Disc moderately narrow, DW ~56% TL; pectoral-fin apex broadly rounded. Snout obtuse and moderately long, length ~11% TL. Teeth in upper jaw 38. Pelvic-fin anterior lobes long, approximately reaching rear edge of posterior lobes. Tail moderately long, length ~57% TL, and moderately broad at base and attenuated distally. No interdorsal space. Thorns numerous and small along orbital rim, in triangular patch over nuchal and scapular area, and in several rows from shoulder to 1st dorsal fin, with thorns in parallel rows larger than those along midline.

Disc tan to medium brown dorsally; white ventrally. Maximum size unknown.

**DISTRIBUTION** South Africa: Western Cape in southeastern Atlantic, to Eastern Cape and KwaZulu-Natal in WIO.

**REMARKS** Likely confused with the abundant *L. wallacei*, and thus overlooked by fisheries biologists; known only from a few juvenile and subadult specimens. Size at hatching <14 cm TL. Size at maturity unknown, but the largest recorded

male was an adolescent measuring ~52 cm TL, and the largest female was immature at ~29 cm TL. Found on upper continental slope, at 383–1 300 m (Weigmann 2016); adults possibly occur deeper.

### Leucoraja wallacei (Hulley 1970)

Yellowspotted skate

PLATE 64

*Raja wallacei* Hulley 1970: 210, Pl. 12, Fig. 19 (off Cape Town, South Africa).

Raja (Leucoraja) wallacei: Hulley 1972\*; SSF No. 25.23\*; Compagno et al. 1991; Ebert et al. 1991.

Leucoraja wallacei: McEachran & Dunn 1998; Compagno 1999, 2005; Compagno & Ebert 2007\*; Ebert *et al.* 2008\*; Last *et al.* 2008.

Disc moderately broad and slightly wider than long, DW 56-62% TL; pectoral-fin apex broadly rounded. Snout short, broad and blunt, with rounded terminal process, length 9.6-11% TL and 2.3-3.1 times interorbital space; angle anterior to spiracles ~110°. Jaws fairly strong; teeth in upper jaw 59-67, cusps sharper in males than in females. Pelvic-fin anterior lobes much shorter than posterior lobes. Tail relatively long, length 53-55% TL, and moderately broad at base and attenuated distally. Interdorsal space small; 1st dorsal fin slightly larger than 2nd. Ventral surface with spinules on snout tip and anterior margins of disc. Dorsal surface with spines on snout tip, anterior margins of disc and tail; 7-10 small orbital thorns, 2 or 3 supraspiracular, on nuchal and scapular areas, ~34 large thorns in median row on disc and tail, plus lateral rows on tail; no interdorsal thorns; mid-dorsal thorns smaller, partially absent or totally absent in large juveniles and adults. Egg cases medium-sized, ~8 cm long excluding horns; surface smooth, with fine striations and no fibres; no lateral keels; posterior apron ~50% wider than anterior apron; horns taper to filamentous tips, and posterior horns about twice as long as anterior horns. Predorsal caudal vertebrae 70.

Disc yellowish brown dorsally, scattered with bright yellow spots mostly in rosettes or whorls (some specimens greyish brown with whitish spots); whitish ventrally. Attains 100 cm TL (females reach larger sizes than males).



Leucoraja wallacei, 51 cm TL (South Africa). O Alvheim © IMR



Leucoraja wallacei, 52 cm DW, 84 cm TL, male (South Africa). Source: SSF

**DISTRIBUTION** Southern Africa: Namibia (Lüderitz) in southeastern Atlantic, to Agulhas Bank (South Africa) and Mozambique in WIO.

**REMARKS** Size at hatching <16 cm TL. Males mature at 64–78 cm TL, and females at 65–81 cm TL, both sexes at ~9 years. Common; found on continental shelf to upper slope, at 70–517 m (usually 150–300 m). Feeds mostly on prawns, but also bony fishes, including dragonets (Callionymidae) and ophichthid eels. Caught incidentally in hake trawl fisheries and discarded. IUCN Red List conservation status Vulnerable.

### GENUS Neoraja McEachran & Compagno 1982

Disc heart-shaped and relatively narrow, DW usually 50-60% TL; pectoral-fin apex broadly rounded; anterior margins of disc strongly concave to straight. Snout moderately short, length usually 8-10% TL; broadly rounded or angular, and tip slightly produced as small triangular process; anteriormost extension of pectoral fins (nearly abutting snout tip) not separated from snout by semi-translucent area. Eves relatively large, eve diameter equal to or greater than interorbital space. Pelvic-fin anterior lobes relatively long, but not reaching rear edge of posterior lobes. Tail moderately long, length 55-60% TL, and moderately slender at base and tapering distally. Dorsal surface densely covered with fine denticles; small thorns on orbital margin, on nuchal and scapular areas (but not forming triangular patch), and on mid-dorsal disc, becoming median row and 1-3 irregular lateral rows on tail. Predorsal caudal vertebrae 65-74. Brown to greyish brown dorsally; pale with wide margin of brown to greyish brown ventrally. Dwarf skates, adults attain up to ~35 cm TL. Found on outer continental shelf and slope, at 295-1 640 m. Occurs in western central and eastern Atlantic Ocean, extending to South Africa (Eastern Cape) in WIO. Five species, 1 in WIO.

### Neoraja stehmanni (Hulley 1972)

South African pygmy skate

*Breviraja stehmanni* Hulley 1972: 254, Figs. 1–5 (Cape Basin, Atlantic Ocean).

Neoraja stehmanni: McEachran & Stehmann 1984; SSF No. 25.5\*; McEachran & Dunn 1998; Compagno 1999, 2005; Compagno & Ebert 2007\*; Ebert *et al.* 2008\*.

PLATES 64 & 65

Very small-sized, DW 54–58% TL; pectoral-fin apex broadly rounded. Snout obtuse and moderately short, length 8.9–11.5% TL and 2.9–3.8 times interorbital space; angle anterior to spiracles 115°–130°. Eyes large and close-set, eye diameter slightly greater than interorbital space. Pelvicfin anterior lobes relatively long, nearly reaching rear edge of posterior lobes. Tail relatively long, length 58–60% TL, and moderately slender at base and attenuated distally. No interdorsal space; dorsal fins similar in size and shape. Jaws small; teeth in upper jaw 38–44, teeth blunt and flat in females and juveniles, and with single long, acute cusp in males. Ventral surface smooth. Dorsal surface of disc covered with denticles, also 2–10 large orbital thorns, 1–4 nuchal, and 11–38 in interrupted median row from shoulder to 1st dorsal fin. Egg cases very small; surface smooth and lacking fibres; broad lateral keels; posterior horns taper to acute inwardly pointing tips. Predorsal caudal vertebrae 65–74.

Disc greyish brown dorsally; pale ventrally, but darker on front and rear margins, on snout tip and underside of tail; 6 or 7 dark crossbars on tail. Attains 21 cm DW, 37 cm TL.

**DISTRIBUTION** South Africa: from Orange River mouth in southeastern Atlantic to Agulhas Bank in WIO.

**REMARKS** Size at hatching <15 cm TL. Matures at ~30 cm TL. Occurs on outer continental shelf, on upper to deep slope, at 102–1 025 m (usually >600 m). Its known distribution is unusually localised compared with other offshore southern African skates, but this may change with further deep-slope exploration. Diet includes shrimp. Probably taken incidentally in hake bottom-trawl fisheries.



Neoraja stehmanni, 20 cm DW, 33 cm TL (South Africa). Source: SSF

### GENUS Okamejei Ishiyama 1958

This genus is in need of revision as most characters currently used to distinguish Okamejei from Dipturus are not useful. Disc quadrangular and moderately broad, DW usually 55–65% TL; pectoral-fin apex weakly angular. Snout acutely angled, slightly to moderately extended and moderately long, length usually 13-17% TL. Eyes moderately small, eye diameter roughly equivalent to interorbital space. Pelvic-fin anterior lobes much shorter than posterior lobes. Tail moderately long, length 44-56% TL, and moderately narrow at base and slightly tapering distally. Dorsal surface largely free of denticles; several moderate-sized thorns on orbital margin, 1 to several nuchal thorns, rarely in median row between nuchal area and base of tail, but in one to several regular or irregular rows on tail; ventral surface smooth. Predorsal caudal vertebrae 35-55. Dark brown dorsally, plain or patterned with yellowish or dark spots, rings, ocelli or reticulations; whitish ventrally, with dark margins and darkly pigmented pores. Small-sized, usually <55 cm TL. Occurs over shallow continental shelf to slope, throughout tropical to warm-temperate waters of Indo-Pacific. Twelve species, 2 in WIO.

#### **KEY TO SPECIES**

- 1a Dorsal surface of disc with numerous brown spots and streaks but no ocelli; complete median row of thorns from nuchal area to 1st dorsal fin, and 1 interdorsal thorn; prenasal length <10% TL; interspiracular distance >7% TL; distance between 5th gill slits >9% TL
- 1b Dorsal surface of disc with or without spots and streaks, and with 2 or more ocelli; no median row of thorns between nuchal area and 1st dorsal fin, 2 interdorsal thorns; prenasal length >12% TL; interspiracular distance <6% TL; distance between 5th gill slits <8.8% TL</p>

### Okamejei heemstrai (McEachran & Fechhelm 1982)

#### East African skate

PLATE 64

Raja (Okamejei) heemstrai McEachran & Fechhelm 1982: 441, Figs. 1–5 (Kenya); Séret 1989; Fricke & Al-Hassan 1995.

Okamejei heemstrai: McEachran & Dunn 1998; Compagno 1999, 2005; Jeong *et al.* 2007; Compagno & Ebert 2007\*; Ebert *et al.* 2008.

Disc moderately narrow, DW 56–63% TL; anterior margins not convex. Snout very acute and relatively long, length 15–17% TL. Teeth in upper jaw 31–35. Eye diameter greater than interorbital space. Pelvic-fin anterior lobes relatively long, not reaching rear edge of posterior lobes. Tail moderately long, length 52–56% TL, and slender at base and attenuated distally. Interdorsal distance greater than length of 1st dorsal-fin base; 1st dorsal fin anteriorly set. Dorsal surface with 1–4 small orbital thorns, 1 interspiracular, 1 or 2 nuchal, in 3–5 irregular rows on tail between base and 1st dorsal fin, and 2 interdorsal. Predorsal caudal vertebrae 47–53.

Disc dark brown dorsally, with symmetrically arranged, small, bicoloured ocelli and sooty blotches on disc, pelvic fins and tail; pale grey ventrally. Attains at least 61 cm TL.



Okamejei heemstrai (Mozambique). © P Bouchet, MNHN

DISTRIBUTION WIO: Mozambique to Kenya.

**REMARKS** Known from few specimens; 2 mature females measured 60 and 61 cm TL. Occurs on outer continental shelf and uppermost slope, at ~200–500 m.

### Okamejei pita (Fricke & Al-Hassan 1995)

#### Pita skate

*Raja* (*Okamejei*) *pita* Fricke & Al-Hassan 1995: 3, Figs. 1–2 (Fao, Iraq, Persian/Arabian Gulf).

Okamejei pita: McEachran & Dunn 1998; Compagno 1999, 2005; Jeong *et al.* 2007; Last & Gledhill 2008.

Holotype: disc moderately broad, DW ~64% TL; anterolateral margins slightly convex. Snout moderately acute and not extended, and moderately long, length ~15% TL. Teeth in upper jaw 46. Eye diameter less than interorbital space. Pelvic-fin anterior lobes  $\sim$ <sup>1</sup>/<sub>4</sub> length of posterior lobes. Tail length ~50% TL, and relatively broad at base and attenuated distally. Interdorsal space narrow, <1/2 length of 1st dorsal-fin base. Dorsal surface covered with denticles except for area on centre of pectoral fins; 5 small orbital thorns, 3 medial to spiracle, ~9 nuchal, 2 scapular, ~30 in median row on mid-dorsal disc

PLATE 65

to tail, and 1 interdorsal. Predorsal caudal vertebrae 58.

Disc pale brown dorsally, with small dark brown blotches on disc and pelvic fins; pale brownish ventrally. Attains at least 46 cm TL.



*Okamejei pita*, 46 cm TL, female holotype (Iraq). Source: Fricke & Al-Hassan 1995

**DISTRIBUTION** Known only from the holotype (female) in the Persian/Arabian Gulf.

**REMARKS** Collected over mud bottom in shallow water (<15 m deep). The genus for this species is uncertain, and its likely it will eventually be placed in the genus *Leucoraja* or *Rajella*. IUCN Red List conservation status Critically Endangered.

### Orbiraja powelli (Alcock 1898)

Indian ring skate

*Raja powelli* Alcock 1898: 145 (Gulf of Martaban, Myanmar); Alcock 1899 [Fig.].

*Raja* (*Okamejei*) *powelli*: Stehmann 1976; McEachran & Fechhelm 1982; Séret 1989; Fricke & Al-Hassan 1995.

*Okamejei powelli*: McEachran & Dunn 1998; Compagno 1999, 2005; Manilo & Bogorodsky 2003; Jeong *et al.* 2007; Last & Gledhill 2008. *Orbiraja powelli*: Last *et al.* 2016\*.

Small-sized with moderately broad disc, DW 61–63% TL; anterolateral margins slightly convex. Snout very acute, extended and relatively long, length 15–16% TL. Teeth in upper jaw 72–75. Eye diameter less than interorbital space. Pelvic-fin anterior lobes ~¾ length of posterior lobes. Tail moderately long, length ~52% TL, and slender at base and attenuated distally. Interdorsal space greater than length of 1st dorsal-fin base; 1st dorsal fin anteriorly set. Dorsal surface with 3–5 small orbital thorns, 3 nuchal, 24–33 in irregular row on tail, and 5 or 6 interdorsal. Predorsal caudal vertebrae 49–55.

Disc orange-brown to dark brown dorsally, with scattered paler spots overall, and large circular target-like eyespot at each pectoral-fin base. Attains 53 cm TL (Last *et al.* 2016).



GENUS Orbiraja Last, Weigmann & Dumale 2016

Similar to and until recently included in the genus *Okamejei*. Tail with 3 well-developed rows of thorns along the midline, closely spaced and distant from lateral thorn rows. Tooth rows in upper jaw 61–76. Disc brown dorsally; pectoral fins with a pair of moderately large, ring-like ocelli. Ventral head without dark-edged pores. Small-sized, <55 cm TL. Occurs over continental shelf to slope, typically below 100 m depth, in WIO (Gulf of Aden, India) and Philippines. Three nominal species, 1 in WIO but additional undescribed species are suspected. Orbiraja powelli (India). Source: Alcock 1899

**DISTRIBUTION** Indian Ocean: Gulf of Aden (Yemen) and India (Travancore coast).

**REMARKS** Possibly the same species as *Orbiraja philipi* (Lloyd 1906) from the Gulf of Aden (collected from ~250 m). Poorly known; occurs on continental shelf and upper slope, at 15–460 m. Caught rarely in demersal longlines and used for its meat. IUCN Red List conservation status Near Threatened.

### GENUS Raja Linnaeus 1758

Disc quadrangular, moderately broad to broad, DW generally 63-75% TL; pectoral-fin apex weakly angular to angular. Snout obtusely angled to rounded, slightly extended and moderately long. Eyes relatively small, eye diameter slightly less than interorbital space. Pelvic-fin anterior lobes considerably shorter than posterior lobes. Tail moderately short, and moderately broad at base and attenuated distally. Dorsal surface largely free of denticles; 2-6 orbital thorns (usually separated into anterior and posterior groups), 1-4 nuchal, 0-1 scapular, in median row on mid-dorsal disc to tail (row on disc may be interrupted or absent), and generally 1 or 2 interdorsal. Predorsal caudal vertebrae 45-69. Disc generally brown to dark brown dorsally, and patterned with pale or dark spots, dark bars or ocelli; white to dusky ventrally, with or without darker disc margins. Size range ~50-120 cm TL. This genus is in need of revision. Raja proper occurs in the eastern Atlantic and southwestern Indian oceans, and Mediterranean Sea; another assemblage occurs in the North Pacific, and another amphi-American group in the eastern Pacific and western Atlantic oceans. Currently 27-30 species recognised, 2 in WIO.

#### **KEY TO SPECIES**

- 1a
   Dorsal surface of disc scattered with faint brown spots, particularly along fin margins; circular or nearly circular, blue-black eyespot, with 3 distinct rings of colour, at each pectoral-fin base

   *R. ocellifera*

### Raja ocellifera Regan 1906

#### Twineye skate

PLATE 65

Raja miraletus (non Linnaeus 1758): Wallace 1967\*; Hulley 1969\*;

McEachran & Dunn 1998; Compagno 1999, 2005; Heemstra & Heemstra 2004; Compagno & Ebert 2007\*; Ebert *et al.* 2008\*.

Raja (Raja) miraletus (non Linnaeus 1758): Hulley 1972\*; Lloris 1982; SSF No. 25.14\*.

Raia ocellifera Regan 1906: 2, Pl. 2 (Algoa Bay, South Africa); SFSA No. 69\*; Smith 1964.

Raja ocellifera: Last et al. 2016\*.

Small-sized with angular disc; DW ~1.3 times disc length. Snout short, with rounded terminal process, length 2.2–2.5 times interorbital space; angle anterior to spiracles 110°–116°. Teeth in upper jaw 36–46. Eyes close-set. Distance from mid-vent to 1st dorsal-fin origin 1.1–1.4 times distance from snout tip to mid-vent. Juveniles with some scapular thorns (diminishing with growth), but largely without spines or denticles on dorsal and ventral surfaces. Thorns in adults: 2 or 3 preorbital and 2 post-orbital, 2 or 3 nuchal, 20–23 in median row on tail (from pelvic-fin area to 1st dorsal fin), and 0–2 interdorsal; adults with spinules on ventral surface of snout, otherwise dorsal and ventral disc surfaces smooth. Females with 2 lateral rows of large thorns, one on either side of tail. Egg cases oblong and very small, <5 cm long excluding horns; surface smooth but densely covered in fibres with fine striations beneath; no lateral keels; posterior apron ~1.3 times length of anterior apron; horns stout and equal in length; posterior horns taper to filamentous tips, anterior horns hooklike and taper to acute tips. Predorsal caudal vertebrae 46–48.

Disc reddish brown dorsally, with scattered faint brown spots, particularly along fin margins; large blue-black eyespot with 3 distinct rings of colour at each pectoral-fin base; whitish ventrally, sometimes with small dark spot on snout tip. Attains  $\sim$ 50 cm TL.



Raja ocellifera, 28 cm DW, 42 cm TL, male (South Africa). Source: SSF

**DISTRIBUTION** WIO: South Africa (Cape Point, Western Cape, to Thukela Bank, KwaZulu-Natal, and abundant on Agulhas Bank).

**REMARKS** Smallest free-swimming specimen ~27 cm TL; hatchling size unknown. Males mature at 38–43 cm TL, and females at 40–45 cm TL, both sexes at ~2–3 years. Occurs on continental shelf to upper slope, over a range of soft bottom (sand-mud and seagrass beds), from 10–106 m. Feeds on various benthic animals and fish offal. Abundant and exploited throughout much of its range; caught by ski-boat anglers and likely a bycatch of deepwater fisheries. IUCN Red List conservation status Endangered.

### Raja straeleni Poll 1951

#### Biscuit skate

PLATE 65

Raja straeleni Poll 1951: 118, Fig. 54, Pls. 8, 13 (West Africa); Hulley 1970\*;
McEachran & Dunn 1998; Compagno 1999, 2005; Heemstra & Heemstra 2004\*; Compagno & Ebert 2007\*; Ebert et al. 2008\*.

Raja (Raja) straeleni: Hulley 1972\*; Lloris 1982; SSF No. 25.22\*. Raja clavata (non Linnaeus 1758): SSF No. 25.8\*.

Disc broad and angular; DW 1.2-1.5 times disc length. Snout moderately long, length 2.8-3.2 times interorbital space; tip triangular and broad-based, angle anterior to spiracles 95°-100°. Teeth in upper jaw 35-45. Distance from mid-vent to 1st dorsal-fin origin 1.2-1.5 times distance from snout tip to mid-vent. Ventral surface with spines on snout tip and anterior margins of disc. Dorsal surface largely free of denticles, but with strong thorns at all stages: 1 or 2 preorbital and 1-3 postorbital, 0-1 supraspiracular, 0-2 scapular, 4 or 5 nuchal, 20-52 in median row from mid-dorsal disc to 1st dorsal fin, and 0-6 interdorsal; also, a few lateral thorns on front portion of tail, with more hook-like thorns towards tip. Males entirely covered with spinules dorsally; females with spinules only on snout tip, anterior margins of disc, mid-dorsal disc and tail. Egg cases medium-sized, ~7.5 cm long excluding horns; surface smooth but densely covered in fibres; broad lateral keel; length of posterior apron ~1.3 times anterior apron; horns taper to filamentous tips, and length of posterior horns ~1.2 times anterior horns. Predorsal caudal vertebrae 48-52.

Disc pale brown to grey dorsally; adults with scattered small to large black spots, sometimes forming rosettes or whorls (no dark spots in juveniles); often with transversely oblong gold and black eyespot at each pectoral-fin base; white or mottled grey ventrally, and pores not pigmented. Attains ~91 cm TL (Last *et al.* 2016); females reach larger sizes than males.

**DISTRIBUTION** Eastern Atlantic (Mauritania to South Africa), and to South Africa (Eastern Cape) in WIO, and possibly also Mozambique, southern Madagascar and Mauritius.

**REMARKS** Has been confused with *R. clavata* from Northern Hemisphere, which needs further taxonomic study. Size at hatching <16 cm TL. Males mature at 56–68 cm TL, and females at 55–71 cm TL. Very common in southern Africa; found in shallow bays and near shore to continental slope, to ~820 m deep (usually above 200 m). Migrates inshore during the warm season and towards deeper offshore waters in winter. Feeds on bony fishes, benthic invertebrates and fish offal. Caught by shore and ski-boat anglers and hake trawlers for their edible pectoral fins.



Raja straeleni, 36 cm DW, female (South Africa). Source: SSF



Raja straeleni, 60 cm TL (South Africa). O Alvheim © IMR

### GENUS Rajella Stehmann 1970

Disc heart-shaped to rhomboidal, narrow to moderately narrow, DW generally 49-61% TL; pectoral-fin apex obtusely angled to broadly rounded. Snout slightly to moderately extended, acutely angled to broadly rounded, and moderately short to moderately long. Eyes moderately sized, eye diameter slightly less than to greater than interorbital space. Pelvicfin anterior lobes short to moderately long but considerably shorter than posterior lobes. Tail moderately long to long, length generally 51-60% TL, and moderately broad at base and attenuated distally. Dorsal surface largely covered with denticles and relatively densely covered with medium to large thorns; generally with complete row of orbital thorns, triangular patch on nuchal and scapular area, in 1 to several rows from shoulder area to 1st dorsal fin, and 1 to several interdorsal thorns; size of thorns in median row equal to or larger than those in lateral rows. Predorsal caudal vertebrae 55-73. Disc pale grey,

brown or brownish black dorsally; whitish to nearly black ventrally. Small-sized, adults attain ~70–90 cm TL. Occurs on outer continental shelf and slope to deep slope. Distributed throughout the Atlantic and southwestern Indian oceans, but also one member each from off Indonesia, the Galápagos, and Australia in the Pacific Ocean. Fifteen species, 5 in WIO, two of which are from deep water.

#### **KEY TO SPECIES**

- 2b Median row of thorns on disc interrupted or thorns reduced in size; tail grey ventrally, with white dorsal tip ...... *R. dissimilis* [deep water, from 420–1 640 m; Plate 66]

### Rajella barnardi (Norman 1935)

Bigthorn skate

PLATE 66

*Raja barnardi* Norman 1935: 43, Fig. 14 (off Cape Town, South Africa). *Raja confundens* Hulley 1970: 203, Pl. 11, Fig. 17 (off Cape Columbine, South Africa).

Raja (Rajella) confundens: Hulley 1972\*; SSF No. 25.9\*; Ebert et al. 1991. Raja (Rajella) barnardi: Stehmann 1995.

Rajella barnardi: McEachran & Dunn 1998; Compagno 1999, 2005;

Long & McCosker 1999; Compagno & Ebert 2007\*; Ebert *et al.* 2008\*; Last & Stehmann 2008.

Disc angular, DW 1.2–1.3 times disc length; pectoral-fin apex rounded. Snout moderately long, length 2.5–3.3 times interorbital space; tip not attenuating but continuous with anterior margins of disc, and with short rounded terminal process; angle anterior to spiracles 99°–120°. Mouth small and jaws weak, with blunt teeth; teeth in upper jaw 39–45. No interdorsal space. Ventral surface smooth. Dorsal surface with 5–10 circumorbital thorns, 1 or 2 supraspiracular and 1 or 2 interspiracular, 2–8 lateral nuchal and 2–9 medial nuchal, 2 or 3 scapular, and 17–24 in median row from nuchal area to 1st dorsal fin, flanked by 2 lateral rows on tail; thorns with stellate bases over rostral cartilage and anterior margins of disc. Egg cases rather small, ~5.5 cm long excluding horns; surface smooth with fine striations; lateral keels broad and feathery, extending length of case (including horns); posterior apron ~1.5 times width of anterior apron; horns robust and tapered; posterior horns ~1.7 times length of anterior horns, anterior horns with attachment fibres near tips. Predorsal caudal vertebrae 55–63.

Body grey dorsally and ventrally, with darker patches along posterior margins of disc and pelvic fins, around vent, and along tail. Attains 75 cm TL.



Rajella barnardi, 44 cm DW (South Africa). Source: SSF

**DISTRIBUTION** Primarily eastern Atlantic (Mauritania to South Africa), and fewer records to South Africa (Eastern Cape) in WIO.

**REMARKS** Size at hatching <11 cm TL. Males mature at 60–62 cm TL, and females at 59–65 cm TL. Common, with wide depth range and concentrations in some areas; occurs on outer continental shelf to deep slope, at 128–1 809 m (usually >200 m). Feeds mainly on small crustaceans (mysids), also polychaetes, bony fishes and squid. Caught incidentally in deep-trawl fisheries.

### Rajella caudaspinosa (Von Bonde & Swart 1923)

#### Munchkin skate

PLATE 66

Raia caudaspinosa Von Bonde & Swart 1923: 8, Pl. 21,

- Fig. 1 (KwaZulu-Natal, South Africa); Hulley 1970\*.
- *Raia albalinea* Von Bonde & Swart 1923: 6, Pl. 20, Fig. 1 (KwaZulu-Natal, South Africa).

Raja (Rajella) caudaspinosa: Hulley 1972\*; SSF No. 25.7\*.

Rajella caudaspinosa: McEachran & Dunn 1998; Compagno 1999, 2005; Long & McCosker 1999; Compagno & Ebert 2007\*; Ebert *et al.* 2008\*; Last & Stehmann 2008.

Small-sized with rough tail and thick disc, DW 1.2-1.4 times disc length; pectoral-fin apex broadly rounded. Snout obtusely pointed and with rounded terminal process, and short, length 2-2.6 times interorbital space; angle anterior to spiracles 125°-130°. Eyes large, bulging and close-set. Mouth small and jaws weak, with blunt teeth; teeth in upper jaw 32-36. Tail thick and longer than disc; no interdorsal space. Ventral surface smooth. Dorsal surface very rough: 5-9 circumorbital thorns, 0-1 interorbital, 1 or 2 interspiracular, 3 or 4 scapular, 4 or 5 median nuchal, 22-23 in mid-dorsal row from nuchal area to 1st dorsal fin, thorns decreasing in size towards tail, and flanked on each side by 1 row of thorns on back and 2 rows on tail; large thorns with stellate bases in 4-6 staggered rows on anterior margins of disc. Egg cases poorly known, but halfdeveloped specimen was small, ~3 cm long excluding horns; surface finely striated and smooth beneath fibrous covering, with narrow lateral keels, and posterior horns tapering to acute tips. Predorsal caudal vertebrae 66-73.

Disc pale grey to brownish dorsally, with or without scattered darker spots, and juveniles with distinctive white bars; whitish ventrally. Attains 32+ cm DW, 65 cm TL.



Rajella caudaspinosa, 17 cm DW, 35 cm TL, juvenile holotype of Raia albalinea (South Africa). Source: Von Bonde & Swart 1923



*Rajella caudaspinosa*, 26 cm DW, female (top); 29 cm DW, male (bottom) (South Africa). Source: SSF

**DISTRIBUTION** Southern Africa: most common from Namibia to South Africa (Cape Agulhas) in southeastern Atlantic; rare off south Cape coast of South Africa in WIO.

**REMARKS** Size at hatching <10 cm TL. Males mature at 53–59 cm TL, and females at 50–57 cm TL. Similar in appearance to other members of the genus. Probably uncommon; occurs on outer continental shelf and upper slope, at 102–1 098 m deep (usually >200 m). Feeds mainly on small crustaceans (mysids), also lightfishes and polychaetes. Taken incidentally in hake trawl fisheries.

### Rajella leopardus (Von Bonde & Swart 1923)

Leopard skate

PLATE 66

*Raia leopardus* Von Bonde & Swart 1923: 7, Pl. 20, Fig. 2 (KwaZulu-Natal, South Africa).

*Raja* (*Rajella*) *leopardus*: Hulley 1972\*; SSF No. 25.13\*; Stehmann 1990, 1995; Compagno *et al.* 1991.

*Rajella leopardus*: McEachran & Dunn 1998; Compagno 1999; 2005; Long & McCosker 1999; Compagno & Ebert 2007\*; Ebert *et al.* 2008\*; Last & Stehmann 2008.

Disc angular, barely wider than long; pectoral-fin apex rounded. Snout obtusely pointed with triangular terminal process and moderately long, length 2.8-4.4 times interorbital space; angle anterior to spiracles 100°-110°. Tail about as long as disc, with thick base. Jaws large, teeth blunt; teeth in upper jaw 52-70. Dorsal surface in juveniles and adults with spinules on snout, anterior margins of disc and sides of tail. Thorns in juveniles: 4 or 5 circumorbital, none or 1 supraspiracular, 1 interspiracular, 1 or 2 scapular, 3-5 median nuchal, 25-30 mid-dorsal thorns from nuchal area to 1st dorsal fin, and 3-6 small thorns on each side of tail at pelvic fins. Thorns in adults: 5–13 circumorbital and supraspiracular, 1 or 2 interspiracular, 2 or 3 scapular, 4–9 median nuchal thorns forming triangular patch of 1 or 2 rows, and 19-29 in middorsal row from mid-disc to 1st dorsal fin, flanked on each side by 1 row along back and 2 or 3 rows on tail. Ventral surface in adults with spines on snout tip and anterior margins of disc. Egg cases small, ~5.5 cm long excluding horns; surface smooth; lateral keels broad and feathery in appearance, extending full length, including horns; posterior horns ~1.2 times as long as anterior horns, anterior horns with fibrous tendril. Predorsal caudal vertebrae 55-58.



*Rajella leopardus*, 38 cm DW (South Africa). Redrawn from Hulley 1970

Disc grey to brown dorsally, with numerous dark spots (more common in juveniles); pale ventrally, sometimes mottled with dusky blotches and patches. Attains 93 cm TL.

**DISTRIBUTION** Southern Africa: Namibia to Cape of Good Hope in southeastern Atlantic, to South Africa (Eastern Cape) in WIO.

**REMARKS** Size at hatching <15 cm TL; matures at 61–73 cm TL. Poorly known, yet the most common skate off southern Africa, along with *R. barnardi*. Occurs on upper continental slope, at 73–1 026 m (usually 300–500 m). Feeds mostly on decapod crustaceans and bony fishes (including dragonets, rattails, hake and lanternfish), also sea pens, cuttlefish and polychaetes. Caught incidentally in hake trawl fisheries.

**EDITORS' NOTE** In the original description it was not specified if *leopardus* was used as a noun or an adjective. We follow *Eschmeyer's Catalog of Fishes* in assuming it is a noun. Other publications have used the adjectival form, *Rajella leoparda*.



Rajella leopardus, 15 cm DW, 25 cm TL, type (South Africa). Source: Von Bonde & Swart 1923

### GENUS **Rostroraja** Hulley 1972

One species.

### Rostroraja alba (Lacepède 1803)

White skate

PLATE 67

Raja alba Lacepède 1803: 661, 663, Pl. 20, Fig. 1 (France, English Channel); Hulley 1966\*, 1970\*; Wallace 1967\*.

Raja (Rostroraja) alba: Hulley 1972\*; SSF No. 25.6\*.

*Rostroraja alba*: McEachran & Dunn 1998; Heemstra & Heemstra 2004; Compagno 2005; Compagno & Ebert 2007\*; Ebert *et al.* 2008\*.

Large-sized with broad disc, DW 1.4-1.5 times disc length; pectoral-fin apex angular. Snout broad-based and attenuating to long point covered with small thorns; snout length 2.5-3.2 times interorbital space; angle anterior to spiracles ~105°. Jaws powerful, with sharp teeth; teeth in upper jaw 40–45. Ventral surface in juveniles with spinules on snout, internasal area, and anterior margins of disc; ventral surface in adults with spinules on snout, internasal area, anterior margins of disc, gill slits, abdomen and tail. Dorsal surface in juveniles with 1 preorbital and 0-1 post-orbital thorns, 10-16 from pelvic fins to 1st dorsal fin, 7-17 in lateral row on tail, and 0-2 interdorsal. Dorsal surface in adults with ~6 circumorbital thorns, 16-30 in median row from pelvic fins to 1st dorsal fin, 17-29 in lateral row on tail, and 0-2 interdorsal; no nuchal or scapular thorns. Egg cases oblong and very large, >12 cm long excluding horns; surface coarsely striated and rough; broad lateral keels; posterior horns very long, ~3 times length of anterior horns, and tapering and flattening towards tips; anterior horns short, tapered, and with attachment fibres. Predorsal caudal vertebrae 62-67.

Adults grey to brown dorsally, usually with many small white spots (no dark spots); white ventrally. Juveniles brown dorsally, with darker margins on disc and fins; white ventrally, with grey margins on disc. Hatchlings reddish brown dorsally, often with blue spots. Attains 160 cm DW, 240 cm TL (females reach larger sizes than males).

**DISTRIBUTION** Mediterranean Sea, eastern Atlantic (British Isles to South Africa), and rounding Cape of Good Hope (South Africa) to central Mozambique in WIO.

**REMARKS** Size at hatching <40 cm TL. Males mature at ~120 cm TL, and females at ~130 cm TL (Last *et al.* 2016). Bottom predator, feeds mainly on bony fishes but also other elasmobranchs, fish offal, crustaceans and cephalopods. Found on continental shelf and upper slope, over sandy and rock-sand bottom, at 10–750 m (usually 50–500 m); once frequently captured but now rare in many parts of its range. Common in

trawls off South Africa and occasionally taken by anglers off sandy beaches. IUCN Red List conservation status Endangered.



Rostroraja alba, 30 cm DW, 30 cm TL, juvenile (South Africa). Source: SSF



Rostroraja alba, 66 cm DW (South Africa). Source: SSF

# FAMILY ARHYNCHOBATIDAE

#### Softnose skates

Neil C Aschliman and David A Ebert

Large group of morphologically diverse skates, yet distinguished by basihyal cartilage supporting the floor of mouth with lateral projections, and external clasper glans components relatively few and similar in most species. (External morphology is otherwise difficult to use in distinguishing softnose skates from hardnose skates of family Rajidae.) Disc quadrangular to rhomboidal. Snout tip typically flexible due to supporting rostral cartilage being very slender, uncalcified, segmented, and/or separated from the neurocranium (pattern of these features varying between genera). Mouth transverse to arched; teeth numerous. Tail slender, with lateral skin folds, weak electric organs, usually 2 posteriorly set small dorsal fins, and reduced caudal fin. Skin often prickly, particularly along dorsal midline.

Reproductive mode oviparity; females lay horny egg cases with long tips. Benthic in shallow tropical waters. Rare, although some species are of commercial interest as skate 'wings' are considered good eating. Distributed mostly in the Pacific, southwestern Atlantic, and southern Indo-Pacific; uncommon in WIO, being largely replaced by hardnose skates.

About 14 genera and at least 111 species; 2 genera with 3 species in WIO, 2 of which are deepwater species, *Notoraja hesperindica* Weigmann, Séret & Stehmann 2021 and *Bathyraja tunae* Stehmann 2005, both found off Mozambique and Madagascar.

GENUS Bathyraja Ishiyama 1958

Softnose skates with sub-rhomboidal disc and uncalcified flexible snout. Small-sized to giant, size range ~50–175 cm TL. Distributed mostly throughout northern and southeastern Pacific, southwestern Atlantic, and Southern Ocean, with few species occurring in the Atlantic or WIO. Some species occur at tremendous depths, over 3 000 m. At least 50 species; 2 species in WIO, but *Bathyraja tunae* from the Mozambique Channel is known only from a deepwater slope, at >1 700 m.

### Bathyraja smithii (Müller & Henle 1841)

Softnose skate

PLATE 67

Raja smithii Müller & Henle 1841: 150, Pl. 49 (South Africa). Raia smithii: Von Bonde & Swart 1923.

Raia smithi: Barnard 1925\*.

Breviraja smithii: Hulley 1970\*, 1972\*.

Bathyraja smithii: SSF No. 25.1\*; Stehmann 1986; Compagno et al. 1989\*,

1991; Ebert *et al.* 1991, 2008; Compagno 2005; Compagno & Ebert 2007\*.

Mouth broad, with strong jaws and sharp teeth. Dorsal-fin bases separated. Disc width 1.3–1.4 disc length (Last *et al.* 2016); tail shorter than disc length. Preorbital snout length 2–2.6 interorbital width; angle of snout anterior to spiracles 90°–100°.

Tooth count 23–28 in upper jaw. Skin spinulose dorsally and smooth ventrally at all life stages; thorns on dorsal disc lost in adults, except for single median row of 14–19 large thorns on tail, sometimes 1 interdorsal thorn, but no lateral thorns on tail; thorns on dorsal surface of disc in juveniles: 1 preorbital, 1 or 2 post-orbital, 3 or 4 scapular, 2 nuchal, ~30 median from nuchal region to 1st dorsal fin, and 1 interdorsal. Predorsal vertebrae 68–71.

Body greyish dorsally, sometimes with white spots; mostly white ventrally, with dark grey or black blotches around gill slits and vent, along disc margins, and underside of tail. Attains 120 cm TL.



Bathyraja smithii, 62 cm DW (South Africa). Source: SSF

**DISTRIBUTION** Southern Africa: Namibia in southeastern Atlantic, to South Africa (Agulhas Plateau and Eastern Cape) in WIO.

**REMARKS** Size at hatching <13 cm TL. Males mature at 95–97 cm TL, and females at ~80–90 cm TL (Last *et al.* 2016). Egg cases probably very large: an incomplete specimen, only one-third developed, was ~8 cm long excluding horns, with striated surface covered with dense fibres, broad lateral keels, and very long and thin horns tapering to filamentous tips. Moderately abundant on deep slopes, known from 250–1 020 m (one report from 40 m requires validation, but mostly >600 m, and most records of juveniles). Feeds on bony fishes (particularly hake, jacopever, barracudina and dragonet) and crustaceans, including shrimp and crabs, and sometimes squid and octopus in deeper waters. Incidentally caught in deepwater hake trawl fisheries, but considered of no commercial value.

# FAMILY ANACANTHOBATIDAE

### Legskates

Neil C Aschliman and David A Ebert

Small to medium-sized skates (to ~70 cm TL) with subcircular disc; snout long, often with small terminal fleshy filament; pelvic fins with a unique deep incision completely separating the lobes, anterior lobes slender and limb-like (enabling them to 'walk' across the benthos); tail short, uniformly slender, dorsal fins absent, and caudal fin rudimentary. Eyes small to large; spiracles small, subcircular. Mouth narrow. Skin usually smooth (except median denticle row on tail of small juveniles of *Sinobatis melanosoma*), or with patches of thorns confined to alar region in adult males.

Reproductive mode oviparity. Restricted distributions in deep water, over outer continental shelf and slope of western Atlantic, western Pacific and southwestern Indian oceans, with known depth range 150–1 725 m. Some species known only from a few specimens (some of which have been subsequently lost). Five genera and 14 species recognised; 3 genera and 3 species in WIO (of which 2 genera and 2 species known from deep water).

#### **KEY TO GENERA**

1a	Tail short, 35–36% TL, ~½ body length; dorsal disc pale
	grey-brown, paler ventrally
	[one deepwater species, Sinobatis brevicauda Weigmann & Stehmann 2016,
	from 960–1 130 m, Saya de Malha Bank, in WIO]
1b	Tail long, more than ½ body length to greater than body
	length; dorsal disc dark grey-brown or reddish brown with

ocelli and pale spots

# GENUS Anacanthobatis

Von Bonde & Swart 1923

Legskates with terminal filament extending from small rounded projection at tip of snout. Tail slender, and shorter or longer than disc; no dorsal fins; caudal fin weakly developed, but with lower lobe which is smaller than upper lobe; pelvicfin posterior lobes fused to base of tail along length. Skin smooth. Occurs on continental shelf and slope, in tropical and subtropical waters of western central Atlantic, western Pacific and WIO. Ten species, 1 in WIO.

### Anacanthobatis marmorata

Von Bonde & Swart 1923

Spotted legskate

Leiobatis marmoratus Von Bonde & Swart 1923: 18, Pl. 23 (KwaZulu-Natal,

PLATE 68

South Africa); Wallace 1967\*; Hulley 1973\*. Anacanthobatis marmoratus: SSF No. 26.1\*; Compagno et al. 1989\*;

Compagno 2005; Compagno & Ebert 2007\*; Ebert 2014\*.

Anacanthobatis marmorata: Last et al. 2016\*.

Diagnosis as for genus. Disc width ~1.2 times disc length. Tail shorter than disc length, with narrow lateral skin folds. Snout angle anterior to spiracles ~102°. Internasal flap fringed and overlapping corners of mouth. Tooth count 28–35 in upper jaw. Dorsal surface with dermal papillae; dorsal and ventral surfaces otherwise smooth, without dermal denticles, but adult males with alar thorn patch.

Body mottled pale brown and white dorsally, with scattered ocelli; terminal filament and dermal papillae dark brown; pale ventrally. Attains at least 29 cm TL (Last *et al.* 2016).

**DISTRIBUTION** WIO: South Africa (KwaZulu-Natal) to southern Mozambique.

**REMARKS** Possibly the smallest southern African skate, although known from few records. Occurs on edge of outer continental shelf and upper slope, with reliable depth records of 162–433 m.



*Anacanthobatis marmorata*, 11 cm DW, ventral view of juvenile (S Mozambique).



*Anacanthobatis marmorata*, 17 cm DW, male (S Mozambique). Source: SSF

# FAMILY **GURGESIELLIDAE**

### Pygmy skates

Neil C Aschliman and David A Ebert

Small-sized skates (to ~60 cm TL) with subcircular to rhomboidal disc; snout short to moderately long, rigid to semi-rigid, with or without small process at tip; pelvic fins deeply notched, anterior lobes slender and limb-like (lobes fused in *Gurgesiella* species) enabling a 'walking' motion across the benthos; tail slender and long, usually with 2 posteriorly set small dorsal fins, or 1 fin, or dorsal fins absent, and caudal fin small. Eyes large; spiracles small, subcircular. Mouth narrow. Skin with sparse small denticles, and thorn patterns variable between genera and species, but adult males of all genera with alar and malar thorns.

Reproductive mode oviparity. Restricted distributions over outer continental shelf and slope of western Atlantic,

western Pacific and southern Indian oceans, known to 1 095 m (Last *et al.* 2016), and inshore to 39 m, off South Africa. Some species are well recorded and others are known only from one or several specimens (some of which have been subsequently lost). Three genera and 19 species recognised worldwide with 2 genera and 5 species in WIO. One deepwater WIO genus, with 2 species, *Fenestraja maceachrani* (Séret 1989) from off Madagascar in 600–765 m, and *F. mamillidens* (Alcock 1889) from Gulf of Mannar and western Sri Lanka known from ~1 090 m.

### GENUS Cruriraja Bigelow & Schroeder 1948

Legskates without terminal filament on snout. Tail stout at base, and with lateral skin folds; 2 dorsal fins; caudal fin weakly developed, lower lobe very weakly developed or absent. Dorsal surface of disc with sparse thorns and sometimes with rows of thorns on midback or tail; ventral surface of disc and tail smooth. Occurs on continental slope, in tropical and subtropical waters of western central to southwestern and southeastern Atlantic and WIO, at ~150 to >1 000 m deep. Eight species, 3 in WIO.

#### **KEY TO SPECIES**

1a 1b	Snout with ~15 rostral thorns <i>C. andamanica</i> Snout with 3–7 rostral thorns 2
2a	No thorns on midback; tips of pelvic-fin anterior lobes pointed
2b	Thorns in continuous rows on midback; tips of pelvic-fin anterior lobes spatulate

### Cruriraja andamanica (Lloyd 1909)

#### Andaman pygmy skate

*Raia andamanica* Lloyd 1909: 140, Pl. 46, Fig. 2 (Andaman Sea). *Cruriraja andamanica*: Stehmann 1976\*; McEachran & Fechhelm 1982; Fischer *et al.* 1990; Compagno 2005; Ebert 2014\*.

Diagnosis as for genus. Snout with ~15 rostral thorns; scapular thorns present; thorns in single row from nuchal to mid-disc region, and in irregular row on tail.

Body uniformly slate grey dorsally and ventrally. Attains at least 50 cm TL (Last *et al.* 2016).



*Cruriraja andamanica*, 21 cm TL, male holotype (Andaman Sea). Source: Lloyd 1909

**DISTRIBUTION** WIO: Known from few specimens, collected off Tanzania, and also from Andaman Sea and possibly off Indonesia (Last *et al.* 2016).

**REMARKS** Taken in tropical water over continental slope, at 274 m (Tanzania) and 511 m (Andaman Sea).

#### Cruriraja hulleyi Aschliman, Ebert & Compagno 2010

Roughnose legskate

PLATE 68

*Cruriraja hulleyi* Aschliman, Ebert & Compagno 2010: 364, Figs. 1–3 (south coast of South Africa); Ebert 2014\*.

*Cruriraja parcomaculata*: Smith 1964\*; Wallace 1967\*; Hulley 1970\*, 1972\*; Compagno *et al.* 1989\*; Compagno 2005; Compagno & Ebert 2007\*.

Disc quadrangular, ~1.2 times wider than long (DW ~57% TL). Snout short, tip pointed, without terminal filament; snout angle anterior to spiracles 80°-90°. Teeth blunt and flat in females and juveniles, and acute with single cusp in adult males; tooth count 37-47/38 or 39. Pelvic-fin anterior lobe tips spatulate. Tail longer than disc, broad, depressed and tapering to 1st dorsal fin; lateral skin folds long, extending to tail tip, and broadest at level of dorsal fins. Interdorsal distance greater than half (but not exceeding) length of dorsal-fin bases. Dorsal fins moderate and equally sized; 1st dorsal-fin tip rounded, 2nd dorsal-fin tip angular; interdorsal distance greater than half (but not exceeding) length of dorsal-fin bases. Dorsal surface generally smooth other than areas of thorns: rostral thorns 4 or 5; orbital thorns 9, in regularly spaced medial semicircle from preorbital area to spiracle; post-orbital thorns 2; scapular thorns in tight irregular patches of 4-9 thorns each; nuchal thorns 1-3; thorns

in 1–5 continuous rows on midback, followed by 1 or more rows of well-developed and strongly recurved tail thorns; interdorsal thorns 6. Ventral surface entirely smooth. Precaudal vertebrae 66–69.

Body yellowish brown dorsally, with large darker brown blotches spanning eyes and along rows of thorns on midback; dorsal surface of claspers and tail duskier and darker brown; uniformly pale ventrally. Juvenile specimen densely mottled dorsally, with circular brown spots on pale field, and tail with brown banding interspersed with smaller brown spots. Attains 60 cm TL.



Cruriraja hulleyi, 33 cm DW (South Africa). Source: SSF

**DISTRIBUTION** Southern Africa: Namibia (Lüderitz) to South Africa (East London, Eastern Cape).

**REMARKS** Individuals tend to be larger on west coast of southern Africa than on south coast. Size at hatching ~10 cm TL; matures at ~47 cm TL. Egg cases small, vase-like, <5 cm long excluding horns, with coarsely striated surface and very narrow lateral keels; posterior horns longer than anterior horns, tapering to thin tips, curving inwards and with fine attachment fibres; anterior horns hook-shaped, with acute tips. Moderately common on outer continental shelf and upper slope, at 39–545 m (most commonly 200–500 m). Feeds mostly on mysid shrimp, but also small bony fishes and various invertebrates, including crustaceans, squid, polychaetes and flatworms. Caught incidentally by hake bottom trawlers.

### Cruriraja parcomaculata (Von Bonde & Swart 1923)

#### Triangular legskate

PLATE 68

Raia parcomaculata Von Bonde & Swart 1923: 9, Pl. 21, Fig. 2 (KwaZulu-Natal, South Africa).

*Cruriraja triangularis*: Smith 1964\*; Wallace 1967\*; Hulley 1970, 1972; Compagno *et al.* 1989\*; Compagno 2005; Compagno & Ebert 2007\*. *Cruriraja parcomaculata*: Aschliman *et al.* 2010; Ebert 2014\*.

Disc ~1.3 times wider than long (DW up to 62% TL). Snout short, tip pointed, without terminal filament; snout angle anterior to spiracles 60°–70°. Tooth count 39–42/34–42. Pelvicfin anterior lobe tips pointed. Interdorsal distance less than half length of dorsal-fin bases. Dorsal surface generally smooth other than areas of thorns: rostral thorns 3–7; post-orbital thorn 1; at most 1 scapular thorn (lacks patches of scapular thorns); nuchal thorn 1; no median row of thorns; interdorsal thorns 4. Ventral surface smooth, except for denticles on underside of claspers.

Body grey-brown dorsally, with darker patches alongside midline; pale ventrally; no bands on tail. Attains 43 cm TL.



Cruriraja parcomaculata, ~10 cm DW, 18 cm TL, juvenile type (South Africa). Source: Von Bonde & Swart 1923



*Cruriraja parcomaculata*, 16 cm DW, female holotype of *C. triangularis*, dorsal view; 12 cm DW, paratype, ventral view of pelvic fins (South Africa). Source: SSF

**DISTRIBUTION** WIO: South Africa (KwaZulu-Natal) to Mozambique.

**REMARKS** Smaller than *C. hulleyi* but of similar proportions; however, the snout is longer and has more acute angle. Males mature at 35–37 cm TL and attain >39 cm TL; females mature at 32–37 cm TL and attain ~43 cm TL. Size at hatching ~9 cm TL. Egg cases small, boxy, not vase-like, <5 cm long excluding horns, with coarsely striated surface, prominent longitudinal ridges and covered by thin fibrous layer, and without lateral keels; posterior horns very long, tapering towards tips and curved inwards, their length greater than egg case proper. Found on outer continental shelf and upper slope, on mud bottom, at 65–680 m (most records from >200 m).

# ORDER MYLIOBATIFORMES

# FAMILY HEXATRYGONIDAE

### Sixgill stingray

David A Ebert and Neil C Aschliman

Unusual ray with 6 pairs of gill slits on underside, flabby body, and long triangular fleshy snout with pointed flexible tip (which changes shape with growth). Disc widest just behind snout; disc length greater than disc width. Eyes small; spiracles far back from eyes and about twice eye diameter. Nostrils far apart; no oronasal grooves; internasal flaps very short, broad, fleshy, and not joined as flap reaching mouth. Mouth broad and nearly transverse. Teeth small, bluntly rounded, and exposed when mouth closed. Pelvic fins small, fleshy and broadly rounded. Tail shorter than disc length, with 1 or 2 long serrated stinging spines, no dorsal fins, and low caudal fin. Skin smooth, without dermal denticles or thorns.

Monotypic.



*Hexatrygon bickelli*, 23 cm DW, prenatal pup. The skin over the cranial cavity has been removed to show the size of the brain which does not increase with growth (South Africa). Source: SSF 1995

### GENUS Hexatrygon Heemstra & Smith 1980

Diagnosis as for family. Although at least 5 species have been described, the genus is considered monotypic as all specimens appear referable to one species, widely distributed in Indo-West to central Pacific.

### Hexatrygon bickelli Heemstra & Smith 1980

#### Sixgill stingray

PLATE 69

Hexatrygon bickelli Heemstra & Smith 1980: 6, Figs. 1–13, 15 (beach at Port Elizabeth, Eastern Cape, South Africa); Compagno *et al.* 1989\*; SSF No. 31.1\* [1995]; Last & Stevens 2009\*; Babu *et al.* 2011; Ebert 2014\*.

Diagnosis as for family. Tooth count 44–102 in both jaws (varies with growth); vertebrae 101; spiral valve turns 13 or 14.

Body brownish to dark violet-blue or pinkish dorsally, and snout uniformly paler; paler or white ventrally, except darker along disc margins; caudal fin dark brown to black. Attains 168 cm TL.



*Hexatrygon bickelli*, 50 cm DW, female holotype (South Africa). Source: Heemstra & Smith 1980

**DISTRIBUTION** Indo-Pacific. WIO: South Africa (south coast), Mozambique, India and Sri Lanka.

**REMARKS** Viviparous; litters of 3–5; size at birth ~50 cm TL. Matures at 105–113 cm TL. Rarely seen, deepsea ray of continental slope, at 300–1 120 m, found close to bottom but may venture into midwater; a few records are of strandings on beaches in South Africa, and once observed and photographed by divers in relatively shallow water (~30 m) off Japan.

# FAMILY **PLESIOBATIDAE**

Deepwater stingray

David A Ebert and Neil C Aschliman

Giant stingray with oval disc (slightly longer than wide), broadly pointed snout, and small eyes. Head not elevated above pectoral fins. Mouth large, with numerous small teeth, and no oral papillae. Nostrils almost circular; internasal flap broad, rectangular, and posterior edge with well-developed fringe. Tail slightly shorter than disc length, not whip-like, without lateral skin folds, and with 1–3 long serrated stinging spines near midpoint of tail; no dorsal fins; caudal fin slender, long and well developed. Dorsal surface of disc and tail covered with small close-set acutely pointed denticles; ventral surface similar, except smooth on disc edges, pelvic-fin margins, internasal flap and small area immediately behind mouth.

Monotypic.

### GENUS Plesiobatis Wallace 1967

Diagnosis as for family. One species, previously placed in family Urolophidae (round stingrays), patchily distributed in Indo-West and central Pacific.

### Plesiobatis daviesi (Wallace 1967)

Deepwater stingray

PLATE 69

*Urotrygon daviesi* Wallace 1967: 8, Figs. 3–4 (Mozambique Channel, off Limpopo River mouth); SSF No. 30.16\*; Compagno *et al.* 1989\*. *Plesiobatis daviesi*: Ebert *et al.* 2002; Last & Stevens 2009\*; Ebert 2014\*.

Diagnosis as for family. Tooth count 30–62 in both jaws; spiral-valve turns 14 or 15.

Body grey-black to brownish or purplish brown dorsally, and on pelvic fins, caudal fin and ventral margins of disc; white ventrally. Attains 270 cm TL, 151 cm DW.

**DISTRIBUTION** Indo-Pacific. WIO: South Africa (KwaZulu-Natal) to southern Mozambique, and southern India to Sri Lanka; elsewhere to Philippines, Taiwan, Kyushu-Palau Ridge (Philippine Sea), Ryukyu Is., Australia and Hawaii.

**REMARKS** Biology poorly known; research is needed on the different populations. Presumed viviparous with yolk sac; size at birth ~50 cm TL. Males mature at 130–172 cm TL, and females at >200 cm TL. Possibly rare; demersal on soft bottom of outer continental shelf and upper slope, from 44–700 m, mostly

>275 m. May migrate to midwater to feed. Diet includes bony fishes, cephalopods and crustaceans. Specimens observed with wounds from bites of cookiecutter shark *Isistius brasiliensis*.



Plesiobatis daviesi, 90 cm DW (S Mozambique). Source: SSF

# FAMILY **DASYATIDAE**

#### Stingrays

B Mabel Manjaji-Matsumoto, Marcelo R de Carvalho, Hugo RS Santos, Ulisses L Gomes and Peter R Last

Small- to very large-sized (~30 cm to 3 m DW) with head, trunk and broadly expanded pectoral fins forming flattened trapezoidal, rhomboidal or oval disc (disc width at most 1.3 times disc length). Snout variably obtuse, rounded, more or less pointed or acute; eyes and spiracles dorsally situated. Mouth transverse and ventral, and mouth floor usually with fleshy oral papillae; teeth small and numerous, in rows forming bands in both jaws. Five pairs of gill slits on underside. Internasal flap well-developed, posterior edge fringed. Tail slender to whip-like, sometimes significantly longer than disc, usually with skin folds (no skin folds in Urogymnus and Himantura); usually 1 or 2 long, serrated, venomous spines posteriorly set on tail (no stinging spine in Urogymnus) (stinging spines are often dropped, leaving a scar, and subsequently replaced); no dorsal fin; no caudal fin. Dorsal surface smooth or covered with small denticles, and variably with patches or rows of enlarged denticles, thorns, tubercles or other dermal derivatives (their size, density and distribution varying with growth and sex). Body uniformly coloured or with markings dorsally, some species ornately patterned; generally pale to whitish ventrally, and disc margins sometimes darker.

Reproductive mode viviparity with yolk-sac placenta and histotroph, wherein uterine secretions nourish the embryos;

litters of few young (<8). Worldwide, primarily in tropical to warm-temperate waters; usually found in shallow coastal marine waters, brackish lagoons and estuaries, and occasionally in freshwater, but a few species occur deeper than 100 m. Several species of *Dasyatis* and *Himantura* are restricted to freshwater in tropical regions of Africa, Southeast Asia and Australia. Most species are benthic or demersal, sometimes remaining partially buried in soft substrates for prolonged periods, and a few are semipelagic or pelagic. Abundant in certain localities and sold locally in markets as the thickest part of the disc is esteemed as meat in some regions. The venomous stinging spine can inflict a painful or fatal injury, although these rays are not naturally aggressive towards people. A few species are popular with aquarists because of their grace and beauty.

Nineteen genera and ~90 species; 14 genera and at least 27 species in WIO.



Continued ...

#### **KEY TO GENERA**

4a	Trunk thin, thickness less than 2 times snout length; small rays, hirth size 5–10 cm disc width
4b	Trunk relatively thicker, robust, thickness more than 2 times snout length; medium to large rays, birth size typically at 15–20 cm disc width, some species born at more than 30 cm disc width
5a	Snout rather long and pointed, length exceeding 3 times combined orbit and spiracle length
5b	Snout shorter, length less than 3 times combined orbit and spiracle
ба	Tail with dorsal and ventral skin folds, folds rudimentary to low; dorsal disc surface smooth, except for large adults with sparse denticles
6b	Tail with ventral skin fold, fold distinct; dorsal disc surface granular or with 1 or 2 median rows of thornlets
	6a 6b
7a	Tail beyond sting very thorny; dorsal disc colour uniform plain (row of pores on anterior disc white in one species)
7b	Tail only sparsely covered with denticles, usually as prickles confined to before sting and in large adults; dorsal disc colour uniform plain to blue marbling centrally <i>Dasyatis</i>
8a	Disc circular to oval; ventral skin fold reaching tip of tail;
8b	Disc rhomboidal; ventral skin fold not reaching tip of tail (when tail undamaged)
9a	Disc oval; dorsal disc with dark specks, or blue spots Taeniura
9b	Disc circular; dorsal disc with black and white mottling (may appear uniform grey due to
	sediment covering disc)
10a	Ventral skin fold on tail deep and prominent, its height 2–3 times tail height above fold; dorsal surface with a distinct denticle band
10b	Ventral skin fold on tail low, its height less than tail height above fold; dorsal surface without a distinct denticle band Megatrygon

Continued ...

#### **KEY TO GENERA**

11a	Disc subcircular to oval; dorsal disc uniform brown to grey, without any spotting or colour pattern; tail not banded 12
11b	Disc suboval to rhomboidal; dorsal disc plain, with speckles, spots, rings (large adults with reticulations); tail usually banded in young, becoming weakly banded to plain in adults 13
12a	Dorsal surface sparsely to densely covered with denticles, without a distinct denticle band
12b	Dorsal surface densely covered with denticles within a
	distinct denticle band Pateobatis
13a	Dorsal disc plain or with dark speckles, white or
	black spots
13b	Dorsal disc with black spots, rings (spots and/or rings may coalesce in large adults to form reticulations)

### GENUS Bathytoshia Whitley 1933

Enormous stingrays (adults to 3 m DW) with disc rhomboidal, dorsal surface smooth or rough; tail usually with 1 stinging spine, dorsal skin fold low or absent, ventral fold long or short; median thorns absent, or in row along disc onto tail; tail spiny, adults with bucklers and tubercles on dorsal midline; body white ventrally and sometimes with dark margins, plain dorsally. Three species, 2 in WIO.

#### **KEY TO SPECIES**

### Bathytoshia brevicaudata (Hutton 1875)

#### Short-tail stingray

PLATE 69

# *Trygon brevicaudata* Hutton 1875, Vol. 16: 317 (Otago Harbour, Dunedin, New Zealand).

Trygon brevicaudatus: Russell 1996; Fricke 1999.

Trygon schreineri Gilchrist 1913: 33, Fig. (False Bay, South Africa).

Dasyatis brevicaudata: Wallace 1967\*; SSF No. 30.1\*.

Bathytoshia brevicaudata: Last et al. 2016\*.

Disc slightly wider than long; snout broadly triangular; trunk thick. Internasal flap broad, short; posterior edge fringed. Tail shorter than disc length, dorsal skin fold rudimentary, ventral skin fold long, low. Dorsal surface of disc smooth; tail with scattered small denticles, densest towards tip, and median row of large thorns before stinging spine (juveniles sometimes with only 1 stout, oval-shaped thorn immediately before spine). Tooth count 26–45/27–50; oral papillae 5–7.

Body greyish brown dorsally, but blackish above eyes, and with paired row of white pores directed anterolaterally on pectoral-fin bases; whitish ventrally, with greyish margins; tail blackish towards tip. Attains 3 m DW, 4.3+ m TL.



Bathytoshia brevicaudata, 77 cm DW (South Africa). Source: SSF

**DISTRIBUTION** Southeastern Atlantic and southern Indo-Pacific. WIO: South Africa (False Bay) to Mozambique and Mauritius.

**REMARKS** Possibly the largest stingray; common to abundant but not well known. Size at birth ~36 cm DW. Occurs in temperate waters, primarily offshore, on outer shelves, uppermost slopes and around offshore islands, but also inshore, near rocky reefs, on open sandy bottoms and in bays and estuaries. Midwater aggregations observed near New Zealand. Feeds on benthic invertebrates (crabs, bivalves) and small bony fishes; preyed on by orca whales and large sharks. Taken as a bycatch but usually discarded.

### Bathytoshia lata (Garman 1880)

#### Thorntail stingray

Trygon lata Garman 1880: 170 (Hawaiian Is.).

*Dasyatis thetidis* Ogilby *in* Waite 1899: 46 (New South Wales, Australia); Wallace 1967\*; SSF No. 30.4\*; Compagno *et al.* 1989\*; Fricke 1999; Fricke *et al.* 2009.

PLATE 69

*Dasybatis agulhensis* Barnard 1925: 78 (Agulhas Bank, South Africa). *Dasyatis lubricus* Smith 1957: 429, Pl. 15 (Algoa Bay, South Africa). *Bathytoshia lata*: Last *et al.* 2016\*.

Disc slightly wider than long; anterior margins nearly straight to weakly undulate, lateral apices narrowly rounded, and posterior margins nearly straight to weakly convex. Snout broadly triangular, tip slightly protruding. Eyes small. Internasal flap broad, short, and posterior edge fringed. Tail longer than disc length, broad-based and tapering; dorsal skin fold minute, and ventral skin fold long, low; usually 1 stinging spine. Pelvic fins small, broadly rounded. Dorsal surface of disc granular, with sparse enlarged pointed denticles centrally, from nape to stinging spine; tail uniformly very thorny beyond stinging spine in adults. Juveniles mostly smooth, with few thorns along midline from snout to base of tail. Tooth count 25–43/29–48; oral papillae 3–5.

Body dark grey to olive-brown dorsally; whitish ventrally. Attains 2 m DW, at least 4 m TL.





**DISTRIBUTION** Indo-Pacific. WIO: South Africa (Eastern Cape) to Mozambique and Réunion.

**REMARKS** Poorly known. Most common inshore, demersal, on soft bottoms, around reefs, and in estuaries and lagoons, but also well offshore, to ~440 m deep. Feeds on invertebrates (crabs, mantis shrimp, bivalves, polychaetes) and fishes (conger eels). Occasionally caught as commercial bycatch and by recreational anglers.

### GENUS Brevitrygon

Last, Naylor & Manjaji-Matsumoto 2016

Disc small, oval to suboval. Tail short with stinging spine close to tail base, dorsal and ventral skin folds usually absent or with rudimentary ventral skin fold; median disc thorns weak, larger if present on tail, scapular thorns absent. Body white ventrally with dark margin, dorsal surface plain; posterior tail with or without pale lateral stripe. Four species, 2 in WIO.

#### **KEY TO SPECIES**

### Brevitrygon imbricata (Bloch & Schneider 1801)

Bengal whipray

PLATE 70

Raja imbricata Bloch & Schneider 1801: 366, 553 (Tharangambadi, India). Raja obtusa Ehrenberg in Klunzinger 1871: 680 (Red Sea). Dasyatis imbricata: Dor 1984.

*Himantura imbricata*: Compagno & Roberts 1982; Randall 1995\*; Manilo & Bogorodsky 2003.

Brevitrygon imbricata: Last et al. 2016\*.

Disc oval, about as wide as long; pectoral-fin anterior margins deeply concave, lateral apices evenly rounded, and posterior margins weakly convex. Snout with broad apical lobe; distance from preorbital snout to maximum disc width 42–50% DW. Eyes smaller than spiracles. Tail stout, shorter than disc, with rudimentary ventral skin fold. Dorsal disc with band of denticles, largest denticles medially and decreasing in size laterally; row of enlarged but low to flattened spear-shaped thorns, confined to midline above head region and on tail from base to stinging spine. Oral papillae 2.

Disc uniformly dark brownish dorsally, with paler margins; whitish ventrally, with darker margins. Attains 30 cm DW, 65 cm TL.

**DISTRIBUTION** Indian Ocean (patchy, poorly defined). WIO: Red Sea, Pakistan, India, Sri Lanka and Mauritius.

**REMARKS** Small-sized; poorly known. Often confused with *H. walga.* Size at birth ~10 cm DW. Found inshore; demersal. Feeds on benthic invertebrates. Caught in large numbers by demersal trawlers; used for human consumption.

### Brevitrygon walga (Müller & Henle 1841)

Scaly whipray

PLATE 70

*Trygon walga* Müller & Henle 1841: 159 (Red Sea). *Trygon nuda* Günther 1870: 476 (Indian Seas). *Dasybatis uylenburgi* Giltay 1933: 13 (East Indies). *Brevitrygon walga*: Last *et al.* 2016\*.

Disc oval, about as wide as long; pectoral-fin anterior margins broadly concave, lateral apices evenly rounded, and posterior margins weakly convex. Snout with broad apical lobe; distance from preorbital snout to maximum disc width 40–43% DW. Eyes smaller than spiracles. Tail stout, shorter than disc, with rudimentary ventral skin fold. Dorsal disc with band of denticles, mid-scapular denticle small but largest, narrow heart-shaped, largest denticles medially and decreasing in size laterally; row of enlarged but low to flattened spear-shaped thorns, confined to midline above head region and on tail from base to stinging spine. Oral papillae 2.

Disc uniformly dark brownish dorsally, with paler margins; whitish ventrally, with darker margins. Attains 30 cm DW, 70 cm TL.

**DISTRIBUTION** WIO: Red Sea, Persian/Arabian Gulf, Pakistan, India.

**REMARKS** Small-sized; poorly known. Often confused with *Brevitrygon imbricata*. Size at birth ~7–10 cm DW. Found inshore; demersal. Feeds on benthic invertebrates. IUCN Red List conservation status Near Threatened.

### GENUS Dasyatis Rafinesque 1810

Anterior margin of disc more or less angular, never convex or broadly rounded; dorsal surface of disc and tail relatively smooth, or with numerous small or large denticles. Tail usually with 1 stinging spine; tail base somewhat broad, tail not whiplike beyond stinging spine; ventral skin fold low (never close to twice height of tail at its deepest section) and not normally extending to tail tip. Teeth rhomboidal, thin-crowned. Body pale to whitish ventrally. Five species, 1 in WIO.

### Dasyatis chrysonota (Smith 1828)

Blue marbled stingray

PLATE 70

*Trygon chrysonota* Smith 1828: 2 (off Gamtoos River, Eastern Cape, South Africa).

Dasyatis pastinaca: Wallace 1967\*; Van der Elst 1981\*; SSF No. 30.3\*; Cowley & Compagno 1993\*.

Dasyatis chrysonota: Heemstra & Heemstra 2004\*; Last et al. 2016\*.

Disc rhomboidal, slightly longer than wide; snout triangular. Tail subequal to disc length; dorsal skin fold indistinct, and ventral skin fold long, low. Dorsal surface of disc smooth, without thorns, but sometimes with small, flattened, conical denticles near front margin. Oral papillae 5.

Disc with unique pattern of conspicuous bright blue finger-like reticulations on golden brown or greenish golden background dorsally; whitish ventrally, but with grey margins in juveniles; tail without banding. Attains 75 cm DW, ~160 cm TL.



*Dasyatis chrysonota*, 46 cm DW, male (South Africa). PC Heemstra © NRF-SAIAB

**DISTRIBUTION** Southern Africa: Angola and Namibia in southeastern Atlantic, to South Africa (KwaZulu-Natal) and possibly Mozambique in WIO.

**REMARKS** Cowley & Compagno (1993) re-evaluated the taxonomic status of the blue stingray, usually misleadingly referred to as Dasyatis pastinaca (Linnaeus 1758), identifying the African forms as separate from the northeastern Atlantic and Mediterranean form; they tentatively ranked it as subspecies D. chrysonota marmorata (Steindachner 1892) (from Senegal), itself separate from D. chrysonota chrysonota (from South Africa, Namibia and Angola). Here, both subspecies are recognised as valid species. Occurs from surf zone to ~110 m deep, usually close inshore, along sandy beaches, near river mouths and sometimes on rocky reefs, and deeper offshore during winter. Litters of 1-7, females produce up to 20 young during their lifetime; size at birth 15-20 cm DW; gestation period ~9 months. Males mature at ~39 cm DW, at ~4 years; females mature at ~51 cm DW, at ~7 years. Feeds mostly on benthic invertebrates; smaller individuals (<30 cm DW) prefer polychaetes and amphipods, and larger individuals consume mostly crabs and mantis shrimp. A bycatch of trawl fisheries and an important sport fish.

### GENUS Himantura Müller & Henle 1837

Disc rounded to oval, rhomboidal or quadrangular. Snout broad or broadly angular to blunt; snout tip with or without small protrusion (apical lobe). Tail tapering and whip-like, without skin folds. Dorsal surface of disc and tail smooth or granular, often with areas of thorns or tubercles in adults. Oral papillae on floor of mouth 0–7. Four species, 3 in WIO, plus 1 other undescribed.

#### **KEY TO SPECIES**





#### Himantura leoparda Manjaji-Matsumoto & Last 2008

#### Leopard whipray

PLATE 70

Himantura leoparda Manjaji-Matsumoto & Last 2008: 293, Figs. 1–5 (Gulf of Carpentaria, Queensland, Australia); Last *et al.* 2016\*.

Dasyatis uarnak (non Gmelin 1789): Wallace 1967\*.

Himantura fava (non Annandale 1909): Compagno & Roberts 1982.

Himantura uarnak (non Gmelin 1789): Van der Elst 1981\*; SSF No. 30.10\*; Compagno *et al.* 1989\*.

Himantura sp. 1: Gloerfelt-Tarp & Kailola 1984.

Himantura undulata (non Bleeker 1852): Last & Stevens 1994.

Disc quadrangular or rhomboidal, slightly wider than long; pectoral-fin anterior margins concave, lateral apices moderately angular to narrowly rounded, and posterior margins convex. Snout broadly triangular, tip acutely pointed; distance from preorbital snout to maximum disc width ~38% DW. Tail long, 2.5–3.7 times DW, usually with 1 stinging spine. By ~33 cm DW, denticles form sub-rectangular band on disc, tapering from interorbital area to base of tail; row of up to 15 prominent heart-shaped thorns on midback (usually 2 of these enlarged); no thorns on midline of tail. Tooth count ~59 in upper jaw; oral papillae 4.

Disc sandy brown background dorsally, covered with leopard-like spots: thick dark brown irregular rings, each ring yellowish inside; juveniles greyish brown with moderately large dark brown polygonal spots; white ventrally; tail of young with dorsolateral spots to stinging spine, and banded dark and pale beyond. Attains 110 cm DW, ~4.7 m TL.



Himantura leoparda, 34 cm DW, immature male (South Africa). Source: SSF



Himantura leoparda (South Africa). © AR Thorpe

**DISTRIBUTION** Indo-Pacific (widespread). WIO: South Africa (KwaZulu-Natal) to Oman, and probably to Sri Lanka.

**REMARKS** Biology not well known due to confusion with other species; often confused with *H. uarnak* and *H. undulata*. Demersal on soft substrates; prefers shallow coastal waters, but found to at least 70 m deep. Probably heavily caught in coastal fisheries. IUCN Red List conservation status Vulnerable.

### Himantura uarnak (Gmelin 1789)

Reticulate whipray

PLATE 70

Raja sephen var. uarnak Forsskål in Niebuhr 1775: 18, viii (Red Sea) [name not available].

Raja uarnak Gmelin 1789: 1509 [Red Sea].

Pastinachus uarnak: Rüppell 1835\*.

Trygon uarnack: Richardson 1846.

Leiobatis (Himantura) uarnak: Bleeker 1877\*.

Trygon narnak: Tillier 1902.

Dasyatis uarnak: SFSA No. 79b\*; Wallace 1967\*.

Himantura uarnak: Day 1865; Van der Elst 1981\*; Compagno & Heemstra 1984\*; SSF No. 30.10\*; Randall 1995\*; Heemstra & Heemstra 2004\*; Last & Stevens 2009\*; Last *et al.* 2016\*.

Disc rhomboidal, about as long as wide; pectoral-fin anterior margins weakly concave, lateral apices narrowly rounded to angular, and posterior margins broadly convex. Snout broadly triangular, with small apical lobe; distance from snout tip to maximum disc width 33–43% DW. Tail extremely long, 2.9–3.5 times DW, usually with 1 stinging spine. Disc mostly smooth, with band of small flat denticles extending from head onto tail; central disc with 1 or 2 enlarged heart-shaped thorns. Tooth count 26–40/27–44; oral papillae 4 or 5 in two transverse rows.

Disc pale brown dorsally, with intricate pattern changing with growth: close-set, fine dark brown polygonal spots in juveniles, and broad dark brown reticulations and/or short narrow irregular bars in adults; spots distributed over brown background, and bars or reticulations separated by narrow yellowish brown lines; pale ventrally, with moderately wide brownish margins; tail banded dark (brown to blackish) and pale (white to yellow) in juveniles, and with reticulations and blackish towards tip in adults. Attains ~200 cm DW, ~6 m TL or more.

**DISTRIBUTION** Indo-Pacific (widespread). WIO: South Africa (Eastern Cape) to Red Sea, eastern Mediterranean Sea, Persian/Arabian Gulf, Pakistan, India, Sri Lanka, Madagascar, Seychelles and Mauritius. **REMARKS** Often confused with *H. undulata* and other reticulate-patterned rays of the '*uarnak*' species-complex. Found inshore to ~50 m deep; common off sandy beaches and in shallow estuaries and lagoons, also over coral reefs. Litters of up to 5; size at birth 21–28 cm DW. Feeds on invertebrates (crabs, shrimp, mantis shrimp, bivalves, gastropods, worms, jellyfish) and bony fishes. Caught as a utilised bycatch of commercial fisheries, and also in artisanal fisheries and by shore anglers. IUCN Red List conservation status Vulnerable globally.

### Himantura undulata (Bleeker 1852)

Honeycomb whipray

PLATE 71

*Trygon undulata* Bleeker 1852: 70 (Jakarta and Semarang, Java, Indonesia). *Trygon russellii (non* Gray 1834): Blyth 1860. *Leiobatis (Himantura) undulatus*: Bleeker 1877. *Trygon favus* Annandale 1909: 25, Pls. 1, 3 (off Odisha, India). *Himantura undulata*: Compagno 1999; Last *et al.* 2016\*.

Disc weakly quadrangular, about as long as wide, and thick; anterior margins of disc strongly concave, lateral apices broadly rounded, and posterior margins broadly convex. Preorbital snout moderately long, with distinct apical lobe; distance from snout tip to maximum disc width 45–46% DW. Tail length ~3 times DW. Teeth subequal in upper and lower jaws, coneshaped with blunt peak and prominent horizontal groove; oral papillae 2–4, broad-based, distally pointed, the middle two connected by low serrated ridge. Disc with well-defined band of denticles extending from head onto tail; dominant pearl-shaped suprascapular denticle, followed by 2 or 3 slightly smaller denticles, and small stellate-based denticles present outside of band (less obvious in adults).

Dorsal surface of disc with large dark brown polygonal spots in young, and broad yellowish brownish undulations and elongated loops or large and hollow polygonal spots in larger specimens and adults; spots or loops separated by slightly narrower or equally wide paler lines; disc white ventrally, often with dusky margin; tail in young laterally banded posterior to stinging spine, dark bands about 3 times wider than pale bands, and tail uniformly pale ventrally; tail in larger specimens and adults with fine dark brown reticulations on pale background. (Dorsal disc colouration appears sexually dimorphic based on the few specimens examined.) Attains 200 cm DW.



*Himantura undulata*, ~ 1 m DW (Malaysia). M Manjaji-Matsumoto © BMRI-UMS

**DISTRIBUTION** Indo-Pacific (possibly widespread but range not well defined). WIO: southern Africa to Red Sea and Lessepsian migrant to eastern Mediterranean Sea.

**REMARKS** Often confused with other species in the *'uarnak'* species-complex. Litters of up to 5; size at birth 21–28 cm DW. Found inshore, to ~50 m deep; common along sandy beaches and in shallow estuaries and lagoons; also found on coral reefs and may enter freshwater. A commercially valuable bycatch in a range of artisanal and commercial fisheries. Feeds on invertebrates (crabs, shrimps, mantis shrimps, bivalves, gastropods, worms, jellyfish) and small bony fishes. IUCN Red List conservation status Endangered.

### GENUS Maculabatis Müller & Henle 1837

Disc suboval to rhomboidal; tail long and whip-like with stinging spine near tail base, dorsal and ventral fin folds absent; 1–3 mid-scapular thorns, or thorns in row on nape; welldeveloped denticle band, rest of disc naked or with patchy denticles; body white ventrally, plain or with spots dorsally, disc may have yellow or dark margins; juveniles usually with distal tail half or fully banded. Six species and 3 undescribed, 5 species in WIO.

#### **KEY TO SPECIES**

1a 1b	Dorsal disc with white spots in young and adults <i>M. gerrardi</i> Dorsal disc plain in young, sometimes with faded dark speckles in adults
2a	Base of tail forward of caudal sting with white spots
2b	Base of tail forward of caudal sting plain
3a	Enlarged mid-scapular denticle preceded by a row of distinct denticles; internasal space 8.4–9.6% disc width <i>M. arabica</i>
3b	Enlarged mid-scapular denticle not preceded by a row of distinct denticles; internasal space narrower, 7.1–8.1% disc width
4a	1 or 2 mid-scapular denticles, small, oval to heart-shaped; tail dark or with weak saddles on dorsal half in young and adults; banded in neonates
4b	5 mid-scapular denticles, large, longer than wide; tail uniform greyish or brownish in adults; banded on dorsal half in young, dark band broadest

### Maculabatis ambigua

Last, Bogorodsky & Alpermann 2016

Baraka's whipray

PLATE 71

Himantura gerrardi (non Gray 1851): Bogorodsky et al. 2014.
Himantura sp. 1: Last et al. 2016\*.
Maculabatis ambigua Last, Bogorodsky & Alpermann 2016: 67, Figs. 1–6 (Red Sea); Last et al. 2016\*.

Disc rhomboidal, slightly wider than long; pectoral-fin anterior margins weakly concave, lateral apices moderately angular, and posterior margins broadly convex. Snout moderately long, tip acutely pointed; preoral snout length 19–21% DW; distance from snout tip to maximum disc width 33–35% DW. Tail very long, 2.3–2.5 times DW. Dermal denticles in distinct band on disc, small, flat, somewhat rounded, extending onto tail; 5 scapular thorns on midback. Tooth count 36 (upper jaw); oral papillae 4.

Disc greyish or brownish dorsally, sometimes dark margin in young; disc white ventrally; tail uniform greyish in adults, weak band on tail posterior to stinging spine in young (but bands fading with size). Attains 90 cm DW, ~290 cm TL. **DISTRIBUTION** WIO: South Africa (Eastern Cape) to Red Sea, Tanzania (Zanzibar), Pakistan, India and Sri Lanka.

**REMARKS** Resembles *M. randalli*, also found in the WIO. Biology unknown.

#### Maculabatis arabica Manjaji-Matsumoto & Last 2016

Pakistan whipray

PLATE 71

Himantura sp. C: Manjaji 2004. Himantura sp.: Henderson *et al.* 2016. Himantura sp. 2: Last *et al.* 2016. Maculabatis sp. 2: Last *et al.* 2016. Maculabatis arabica Manjaji-Matsumoto & Last 2016: 337, Figs. 1–6 (west of Turshian Creek, Pakistan); Last *et al.* 2016\*.

Disc rhomboidal, slightly wider than long; pectoral-fin anterior margins weakly concave, lateral apices moderately angular, and posterior margins broadly convex. Snout moderately long, tip acutely pointed; preoral snout length 21% DW; distance from snout tip to maximum disc width 35% DW. Tail very long, 2.2–2.6 times DW. Dermal denticles in distinct band on disc, small, flat, somewhat rounded, extending onto tail; 1–3 scapular thorns on midback, preceded by row of up to 10 smaller enlarged denticles. Oral papillae 2–4.

Disc pale brown to greenish dorsally, paler disc margin and denticle band; disc white ventrally; tail uniform pale brownish in adults, band on dorsal half of tail posterior to stinging spine in young (dark bands broader). Attains 61 cm DW, >200 cm TL.

**DISTRIBUTION** Northwestern Indian Ocean: Arabian Sea (off Pakistan and western India).

**REMARKS** Attains similar size as *M. randalli*, also found in the WIO, but has a narrower internasal space. Probably feeds on small invertebrates.

### Maculabatis bineeshi Manjaji-Matsumoto & Last 2016

#### Short-tail whipray

PLATE 71

Dasyatis uarnak (non Gmelin 1789): Jones & Kumaran 1970, Fig. 16, two specimens 425 and 480 mm DW (compiled distribution partly in error). ?*Himantura gerrardi (non* Gray 1851): Ishihara *et al.* 1998, 44, Pl. 3,

brief description of two specimens, MTUF 30005.

Himantura sp. D: Manjaji 2004.

Himantura sp. 3: Last et al. 2016.

Maculabatis sp. 3: Last et al. 2016.

Maculabatis bineeshi Manjaji-Matsumoto & Last 2016: 344, Figs. 7–13 (west of Turshian Creek, Pakistan).

Maculabatis bineeshi: Last et al. 2016\*.

Disc rhomboidal, slightly wider than long; pectoral-fin anterior margins weakly concave, lateral apices moderately angular, and posterior margins broadly convex. Snout moderately long, tip acutely pointed; preoral snout length 21% DW; distance from snout tip to maximum disc width 26% DW. Tail long, 2.3–2.6 times DW. Dermal denticles in distinct band on disc, small, flat, somewhat rounded, extending onto tail; 1–3 scapular thorns on midback. Oral papillae 4.

Disc pale brown; white ventrally, disc and pelvic fins with broad yellowish margins; tail dark, weak band on dorsal half of tail posterior to stinging spine. Attains 42 cm DW.

**DISTRIBUTION** Northern Indian Ocean: Persian/Arabian Gulf, Arabian Sea (off Pakistan) and Bay of Bengal (Odisha, India).

**REMARKS** The secondary denticle band is well developed by 24 cm DW. Probably feeds on small invertebrates.

### Maculabatis gerrardi (Gray 1851)

Whitespotted whipray

Trygon gerrardi Gray 1851: 116 (India). Leiobatis (Himantura) gerrardi Bleeker 1877: Fig. 1, Pl. 559. Trygon alcockii Annandale 1909: 27, Fig. 3 (Puri, Odisha, India). Trygon uarnak (non Gmelin 1789): Anonymous 1955\*. Himantura gerrardi: Compagno & Heemstra 1984; SSF No. 30.9\*; Randall 1995\*; Manilo & Bogorodsky 2003; Bonfil & Abdallah 2004;

Heemstra & Heemstra 2004; Bogorodsky *et al.* 2014. *Maculabatis gerrardi*: Last *et al.* 2016\*.

Disc rhomboidal, slightly wider than long; pectoral-fin anterior margins weakly concave, lateral apices moderately angular, and posterior margins broadly convex. Snout moderately long, tip acutely pointed; preoral snout length 19–23% DW; distance from snout tip to maximum disc width 33–43% DW. Tail very long, 2.7–3.4 times DW. Dermal denticles in distinct band on disc, small, flat, somewhat rounded, extending onto tail; a few small scapular thorns on midback. Tooth count 22–26/25–30; oral papillae 2–5.

Disc greyish brown to pale olive-brown dorsally, sparsely spotted or with white spots or blotches scattered over entire surface or confined to rear half of disc; spots extending along sides of tail to stinging spine; disc whitish ventrally, sometimes with narrow brown margins posteriorly; tail with pale and dark brown bands beyond stinging spine in both young and adults (but bands fading with size). Attains 100 cm DW, >200 cm TL.

606 COASTAL FISHES OF THE WESTERN INDIAN OCEAN | VOLUME 1

PLATE 72


Maculabatis gerrardi, 19 cm DW, 60 cm TL (South Africa). Source: SSF

**DISTRIBUTION** Indo-Pacific (widespread). WIO: South Africa (Eastern Cape) to Red Sea, Tanzania (Zanzibar), Pakistan, India and Sri Lanka.

**REMARKS** Often confused with *H. uarnak*. Size at birth 18–21 cm DW. Males mature at 46–48 cm DW; females mature by ~64 cm DW. Common, demersal, in coastal regions including estuaries, to at least 60 m deep. Feeds on invertebrates (bivalves, crustaceans) and small fishes. Often hooked by shore anglers. IUCN Red List conservation status Endangered.

#### Maculabatis randalli

(Last, Manjaji-Matsumoto & Moore 2012)

Arabian banded whipray

PLATE 72

Himantura randalli Last, Manjaji-Matsumoto & Moore 2012: 21, Figs. 1–6 (Kuwait City fish market, Persian/Arabian Gulf). Trygon gerrardi (non Gray 1851): Blegvad & Løppenthin 1944. Himantura gerrardi (non Gray 1851): Randall 1995\*. Himantura sp. B: Manjaji 2004. Maculabatis randalli: Last et al. 2016\*.

Disc rhomboidal, slightly broader than long; anterior margins almost straight to weakly concave, lateral apices rounded, and posterior margins broadly convex. Snout moderately long, preoral length 1.5–1.6 times interorbital width; apical lobe weak. Tail 2.2–2.4 times disc width, slender and tapering gently towards stinging spine and less so beyond to tail tip; tail behind stinging spine subcircular with deep longitudinal ventral groove and prominent midlateral ridge in juveniles, and weakly depressed in adults. Denticle band covering entire dorsal surface of tail in adults; small, broadly heart-shaped to seed-shaped denticles on midback. Tooth rows ~34/40; oral papillae 2. Body uniformly pale brown dorsally, occasionally with dark flecks (specimens <25 cm DW), and disc margins sometimes paler; disc whitish ventrally; tail dark dorsally and sharply demarcated from paler surface ventrally (in adults), or with conspicuous white saddles and almost uniformly dark distally (in neonates and juveniles). Attains 54 cm DW (males) to 62 cm DW (females).

#### DISTRIBUTION WIO: Persian/Arabian Gulf.

**REMARKS** So far recorded only from the Persian/Arabian Gulf, where it is possibly endemic. Found on soft substrates, to ~60 m deep. Presumably feeds on invertebrates (particularly sergestid shrimp *Acetes*).

#### GENUS Megatrygon

Last, Naylor & Manjaji-Matsumoto 2016

Disc rhomboidal. Tail short, broad at base, tapering abruptly, and whip-like beyond stinging spine; dorsal skin fold indistinct, and ventral skin fold long, low. Denticle band absent. One species.

#### Megatrygon microps (Annandale 1908)

Small-eye stingray

PLATE 72

*Trygon microps* Annandale 1908: 393, Pl. 27 (off Chittagong coast, India, Bay of Bengal).

Dasyatis microps: Pierce et al. 2008\*; Last & Stevens 2009\*; Bineesh et al. 2014. Megatrygon microps: Last et al. 2016\*.

Disc broad and angular; pectoral-fin anterior margins almost straight, tips angular; snout weakly rounded. Eyes small; spiracles large. Tail slightly shorter than disc width; usually 1 stout stinging spine. Pelvic fins relatively large. Dorsal surface covered with minute stellate denticles, larger denticles on snout tip and between eyes and spiracles, and slightly larger denticles covering base and sides of tail to stinging spine. Oral papillae 5.

Body brown to reddish or pinkish brown dorsally, darkening towards tail tip; row of white pores on either side of snout near anterior margin; large pale spots near eyes and spiracles, and in loose rows surrounding centre of disc, and distinctive paired row on outer pectoral fins; row of small white spots laterally at base of tail; tail blackish beyond stinging spine; disc whitish ventrally, but margins and tail dusky. Attains 220 cm DW, 320 cm TL.



*Megatrygon microps*, ~2 m DW (S Mozambique).

**DISTRIBUTION** Indo-Pacific (patchy). WIO: Mozambique, southern India and Maldives.

**REMARKS** Large-sized stingray. Litters of 1; size at birth 31–33 cm DW. Depth range and habitat poorly known; inhabits shallow coastal areas, reefs, estuaries and river mouths, and probably deeper waters; assumed to be predominantly demersal, but also semipelagic. Probably an occasional bycatch throughout its range.

# GENUS Neotrygon Castelnau 1873

Orbital region with dark mask-shaped pattern. Thorn-like denticles confined to single variable-length row on midline of disc. Tail short, with generally 2 stinging spines, relatively broad-based and slender beyond stinging spines, and with short dorsal skin fold and long low ventral skin fold. Ten species, 1 in WIO. *Neotrygon indica* Pavan-Kumar *et al.* 2018, from the Gulf of Mannar (species based on DNA and spot colour pattern), has not been examined or included.

#### Neotrygon caeruleopunctata

Last, White & Séret 2016

Blue-spotted maskray

PLATE 72

*Dasyatis kuhlii* (non Müller & Henle 1841): Compagno & Heemstra 1984\*; SSF No. 30.2\*; Last & Stevens 2009\*.

*Neotrygon caeruleopunctata* Last, White & Séret 2016: 546, Figs. 5c, 6c, 7c, 10 (Kedonganan fish market, Bali, Indonesia); Last *et al.* 2016\*.

Disc rhomboidal, wider than long, and trunk thick; pectoralfin apices angular. Snout blunt, preoral length 13–15% DW. Pelvic fins moderately large. Tail subequal to or slightly longer than disc, broad-based, with usually 2 stinging spines, and short dorsal skin fold and long low ventral skin fold. Short row of posteriorly directed, flattened thorns in scapular region (juveniles often lacking thorns); tail smooth. Tooth count 25–32 in each jaw; oral papillae 2.

Disc greyish to reddish brown dorsally, with prominent blue ocelli with brighter blue centres, and scattered fine black spots; prominent dark transverse bar across orbital region; whitish ventrally, with dusky margins; tail banded black and white beyond stinging spines; skin folds on tail pale with darker outer margins. Attains at least 47 cm DW, 76 cm TL.



*Neotrygon caeruleopunctata*, 36 cm DW, adult (South Africa). Source: SSF

**DISTRIBUTION** Indo-Pacific, but probably a species-complex. WIO: South Africa, Mozambique, Madagascar, Mauritius, Red Sea, India and Sri Lanka.

**REMARKS** The taxonomic status of this species requires investigation, as multiple colour morphs exist which may represent different species. Litters of 1 or 2; size at birth ~16 cm DW. Males mature at ~25 cm DW. Common inshore, typically in deep water, to ~90 m deep, but moves into shallow lagoons or onto reef flats at high tide; demersal, over sandy bottoms near rocky or coral reefs, and sometimes lies buried in sand with only its eyes and tail showing. Feeds on invertebrates (crabs, shrimp) and small fishes. A utilised bycatch in many parts of its range.

# GENUS Pastinachus Rüppell 1829

Disc robust and strongly rhomboidal; distance from cloaca to stinging spine  $>\frac{1}{2}$  DW. Tail broad-based; ventral skin fold long and prominent, its depth up to 2–6 times tail height over fold; no dorsal skin fold. Nuchal thorns 1–3; no enlarged thorns elsewhere on disc or tail. Five species, 2 in WIO.

#### **KEY TO SPECIES**

# the second secon

#### Pastinachus ater (Macleay 1883)

Broad cowtail ray

PLATE 72

*Taeniura atra* Macleay 1883: 598 (New Guinea). *Pastinachus ater*: Last *et al*. 2016\*.

Disc rhomboidal, 1.2–1.3 times wider than long; pectoralfin anterior margins nearly straight or weakly convex, lateral apices rounded to angular, and posterior margins weakly convex. Snout broadly rounded and blunt, often with slightly produced tip. Tail long, up to twice disc length, with deep ventral skin fold not reaching tail tip; usually 1 stinging spine, posteriorly set. Dorsal surface with broad band of fine denticles from near snout tip to tail, excluding disc margins; disc entirely smooth in neonates, and juveniles develop up to 4 circular tubercles at centre, which may become indistinct in adults. Tooth count ~20 in each jaw; oral papillae 5.

Disc dark greyish brown dorsally; whitish ventrally; tail and ventral skin fold black. Attains 200 cm DW, 3 m TL or more.

**DISTRIBUTION** Indo-Pacific (widespread but scattered). WIO: Persian/Arabian Gulf, Oman, Red Sea to South Africa (KwaZulu-Natal), Seychelles and Maldives.

**REMARKS** Biology and range poorly known. Demersal on continental and insular shelves. Ventures into estuaries and freshwater. Feeds on invertebrates (crustaceans, polychaetes, sipunculids, molluscs) and bony fishes. Caught in various fisheries and used for its skin and meat.

Pastinachus ater, 45 cm DW, female (South Africa). Source: SSF

# Pastinachus sephen (Fabricius 1775)

Cowtail stingray

Raja sephen Fabricius in Niebuhr (ex Forsskål) 1775: 17, viii (Red Sea).
Trigon forskalii Rüppell 1829: 53, Pl. 13 (Red Sea).
Dasyatis sephen: Smith 1957\*, 1961; Wallace 1967\*.
Hypolophus sephen: SSF No. 30.12\*.
Dasybatus (Pastinachus) gruveli Chabanaud 1923: 45 (Gulf of Thailand).
Pastinachus sephen: Bonfil & Abdallah 2004\*; Last et al. 2016\*.

Disc rhomboidal, up to ~1.3 times wider than long; pectoralfin anterior margins nearly straight or weakly convex, lateral apices rounded to angular, and posterior margins weakly convex. Snout broadly rounded and blunt, often with slightly produced tip. Tail long, up to twice disc length, with deep ventral skin fold not reaching tail tip; usually 1 stinging spine, posteriorly set. Dorsal surface with broad band of fine denticles from near snout tip to tail, excluding disc margins; disc entirely smooth in neonates, and juveniles develop up to 4 circular tubercles at centre, which may become indistinct in adults. Tooth count ~20 in each jaw; oral papillae 5.

Disc greyish brown to almost black dorsally; whitish ventrally, but with dark margins on disc and pelvic fins in young; tail and ventral skin fold dark brown or black. Attains 180 cm DW, 3 m TL or more.

PLATE 73

**DISTRIBUTION** Northwestern Indian Ocean. WIO: Persian/ Arabian Gulf, Red Sea to Pakistan.

**REMARKS** Biology and range poorly known; two distinct species (thick-fold and thin-fold tail forms) have been confused. Size at birth ~18 cm DW. Prefers shallower water, but found to ~60 m deep, on reef flats, in estuaries and intertidal lagoons, and far up rivers in freshwater. Feeds on invertebrates (crustaceans, polychaetes, sipunculids, molluscs) and bony fishes. Caught in various fisheries and used for its skin and meat. IUCN Red List conservation status Near Threatened.

# GENUS Pateobatis

#### Last, Naylor & Manjaji-Matsumoto 2016

Disc suboval to rhomboidal. Tail short, to very long and whip-like, with 1–3 stinging spines close to tail base, dorsal and ventral skin folds absent; 1–3 heart- or pearl-shaped mid-scapular thorns, or thorns in row on nape. Dorsal body plain, white ventrally, often with dark margin; tail plain. Five species, 3 in WIO.

#### **KEY TO SPECIES**

1a	Disc subcircular; snout produced and narrowly triangular <i>P. bleekeri</i>
1b	Disc rhomboidal; snout broadly triangular or convex
2a	Snout broadly convex; tail very long, 2–2.6 times disc width; dorsal surface of disc with small denticles but no enlarged thorns along midline of disc and tail base
2b	Snout broadly triangular; tail slightly longer than disc width; 1 or 3 rows of enlarged upright thorns along midline of disc and tail base, to stinging spine

#### Pateobatis bleekeri (Blyth 1860)

Bleeker's whipray

PLATE 73

*Trygon bleekeri* Blyth 1860: 41 (Calcutta fish market, India). *Pateobatis bleekeri*: Last *et al*. 2016\*.

Disc subcircular, slightly wider than long; pectoral-fin anterior margins straight, lateral apices broadly rounded, and posterior margins broadly convex. Snout produced, preoral snout length ~1/3 disc length, or twice interorbital width. Tail whip-like, ~3 times disc length. Adults with sharply defined band of

denticles extending from snout to base of tail, and 1 or 2 larger pearl-like thorns on midback. Oral papillae 2.

Body usually uniformly greyish or brownish; young often with small dark blotches on dorsal disc, more conspicuous on central disc and head, fading towards pectoral-fin margins; creamy white ventrally. Attains 105 cm DW, ~4 m TL.

**DISTRIBUTION** Indo-Pacific. WIO: Pakistan, India and Sri Lanka.

**REMARKS** The Indian record is based on a colour photograph of a juvenile specimen from "India" identified as *H. bleekeri* in FishBase (Froese & Pauly 2014). Found inshore, to at least 30 m deep, over soft bottoms, and also enters estuaries. Feeds on invertebrates (crustaceans, molluscs, polychaetes) and small fishes. Probably taken in moderate quantities in demersal fisheries. IUCN Red List conservation status Endangered.

#### Pateobatis fai (Jordan & Seale 1906)

#### Pink whipray

PLATE 73

Himantura fai Jordan & Seale 1906: 184, Fig. 2 (Apia, Upolu I., Samoa); Last & Compagno 1999; Bonfil & Abdallah 2004\*; Last & Stevens 2009\*.

*Trygon purpurea* Smith (ex Müller & Henle) 1841: 160, Pl. 52 [South Africa].

*Trygon liocephalus* Klunzinger 1871: 678 (Al-Qusayr, Egypt, Red Sea). *Dasyatis purpureus*: Wallace 1967\*. *Himantura* sp.: SSF No. 30.11\*.

Pateobatis fai: Last et al. 2016\*.

Disc rhomboidal, trunk thick; pectoral-fin anterior margins broadly convex, lateral apices moderately angular, and posterior margins broadly convex. Snout broadly convex with feeble tip; preoral snout length 2.3–3 in mouth width; distance from snout tip to maximum disc width 32–39% DW. Tail narrow, very long (2–2.6 times DW), whip-like beyond stinging spine; usually 1 serrated stinging spine. Dorsal surface, from centre of disc to entire tail, covered with small rounded denticles; few small sharp thorns along midline, becoming densest on tail; juveniles mostly smooth or with sparse covering of small, flat, heart-shaped denticles from interorbital area to tail. Tooth count 20–22/28–31; oral papillae 4.

Disc uniformly pale brown to greyish dorsally, with pinkish margins and darker pelvic fins; sometimes with small white blotch in front of eyes and spiracles; pale ventrally; tail dark grey to black beyond stinging spine. Attains 184 cm DW, ~5 m TL (possibly more).



Pateobatis fai, 77 cm DW, female (South Africa). Source: SSF

**DISTRIBUTION** Indo-Pacific (range poorly defined). WIO: South Africa, Egypt (Suez Canal), Pakistan, India and Maldives.

**REMARKS** Often confused with *P. jenkinsii*. Current WIO records of *P. fai* are based on photographs alone. Occurs in intertidal zone to ~200 m deep (but usually above 70 m); prefers soft sandy bottoms. Aggregations (of sometimes stacked individuals) have been observed. Feeds on small benthic invertebrates. A bycatch of commercial fisheries, and sold as food.

#### Pateobatis jenkinsii (Annandale 1909)

Jenkins whipray

PLATE 73

*Trygon jenkinsii* Annandale 1909: 28, Fig. 4 (Ganjam coast, Odisha, India). *Dasyatus jenkinsii*: Morrow 1954.

Himantura draco Compagno & Heemstra 1984: 6, Figs. 1–8 (off Durban, KwaZulu-Natal, South Africa); SSF No. 30.8\*.

*Himantura jenkinsii*: Randall 1995\*; Heemstra & Heemstra 2004; Last & Stevens 2009\*.

Pateobatis jenkinsii: Last et al. 2016\*.

Disc rhomboidal, slightly wider than long, trunk thick; pectoral-fin anterior margins straight, lateral apices broadly rounded, and posterior margins convex. Snout broadly triangular, tip feebly pointed; distance from preorbital snout to maximum disc width 31–37% DW. Pelvic fins small. Tail slightly longer than disc width, with up to 3 stinging spines. Conspicuous band of imbricated, flat, narrow, heart-shaped denticles on head and scapular region, and 1 or 3 irregular rows of enlarged wedge-shaped thorns on midline from centre of disc to stinging spine; flat and spinulose denticles interspersed on all surfaces of tail posterior to stinging spine; small granulations elsewhere on disc. Tooth count 28/31; oral papillae 2–4.

Dorsal surface of disc uniformly yellowish brown, but some specimens with small black spots along posterior margins; disc and margins white ventrally; tail greyish beyond stinging spine. Attains at least 104 cm DW, ~200 cm TL.



Pateobatis jenkinsii, 56 cm DW, immature male holotype of Himantura draco (South Africa). Source: SSF

**DISTRIBUTION** Indo-Pacific (patchy). WIO: South Africa (KwaZulu-Natal), Mozambique, northwestern Madagascar, Socotra I., Oman, India and Sri Lanka.

**REMARKS** Likely misidentified as *H. fai* in some literature. Size at birth 20–27 cm DW; males mature at ~70 cm DW. Found inshore to ~50 m deep, typically demersal on sandy substrates. Feeds on crustaceans and small fishes. Caught in various inshore fisheries; sold as food and valued for its skin.

# GENUS **Pteroplatytrygon** Fowler 1910

Disc trapezoidal, ~<sup>1</sup>/<sub>3</sub> wider than long; pectoral-fin anterior margins convex, posterior margins weakly convex to straight. Snout short, with small rounded tip. Eyes almost lateral, not protruding above disc. Internasal flap laterally expanded, exceeding edges of nostrils. Teeth with 1 or 2 pointed cusps. Tail with well-developed ventral skin fold. Dorsal and ventral surfaces of disc both dark. One species.

### Pteroplatytrygon violacea (Bonaparte 1832)

Pelagic stingray

*Trygon violacea* Bonaparte 1832, fasc. 1, punt. 6, Pl. 155 (Italy, Mediterranean Sea).

Trygon purpurea: Müller & Henle 1841\*.

Dasyabatus purpureus: Barnard 1925\*.

Pteroplatytrygon violacea: McEachran & Fechhelm 1998; Compagno 1999; Fricke et al. 2009; Last et al. 2016\*.

PLATE 73

Anterior margins of disc evenly rounded. Snout broad, preorbital snout length 11–17% DW. Eyes small, not protruding above disc. Tail broad-based and long, can exceed twice disc length when undamaged; 1 or 2 stinging spines; ventral skin fold long and low. Denticles on dorsal surface randomly and widely spaced; interorbital area with minute denticles; short thorns along midline of disc and tail; males sometimes with denticles on ventral surface of disc. Tooth count 25–39/25–38; oral papillae 5–15.

Disc dark purplish to bluish black or sometimes dark bluegreen dorsally; slightly paler brownish black to dark grey ventrally; tail and ventral skin fold black. Attains 80 cm DW (usually <60 cm DW), 160 cm TL.



Pteroplatytrygon violacea, 35 cm DW (Cook Is.). Source: SSF

**DISTRIBUTION** Almost circumglobal in tropical to warmtemperate seas, including Mediterranean Sea. WIO (scattered records): Kenya, Mozambique, South Africa, Madagascar and Réunion. **REMARKS** Males mature at 35–40 cm DW, and females at 40–50 cm DW. Litters of 4–9; size at birth 15–25 cm DW. Perhaps the only truly pelagic stingray, as it occurs in the epipelagic zone of open ocean as well as in deep bays near land; usually hooked above 100 m over deeper water, but recorded to 238 m deep. Feeds on invertebrates (krill, amphipods, shrimp, jellyfish, squid, octopus) and pelagic fishes. A common bycatch of pelagic longliners and mostly discarded.

#### GENUS Taeniura Müller & Henle 1837

Disc oval; snout short; tail broad and compressed, with 1 or 2 posteriorly located stinging spines, and low ventral skin fold tapering and reaching tail tip. Skin mostly smooth. One or 2 species, 1 in WIO.

#### Taeniura lymma (Fabricius 1775)

Blue-spotted ribbontail ray

Raja lymma Fabricius in Niebuhr (ex Forsskål) 1775: 17,

viii (Yemen, Red Sea).

Raia lymnia: Bonnaterre 1788\*.

*Raja lymna*: Bloch & Schneider 1801.

*Taeniura lymma*: Wallace 1967\*; SSF No. 30.13\*; Heemstra & Heemstra 2004\*; Last & Stevens 2009\*; Last *et al.* 2016\*.

Disc oval and elongated; anterior margins weakly convex, lateral apices and posterior margins broadly rounded. Snout angular and bluntly rounded, tip not pointed; dorsal midline with low fleshy ridge. Internasal flap narrow, posterior edge fringed. Eyes moderately large, protruding; interorbital space narrow. Tail stout, relatively short (<½ TL); 1 or 2 stinging spines set beyond midlength of tail; deep ventral skin fold extending to tail tip. Skin mostly smooth, but a few rows of widely spaced thorns in nuchal area, and short row of small flat denticles on midback in adults. Tooth count 15–24 in each jaw; oral papillae 2.

Body yellowish brown or greyish brown dorsally, with large bright blue spots on most of disc (excluding margins) and pelvic fins; mostly white ventrally, sometimes with brown disc margins; tail usually with pair of blue lateral stripes (no stripes in southern African specimens) and bluish towards tip. Attains at least 35 cm DW, at least 75 cm TL.

PLATE 74



Taeniura lymma, 17 cm DW (South Africa). Source: SSF

**DISTRIBUTION** Indo-Pacific (widespread). WIO: South Africa (KwaZulu-Natal) to Red Sea, Madagascar, Mauritius, Seychelles, Tanzania (Zanzibar), Arabian Sea, Gulf of Oman, Persian/Arabian Gulf, Pakistan, India, Sri Lanka and Maldives.

**REMARKS** Poorly known although abundant in some areas. Litters of up to 7. Males mature by ~21 cm; females still immature at 24 cm DW. Benthic on shallow coral reefs and nearby sandy areas, to ~30 m deep; moves in large groups onto sandy flats during rising tide to feed on benthic invertebrates (molluscs, worms, shrimps, crabs) and small bony fishes. Commonly taken by artisanal and small commercial fisheries near coral-reef habitats.

#### GENUS **Taeniurops** Garman 1913

Disc circular; tail compressed, with 1 posteriorly positioned stinging spine, and deep ventral skin fold not tapering but reaching tail tip. Two species, 1 in WIO.

#### Taeniurops meyeni (Müller & Henle 1841)

#### Blotched fantail ray

PLATE 74

*Taeniura meyeni* Müller & Henle 1841: 172, Pl. 75 (Mauritius, Mascarenes); Heemstra & Heemstra 2004.

Taeniura melanospilos: Wallace 1967\*; SSF No. 30.14\*.

Taeniurops meyeni: Last & Stevens 2009\*; Last et al. 2016\*.

Disc nearly circular, slightly wider than long, all outer margins evenly rounded. Internasal flap short, very broad, posterior edge with short fringe. Tail relatively short, less than disc width; usually 1 stinging spine; ventral skin fold long and broad, extending to tail tip. Pelvic fins small. Dorsal surface of disc and tail uniformly covered with small stellate denticles in adults, and smooth in young; no true thorns, but a few rows of enlarged denticles along midline of disc and tail. Tooth count 37–46/39–45; oral papillae 5–7.

Body usually dark purplish, bluish or brownish grey dorsally, or pale with scattered irregular dark spots or mottling and dusky specks or streaks; disc whitish ventrally, with darker margins; tail black beyond stinging spine. Attains 180 cm DW, 330 cm TL.



*Taeniurops meyeni*, 144 cm DW, female (South Africa). Source: Smith & Smith 1963

**DISTRIBUTION** Indo-Pacific (widespread). WIO: South Africa (KwaZulu-Natal) to Red Sea, Madagascar, Mauritius, Seychelles, Arabian Sea, Persian/Arabian Gulf, Gulf of Oman, Pakistan, India, Sri Lanka and Maldives.

**REMARKS** Size at birth ~35 cm DW; males mature at 100–110 cm DW. Bottom-dwelling in lagoons, estuaries and on reefs; common close to shore, but recorded to 439 m deep. Feeds on invertebrates (bivalves, crabs, shrimp) and small benthic fishes. Caught throughout its range by line gear and trawls, as a targeted or bycatch species, and also by recreational anglers in southern Africa. IUCN Red List conservation status Vulnerable globally; Least Concern in Maldives.

# GENUS Telatrygon

#### Last, Naylor & Manjaji-Matsumoto 2016

Disc sub-rhomboidal to rhomboidal; dorsal surface of disc smooth, with row of median thorns on nape; snout elongate, acutely angular. Tail long, and filamentous distally; caudal sting close to tail base; tail with thorns absent, and denticles rarely present. Body white ventrally, disc usually with dark edge; dorsal disc and tail plain. Three species and one undescribed; 1 species in WIO.

# Telatrygon crozieri (Blyth 1860)

Indian sharpnose ray

PLATE 74

*Trygon crozieri* Blyth 1860: 45 (Arakan coast, India) *Telatrygon crozieri*: Last *et al.* 2016\*.

Disc rhomboidal, slightly longer than wide; anterior margins concave, posterior margins convex. Snout triangular, elongated; head more than half disc length. Eyes small. Tail slightly longer than disc width; dorsal skin fold indistinct/ rudimentary, and ventral skin fold long, very low. Dorsal surface with median row of up to 12 variably enlarged thorns before stinging spine in adults, but smooth in juveniles. Tooth count 40–55 in each jaw; no oral papillae.

Disc brown dorsally; whitish ventrally, with brown margins and dark brown ventral skin fold on tail. Attains 29 cm DW, 75 cm TL.

**DISTRIBUTION** Indo-Pacific: southern India in WIO, and eastwards.

**REMARKS** Resembles *T. zugei* from western Pacific. Smallsized. Litters of 1–3; size at birth 7–10 cm DW. Males mature at ~18 cm DW, and females at ~19 cm DW. Occurs on continental and insular shelves; benthic, typically in shallow waters <100 m deep, over flat sandy areas and in estuaries. Feeds on small crustaceans (prawns) and fishes. A sizeable component of the elasmobranch bycatch in parts of its range. IUCN Red List conservation status Near Threatened.

# GENUS Urogymnus Müller & Henle 1837

Disc suboval to circular; snout rounded to angular; dorsal surface often covered with large prickly denticles; 1–3 mid-scapular thorns, or thorns absent. Tail short to long, stinging spine on tail absent, or in normal position, or set well back; dorsal and ventral skin folds absent. Ventral body white,

plain dorsally, disc often dark edged, tail plain, not banded. Five species, 3 in WIO.

#### **KEY TO SPECIES**

1a	Snout evenly rounded; tail subequal to disc length, stinging spine on tail absent
1b	Snout pointed; tail longer than disc length, 1 or 2 stinging spines on tail
2a	Tail 1.5–2 times disc length; disc with scattered small white spots dorsally, tail white beyond stinging spine <b>U. granulatus</b>
2b	Tail 1.9–2.4 times disc length; disc plain dorsally, tail blackish beyond stinging spine

# Urogymnus asperrimus (Bloch & Schneider 1801)

Porcupine ray

PLATE 75

*Raja asperrima* Bloch & Schneider 1801: 367 (off Mumbai, India). *Raja africana* Bloch & Schneider 1801: 367 (Guinea, West Africa [?]). *Urogymnus asperrimus*: SSF No. 30.15\*; Bonfil & Abdallah 2004\*;
Last & Stevens 2009\*; Last *et al.* 2016\*.

Disc oval, almost as wide as long, and greatly elevated at centre (convex appearance in lateral view). Snout tip evenly rounded, barely protruding. Pelvic fins small, slender. Tail about equal to disc length; no stinging spine. Dorsal surface of central disc and tail covered with plate-like denticles and very sharp enlarged thorns (especially in adults). Tooth count 48 in each jaw; oral papillae 3–5.

Disc pale to dark grey-brown dorsally, without markings; whitish ventrally; tail tip blackish. Attains 147 cm DW,  $\sim$ 330 cm TL.



Urogymnus asperrimus, 94 cm DW (Kenya). Source: SSF

**DISTRIBUTION** Eastern Mediterranean Sea and Indo-Pacific (scattered); records from eastern Atlantic are of the closely related *U. ukpam.* WIO: South Africa to Red Sea, Madagascar, Seychelles, Arabian Sea, Persian/Arabian Gulf, Gulf of Oman, Pakistan, India, Sri Lanka, Maldives and Chagos.

**REMARKS** Unusual and poorly known; relatively uncommon compared to other sympatric stingrays. Benthic, usually close to shore, over sand, coral gravel, seagrass beds and near reefs, to ~30 m deep. Matures at 90–100 cm DW. Observed aggregating when feeding on benthic invertebrates (sipunculids, polychaetes, crustaceans) and small fishes by ploughing the bottom and expelling sand through its spiracles. Presumably a bycatch of some coastal fisheries. IUCN Red List conservation status Vulnerable.

#### Urogymnus granulatus (Macleay 1883)

Mangrove whipray

PLATE 75

*Trygon granulata* Macleay 1883: 598 (Port Moresby, Papua New Guinea). *Trygon ponapensis* Günther 1910: 493, Pl. 180 (Kabary, Pohnpei, CarolineIs.). *Himantura marginatus (non* Blyth 1860): James 1980.

Himantura granulata: Last & Stevens 1994\*; Bineesh et al. 2014; Bogorodsky et al. 2014\*; Last et al. 2016\*.

Disc oval, slightly longer than wide, and trunk thick; pectoralfin anterior margins straight to weakly concave, lateral apices evenly rounded, and posterior margins broadly convex. Snout tip narrowly pointed; distance from preorbital snout to maximum disc width 42–51% DW. Eyes moderately large, protruding above disc; interorbital area broad. Internasal flap broad, short; posterior edge with fine fringe. Pelvic fins small, slender. Tail moderately long, 1.5–2 times disc length, and with 1 or 2 stinging spines. Dorsal surface of disc smooth in juveniles (<23 cm DW), and main band of denticles in adults with small and large denticles, largest over branchial area and along midline of back and tail as far as stinging spine; ventral surface entirely smooth. Tooth count 40–50/38–50; oral papillae 2–7.

Body brownish or greyish dorsally (often covered in black mucus), with darkened outer margins; small white spots speckled over disc, pelvic fins, and tail before stinging spine; adults pale ventrally, with dark blotches on disc margins, and juveniles uniformly pale or with greyish yellow margins; tail entirely white beyond stinging spine. Attains 141 cm DW, 3.5 m TL.

**DISTRIBUTION** Indo-Pacific (patchy, poorly defined). WIO: Gulf of Aqaba and central Red Sea (Al Lith, Saudi Arabia), Gulf of Aden and Maldives; also Gulf of Mannar (India and Sri Lanka).

**REMARKS** Large-sized; locally rare. Size at birth ~14 cm DW. Prefers shallow inshore habitats, but adults found to at least 85 m deep; demersal, over hard bottoms, rocky areas and corals, broken rocky-sandy substrate, sand flats, and in estuaries and mangroves. Feeds on crustaceans and small fishes. Caught by inshore artisanal and subsistence fisheries and some coastal commercial fisheries. IUCN Red List conservation status Vulnerable.

# Urogymnus polylepis (Bleeker 1852)

Giant freshwater whipray

PLATE 75

*Trygon polylepis* Bleeker 1852: 73 (Jakarta, Java, Indonesia). *Leiobatis (Himantura) polylepis*: Bleeker 1877\*. *Urogymnus laevior* Annandale 1909: 37 (Malabar coast, India). *Trygon fluviatilis (non* Hamilton 1822): Annandale 1910\*. *Himantura chaophraya* Monkolprasit & Roberts 1990: 204,

Fig. 1 (Chao Phraya River, upriver from Ayutthaya, Thailand). Himantura polylepis: Last & Manjaji-Matsumoto 2008. Urogymnus polylepis: Last et al. 2016\*.

Disc subcircular, longer than wide; pectoral-fin anterior margins truncate, lateral apices broadly rounded, and posterior margins broadly convex. Snout long, 41–50% DW, tip with large triangular apical lobe. Tail long, slender, 1.9–2.4 times DW. Band of denticles along midtrunk moderately sparse, without well-defined margin; mid-scapular denticles small or inconspicuous; tail only sparsely covered with denticles, but denser in large adults. Tooth count 22–23/19–23; oral papillae 4–7.

Disc uniformly pale brown or greyish dorsally; white ventrally, with broad dark brown marginal band, inner part of band with dark brown blotches; tail darker brown to blackish dorsally, and tail base white ventrally, becoming blackish beyond stinging spine. Attains 200 cm DW, at least 4 m TL.



Urogymnus polylepis. Source: Bleeker 1877

**DISTRIBUTION** Widespread in large rivers of Southeast Asia (India to eastern Indonesia) and also Malabar coast, India, in WIO.

**REMARKS** The description of *Himantura chaophraya* from Thailand failed to take into account the description of *H. polylepis* (Bleeker 1852). Last & Manjaji-Matsumoto (2008) compared the holotype of *H. polylepis* (from Java, Indonesia) with material from the Chao Phraya River (Thailand), Sabah (Malaysia) and India, and confirmed *polylepis* as the senior name. Even so, the taxonomic status of subpopulations in different drainages requires further research. Litters of up to 4; size at birth ~30 cm DW. Males mature at 90–130 cm DW, and females at ~80 cm DW. Found mostly in freshwaters of large rivers with muddy or sandy bottom, and less often in estuarine waters. Feeds mostly on freshwater and estuarine crustaceans and fishes. Caught incidentally in artisanal fisheries. IUCN Red List conservation status Endangered; subpopulation in Thailand assessed Critically Endangered.

# FAMILY GYMNURIDAE

#### Butterfly rays

Brett A Human, Leandro Yokota and Marcelo R de Carvalho

Small- to large-sized (to ~260 cm DW) with extremely flattened body; head depressed and not elevated above disc, anterior margin not indented; snout short and obtuse,

sometimes with small knob at tip. Pectoral-fin origins anterior to nostrils and confluent with head and body, forming greatly laterally expanded rhomboidal or diamond-shaped disc (disc width 1.5-2.2 disc length); pectoral-fin tips narrowly rounded, rear margins straight or convex. Dorsal surface smooth, without thorns. Five pairs of short gill slits on underside; no electric organs. Eyes small, widely separated; spiracles with or without posteriorly projecting dermal tentacle on inner rear margin. Internasal flap broad, short and reaching mouth; posterior edge smooth or weakly fringed. Mouth transverse or slightly arched, with numerous small teeth. Pelvic fins small, angular, not bilobed; claspers short and stout. Tail usually banded, filamentous and flexible, cylindrical or slightly depressed, shorter than disc, and with low dorsal or ventral skin folds variably present or absent; dorsal fin absent or small, low, rounded or angular, its origin just behind pelvic-fin insertions or beyond; 0-2 stinging spines; no caudal fin. Tooth rows 52-130/52-130; spiral valve turns 6-12.

Historically, the family Gymnuridae was separated into two genera based on the presence (Aetoplatea) or absence (Gymnura) of 1 small dorsal fin. However, recent studies have shown that this character varies even within species (Jacobsen & Bennett 2009; Nunes & Piorski 2009; Yokota & Carvalho 2017), hence it cannot be used as a generic character. Aetoplatea is thus a junior synonym of Gymnura. Furthermore, the development of dorsal and ventral skin folds on the tail has been shown to be a sexually dimorphic trait in G. marmorata and G. crebripunctata. Similarly, the presence or absence of stinging spines is inconsistent in some of the species. Although some regional revisions of gymnurids have been conducted recently, a review of all species globally is needed to develop characters that separate them as well as to better understand their distributions and biology. Possibly vary their colour pattern to match the sea bottom. Reproductive mode is viviparity with histotroph, wherein uterine secretions nourish the embryos (by being absorbed through the spiracles and delivered into the pharynx of the young). A common bycatch in commercial fisheries, although the flesh is not highly regarded.

One genus and 12 species recognised pending a revision; 3 or 4 described species and possibly 1 undescribed species in WIO.

#### GENUS Gymnura Van Hasselt 1823

Diagnosis as for family. Found in tropical and warm-temperate seas; 12 species, 3 or 4 in WIO (as *Gymnura zonura* is recorded in the Bay of Bengal, the possibility of it occurring in the WIO was considered). In addition, frequent reference in the literature to a ray similar to *G. micrura* in the WIO may be an undescribed species or misidentifications (and likewise for records of *G. micrura* elsewhere in Indo-Pacific).

#### **KEY TO SPECIES**



#### Gymnura natalensis (Gilchrist & Thompson 1911)

Diamond ray

PLATE 76

*Pteroplatea natalensis* Gilchrist & Thompson 1911: 56 (KwaZulu-Natal, South Africa); Gilchrist & Thompson 1916; Von Bonde & Swart 1923; Barnard 1925.

Pteroplatea micrura: Barnard 1925.

Gymnura natalensis: Smith 1949\*; Wallace 1967\*; SSF No. 30.7\*;

Compagno *et al.* 1989\*; Cliff & Wilson 1994\*; Heemstra & Heemstra 2004\*; Jacobsen & Bennett 2009.

Spiracles with rudimentary dermal tentacle on inner rear margin. No dorsal fin; tail with dorsal and ventral skin folds, and 1 or 2 stinging spines; tail short, post-cloacal length 35–50% disc length (tail usually longer in embryos and small juveniles: post-cloacal length 50–60% disc length). Tooth count 68–93/70–93.

Disc grey to olive-green, often with large dark blotches and smaller yellowish green spots; pale ventrally; tail with pale and dark bands (most prominent in juveniles); has limited ability to change colouration to match substrate. Attains 250 cm DW.

**DISTRIBUTION** Southern Africa: Namibia in southeastern Atlantic, to South Africa and southern Mozambique in WIO.

**REMARKS** Displays marked ontogenetic differences in morphology: disc almost triangular in juveniles, and rhomboidal in adults. Specimens from South Africa identified as G. japonica are misidentified specimens of G. natalensis. Occurs in temperate to warm-temperate waters, inshore, commonly off sandy beaches, to ~75 m deep; usually solitary on seabed, although observed swimming in shoals in midwater. Females enter inshore areas and estuaries in summer to give birth; litters of 2–9; size at birth ~35 cm DW; gestation period 12 months. Males mature at 100-110 cm DW, and females at 145-150 cm DW; females attain larger sizes than males. Diet includes bony fishes, crabs and worms. A common bycatch of prawn trawl fisheries and in anti-shark nets, and regularly taken by shore anglers. Although the spine at the base of the tail normally poses little threat, the authors can attest to the potency of the venom.



Gymnura natalensis, 67 cm DW, female (South Africa). Source: SSF



Gymnura natalensis, 185 cm DW (South Africa). Source: SSF

# Gymnura poecilura (Shaw 1804)

#### Longtail butterfly ray

PLATE 76

*Raja poecilura* Shaw 1804: 291 (Visakhapatnam, India) [based on Russell 1803, Pl. 6].

Pteroplatea micrura: Day 1865, 1889\*; Wood-Mason & Alcock 1891\*; Alcock 1892\*.

*Gymnura poecilura*: James 1966\*; Bianchi 1985\*; Compagno & Randall 1987; Al-Abdessalaam 1995\*; Randall 1995\*; Carpenter *et al.* 1997\*; Morón *et al.* 1998; Compagno & Last 1999\*; Sujatha 2002; Bonfil & Abdallah 2004\*; White & Dharmadi 2007; Jacobsen & Bennett 2009.

*Gymnura hormosensis* Vossoughi & Vosoughi 1999: 304 [validity uncertain].

Gymnura japonica: Sujatha 2002.

Spiracles without dermal tentacle on inner rear margin. No dorsal fin; tail with dorsal and ventral skin folds (dorsal fold more prominent), and no or 1 stinging spine (rarely 2 spines); tail long, post-cloacal length 50–100% disc length, with larger individuals presenting proportionately longer tail.

Disc uniformly olive-green to greyish brown dorsally or peppered with minute darker spots or pale spots; pale ventrally (whitish to brownish or coppery); tail usually with prominent pale and dark bands in repeating pattern of long dark (almost black) band, short white band, short dark band, short white band, and long dark band (total 8–12 dark bands). Attains 100 cm DW.



*Gymnura poecilura*, 89 cm DW, 75 cm TL, female (Red Sea). © SV Bogorodsky

**DISTRIBUTION** Indo-Pacific (widespread but disjunct). WIO: Somalia, Red Sea (common), Oman, Gulf of Oman, Persian/ Arabian Gulf, Pakistan, India and Sri Lanka.

**REMARKS** Embryos and juveniles have almost triangular disc as compared with rhomboidal disc of adults. A new species (*G. hormosensis*) from the Strait of Hormuz (Persian/

Arabian Gulf) was proposed with uncertainty based on minor colouration differences; the illustration provided closely agrees with G. poecilura, particularly the banding pattern on tail, and the few characters provided for it are identical to those for G. poecilura (including no tentacle on spiracles). Therefore, this work follows Yoshigou et al. (2004) in placing G. hormosensis as a junior synonym of G. poecilura. However, it is possible that the specimens from the WIO are of a species similar to *G. poecilura* (i.e. *G. cf. poecilura*). Biology poorly known. Litters of up to 7; size at birth 18-21 cm DW; females mature at ~41 cm DW. Occurs inshore, mostly over sandy and muddy bottoms, with narrow depth range to just ~30 m deep. Diet includes bony fishes, crustaceans and molluscs. A bycatch of commercial trawls and gillnets, and targeted by artisanal and commercial fisheries as the fins are retained for consumption. IUCN Red List conservation status Near Threatened.

# Gymnura tentaculata (Valenciennes 1841)

Tentacled butterfly ray

PLATE 76

*Aetoplatea tentaculata* Valenciennes *in* Müller & Henle 1841: 175 (Red Sea? [possibly Malabar coast of India]); Bianchi 1985;

Compagno & Randall 1987; Kailola 1987\*; Yoshigou *et al.* 2004. *Pteroplatea tentaculata*: Annandale 1909\*.

Gymnura tentaculata: Wallace 1967; Jacobsen & Bennett 2009.

Spiracles with dermal tentacle on inner rear margin. Tail with low dorsal fin; dorsal and ventral skin folds variably present or absent; stinging spine minute or absent; tail (post-cloacal length) 45–60% disc length.

Disc dark olive dorsally, frequently with yellowish green blotches (disc green dorsally, with dark brown spots and reticulations in young); white or marbled ventrally; tail without banding. Attains at least 76 cm DW.

**DISTRIBUTION** Indian Ocean: Red Sea to southern India and to Bay of Bengal; records from elsewhere in northern WIO require confirmation.

**REMARKS** Apparently rare, known from very few specimens. Morphologically very similar *G. australis* which otherwise has distinct tail banding; records from New Guinea in western Pacific are misidentifications of *G. australis*. Biology virtually unknown. Presumably a bycatch of inshore fisheries throughout its range. IUCN Red List conservation status Data Deficient.

#### Gymnura zonura (Bleeker 1852)

Zonetail butterfly ray

PLATE 77

Aetoplatea zonurus Bleeker 1852: 79 (Jakarta, Java, Indonesia).

?Pteroplatea micrura: Annandale 1909.

Pteroplatea zonura: Annandale 1909\*.

Aetoplatea zonura: Compagno & Last 1999\*.

*Gymnura zonurus*: Sujatha 2002.

*Gymnura japonica*: White & Dharmadi 2007.

Gymnura zonura: White & Dharmadi 2007; Jacobsen & Bennett 2009.

Spiracles without dermal tentacle on inner rear margin. Tail usually with low dorsal fin (fin may be absent); ventral skin folds variably present; usually 1 stinging spine (absent in some specimens); tail (post-cloacal length) 40–65% disc length.

Disc medium to dark olive-green dorsally, often presenting speckled with minute darker spots and larger whitish spots; pale ventrally; tail with prominent pale and dark bands in repeating pattern of long dark (almost black) bands and shorter white bands (total 6–10 dark bands). Attains 108 cm DW.

**DISTRIBUTION** Tropical waters of eastern Indian Ocean and western Pacific, and possibly west coast of India in WIO.

**REMARKS** Biology unknown in WIO; elsewhere males mature at 46–50 cm DW, and females are immature at 51 cm DW, but mature by 78 cm DW. Litters of 2–5; size at birth probably 14–28 cm DW. Narrow depth range, to ~40 m deep, over soft substrates. Heavily caught in demersal gillnet fisheries in Indonesia and probably throughout its range. IUCN Red List conservation status Vulnerable.

# FAMILY MYLIOBATIDAE

#### Eagle rays

Marcelo R de Carvalho and Mateus C Soares

Moderate- to large-sized rays (to 9 m TL, 3.3 m DW; most species attain <200 cm DW) with rhomboidal or lozengeshaped disc much wider than long; head projecting beyond disc, snout rounded; pectoral fins meet head below eyes, angular with concave posterior margins. Fleshy lateral ridge supporting snout, either continuous with pectoral-fin origins (*Myliobatis*) or not continuous and situated well below pectoral-fin origins at level of eyes (*Aetomylaeus*). Head relatively narrow to somewhat broad (*Myliobatis*), thick, and elevated from disc. Snout varying from short (*Myliobatis*) to relatively elongate (*Aetomylaeus*), with additional fleshy rostral lobe (resembling duck's bill) or small knob. Eyes lateral, medium to moderately large, anteromedial to spiracles; spiracles elongated, lateral, margins smooth and without papillae, rims not elevated. Mouth narrow to relatively broad, transverse, straight, and without prominent knobs, depressions or lip folds. Nostrils relatively small, usually slightly elongate, only slightly anterior to mouth, connected to mouth by broad oronasal grooves; internasal flap broad, overlapping mouth, straight or slightly undulate, but not notched. Teeth large and blunt, greatly flattened, laterally expanded, plate-like, roughly hexagonal; 6–10 tooth rows in each jaw (sometimes more; lateral teeth may be lacking in adults of *Aetomylaeus vespertilio*). Five pairs of small gill slits on underside of front half of disc. No electric organs.



Aetomylaeus bovinus, 68 cm DW, ventral head with internasal flap margin straight (South Africa).

Pelvic fins slightly longer than wide, not bilobed, projecting past disc, and posterior margins rounded or straight. Tail long (~1.5–3 times disc length), filamentous and whip-like, cylindrical and without lateral skin folds; 1 small dorsal fin, relatively high, angular or rounded-angular, set over or just behind pelvic-fin insertions, and rear tip joined to tail surface; prominent stinging spine close behind dorsal fin in most species (stinging spines in only 1 species of *Aetomylaeus*); no caudal fin. Dorsal surface with minute dermal denticles, or smooth above and below with only a few reduced thorns or tiny denticles on disc midline or around eyes. Body uniformly brown, greenish, grey or yellowish to blackish dorsally, or with spots, small ocelli, faint banding, or mildly reticulated; generally pale to whitish ventrally.

Circumglobal in temperate and tropical waters, inhabiting relatively shallow waters of continental and insular shelves; semipelagic, in schools of a few to hundreds of individuals (*Myliobatis*). The expanded, wing-like pectoral fins of eagle rays permit great propulsion, allowing them to reach high speeds and leap from the water; also found resting on the bottom, raising their body with their pectoral fins. Reproductive mode is viviparity with histotroph (uterine secretions that nourish embryos), with litters of 2–10. Females enter shallow water or estuaries to give birth, observed leaping to facilitate parturition; copulation has been recorded in captivity. Feed on hard-shelled prey items (molluscs and crustaceans), which they crush with pavement-like tooth plates, and also on bony fishes, worms and squid; often use

their fleshy rostral lobe to expose buried prey. Species with a stinging spine are usually harmless to people due to its anterior position at base of tail, but larger species have powerful jaws. A few species are common in public aquariums.

Two genera and 18 species; both genera and 6 species in WIO.

#### **KEY TO GENERA**



#### GENUS Aetomylaeus Garman 1908

Fleshy lateral ridge around snout well beneath pectoral-fin origins, not continuous, at level of eyes; rostral lobe fleshy, long, duck-bill-shaped. Posterior edge of internasal flap relatively straight. Both jaws with 7 rows of plate-like teeth, centre row widest, and 3 rows of diamond-shaped teeth at each side. Stinging spine lacking except in 1 species in WIO. Currently 6 species recognised, 5 in WIO; in addition, a probable undescribed species has been photographed in the Maldives (referred to as *Aetomylaeus vespertilio* in Debelius 1999, and *Aetomylaeus* sp. in Hennemann 2001).

#### **KEY TO SPECIES**

1a	Stinging spines present A. bovinus
1b	No stinging spines2
2a	Dorsal-fin origin just behind (rarely over) pelvic-fin
2b	Dorsal-fin origin just before or over pelvic-fin insertions 4

Continued . . .

#### **KEY TO SPECIES**

3a	Dorsal surface of disc with pattern of broken, transverse, darker stripes anteriorly and darker reticulate pattern posteriorly
3b	Rear half of dorsal surface of disc with more or less evenly scattered whitish spots and incomplete ocelli <i>A. maculatus</i>
4a	Dorsal surface of mid-disc with darker stripes, and rear half of disc with numerous ocelli of varying size
4b	Dorsal surface, from head to rear of disc, with pale, evenly spaced crossbars (faded in larger individuals)

#### Aetomylaeus bovinus (Geoffroy Saint-Hilaire 1817)

#### Bull ray

*Myliobatis bovina* Geoffroy Saint-Hilaire 1817: no page number, Pl. 26, Fig. 1 (off Alexandria, Egypt, Mediterranean Sea).

- ?*Aetomylus huletti* Smith 1953: 513, Fig. 77a (KwaZulu-Natal, South Africa).
- *Pteromylaeus bovinus*: Barnard 1925; Smith 1949\*; Wallace 1967\*; Van der Elst 1988\*; Compagno *et al.* 1989\*; SSF No. 28.3\* [1995]; Heemstra & Heemstra 2004\*.

Aetomylaeus bovinus: Weigmann 2016; White & Last 2016.

Pectoral-fin anterior margins strongly convex. Snout elongate, relatively slender, and rostral lobe fleshy, duck-bill shaped. Posterior edge of internasal flap straight or shallowly notched; 7 rows of plate-like teeth in both jaws: central row widest, and 3 rows of small diamond-shaped teeth at each side. Pelvic fins narrow. Dorsal-fin origin over pelvic fins. Tail 1.6–1.8 times disc length; 1–4 small serrated stinging spines just behind dorsal fin. Skin rough to touch on head, tail and along midback, but smooth on snout, outer angles and posterior margins of disc, and pelvic fins. Males with short conical horn above each eye.

Body brown dorsally, with 7 or 8 irregular transverse pale blue-grey crossbars over disc and head (faded in adults and darker in juveniles, stripes reportedly become more intense upon capture due to stress); uniformly white ventrally. Attains 180 cm DW.

**DISTRIBUTION** Mediterranean Sea and southern Africa to East Africa: Namibia in southeastern Atlantic, to South Africa, Mozambique and Tanzania (Zanzibar) in WIO.

PLATE 78





Aetomylaeus maculatus. Source: Bleeker 1852

**DISTRIBUTION** Indo-Pacific. WIO: Arabian Sea, Persian/ Arabian Gulf (Iran), India and possibly Sri Lanka.

**REMARKS** Little known. Males mature at 54–72 cm DW. Occurs predominantly inshore; once commonly found near reefs and in estuaries and mangroves, but has become uncommon in parts of its range. Incidental catch in inshore demersal fisheries; sold in fish markets in Southeast Asia. IUCN Red List conservation status Endangered.

#### Aetomylaeus milvus (Valenciennes 1841)

Ocellate eagle ray

PLATE 77

PLATE 77

*Myliobatis milvus* Müller & Henle (ex Valenciennes) 1841: 178 (Red Sea); Klunzinger 1871.

Aetomylaeus maculatus: ?Munro 1955\*.

Aetomylaeus milvus: Compagno & Last 1999\*; Bonfil & Abdallah 2004\*; White & Last 2016.

Disc width about twice disc length, with concave posterior margins and broadly convex anterior margins. Head relatively wide, but rostral lobe blunt, subtriangular, not greatly protruding. Eyes proportionally large. Pelvic fins slightly protruding beyond posterior margin of disc. Dorsal-fin origin just before or over pelvic-fin insertions.

Body brownish dorsally, with transverse darker bands at mid-disc, extending from centre to outer margins of pectoral fins, and with numerous greenish irregular ocelli posteriorly; uniformly white ventrally; tail strongly banded. Attains at least 37 cm DW.

Aetomylaeus bovinus. (South Africa).

**REMARKS** Until recently this species was placed in the genus *Pteromylaeus*, now considered a synonym of *Aetomylaeus* (White 2014). Litters of 4–6; size at birth ~48 cm DW. Males mature at ~95 cm DW, and females at ~110 cm DW. Found over sandy and muddy bottoms, from shoreline to ~150 m deep; also enters estuaries and lagoons. Feeds on bony fishes, crustaceans (crabs), squid and molluscs. A bycatch in various artisanal and commercial fisheries, and often caught by shore anglers; powerful swimmer able to leap from water.

### Aetomylaeus maculatus (Gray 1834)

Mottled eagle ray

*Myliobatus maculatus* Gray 1834: no page number, Pl. 101 (Penang, Malaysia).

*Myliobatis maculatus*: Annandale 1909; Blegvad & Løppenthin 1944. *Aetomylaeus maculatus*: ?Munro 1955\* [probably *A. milvus*];

Compagno & Last 1999\*; White et al. 2006\*.

Pectoral fins falcate, with posterior margins concave and anterior margins slightly convex. Rostral lobe triangular, relatively short. Eyes slightly bulging, slightly smaller than spiracles. Pelvic fins slender, protruding beyond posterior margin of disc. Dorsal-fin origin just behind (rarely over) pelvic-fin insertions. Tail whip-like, very long.

Body brown dorsally, with whitish spots and irregular ocelli on rear half of disc and pelvic fins; uniformly white ventrally; tail with alternating pale and dark bands. Attains at least 78 cm DW.



Aetomylaeus milvus. Source: Bleeker 1852

**DISTRIBUTION** Indo-Pacific. WIO: Red Sea, Persian/Arabian Gulf, Pakistan, India and possibly Sri Lanka.

**REMARKS** Small-sized, sparsely recorded and poorly known. Found inshore. Of questionable taxonomic validity, as it may represent the juvenile stage of *A. vespertilio* or *A. maculatus*.

#### Aetomylaeus nichofii (Bloch & Schneider 1801)

Banded eagle ray

PLATES 77 & 78

*Raja niehofii* Bloch & Schneider 1801: 364 ('East Indies')
[spelled *nichofii* in text but corrected in corrigenda, p. 579]. *?Raja fasciata* Shaw 1804: 286, Pl. 143 [no locality given]. *Myliobatis nieuhofii*: Annandale 1909.

*Myliobatis nieuhofii cornifera* Annandale 1909: 52, Pl. 2, Fig. 4a (India). *Myliobatis nichofii*: Blegvad & Løppenthin 1944\*.

?Aetomylus huletti Smith 1953: 513, Fig. 77a (KwaZulu-Natal, South Africa).

Aetomylaeus nichofii: Munro 1955\*; Qureshi 1972\*; Randall 1995\*;

Compagno & Last 1999\*; White *et al.* 2006\*; Last & Stevens 2009\*; White & Last 2016.

Disc width almost twice disc length. Internasal flap skirtshaped with fine fringe. Dorsal-fin origin just before or over pelvic-fin insertions. Tail long, whip-like, 1.5–2 times disc length. Skin normally smooth, without thorns; large adults may have minute, widely spaced denticles on disc.

Body bronze or greyish brown background dorsally, with transverse bands of pale colour on disc (usually 5 bands) and head (usually 2 bands) in adults (bands not evident in preserved specimens), and juveniles with darker, broader, more irregular bands; white ventrally; tail grey at base, darker towards tip. Attains 72 cm DW, 100 cm TL.



Aetomylaeus nichofii, 51 cm DW, 95 cm TL, adult male (Pakistan). Source: Capapé & Desoutter 1979

**DISTRIBUTION** Indo-Pacific. WIO: Mozambique, Persian/ Arabian Gulf (Iran and Oman), Pakistan, India, Sri Lanka and Maldives.

**REMARKS** Litters of 4 or more; size at birth ~17 cm DW. Males mature at 39–42 cm DW. Wide-ranging but increasingly uncommon; occurs inshore and offshore, to at least 70 m deep, and sometimes in brackish waters. Feeds on worms, crustaceans, snails and bony fishes. Marketed throughout most of its range. IUCN Red List conservation status Vulnerable.

### Aetomylaeus vespertilio (Bleeker 1852)

Ornate eagle ray

Myliobatis vespertilio Bleeker 1852: 85 (Jakarta, Java, Indonesia). Aetomylaeus vespertilio: SSF No. 28 [1995]; Compagno 1999; Compagno & Last 1999\*; Hennemann 2001\*; Bonfil & Abdallah 2004\*; White *et al.* 2006\*; Last & Stevens 2009\*; Bineesh *et al.* 2014;

PLATE 77

White & Last 2016.

Disc width about twice disc length. Pectoral-fin posterior margins deeply concave. Internasal flap skirt-shaped with indistinct fringe. Pelvic fins narrowly rounded. Dorsal-fin origin just behind or over pelvic-fin insertions. Tail extremely long, whip-like, up to 3 times disc length. Skin smooth, except adults with narrow band of tiny flat denticles along midline.

Body pale to dark brown dorsally, with irregular darker brown to black transverse lines on front half of disc and dark reticulate pattern on rear half of disc, head with few black spots and stripes, and white spots on pelvic fins and base of tail; white ventrally; tail mainly blackish. Attains 240 cm DW, at least 3.5 m TL.



Aetomylaeus vespertilio (SW India). KK Bineesh © CMFRI

**DISTRIBUTION** Indo-Pacific (widespread but scattered). WIO: Maldives, South Africa (KwaZulu-Natal) to Arabian Sea and Red Sea.

**REMARKS** Large-sized; naturally uncommon and rarely observed, hence poorly known. Males mature at ~170 cm DW. Occurs inshore on continental shelf, over sandy bottoms, in muddy bays and on coral reefs, to ~110 m deep. Caught in low numbers in demersal fisheries and marketed. IUCN Red List conservation status Endangered.

#### GENUS Myliobatis Cuvier 1816

Fleshy lateral ridge supporting snout continuous with pectoralfin origins; pectoral-fin tips blunt. Posterior edge of internasal flap relatively straight or slightly concave, not deeply notched; 7 rows of plate-like teeth in each jaw: central row widest, and 3 rows of diamond-shaped teeth at each side. Dorsal-fin origin behind pelvic-fin tips; prominent stinging spine just behind dorsal fin. About 11 species, 1 in WIO.

### Myliobatis aquila (Linnaeus 1758)

#### Common eagle ray

PLATE 78

- *Raja aquila* Linnaeus 1758: 232 (Mediterranean Sea and northeastern Atlantic).
- *Myliobatis cervus* Smith 1935: 169, Fig. 1 (Knysna, Western Cape, South Africa).
- *Myliobatis aquila*: Barnard 1925\*; Smith 1949; Wallace 1967\*;
- SSF No. 28.2\*; Van der Elst 1988\*; Compagno *et al.* 1989\*; Heemstra & Heemstra 2004\*; White & Last 2016.

Disc width about twice disc length, with mostly straight anterior margins and slightly concave posterior margins; pectoral-fin apices slightly rounded. Snout short, very broad, rounded, and with small anterior protuberance. Pelvic fins large, truncate. Dorsal fin entirely behind pelvic fins, rounded at apex. Tail whip-like, about twice disc length; stinging spine slender and long. Males with small conical horn above each eye.

Body dark brown, olive-green or blackish dorsally, occasionally with irregular dusky black spots; white ventrally. Attains 150 cm DW.



Myliobatis aquila, 36 cm DW (South Africa). Source: SSF

**DISTRIBUTION** Mediterranean Sea, eastern Atlantic and Indian Ocean. WIO: South Africa (Western Cape) to southern Mozambique (Bazaruto), and possibly western Mascarenes.

**REMARKS** Common, but poorly known; a systematic review of populations is needed. Litters of 3–7, born in shallow water; size at birth 20–23 cm DW; gestation period 6–8 months. Males probably mature by ~55 cm DW, and females by ~60 cm DW. Found inshore and offshore, but normally in shallow waters of bays and estuaries and on rocky reefs, but recorded to 537 m deep off southern Africa; semipelagic, observed swimming in small groups near bottom. Feeds on crustaceans (crabs), molluscs, worms and bony fishes. A bycatch of various fisheries and a good sport fish; excellent for eating.

# FAMILY **AETOBATIDAE**

#### Pelagic eagle rays

Marcelo R de Carvalho and Mateus C Soares

Moderate-sized to very large rays (to ~9 m TL, 3 m DW; most species attain <200 cm DW) with lozenge-shaped disc much wider than long; head robust, projecting beyond disc; pectoral fins meet head at level of eyes, angular, with concave posterior margins and broadly rounded free rear tips. Fleshy lateral ridge supporting snout not continuous and situated well below pectoral-fin origins at level of eyes. Head relatively narrow, thick and elevated from disc. Snout elongate and oval, with fleshy rostral lobe. Eyes lateral, moderately large and anteromedial to spiracles; spiracles dorsolateral and elongated, visible in dorsal view, margins smooth without papillae or elevated rims. Mouth narrow, transverse, straight, and without prominent knobs, depressions or lip folds. Nostrils relatively small and slightly elongate, just anterior to mouth, and connected to mouth by rather large oronasal grooves; internasal flap broad, overlapping mouth, posterior edge deeply notched with inverted V-shape. Teeth large and blunt, greatly flattened, laterally expanded, plate-like, and chevron-shaped; both jaws with a single tooth row in postnatal specimens, lacking lateral teeth.



Aetobatus narinari, 60 cm DW, ventral head with V-shaped notch on internasal flap (Mozambique).

Five pairs of small gill slits on underside of front half of disc. No electric organs. Pelvic fins slightly longer than wide, not bilobed, projecting past disc, and posterior margins rounded. Tail long (~1.5–3 times disc length), filamentous and whiplike, cylindrical and without lateral skin folds; 1 small dorsal fin, raked and with rounded-angular apex, positioned just behind pelvic-fin insertions, with short free rear tip. Stinging spine(s) close behind dorsal fin, relatively long. No caudal fin. Dorsal surface almost entirely smooth above and below. Body uniformly brown to greenish, or grey dorsally with white spots or ocelli; generally pale to whitish ventrally.

Circumglobal in most tropical and subtropical seas, inhabiting relatively shallow waters of continental and insular shelves; pelagic, solitary, in small groups, or schools of up to~200 individuals. The powerful expanded, wing-like pectoral fins of these active rays allows the acceleration required to reach high speeds and leap from the water; also found resting on the bottom, raising their body with their pectoral fins. Viviparous with histotroph (uterine secretions that nourish embryos), with litters of usually up to 4. In general biology, feeding and reproduction, this family is very similar to Myliobatidae (see Myliobatidae for details). Species of the *Aetobatus narinari* complex are common in public aquariums.

One genus with 5 species; 3 species in WIO.

#### GENUS Aetobatus Blainville 1816

See family account.

#### **KEY TO SPECIES**

1a 1b	Dorsal surface uniformly coloured, without conspicuous markings; snout elongate, tapering and acute <i>A. flagellum</i> Dorsal surface with spots or ocelli; snout less elongate, less acute
2a	Dorsal background colour yellowish brown; entire surface of disc covered with spots or ocelli
2b	Dorsal background colour greenish brown or greenish grey; sometimes with spots or ocelli only on rear margins of disc

#### Aetobatus flagellum (Bloch & Schneider 1801)

Longheaded eagle ray

PLATE 79

*Raja flagellum* Bloch & Schneider 1801: 361, Pl. 73 (Coromandel coast, India).

Aetobatis flagellum: Annandale 1909\*.

Aetobatus flagellum: Compagno & Last 1999\*; Bonfil & Abdallah 2004\*; White *et al.* 2006\*; White & Last 2016.

Disc diamond-shaped, with angular pectoral fins, much broader than long, and with concave posterior margins. Snout conspicuously elongate and pointed, narrowly tapering ahead of eyes. Eyes small, slightly oval, ventrolateral; spiracles large, elliptical. Internarial flap long, with pronounced median notch posteriorly. Teeth in up to 6/13 rows; lower tooth plate very broad, ~<sup>2</sup>/<sub>3</sub> mouth width. Pelvic fins protrude beyond posterior margin of disc. Dorsal fin small, its origin behind pelvic-fin insertions. Tail very long and slender, whip-like; stinging spine elongate, slender.

Body uniformly dark greenish brown to brown dorsally, unspotted, without conspicuous markings; uniformly white ventrally. Attains at least 57 cm DW, 126 cm TL. **DISTRIBUTION** Indo-Pacific (disjunct); records from eastern Atlantic and Hawaii in central Pacific require verification. WIO: Red Sea, Persian/Arabian Gulf (Kuwait) and India.

**REMARKS** Naturally uncommon and little known, but biology probably similar to *A. narinari*. Found inshore on continental shelf and in brackish water. Likely caught in local demersal fisheries. IUCN Red List conservation status Endangered.

#### Aetobatus narinari (Euphrasen 1790)

Spotted eagle ray

PLATES 78 & 79

*Raja narinari* Euphrasen 1790: 217, Pl. 10 (St Barthélemy I., West Indies; Brazil).

Aetobatis narinari: Klunzinger 1871; Day 1878\*; Annandale 1909;

Gilchrist & Thompson 1911; Barnard 1925; Blegvad & Løppenthin 1944\*; Randall 1995\*.

Stoasodon narinari: Smith 1949\*.

Aetobatus narinari: Munro 1955\*; Wallace 1967\*; Qureshi 1972\*;
SSF No. 28.1\*; Van der Elst 1988\*; Compagno *et al.* 1989\*;
Debelius 1993\*; Randall 1995\*; Compagno & Last 1999\*; Debelius 1999\*;
Hennemann 2001\*; Bonfil & Abdallah 2004\*; Heemstra & Heemstra 2004; White *et al.* 2006\*; Fricke *et al.* 2009; Last & Stevens 2009\*;
White 2014; White & Last 2016.

Head and body thick; snout and rostral lobe relatively long. Pectoral fins angular; anterior margins oblique to body axis, posterior margins slightly concave. Spiracles large, set over pectoral-fin origins. Posterior edge of internasal flap notched, inverted V-shape, and fringed; 1 row of plate-like teeth in each jaw, lower teeth protruding. Pelvic fins narrowly rounded. Dorsal fin over pelvic fins. Tail extremely long, up to 3 times disc length. Skin smooth. Body blue to black dorsally (in WIO region), with many white spots or rings; uniformly white ventrally; tail black. Attains 3.3 m DW, possibly more; ~8.8 m TL (assuming undamaged tail).

**DISTRIBUTION** Circumglobal in most tropical and subtropical seas (likely a species-complex). WIO: South Africa to Red Sea, Gulf of Oman, Persian/Arabian Gulf, Pakistan, India, Sri Lanka, Maldives, Madagascar, Seychelles and Réunion.

**REMARKS** Recent molecular studies suggest A. narinari may represent two or more species, and colour differences can be noted between populations in Atlantic, Indian and Pacific oceans. The nominal species Myliobatus ocellatus Kuhl 1823 is now employed as a senior synonym of A. narinari in the Indo-Pacific, but it is not clear whether this should apply to the species in WIO. Biology poorly known. Females mate with up to 4 different males within 1 hour; litters of up to 4; size at birth 17-36 cm DW; gestation period 12 months or less. Males mature at ~100 cm DW, and females at ~214 cm DW, at age 4-6 years for both sexes. Occurs near reefs, beaches, estuaries and bays, to ~60 m deep, and occasionally found in open ocean or in estuaries and lagoons. Active and agile swimmer near surface and able to rapidly accelerate and leap from water; observed singly, in small groups, as well as in schools of over 200 individuals. Uses its rostral lobe and exposed tooth plate to hunt benthic invertebrates (shrimps, crabs, clams, snails, worms, ovsters, octopus, squid and sea urchins). Preved on by sharks which tend to follow pregnant eagle rays to feed on newborn pups. Caught in artisanal fisheries throughout its range. A powerful fighter for anglers, but its meat is not much appreciated. Common in public aquariums. IUCN Red List conservation status Near Threatened.



Aetobatus narinari. Source: Bleeker 1877

# Aetobatus ocellatus (Kuhl 1823)

#### Ocellated eagle ray

PLATE 79

Myliobatus ocellatus Kuhl in Van Hasselt 1823: 316 (Jakarta, Java, Indonesia). Aetobatis guttata: Annandale 1909. Aetobatus flagellum: Misra 1962\*. Aetobatus guttatus: Compagno & Last 1999\*. Aetomylaeus ocellatus: Sujatha 2002.

Aetobatus ocellatus: White & Last 2016.

Disc much broader than long, acute at tips. Head broad, much broader than snout; snout moderately elongate and not very acute. Pectoral-fin anterior margins relatively straight, nearly perpendicular to body axis, or slightly convex; posterior margins slightly concave; fin bases rather narrow. Spiracles large. Lower tooth band ~½ mouth width. Pelvic fins protrude beyond posterior margin of disc. Dorsal fin behind pelvicfin insertions. Tail long and whip-like; stinging spine slender, elongate.

Body greenish brown to grey dorsally, with white spots usually more numerous on rear half of disc, but pattern of spots highly variable; uniformly white ventrally. Attains at least 125 cm DW, 4 m TL.

**DISTRIBUTION** Indian Ocean and western central Pacific. WIO: Pakistan, India and Sri Lanka.

**REMARKS** The name *Aetobatus ocellatus* was recently resurrected to be used in place of *A. narinari* in the Indo-Pacific region. However, whether the WIO specimens are conspecific with *A. ocellatus* from the western central Pacific, from where it was described, needs to be determined. Poorly known, but biology probably similar to other myliobatids.

# FAMILY RHINOPTERIDAE

#### Cownose rays

Brett A Human

Moderate- to large-sized (up to 210 cm DW), heavy-bodied rays with head depressed but elevated above disc, and forehead strongly indented with associated pair of lobe-like rostral flaps originating under spiracles and opposite mouth, continuing to front of head and uniting at front indentation. Disc rhomboidal, much wider than long (disc width 1.5–2 times disc length). Pectoral-fin origins posterior to nostrils and spiracles, fin tips acutely angular, and posterior margins concave. Eyes and spiracles lateral; 5 pairs of short gill slits on underside of head. Snout short, indented, and without rostral cartilage; internasal flap reaches mouth, with serrate posterior edge. Mouth ventral, transverse; teeth plate-like, arranged in 7 or more rows. Pelvic fins rounded or angular, and not bilobed; claspers short and stout. Dorsal fin small, prominent, its origin at base of tail, near pelvic-fin insertions. Tail filamentous, cylindrical, subequal to or much longer than disc, and without skin folds; no caudal fin. Dorsal surface smooth and without thorns, or with small denticles; 1 or more short stinging spines at base of tail and just behind dorsal-fin free rear tip (1 spine in WIO representatives). No electric organs.

Reproductive mode viviparity with histotroph, with uterine secretions nourishing few embryos. Highly social rays, schools of hundreds to even thousands of individuals have been observed, with such schools so far associated with breeding and dominated by females (when sex ratios have been reported). Some species are known to undertake long migrations. The gregarious nature of cownose rays makes them susceptible to various commercial net-gear fisheries (trawls, gillnets, seine nets), with accounts of thousands in a single catch; they are also caught by recreational anglers and exploited for the aquarium trade.

Poorly known group taxonomically, although easily recognisable by their blunt, anteriorly indented heads and odd-looking rostral flaps. A revision of this family would probably result in fewer valid species as some species currently recognised may be merely geographic variants of others. The primary character historically used to separate species (the number of tooth rows) has proven to be highly variable within a species; moreover, dentition is likely to be sexually dimorphic (as in *Rhinoptera javanica*), particularly in reference to tooth width versus length in the middle rows in both jaws, with females probably having wider teeth than males. However, head width and tail length may prove to be useful diagnostic characters. Circumglobal in tropical to warm-temperate seas. One genus with ~10 species; 2 species in WIO.

#### GENUS Rhinoptera Kuhl 1829

Diagnosis as for family. A taxonomically perplexing group, with  $\sim 10$  species currently recognised, but probably fewer are valid. The following key is highly provisional pending the discovery of more reliable characters.

#### **KEY TO SPECIES**

- 1b
   Tail length equal to or greater than disc length;

   tooth width <5.6 times tooth length in middle row</td>
   of upper jaw

   *R. javanica*



Rhinoptera javanica, 65 cm DW (S Mozambique).

#### Rhinoptera javanica Müller & Henle 1841

#### Flapnose ray

PLATE 79

Rhinoptera javanica Müller & Henle 1841: 182, Pl. 58 (Java, Indonesia [Malabar coast, India]); Day 1878\*, 1889\*; James 1962\*, 1970\*; Wallace 1967\*; Srinivasarengan 1981; Bianchi 1985\*; SSF No. 28.4\*; Séret & McEachran 1986; Compagno et al. 1989\*; Cliff & Wilson 1994\*; Morón et al. 1998; Compagno & Last 1999\*; Vossoughi & Vosoughi 1999\*; Sujatha 2002; Bonfil & Abdallah 2004\*; White et al. 2006\*; White & Dharmadi 2007; Last & Stevens 2009\*.

Rhinoptera adspersa Müller & Henle (ex Valenciennes) 1841: 183 (India); Day 1878, 1889; Compagno & Last 1999\*.

Disc width 1.5–2 times disc length; tail 1–3 times disc length. Tooth width 2.5–5.6 times tooth length in middle row of upper jaw; tooth count 7-11/7-10.

Body uniformly dark brown to olive dorsally; white ventrally. Attains 150 cm DW.

**DISTRIBUTION** Indo-Pacific (widespread but range not well defined). WIO: South Africa, Mozambique, Seychelles, Arabian Sea, Gulf of Aden, Red Sea, Persian/Arabian Gulf, Pakistan, India and Sri Lanka.

**REMARKS** The diagnostic features provided here have been gathered from western and northern Indian Ocean references, as the morphometric values provided for this species from Australia are very different (e.g., tail 2.6-3.4 times disc length; tooth width 10-13 times tooth length in middle row of upper jaw). Apparently common throughout its range, but poorly known. Litters of up to 6 (generally 1 or 2); size at birth 31-61 cm DW, born in January and February in India. Pelagic; inshore to ~30 m deep, including in bays, estuaries, mangroves and near reefs. Migrates inshore daily to feed; diet

includes bivalves (clams, oysters) and crustaceans; can occur in large schools. Taken in artisanal fisheries and as bycatch in commercial net-gear fisheries and anti-shark nets; widely utilised. IUCN Red List conservation status Vulnerable.

# Rhinoptera jayakari Boulenger 1895

Short-tail cownose ray

PLATE 79

Rhinoptera javakari Boulenger 1895: 141 (off Muscat, Gulf of Oman); Randall 1995\*; Manilo & Bogorodsky 2003; Bonfil & Abdallah 2004; Last et al. 2010\*; Bogorodsky et al. 2014\*.

Rhinoptera javanica: Schwartz 1990; Al-Abdessalaam 1995\*. Rhinoptera sp. 1: White et al. 2006\*; White & Dharmadi 2007.

Disc width ~1.75 times disc length; tail ~<sup>2</sup>/<sub>3</sub> times disc length. Tooth width ~8 times tooth length in middle row of upper jaw; tooth count 9/9.

Body uniformly pale grey to olive-green dorsally; pale ventrally; tail darker. Attains 80 cm DW.



Rhinoptera jayakari (Mozambigue).

**DISTRIBUTION** Indo-Pacific. WIO: southern Red Sea (Jizan, Saudi Arabia), Gulf of Aden to Gulf of Oman, and southward to Mozambique and South Africa (KwaZulu-Natal); elsewhere to Indonesia, Japan and possibly Australia.

**REMARKS** The taxonomic validity of this species is questioned in Compagno's (1999) checklist of living elasmobranchs. Locally rare. Sometimes forms schools; observed in Arabian Sea off Oman apparently basking at surface, in groups of tens and amounting to several hundred individuals within a few nautical miles, presumably feeding on benthos in nearly anoxic waters created by Somali current, which may force the rays to the surface to recover. Extent of its range unknown due to confusion with *R. javanica*. Historically a locally discarded bycatch of artisanal and commercial fisheries, but the pectoral fins have become valuable in the shark-fin trade. Presumably feeds on molluscs and other benthic invertebrates.

# FAMILY MOBULIDAE

#### Devilrays

Andrea D Marshall

Small-sized to enormous (1–8 m DW), with rhomboidal or chevron-shaped disc, broader than long; head trapezoidal, protruding, with prominent forward-projecting cephalic lobes (extensions of pectoral fins, which are curled while swimming and unfurled when feeding); pectoral-fin tips acute, posterior margins concave; pelvic fins small and barely projecting beyond disc; tail whip-like, and some species with very small serrated spine at base; dorsal fin small, angular, at base of tail and over pelvic fins; no skin folds or caudal fin. Eyes and spiracles lateral; 5 pairs of short to wide gill slits on underside, and internal gill openings with unique fringed filter plates. Mouth broad, terminal or subterminal; teeth small, peg-like, embedded in bands on both jaws or on lower jaw only. Skin on dorsal and ventral surfaces usually covered with small denticles.

Pelagic rays with modified branchial filter plates enabling them to feed on small planktonic crustaceans, gastropods, coral and fish spawn, as well as small fishes. Typically inhabit shallow inshore waters, around reefs, bays, atolls and island groups, but also offshore reefs and seamounts; graceful, powerful swimmers, observed singly or in small or large groups, often feeding or basking at surface or interacting around aggregation sites. Their movement patterns and migratory habits are not well understood. Reproductive mode viviparity with histotroph, wherein embryos are nourished by uterine secretions; very low fecundity, with litters of only 1 or 2; the reproductive season often coincides with summer when water temperatures are highest. Presumably, mobulids cannot sustain targeted fisheries for long due to low reproductive potential and relatively small population sizes, yet there are increasing catches in directed fisheries, as bycatch, and incidentally in trawl fisheries. Urgent conservation measures for most species are needed. The meat is used for food, the skin for leather, and the branchial filter plates have particularly high value as medicinal products in Asia. Increasingly, some species are the subject of dive tourism (although human interactions without codes of conduct may affect their natural behaviour).

Mobulidae species occur worldwide in tropical and temperate seas. A complicated nomenclatural history, gaps in taxonomic and genomic sampling and poor representations of mobulids in museum collections has contributed to taxonomic confusion with species being regularly confused. Recent efforts to clarify long-standing taxonomic and phylogenetic uncertainties have resulted in significant changes to the taxonomic arrangement of the family and several species being synonymised. There are at least 10 distinct species in the currently recognised single genus *Mobula*, of which 7 occur in WIO. However, *Mobula* has been found to be paraphyletic, but additional research is still warranted to fully resolve the taxonomic uncertainty.

# GENUS Mobula Rafinesque 1810

See family account.

#### **KEY TO SPECIES**



#### **KEY TO SPECIES**



# Mobula alfredi (Krefft 1868)

#### Reef manta ray

PLATE 80

*Deratoptera alfredi* Krefft 1868: 3, 9, Fig. (entrance to Sydney Harbour, Australia).

Manta fowleri Whitley 1936: 182 (Line Is., central Pacific).

*Manta pakota* Whitley 1936: 183 (Marquesas Is.) [based on '*Pakoka*' in Pinchot 1930\*].

Manta alfredi: Marshall et al. 2009\*.

Mobula alfredi: White et al. 2017.

Disc width more than twice disc length. Spiracles small and furrow-like; branchial filter plates fused. Head very short, broad; cephalic lobes long, large, and ventral margin overlapping dorsal margin when rolled; rostral margin straight. Mouth terminal; lower-jaw teeth small, cusped, not overlapping, with ~6–8 rows, 142–182 files across width of tooth band, and total tooth count 900–1 500; upper jaw without teeth or rows of tiny peg-like denticles. Tail whip-like, short (<½ DW). Calcified mass with embedded stinging spine at base of tail greatly reduced and in most cases absent. Dermal denticles small, knob-like, evenly distributed.

Body black dorsally, with variable pale to white shoulder patches (anterior distal edges emanate posteriorly from spiracles, generally forming anterior-facing hook shape); mouth white to pale grey; cream to white ventrally, with variable blue-grey to black spots and shading across posterior half of disc and often medially between gill slits. Melanistic colour morph entirely black dorsally and predominantly black ventrally, except for variably sized white blaze along ventral midline (reports from Maldives). Leucistic colour morph extremely pale dorsally and with greatly reduced markings and shading ventrally (reports from southern Mozambique and Maldives). Attains 5 m DW.



*Mobula alfredi*, ~2.4 m DW, ventral and dorsal view (South Africa). Source: SSF (lower dorsal view)

**DISTRIBUTION** Fragmented populations occur in tropical to subtropical seas throughout the Indian Ocean and Pacific Ocean west of the Hawaiian Is. and French Polynesia. WIO: Red Sea to South Africa (Port St Johns, Eastern Cape), Madagascar, Oman to Persian/Arabian Gulf, India and Sri Lanka to Maldives and Chagos.

**REMARKS** Litters of 1, rarely 2; size at birth ~150 cm DW; gestation period ~1 year. Males mature at ~3 m DW, and females at ~3.5 m DW (maturity varies with region); reproductive periodicity commonly biennial or triennial; relatively long-lived, up to ~40 years. Found more commonly in shallow inshore waters, especially in association with areas of upwelling and high biological productivity, around coral and rocky reefs, bays, atolls and island groups, but also offshore reefs and seamounts. Sometimes seen in large groups; less migratory and with smaller home ranges than *M. birostris*, but can move seasonally in response to food abundance. IUCN Red List conservation status Vulnerable.

# Mobula birostris (Walbaum 1792)

#### Giant manta ray

Raja birostris Walbaum 1792: 535 [no locality given].

Ceratoptera ehrenbergii Müller & Henle 1841: 187 (Red Sea). Manta birostris: Bigelow & Schroeder 1953\*; Wallace 1967\*; SSF No. 29.1\*;

Compagno *et al.* 1989\*; Randall 1995\*; Ishihara *et al.* 2001\*; Manilo & Bogorodsky 2003; Bonfil & Abdallah 2004\*; Marshall *et al.* 2009\*. *Mobula birostris*: White *et al.* 2017.

Disc width more than twice disc length. Spiracles small and furrow-like; branchial filter plates fused. Head short, broad; cephalic lobes long, large, with ventral margin overlapping dorsal margin when rolled; rostral margin straight to slightly concave. Mouth terminal; teeth small, cusped, slightly overlapping, with ~12–16 rows, 220–250 files across width of tooth band, and total tooth count ~3 000–4 000; upper jaw without functional teeth but with at least 2 rows of tiny rough peg-like denticles. Tail whip-like, short (<½ DW). Calcified mass present at base of tail with embedded vestigial stinging spine with tip protruding 1–2 mm. Denticles small, dense, overlapping, forming long sagittally-oriented raised ridges and furrows on skin.

Body predominantly black or greyish blue dorsally, with angular shoulder patches with posterior-facing hook shapes; mouth dark grey to black; cream to white ventrally, with irregular dark spots or patches centrally on abdomen but without dark spots or patches medially between gill slits, and posterior margins of pectoral fins typically charcoal grey for length of fin. Melanistic colour morph entirely black dorsally and predominantly black ventrally, except for variably sized white blaze along ventral midline. Leucistic colour morph with white mouth, lighter colouration dorsally and little colouration ventrally (reports from southern Mozambique). Attains up to 7.1 m DW (anecdotal reports to ~9 m DW).



Mobula birostris, ~5 m DW, dorsal view (S Mozambique).



*Mobula birostris*, ~5 m DW, ventral view, different specimen from dorsal view (S Mozambigue).

**DISTRIBUTION** Circumglobal and widespread in tropical to temperate seas. WIO: from South Africa (East London) and Madagascar to the Red Sea, Oman to India to the south of the Maldives.

**REMARKS** The largest living ray. Litters of 1; estimated size at birth ~150 cm DW. Males mature at ~3.8 m DW, and females at ~4 m DW. Most often sighted along productive coastlines with regular upwelling, near oceanic island groups, and around offshore pinnacles and seamounts. Telemetry studies indicate M. birostris spends considerable time in deeper offshore waters, between 50-200 m, and is capable of periodic dives below 1 000 m. Typically encountered in shallow inshore waters >30 m. Appears to be migratory but often revisits aggregation areas (sightings in specific locations may be common, seasonal or sporadic). Not often found in large groups, and encountered by divers less frequently than M. alfredi. Sympatric with M. alfredi in a few locations, such as Mozambique and South Africa, but the two species exhibit different movement patterns and habitat use. Has high value in directed and artisanal fisheries, and taken as bycatch; increasingly the subject of dive tourism. IUCN Red List conservation status Endangered.

# Mobula eregoodoo Cantor 1949

Longhorned devilray

PLATE 81

*Dicerobatis eregoodoo* Cantor 1849: 1420 (Penang, Malaysia; Coromandel, India).

Mobula eregoodootenkee (non Bleeker 1859): Bonfil & Abdallah 2004\* [as *M. eregoodootenke*]; Last & Stevens 2009\*.

Mobula eregoodootenkee (non Cuvier 1829): Notarbartolo di Sciara 1987\*;
SSF No. 29.3 [1995]; Manilo & Bogorodsky 2003 [as *M. eregoodootenke*].
Mobula kuhlii (non Valenciennes 1841): White *et al.* 2017\* (in part).
Mobula eregoodoo: Notarbartolo di Sciara *et al.* 2019\*.

Disc wider than long; pectoral-fin anterior margins straight to weakly convex, posterior margins concave; head elongated, cephalic lobes long; rostral margin deeply concave. Spiracles very small, nearly circular, set below level of pectoral fins. Branchial filter plates reduced, each with 4–6 short lateral lobes; terminal lobe elongated, leaf-shaped and without ridges. Teeth with 1–3 (mostly 2) small narrow-based cusps on buccal edge of crown. Tail shorter than disc length, and base quadrangular; no stinging spine. Skin smooth, except some denticles at dorsal-fin base.

Body dark greyish brown dorsally; dorsal fin uniformly dark; white ventrally, but pectoral-fin anterior margins with dark blotch midway. Attains 130 cm DW.

**DISTRIBUTION** Indo-Pacific (widespread). WIO: Madagascar, South Africa (KwaZulu-Natal), to Red Sea, Oman, Persian/Arabian Gulf, Pakistan, India and Sri Lanka; elsewhere to Malaysia, Indonesia, Taiwan and Australia.

**REMARKS** Previously misidentified as *M. diabolus (non* Shaw 1804). *Mobula eregoodoo* was then synonymised with *M. kuhlii*, but Notarbartolo di Sciaria *et al.* (2019) recognised it as a valid species. Usually litters of one. Locally common but little known; found in coastal continental waters, and not recorded from oceanic islands or epipelagic zone. Appears to mostly feed on small fishes. Likely a bycatch of several fisheries. IUCN Red List conservation status Endangered.

#### Mobula kuhlii (Valenciennes 1841)

Pygmy or lesser devilray

PLATES 80 & 81

*Cephaloptera kuhlii* Valenciennes (ex Müller & Henle) 1841: 185, Pl. 59 (India); Duméril 1865\*.

*Mobula kuhlii*: Notarbartolo di Sciara 1987\*; Compagno *et al.* 1989\*; Randall 1995\*; Manilo & Bogorodsky 2003; White *et al.* 2006\*; Notarbartolo di Sciara *et al.* 2017; White *et al.* 2017\* (in part).

Disc width approximately twice length; pectoral-fin anterior margins straight, posterior margins concave; broad head with short cephalic lobes. Ventral margin of lobes not overlapping dorsal margin when rolled; rostral margin straight; mouth subterminal. Spiracles small, nearly circular, set below level of pectoral fins. Branchial filter plates with leaf-shaped terminal lobes. Tail can be longer than disc length when unbroken, and base quadrangular; no stinging spine. Skin distinctly smooth.

Body dark brown or grey dorsally often with a darker collar over posterior part of the head above spiracles (fades in dead specimens), white ventrally with or without pale brown to grey pectoral-fin tips; pectoral-fin anterior margins can be grey and sometimes with dark blotch mid-way; dorsal fin often whitetipped but can also be uniformly dark; tail counter-shaded dark dorsally and white ventrally. Attains 120 cm DW.



Mobula kuhlii, 111 cm DW, male (South Africa). Source: SSF

**DISTRIBUTION** Indo-Pacific (widespread but patchy). WIO: South Africa (KwaZulu-Natal) to Mozambique, Tanzania (Zanzibar), Somalia, Seychelles, Oman, India to Sri Lanka.

**REMARKS** Litters of 1; size at birth ~31 cm DW. Males mature at ~115 cm DW. Common; occurs inshore over continental shelf but not into epipelagic zone. Feeds on planktonic crustaceans, and possibly small fishes and cephalopods. Likely a targeted catch or utilised bycatch throughout its range. IUCN Red List conservation status Endangered.

#### Mobula mobular (Bonnaterre 1788)

Spinetail devilray

PLATE 81

Raia mobular Bonnaterre 1788:5 (Montredon, near Marseille, France). Cephaloptera japanica Müller & Henle 1841: 185 ([Nagasaki] Japan). Mobula japanica: Notarbartolo di Sciara 1987\*, 1988; SSF No. 29.4 [1995];

Manilo & Bogorodsky 2003; Bonfil & Abdallah 2004\*; White *et al.* 2006\*; Last & Stevens 2009\*.

Mobula mobular: White et al. 2017\*.

Disc width twice length; pectoral-fin anterior margins straight to slightly convex, posterior margins concave; head and cephalic lobes short, and ventral margin of lobes not overlapping dorsal margin when rolled; rostral margin relatively straight; mouth subterminal. Spiracles ovoid, transverse slits, set above level of pectoral fins. Branchial filter plates separate from each other; 3rd arch with 85–95 plates on each face, each with 18–25 lateral lobes, and terminal lobe leafshaped with median longitudinal ridge. Pelvic fins distinctly wedged-shaped; conspicuous abdominal pores near pelvic-fin insertions. Dorsal fin high, relatively advanced on body. Tail very long (subequal to or longer than DW), with prominent white denticles entire length; serrated stinging spine exposed (~3–5 cm long) and base embedded in calcified mass. Dermal denticles dense, but sparser on ventral surface of pectoral fins.

Body bluish black dorsally, with pale crescent-shaped shoulder patches (distinct in juveniles, fading in adults); cephalic lobes grey with black tip, outer surface white; dorsalfin tip white; white denticles laterally on tail; white ventrally, sometimes with dark patches in adults (but pectoral-fin anterior margins never dark). Attains 320 cm DW.

**DISTRIBUTION** Probably circumglobal in tropical to warmtemperate seas (distribution not completely defined). WIO: South Africa (KwaZulu-Natal), Gulf of Aden, Persian/Arabian Gulf, Oman to Pakistan, India and Sri Lanka.

**REMARKS** Biology poorly known. Litters of 1. Occurs inshore and offshore and is capable of exploiting deepwater habitats. Feeds largely on planktonic crustaceans and occasionally small fishes. A bycatch of other fisheries and increasingly targeted for human consumption. IUCN Red List conservation status for *Mobula mobular* in the Mediterranean is Endangered, and before *Mobula japonica* became a junior synonym of *M. mobular*, it was listed as Near Threatened globally and Vulnerable throughout Southeast Asia.

#### Mobula tarapacana (Philippi 1892)

Sicklefin devilray

PLATE 81

*Cephaloptera tarapacana* Philippi 1892: 8, Pl. 3, Fig. 2 (Tarapacá Region, Chile).

*Mobula tarapacana*: Notarbartolo di Sciara 1987\*, 1988; Compagno *et al.* 1989\*; SSF No. 29.5 [1995]; Bonfil & Abdallah 2004\*; White *et al.* 2006\*; White *et al.* 2017.

Disc wider than long; pectoral-fin anterior margins weakly concave, becoming broadly convex towards slender apex, and posterior margins concave (fins sickle-shaped); head long; cephalic lobes short and thick, and ventral margin of lobes not overlapping dorsal margin when rolled; rostral margin broadly concave; mouth subterminal. Spiracles longitudinal slits, set above plane of pectoral fins, and inside surface corrugated by several transverse skin folds; crease from eye to origin of spiracle. Branchial filter plates fused; 3rd gill arch with 50-65 plates, each with 15-22 lateral lobes, and terminal lobe broadly rounded distally and concave proximally. Teeth relatively large and tessellated (not overlapping). Pelvic-fin area uniquely narrow, elongated, with conspicuous abdominal pores near pelvic-fin insertions. Tail short (<1/2 DW), compressed at base and distinctly rounded towards tip; no stinging spine. Skin rough, covered with long, thick, posteriorly pointing thorn-like denticles.

Body olive-green to bronze dorsally; dorsal fin uniformly dark; ventral surface white anteriorly and grey posteriorly

PLATE 81

(separated by irregular demarcation line), underside of rostrum to lower jaw silvery, inner cephalic lobes grey with black tip, dark eye-spots sometimes near pectoral-fin tips, and rarely with dark patches on gill slits.Attains 3.28 m DW (unconfirmed reports to 3.70 m DW).



Mobula tarapacana (WIO). Composite

**DISTRIBUTION** Probably circumglobal (but scattered) in tropical to warm-temperate seas. WIO: South Africa (Eastern Cape) to Red Sea, India, Sri Lanka and Maldives.

**REMARKS** Litters of 1; size at birth 105–132 cm DW. Males mature at 240–250 cm DW, and females at 270–280 cm DW. Undertakes long migrations and, like some other species of *Mobula*, can make deep dives >1 800 m. More oceanic pelagic occurrence that other mobulids, but occasionally coastal. Feeds on planktonic crustaceans and small fishes; larger mesh size of branchial filter plates suggests it is ichthyophagous or a more general feeder than other mobulids. A bycatch of pelagic gillnet, purse-seine and longline fisheries, and a minor component in the increasing mobulid catches elsewhere in its range. IUCN Red List conservation Endangered.

#### Mobula thurstoni (Lloyd 1908)

#### Bentfin devilray

*Dicerobatis thurstoni* Lloyd 1908: 179, Fig. 3, Pl. 4 (India, northern Indian Ocean).

*Mobula thurstoni*: Notarbartolo di Sciara 1987\*, 1988\*; Compagno *et al.* 1989\*; Randall 1995\*; SSF No. 29.6 [1995]; Manilo & Bogorodsky 2003; White *et al.* 2006\*; Last & Stevens 2009\*; White *et al.* 2017.

Disc width approximately twice length; pectoral-fins sinusoidal (margins somewhat wavy), anterior margins convex and posterior margins notably concave (fin with prominent double curvature); head and cephalic lobes short, and ventral margin of lobes not overlapping dorsal margin when rolled; rostral margin relatively straight; mouth subterminal. Spiracles small, nearly circular, set below level of pectoral fins. Branchial filter plates 70–75, separate, each with 15–20 lateral lobes, and terminal lobe elliptical or leaf-shaped. Teeth small and widebased, overlapping in adults. Tail ~60% disc length, flattened at base but overall ovoid in cross-section; no stinging spine. Dermal denticles sparse.

Body dark blue to black dorsally, and silvery grey around eyes; anterior margins of pectoral-fin sometimes with white stripe, and trailing edge of ventral surface of fin sometimes with dark metallic green patch (not in Indo-Pacific and Atlantic specimens); ventral surface of pectoral and pelvic fins often with dark posterior margins; often with 2 conspicuous abdominal pores at level of pelvic insertion; dorsal fin whitetipped; mostly white ventrally (of variable intensity), and often silvery near pectoral-fin tips; tail counter-shaded dark dorsally and white ventrally. Attains 180 cm DW.

**DISTRIBUTION** Probably circumglobal (scattered) in tropical to subtropical seas. WIO: South Africa (Eastern Cape and KwaZulu-Natal), Madagascar, Maldives to Red Sea, Oman to India and Sri Lanka.

**REMARKS** Litters of 1; size at birth 65–85 cm DW; gestation period probably 1 year. Matures at ~150 cm DW; reproductive periodicity reportedly every 2 or more years. Most commonly found in shallow neritic waters, to ~100 m deep, but sometimes seen several kilometres offshore over deeper waters; not schooling, observed singly or in small groups. Feeds on planktonic crustaceans (euphausiids, mysids). IUCN Red List conservation status Endangered globally but suspected to be Vulnerable in the Indian Ocean and throughout Southeast Asia.

# ORDER CHIMAERIFORMES

# FAMILY CALLORHINCHIDAE

#### Elephantfishes

Jenny M Kemper and David A Ebert

Highly distinctive cartilaginous fishes with relatively elongate, torpedo-like body (to 125 cm TL); prominent, fleshy, ploughshaped snout, terminal end transverse and leaf-shaped. Single pair of gill openings covered by soft flap; head with well-developed system of mucous canals and sensory pores; lateral-line canals enclosed, pored (not open grooves), readily visible and continuous from behind eyes to peduncle. Eyes small, set high on head; no spiracles. Mouth small, directed anteroventrally, with plate-like dentition; tooth plates large and robust, 2 pairs in upper jaw, 1 pair in lower jaw. Pectoral fins large, fan-like; bases of paired fins thick and fleshy, and outer fin webs thin and flexible. Two widely separated dorsal fins: 1st dorsal fin erectile, high and triangular, preceded by long, serrated, moveable and mildly venomous spine; 2nd dorsal fin short-based (relative to other Chimaeriformes), its length nearly equal to pectoral-fin to pelvic-fin space, anterior onethird of fin tall, sloping posteriorly to a low, evenly tall fin. Anal fin tall, narrow-based, separated from caudal fin only by deep notch. Caudal fin heterocercal, narrow and arched, about as long as pectoral fins; no caudal filament. Skin smooth.

Reproductive mode oviparity; females produce large, horny, spindle-shaped egg cases. Elephantfishes display relatively fast growth, early maturity and high fecundity. Occur close inshore to continental slope. Feeble swimmers; the distinctive snout is used to probe the bottom for feeding on invertebrates and small fishes. Caught commercially, as bycatch, and by shore anglers. Common in temperate oceans of Southern Hemisphere. One genus, *Callorhinchus* Lacepède 1798, with 3 species, 1 in WIO.

# Callorhinchus capensis Duméril 1865

Cape elephantfish

PLATE 82

Callorhynchus capensis Duméril 1865: 695, Pl. 13, Fig. 5 (Cape of Good Hope, South Africa); SFSA No. 95\*; Krefft 1990.

*Callorhinchus capensis*: SSF No. 34.1\*; Compagno, Ebert & Smale 1989\*; Compagno, Ebert & Cowley 1991; Bianchi & Carpenter *in* Bianchi *et al.* 1993\*; Freer & Griffiths 1993; Heemstra & Heemstra 2004\*.

Diagnosis as for family. Pelvic claspers simple, rod-like, without dilated tips or denticles; frontal tenaculum club-like, with small spines near apex; pre-pelvic tenacula relatively complex, spoon-shaped, with medial barbs on inner surface. Egg cases elongate, oval, with broad lateral flanges.

Body silvery, with dusky spots or blotches on head and trunk; fin webs brown. Attains 122 cm TL.



*Callorhinchus capensis*, 22 cm TL, male, ventral view, and egg case. Source: SFSA



*Callorhinchus capensis*, 42 cm to rear of 2nd dorsal fin, male (South Africa). Source: SSF

**DISTRIBUTION** Southern Africa: Namibia to Cape of Good Hope in southeastern Atlantic, to South Africa (abundant on south coast, rarely to KwaZulu-Natal) in WIO.

**REMARKS** Found from near shore and in shallow bays to continental slope, near bottom, usually over soft substrate, to 366 m deep (mostly <120 m). One egg per oviduct, up to 22 eggs per female per season; size at birth ~13 cm TL. Males mature at ~43 cm TL, and females at 46–50 cm TL; reproductive periodicity year-round, peaking in summer. Feeds on invertebrates (echinoderms [urchins], bivalves, gastropods, crustaceans [mantis shrimp], polychaetes) and small bony fishes. Caught in directed bottom-set gillnet fisheries, and a bycatch of demersal trawl fisheries; sold for human consumption. In South Africa, commonly known as St Joseph.

# FAMILY RHINOCHIMAERIDAE

#### Longnose chimaeras

Jenny M Kemper and David A Ebert

Medium- to large-sized cartilaginous fishes with somewhat compressed, elongate body (to ~130 cm TL), tapering to narrow diphycercal tail; snout elongated into cylindrical or flattened pointed projection, not plough-shaped. Mouth ventral, with beak-like tooth plates: two pairs in upper jaw, one pair in lower jaw; dentition smooth, either without hypermineralised tritors (grinding surfaces) or with ridged hypermineralised tritors. Pair of single gill openings covered by soft flap; lateral-line canals open grooves. Two dorsal fins: 1st dorsal fin erectile, triangular, and preceded by tall, smooth or serrated spine; 2nd dorsal fin long and low, with straight or slightly undulating distal margin. Anal fin high and narrowbased, or absent. Caudal fin lanceolate; terminal filament present or absent; upper caudal margin with or without series of tubercles (or denticles in males). Pelvic claspers slender and rod-like, with small, fleshy but denticulate tips; adult males also with single frontal tenaculum and paired pre-pelvic tenacula. Skin smooth and deciduous.

Reproductive mode oviparous; egg cases ovoid and large, with spindle-shaped capsule; fan-like lateral flanges, narrower than that of the callorhinchids, with a constricted central spindle. Rare to moderately common, benthopelagic, over continental slope and abyssal plains, generally inhabit deep water at ~100–2 000+ m; adults and juveniles may occupy different habitats. Minimal interest to fisheries, primarily as bycatch in bottom-trawl fisheries. Worldwide but scattered, in tropical to temperate seas. Three genera and 8 species; all 3 genera and 5 species in WIO, but only 2 genera, with at least 2 and possibly 3 species in WIO at depths of <200 m.





#### GLOSSARY

**tenaculum (pl. tenacula)** – an appendage on the forehead of male chimaeroid fishes, weakly differentiated to ridge-like in male juveniles (frontal tenaculum); additional small, retractable appendage on each side just in front of the pelvic fins with intromittent organs (claspers), of chimaeroid males, only partially differentiated in male juveniles, and hidden in a slit when retracted (pre-pelvic tenaculum).

#### GENUS Harriotta Goode & Bean 1895

Body elongate, tapering posteriorly to caudal filament; snout very long, relatively slender, with pointed and often upturned tip, and adult males often with rounded knobs on tip. Head convex in lateral profile; eyes small to moderately large. Mouth located below eyes; tooth plates thick, hypermineralised, with transverse ridges and tritors (except in small juveniles). Pectoral fins broad, tips sometimes reaching beyond pelvic-fin origins when depressed. First dorsal fin short-based, high, preceded by tall spine with serrations; spine as high as 1st dorsal fin and almost entirely free from its margin; 2nd dorsal fin low and long, either widely separated from or basally connected to 1st dorsal fin, and its distal margin convex to straight. No anal fin. Upper and lower margins of caudal fin either similar or lower margin longer and taller; upper edge without tubercles. Two species, 1 in WIO.

### Harriotta raleighana Goode & Bean 1895

#### Narrownose chimaera

Harriotta raleighana Goode & Bean 1895: 472, Pl. 19 (northwestern Atlantic, Gulf Stream); Jordan & Evermann 1900\*; Garrick 1971\*; Garrick & Inada 1975; Stehmann & Bürkel *in* Whitehead *et al.* 1984\*; SSF No. 33.1\*; Compagno *et al.* 1989\*; Krefft 1990; Compagno *et al.* 1991; Last & Stevens 1994\*; Didier 1995; Gomon 2008\*.

Relatively small-bodied; snout moderately long, slightly flattened with blunt tip, often bent upwards, its width greatest at midlength. Eyes moderately large, diameter about half distance between eye and dorsal spine. Mouth in front of or below eyes. Pectoral fins broad and short, apices rather angular and reaching beyond pelvic-fin origins when depressed. First dorsal fin tall, spine relatively straight and usually taller than 1st dorsal fin, and fin tip reaching at least to 2nd dorsal-fin origin when depressed; interdorsal space short; 2nd dorsal-fin mainly straight, base more than twice length of pelvic fins. Caudal-fin upper portion shorter than lower portion, but both distal margins similar in height; origin of lower margin approximately opposite 2nd dorsal-fin insertion; caudal filament long. Frontal tenaculum small; pre-pelvic tenacula with large spines along inner margin.

Body uniformly chocolate brown and skin deciduous; fin edges darker; pelvic fins brownish black. Attains 120 cm TL (including caudal filament).

**DISTRIBUTION** Atlantic and Pacific oceans (widespread), and Indian Ocean (not widely recorded). WIO: South Africa (off Cape Point) and Mozambique Seamount.

**REMARKS** Biology poorly known. Size at birth ~13 cm (body length without caudal filament); few juveniles ever collected. Primarily deep water, over upper continental slope, on or near bottom, at ~380–2 600 m, but with an unconfirmed report from 100 m deep; apparently fairly abundant where they occur. Feeds mostly on benthic invertebrates. Minor bycatch of deepwater bottom-trawl fisheries; used for its meat and as fishmeal.



*Harriotta raleighana*, male juvenile. Source: SSF, drawn from Fischer in Bigelow & Schroeder 1952

#### GENUS Neoharriotta Bigelow & Schroeder 1950

Diagnosis as for family. Snout tip without knobs or tubercles; eyes large; tooth plates thick and ridged with hypermineralised tritors. Anal fin separated from lower caudal fin only by deep notch; upper caudal margin without denticles in males. Three species, 2 in WIO.

#### **KEY TO SPECIES**

PLATE 82

- 1a
   Head and body uniform chocolate brown; caudal-fin filament short, less than snout length
   N. pinnata

#### Neoharriotta pinnata (Schnakenbeck 1931)

Sicklefin chimaera

PLATE 82

Harriotta pinnata Schnakenbeck 1931: 40, Figs. 6–9 (Walvis Bay, Namibia).
Neoharriotta pinnata: SSF No. 33.2\*; Compagno et al. 1989\*; Krefft 1990;
Compagno et al. 1991; Didier 1995; Didier & Stehmann 1996\*;
Manilo & Bogorodsky 2003; Ali et al. 2009\*; Akhilesh et al. 2011.

Diagnosis as for genus. Snout moderately slender, elongate, slightly flattened and blunt-tipped. Oral and preopercular lateral-line canals separated by large space. Pectoral fins short and broad. First dorsal-fin spine serrated; height of 2nd dorsal fin uniform. Pelvic fins rounded. Anal fin tall, curved. Caudal fin with short terminal filament. Frontal tenaculum of adult males very large.

Body uniformly chocolate brown. Attains 147 cm TL (including caudal filament).

**DISTRIBUTION** Eastern Atlantic (apparently widespread: Western Sahara to Namibia) and Indian Ocean. WIO: South Africa (off Cape Point), and Oman to India.

**REMARKS** Biology unknown and few adult voucher specimens have ever been collected. Probably matures at 50–55 cm (body length without caudal filament). Found in relatively shallower habitats, on continental shelf and upper slope, than other members of the family, on or near bottom, at 113–760 m. Likely feeds on small benthic invertebrates and fishes. Possibly a bycatch of deepwater fishery trawls; of some fisheries interest in India.

> *Neoharriotta pinnata*, 76 cm (to rear of 2nd dorsal fin), male (Namibia). Source: SSF

# Neoharriotta pumila Didier & Stehmann 1996

Arabian sicklefin chimaera

PLATE 82

Neoharriotta pumila Didier & Stehmann 1996: 957, Figs. 2-3 (off Socotra I., Arabian Sea); Manilo & Bogorodsky 2003.

Diagnosis as for genus. Small-bodied and tapering to caudal fin with a long filament. Snout broad at base and tapering to blunt point. Eyes large and horizontally ovoid. Preopercular and horizontal lateral-line canals separated by narrow space. Pectoral fins large, rounded, broad-based. First dorsal-fin spine serrated, upper half of spine free from fin, and fin as tall as or taller than spine; 2nd dorsal fin much lower than 1st, sloping posteriorly; interdorsal space short. Dermal denticles in space between 2nd dorsal fin and upper caudal lobe in adults of both sexes. Pelvic fins more rectangular than rounded, with straight distal margin. Anal fin slender with blunt tip, fin origin behind 2nd dorsal-fin insertion. Caudal fin with long terminal filament; upper portion of fin uniform in height, and margin without denticles; lower portion taller anteriorly, its origin before 2nd dorsal-fin insertion. Pre-pelvic tenacula each with 5 small denticles on medial edge, and four rows of 4 or 5 small curved denticles on ventral surface.

Body mottled grey-brown, snout and head darker; fins dark grey to black; paler ventrally and at bases of paired fins. Attains 65 cm TL (including terminal filament).

**DISTRIBUTION** WIO: Somalia and Socotra (Gulf of Aden and Arabian Sea), and possibly southwestern India.

**REMARKS** Rare, known only from localised populations, but may have a wider distribution at greater depths; inhabits outer continental shelf and upper to middle slope, on or near bottom, at 100–1 120 m. Matures at ~16 cm (body length without caudal filament). Presumably feeds on benthic invertebrates (bivalves and polychaetes) and fishes.



*Neoharriotta pumila*, ~45 cm TL (including caudal filament), female paratype (Socotra). Source: Didier & Stehmann 1996